Participation of Hydrogen Peroxide and Nitric Oxide in Improvement of Seed Germination Performance Under Unfavourable Conditions

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Contents

Abstract Seed germination is a complex process. Upon imbibition, many factors including phytohormones (gibberellin and abscisic acid) and reactive oxygen and nitrogen species [hydrogen peroxide (H_2O_2) and nitric oxide (NO), respectively] are involved in a complicated web of interactions. While there are some impressive recent progresses made in our understanding of these interactions, it is also of great interest to investigate treatments that help seeds with difficulties to germinate under unfavourable conditions including abiotic stress factors such as chilling and heavy metals. In this chapter, an update and critical interpretations of some recent investigations into the relationships among H_2O_2 , NO, catalase activity and gene expression in cold stratification, light signal and abiotic stress are provided.

Keywords Abscisic acid $(ABA) \cdot$ Catalase \cdot Chilling stress \cdot Cold stratification \cdot Gibberellic acid · Heme oxygenase · NADPH oxidase · NO donor · Phytochrome

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1 Introduction

Emergence of the radicle through the seed coat and other seed structures that may be enclosing or associated with the embryo is the first visible sign that a seed has completed germination (Bewley et al. [2013\)](#page-10-2). This is a critical step towards the successful natural regeneration of plants in the wild as well as crop production. Seeds from some plant species may exhibit a low germination rate under otherwise favourable germination conditions for many other species. This problem has long been the subject of many studies. The possible reasons for a low rate of seed germination can be varied (Finch-Savage and Leubner-Metzger [2006](#page-10-3); Bewley et al. [2013\)](#page-10-2). For example, there may be a need for tuning in with certain environmental factors including light and cold temperature before the germinationassociated processes in the hydrated seeds can be initiated and eventually culminated in radicle protrusion through the surrounding seed structures. Seeds may be sown under unfavourable growth conditions or stress such as chilling, salinity or heavy metals, and therefore their germination performance may be impaired. The storage conditions of seeds after harvest may not be appropriate and could lead to impairment in seed germination performance. In the literature, there is a classical view about the antagonism of two phytohormones, gibberellins (GA) and abscisic acid (ABA), in the decision for a seed to germinate or not (Nelson and Steber [2017\)](#page-11-0). In the seeds with low germination performance for whatever reasons, the metabolism and signalling of ABA may predominate and ensure the no germination state (Finkelstein and Lynch [2000](#page-11-1)).

The focus on a few seeds as model lab systems, including Arabidopsis thaliana, has recently led to some rapid and impressive advances in our understanding of the interplay between reactive nitrogen species, particularly nitric oxide (NO) signalling and crosstalk with GA and ABA, occurring prior to the completion of seed germination (Nonogaki [2014](#page-11-2), [2017\)](#page-11-3). There has also been a great deal of investigations into the involvement of hydrogen peroxide (H_2O_2) in the regulation of seed germination. There may be a universal germination mechanism in different seeds. It is of great practical interest to be able to apply this knowledge to improve germination of seeds that is adversely affected by abiotic stress or less than favourable germination conditions. In this chapter, the objective is to provide an update of insights gained from the most recent papers (mainly published in between 2017 and 2018) on the participation of NO and H_2O_2 in seed germination under these conditions.

2 Cold Stratification

Freshly harvested Arabidopsis thaliana seeds (from wild-type plants of ecotype Col-0) were dormant $(1\%$ germinable seeds), but a majority of the seeds germinated after cold stratification at 4 \degree C for a few days (Bethke et al. [2004](#page-10-4)). However, supplementation of 4 or 5 mM hydrogen peroxide (H_2O_2) to a plant tissue culture

medium for seed germination led to a reduction in Arabidopsis seed germination by about 20 and 50%, respectively (Bi et al. [2017](#page-10-5)). Similar to the exogenous application of H_2O_2 , supplementation of $3-10$ mM of an inhibitor of catalase activity, 3-amino-1,2,4 triazole (3-AT), led to inhibition of the germination of the wild-type Arabidopsis seeds by about 30% (Bi et al. [2017](#page-10-5)). Catalase activity and H_2O_2 content in wild-type seeds treated with 5 mM 3-AT were only one-third and 80%, respectively, of those in the seeds germinated in the absence of the catalase activity inhibitor. This suggests that high catalase activity seems to be associated with a positive role in Arabidopsis seed germination. However, it is more complex to interpret the possible role of H_2O_2 content in cold-stratified Arabidopsis seed germination based on this result. Firstly, it is not readily clear why inhibiting catalase activity by 3-AT could lead to a reduction in endogenous H_2O_2 content in coldstratified Arabidopsis seeds as the opposite would be expected to happen.

Since abscisic acid (ABA) is well-known to be associated with germination inhibition, it is of interest to probe the relationship of catalase activity and endogenous H_2O_2 content with ABA signalling in germinating Arabidopsis seeds after cold stratification (Bi et al. [2017](#page-10-5)). Abscisic acid (ABA)-insensitive 5 (ABI5) is a basic leucine zipper transcription factor. It is the core component of the ABA signalling pathway and has been shown to affect the expression of several ABA-responsive genes (Skubacz et al. [2016\)](#page-11-4). In some earlier studies, for example, ABI5 was found to interact with the promoters of some ABA responsive genes including several late-embryogenesis-abundant genes during seed germination (Finkelstein and Lynch [2000\)](#page-11-1). Recently, it was revealed that ABI5 could bind directly to the promoter of a catalase gene, *CAT1* but not *CAT2*, in the *Arabidopsis* genome. Furthermore, the expression of CAT1was also increased by ABI5 (Bi et al. [2017\)](#page-10-5).

The possible roles of catalase activity and H_2O_2 content in cold-stratified Arabidopsis seeds were evaluated further using Arabidopsis plant lines with perturbation in sensitivity to ABA and the ABA signalling pathway. The plant lines that were investigated included mutant plants called *abi*5-1 and *abi7*, which harbour mutations in the ABI5 gene loci, and ABI5-overexpression lines. Based on the results obtained, it seems that the sensitivity of cold-stratified Arabidopsis seeds to exogenous H_2O_2 was dependent on ABA signalling (Bi et al. [2017](#page-10-5)). The cold-stratified seeds of *abi5-1* and *abi7* exhibited significantly lower germination percentages than wild type in response to exogenous application of $3-5$ mM of H_2O_2 . Conversely, overexpression of ABI5 seemed to be associated with the insensitivity to the germination inhibitory effect of exogenous application of H_2O_2 . These results would seem to be consistent with the possibility that catalase activity played a positive role in the germination of the cold-stratified Arabidopsis seed. Based on the link between ABI5 and catalase gene expression, there would be an elevated level of catalase activity in ABI5-overexpression lines. Therefore any excess H_2O_2 from exogenous application could be decomposed so that germination process in the cold-stratified seeds was insensitive to the excess H_2O_2 (Bi et al. [2017\)](#page-10-5).

Consistent with this, the seeds of the ABI5-overexpression lines and the abi5-1 mutant plants exhibited a higher and lower catalase activity, respectively, than the

wild type (Bi et al. [2017](#page-10-5)). This finding was correlated with that of exogenous application of H_2O_2 having little or more germination inhibitory effect on the overexpression lines or the mutant plants, respectively, than the wild type. This result was in agreement with the results on the germination responses to 3 to 10 mM 3-AT of the cold-stratified seeds of abi5-1 and abi7 and ABI5-overexpression lines compared to wild type. Their germination responses mirrored those of the plant lines exposed to exogenous application of H_2O_2 (Bi et al. [2017\)](#page-10-5). Therefore, it seemed that inhibition of catalase activity by 3-AT, like the seed germination response to exogenous application of H_2O_2 , was also dependent on ABI5 expression levels (Bi et al. [2017](#page-10-5)). Catalase in cold-stratified Arabidopsis seeds would be working downstream of ABI5 in germinating seeds.

There is, however, evidence that calls for some caution in accepting a positive role for catalase in Arabidopsis seed germination. ABI5 expression levels were linked to gene expression of CAT1 which was higher in the cold-stratified seeds of the ABI5-overexpression lines than wild type. The cold-stratified abi5-1 seeds exhibited a lower level of CAT1 expression than wild type. As expected from the lack of interaction between ABI5 and the promoter of CAT2 in a yeast-one hybrid system, there was no difference in the expression levels of CAT2 in the wild type and the abi5-1 and ABI5-overexpression lines. Interestingly, the remaining member of the CAT gene family, CAT3, exhibited the lowest and highest expression levels in the seeds of the wild type and *abi5-1*, respectively. As far as germination percentage was concerned, there was, however, no difference exhibited by the seeds of wild type, abi5-1 and ABI5-overexpression lines when the cold-stratified seeds were sown in a plant tissue culture medium in the absence of exogenous H_2O_2 . Therefore, it is not clear how the cold stratification treatment would enable seeds to germinate independently of their different ABI5 expression levels. The relative significance of the varying catalase gene expression levels in the germination process must also await further elucidation.

Many seeds require a prior exposure to cold stratification conditions before they can germinate at a higher temperature. It would be interesting to investigate the changes in the catalase activity and endogenous H_2O_2 content in other cold stratification-requiring seeds to ascertain whether the finding from the studies on the Arabidopsis model system is translational to other non-model seeds. However, a survey of the recent literature shows a gap in our knowledge in this direction. For example, about 30% of Zanthoxylum nitidum seeds (of the Rutaceae family, a common Chinese medicinal plant) germinated at $15 \degree C$ after cold stratification at 4° C for 3 months, while less than 3% of the non-cold-stratified seeds germinated (Lu et al. [2018](#page-11-5)). As expected from the classical literature about the involvement of gibberellins in the control of seed germination, the seeds without cold stratification but treated with gibberellic acid (300 mg L^{-1}) exhibited about 22% germination which was significantly higher than control. Based on the proteomic analytical approach iTRAQ-coupled LC-MS/MS, 484 proteins in the cold-stratified seeds were more abundant than the non-cold-stratified seeds (Lu et al. [2018](#page-11-5)). Only about eight proteins in the category of detoxification function, which could potentially include catalase gene, were found to be either up- or downregulated in association

with dormancy release. This is highly speculative but it is worthwhile to validate this further through direct measurement of catalase activity and determination of H_2O_2 content in the cold-stratified, GA-treated seeds and the control.

3 Abiotic Stress-Related Suppression of Seed Germination

3.1 Chilling Stress

At 10 $^{\circ}$ C for 10 days, about 20% of *Hedysarum scoparium* seeds germinated compared to 80% germination at 25 °C (Su et al. [2016](#page-11-6)). Interestingly, about 80% of the seeds could also germinate after 10 days at 10 \degree C provided the seeds were prior treated with 10 days of cold stratification (seeds mixed with sand at 4° C). Therefore, the prior cold stratification aided the seeds to overcome chilling stress on seed germination. Exogenous application of 50 mM H_2O_2 promoted maximum germination (80%) of the non-cold stratified seeds at 10 $^{\circ}$ C. Exogenous application of an antioxidant, 50 mM N-acetyl cysteine (NAC), effectively prevented the stimulation of germination by 50 mM H_2O_2 . This result is contrary to that implicating the need for preventing excess H_2O_2 content in cold-stratified, germinating Arabidopsis seeds (Bi et al. [2017](#page-10-5)).

In situ detection using confocal microscopy of H_2O_2 production at the tip of the embryonic axes of H. scoparium after 24 h of imbibition showed that there was only slightly detectable H_2O_2 content in the non-cold-treated seeds, but there was elevated accumulation of H_2O_2 in cold-stratified seeds imbibed in water and non-coldstratified seeds treated with 50 mM exogenous H_2O_2 (Su et al. [2016](#page-11-6)). The treatment of the cold-stratified seeds with 50 mM NAC effectively nullified the endogenous H2O2 production. Hence, ROS generation following cold stratification of the seeds plays a positive role in seed germination. It would be worthwhile in future studies to evaluate the status of the catalase activity and gene expression in this system.

The cold-stratified H. scoparium seeds had lower ABA content than non-coldstratified seeds that germinated poorly at $10\degree C$. There was an inverse correlation between H_2O_2 and ABA contents in the embryonic axes of H. scoparium (Su et al. [2016\)](#page-11-6). Exogenous application of ABA prevented cold-stratified seeds from germinating and lowered H_2O_2 accumulation. This result suggests that ABA could influence upstream of ROS metabolism which was in turn linked to the completion of the germination process. However, exogenous application of H_2O_2 that enabled non-cold-stratified seeds to germinate led to a 20% reduction in ABA contents in the seeds within 24 h of imbibition. Conversely, NAC treatment of cold-stratified seeds led to an increase in ABA correlating with inhibition of seed germination. Hence, the precise relationship between changes in ABA and H_2O_2 contents in relation to the decision to germinate or not is still not completely clear (Su et al. [2016](#page-11-6)).

Seeds that normally germinate at a higher temperature can be prevented from germination at low temperatures. For example, 96% maize seeds germinated after 3–4 days at 25 °C, but after 3 and 4 days at 13 °C, only 15% germinated, respectively (Li et al. [2017b\)](#page-11-7). The seeds at 25 °C were found to have a higher endogenous H_2O_2 content as well as activities of catalase and other antioxidative enzymes than those at 13 °C after 48 h from sowing. Priming maize seeds with 50 mM H_2O_2 for 24 h enabled 50 and 90% of the seeds germinated at 13 \degree C after 3 and 4 days, respectively (Li et al. [2017b](#page-11-7)). Seed priming with H_2O_2 also led to a higher catalase activity at 48 h from sowing the primed seeds under chilling stress than non-primed seeds. The activities of other antioxidative enzymes assayed including ascorbate peroxidase, another enzyme beside catalase that can decompose H_2O_2 , were also increased following seed priming with 50 mM H_2O_2 . These results suggest that the net accumulation of H_2O_2 and probably other ROS was an important force for maize seed germination to occur.

There was no difference in the expression of $ZmNCED1$ (a key gene for ABA biosynthesis in maize) and ZmCYPT707A2 (a key gene for ABA catabolism) between non-primed and H_2O_2 -primed maize seeds after 48 h of imbibition. However, the H_2O_2 -primed maize seeds exhibited higher levels in the expression of the two genes after 72 h at 13 °C (Li et al. [2017b](#page-11-7)). Based on these results, it is difficult to interpret the relative contribution of ABA biosynthesis and ABA catabolism in H_2O_2 -primed maize seeds without determination of ABA contents in the seeds. In contrast, the positive involvement of increased GA biosynthesis with concomitant reduction in GA catabolism in germination of H_2O_2 -primed maize seeds was clearly evident early (6–24 h) during imbibition. Nevertheless, since higher levels of H_2O_2 and catalase activity were found in H_2O_2 -primed than in non-primed maize seeds after 48 h of imbibition, this suggests that ROS homeostasis-related to seed germination might work upstream of ABA metabolism but downstream of GA metabolism (Li et al. [2017b\)](#page-11-7).

3.2 Salinity and Heavy Metal Stress

Germination of alfalfa seeds in water and 150 mM NaCl (salinity stress) was about 98 and 45%, respectively (Amooaghaie and Tabatabaie [2017\)](#page-10-6). Pretreatment of the seeds with 2 mM H_2O_2 or 300 mg L⁻¹ polyethylene glycol (PEG 6000, osmopriming) for 6 h enabled more seeds (about 70%) to germinate in the presence of the salinity stress. Both priming with exogenous H_2O_2 and PEG 6000 led to a higher level of endogenous H_2O_2 than non-primed alfalfa seeds after 6 h from exposure to 150 mM NaCl. Application of N,N-dimethylthiourea (DMTU, a chemical absorbent of H_2O_2 and a ROS scavenger) prevented the stimulation of germination under salinity stress by priming with H_2O_2 or PEG 6000. In addition, application of DMTU suppressed an increase in endogenous H_2O_2 by priming the alfalfa seeds with H_2O_2 or PEG 6000. Interestingly, there was an increase in heme oxygenase activity and heme-oxygenase gene $(HO-I)$ expression. These results suggest a sequence of signalling events involving endogenous H_2O_2 and heme oxygenase for osmopriming to overcome salinity stress on alfalfa seed germination (Fig. [1\)](#page-6-0). Heme oxygenase coupled with carbon monoxide signalling has been

thought to be part of the response of plant cells under different environmental stress and ABA and may play a cytoprotective role (Shekhawat and Verma [2010](#page-11-8)).

Germination of maize seeds in water and 5 mM PbCl_2 (under Pb stress) was about 98 and 25% after 72 h from sowing (Zhang et al. [2018\)](#page-11-9). The Pb stress-induced inhibition of seed germination could be reversed by an inhibitor (imidazole or diphenyliodonium) of NADPH oxidase (NOX) activity which is responsible for the production of superoxide free radical (O_2) . Superoxide may be scavenged by superoxide dismutase to generate H_2O_2 . Interestingly, EDTA (a metal chelator) or DMTU (a trap for H_2O_2) was not able to counteract the Pb stress-induced inhibition of seed germination. This suggests that although endogenous H_2O_2 content in maize seeds during imbibition was also increased under Pb stress, the Pb stress-triggered production of O_2 ⁻⁻ is a key suppressor of seed germination. Similar results were obtained in rice seeds germinated in the absence of abiotic stress (Li et al. [2017a](#page-11-10)).

3.3 Seed Storage Conditions

Two batches of barley seeds which were from the same harvest but were stored under two different conditions resulted in different germination performances under the same imbibition conditions: the seeds of higher germination performance were stored at 23 °C for 6 months, and those of lower germination performance were stored at -28 °C (Ishibashi et al. [2017](#page-11-11)). The level of H_2O_2 as a marker of reactive oxygen species (ROS) production in the batch of barley seeds exhibiting higher germination performance, both in relation to germination speed and higher germination percentage, was higher than in the other batches of barley seeds of lower germination performance. The significance of this correlation between hydrogen peroxide level and germination performance was supported with exogenous application of hydrogen peroxide and ascorbate (an antioxidant that resulted in a lower level of hydrogen peroxide in the seeds) during seed imbibition. After 48 h of imbibition in water, the germination percentage of high germination performing seeds was about 80%, but it was reduced significantly to about 20% in the presence of 20 mM sodium ascorbate. Conversely, the seeds of low germination performance exhibited a higher germination percentage (45%) when imbibed in 100 mM hydrogen peroxide than that of those imbibed in water (30%) (Ishibashi et al. [\(2017](#page-11-11))).

Higher H_2O_2 and lower ABA contents were found in the barley seeds of higher germination performance compared to the batch of seeds of lower germination performance after 48 h of imbibition (Ishibashi et al. [2017](#page-11-11)). While there was little change in the expression of $HvNECDI$ (a key gene for ABA biosynthesis in barley), that of $HvABA8'$ -OH1 (encoding ABA-8'-hydroxylase for ABA catabolism) was significantly higher in the barley seeds of higher germination performance than those of lower germination performance.

Higher H_2O_2 content in the barley seeds of higher germination performance was correlated with higher NADPH oxidase activity. On the other hand, of the two H_2O_2 scavenger enzymes, ascorbate peroxidase activity was higher in the barley seeds of high germination performance than those of low germination performance, while catalase activity showed the opposite relationship with the two batches of seeds during imbibition. It seems difficult to decipher the relative contributions of these two antioxidative enzymes on H_2O_2 signalling in seed germination. Further studies might include simultaneous investigations into the effect of exogenous ABA on the status of both ascorbate peroxidase and catalase at the levels of gene expression and enzyme activity in relation to the germination of the two batches of barley seeds.

4 The Scientific Basis for Improving Seed Germination by Exogenous Nitric Oxide

In many studies, application of exogenous NO has been found to improve seed germination performance due to diverse circumstances, although only some studies validated the effect with a NO-specific scavenger and evaluate the physiological significance of endogenous NO content (Table [1](#page-9-0)). For example, isolated embryos of mountain ash (Sorbus pohuashanesis) placed on filter paper wetted with distilled water for 8 days exhibited 42% germination. The promoter effect of pretreatment with 2 mM sodium nitroprusside (SNP, a nitric oxide donor) for 3 h, compared to pretreatment with water only, on germination of isolated embryos of mountain ash (S. pohuashanesis) on filter paper wetted with distilled water has been demonstrated to be 80 and 42%, respectively (Yang et al. [2018](#page-11-12)). The embryos treated with a combination of 2 mM SNP and 0.3 mM cPTIO (a NO-scavenger) did not exhibit dormancy release as only 42% of the embryos germinated. Furthermore, germination of embryos treated with 0.3 mM cPITO alone was slightly lower than imbibition in water only.

Upon seed imbibition, ABA in seeds could stimulate the expression of ABI5 which is a key repressor of seed germination. There are four ways that exogenous application of NO or endogenous NO produced after start of seed imbibition can interfere with regulation of ABA in seed germination. For example, NO or other RNS (reactive nitrogen species) could inactivate the protein signalling intermediates between ABA and ABI5 by tyrosine nitration in these proteins (Signorelli and Considine [2018](#page-11-13)). As a result, the ABI5 expression level would be reduced and hence has a less stronghold on suppression of seed germination.

In lettuce seeds, it was shown that light could stimulate NO production (An and Zhou [2017](#page-10-7)). Then NO was thought to in turn stimulate the activity of phospholipase D activity which is responsible for the hydrolysis of phospholipids to phosphatidic acid. Presumably, the phosphatidic acid formed could interact with phytohormones such as GA and ABA to allow lettuce seed germination in the light. The lightstimulated NO production in Arabidopsis thaliana seeds seems to be able to interact with the well-known phytochrome signalling in light-sensitive seed germination (Li et al. [2018](#page-11-14)). Further investigations are warranted to investigate between phosphatidic acid formation and phytochrome signalling in light-regulated seed germination.

NO could also interact with GA, a well-established germination promoter, and other phytohormones to release seed dormancy. For example, less than 10% of Amaranthus retroflexus seeds germinated at 25° C in light. However, exogenous application of potassium nitrite (KNO_{2, a} NO donor) for 5 h or 10^{-3} M GA₃ led to 60% seed germination (Kepczynski et al. [2017](#page-11-15)). Likewise, exogenous application of ethylene or an ethylene biosynthesis precursor stimulated seed germination. Ethylene production seemed to be associated with the action of GA or NO in promoting germination. It was validated further that NO worked with GA or ethylene in germination stimulation because cPTIO blocked the effect of GA or ethylene.

Table 1 Some recent examples of seed germination stimulation following NO pretreatment Table 1 Some recent examples of seed germination stimulation following NO pretreatment

n.a. =
SNP = not determined sodium nitroprusside

¼SNAP

S-nitroso-N-acetylpenicillamine

154 D. W. M. Leung

The relative effectiveness of exogenous application of H_2O_2 and NO (SNP as NO donor) on promoting coriander and carrot seed germination was studied (Panngom et al. [2018\)](#page-11-17). About 40% seeds germinated in water, but pretreatment with 25 mM H₂O₂ or 12.5 μM SNP led to 90 and 80% seed germination, respectively. Pretreatment of carrot seeds with $25-200$ mM H₂O₂ resulted in about 70% seed germination compared to 60% germination in water. Pretreatment of carrot seeds with $0-100 \mu M$ SNP did not have any effect seed germination. This suggests that the sensitivity of different seeds to NO and H_2O_2 signalling may vary as far as seed germination is concerned.

5 Conclusion

Seed germination research is justified given its agricultural importance and implication for natural plant regeneration to maintain survival and propagation of wild plants. During imbibition, the current seed germination research communities seem to have fully embraced the concept that H_2O_2 and NO are important signal partners in the complex web of molecules including the well-known germination promoter and inhibitor played by GA and ABA, respectively. The details as to how either $H₂O₂$ or NO interacts with these key germination regulators are being revealed in different studies including the improvement in seed germination under unfavourable germination conditions or abiotic stress. There is yet no comprehensive study to investigate in detail the interrelationships of both H_2O_2 and NO and how they would interact with GA and ABA metabolism and signalling in the same seed during imbibition.

References

- Amooaghaie R, Tabatabaie F (2017) Osmopriming-induced salt tolerance during seed germination of alfalfa most likely mediates through H_2O_2 signaling and upregulation of heme oxygenase. Protoplasma 254:1791–1803
- An CF, Zhou CJ (2017) Light induces lettuce seed germination through promoting nitric oxide production and phospholipase D-derived phosphatidic acid formation. South Afr J Bot 108:416–422
- Bethke PC, Gubler F, Jacobsen JV, Jones RL (2004) Dormancy of Arabidopsis seeds and barley grains can be broken by nitric oxide. Planta 219:847–855
- Bewley DJ, Bradford KJ, Hilhorst HWM, Nonogaki H (2013) Seeds: physiology of development, germination, and dormancy. Springer, New York
- Bi C, Ma Y, Yu YT, Liang S, Lu K, Wang XF (2017) Arabidopsis ABI5 plays a role in regulating ROS homeostasis by activating CATALASE 1 transcription in seed germination. Plant Mol Biol 94:197–213
- Finch-Savage WE, Leubner-Metzger G (2006) Seed dormancy and control of seed germination. New Phytol 171:501–523
- Finkelstein RR, Lynch TJ (2000) The Arabidopsis abscisic acid response gene ABI5 encodes a basic leucine zipper transcription factor. Plant Cell 12:599–609
- Ishibashi Y, Aoki N, Kasa S, Sakamoto M, Kai K, Tomokiyo R, Watabe G, Yuasa T, Iwaya-Inoue MI (2017) The interrelationship between abscisic acid and reactive oxygen species plays a key role in barley seed dormancy and germination. Front Plant Sci 8:275
- Kepczynski J, Cembrowska-Lech D, Sznigir P (2017) Interplay between nitric oxide, ethylene, and gibberellic acid regulating the release of Amaranthus retroflexus seed dormancy. Acta Physiol Plant 39:254
- Li WY, Chen BX, Chen ZJ, Gao YT, Chen Z, Liu J (2017a) Reactive oxygen species generated by NADPH oxidases promote radicle protrusion and elongation during rice seed germination. Int J Mol Sci 18:110
- Li Z, Xu J, Gao Y, Wang C, Guo G, Luo Y, Huang Y, Hu W, Sheteiwy MS, Guan Y, Hu J (2017b) The synergistic priming effect of exogenous salicylic acid and H_2O_2 on chilling tolerance enhancement during maize (Zea mays L.) seed germination. Front Plant Sci 8:1153
- Li R, Jia Y, Yu L, Yang W, Chen Z, Chen H, Hu X (2018) Nitric oxide promotes light-initiated seed germination by PIF1 expression and stabilizing HFR1. Plant Physiol Biochem 123:204–212
- Lu Q, Zhang ZS, Zhan RT, He R (2018) Proteomic analysis of Zanthoxylum nitidum seeds dormancy release: influence of stratification and gibberellin. Ind Crop Prod 122:7–15
- Mao C, Zhu Y, Chen H, Yan H, Zhao L, Tang J, Ma X, Mao P (2018) Nitric acid oxide regulates seedling growth and mitochondrial responses in aged oat seeds. Int J Mol Sci 19:1052
- Nelson SK, Steber CM (2017) Transcriptional mechanisms associated with seed dormancy and dormancy loss in gibberellin-insensitive sly1-2 mutant of *Arabidopsis thaliana*. PLoS One 12: e0179143
- Nonogaki H (2014) Seed dormancy and germination-emerging mechanisms and new hypotheses. Front Plant Sci 5:233
- Nonogaki H (2017) Seed biology updates-highlights and new discoveries in seed dormancy and germination research. Front Plant Sci 8:524
- Panngom K, Chuesaard T, Tamchan N, Jiwchan T, Srikongsritong K, Park G (2018) Comparative assessment for the effects of reactive species on seed germination, growth and metabolisms of vegetables. Scient Hort 227:85–91
- Shekhawat GS, Verma K (2010) Haem oxygenase (HO): an overlooked enzyme of plant metabolism and defence. J Exp Bot 61:2255–2270
- Signorelli S, Considine MJ (2018) Nitric oxide enables germination by a four-pronged attack on ABA-induced seed dormancy. Front Plant Sci 9:296
- Skubacz A, Daszkowska-Golec A, Szarejko I (2016) The role and regulation of ABI5 (ABA-Insensitive 5) in plant development, abiotic stress responses and phytohormone crosstalk. Front Plant Sci 7:1884
- Su L, Lan Q, Pritchard HW, Xue H, Wang X (2016) Reactive oxygen species induced by cold stratification promote germination of *Hedysarum scoparium* seeds. Plant Physiol Biochem 109:406–415
- Yang L, Zhang D, Liu H, Wei C, Wang J, Shen H (2018) Effects of nitric oxide donor and nitric oxide scavengers on Sorbus pohuashanensis. J For Res 29:631–638
- Zhang Y, Deng B, Li Z (2018) Inhibition of NADPH oxidase increases defense enzyme activities and improves maize seed germination under Pb stress. Ecotoxicol Environ Saf 158:187–192