

Insect Allergy: A Review of Diagnosis and Treatment **30**

James M. Tracy and Jeffrey G. Demain

Contents

30.1	Introduction: Terminology, Types of Reactions, and History				
30.2	Insect Biology, Terminology, and Identification	680			
30.3	Diagnosis	684			
30.4 30.4.1	Diagnostic Testing	685 685			
30.5	Treatment	686			
30.6	Large Local Reactions and VIT	688			
30.7	Duration of VIT	688			
30.8	Recent Developments in Insect Allergy	688			
30.9	Conclusion	689			
References					

J. M. Tracy (🖂)

Allergy, Asthma and Immunology Associates, P.C, Omaha, NE, USA

Division of Allergy and Immunology, Creighton University College of Medicine, Omaha, NE, USA

Creighton University, Omaha, NE, USA e-mail: jmtracy@cox.net

J. G. Demain

Department of Pediatrics/Allergy Asthma and Immunology Center of Alaska, University of Washington, Anchorage, AK, USA

WWAMI School of Medical Education, University of Alaska, Anchorage, AK, USA e-mail: jdemain@allergyalaska.com

© Springer Nature Switzerland AG 2019 M. Mahmoudi (ed.), *Allergy and Asthma*, https://doi.org/10.1007/978-3-030-05147-1_31

Abstract

Insect allergy is the third most common cause of the life-threatening condition anaphylaxis, following food and medications. Insect allergy anaphylaxis poses risk of considerable morbidity and mortality. Avoidance of the offending agent is the cornerstone to the management anaphylaxis regardless of the cause. However, unlike food and medication allergy, insect allergy has been effectively treated, using well-established protocols for many years. Hymenoptera are the insects most associated with allergy and anaphylaxis with at least 40 deaths per year attributed to insect stings in the United States. It is critical that healthcare professionals and the public understand the proper diagnosis as well as the longterm treatment of this potentially life-threatening allergy. Insect allergy from Hymenoptera, managed prospectively using venom immunotherapy, conveys up to 98% protection of anaphylaxis with future stings. Insects of the order Hymenoptera include bees, wasps, hornets, yellow jackets, and stinging ants. Stinging ant allergy will not be reviewed in this chapter. An understanding of the biology and habitat of the various Hymenoptera species is helpful in recommending insect avoidance strategies. The diagnosis of insect allergy relies on a history of a systemic allergic reaction followed by appropriate testing for venom-specific IgE. If the history of a generalized anaphylactic reaction to an insect sting and the presence of venom-specific IgE are confirmed, venom immunotherapy is indicated. It is venom immunotherapy, a disease modifying therapy, that provides the most effective protection against future sting reactions. Ultimately, recognition and lifesaving management is critical. Subsequently, evaluation and potentially long-term management of insect allergy include appropriate referral to an allergist familiar with insect allergy and, if indicated, venom immunotherapy.

Keywords

Insect · Hymenoptera · Anaphylaxis · Epinephrine

Insects are one of the three most common allergic triggers for anaphylaxis, the others being foods and medications (Simons 2008; Simons et al. 2007; Sampson et al. 1992; Simons and Sampson 2008). Insect allergy results in significant morbidity and mortality, with potentially life-threatening systemic reactions occurring in 0.4% to 0.8% of children and up to 3% of adults, and accounts for at least 40 deaths annually in the United States (Graft 2006; Schwartz et al. 1995). Under recognition and treatment may actually underestimate the true mortality from insect anaphylaxis (Graft 2006; Schwartz et al. 1995; Golden et al. 2011). With proper evaluation and treatment, the risk of a

severe event with a subsequent sting can be dramatically diminished. Venom immunotherapy (VIT) can provide up to a 98% level of protection from future insect-related anaphylactic events (Golden et al. 2011; Valentine 1984; Hunt et al. 1978; Reisman and Livingston 1992). This chapter will address the current state of knowledge about insect allergy, including insect identification, diagnosis, and evaluation, as well as longand short-term evaluation and treatment.

30.1 Introduction: Terminology, Types of Reactions, and History

Insects belonging to the order Hymenoptera account for the majority of serious sting-related reactions. Within this order, three families are medically relevant. These include the Apidae, Vespidae, and Formicidae families. The Apidae family includes honeybees and bumblebees; the Vespidae family includes yellow jackets, white-faced hornets, yellow hornets, and wasps; the Formicidae family includes primarily imported fire ants and harvester ants (Gurlanick and Benton 2003; Goddard 2003). The family Vespidae includes the genus Polistes or wasps. In North America P. annularis, P. fuscatus, P. metricus, and P. exclamans are the predominant species. In Europe P. dominulus, P. gallicus, and P. nimphus are widespread. Although there is some cross-reactivity between American and European Polistes species, there are significant differences to warrant different testing and treatment venoms (Severino et al. 2006).

Anaphylaxis to stings of the imported fire ant and to bites from reduviids and mosquitos is reviewed in a separate chapter. Non-Hymenoptera stinging and biting arthropods, such as scorpions and spiders, are more extensively reviewed elsewhere and will not be the focus of this work (Demain 2003; More et al. 2004).

30.2 Insect Biology, Terminology, and Identification

Knowledge of these Hymenoptera insects, their biology, habits, and dwellings, can assist in recognition of the insect and circumstance of sting, though this information should not be relied upon solely in identification of the offending insect. This knowledge of the circumstance and the suspect insect can be helpful for the diagnosis and treatment of insect allergy (Gurlanick and Benton 2003; Goddard 2003).

Yellow jackets can be either ground dwelling or in nests above ground. *Vespula vulgaris* are generally ground-dwelling yellow jackets, commonly encountered during outdoor activities. *V. vulgaris* can be very aggressive after even minimal provocation, particularly with vibration, such as a leaf blower or weed whacker (Fig. 1). A second species of yellow jacket (*Dolichovespula arenaria*) nests above ground, usually in shrubs and trees. Yellow jackets are carnivorous, have smooth bodies with straight barbless stingers, and can sting multiple times.

Wasps (*Polistes*) are also carnivorous and smooth bodied. The nests of wasps can be distinguished from yellow jackets by the triangular, open-celled configuration without the outer paper encasement typical of other vespids (Fig. 2). Wasp nets are frequently found under the eaves of houses and barns.

Domestic or European honeybees are herbivorous with hairy bodies and have a barbed stinger that results in evisceration and their death after the sting. Typically, they are nonaggressive unless protecting their hives; as a result, honeybee stings are often accidental and occur in children and adults who, while barefoot, inadvertently step on them in the grass (Fig. 3). Africanized honeybees were imported to South America from Africa and have been migrating north to the United States. Unlike their domestic counterparts, they are very aggressive. The venom from Africanized honeybees is identical to their domestic cousins, and the venom volume per sting is similar. However, unlike the single sting of a domestic honeybee, Africanized honeybees often sting in large numbers and will pursue their victim for much longer distances. The domain of the Africanized honeybee is currently limited in the United States to Texas, New Mexico, Arizona, Nevada, and California (Golden et al. 2011) (Fig. 4). Imported fire ants (Formicidae), which are discussed in other chapters, also have limited, but similar domains in



Fig. 1 Yellow jacket (Photograph courtesy of Dr. Jeffrey G. Demain)



Fig. 2 Wasp (Photograph courtesy of Dr. Jeffrey G. Demain)



Fig. 3 Honeybee (Photograph courtesy of Dr. Jeffrey G. Demain)

the Southern United States (Golden et al. 2011) (Fig. 5).

Unfortunately, the absolute identification of the culprit insect usually cannot be confirmed, so testing with each of the common venoms is







Fig. 5 Imported fire ant quarantine. (Regularly updated maps of the fire ant range and agriculture quarantine areas within the United States (Golden et al. 1989))

warranted in almost all cases, which will be discussed later. There are some circumstances where the offending insect is more obvious. As mentioned, honeybees have barbed stingers, and usually their venom sac can become lodged in the skin following a sting event. While this can be helpful in identification, it is important to note that yellow jackets may also leave the stinger embedded in the skin. In the case of imported fire ants, the presence of a pseudo-pustule up to 24 h later is virtually diagnostic of a fire ant sting (Golden et al. 2011; Moffitt 2003). When taking a history, it is important to take into account historical elements such as the person's activity at the time of the sting, insect activity in the area where the patient was stung, time of the year, and/or geographical considerations (Moffitt 2003).

The amount of venom delivered with a single sting varies between species. A single imported

fire ant sting may contain up to 100 ng of venom, while in the case of honeybees, yellow jackets, hornets, and wasps, each sting can range from 20 to 50 mcg (Hoffman and Jacobson 1984). Hymenoptera venoms contain a variety of peptide and protein components. It is these components that cause the characteristic local reactions consisting of redness, swelling, and pain. Individuals having been previously stung may have generated venom-specific IgE antibodies, placing that individual at risk for a potential life-threatening anaphylaxis with subsequent stings. Individual Hymenoptera species contain some shared venom antigenic components. There is considerable immunologic cross-reactivity and sensitization between hornet and yellow jacket venoms, though there is much less between yellow jacket and hornet with wasp venoms. The immunogenic cross-reactivity and sensitization are even less common between honeybee and the other venoms (Hoffman 1993; King et al. 1985; Reisman et al. 1982). Bumblebee (Bombus terrestris) venom has variable cross-reactivity and sensitization with honeybee venom, though at least two antigens are unique. Because bumblebees are nonaggressive, allergic reactions to bumblebee field stings are rare in the United States compared to other Hymenoptera stings. In Europe, bumblebees are used for pollination in greenhouses; therefore more frequent allergic reactions have been reported, particularly among greenhouse workers. Specific venom to bumblebee would be optimal for skin testing and immunotherapy but is currently not available in the United States (Franken et al. 1994; Hoffman et al. 2001; Freeman 2004; De Root 2006) (Table 1).

30.3 Diagnosis

Common

names

Diagnosis of Hymenoptera allergy is based upon a comprehensive clinical history, the presence of allergic symptoms consistent with anaphylaxis, and objective evidence of venom-specific IgE antibodies. Accurate diagnosis is critical as once

Nesting habits

the thorough history supports that a generalized systemic reaction to a sting occurred, and the presence of venom-specific IgE is confirmed, the patient becomes a candidate for venom-specific immunotherapy (VIT) (Franken et al. 1994; Hoffman et al. 2001; Freeman 2004; De Root 2006). Proper treatment with VIT can result in up to 98% protection from future life-threatening sting events.

In the majority of cases, the insect sting is reported by the patient; however, it is important to know that there are reports of systemic events occurring without the patient realizing they have been stung. Following an insect sting, the initial diagnostic question is to determine whether the sting reaction is localized, cutaneous such as hives or angioedema, or a more severe systemic reaction (Golden et al. 2006). After a sting, most people develop only minor local symptoms, limited to local pain, tenderness, and swelling; these reactions are self-limited, lasting between 48 and 72 h. A local reaction is defined as a reaction in which the swelling and redness are confined to the tissues contiguous to the sting site. Large local reactions are based on size and vary from 5 to 8 cm to greater than 10-16 cm. It is estimated that large local reactions make up 5-15% of sting events

Avoidance strategies

Feeding habit

 Table 1
 Hymenoptera biology and habitat
Taxonomic

classification

Honeybee	Family Apidae	Commercial hives	Nectar and pollen flowering	Print clothing and wearing floral scents;
Yellow jacket	Family Vespidae	Multilayered, usually underground; although there is	Scavengers, aggressive	Avoid open food sources, picnic areas, garbage;
5	Vespula species	also an aerial yellow jacket: Dolichovespula arenaria ^b	Carnivorous	destroy in-ground nests
Paper wasp	Family Vespidae <i>Polistes</i> species	Hangs from eaves and porches	Nectar and arthropods	Avoid flower-print clothing and wearing floral scents; remove nests when possible
White- faced hornet	Family Vespidae Dolichovespula species	Multilayered, open areas	Nectar and arthropods	Avoid flower-print clothing and wearing floral scents; remove nests when possible
Fire ant	Family Formicidae	Earthen mounds in Southern United States	Omnivorous	Avoid mounds; wear shoes, sock, and gloves
^a A subspecies	of honevbee exists i	n South Texas. Central and South An	nerica called "Africar	nized." It is more aggressiv

e than local species and is clinically relevant in regions of infestation

^bEuropean species include P. dominulus, P. gallicus, and P. nimphus

(Golden et al. 2011). By contrast, systemic reactions, though occasionally delayed, are generally immediate-type hypersensitivity, mediated by venom-specific IgE. Systemic reactions involve signs and symptoms distant from the immediate sting site; the symptoms may range from mild to life-threatening. Mild systemic reactions, also termed cutaneous reactions, are typically limited to minimal flushing, urticaria, or angioedema. While some serious reactions may begin 15-30 min or longer after the sting, most serious reactions occur within minutes of the sting event. Generalized systemic reactions may include bronchospasm, gastrointestinal symptoms, hypotension, diaphoresis, shock, and - the most common cause of fatalities laryngeal edema.

30.4 Diagnostic Testing

Once the history of a systemic reaction to an insect sting has been established, the next step is to discern the presence of venom-specific IgE. It is important to note that up to 27% of the general population may have detectable levels of venomspecific IgE, so the presence of venom-specific IgE without a history of a systemic reaction may not be predictive of a future insect-related anaphylactic event (Golden et al. 1989). As a result, skin testing is not indicated unless the patient has a history of a systemic allergic reaction other than hives to an insect sting. All individuals, regardless of age, with a history of a systemic or anaphylactic reaction, beyond hives and/or angioedema, following an insect sting should be tested (Golden et al. 2006, 2011; Light et al. 1977; Reisman 2005). Recently new guidance has emerged regarding testing and VIT in individuals with systemic anaphylactic reactions limited to cutaneous involvement (Golden et al. 2011). In the 2017, Golden et al. outlined changes for individuals with limited cutaneous systemic reactions to stinging insects and who required testing and ultimately therapy. Adults and children who have reactions limited to the skin, such as hives and angioedema, appear not to have a significant risk for more severe reactions in the future, and therefore testing is not warranted (Georgitis and Reisman 1985; Golden et al. 2017). This is a change from the previous recommendations, where adults, but not children younger than 16 years, warranted testing for hives and/or angioedema (Golden et al. 1997, 2011, 2017; Georgitis and Reisman 1985). Sensitivity can persist for many years, even in cases of an intervening sting without a reaction; as a result, testing should be performed regardless of when the systemic sting event occurred.

30.4.1 Methods

The next consideration is the selection of the method for allergy testing. Skin testing to specific venom is the gold standard for identifying venomspecific IgE. In general, skin testing is preferred over in vitro methods for initial assessment because skin testing is more sensitive and usually less costly (Hamilton 2001, 2004) and should be performed by an allergist/immunologist who has training and experience in the diagnosis and treatment of insect allergy (Golden et al. 2011). Skin testing for Hymenoptera venom is most commonly performed using a combination of epicutaneous (prick/puncture) and intracutaneous (intradermal) methods accompanied by appropriate positive and negative controls. Testing for Hymenoptera venoms usually begins with skin prick testing at 100mcg/ml concentrations and if negative followed by intracutaneous testing starting at venom concentration of between 0.001 and 0.01 mcg/ml. At intervals of 20-30 min, the skin tests are preformed using tenfold increase in concentration until a positive skin test response occurs - or a maximum concentration of 1.0 mcg/ml is administered. Venom concentrations greater than 1.0 mcg/ml are associated with an increase in irritant skin reactions or falsely positive results. A positive skin test reaction at a concentration $\leq 1.0 \text{ mcg/ml}$ confirms the presence of venom-specific IgE antibodies (Georgitis and Reisman 1985; Golden et al. 2017). Whole-body extract is the only reagent available for testing in imported fire ant patients suspected of having fire ant hypersensitivity and is discussed in later chapters. Venom skin testing is

positive in 70-90% of patients with a significant history of a systemic reaction (Valentine 1984; Hunt et al. 1978; Reisman 2005; Golden et al. 1997; Parker et al. 1982). Since the stinging insect cannot always be reliably identified, physicians should test all relevant insects for the geographic area in question. For most areas in the United States, skin testing should include testing for honeybee, yellow jacket, yellow hornet, white-faced hornet, and wasp. Discussed in detail elsewhere, in areas of the Southern United States, testing for venomous ants including the imported fire ants should be considered. Many individuals experience reduced sensitivity to venom testing in the first few weeks after a systemic sting reaction; therefore testing should be deferred for 4 to 6 weeks, as the potential of a false-negative reaction may be greater within 4-6 weeks of anaphylaxis (Goldberg and Confino-Cohen 1997).

A negative skin test result with a convincing history of sting reaction should be interpreted with caution (Golden et al. 2001; Reisman 2001). If the initial percutaneous and intradermal tests are negative, an in vitro test, measuring sIgE for venoms, such as Immunocap Assay[®], is indicated. A serum basal tryptase level should also be ordered to assess for possible underlying mast cell disease (discussed later) (Georgitis and Reisman 1985; Golden et al. 2017). If both initial skin testing and in vitro testing are negative, then the testing should be repeated in 6–12 weeks (Georgitis and Reisman 1985; Golden et al. 2017).

As previously noted, there is some antigen cross-reactivity between the various Hymenoptera species. This could be secondary to crossreacting carbohydrate determinants, which not thought to be clinically relevant (Hoffman 1993; King et al. 1985; Reisman et al. 1982). Neither the size of the skin test reaction nor the measured level of venom-specific IgE antibodies is reliable indicators of future sting reaction severity (Hoffman 1993; Golden et al. 2001; Reisman 2001).

Periodically, falsely positive and falsely negative reactions may occur. False-positive reactions are usually caused by the inherent, nonspecific irritant effect of the venom, usually at concentrations above 1 mcg/ml (Hoffman 1993). The combination of venom skin testing and complementary in vitro testing detects 98% of sensitized individuals (Hamilton 2001, 2004). However, occasionally, an individual with a convincing history of a systemic Hymenoptera sting reaction has both negative skin and in vitro testing (Golden et al. 2001, 2003; Reisman 2001). Again, a negative venom test should be interpreted with caution. Occurrences of anaphylaxis have been reported in individuals who tested negative to both venom skin testing and in vitro methods (Hamilton 2004; Golden et al. 2001; Reisman 2001). In such cases, mast cell disorders, such as occult or indolent mastocytosis or mast cell activation syndromes, should be considered. A basal serum tryptase level is recommended in subjects with negative testing and convincing history. The role and utility of serum tryptase in the evaluation of Hymenoptera allergy and occult or indolent mast cell disorders is evolving. A baseline serum tryptase level of >11.4 ng/ml after a fully subsided reaction suggests an underlying mast cell disorder (Bonadonna et al. 2010). Serum tryptase levels of greater than 20 ng/ml would warrant consideration of additional testing, including bone marrow biopsy (González de Olano et al. 2008; Brockow et al. 2008; Rueff et al. 2009; Bonadonna et al. 2010). Individuals with underlying or occult mast cell disorders are at greater risk for anaphylaxis, particularly insect anaphylaxis (Rueff et al. 2009; Bonadonna et al. 2009, 2010). The protective level of VIT may be lower than that in the general population, and the safety of VIT may also be lower in individuals with mast cell disorders (Oude Elberink et al. 1997; Niedoszytko et al. 2009). However, VIT is recommended as affected subjects are at greater risk without treatment.

30.5 Treatment

Hymenoptera stings are usually acutely painful and the event is obvious. Local reactions – those that are limited to the area contiguous to the sting site – are treated symptomatically. If a stinger is embedded, it should be removed by flicking it out and not squeezing the attached venom sac. The rate of venom delivery can be very rapid. In honeybees 90% of the venom is delivered in 20 s, and by 1 min nearly the entire venom sac has been emptied suggesting that the removal of the venom sac must occur within seconds to reduce the potential of anaphylaxis. Otherwise, icing the affected area, using age-appropriate analgesia and oral antihistamines, is the mainstay of treatment. Although considerable pain, erythema, and swelling may exist, even in the case of large local reactions, secondary infection is rare (Schumacher et al. 1994a).

Anaphylaxis due to insect venom is managed the same as anaphylaxis caused by any other allergen (Kosnik and Korosec 2011). Initial treatment of choice is an intramuscular injection of epinephrine, preferably into the anterior, upper, and outer aspect of the thigh. Other medications, such as oral or intravenous corticosteroids and/or H-1 and H-2 histamine receptor blockers, are secondary medications that do not substitute for epinephrine. These are secondary therapies and should be administered only after epinephrine. This is regardless of the patients' age, health status, or comorbid medical conditions (Golden et al. 2011). The time interval between the onset of anaphylactic symptoms and the first dose of epinephrine is the best indicator of a successful outcome, and delayed use is a risk factor for death. Regrettably, underuse of epinephrine in the outpatient and emergency department settings remains problematic (Simons 2008; Manivannan et al. 2009; Bilò and Bonifazi 2008; Demain et al. 2010).

Once the patient is stabilized and the effects of the initial sting event are addressed, further intervention may be necessary. If the reaction is limited to a local reaction, regardless of how large, the patient should be reassured that the risk of a more severe future reaction is small (5-10%) (Graft et al. 1984; Mauriello et al. 1984). Generally, in cases where the sting event reaction was limited to local signs and symptoms, an epinephrine autoinjector is not warranted. In rare cases, where the patient has significant anxiety about a future sting event, an epinephrine auto-injector may contribute to an improved quality of life. This requires careful consideration and should be evaluated on a case-by-case basis. If the reaction included more generalized symptoms, such as bronchospasm, gastrointestinal symptoms, hypotension, or laryngeal edema, provision of and detailed training on

the utilization of an epinephrine auto-injector is recommended. The patient and/or family should be able to demonstrate understanding of appropriate utilization. Avoidance is the mainstay of the management of all allergic diseases. This is certainly true of Hymenoptera allergy, regardless whether the reaction was local or systemic. The individual or family should be counseled on the insect-appropriate avoidance strategies and the benefits of following these strategies. If the sting event resulted in systemic signs and symptoms, the appropriate next step is to refer the patient to an allergy specialist for further evaluation, where the insect allergy will be evaluated and VIT considered (Golden et al. 2011).

VIT should be considered and offered to any patient with a history of a systemic allergic reaction to a Hymenoptera sting and evidence by skin test or in vitro methods of venom-specific IgE antibodies. VIT can provide an up to 98% protection against future sting events. VIT consists of gradually increasing doses of venom, usually beginning at 0.1 to 1.0 mcg/ml. Using current guidelines, the venom for winged Hymenoptera is given subcutaneously until a total dose of 100 mcg is achieved for each of the venoms being treated (Bonifazi et al. 2005; Reisman and Livingston 1992; Golden et al. 1981). The usual venom exposure from most Hymenoptera stings is 20-50 mcg; therefore, a treatment dose of 100 mcg for each venom would represent a protective dose approximating two to five stings (Schumacher et al. 1994b). This maintenance dose was based upon published protocols and is the manufacturers' recommended dosing per the FDA-approved package inserts. A maintenance VIT dose of 100 mcg provides a protection from anaphylaxis in up to 98%, whereas a maintenance VIT dose of 50 mcg/ml, recommended by a single investigator, can provide protection in approximately 80–90% of stings (Bonifazi et al. 2005; Graft et al. 1998). In few cases, the patient experiences local and/or systemic reactions during treatment, resulting in difficulty achieving a full maintenance dose of 100 mcg. In such cases, a maintenance dose of 50 mcg, though suboptimal, may provide adequate protection. In most cases, local reactions should not prevent the achievement

of a full 100 mcg dose, and every effort should be made to achieve this dose. There have been no long-term safety or toxicity issues associated with VIT, including in young children and pregnancy.

The physician should monitor VIT patients at regular intervals of 6 to 12 months. During treatment with VIT, between 3% and 12% of patients will experience a systemic reaction, mostly during the early build-up phase (Golden et al. 2011). These reactions are usually mild. Honeybeeallergic patents and those patients with elevated baseline serum tryptase seem to be at a somewhat higher risk of a systemic reaction during VIT. In addition, patients on beta-blockers or ACE inhibitors have a somewhat higher risk (Rueff et al. 2009). Local reactions to VIT present an important, frequent but generally less serious problem than systemic reaction during VIT. Approximately one-third of venom-allergic patients on VIT will experience local reactions during treatment. Although troublesome to the patient, these local reactions for VIT do not predict an increased risk for future, systemic reactions to VIT. These reactions can be uncomfortable, and as a result, the physician may make adjustments in dosing. It is important to recognize that these adjustments in VIT are primarily made for comfort, not for safety.

30.6 Large Local Reactions and VIT

Large local reactions to Hymenoptera stings are often caused by an IgE-mediated late-phase response. These reactions are not considered lifethreatening and are associated with no more than a 5-10% risk of a future sting, systemic allergic reaction. Venom allergy testing is generally not indicated (Bilò and Bonifazi 2008). However, there are data to suggest that in some patients where the reactions are debilitating, or progressively worsening, VIT may be a consideration to reduce the severity of the local reactions (Demain et al. 2010). An example would be severe facial swelling in a mailman following wasp stings. So, in special circumstances, venom testing and VIT are indicated in patients with large local sting reactions.

30.7 Duration of VIT

The duration of VIT for venom-allergic patients is unclear (Bonifazi et al. 2005; Graft et al. 1998; Golden et al. 1996, 2000; Muller et al. 1991). The majority of patients are sufficiently protected after completing a 5-year treatment plan; however some authors suggest that lifetime therapy may be warranted. Some experts suggest that repeat venom skin testing can be helpful for determining who may discontinue VIT (Forester et al. 2007; Muller et al. 1992). Although this information may be helpful, the loss of skin test reactivity is not a guarantee of an absence of risk to venominduced anaphylaxis. Lifelong VIT should be considered in individuals who have experienced a previous life-threatening event; have honeybee allergy, mast cell disease, and comorbid conditions; or have had a systemic reaction during VIT (Georgitis and Reisman 1985; Golden et al. 1998, 2017; Lerch and Muller 1998). Those patients requiring a higher than usual venom dose, having severe anxiety concerning future stings, or having high risk for recurrent stings should also consider lifelong VIT.

30.8 Recent Developments in Insect Allergy

Advances in our understanding of the role of clonal mast cell disorders, basophil biology, and utility of serum tryptase have enhanced the evaluation of Hymenoptera sting allergy. Many of these advances will contribute to improved diagnosis and management of insect-allergic individuals. For example, the effective management of patients with a compelling history of insectinduced anaphylaxis, yet are skin and blood test negative for venom-specific IgE, has been a challenge. Occult mastocytosis or other mast cell disorders are now recognized as a potential explanation. A multicenter study of predictors of severe anaphylaxis reported elevated serum tryptase is one of the predictors (Bonadonna et al. 2010; Oude Elberink et al. 1997; Alvarez-Twose et al. 2010). Hymenoptera allergy is a frequent finding in individuals with mastocytosis. The effectiveness

of VIT is less in subjects with mastocytosis or clonal mast cell disorders (Rueff et al. 2010). The 2017 insect allergy practice parameter more thoroughly addresses the role for obtaining tryptase levels in the evaluation of insect allergy and supports the role of VIT in patients with clonal mast cell disease (Georgitis and Reisman 1985; Golden et al. 2017). Finally, several recent cases have reported the usefulness of the immunomodulatory effects of omalizumab, a monoclonal antibody specific for IgE antibody, in the management of difficult to treat insect anaphylaxis in subjects with indolent or occult mastocytosis (Galera et al. 2009; Kontou-Fill et al. 2010). Though not an FDA-approved indication, in special circumstances, omalizumab may be a consideration (Georgitis and Reisman 1985; Golden et al. 2017).

In addition to the evolving understanding of clonal mast cell disorders and Hymenoptera allergy, the role of basophils in the diagnosis and management of insect anaphylaxis is also expanding. Although not commonly used in the United States, the basophil activation test may be informative in managing individuals with a history of systemic reactions to insect stings without specific IgE (Kruse et al. 2009; Kosnik and Korosec 2011; Peternelj et al. 2009).

30.9 Conclusion

Insect allergy is one of the three most common triggers of life-threatening anaphylaxis and is by far the most treatable. It is crucial that physicians and the public understand proper diagnosis, treatment, and management of this potentially lifethreatening allergy. While the other two causes, food and medication anaphylaxis, are managed primarily by avoidance, Hymenoptera allergy can be managed prospectively with VIT, which provides up to 98% protection from subsequent sting anaphylaxis. Effective management of the acute event, a thorough history of the sting circumstances, recognition of the likely culprit insect, appropriate venom testing, VIT, and optimal use of auto-injector epinephrine are necessary for ideal outcomes. Acute management includes establishing the presence of a Hymenoptera stingrelated anaphylactic event, followed by appropriate use epinephrine. Occult mast cell disease may be playing an important role in Hymenoptera sting reactions, and a basal tryptase level may be very helpful. However, long-term management does not end with the dispensing of an epinephrine auto-injector but includes appropriate referral, determination of venom-specific IgE, and, if indicated, IT.

References

- Álvarez-Twose I, González de Olano D, et al. Clinical, biological and molecular characteristics of clonal mast cell disorders presenting with mast cell activation symptoms. J Allergy Clin Immunol. 2010;125:1269–78.
- Bilò BM, Bonifazi F. Epidemiology of insect venom anaphylaxis. Curr Opin Allergy Clin Immunol. 2008;8:330–7.
- Bonadonna P, Perbellini O, Passalacqua G, et al. Clonal mast cell disorders in patients with systemic reactions to Hymenoptera stings and increased serum tryptase levels. J Allergy Clin Immunol. 2009;123:680–6.
- Bonadonna P, Zanotti R, Müller U. Mastocytosis and insect venom allergy. Curr Opin Allergy Clin Immunol. 2010;10:347–53.
- Bonifazi F, Jutel M, Bilo BM, Birnbaum J, Muller U, EAACI. Prevention and treatment of Hymenoptea venom allergy: guidelines for clinical practice. Allergy. 2005;60:1459–70.
- Brockow K, Jofer C, Behrendt H, Ring J. Anaphylaxis in patients with mastocytosis: a study on history, clinical features and risk factors in 120 patients. Allergy. 2008;63:226–32.
- De Root H. Allergy to Bumblebee. Curr Opin Allergy Clin Immunol. 2006;6:294–7.
- Demain JG. Papular urticaria and things that bite in the night. Curr Allergy Asthma Rep. 2003;3(4):291.
- Demain JG, Minaei AA, Tracy JM. Anaphylaxis and insect allergy. Curr Opin Allergy Clin Immunol. 2010;10: 318–22.
- Forester JP, Johnson TL, Arora R, Quinn JM. Systemic reaction rates to field stings among imported fire ant sensitive patients receiving >3 years of immunotherapy versus <3 years of immunotherapy. Allergy Asthma Proc. 2007;28:485–8.
- Franken HH, Dubois AE, Minkema HJ, et al. Lack of reproducibility of a single negative sting challenge response in the assessment of anaphylactic risk in patients with suspected yellow jacket hypersensitivity. J Allergy Clin Immunol. 1994;93:431.
- Freeman TM. Clinical practice. Hypersensitivity to hymenoptera stings. N Engl J Med. 2004;351:1978.
- Galera C, Soohun N, Zankar N, et al. Severe anaphylaxis to bee venom immunotherapy: efficacy of pretreatment and concurrent treatment with omalizumab. J Investig Allergol Clin Immunol. 2009;19:225–9.

- Georgitis JW, Reisman RE. Venom skin tests in insectallergic and insect-nonallergic populations. J Allergy Clin Immunol. 1985;76:803.
- Goddard J. Physician's guide to arthropods of medical importance. 4th ed. Boca Raton: CRC Press; 2003. p. 4.
- Goldberg A, Confino-Cohen R. Timing of venom skin tests and IgE determinations after insect sting anaphylaxis. J Allergy Clin Immunol. 1997;100:182.
- Golden DBK, Kagey-Sobotka A, Valentine MD, Lichtenstein LM. Dose dependence of Hymenoptera venom immunotherapy. J Allergy Clin Immunol. 1981;67:370–4.
- Golden DB, Marsh DG, Kagey-Sobotka A, et al. Epidemiology of insect venom sensitivity. JAMA. 1989;262:240.
- Golden DBK, Kwiterovich KA, Kagey-Sobotka A, Valentine MD, Lichtenstein LM. Discontinuing venom immunotherapy: outcome after five years. J Allergy Clin Immunol. 1996;97:579–87.
- Golden DB, Marsh DG, Freidhoff LR, et al. Natural history of Hymenoptera venom sensitivity in adults. J Allergy Clin Immunol. 1997;100:760.
- Golden DBK, Kwiterovich KA, Addison BA, Kagey-Sobotka A, Lichtenstein LM. Discontinuing venom immunotherapy: extended observations. J Allergy Clin Immunol. 1998;101:298–305.
- Golden DBK, Kagey-Sobotka A, Lichtenstein LM. Survey of patients after discontinuing venom immunotherapy. J Allergy Clin Immunol. 2000;105:385–90.
- Golden DB, Kagey-Sobotka A, Norman PS, et al. Insect sting allergy with negative venom skin test responses. J Allergy Clin Immunol. 2001;107:897.
- Golden DB, Tracy JM, Freeman TM, et al. Negative venom skin test results in patients with histories of systemic reaction to a sting. J Allergy Clin Immunol. 2003;112:495.
- Golden DB, Breisch NL, Hamilton RG, et al. Clinical and entomological factors influence the outcome of sting challenge studies. J Allergy Clin Immunol. 2006;117: 670.
- Golden DB, Moffitt JE, Nicklas RA, et al. Stinging insect hypersensitivity: a practice parameter update 2011. J Allergy Clin Immunol. 2011;127:852–4.
- Golden DB, Demain J, Freeman T, Graft D, Tankersley M, Tracy J, et al. Stinging insect hypersensitivity: a practice parameter update 2016. Ann Allergy Asthma Immunol. 2017;118(1):28–54.
- González de Olano D, Alvarez-Twose I, Esteban-López MI, et al. Safety and effectiveness of immunotherapy in patients with indolent systemic mastocytosis presenting with Hymenoptera venom anaphylaxis. J Allergy Clin Immunol. 2008;121:519.
- Graft DF. Insect sting allergy. Med Clin N Am. 2006;90: 211–32.
- Graft DF, Schuberth KC, Kagey-Sobotka A, et al. A prospective study of the natural history of large local reactions after Hymenoptera stings in children. J Pediatr. 1984;104:664.
- Graft DF, Golden D, Reisman R, Valentine M, Yunginger J. The discontinuation of Hymenoptera venom

immunotherapy. Report from the Committee on Insects. J Allergy Clin Immunol. 1998;101:573–5.

- Gurlanick MW, Benton AW. Entomological aspects of insect sting allergy. In: Levine MI, Lockey RF, editors. Monograph on insect allergy. 4th ed. Pittsburgh: Dave Lambert Associates; 2003. p. 11.
- Hamilton RG. Responsibility for quality IgE antibody results rests ultimately with the referring physician. Ann Allergy Asthma Immunol. 2001;86:353.
- Hamilton RG. Diagnostic methods for insect sting allergy. Curr Opin Allergy Clin Immunol. 2004;4:297.
- Hoffman DR. Allergens in Hymenoptera venom. XXV. The amino acid sequence of Antigen 5 molecules. The structural basis of antigenic crossreactivity. J Allergy Clin Immunol. 1993;92:707–16. (III)
- Hoffman DR, Jacobson RS. Allergens in Hymenoptera venom. XII. How much protein in a sting? Ann Allergy. 1984;52:276–8.
- Hoffman DR, El-Choufani SE, Smith MM, et al. Occupational allergy to bumblebee: allergens of *Bombusterrestris*. J Allergy Clin Immunol. 2001; 108:855–60.
- Hunt KJ, Valentine MD, Sobotka AK, Benton AW, Amodio FJ, Lichtenstein LM. A controlled trial of immunotherapy in insect hypersensitivity. N Engl J Med. 1978;299:157–61.
- King TP, Joslyn A, Kochoumian L. Antigenic crossreactivity of venom proteins from hornets, wasps and yellow jackets. J Allergy Clin Immunol. 1985;75: 621–8. (III)
- Kontou-Fill K, Fillis CI, Voulgari C, Panayiotidis PG. Omalizumab monotherapy for bee sting and unprovoked 'anaphylaxis' in a patient with systemic mastocytosis and undetectable specific IgE. Ann All Asthma Immunol. 2010;104:537–9.
- Kosnik M, Korosec P. Importance of basophil activation testing in insect venom allergy. Allergy Asthma Clin Immunol. 2011;5:11.
- Kruse P, Erzen R, Silar M, et al. Basophil responsiveness in patients with insect sting allergies and negative venomspecific immunoglobulin E and skin prick test results. Clinical Exp Allergy. 2009;39:1730–7.
- Lerch E, Muller U. Long-term protection after stopping venom immunotherapy. J Allergy Clin Immunol. 1998;101:606–12.
- Light WC, Reisman RE, Shimizu M, Arbesman CE. Unusual reactions following insect stings. Clinical features and immunologic analysis. J Allergy Clin Immunol. 1977;59:391.
- Manivannan V, Campbell RL, Bellolio MF, et al. Factors associated with repeated use of epinephrine for the treatment of anaphylaxis. Ann Allergy Asthma Immunol. 2009;103:395–400.
- Mauriello PM, Barde SH, Georgitis JW, Reisman RE. Natural history of large local reactions from stinging insects. J Allergy Clin Immunol. 1984;74:494.
- Moffitt JE. Allergic reactions to insect stings and bites. South Med J. 2003;96:1073–9.

- More D, Nugent J, Hagen L, et al. Identification of allergens in the venom of the common stripped scorpion. Ann Allergy Asthma Immunol. 2004;93:493–8.
- Muller U, Berchtold E, Helbling A. Honeybee venom allergy: results of a sting challenge 1 year after stopping venom immunotherapy in 86 patients. J Allergy Clin Immunol. 1991;87:702–9.
- Muller U, Helbling A, Berchtold E. Immunotherapy with honeybee venom and yellow jacket venom is different regarding efficacy and safety. J Allergy Clin Immunol. 1992:89:529–35.
- Niedoszytko M, de Monchy J, van Doormaal JJ, et al. Mastocytosis and insect venom allergy: diagnosis, safety and efficacy of venom immunotherapy. Allergy. 2009;64:1237–45.
- Oude Elberink JNK, deMonchy JGR, Kors JW, et al. Fatal anaphylaxis after a yellow jacket sting, despite venom immunotherapy, in two patients with mastocytosis. J Allergy Clin Immunol. 1997;100:11–5.
- Parker JL, Santrach PJ, Dahlberg MJ, Yunginger JW. Evaluation of Hymenoptera-sting sensitivity with deliberate sting challenges: inadequacy of present diagnostic methods. J Allergy Clin Immunol. 1982;69:200.
- Peternelj A, Silar M, Bajrovic N, et al. Diagnostic value of the basophil activation test in evaluating Hymenoptera venom sensitization. Wien Klin Wochenschr. 2009;121: 344–8.
- Regularly updated maps of the fire ant range and agriculture quarantine areas within the United States. www. aphis.usda.gov/plant_health/plant_pest_info/fireants/ downloads/fireant.pdf. Accessed 14 Mar 2018.
- Reisman RE. Insect sting allergy: the dilemma of the negative skin test reactor. J Allergy Clin Immunol. 2001;107:781.
- Reisman RE. Unusual reactions to insect stings. Curr Opin Allergy Clin Immunol. 2005;5:355.
- Reisman RE, Livingston A. Venom immunotherapy: 10 years of experience with administration of single venoms and 50 micrograms maintenance doses. J Allergy Clin Immunol. 1992;89:1189–95.

- Reisman RE, Mueller U, Wypych J, Eliott W, Arbesman CE. Comparison of the allergenicity and antigenicity of yellow jacket and hornet venoms. J Allergy Clin Immunol. 1982;69:268–74. (III)
- Rueff F, Przybilla B, Bilo MB, Muller U, Scheipl F, Aberer W, et al. Predictors of severe systemic anaphylactic reactions in patients with Hymenoptera venom allergy: importance of baseline serum tryptase – a study of the EAACI Interest Group on Insect Venom Hypersensitivity. J Allergy Clin Immunol. 2009;124:1047–54.
- Rueff F, Przybilla B, Bilo MB, Muller U, et al. Predictors of side effects during build-up phase of venom immunotherapy for Hymenoptera venom allergy: the importance of baseline serum tryptase. J Allergy Clin Immunol. 2010;126:105–11.
- Sampson HA, Mendelson L, Rosen JP. Fatal and near-fatal anaphylactic reactions to food in children and adolescents. N Engl J Med. 1992;327:380–4.
- Schumacher MJ, Tveten MS, Egan NB. Rate and quantity of venom from honeybee stings. J Allergy Clin Immunol. 1994a;93:832–5.
- Schumacher MJ, Tveten MS, Egen NB. Rate and quantity of delivery of venom from honeybee stings. J Allergy Clin Immunol. 1994b;93:831–5.
- Schwartz HJ, Yunginger JW, Schwartz LB. Is unrecognized anaphylaxis a cause of sudden unexpected death? Clin Exp Allergy. 1995;25:866–70.
- Severino MG, Campi P, Macchia D, Manfredi M, et al. European Polistes venom allergy. Allergy. 2006;61: 860–3.
- Simons FE. Anaphylaxis. J Allergy Clin Immunol. 2008;121(2 Suppl):S402–7.
- Simons PER, Sampson HA. Anaphylaxis epidemic: fact or fiction? J Allergy Clin Immunol. 2008;122:1166–8.
- Simons FER, Frew AJ, Ansotegui IL, Bochner BS, Finkelman F, Golden DBK, et al. Risk assessment in anaphylaxis: current and future approaches. J Allergy Clin Immunol. 2007;120:S2–24.
- Valentine M. Insect venom allergy: diagnosis and treatment. J Allergy Clin Immunol. 1984;73:299–304.