



Pathogenesis of Gestational Diabetes Mellitus

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Introduction

Gestational diabetes mellitus (GDM) constitutes the most common metabolic disease of pregnancy, with a continuously increasing prevalence [1, 2]. It has been associated with several maternal and fetal/neonatal complications [3, 4]. Increased maternal age, increased pre-pregnancy body mass index (BMI), excessive weight gain during pregnancy, Aboriginal Australian, Middle Eastern and Pacific Islander ethnicity, positive family history of GDM, and parity are established risk factors for the development of GDM [5, 6]. GDM, similarly to type 2 diabetes mellitus (T2DM), is a multifactorial disease; its pathogenetic mechanisms are not yet fully understood. Genetic and acquired factors that affect insulin sensitivity and insulin secretion have been implicated to GDM development and determine the disease severity [7]. Hormonal, inflammatory, and immunologic factors contribute to GDM pathogenesis. Suboptimal lifestyle, such as hypercaloric diet, unhealthy nutritional habits, and reduced physical activity, contributes to central obesity, a triggering factor for GDM [8, 9].

Insulin Action and Sensitivity

A major pathogenetic mechanism for GDM is the reduced insulin sensitivity that occurs in normal pregnancy due to placental and maternal hormonal action. Insulin action is impaired at hepatic, muscle, and adipose tissue level [10–12]. Impaired post-receptor insulin signaling is mainly responsible for pregnancy-induced insulin resistance. Experimental studies showed impaired mRNA or protein expression of insulin signaling cascade components, such as insulin receptor substrate (IRS)-1 and (IRS)-2, as well

as glucose transporter (GLUT)-1 and (GLUT)-4 in adipose tissue and muscle of women whose pregnancies were complicated by GDM. Decreased IRS-1 tyrosine phosphorylation, decreased GLUT-4 insulin-induced translocation to the cell surface, and decreased glucose transport into the cell were also found in muscle and adipose tissue of women with GDM [13–16]. Similar post-receptor insulin defects have been found in the placenta of GDM-affected pregnancies [17]. Chronic low-grade inflammation that characterizes obesity, which often accompanies GDM pregnancies, contributes to insulin signaling impairment [18], as well as oxidative stress [19].

Placental Hormones

Placental hormones, such as human placental lactogen (HPL) and placental growth hormone (GH), are opposed to insulin action [20]. HPL is produced by syncytiotrophoblast and is gradually increased during pregnancy until about 30th gestational week, when it reaches a plateau. It is correlated to fetal weight and well-being as well as placental function [21]. HPL is the main insulin resistance mediator during pregnancy. It acts as an “anti-insulin” agent in order to ensure adequate glucose supply to the embryo [22, 23]. HPL results in raised maternal blood glucose concentrations by increasing insulin resistance and raised free fatty acids concentrations by increasing lipolysis [24]. A sudden drop to HPL concentrations could indicate fetal distress [25–27]. Growth hormone (GH) is an anabolic hormone, involved in carbohydrate and lipid metabolism, and, when in excess, has diabetogenic properties, opposing insulin action [28]. Human placental GH is the main GH molecule produced during pregnancy, having an effect on maternal insulin sensitivity [29]. It is produced mainly by placental syncytiotrophoblastic cells, and it is gradually increased by midpregnancy to term. Studies in transgenic mice showed severe insulin resistance induction by placental GH [30].

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Maternal Hormones

Maternal serum GH, other growth factors, such as insulin like growth factor-1 (IGF-1), their binding proteins, prolactin (PRL), progesterone, and cortisol are altered in women whose pregnancies are complicated by as compared with unaffected pregnant women [31, 32]. PRL is produced, mainly, by anterior pituitary lactotroph cells and, secondary, by the central nervous system, immune cells, nonpregnant uterus, placenta, amnion, decidua, and the mammary gland. The most well-known action of PRL is lactation. Other PRL effects are mammary epithelial proliferation, corpus luteum function, and immune response [33, 34]. Evidence about PRL's effect on insulin sensitivity is contradictory. Hyperprolactinemia, as in patients with a prolactinoma, exacerbates insulin resistance to the nonpregnant state [35, 36]. The latter effect regresses after treatment with dopaminergic receptor agonist [37]. In contrary, studies in nonpregnant healthy women (with normal prolactin concentrations) showed that lower prolactin concentrations were correlated to decreased insulin sensitivity and increased risk for diabetes [38, 39]. During pregnancy, PRL is also produced by decidual cells and fetal pituitary. Maternal PRL is increased gradually by conception to term [40]. In pregnant rats, increased prolactin concentrations have been correlated to a post-receptor insulin defect [20]. In humans, higher concentrations of PRL during the third trimester of pregnancy have been associated with decreased glucose tolerance, implying a causative relationship between hyperprolactinemia and GDM [41]. In contrary, in another study, no difference in PRL concentrations has been found between GDM and controls [42]. Maternal, placental, and fetal adrenal steroids, progesterone, cortisol, estrogen, and androgens, also contribute to pregnancy-induced insulin resistance [43]. Progesterone, produced initially by the corpus luteum and later by the placenta, inhibits insulin action *in vivo* and *in vitro*, mainly by inhibiting the PI3-kinase pathway of the insulin signaling cascade in the adipocytes [44]. Cortisol can also induce insulin resistance through post-receptor insulin defect [20]. Androgen receptors are overexpressed in placentas of GDM-affected pregnancies as compared to controls [45]. Although it is known that estrogens regulate carbohydrate metabolism, the underlying mechanisms are not fully understood. In the nonpregnant state, estradiol (E₂) partially affects insulin signaling through modification of mitochondrial function [46]. In GDM-affected pregnancies, estrogen concentrations are lower as compared to unaffected pregnant women [47].

Maternal Adipokines

Maternal adipokines have a significant effect on insulin action. Adiponectin, an adipose tissue-derived plasma protein, has a beneficial effect on carbohydrate metabolism

by increasing insulin sensitivity [48, 49]. It is produced mainly by white adipose tissue (WAT). Adiponectin seems to express protective properties for the vascular endothelium and the heart through anti-inflammatory action and suppression of the atherosclerotic processes [50–52]. Higher concentrations of adiponectin have been associated with lower risk of T2DM development in nonpregnant women [53]. In pregnancy, evidence about adiponectin concentrations is not consistent; placental production of adiponectin has not been confirmed by all investigators [54, 55]. Some studies have demonstrated an increase in adiponectin concentrations in early pregnancy and a gradual decrease thereafter compared with the prepregnancy state [56, 57]. Although evidence regarding gestational concentrations of adiponectin and carbohydrate metabolism is less clear, a link between hypoadiponectinemia and insulin resistance exists [57, 58], as pregnant women with GDM have lower adiponectin levels than healthy controls [59].

Another adipokine, leptin, is strongly involved to metabolic issues affecting insulin secretion and action as well as tissue insulin sensitivity [60, 61]. Leptin is produced mainly by WAT adipocytes, proportionally to adipose tissue mass [62]. In a lesser degree, it is produced by brown adipose tissue (BAT), placenta, skeletal muscle cells, ovaries, and gastric cells. Leptin's primary action is the regulation of energy homeostasis [63]. Leptin reduces insulin synthesis and secretion, whereas it increases insulin sensitivity [61, 64]. Obesity is associated with resistance to leptin action [65]. During pregnancy, placenta-derived leptin results in nearly a 100% increase in maternal serum concentrations [66, 67]. Further increased leptin concentrations have been found in GDM-affected women as compared to non-affected pregnant women [68, 69]. Both adiponectin and leptin gene polymorphisms have been correlated to GDM occurrence [70]. Low adiponectin and high leptin concentrations during the first trimester may predict GDM occurrence during later pregnancy [71, 72].

Fetuin B, a recently identified adipokine, impairs insulin action. Women with GDM-affected pregnancies have higher fetuin B concentrations as compared with controls [73]. Data on resistin, visfatin and apelin concentrations, and their association with GDM are not consistent. Other novel adipokines, such as omentin and chemerin, have been associated to GDM development, and a causal effect is implied by some investigators [74].

Immunological Changes and Low-Grade Inflammation

Normal pregnancy is accompanied by immunological changes and a low-grade inflammation that is occurred to the benefit of the fetus [75, 76]. Inflammation is exacerbated by obesity, a common risk factor of GDM, and

affects insulin sensitivity through post-receptor signaling defect. Inflammatory cytokines, such as tumor necrosis factor- α (TNF- α), have a direct adverse effect on insulin action in healthy nonpregnant women, inducing insulin resistance [77]. TNF- α is a transmembrane protein produced mainly by activated macrophages in response to immunological stimulus [78]. In a lesser degree, it is expressed by other cells, such as lymphoid cells, cardiac myocytes, endothelial cells, and adipocytes. It expresses a cytotoxic effect on many cells; simultaneously, it has a regenerative effect on tissues [79, 80]. TNF- α induces phosphorylation of the IRS-1, thus preventing the interaction of insulin with the insulin receptor and impairing insulin action. Interleukin-6 (IL-6) is a pro-inflammatory cytokine and an anti-inflammatory myokine expressed by immune cells, such as T-cells and macrophages, visceral adipocytes, osteoblasts, and vascular smooth muscle cells. It is the main stimulator of the production of many acute-phase proteins. It impairs insulin-induced insulin receptor and IRS-1 phosphorylation, resulting to inhibition of the insulin signaling cascade [81]. C-reactive protein (CRP) is an acute-phase protein of hepatic origin that is increased in response to inflammation and IL-6 secretion. It acts through activation of the complement system, triggering phagocytosis by immune cells. CRP is associated to insulin resistance in healthy individuals; high concentrations of high-sensitivity (hs)-CRP are indicative of higher risk for metabolic, cardiovascular, and cerebrovascular disease [82]. The Generation R Study showed that increased CRP concentrations during early gestation are associated to high risk of neonatal complications [83]. During normal pregnancy, low-grade inflammatory markers, such as CRP, IL-6, TNF- α , and GlycA, have found to be increased, suggesting an upregulation of systemic maternal inflammation [75, 84]. In contrast to this normal maternal adaptation, a further increase of some inflammatory markers is considered a risk factor for adverse pregnancy outcomes, including GDM. Specifically, it has been shown that women with GDM-affected pregnancies have increased IL-6 concentrations as compared to controls [85]. In a recent meta-analysis, TNF- α has been found to be higher in GDM pregnancies compared to controls, independently of BMI [69]. CRP has been associated with GDM; an increase in its concentrations during early pregnancy is predictive of GDM development later in pregnancy [86, 87].

Oxidative Stress

Normal pregnancy is considered a condition of increased oxidative stress. Several pathologic conditions during pregnancy, including GDM, are associated with a further aggravation of oxidative stress. It is believed that oxidative stress is caused either by increased reactive oxygen species (ROS)

production or by a reduction of the antioxidant capacity [19]. Both an increase in oxidative stress markers and a decrease in antioxidative factors have been found in GDM-affected pregnancies. ROS induce inflammatory response and inflammatory protein expression, aggravating the normal low-grade inflammation and insulin resistance during pregnancy. Furthermore, increased protein oxidation due to enhanced oxidative stress could be implicated to GDM pathogenesis [87, 88].

β -Cell Dysfunction and Insulin Secretion

β -Cell Dysfunction

During normal pregnancy, pancreatic cell adaptation occurs to compensate for the increased need for insulin. β -cell expansion and hyperfunctioning occur early in pregnancy in order to cope with the decreased insulin sensitivity that occurs after the second half of pregnancy [89]. GDM is characterized by decreased insulin response to oral glucose and protein, sluggish first-phase insulin secretion, and delayed peak insulin secretion [90]. Subclinical pre-existing β -cell dysfunction, rather than a gradual decline of β -cell function during pregnancy, and the effect of maternal hormones and inflammatory mediators (see Sect. 2) on β -cell function constitute main mechanisms for the occurrence of GDM [91, 92]. Pre-existing β -cell dysfunction, due to genetic predisposition, does not allow for compensatory pancreatic β -cell hyperfunction to counter-regulate for the increased insulin resistance of pregnancy (Fig. 14.1) [93]. β -cell dysfunction in pregnancies complicated by GDM persists postpartum as compared to controls. Given the normalization of insulin sensitivity after delivery, only a small percentage of women with GDM remain within diabetic ranges; nevertheless, the risk for developing T2DM in later life remains increased (Fig. 14.2) [94].

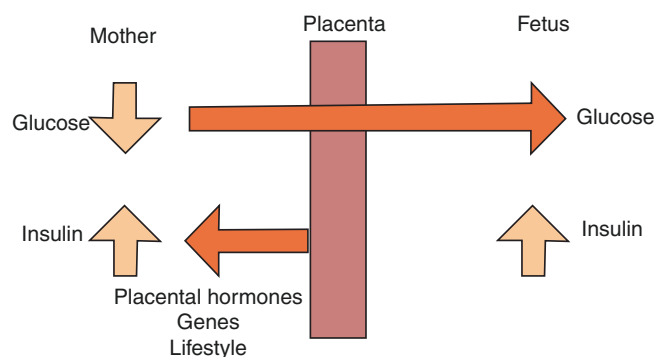
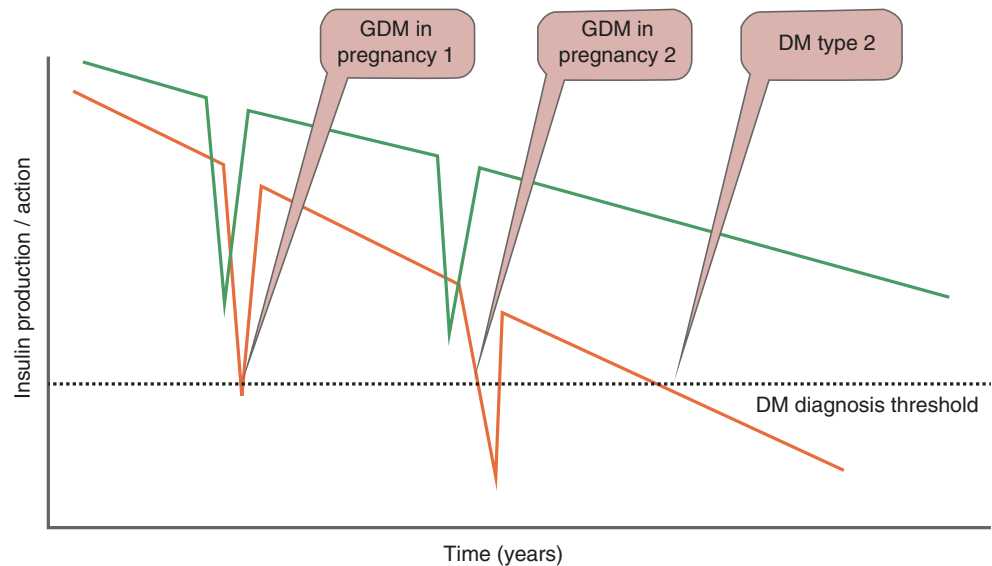


Fig. 14.1 Pathogenesis of GDM: combination of maternal and placental hormonal alteration, genetic predisposition, and suboptimal lifestyle. (Adapted from: Poulakos et al. [93])

Fig. 14.2 T2DM development in women with prior GDM. Women with a history of GDM have increased probability of developing GDM in later life due to genetic predisposition and suboptimal lifestyle. (Adapted from: Poulakos et al. [93])



Vitamin D Deficiency

Hypovitaminosis D, defined as low serum concentrations of 25-hydroxy- vitamin D₃ [25(OH)D₃], has been correlated to β -cell dysfunction in the nonpregnant state; vitamin D supplementation has been shown to improve insulin secretion in rats [95, 96]. During pregnancy, lower 25(OH)D₃ concentrations have been associated with GDM [97]. Moreover, lower 25(OH)D₃ concentrations postpartum have been associated with impaired β -cell function in women with a history of GDM [98]. Increased parathyroid hormone (PTH) concentrations have been implicated to GDM pathogenesis, partially through insulin secretion impairment [99].

Maternal Hormones

The effect of maternal hormones on β -cell function and proliferation during pregnancy is still not completely understood, and some results are contradictory. Despite that PRL is considered as a major regulator of β -cell expansion and hyperfunction during pregnancy, higher prolactin concentration has been correlated to decreased glucose tolerance during late pregnancy [41, 100]. PRL receptor-null mice have shown β -cell maladaptation during pregnancy [101]. Moreover, PRL has been found to reduce menin concentrations, a known tumor suppression factor that also suppresses β -cell proliferation and may be implicated to GDM development in pregnant mice [102]. 17 β -estradiol is seemed to be involved to β -cell adaptation and insulin secretion during pregnancy, specifically β -cell survival [103, 104]. Progesterone receptor-knockout mice

have increased insulin secretion probably due to increased β -cell mass [105]. The latter is in accordance with another experimental study that showed an apoptotic action of progesterone to pancreatic β -cells through an oxidative stress-dependent mechanism [106]. HPL stimulates insulin secretion and may have a central role to regulation of islet function during pregnancy [107]. Recent data suggest a leptin-induced decrease of insulin secretion by direct action on β -cells. Moreover, leptin affects β -cell proliferation and apoptosis and inhibits insulin gene expression [108].

Low-Grade Inflammation

As mentioned above, the low-grade inflammation that characterizes GDM affects glucose metabolism through an increase to insulin resistance. Additionally, an impairment on adipokines production, possibly due to this inflammation, has also been correlated to β -cell dysfunction and decreased insulin secretion [69, 109]. Specifically, GDM-affected women have lower adiponectin concentrations as compared with controls [69, 110]. This hypo adiponectinemia of GDM pregnancy has been associated to β -cell dysfunction [111]. As part of the low-grade inflammation, GDM-affected women have increased TNF- α concentrations [69]. Beyond insulin resistance, TNF- α has a pro-apoptotic effect on β -cells [112]. The latter could contribute to the reduced insulin secretion of GDM. As mentioned above, GDM-affected women have lower concentrations of 25(OH)D₃. Vitamin D deficiency has also been associated to increased concentrations of inflammatory markers that could further deteriorate β -cell function [113].

Oxidative Stress

Beyond insulin resistance, oxidative stress per se or as a consequence of inflammation and hyperglycemia has been linked to decreased insulin secretion during the nonpregnant state [114]. GDM is characterized by increased oxidative stress as it is determined by increased concentrations of advanced glycosylated end-products (AGEs) and other markers of oxidative lipid and protein damage [88, 115]. Recently, a furan fatty acid metabolite, 3-carboxy-4-methyl-5-propyl-2-furanpropanoic acid (CMPF), has been recognized as a possible negative regulator of β -cell function, inhibiting insulin synthesis and secretion through oxidative stress and mitochondrial dysfunction in human and mouse islets. Women with GDM have increased concentrations of CMPF as compared with controls [116]. Moreover, in GDM-affected women, CMPF predicted lower β -cell function indices [92].

Autoimmunity

A rare cause of GDM is an autoimmune destruction of pancreatic β -cells, similar to that of type 1 diabetes mellitus (T1DM). Autoimmune GDM consists in less than 10% of cases. GDM-affected women with autoimmune form of diabetes often develop T1DM soon after pregnancy or latent autoimmune diabetes of adulthood (LADA) some years after delivery [117]. In a Swedish population, antibodies implicated in T1DM pathogenesis [glutamic acid decarboxylase antibodies (GADA), islet cell antigen-2 antibodies (ICA)/tyrosine phosphatase antibodies (IA2)] have been detected in 6% of women with GDM [118]. Specifically, the prevalence of GADA in GDM-affected women has been shown to extend between 0% and 11%, of ICAs between 1% and 35%, of insulin autoantibodies (IAA) between 0% and 6%, and that of anti-IA2 between 0% and 6% [119]. Moreover, pancreatic autoantibodies may be developed in some GDM women postpartum [117]. GADA were positively associated with postpartum development of diabetes in women diagnosed with GDM [120]. As a consequence, positive GADA and other pancreatic autoantibodies in GDM-affected women can be predictive of postpartum T1DM development [121]. A recent meta-analysis has shown an association between HLA class II variants, which consists of up to 30–50% of the pathogenesis of T1DM, and GDM. Specifically, DQB1*02 and DRB1*1302 alleles have been significantly associated with increased risk of developing GDM. In contrary, DQB1*0602 seems to be a protective allele against GDM development [122]. HLA-DR6 alleles were also positive correlated to GDM development, whereas other haplotypes, such as HLA-DR2 and HLA-DR51, seem to be protective.

Besides HLA-DR3 gene and HLA-DR6/DR9 heterozygote were associated to GDM severity and prognosis [123]. Other studies found no significant differences to the distribution of HLA class II polymorphism between GDM, impaired glucose tolerance (IGT), and unaffected pregnant women [124]. It is obvious that the evidence about the relationship between GDM and autoimmunity is still controversial and more studies are needed to establish it.

Genetic Causes

A rare cause of GDM is maturity-onset diabetes of the young (MODY) gene mutations. Several MODY gene mutations are present in GDM-affected women. MODY is an inherited form of diabetes resulting by a mutation of a single, autosomic, dominant gene that disrupts insulin secretion. It may be inherited to the offspring by both maternal and paternal origin; less frequently, it can be caused by de novo gene mutation. Nowadays, several types of MODY have been recognized. Genes that are implicated to MODY development are hepatocyte nuclear factor-1 homeobox a (HNF1a) gene that is responsible for MODY 3 development, glucokinase (GCK) gene for MODY 2, hepatocyte nuclear factor-4 homeobox a (HNF4a) gene for MODY 1, hepatocyte nuclear factor-1 homeobox b (HNF1b) gene that cause diabetes and renal cysts (MODY 5), insulin promoter factor (HPF1) gene for MODY 4, insulin gene for MODY 10, ABCC8 gene [sulfonylurea receptor-1 (SUR1) subunit] for MODY 12, potassium inwardly rectifying channel subfamily J member 11 (KCNJ11) gene for MODY 13, neurogenic differentiation-1 gene (NEUROD1) for MODY 6, kruppel-like factor 11 (KLF 11) gene for MODY 7, carboxyl ester lipase (CEL) gene for MODY 8, paired box-4 (PAX4) gene for MODY 9, and BLK gene for MODY 11 [125–133]. These monogenic forms of diabetes constitute less than 10% of GDM; MODY 2 has been recognized as the most frequent type associated with GDM [134]. Several other mutations of MODY genes have been detected in GDM women such as HNF1a, IPF1, insulin gene, and KCNJ11 gene [135–139]. However, a causal relationship between MODY and GDM has not been established yet. Further investigation is needed regarding the possible clinical implications of MODY gene mutations on maternal and fetal health [134].

Conclusions

GDM is the most common metabolic complication of pregnancy. Its prevalence has been increasing over the years and parallels the increasing obesity trend. The main pathogenetic mechanism is insulin resistance as a result of maternal and

placental hormone alteration, maternal adipokine alteration, low-grade inflammation, and oxidative stress that accompany both pregnancy and obesity. An additional pathogenetic mechanism is β -cell dysfunction either pre-existing, as a result of occult genetic predisposition, or due to hormonal and inflammatory effect of pregnancy and obesity. Less frequent causes of GDM are autoimmune destruction of pancreatic β -cells (similarly to T1DM) and impaired insulin secretion caused by gene mutations, such as MODY.

Multiple-Choice Questions

- Gestational diabetes mellitus constitutes:
 - A rare disease
 - The most common metabolic disease of pregnancy
 - A disease that begins when healthy blood cells change and grow uncontrollably
 - The onset of Type 2 diabetes in pregnancy
 - A monogenic form of diabetes occurring in pregnancy
- In gestational diabetes mellitus-affected pregnancies, estrogen concentrations are:
 - Equal as compared to unaffected pregnant women
 - Higher as compared to unaffected pregnant women
 - Lower as compared to unaffected pregnant women
 - Abolished during pregnancy
 - Are highly dependent of insulin concentrations
- The Generation R Study showed that increased CRP concentrations during early gestation are associated to high risk of:
 - Asthma
 - Neonatal complications
 - Weight loss
 - Hypoglycemia
 - Maternal cardiovascular disease
- Which is the main pathogenetic mechanism of gestational diabetes mellitus?
 - Insulin resistance as a result of maternal and placental hormone alteration, maternal adipokine alteration, low-grade inflammation, and oxidative stress that accompany both pregnancy and obesity
 - Insulin as a result of maternal and placental hormone alteration, maternal adipokine, low-grade inflammation and oxidative stress that accompany the obesity
 - Autoimmune destruction of pancreatic β -cells (similarly to T1DM) and impaired insulin secretion caused by gene mutations, such as MODY
 - Insulin resistance in skeletal muscle resulting from physical inactivity during pregnancy
 - High levels of counter-regulatory hormones
 - Maternal overweight and obesity
- Placental hormones, such as human placental lactogen (HPL) and placental growth hormone (GH) are:
 - Opposed to insulin action
 - Excellent drugs to treat gestational diabetes mellitus
 - Acts as a “pro-insulin” agent
 - Opposed to glucagon action
 - Supportive to insulin action
- Inflammatory cytokines, such as tumor necrosis factor- α (TNF- α), have a direct adverse effect on insulin action in healthy nonpregnant women, inducing:
 - Insulin resistance
 - Gestational diabetes mellitus
 - C-reactive protein decrease
 - Lower risk for metabolic, cardiovascular, and cerebrovascular disease
 - Beta-cell failure
- Maternal, placental, and fetal adrenal steroids, progesterone, cortisol, estrogen, and androgens, also contribute to pregnancy-induced insulin resistance.
 - False
 - True
- An anabolic hormone, involved in carbohydrate and lipid metabolism, when in excess, has diabetogenic properties, opposing insulin action.
 - Growth hormone
 - Epinephrine
 - Estrogens
 - Progesterone
 - Leptin
- Inflammation resulting from impaired adipokine synthesis has been correlated to β -cell dysfunction and decreased insulin secretion.
 - False
 - True
- In nonpregnant healthy women (with normal prolactin concentrations), lower prolactin concentrations are associated with:
 - Decreased insulin sensitivity and lower risk for diabetes.
 - Decreased insulin sensitivity and high risk for diabetes.
 - Increased insulin sensitivity and low risk for diabetes.
 - Increased insulin sensitivity and high risk for diabetes.
 - No associations have been documented.

Correct Answers

- (b) The most common metabolic disease of pregnancy
- (c) Lower as compared to unaffected pregnant women
- (b) Neonatal complications

4. (a) Insulin resistance as a result of maternal and placental hormone alteration, maternal adipokine alteration, low-grade inflammation, and oxidative stress that accompany both pregnancy and obesity
5. (a) Opposed to insulin action
6. (a) Insulin resistance
7. (b) True
8. (a) Growth hormone
9. (b) True
10. (b) Decreased insulin sensitivity and high risk for diabetes

References

1. Ferrara A. Increasing prevalence of gestational diabetes mellitus: a public health perspective. *Diabetes Care*. 2007;30:141–6.
2. Negrato CA, Gomes MB. Historical facts of screening and diagnosing diabetes in pregnancy. *Diabetol Metab Syndr*. 2013;5:22.
3. Mitánchez D. Foetal and neonatal complications in gestational diabetes: perinatal mortality, congenital malformations, macrosomia, shoulder dystocia, birth injuries, neonatal complications. *Diabetes Metab*. 2010;36:617–27.
4. Lowe LP, Metzger BE, Dyer AR, Lowe J, McCance DR, Lappin TR, Trimble ER, Coustan DR, Hadden DR, Hod M, Oats JJ, Persson B. Hyperglycemia and Adverse Pregnancy Outcome (HAPO) Study: associations of maternal A1C and glucose with pregnancy outcomes. *Diabetes Care*. 2012;35:574–80.
5. Di Cianni G, Volpe L, Lencioni C, Miccoli R, Cuccuru I, Ghio A, Chatzianagnostou K, Bottone P, Teti G, Del Prato S, Benzi L. Prevalence and risk factors for gestational diabetes assessed by universal screening. *Diabetes Res Clin Pract*. 2003;62:131–7.
6. Yuen L, Wong VW. Gestational diabetes mellitus: challenges for different ethnic groups. *World J Diabetes*. 2015;6:1024–32.
7. Baz B, Riveline JP, Gautier JF. Endocrinology of pregnancy: gestational diabetes mellitus: definition, aetiological and clinical aspects. *Eur J Endocrinol*. 2016;174:43–51.
8. Oken E, Ning Y, Rifas-Shiman SL, Radesky JS, Rich-Edwards JW, Gillman MW. Associations of physical activity and inactivity before and during pregnancy with glucose tolerance. *Obstet Gynecol*. 2006;108:1200–7.
9. Zhang C, Ning Y. Effect of dietary and lifestyle factors on the risk of gestational diabetes: review of epidemiologic evidence. *Am J Clin Nutr*. 2011;94:1975S–9S.
10. Catalano PM, Huston L, Amini SB, Kalhan SC. Longitudinal changes in glucose metabolism during pregnancy in obese women with normal glucose tolerance and gestational diabetes mellitus. *Am J Obstet Gynecol*. 1999;180:903–16.
11. Qu HM, Ye YH, Peng W, Zhan Y. Relationship between tyrosine phosphorylation and protein expression of insulin receptor substrate-1 and insulin resistance in gestational diabetes mellitus. *Zhonghua Fu Chan Ke Za Zhi*. 2007;42:249–52.
12. Tumurbaatar B, Poole AT, Olson G, Makhlof M, Sallam HS, Thukuntla S, Kankanala S, Ekhaese O, Gomez G, Chandalia M, Abate N. Adipose tissue insulin resistance in gestational diabetes. *Metab Syndr Relat Disord*. 2017;15:86–92.
13. Friedman JE, Ishizuka T, Shao J, Huston L, Highman T, Catalano P. Impaired glucose transport and insulin receptor tyrosine phosphorylation in skeletal muscle from obese women with gestational diabetes. *Diabetes*. 1999;48:1807–14.
14. Colomiere M, Permezel M, Lappas M. Diabetes and obesity during pregnancy alter insulin signalling and glucose transporter expression in maternal skeletal muscle and subcutaneous adipose tissue. *J Mol Endocrinol*. 2010;44:213–23.
15. Garvey WT, Maianu L, Hancock JA, Golichowski AM, Baron A. Gene expression of GLUT4 in skeletal muscle from insulin-resistant patients with obesity, IGT, GDM, and NIDDM. *Diabetes*. 1992;41:465–75.
16. Garvey WT, Maianu L, Zhu J-H, Hancock JA, Golichowski AM. Multiple defects in the adipocyte glucose transport system cause cellular insulin resistance in gestational diabetes: heterogeneity in the number and a novel abnormality in subcellular localization of GLUT4 glucose transporters. *Diabetes*. 1993;42:1773–85.
17. Colomiere M, Permezel M, Riley C, Desoye G, Lappas M. Defective insulin signaling in placenta from pregnancies complicated by gestational diabetes mellitus. *Eur J Endocrinol*. 2009;160:567–78.
18. de Luca C, Olefsky JM. Inflammation and insulin resistance. *FEBS Lett*. 2008;582:97–105.
19. Lappas M, Hiden U, Desoye G, Froehlich J, Hauguel-de Mouzon S, Jawerbaum A. The role of oxidative stress in the pathophysiology of gestational diabetes mellitus. *Antioxid Redox Signal*. 2011;15:3061–100.
20. Ryan EA, Enns L. Role of gestational hormones in the induction of insulin resistance. *J Clin Endocrinol Metab*. 1988;67:341–7.
21. Handwerger S, Freemark M. Role of placental lactogen and prolactin in human pregnancy. In: Mahesh VB, Dhindsa DS, Anderson E, Kalra SP, editors. *Regulation of ovarian and testicular function*. Boston, MA: Springer US; 1987. p. 399–420.
22. Beck P, Daughaday WH. Human placental lactogen: studies of its acute metabolic effects and disposition in normal man. *J Clin Invest*. 1967;46:103–10.
23. Walker WH, Fitzpatrick SL, Barrera-Saldana HA, Resendez-Perez D, Saunders GF. The human placental lactogen genes: structure, function, evolution and transcriptional regulation. *Endocr Rev*. 1991;12:316–28.
24. Mochizuki M, Morikawa H, Ohga Y, Tojo S. Lipolytic action of human chorionic somatomammotropin. *Endocrinol Jpn*. 1975;22:123–9.
25. Higgins LE, Rey de Castro N, Addo N, Wareing M, Greenwood SL, Jones RL, Sibley CP, Johnstone ED, Heazell AE. Placental features of late-onset adverse pregnancy outcome. *PLoS One*. 2015;10:e0129117.
26. Bersinger NA, Odegard RA. Second- and third-trimester serum levels of placental proteins in preeclampsia and small-for-gestational age pregnancies. *Acta Obstet Gynecol Scand*. 2004;83:37–45.
27. Olszewski J, Szczurowicz A, Wojcikowski C. Changes in levels of human placenta lactogen (hPL), progesterone, and estriol in blood serum and estrogens in urine during gestational diabetes mellitus. *Ginekol Pol*. 1995;66:145–50.
28. Moller N, Jorgensen JO, Abildgard N, Orskov L, Schmitz O, Christiansen JS. Effects of growth hormone on glucose metabolism. *Horm Res*. 1991;36:32–5.
29. McIntyre HD, Zeck W, Russell A. Placental growth hormone, fetal growth and the IGF axis in normal and diabetic pregnancy. *Curr Diabetes Rev*. 2009;5:185–9.
30. Barbour LA, Shao J, Qiao L, Pulawa LK, Jensen DR, Bartke A, Garrity M, Draznin B, Friedman JE. Human placental growth hormone causes severe insulin resistance in transgenic mice. *Am J Obstet Gynecol*. 2002;186:512–7.
31. Grissa O, Yessoufou A, Mrisak I, Hichami A, Amoussou-Guenou D, Grissa A, Djrolo F, Moutairou K, Miled A, Khairi H, Zaouali M, Bougmiza I, Zbidi A, Tabka Z, Khan NA. Growth factor concentrations and their placental mRNA expression are modulated in gestational diabetes mellitus: possible interactions with macrosomia. *BMC Pregnancy Childbirth*. 2010;10:7.
32. Luthman M, Stock S, Werner S, Bremme K. Growth hormone-binding protein in plasma is inversely correlated to placental lac-

- togen and augmented with increasing body mass index in healthy pregnant women and women with gestational diabetes mellitus. *Gynecol Obstet Investig.* 1994;38:145–50.
33. Horseman ND, Gregerson KA. Prolactin actions. *J Mol Endocrinol.* 2014;52:95–106.
 34. Freeman ME, Kanyicska B, Lerant A, Nagy G. Prolactin: structure, function, and regulation of secretion. *Physiol Rev.* 2000;80:1523–631.
 35. Serri O, Beauregard H, Rasio E, Hardy J. Decreased sensitivity to insulin in women with microprolactinomas. *Fertil Steril.* 1986;45:572–4.
 36. Yavuz D, Deyneli O, Akpinar I, Yildiz E, Gozu H, Sezgin O, Haklar G, Akalin S. Endothelial function, insulin sensitivity and inflammatory markers in hyperprolactinemic pre-menopausal women. *Eur J Endocrinol.* 2003;149:187–93.
 37. Inancli SS, Usluogullari A, Ustu Y, Caner S, Tam AA, Ersoy R, Cakir B. Effect of cabergoline on insulin sensitivity, inflammation, and carotid intima media thickness in patients with prolactinoma. *Endocrine.* 2013;44:193–9.
 38. Wagner R, Heni M, Linder K, Ketterer C, Peter A, Bohm A, Hatziaelaki E, Stefan N, Staiger H, Haring HU, Fritsche A. Age-dependent association of serum prolactin with glycaemia and insulin sensitivity in humans. *Acta Diabetol.* 2014;51:71–8.
 39. Wang T, Lu J, Xu Y, Li M, Sun J, Zhang J, Xu B, Xu M, Chen Y, Bi Y, Wang W, Ning G. Circulating prolactin associates with diabetes and impaired glucose regulation: a population-based study. *Diabetes Care.* 2013;36:1974–80.
 40. Jabbour HN, Critchley HO. Potential roles of decidual prolactin in early pregnancy. *Reproduction.* 2001;121:197–205.
 41. Ekinci EI, Torkamani N, Ramchand SK, Churilov L, Sikaris KA, Lu ZX, Houlihan CA. Higher maternal serum prolactin levels are associated with reduced glucose tolerance during pregnancy. *J Diabetes Investig.* 2017;8:697–700.
 42. Skouby SO, Kuhl C, Hornnes PJ, Andersen AN. Prolactin and glucose tolerance in normal and gestational diabetic pregnancy. *Obstet Gynecol.* 1986;67:17–20.
 43. Vejrazkova D, Vcelak J, Vankova M, Lukasova P, Bradnova O, Halkova T, Kancheva R, Bendlova B. Steroids and insulin resistance in pregnancy. *J Steroid Biochem Mol Biol.* 2014;139:122–9.
 44. Wada T, Hori S, Sugiyama M, Fujisawa E, Nakano T, Tsuneki H, Nagira K, Saito S, Sasaoka T. Progesterone inhibits glucose uptake by affecting diverse steps of insulin signaling in 3T3-L1 adipocytes. *Am J Physiol Endocrinol Metab.* 2010;298:E881–8.
 45. Uzelac PS, Li X, Lin J, Neese LD, Lin L, Nakajima ST, Bohler H, Lei Z. Dysregulation of leptin and testosterone production and their receptor expression in the human placenta with gestational diabetes mellitus. *Placenta.* 2010;31:581–8.
 46. Gupte AA, Pownall HJ, Hamilton DJ. Estrogen: an emerging regulator of insulin action and mitochondrial function. *J Diabetes Res.* 2015;2015:916585.
 47. Villarroel C, Salinas A, Lopez P, Kohen P, Rencoret G, Devoto L, Codner E. Pregestational type 2 diabetes and gestational diabetes exhibit different sexual steroid profiles during pregnancy. *Gynecol Endocrinol.* 2016;33:212–7.
 48. Karbowska J, Kochan Z. Role of adiponectin in the regulation of carbohydrate and lipid metabolism. *J Physiol Pharmacol.* 2006;57(Suppl 6):103–13.
 49. Stefan N, Vojarova B, Funahashi T, Matsuzawa Y, Weyer C, Lindsay RS, Youngren JF, Havel PJ, Pratley RE, Bogardus C, Tataranni PA. Plasma adiponectin concentration is associated with skeletal muscle insulin receptor tyrosine phosphorylation, and low plasma concentration precedes a decrease in whole-body insulin sensitivity in humans. *Diabetes.* 2002;51:1884–8.
 50. Matsuzawa Y, Funahashi T, Kihara S, Shimomura I. Adiponectin and metabolic syndrome. *Arterioscler Thromb Vasc Biol.* 2004;24:29–33.
 51. Nanayakkara G, Kariharan T, Wang L, Zhong J, Amin R. The cardio-protective signaling and mechanisms of adiponectin. *AJCD.* 2012;2:253–66.
 52. Shehzad A, Iqbal W, Shehzad O, Lee YS. Adiponectin: regulation of its production and its role in human diseases. *Hormones (Athens).* 2012;11:8–20.
 53. Yamamoto S, Matsushita Y, Nakagawa T, Hayashi T, Noda M, Mizoue T. Circulating adiponectin levels and risk of type 2 diabetes in the Japanese. *Nutr Diabetes.* 2014;4:e130.
 54. Caminos JE, Nogueiras R, Gallego R, Bravo S, Tovar S, Garcia-Caballero T, Casanueva FF, Dieguez C. Expression and regulation of adiponectin and receptor in human and rat placenta. *J Clin Endocrinol Metab.* 2005;90:4276–86.
 55. Corbetta S, Bulfamante G, Cortelazzi D, Barresi V, Cetin I, Mantovani G, Bondioni S, Beck-Peccoz P, Spada A. Adiponectin expression in human fetal tissues during mid- and late gestation. *J Clin Endocrinol Metab.* 2005;90:2397–402.
 56. Mazaki-Tovi S, Kanety H, Pariente C, Hemi R, Wisner A, Schiff E, Sivan E. Maternal serum adiponectin levels during human pregnancy. *J Perinatol.* 2007;27:77–81.
 57. Catalano PM, Hoegh M, Minium J, Huston-Presley L, Bernard S, Kalhan S, Hauguel-De Mouzon S. Adiponectin in human pregnancy: implications for regulation of glucose and lipid metabolism. *Diabetologia.* 2006;49:1677–85.
 58. Ritterath C, Rad NT, Siegmund T, Heinze T, Siebert G, Buhling KJ. Adiponectin during pregnancy: correlation with fat metabolism, but not with carbohydrate metabolism. *Arch Gynecol Obstet.* 2009;281:91.
 59. Retnakaran R, Hanley AJ, Raif N, Connelly PW, Sermer M, Zinman B. Reduced adiponectin concentration in women with gestational diabetes: a potential factor in progression to type 2 diabetes. *Diabetes Care.* 2004;27:799–800.
 60. Tucholski K, Otto-Buczowska E. The role of leptin in the regulation of carbohydrate metabolism. *Endokrynol Pol.* 2011;62:258–62.
 61. Paz-Filho G, Mastrorandi C, Wong M-L, Licinio J. Leptin therapy, insulin sensitivity, and glucose homeostasis. *Indian J Endocrinol Metab.* 2012;16:549–55.
 62. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology.* 2004;145:2273–82.
 63. Margetic S, Gazzola C, Pegg GG, Hill RA. Leptin: a review of its peripheral actions and interactions. *Int J Obes Relat Metab Disord.* 2002;26:1407–33.
 64. Seufert J. Leptin effects on pancreatic β -cell gene expression and function. *Diabetes.* 2004;53:S152–8.
 65. Myers MG, Leibel RL, Seeley RJ, Schwartz MW. Obesity and leptin resistance: distinguishing cause from effect. *Trends Endocrinol Metab.* 2010;21:643–51.
 66. Lepercq J, Hauguel De Mouzo S. Leptin during pregnancy. *J Gynecol Obstet Biol Reprod (Paris).* 2002;31:167–72.
 67. Highman TJ, Friedman JE, Huston LP, Wong WW, Catalano PM. Longitudinal changes in maternal serum leptin concentrations, body composition, and resting metabolic rate in pregnancy. *Am J Obstet Gynecol.* 1998;178:1010–5.
 68. Fatima SS, Alam F, Chaudhry B, Khan TA. Elevated levels of chemerin, leptin, and interleukin-18 in gestational diabetes mellitus. *J Matern Fetal Neonatal Med.* 2016:1–6.
 69. Xu J, Zhao YH, Chen YP, Yuan XL, Wang J, Zhu H, Lu CM. Maternal circulating concentrations of tumor necrosis factor-

- alpha, leptin, and adiponectin in gestational diabetes mellitus: a systematic review and meta-analysis. *ScientificWorldJournal*. 2014;2014:926–32.
70. Pawlik A, Teler J, Maciejewska A, Sawczuk M, Safranow K, Dziedzicko V. Adiponectin and leptin gene polymorphisms in women with gestational diabetes mellitus. *J Assist Reprod Genet*. 2017;34:511.
71. Lain KY, Daftary AR, Ness RB, Roberts JM. First trimester adipocytokine concentrations and risk of developing gestational diabetes later in pregnancy. *Clin Endocrinol (Oxf)*. 2008;69:407–11.
72. Qiu C, Williams MA, Vadachkoria S, Frederick IO, Luthy DA. Increased maternal plasma leptin in early pregnancy and risk of gestational diabetes mellitus. *Obstet Gynecol*. 2004;103:519–25.
73. Kralisch S, Hoffmann A, Lossner U, Kratzsch J, Bluher M, Stumvoll M, Fasshauer M, Ebert T. Regulation of the novel adipokines/hepatokines fetuin A and fetuin B in gestational diabetes mellitus. *Metabolism*. 2017;68:88–94.
74. Pan BL, Ma RM. Correlation of serum omentin-1 and chemerin with gestational diabetes mellitus. *Nan Fang Yi Ke Da Xue Xue Bao*. 2016;36:1231–6.
75. Wang Q, Würtz P, Auro K, Mäkinen V-P, Kangas AJ, Soininen P, Tiainen M, Tynkynen T, Jokelainen J, Santalahti K, Salmi M, Blankenberg S, Zeller T, Viikari J, Kähönen M, Lehtimäki T, Salomaa V, Perola M, Jalkanen S, Järvelin M-R, Raitakari OT, Kettunen J, Lawlor DA, Ala-Korpela M. Metabolic profiling of pregnancy: cross-sectional and longitudinal evidence. *BMC Med*. 2016;14:205.
76. Sargent IL, Borzychowski AM, Redman CWG. NK cells and human pregnancy - an inflammatory view. *Trends Immunol*. 2006;27:399–404.
77. Krogh-Madsen R, Plomgaard P, Møller K, Mittendorfer B, Pedersen BK. Influence of TNF-alpha and IL-6 infusions on insulin sensitivity and expression of IL-18 in humans. *Am J Physiol Endocrinol Metab*. 2006;291:E108–14.
78. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell*. 2001;104:487–501.
79. Sedger LM, McDermott MF. TNF and TNF-receptors: from mediators of cell death and inflammation to therapeutic giants – past, present and future. *Cytokine Growth Factor Rev*. 2014;25:453–72.
80. Wajant H, Pfizenmaier K, Scheurich P. Tumor necrosis factor signaling. *Cell Death Differ*. 2003;10:45–65.
81. Senn JJ, Klover PJ, Nowak IA, Zimmers TA, Koniaris LG, Furlanetto RW, Mooney RA. Suppressor of cytokine signaling-3 (SOCS-3), a potential mediator of interleukin-6-dependent insulin resistance in hepatocytes. *J Biol Chem*. 2003;278:13740–6.
82. Ndumele CE, Pradhan AD, Ridker PM. Interrelationships between inflammation, C-reactive protein, and insulin resistance. *J Cardiometab Syndr*. 2006;1:190–6.
83. Ernst GDS, de Jonge LL, Hofman A, Lindemans J, Russcher H, Steegers EAP, Jaddoe VWV. C-reactive protein levels in early pregnancy, fetal growth patterns, and the risk for neonatal complications: the Generation R Study. *Am J Obstet Gynecol*. 2011;205:e1–e12.
84. Brewster JA, Orsi NM, Gopichandran N, McShane P, Ekbote UV, Walker JJ. Gestational effects on host inflammatory response in normal and pre-eclamptic pregnancies. *Eur J Obstet Gynecol Reprod Biol*. 2008;140:21–6.
85. Kuzmicki M, Telejko B, Szamatowicz J, Zonenberg A, Nikolajuk A, Kretowski A, Gorska M. High resistin and interleukin-6 levels are associated with gestational diabetes mellitus. *Gynecol Endocrinol*. 2009;25:258–63.
86. Fatema N, Deeba F, Akter S, Sultana N, Nasrin B, Ali L, Begum SA. CRP (C-reactive protein) in early pregnancy predictor for development of GDM. *Mymensingh Med J*. 2016;25:271–6.
87. Zhu C, Yang H, Geng Q, Ma Q, Long Y, Zhou C, Chen M. Association of oxidative stress biomarkers with gestational diabetes mellitus in pregnant women: a case-control study. *PLoS One*. 2015;10:e0126490.
88. Li H, Yin Q, Li N, Ouyang Z, Zhong M. Plasma markers of oxidative stress in patients with gestational diabetes mellitus in the second and third trimester. *Int J Gynaecol Obstet*. 2016;2016:3865454.
89. Baeyens L, Hindi S, Sorenson RL, German MS. beta-Cell adaptation in pregnancy. *Diabetes Obes Metab*. 2016;18(Suppl 1):63–70.
90. Kühl C. Insulin secretion and insulin resistance in pregnancy and GDM: implications for diagnosis and management. *Diabetes*. 1991;40:18–24.
91. Buchanan TA, Xiang A, Kjos SL, Watanabe R. What Is gestational diabetes? *Diabetes Care*. 2007;30:S105–11.
92. Retnakaran R, Ye C, Kramer CK, Connelly PW, Hanley AJ, Sermer M, Zinman B. Evaluation of circulating determinants of beta-cell function in women with and without gestational diabetes. *J Clin Endocrinol Metab*. 2016;101:2683–91.
93. Poulakos P, Mintziori G, Tsiro E, Taousani E, Savvaki D, Harizopoulou V, Goulis DG. Comments on gestational diabetes mellitus: from pathophysiology to clinical practice. *Hormones (Athens)*. 2015;14:335–44.
94. Xiang AH, Takayanagi M, Black MH, Trigo E, Lawrence JM, Watanabe RM, Buchanan TA. Longitudinal changes in insulin sensitivity and beta cell function between women with and without a history of gestational diabetes mellitus. *Diabetologia*. 2013;56:2753–60.
95. Kayaniyl S, Vieth R, Retnakaran R, Knight JA, Qi Y, Gerstein HC, Perkins BA, Harris SB, Zinman B, Hanley AJ. Association of vitamin D with insulin resistance and β -cell dysfunction in subjects at risk for type 2 diabetes. *Diabetes Care*. 2010;33:1379–81.
96. Cade C, Norman AW. Rapid normalization/stimulation by 1,25-dihydroxyvitamin D₃ of insulin secretion and glucose tolerance in the vitamin D-deficient rat. *Endocrinology*. 1987;120:1490–7.
97. Zhang MX, Pan GT, Guo JF, Li BY, Qin LQ, Zhang ZL. Vitamin D deficiency increases the risk of gestational diabetes mellitus: a meta-analysis of observational studies. *Nutrients*. 2015;7:8366–75.
98. Shaat N, Ignell C. Glucose homeostasis, beta cell function, and insulin resistance in relation to vitamin D status after gestational diabetes mellitus. *Acta Obstet Gynecol Scand*. 2017;96:821.
99. Kramer CK, Swaminathan B, Hanley AJ, Connelly PW, Sermer M, Zinman B, Retnakaran R. Vitamin D and parathyroid hormone status in pregnancy: effect on insulin sensitivity, beta-cell function, and gestational diabetes mellitus. *J Clin Endocrinol Metab*. 2014;99:4506–13.
100. Huang C, Snider F, Cross JC. Prolactin receptor is required for normal glucose homeostasis and modulation of beta-cell mass during pregnancy. *Endocrinology*. 2009;150:1618–26.
101. Sorenson RL, Brelje TC. Prolactin receptors are critical to the adaptation of islets to pregnancy. *Endocrinology*. 2009;150:1566–9.
102. Karnik SK, Chen H, McLean GW, Heit JJ, Gu X, Zhang AY, Fontaine M, Yen MH, Kim SK. Menin controls growth of pancreatic beta-cells in pregnant mice and promotes gestational diabetes mellitus. *Science*. 2007;318:806–9.
103. Nadal A, Alonso-Magdalena P, Soriano S, Roperio AB, Quesada I. The role of oestrogens in the adaptation of islets to insulin resistance. *J Physiol*. 2009;587:5031–7.
104. Ackermann S, Hiller S, Osswald H, Losle M, Grenz A, Hambrock A. 17beta-Estradiol modulates apoptosis in pancreatic beta-cells

- by specific involvement of the sulfonylurea receptor (SUR) isoform SUR1. *J Biol Chem.* 2009;284:4905–13.
105. Branisteanu DD, Mathieu C. Progesterone in gestational diabetes mellitus: guilty or not guilty? *Trends Endocrinol Metab.* 2003;14:54–6.
 106. Nunes VA, Portioli-Sanches EP, Rosim MP, Araujo MS, Praxedes-Garcia P, Valle MM, Roma LP, Hahn C, Gurgul-Convey E, Lenzen S, Azevedo-Martins AK. Progesterone induces apoptosis of insulin-secreting cells: insights into the molecular mechanism. *J Endocrinol.* 2014;221:273–84.
 107. Brelje TC, Scharp DW, Lacy PE, Ogren L, Talamantes F, Robertson M, Friesen HG, Sorenson RL. Effect of homologous placental lactogens, prolactins, and growth hormones on islet B-cell division and insulin secretion in rat, mouse, and human islets: implication for placental lactogen regulation of islet function during pregnancy. *Endocrinology.* 1993;132:879–87.
 108. Marroqui L, Gonzalez A, Neco P, Caballero-Garrido E, Vieira E, Ripoll C, Nadal A, Quesada I. Role of leptin in the pancreatic beta-cell: effects and signaling pathways. *J Mol Endocrinol.* 2012;49:R9–17.
 109. Vrachnis N, Belitsos P, Sifakis S, Dafopoulos K, Siristatidis C, Pappa KI, Iliodromiti Z. Role of adipokines and other inflammatory mediators in gestational diabetes mellitus and previous gestational diabetes mellitus. *Int J Endocrinol.* 2012;2012:549748.
 110. Pala HG, Ozalp Y, Yener AS, Gerceklioglu G, Uysal S, Onvural A. Adiponectin levels in gestational diabetes mellitus and in pregnant women without glucose intolerance. *Adv Clin Exp Med.* 2015;24:85–92.
 111. Retnakaran R, Hanley AJ, Raif N, Hirning CR, Connelly PW, Sermer M, Kahn SE, Zinman B. Adiponectin and beta cell dysfunction in gestational diabetes: pathophysiological implications. *Diabetologia.* 2005;48:993–1001.
 112. Parkash J, Chaudhry MA, Rhoten WB. Tumor necrosis factor- α -induced changes in insulin-producing beta-cells. *Anat Rec A Discov Mol Cell Evol Biol.* 2005;286:982–93.
 113. Haidari F, Jalali M-T, Shabbazian N, Haghighizadeh M-H, Azadegan E. Comparison of serum levels of vitamin D and inflammatory markers between women with gestational diabetes mellitus and healthy pregnant control. *J Family Reprod Health.* 2016;10:1–8.
 114. Newsholme P, Cruzat VF, Keane KN, Carlessi R, de Bittencourt PI Jr. Molecular mechanisms of ROS production and oxidative stress in diabetes. *Biochem J.* 2016;473:4527–50.
 115. Pertynska-Marczewska M, Glowacka E, Sobczak M, Cypryk K, Wilczynski J. Glycation endproducts, soluble receptor for advanced glycation endproducts and cytokines in diabetic and non-diabetic pregnancies. *Am J Reprod Immunol.* 2009;61:175–82.
 116. Prentice KJ, Luu L, Allister EM, Liu Y, Jun LS, Sloop KW, Hardy AB, Wei L, Jia W, Fantus IG, Sweet DH, Sweeney G, Retnakaran R, Dai FF, Wheeler MB. The furan fatty acid metabolite CMPF is elevated in diabetes and induces β cell dysfunction. *Cell Metab.* 2014;19:653–66.
 117. Lapolla A, Dalfrà MG, Fedele D. Diabetes related autoimmunity in gestational diabetes mellitus: is it important? *Nutr Metab Cardiovasc Dis.* 2009;19:674–82.
 118. Torn C, Gupta M, Sanjeevi CB, Aberg A, Frid A, Landin-Olsson M. Different HLA-DR-DQ and MHC class I chain-related gene A (MICA) genotypes in autoimmune and nonautoimmune gestational diabetes in a Swedish population. *Hum Immunol.* 2004;65:1443–50.
 119. de Leiva A, Mauricio D, Corcoy R. Diabetes-related autoantibodies and gestational diabetes. *Diabetes Care.* 2007;30:S127–33.
 120. Papadopoulou A, Lynch KF, Anderberg E, Landin-Olsson M, Hansson I, Agardh CD, Lernmark A, Berntorp K. HLA-DQB1 genotypes and islet cell autoantibodies against GAD65 and IA-2 in relation to development of diabetes post partum in women with gestational diabetes mellitus. *Diabetes Res Clin Pract.* 2012;95:260–4.
 121. Nilsson C, Ursing D, Torn C, Aberg A, Landin-Olsson M. Presence of GAD antibodies during gestational diabetes mellitus predicts type 1 diabetes. *Diabetes Care.* 2007;30:1968–71.
 122. Guo CC, Jin YM, Lee KK, Yang G, Jing CX, Yang X. The relationships between HLA class II alleles and antigens with gestational diabetes mellitus: a meta-analysis. *Sci Rep.* 2016;6:35005.
 123. Song D, Liu Y, Han Y, Shang G, Hua S, Zhang H, Guo S, Jiao S. Study on the gestational diabetes mellitus and histocompatibility human leukocyte antigen DRB allele polymorphism. *Zhonghua Fu Chan Ke Za Zhi.* 2002;37:284–6.
 124. Vambergue A, Fajardy I, Bianchi F, Cazaubiel M, Verier-Mine O, Goeusse P, Fontaine P, Danze PM. Gestational diabetes mellitus and HLA class II (-DQ, -DR) association: the digest study. *Eur J Immunogenet.* 1997;24:385–94.
 125. Yamagata K, Oda N, Kaisaki PJ, Menzel S, Furuta H, Vaxillaire M, Southam L, Cox RD, Lathrop GM, Boriraj VV, Chen X, Cox NJ, Oda Y, Yano H, Le Beau MM, Yamada S, Nishigori H, Takeda J, Fajans SS, Hattersley AT, Iwasaki N, Hansen T, Pedersen O, Polonsky KS, Bell GI, et al. Mutations in the hepatocyte nuclear factor-1 α gene in maturity-onset diabetes of the young (MODY3). *Nature.* 1996;384:455–8.
 126. Froguel P, Zouali H, Vionnet N, Velho G, Vaxillaire M, Sun F, Lesage S, Stoffel M, Takeda J, Passa P, et al. Familial hyperglycemia due to mutations in glucokinase. Definition of a subtype of diabetes mellitus. *N Engl J Med.* 1993;328:697–702.
 127. Yamagata K, Furuta H, Oda N, Kaisaki PJ, Menzel S, Cox NJ, Fajans SS, Signorini S, Stoffel M, Bell GI. Mutations in the hepatocyte nuclear factor-4 α gene in maturity-onset diabetes of the young (MODY1). *Nature.* 1996;384:458–60.
 128. Horikawa Y, Iwasaki N, Hara M, Furuta H, Hinokio Y, Cockburn BN, Lindner T, Yamagata K, Ogata M, Tomonaga O, Kuroki H, Kasahara T, Iwamoto Y, Bell GI. Mutation in hepatocyte nuclear factor-1 beta gene (TCF2) associated with MODY. *Nat Genet.* 1997;17:384–5.
 129. Stoffers DA, Ferrer J, Clarke WL, Habener JF. Early-onset type-II diabetes mellitus (MODY4) linked to IPF1. *Nat Genet.* 1997;17:138–9.
 130. Meur G, Simon A, Harun N, Virally M, Dechaume A, Bonnefond A, Fetita S, Tarasov AI, Guillausseau P-J, Boesgaard TW, Pedersen O, Hansen T, Polak M, Gautier J-F, Froguel P, Rutter GA, Vaxillaire M. Insulin gene mutations resulting in early-onset diabetes: marked differences in clinical presentation, metabolic status, and pathogenic effect through endoplasmic reticulum retention. *Diabetes.* 2010;59:653–61.
 131. Bonnefond A, Philippe J, Durand E, Dechaume A, Huyvaert M, Montagne L, Marre M, Balkau B, Fajardy I, Vambergue A, Vatin V, Delplanque J, Le Guilcher D, De Graeve F, Lecoer C, Sand O, Vaxillaire M, Froguel P. Whole-exome sequencing and high throughput genotyping identified KCNJ11 as the thirteenth MODY gene. *PLoS One.* 2012;7:e37423.
 132. Bowman P, Flanagan SE, Edghill EL, Damhuis A, Shepherd MH, Paisey R, Hattersley AT, Ellard S. Heterozygous ABCC8 mutations are a cause of MODY. *Diabetologia.* 2012;55:123–7.
 133. Fajans SS, Bell GI. MODY: history, genetics, pathophysiology, and clinical decision making. *Diabetes Care.* 2011;34:1878–84.
 134. Colom C, Corcoy R. Maturity onset diabetes of the young and pregnancy. *Best Pract Res Clin Endocrinol Metab.* 2010;24:605–15.
 135. Kleinberger JW, Maloney KA, Pollin TI. The genetic architecture of diabetes in pregnancy: implications for clinical practice. *Am J Perinatol.* 2016;33:1319–26.

136. Ellard S, Beards F, Allen LI, Shepherd M, Ballantyne E, Harvey R, Hattersley AT. A high prevalence of glucokinase mutations in gestational diabetic subjects selected by clinical criteria. *Diabetologia*. 2000;43:250–3.
137. Lambrinoudaki I, Vlachou SA, Creatsas G. Genetics in gestational diabetes mellitus: association with incidence, severity, pregnancy outcome and response to treatment. *Curr Diabetes Rev*. 2010;6:393–9.
138. Weng J, Ekelund M, Lehto M, Li H, Ekberg G, Frid A, Aberg A, Groop LC, Berntorp K. Screening for MODY mutations, GAD antibodies, and type 1 diabetes--associated HLA genotypes in women with gestational diabetes mellitus. *Diabetes Care*. 2002;25:68–71.
139. Shaat N, Karlsson E, Lernmark A, Ivarsson S, Lynch K, Parikh H, Almgren P, Berntorp K, Groop L. Common variants in MODY genes increase the risk of gestational diabetes mellitus. *Diabetologia*. 2006;49:1545–51.