# Chapter 27 Readily Releasable Stores of Calcium in Neuronal Endolysosomes: Physiological and Pathophysiological Relevance



#### Koffi L. Lakpa, Peter W. Halcrow, Xuesong Chen, and Jonathan D. Geiger

Abstract Neurons are long-lived post-mitotic cells that possess an elaborate system of endosomes and lysosomes (endolysosomes) for protein quality control. Relatively recently, endolysosomes were recognized to contain high concentrations (400-600 µM) of readily releasable calcium. The release of calcium from this acidic organelle store contributes to calcium-dependent processes of fundamental physiological importance to neurons including neurotransmitter release, membrane excitability, neurite outgrowth, synaptic remodeling, and cell viability. Pathologically, disturbances of endolysosome structure and/or function have been noted in a variety of neurodegenerative disorders including Alzheimer's disease (AD) and HIV-1 associated neurocognitive disorder (HAND). And, dysregulation of intracellular calcium has been implicated in the neuropathogenesis of these same neurological disorders. Thus, it is important to better understand mechanisms by which calcium is released from endolysosomes as well as the consequences of such release to inter-organellar signaling, physiological functions of neurons, and possible pathological consequences. In doing so, a path forward towards new therapeutic modalities might be facilitated.

Keywords Endosomes  $\cdot$  Lysosomes  $\cdot$  Endolysosomes  $\cdot$  Calcium  $\cdot$ Store-operated calcium entry  $\cdot$  N-type calcium channels  $\cdot$  Neurodegenerative diseases  $\cdot$  HIV-1 associated neurocognitive disorder  $\cdot$  Alzheimer's disease  $\cdot$  Neurons

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## 27.1 An Evolutionary Perspective on Calcium, Intracellular Organelles and Endolysosomes

Intracellular calcium regulates many essential functions of neurons including neurotransmitter release, excitability, synaptic plasticity, and cell viability [1]. Levels of intraneuronal calcium are very tightly regulated both temporally and spatially by various mechanisms including calcium release from intracellular stores, calcium influx across plasma membranes, and its association with a whole host of calcium binding proteins. Because of its importance both physiologically and pathologically, we start our story about the presence and functional significance of readily releasable stores of calcium in neuronal endolysosomes with a brief evolutionary perspective about calcium and intracellular organelles.

Calcium is well-known to be important for signal transduction in most cells including neurons. Indeed, calcium has been referred to as a universal second messenger in eukaryotic cells. The approximate 10,000-fold gradient of extracellular to intracellular calcium originated evolutionarily because of the gradual rise in calcium levels from about 100 nM during the period when the basic building blocks of life developed in thermal ducts under the ocean floor to about 1 mM during the Pre-Cambrian period when multicellular life evolved [2, 3]. Due to the toxic nature of millimolar levels of calcium, evolutionary pressure was applied such that cellular survival dictated that semipermeable membranes appeared and a variety of mechanisms were formed to maintain appropriate calcium gradients across plasma membranes [3]. Simultaneously, embedded in the plasma membranes were newly developed calcium pumps and calcium binding proteins which helped with calcium homeostasis [3]. Together, in neurons, these evolutionary changes provide unique and complex spatial and temporal handling of calcium that is essential for not only proper cellular signaling but also neuronal cell life and death.

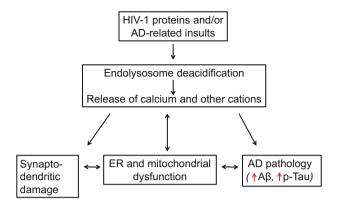
It was also during this billion-year evolutionary period that intracellular organelles began appearing including mitochondria resulting from symbiotic relationships with bacteria and the development of functional endocytic machinery [4]. Mitochondria are integral to the maintenance of cellular energetics and they are important 'sinks' for intracellular calcium [5]. However, when too much calcium is up-taken into mitochondria cellular energetics are compromised and the resulting calcium overload can lead to a cascade of events including increased oxidative stress and cell death. It has also become increasingly appreciated that organelles including endoplasmic reticulum, endosomes and lysosomes (hereafter referred to as endolysosomes) have readily releasable and functionally important pools of intracellular calcium. Although less well known, the approximate 500 µM levels of calcium in endolysosomes are similar to the calcium concentrations present in endoplasmic reticulum [6]. This is a very important concept because endoplasmic reticulum is commonly referred to as the principal intracellular store of readily releasable calcium. Furthermore, as the field of inter-organellar signaling as well as physical and chemical crosstalk between organelles has grown over the past decade it is prudent of us to now posit that this relatively new and highly complicated area of modern cell biology is key to our understanding of the regulation and dysregulation of calcium [7].

With this as a very quick trip across 1 billion years of evolutionary biology, here we embark on a brief but focused summary of findings that neuronal endolysosomes contain readily releasable stores of calcium and once released this calcium can lead to calcium influx into cells, calcium release from other organelles, and calcium dysregulation-induced neurotoxicity. The relevance of such an important upstream store of calcium to the regulation of physiological functions and pathophysiological events is obvious and will be discussed with particular relevance to the pathogenesis of two neurodegenerative disorders; Alzheimer's disease (AD) and HIV-1 associated neurocognitive disorder (HAND).

## 27.2 Endolysosomes Contain Readily Releasable Pools of Calcium

Neurons are long-lived post-mitotic cells that possess an elaborate endolysosome system for quality control especially for proteins. Endolysosomes are well known to be acidic organelles that contain high levels of cations including calcium, iron, zinc and copper. However, for the cation calcium it was not until fairly recently that these organelles were described as being 'acidic calcium stores' because the luminal pH of endolysosomes is acidic and endolysosomes contain high (400–600  $\mu$ M) levels of readily releasable calcium [8, 9].

Neuronal calcium signals display spectacular spatiotemporal complexity and understanding how calcium signals are generated spatially and temporally is necessary to understand calcium-dependent cellular processes. Endolysosome calcium levels are maintained by a variety of uptake and efflux mechanisms. Essential for uptake of calcium into endolysosomes, proton gradients are established mainly by vacuolar H<sup>+</sup>-ATPase (v-ATPase) that pumps H<sup>+</sup> into the lumen and this helps regulate Ca<sup>2+</sup> levels [9–11]. Four main mechanisms for calcium release from endolysosomes have been described including: (1) Calcium release through twopore channels (TPCs) triggered by nicotinic acid adenine dinucleotide phosphate (NAADP) [12–17]; (2) Elevation of endolysosome pH with, for example, the selective v-ATPase inhibitor bafilomycin (BAF) or the alkaline lysosomotropic agents  $NH_4Cl$  and chloroquine [8, 18, 19]; (3) Involvement of TRPML1 mucolipin-type channels and P2X4 receptors [20-22]; and (4) Selective disruption of endolysosome membranes with Gly-Phe- $\beta$ -naphtylamide (GPN) [23, 24]. Of physiological significance, calcium released from endolysosomes has been shown to contribute to a variety of calcium-dependent neuronal processes including neurotransmitter release, neuronal excitability, neurite outgrowth, synaptic remodeling, and cell viability [25-27].



**Fig. 27.1** HIV-1 proteins and other neurotoxic insults can cause deacidification of endolysosomes. Increasing endolysosome pH can release calcium and other cations from endolysosomes. Calcium released from readily releasable stores in endolysosomes can increase the release of calcium from other intracellular stores and can increase the influx of extracellular calcium. Such increases in pH and calcium levels can cause endoplasmic reticulum (ER) and mitochondrial dysfunction, Alzheimer's disease (AD)-like pathology, and synaptodendritic damage

Endolysosomes can release calcium transiently and in a highly localized and distinct fashion [17, 28, 29]. Endolysosome calcium can affect the release of calcium from organellar stores as well as through plasma membrane-based calcium influx mechanisms. The inter-organellar signaling and signaling with the plasma membrane is explained at least in part by findings that endolysosomes are highly mobile in cells, are highly dynamic metabolically, have high rates of biogenesis, and can interact physically and functionally with other intracellular organelles (Fig. 27.1).

At least three models of acidic store-induced calcium signaling mechanisms have been described [9]. (1) Acidic stores of calcium might communicate with endoplasmic reticulum calcium stores such that calcium released from endolysosomes can enhance endoplasmic reticulum calcium loading [30] and calcium-induced calcium release [13, 15]. (2) Changes in endolysosome pH may release calcium from a subgroup of acidic calcium stores and the released calcium may affect other subgroups of acidic stores through mechanisms such as vesicular fusion of late endosomes and lysosomes [9, 15, 31]. (3) Calcium released from acidic calcium stores might depolarize plasma membranes, evoke calcium-dependent currents, and stimulate calcium influx across plasma membranes [12].

#### 27.3 Acidic Store-Operated Calcium Entry in Neurons

Acidic store-operated calcium entry (aSOCE) is a unique mechanism that links readily releasable calcium in endolysosomes with influx of extracellular calcium into neurons. This is a novel means by which intraneuronal stores of calcium can contribute to spatial and temporal integration of calcium signaling. In support of this novel mechanism, we found that calcium could be released from endolysosomes following stimulation of a number of different mechanisms, that the calcium release could be independent of other organellar stores of calcium, that release of calcium from endolysosomes triggered calcium influx, and that the calcium influx was regulated by N-type calcium channels and lysosome exocytosis (Fig. 27.2).

Capacitative influx of calcium into cells was described over 30 years ago [32]. Such calcium influx mechanisms, that are now commonly referred to as store-operated calcium entry (SOCE), are principally initiated by a reduction in

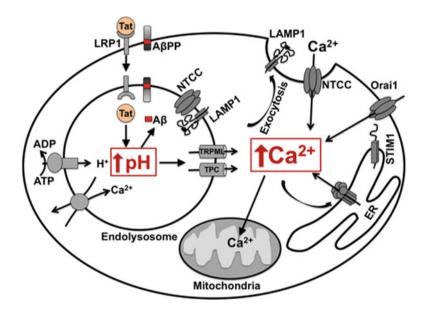


Fig. 27.2 HIV-1 Tat de-acidifies endolysosomes, increases amyloidogenesis, and releases calcium from readily releasable stores in endolysosomes. Calcium released from endolysosomes can affect mitochondrial and endoplasmic reticulum (ER) calcium stores, and increase store operated calcium (SOCE) mechanisms. Mechanistically, following de-acidification endolysosome calcium is released through TRPML1 and two pore channels (TPCs). The calcium signals can be amplified by releasing calcium from other organelles including mitochondria and ER, and by activating ER-based SOCE involving STIM1 and Orai channels as well as acidic store operated calcium entry involving N-type calcium channels (NTCCs)

endoplasmic reticulum calcium stores followed by influx of extracellular calcium in a variety of cells including neurons in order to refill the depleted stores of calcium. Mechanistically, depleting endoplasmic reticulum calcium stores drives the oligomerization and translocation of stromal interaction molecule 1 (STIM1) proteins to endoplasmic reticulum junctions close to the plasma membrane. Such STIM1 translocation induces the clustering of calcium release-activated calcium modulator 1 (Orai1) channels and/or transient receptor potential (TRP) cation channels into plasma membranes thereby enabling extracellular calcium entry [33].

Conceptually, but not mechanistically, we observed similar store-operated calcium entry involving endolysosomes in neurons. Using primary cultures of rat cortical neurons, we found that calcium was released from endolysosomes following treatment with the two-pore channel agonist NAADP-AM, the v-ATPase inhibitor BAF, and the lysosomotropic agent GPN; all of which de-acidify endolysosomes [34]. However, when these experiments were conducted in the absence of extracellular calcium, de-acidification of endolysosomes with NAADP-AM, BAF and GPN increased only slightly levels of free cytosolic calcium. When these same experiments were conducted in the presence of extracellular calcium, NAADP-AM, BAF and GPN all increased significantly the levels of free cytosolic calcium. Although it is not well understood currently, the relatively small release of calcium from endolysosomes causes a much larger influx of extracellular calcium and this might be due to plasma membrane depolarization as is accompanied by NAADP-induced endolysosome calcium release [34, 35]. Besides neurons, phenomena similar to aSOCE have been described in other cell types where NAADP has been found to induce endolysosome calcium release and large influxes of calcium across plasma membranes [12, 36–39]. These observations suggested to us that endolysosome deacidification by three completely different mechanisms led directly or indirectly to an enhanced influx of calcium into neurons. Accordingly, we next tested more specifically the extent to which a store-operated mechanism might control the observed calcium influx across the plasma membrane.

Using approaches similar to those used by others and us, we began studying store-operated calcium mechanisms including the classical endoplasmic reticulumbased capacitative SOCE. Indeed, we confirmed that in the absence of extracellular calcium and following depletion of endoplasmic reticulum calcium with the SERCA pump inhibitor thapsigargin (TG) there was a significant increase in levels of free intracellular calcium only when calcium was re-introduced to the extracellular medium. With this positive control for the functional presence of endoplasmic reticulum-based SOCE in our cultured neurons, we conducted similar experiments with agents that de-acidify endolysosomes and release calcium from endolysosome stores. Even after depleting ER pools of calcium with TG, application of NAADP-AM, BAF and GPN still caused increased influx of extracellular calcium and still induced increased levels of intracellular calcium. Thus, in these neurons there appeared to be at least two separate and functional store-operated calcium mechanisms; one governed by endoplasmic reticulum and the other by endolysosomes.

In testing the distinctive nature of the two store-operated calcium mechanisms governed by endoplasmic reticulum or endolysosomes, we used pharmacological and molecular/genetic strategies. Using siRNA to knock-down protein expression levels of STIM1, a protein that is central to SOCE, and the SOCE blockers SKF-96365 and 2-APB we were able to block significantly TG-induced release of calcium from endoplasmic reticulum, but we were unable to block significantly NAADP-AM-, BAF- and GPN-induced calcium influx. However, we were able to block significantly NAADP-AM-, BAF- and GPN-induced calcium influx with the selective N-type calcium channel (NTCC) blocker ( $\omega$ -conotoxin). The selective and specific nature of this inhibition by  $\omega$ -conotoxin was confirmed further by showing that NAADP-AM-, BAF- and GPN-induced calcium influx was not blocked by inhibitors of L-type (nimodipine, verapamil) and P/Q-type ( $\omega$ -agatoxin) calcium channels. Moreover, we confirmed these pharmacological findings by showing that siRNA knockdown of NTCCs attenuated significantly NAADP-AM-, BAFand GPN-induced calcium influx, but did not affect TG-induced SOCE. Together, the above results demonstrated that calcium released from endolysosomes can be distinct from calcium released from endoplasmic reticulum through SOCE mechanisms and that the calcium released from endolvsosomes is capable of activating cell surface calcium channels to stimulate calcium influx. These findings support and extend earlier findings that calcium released from endolysosomes did not stimulate endoplasmic reticulum-dependent SOCE in MDCK epithelial cells [23]. Accordingly, this new mechanism was termed by us as "acidic store-operated calcium entry" (aSOCE) [34].

## 27.4 Role of Lysosome Exocytosis in Acidic Store-Operated Calcium Entry (aSOCE)

Multiple mechanisms might control aSOCE involving NTCCs. One such mechanism might involve lysosome exocytosis because we have shown using a quantitative biotinylation of surface proteins assay that NAADP-AM, BAF and GPN all increased cell surface protein expression levels of NTCCs and lysosome-associated glycoprotein 1 (LAMP1). Next, we addressed the possibility that lysosome exocytosis and NTCCs were linked directly by conducting co-immunoprecipitation studies and found a physical interaction between NTCCs and LAMP1. Because LAMP1 is critical for lysosome exocytosis [40], those observations suggested to us that lysosome exocytosis might be a functional partner in aSOCE especially because aSOCE was inhibited following siRNA knockdown of protein expression levels of LAMP1. Thus, de-acidification of endolysosomes might be of central importance because NAADP-AM, BAF and GPN through very different initial mechanisms all appeared to enhance lysosome exocytosis and the recycling of NTCCs to the plasma membrane where they participated in calcium influx generally and aSOCE more specifically. Physically, this makes sense as well because of findings that de-acidification of endolysosomes changes cellular distribution patterns of these organelles from a mostly peri-nuclear pattern to one where the endolysosomes migrate close to the plasma membrane [41]. Thus, functionally and physically there is evidence favoring endolysosomes and endolysosome exocytosis in calcium entry.

# 27.5 Physical Interactions and Functional Relevance of Inter-organellar Signaling

In addition to physical interactions between endolysosomes and plasma membranes, it is becoming increasingly clear that endolysosomes physically and functionally interact as well with other intracellular organelles including mitochondria and endoplasmic reticulum. Such recognition has led to an appreciation for dynamic physical and chemical communications between intracellular organelles including those regulated by pH and calcium.

Physical interactions between mitochondria and endoplasmic reticulum were first described about 60 years ago and the functional significance of mitochondriaassociated membranes was first characterized about 30 years ago [42]. Even today, there continues to be work focused on the physical and functional interactions between organelles [43] as well as the role that organellar interactions plays in the pathogenesis of neurodegenerative diseases [44, 45]. As it relates to endolysosomes, it is now known that there are extensive physical interactions between endolysosomes and mitochondria and that these inter-organellar communications participate in lipid and metabolite exchange as well as mitochondrial quality control [46]. Conversely, mitochondrial dysfunction has been found to negatively affect lysosome structure and function through reactive oxygen species-dependent mechanisms [47]. Extensive membrane contact sites have been described between lysosomes and endoplasmic reticulum, that these contact sites were evolutionarily conserved, and that calcium released from lysosomes was sufficient to stimulate endoplasmic reticulum-dependent calcium-induced calcium release [48, 49, 50]. However, only recently was it shown that endolysosomes maintain their 1000-fold calcium concentration gradient in cells in part by refilling endolysosome stores of calcium from IP<sub>3</sub>-regulated stores of calcium in endoplasmic reticulum [51]. Some of the differences in findings as to calcium movements between organelles might be because of cell-specific mechanisms. In addition, the difficult nature of understanding inter-organellar calcium dynamics is highlighted by work showing that STIM1 and STIM2 are expressed in endolysosomes, at least in platelets, and that depletion of acidic organellar stores of calcium can increase protein-protein interactions between STIM proteins with Orai1 and TRPC channels to induce SOCE [52]. It is further complicated by findings that calcium released through endolysosomeresident TRML1 channels can cause calcium release from endoplasmic reticulum and calcium influx [53] and that NAADP has been implicated in this "cross-talk" [54].

# 27.6 Possible Role of Endolysosomes and aSOCE in Pathogenesis of Alzheimer's Disease and HIV-1 Associated Neurocognitive Disorder (HAND)

Disturbances in endolysosome structure and/or function have been noted in a variety of neurodegenerative disorders including Alzheimer's disease (AD) and HIV-1 neurocognitive disorder (HAND) [55–59]. AD is a devastating age-related neurodegenerative disease that is the commonest cause of dementia in people over the age of 65. People with HAND, on the other hand, exhibit neurological complications ranging from mild (mild cognitive impairment) to severe (dementia). In the current era of anti-retroviral therapeutics HIV-1 infected individuals are living almost full life-spans, but are now experiencing a prevalence rate of over 50% for HAND [60, 61]. Clinically and pathologically people living with neuroHIV-1 are exhibiting AD-like symptoms including learning and memory deficits as well as increased amyloidogenesis. Although the pathogenesis of HAND is not fully understood, HIV-1 proteins including the HIV-1 transactivator of transcription protein Tat have been implicated by others and us to be causative virotoxins in HAND [62-69]. Among the HIV-1 viral proteins, HIV-1 Tat is present in brains of HIV-1 infected individuals and its levels stay elevated in CSF even when HIV-1 viral levels are immeasurable [70]. Others and we have shown that HIV-1 Tat directly excites neurons [65, 71, 72], disturbs neuronal calcium homeostasis [64, 73], disrupts synaptic integrity [74, 75], and induces neurotoxicity [68, 76].

Endolysosome dysfunction has been implicated in the development of at least two pathological hallmarks of AD and HAND; A $\beta$  accumulation and neurofibrillary tangle formation. Endolysosomes are very important for amyloidogenic processing of A $\beta$ PP to A $\beta$  because amyloid  $\beta$  precursor protein (A $\beta$ PP) is first endocytosed, the amyloidogenic enzymes BACE-1 and  $\gamma$ -secretase are almost exclusively located in endosomes and lysosomes, the acidic environment of endolysosomes is favorable for amyloidogenic metabolism of A $\beta$ PP, and A $\beta$  can be either accumulated in or released by exocytosis from endolysosomes [77–83]. Tau is a microtubuleassociated protein, and when hyperphosphorylated it aggregates and contributes to the formation of neurofibrillary tangles. Tau aggregates can be degraded by cathepsin D in autophagosomes-lysosomes [84, 85], and endolysosome dysfunction contributes to tau aggregation and neurofibrillary tangle formation [86, 87]. On the other hand, transcriptional activation of lysosome biogenesis can clear aggregated tau [88]. Thus, endolysosomes are important sites for development of these neurological disorders.

Dysregulation of intracellular calcium has also been implicated in the neuropathogenesis of these same neurological disorders. And it is clear (see above) that de-acidification of endolysosomes releases calcium from these acidic stores [28, 89, 90]. We found that HIV-1 Tat protein elevated endolysosome pH and disturbed the structure and function of endolysosomes [74], a prominent and early pathological feature of HAND [57, 58]. Clearly, endolysosome calcium stores

contribute to neuronal calcium signaling and function [91–93] and calcium release from endolysosomes triggers calcium release from endoplasmic reticulum [11, 17] and through plasma membranes via aSOCE (see above).

HIV-1 proteins including HIV-1 Tat, and anti-retroviral therapeutic drugs contribute to the development of AD-like pathology including increases in A $\beta$  levels [94–99]. HIV-1 Tat enters neurons via receptor-mediated endocytosis [100–102]. The Tat-induced de-acidification of endolysosomes and resulting effects on calcium dyshomeostasis likely results from the ability of HIV-1 Tat to decrease the levels and activity of vacuolar-ATPase as well as compensatory increases in cathepsin D and LAMP-1 [103]. The consequences of such alterations in calcium dynamics and homeostasis are synaptic disruption and neurotoxicity [104–106].

Endolysosomes contain physiologically important levels of calcium that is readily releasable by a number of stimuli and insults. The calcium can exit through a number of channels including TRPML and two pore channels. Once released the calcium can signal other organelles to release calcium and for greater influx of calcium through plasma membrane-resident calcium channels especially N-type calcium channels. These effects on endolysosome structure and function have clear implications to the pathogenesis of AD and HAND; neurological disorders that show overlap in terms of clinical and pathological features. We are excited to be part of this emerging area of cell biology focused on inter-organellar signaling and look forward to further studies elucidating physiological and pathological consequences of calcium release from endolysosome stores.

Acknowledgements The authors gratefully acknowledge the funding provided by the NIH for our work; P30GM103329, R01MH100972, R01MH105329, R01MH119000, R01NS065957, and R01DA032444.

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