Chapter 6 An Update on Bioactive Natural Products from Endophytic Fungi of Medicinal Plants



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Abstract Pathogens are developing resistance against the current regime of drugs, which urge the need of novel drugs. This has led scientists to explore natural sources that are safe as well as potent. Microbes have been explored for over decades for natural products and have been the vast reservoir of secondary metabolites of drugs

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potential. Because of their huge diversity and particular habituation, they can act as good resource to obtain bioactive secondary metabolites. Endophytes have been exploited to get drug-like molecules that have antibacterial, antifungal, anticancer, antioxidant, antidiabetic, antileishmaniasis, and antiviral activities. Here we review endophytic fungi as storehouse of naturally occurring bioactive secondary metabolites.

6.1 Introduction

De Bary in 1866 coined the term "endophyte," and according to him, "Endophytes are the microorganisms, which reside inside the plant tissues and are significantly different from those found on the plant surface." Since then a large number of definition were given by various researchers; each definition collectively says "Endophytes are the microbes that reside intra or intercellular in the tissues of host plants and there they do not give rise to any apparent deleterious effects." The endophytes have been found to be an excellent producer of bioactive metabolites of pharmaceutical importance (Bascom-Slack et al. 2009; Pimentel et al. 2011; Brader et al. 2014). Today most of the world population depends on natural products for treating various diseases because of its least side effects. Endophytes are propitious source of bioactive compounds with pharmaceutical importance, and many secondary metabolites of microbial origin are under clinical trials. The natural products were either derived from plants or microorganisms, and among microorganisms, endophytes have the major contribution. Since both plants and microbes have largely been explored for the isolation of compounds, bioprospection of microbes from unique niches would serve to gain the source of new metabolites to combat the upcoming challenges like resistance against the available drugs in the market (Kudo et al. 1998; Zuck 1998; Qin et al. 2010).

Due to extensive diversity of plants, it is expected that microbes residing inside them will act as an excellent source of novel compounds of various bioactivities. An endophytic fungus associates specifically with their host in a facultative or obligate or symbiotic relationship and mediates various signals of mutual relationship (Nair and Padmavathy 2014). These signals depend on the environment which the microbe faces inside the host plant, resulting in the specificity with their host (Dudeja et al. 2012). The endophytes face biotic and abiotic stresses inside the host plant which they overcome through interaction with each other and also with their host plant, resulting to produce diverse secondary metabolites (Liu et al. 2010; Shimizu et al. 2001) (Fig. 6.1). The majority of the bioactive compounds produced by endophytic fungi belong to the different structures and classes, such as quinolones, flavonoids, steroids, phenols, xanthones, terpenoids, etc. (Tan and Zou 2001) exhibiting diverse bioactive potentials including antibacterial, antifungal, antioxidants, antiviral, anticancer, immunosuppressive, etc. (Gunatilaka 2006; Qin et al. 2008). Being a source of structurally diverse and novel metabolites, the endophytic fungi would be



Fig. 6.1 Relationship of endophytic fungi with their host, enhancing growth, promoting resistance, and regulating the production and accumulation of secondary metabolites

promising to obtain the novel compounds if unique niches may explored. A schematic representation to exploit the endophytic fungi for bioactive metabolites has been given in Fig. 6.2.

6.2 Endophytic Fungi as a Storehouse of Bioactive Compounds

Endophytic fungi remain rich sources of many important therapeutic agents. Since decade endophytic fungi have been extensively explored for bioactive natural products. This appears to be emerging area of research which could give a boost to the drug discovery programs. Here we have reviewed and compiled the bioactive compounds reported from the endophytic fungi since the last 5 years. The second-ary metabolites of antimicrobial, anticancer, antioxidant, antidiabetic, antileishmaniasis, and antiviral potential obtained from endophytic fungi of medicinal plants are listed and discussed below in the following sections.



Fig. 6.2 Production of bioactive metabolites in endophytic fungi

6.2.1 Antibacterial Compounds from Endophytic Fungi

Antibacterial compounds can be defined as the molecules which have potential to kill or inhibit the growth of other microorganisms. Since two decades there has been an extensive hike of penicillin-resistant *Enterococcus faecium*, *Staphylococcus pneumonia*, and methicillin-resistant *Staphylococcus aureus* (MRSA). In view of resistance and inadequate number of drugs against the pathogens, there is a need for the hunt of new and diverse drugs that can confront with the upcoming challenges of variety of infections although a huge number of diverse and novel compounds with varied bioactivities have been reported from endophytic fungi but still many more to be discovered. The reviewed literature clearly indicates that the secondary metabolites produced by endophytic fungi belonging to diverse class of compounds such as flavonoids, xanthones, terpenoids, alkaloids, phenols, steroids, and quinines have antibacterial activities (Gao et al. 2018); the recent findings have been enlisted in Table 6.1 and represented in Fig. 6.3.

An endophytic fungus, *Colletotrichum gloeosporioides* of *Lannea coromandelica*, displayed considerable antimicrobial activity. One of the metabolite showed highest zone of inhibition with 25 mm zone formation against *Staphylococcus aureus*. The main compounds responsible for antimicrobial activity were 9-octadecenamide (1), hexadecanamide (2), diethyl phthalate (3), 2-methyl-3methyl-3-hexene (4), and 3-ethyl-2, 4-dimethylpentane (5) (Tayung et al. 2011). In one study, the Di-n-octyl phthalate (6) was reported from *Aspergillus terreus* MP15,

Compound	Endophytic fungi	Host plant	References
9-octadecenamide (1), hexadecanamide (2), diethyl phthalate (3), 2-methyl-3- methyl-3-hexene (4), and 3-ethyl-2,4- dimethyl-pentane (5)	Colletotrichum gloeosporioides	Lannea coromandelica	Tayung et al. (2011)
Di-n-octyl phthalate (6)	Aspergillus terreus	Swietenia macrophylla	Yin et al. (2015)
Ergosterol (7) and cerevisterol (8)	2L-5	Ocimum basilicum	Haque et al. (2005)
Fatty acid and phenolic compound	Diaporthe phaseolorum	Tectona grandis L.f	Kumala et al. (2015)
Tannins, phenols, and amino acid	Stemphylium radicinum	Calyptus	Hussain et al. (2014)
Colletotrichones A (9), colletotrichones B (10), and colletotrichones C (11)	<i>Colletotrichum</i> sp. BS4	Buxus sinica	Wang et al. (2016)
Equisetin (12)	Fusarium sp.	Opuntia dillenii	Ratnaweera et al. (2015)
Asperamide A (13) and cerebroside C (14)	Aspergillus niger	Ipomoea batatas	Shaaban et al. (2013)
Fusarubin methyl ether (15)	Cladosporium sp.	Rauwolfia serpentina	Khan et al. (2016)
Viriditoxin (16)	Paecilomyces variotii	Laguncularia racemosa	Silva et al. (2013)
Purpureone (17)	Purpureocillium lilacinum	Rauvolfia macrophylla	Lenta et al. (2016)
Alternariol 9-methyl ether (18)	Alternaria sp.	Salvia miltiorrhiza	Lou et al. (2016)
18-ethoxycytochalasin J (19), cytochalasins H (20), and cytochalasins J (21)	Phomopsis sp.	Garcinia kola	Jouda et al. (2016)
Phomosine A (22) and Phomosine C (23)	<i>Diaporthe</i> sp.	Siparuna gesnerioides	Sousa et al. (2016)
Terrein (24)	Aspergillus terreus	Achyranthes aspera	Goutam et al. (2017)
(22 <i>E</i> ,24 <i>R</i>)-stigmasta-5,7,22-trien-3-β-ol (25)	Aspergillus terreus	Carthamus lanatus	Elkhayat et al. (2016)
Beauvericin (26)	Epicoccum nigrum	Entada abyssinica	Dzoyem et al. (2017)
Alternariol (27) and 3,7-dihydroxy-9- methoxy-2-methyl-6Hbenzo[c]chromen- 6-one (28)	Alternaria alternata	Grewia asiatica	Deshidi et al. (2017)

 Table 6.1
 Antibacterial compounds derived from endophytic fungi

a fungal endophyte of *Swietenia macrophylla*. The Di-n-octyl phthalate was found to be used as food preservative along with antibacterial potential against foodborne pathogenic bacteria that are *Staphylococcus aureus*, *Bacillus cereus*, *Bacillus spizizenii*, and *Bacillus subtilis* (Yin et al. 2015). Antibacterial compounds can also be used as a food preservative, which acts to control the food spoilage that is the main concern for the problems created in the food chain (Liu and Xu 2008).



Fig. 6.3 Antibacterial compounds (1-28) from endophytic fungi

Similarly, *Ocimum basilicum* was explored, resulting to obtain 23 endophytic fungi from healthy leaf, stem, and roots. One of the endophytic fungi among them has been reported to produce two steroidal compounds, ergosterol (7) and cerevisterol (8). The compounds were found to have antibacterial potential against *Bacillus cereus* and *Staphylococcus aureus* (Haque et al. 2005). Jati tree (*Tectona grandis Lf*) is an herbal plant that acts as host of an endophytic fungus *Diaporthe phaseolorum*. Four fractions were obtained from this fungus by using column chromatography;



consequently the fraction IV was found to have potent antibacterial activity with a zone of inhibition at 11.10 mm when tested against *Staphylococcus aureus*. Further, a fatty acid and a phenolic compound have been reported via GCMS in the bioactive fraction (Kumala et al. 2015).

Stemphylium radicinum an endophytic fungus was reported from the *Calyptus* roots collected from Misan city of Iraq. The extract of the fungus was found to have tannins, phenols, and amino acid as active secondary metabolites and shows potent activity against *Escherichia coli*, *Staphylococcus aureus*, *Proteus vulgaris*, *Klebsiella pneumoniae*, and *Streptococcus pyogenes*. The antibacterial activity was performed using the disc diffusion technique, and the zones of inhibition were reported in the range of 22.5–35.5 mm with MIC of 25.0–100 µg/mL (Hussain et al. 2014).

Besides a reported compound chermesinone B, three novel, colletotrichones A-C (9–11) were isolated from the fungus *Colletotrichum* sp. BS4. The fungus was isolated from the leaves of traditional medicinal plant *Buxus sinica* of China. The compounds were evaluated for antibacterial potential tested with *Escherichia coli*, *Bacillus subtilis*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*. Colletotrichone A (9) was having MIC of 0.1 and 1.0 µg/mL against *B. subtilis* and *E.coli*, respectively, whereas the compound 10 and 11 shows activity with MIC value of 5.0 µg/mL against *S. aureus* and *E. coli*, respectively (Wang et al. 2013).

In one study, eight fungi were isolated from *Opuntia dillenii* and reported first time, among them the *Fusarium* sp. and *Aspergillus niger* were found to be the most bioactive. The fractionation of the extract of *Fusarium* sp., followed by bioactivity, led to the isolation of tetramic acid derivative, named equisetin (**12**). Equisetin exhibited MIC value of 8 μ g/mL against *Bacillus subtilis* and 16 μ g/mL against MRSA and *Staphylococcus aureus*. The research article also concluded that the endophytic fungi isolated from rough and combative environment can also act as a source for diverse and novel bioactive compounds (Ratnaweera et al. 2015).

Aspergillus niger isolated from the leaves of *Ipomoea batatas* (sweet potato) was found to produce eight diverse bioactive compounds. The isolated compounds were characterized as ergosta-7,22-dien- 3β , 5α , 6α -triol, aurasperone B, aurasperone C, aurasperone F, asperamide A (13), cerebroside C (14), linoleic acid, and R(-)-glycerol monolinoleate. Compounds 13 and 14 exhibited weak to moderate antibacterial activities (11–17 mm), among them cerebroside C showed higher activity than those of the non-glycosylated asperamide A. Cytotoxic examination of the isolated compounds against the brine shrimp confirmed their activities to range between moderate (28–35%) and weak (9–11%) (Naureen et al. 2015).

Two naphthoquinones, namely, anhydrofusarubin and methyl ether of fusarubin (**15**), were isolated from an endophytic fungus, *Cladosporium* sp., isolated from the leaves of *Rauwolfia serpentina*. Fusarubin methyl ester at a concentration of 40 μ g/ disc exhibited prominent antibacterial activities against *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, and *Bacillus megaterium* with zone of inhibition of 27 mm, 25 mm, 24 mm, and 22 mm, respectively (Khan et al. 2016).

Viriditoxin (16) was isolated using column chromatography from *Paecilomyces variotii*, an endophytic fungus isolated from the healthy leaves of *Laguncularia racemosa*. Viriditoxin showed antibacterial activity against *Staphylococcus aureus* and *Enterococcus* sp. with MIC of 0.5 and 2 μ g/mL, respectively (Silva et al. 2013). A new ergochromone derivative purpureone (17) was reported from an endophytic fungus *Purpureocillium lilacinum*, isolated from the roots of *Rauvolfia macrophylla*. Purpureone exhibited antibacterial activity against *Bacillus cereus*, *Escherichia coli*, and *Providencia stuartii* with MIC values below 62.6 μ g/mL (Lenta et al. 2016).

Alternariol 9-methyl ether (**18**) isolated from the extract of *Alternaria* sp., an endophytic fungus from the roots of *Salvia miltiorrhiza Bunge*, inhibited the growth of six bacteria, i.e., *Agrobacterium tumefaciens*, *Bacillus subtilis*, *Pseudomonas lachrymans*, *Ralstonia solanacearum*, *Staphylococcus haemolyticus*, and *Xanthomonas vesicatoria* with MIC values ranging from 25 to 75 µg/mL (Lou et al. 2016).

Three reported cytochalasins that are 18-methoxycytochalasin J (19), cytochalasin H (20), and cytochalasin J (21) along with alternariol were reported from *Phomopsis* sp., an endophytic fungus obtained from the nuts of *Garcinia kola*. The cytochalasins exhibited strong activity against *Shigella flexneri*. The antibiotic ampicillin at concentration of 512 μ g/mL was not able to show any inhibition against *Vibrio cholerae* NB2, *Vibrio cholerae* PC2, and *Shigella flexneri*, but interestingly *Shigella flexneri* was sensitive to the cytochalasins (Jouda et al. 2016).

Six compounds, 4H-1-benzopyra-4-one-2,3-dihydro-5-hydroxy-2,8-dimetyl, 4H-1-benzopyran-4-one-2,3-dihydro-5-hydroxy-8-(hydroxy-lmethyl)-2-methyl, 4H-1-benzopyra-4-one-2,3-dihydro-5-methoxyl-2,8-dimetyl, phomosine A (22), phomosine C (23), and phomosine D, were isolated from endophytic fungus *Diaporthe* sp. F2934 of *Siparuna gesnerioides*. Compound 22 at concentration 4 μ g/mL showed 20% higher inhibition zone diameter (IZD) than vancomycin, standard against *Staphylococcus aureus*. It also exhibited activity against both Gram (+ve) and (-ve) bacteria, with respective IZD of 77–90% and 83–92%. Phomosine C showed an IZD against *S. aureus* (90%), *Micrococcus luteus* (60%), and *Streptococcus oralis* (61%) (Sousa et al. 2016). Terrein (24) exhibited IC₅₀ of 20 μ g/mL against *Enterococcus faecalis* and [>]20 μ g/mL against *Staphylococcus aureus* and *Aeromonas hydrophila* (Goutam et al. 2017).

(22E,24R)-stigmasta-5,7,22-trien-3- β -ol (**25**) reported from the *Aspergillus terreus* an endophytic fungus isolated from *Carthamus lanatus* roots exhibited a potent activity against MRSA and *Cryptococcus neoformans* with respective IC₅₀ values of 2.29 and 10.68 μ M (Elkhayat et al. 2016).

In one study, beauvericin (26) isolated from *Epicoccum nigrum*, an endophytic fungus of *Entada abyssinica*, had significant antibacterial activity against *Bacillus cereus* and *Salmonella typhimurium* with MIC values of 3.12 and 6.25 μ g/mL, respectively (Hewage et al. 2014).

Recently, alternariol (27), alternariol 9-methyl ether, and 3,7-dihydroxy-9methoxy-2-methyl-6Hbenzo[c]chromen-6-one (28) has been reported from an endophytic fungus *Alternaria alternata*, isolated from a well-known folk medicinal plant *Grewia asiatica*. This was the first report for the isolation of *Alternaria alternata* from this plant. Compounds were detected and quantified by a new multimode LC-ESI-MS/MS method using multiple reaction monitoring. Both 27 and 28 exhibited antibacterial activity against *S. aureus* and vancomycin-resistant *Enterococci* (VRE) with MIC values of 32 and 128 µg/mL, respectively, whereas 27 exhibited promising activity against methicillin-resistant *S. aureus* (MRSA)-15187 with MIC value of 8 µg/mL comparable to ciprofloxacin in contrast to 64 µg/mL of 28 (Deshidi et al. 2017).

6.2.2 Antifungal Compounds from Endophytic Fungi

The rate of fungal infection increases during graft transfer, allogeneic transplantation of the bone marrow, and chemotherapy in cancer patients. Still, a very less number of antifungal drugs are there in the market for treating lethal diseases like aspergillosis, candidiasis, meningitis, fungal eye infection, ringworm, valley fever caused by *Cryptococcus gattii*, histoplasmosis, and sporotrichosis. Due to the evolving pathogenicity and resistance among the fungal pathogens, there is an immediate need of diverse and novel compounds to combat.

Isolation of microbial secondary metabolites has been a natural alternative for various therapeutic applications. Secondary metabolites of endophytic fungi have a great potential in various agricultural, industrial, and medical fields. There are many antifungal compounds that have been isolated from endophytic fungi (Fig. 6.4) which are listed in Table 6.2 and discussed in the following text.



²⁻phenylethyl 1*H*indol-3-yl-acetate (45)

Fig. 6.4 Structures of antifungal compounds (29-45) from endophytic fungi

Compound	Endophytic fungi	Host plant	References
Terrein (24)	Aspergillus terreus	Achyranthes aspera	Goutam et al. (2017)
α -viridin (29), β -viridin (30), and adenosyl 9a-D-arabinofuranoside (31)	Trichoderma sp.	Centaurea stoebe	Abdou and Abdelhady (2015)
Asperamide A (32) and cerebroside C (33)	Aspergillus niger	Ipomoea batatas	Shaaban et al. (2013)
Outovirin C (34)	Penicillium raciborskii	Rhododendron tomentosum	Kajula et al. (2016)
(2S,3R,4R)-(E)-2-amino-3,4- dihydroxy-2-(hydroxymethyl)-14- oxoeicos-6,12-dienoic acid (35) and myriocin (36)	<i>Mycosphaerella</i> sp.	Eugenia bimarginata	Pereira et al. (2015)
Trichothecinol-A (37)	Trichothecium sp.	Phyllanthus amarus	Taware et al. (2014)
4-hydroxymellein (38) and 6-hydroxymellein (39), tyrosol (40), cervesterol (8)	Penicillium sp.	Senecio flavus	Elkhayat and Goda (2017)
Trichodermin (41)	Trichoderma brevicompactum	Allium sativum	Shentu et al. (2014)
Griseofulvin (42)	Nigrospora oryzae	Emblica officinalis	Rathod et al. (2014)
Cladosporin (43) and isocladosporin (44)	Cladosporium cladosporioides	-	Wang et al. (2013)
2-phenylethyl 1 <i>H</i> -indol-3-yl-acetate (45)	Colletotrichum gloeosporioides	Michelia champaca	Chapla et al. (2014)

 Table 6.2
 Antifungal compounds derived from endophytic fungi

Secondary metabolites α -viridin (29), β -viridin (30), and adenosyl 9a-Darabinofuranoside (31) were reported from an endophytic fungus *Trichoderma* sp. cultured from the stem of the medicinal plant *Centaurea stoebe*. These fungal metabolites (29–31) were tested for antifungal potential which shows MIC of 7.81, 15.63, and 8.52 µg/mL against *Aspergillus terreus*, respectively, while against *Fusarium oxysporum*, these compounds have MIC of 57.50, 75.00, and 63.30 µg/ mL, respectively (Abdou and Abdelhady 2015).

In one study, asperamide A (**32**) and cerebroside C (**33**) displayed moderate to high activities against *Candida albicans* (13–15 mm) and *Mucor miehi* (16–18 mm) and the microalgae *Chlorella vulgaris* (13–14 mm), *Chlorella sorokiniana* (12–14 mm), and *Scenedesmus subspicatus* (13–15 mm) (Naureen et al. 2015).

Three novel epithiodiketopiperazine compounds outovirin A, outovirin B, and outovirin C (**34**) have been reported from the extracts of endophytic fungi *Penicillium raciborskii*, isolated from *Rhododendron tomentosum*. The essential oil and crude extracts of this plant have a number of properties like anti-insecticidal, antioxidant, and antimicrobial; leaves and flower extracts have traditionally been used in the primitive time for the treatment of various infections. Outovirin A is the first reported epimonothiodiketopiperazine, and outovirin C represents first most reported trisul-

fide gliovirin-like compound. Outovirin C inhibited growth of *Fusarium oxysporum*, *Botrytis cinerea*, *and Verticillium dahliae* at a mild concentration of 0.38 mM (207 μ g/mL), but a more significant inhibition of growth was observed at a higher concentration of 0.76 mM (413 μ g/mL) (Kajula et al. 2016).

Similarly, two eicosanoic acid compounds, (2S,3R,4R)-(E)-2-amino-3,4dihydroxy-2-(hydroxymethyl)-14-oxoeicos-6,12-dienoic acid (**35**) and myriocin (**36**), were isolated using bioassay-guided fractionation of crude extract by semiprep HPLC and liquid-liquid partitioning of an endophytic fungus *Mycosphaerella* sp. UFMGCB 2032 cultured from *Eugenia bimarginata* DC. Both the compounds exhibited strong antifungal activity against *Cryptococcus neoformans* and *Cryptococcus gattii* and showed MIC values in the range of 1.3–2.50 µg/mL and 0.5 µg/mL, respectively (Pereira et al. 2015).

In one study trichothecinol-A (**37**) a sesquiterpene was reported for the very first time. The compound was isolated from an endophytic fungus *Trichothecium* sp. cultured from *Phyllanthus amarus*. Also, it exhibited moderate inhibition of *Cryptococcus albidus* var. *diffuens* (NCIM 3371 and 3372) with MIC values of 36 and 20 µg/mL, respectively, and against *Penicillium expansum* (NCIM 939) with MIC of 36 µg/mL (Taware et al. 2014). Recently, four compounds were reported from an endophytic fungus *Penicillium* sp. and characterized as 4-methylmellein, 4-hydroxymellein (**38**), 6-hydroxymellein (**39**), and tyrosol (**40**). These compounds had shown activity against *Candida albicans, Fusarium oxysporum*, and *Aspergillus flavus* at the concentration of 50 µg per disc, with zones of inhibition in the range of 6–8 mm (Elkhayat and Goda 2017).

Trichodermin (41) was characterized from *Trichoderma brevicompactum*, an endophytic fungus strain 0248, reported from garlic. Bioactivity-guided fractionation was used to isolate this compound and identified by spectral analysis and mass spectrometry. Trichodermin showed potent antifungal activity against *Rhizoctonia solani* and *Botrytis cinerea* with an EC₅₀ of 0.25 µg/mL and 2.02 µg/mL, respectively (Shentu et al. 2014).

Similarly, the compound griseofulvin (42) was reported from *Nigrospora oryzae*, an endophytic fungus isolated from *Emblica officinalis*. Griseofulvin exhibited antifungal activity against *Fusarium oxysporum*, *Trichophyton mentagrophytes*, and *Microsporum canis* (Rathod et al. 2014).

The extract of *Fusarium proliferatum* an endophytic fungus cultured from *Syzygium cordatum* showed 100% cytotoxicity at 100 µg/mL against the brine shrimp *Artemia salina*. Upon fractionation seven biologically active colored compounds were isolated, namely, ergosta-5,7,22-trien-3β-ol, nectriafurone-8-methyl ether, 9-*O*-methyl fusarubin, bostrycoidin, bostrycoidin-9-methyl ether, and 8-hydroxy-5,6-dimethoxy-2-methyl-3-(2-oxo-propyl)-1,4-naphthoquinone. Fraction BS749_AR was having structural similarity to nectriafurone-8-methyl ether and labeled as compound 7. Due to the small amount of fraction, confirmation of the proposed structure and measuring its 2D NMR spectra was not possible. This fraction inhibited *Staphylococcus aureus* with zone diameter of 11 mm at 40 µg/paper disc. It also exhibited antifungal activity against *Rhizoctonia solani*

and *Aphanomyces cochlioides* with zone diameter of 11 and 10 mm, respectively (Dame et al. 2016).

Cladosporin (43), isocladosporin (44), 5'-hydroxyasperentin, and cladosporin-8methyl ether were reported from crude extract of an endophytic fungus Cladosporium cladosporioides using bioassay-guided fractionation, and acetylation of 5'-hydroxyasperentin was used synthesize а compound to novel 5',6-diacetylcladosporin. Compound 43 at 30 µM showed 92.7, 90.1, 95.4, and 79.9% growth inhibition of plant pathogens Colletotrichum acutatum, Colletotrichum fragariae, Colletotrichum gloeosporioides, and Plasmopara viticola, respectively, while 44 at the same concentration exhibited 50.4, 60.2, and 83.0% growth inhibition. This was the first report of 5-hydroxyasperentin and cladosporin-8-methyl ether from Cladosporium cladosporioides (Wang et al. 2013). 2-phenylethyl 1H-indol-3-yl-acetate (45) reported from an endophytic fungus Collectorichum gloeosporioides cultured from Michelia champaca showed propitious antifungal activity against Cladosporium cladosporioides and Cladosporium sphaerospermum as same as that of the positive standard nystatin (Chapla et al. 2014).

Terrein (24) was recently isolated and characterized from an endophytic fungus *Aspergillus terreus* JAS-2 cultured from *Achyranthes aspera* traditional medicinal plant. Analysis of NMR data (¹H proton and ¹³C) and Fourier-transform infrared spectroscopy confirmed and characterized the product as 4,5-Dihydroxy-3-(1-propenyl)-2-cyclopenten-1-one (Goutam et al. 2017). Ten microgram/microliter concentration of terrein had shown inhibition of *Bipolaris sorokiniana* (57.14%), *Aspergillus flavus* (52.5%), and *Alternaria alternata* (91.25%) as compared to positive standard; 1 µg/µL was found sufficient for 100% growth inhibition of *Phytophthora drechsleri*. Terrein had also shown antioxidant potential by DPPH scavenging with IC₅₀ value of 112 µg/mL.

6.2.3 Anticancer Compounds from Endophytic Fungi

Through literature it is evident that the search for anticancer natural molecules always remains on priority in bioprospection of fungal endophytes, and there are many new compounds (Fig. 6.5) which were reported in recent 5 years, as listed in Table 6.3.

Taxol is a well-known diterpenoid for its anticancer activity (Wani and Taylor 1971); it was initially reported from the bark of western yew plant, *Taxus brevifolia*. The main function of taxol is to inhibit tubulin depolymerization during cell division. Taxol production from fungal endophytes using large-scale fermentation process is cost-effective and safe (Page et al. 2000). It has been reported from many endophytic fungi of genera *Alternaria*, *Fusarium*, *Monochaetia*, *Pestalotia*, *Pestalotiopsis*, *Pithomyces*, and *Taxomyces* (Strobel et al. 1996), and more than 20 fungal genera were identified for the biosynthesis of anticancer drug paclitaxel and its derivatives (Zhao et al. 2010).

Taxol (46) was also isolated from the endophytic fungus that is *Phomopsis lon*gicolla of Aliyar plant, and approximately 381 μ g/mL amount of it was detected



10-deacetybaccatin (58) Camptothecin (59) Homoharringtonine (60) Pyrrocidine A (61)

Fig. 6.5 Structures of anticancer compounds (46-77) from endophytic fungi

using HPLC. Podophyllotoxin is the precursor for the three clinically approved anticancer drugs, EtoposideTM, EtopophosTM, and TeniposideTM. Occurrence of this compound is very less in nature that cannot meet the requirements for the production of these well-known antineoplastic compounds in the medical industry. The search for the natural production of this compound was started, and this was reported from the endophytic fungus *Alternaria tenuissima* cultured from roots of *Sinopodophyllum*



Fig. 6.5 (continued)

emodi (Wall.). HPLC was used for the isolation and quantification purposes and confirmed by using available authentic standards (Liang et al. 2015). This report concluded that using fermentation, podophyllotoxin can be isolated on large scale to fulfill clinical demands and also confirmed that endophytes isolated from the plant can produce the same compound because of its coevolution with their host due to the uptake of host genomes (Stierle et al. 1993).

Similarly, anhydrofusarubin (47) and methyl ether of fusarubin (15) isolated from the endophytic fungus *Cladosporium* sp. cultured from *Rauwolfia serpentina* leaves showed potent cytotoxicity against human leukemia cells (K-562) with IC₅₀ values of 3.97 µg/mL and 3.58 µg/mL, respectively (Khan et al. 2016).

Compounds	Endophytic fungus	Host plant	References
18-methoxycytochalasin J (19), cytochalasin H (20), cytochalasin J (21), and alternariol (27)	Phomopsis sp.	Garcinia kola	Jouda et al. (2016)
Terrein (24)	Aspergillus terreus	Achyranthes aspera	Goutam et al. (2017)
α -viridin (29) and β -viridin (30)	Trichoderma species	Centaurea stoebe	Abdou and Abdelhady (2015)
Trichothecinol-A (37)	Trichothecium sp.	Phyllanthus amarus	Taware et al. (2014)
4-hydroxymellein (38) and 6-hydroxymellein (39)	Penicillium sp.	Senecio flavus	Elkhayat and Goda 2017
Anhydrofusarubin (47) and fusarubin methyl ether (15)	Cladosporium sp.	Rauwolfia serpentina	Khan et al. (2016)
Cytochalasin D (48)	<i>Xylaria</i> sp.	Seaweed	de Felício et al. (2015)
Phomocytochalasin (49), cytochalasin H (19), cytochalasin N (50), RKS-1778 (51), dankasterone B (52), and Cyclo(L-Ile-L-Leu) (53)	Phomopsis theicola	Litsea hypophaea	Hsiao et al. (2016)
Vinblastine and vincristine (54)	<i>Eutypella</i> sp.	Catharanthus roseus	Hani and Eman (2015)
11-dehydroxy epoxyphomalin A (57)	Peyronellaea coffeae-arabicae	Pritchardia lowreyana	Li et al. (2015b)
10-deacetylbaccatin III (58)	Trichoderma sp.	Taxus wallichiana	Li et al. (2015b)
Camptothecin (59)	Fusarium oxysporum	Nothapodytes foetida	Musavi et al. (2015)
Homoharringtonine (60)	Alternaria tenuissima	Cephalotaxus hainanensis Li	Hu et al. (2016)
Pyrrocidine A (61)	Neonectria ramulariae	<i>Cylindrocarpon</i> sp. and <i>Acremonium zeae</i>	Uesugi et al. (2016)
Eremofortin F (62) and mycoepoxydiene (63)	Sabicea cinerea	Rubiaceae sp.	Mandavid et al. (2015)
GKK1032B (64)	Penicillium citrinum	G. mangostana	Qader et al. (2015)
TMC-264 (65) and PR-toxin (66)	Penicillium Chermesinum	Heritiera Littoralis	Darsih et al. (2015)
Depsipeptide (PM181110) (67)	Phomopsis glabrae	Pongamia pinnata	Verekar et al. (2014)
3- <i>epi</i> -steperoxide A (68), steperoxide A (69), merulin B (70), and merulin C (71)	Pseudolagarobasidium acaciicola	Bruguiera gymnorrhiza	Wibowo et al. (2014)

 Table 6.3 Anticancer compounds derived from endophytic fungi

(continued)

Compounds	Endophytic fungus	Host plant	References
Versicoumarin A (72)	Aspergillus versicolor	Paris marmorata	Ye et al. (2014)
Naphthoquinone (herbarin) (73)	Dendryphion nanum	Ficus religiosa	Mishra et al. (2013)
Brefeldin A (74)	Penicillium sp.	Panax notoginseng	Xie et al. (2017)
Xylarione A (75) and (-) 5-methylmellein (76)	Xylaria psidii	Aegle marmelos	Arora et al. (2016)
Jammosporin A (77)	Rosellinia sanctae-cruciana	Albizia lebbeck	Sharma et al. (2018)
E2.2	Colletotrichum gloeosporioides	Phaleria macrocarpa	Gasong and Tjandrawinata (2016)
α -Pyrone derivative	Pestalotiopsis microspore	Taxus chinensis	Li et al. (2015a)

Table 6.3 (continued)

In one study an endophytic fungus, *Colletotrichum gloeosporioides* of *Phaleria macrocarpa*, was reported to yield E2.2 compound. This compound is responsible for its anticancer activity toward MDA-MB-231 and MCF-7 human breast adenocarcinoma cell lines. Phloroglucinol was found to better enhance the production of this compound. Production was found high when incubated in PDB medium, pH 5.0, and kept for 15 days in incubation (Gasong and Tjandrawinata 2016).

A well-known anticancer and antibiotic, cytochalasin D (48), was isolated from Xylaria sp. This was the first report for the isolation of cytochalasin from marine seaweed endophyte (de Felício et al. 2015). Recently, three reported cytochalasins, 18-methoxycytochalasin J (19), cytochalasin H (20), and cytochalasin J (21), along with one more compound named alternariol (27) characterized from an endophytic fungus Phomopsis sp. showed promising cytotoxic activity against human cancer cells with IC₅₀ in the range of 3.66–35.69 µg/mL and without any toxicity against normal cell lines (Jouda et al. 2016). A new cytochalasin along with five reported cytochalasins has been characterized from the solid-state fermentation of Phomopsis theicola BCRC 09F0213, an endophytic fungus isolated from the leaves of Litsea hypophaea Hayata. In one study, a new cytochalasin named as Phomocytochalasin (49) and five others were reported as cytochalasin H (20), cytochalasin N (50), RKS-1778 (51), dankasterone B (52), and cyclo(L-Ile-L-Leu) (53). Among the isolated compounds, cytochalasin N showed nitric oxide inhibitory activity with IC_{50} values of 77.8 μ M, whereas the other cytochalasins have shown IC_{50} values >100 μ M. All the isolates were also tested for interleukin-6 (IL-6) inhibitory activity and showed IC₅₀ values of >100 μ M. The cytochalasin H showed progesterone receptor antagonism with IC₅₀ values of 1.42 µM compared with positive standard RU486 with IC_{50} of 0.063 nM (Hsiao et al. 2016).

Catharanthus roseus is a well-known herbal plant for the synthesis of two valuable anticancer compounds, vincristine and vinblastine. Recently, the production of vincristine (**54**) was reported from an endophytic fungus *Eutypella* sp. CrP14 isolated from the stem of *Catharanthus roseus*. The presence of vincristine was confirmed using chromatographic and spectroscopic analysis. Vincristine showed potent cytotoxic activity in the MTT assay against human squamous carcinoma cells – A431. It was also able to induce apoptosis and causes loss of mitochondrial membrane potential, DNA fragmentation, and generation of reactive oxygen species (Kuriakose et al. 2016).

In one study methyl 9-dihydro-8-trihydroxy-9-oxo-Hxanthene-1-carboxylate (55) and (E)-methyl 2-hydroxy-6, 6-dimethyl hept-3-enoate (56) were reported from the *Chaetomium globosum*, which suppress the growth of Michigan Cancer Foundation-7 (MCF-7) breast cancer cell line and human liver carcinoma cancer cell line (HepG-2) (Hani and Eman 2015).

The fungus *Peyronellaea coffeae-arabicae* isolated from *Pritchardia lowreyana* was explored, from which peyronellins A–C and 11-dehydroxy epoxyphomalin A (**57**) were obtained. The compound **57** was found to inhibit STAT3 at 5 μ m along with potent anticancer activity (IC₅₀ 0.5 μ M) against ovarian cancer cell lines (OVCAR3) (Li et al. 2016).

Taxol is produced in a semisynthetic method using two precursors baccatin III and 10-deacetylbaccatin III; vast literature is available where these two precursors are produced along with taxol. Endophytic fungi add special references in the production of 10-deacetylbaccatin III (**58**) by *Trichoderma* sp. isolated from *Taxus wallichiana* (Li et al. 2015b).

Camptothecin (CPT) is a topoisomerase I-DNA inhibitor and was first reported from *Camptotheca acuminata* Decne (Nyssaceae), a deciduous tree of South China, which has attained inordinate attention due to its potential antitumor activities in experimental studies. In the last decade, camptothecin (CPT) (**59**) along with its analogues such as 9-methoxycamptothecin (MCPT) and 10-hydroxycamptothecin (HCPT) was found to be produced by many endophytic fungi isolated from plant *Camptotheca acuminata* and many plants belonging to the family *Icacinaceae* (Musavi et al. 2015). There are a number of problems regarding production of these compounds by fermentation, as in many endophytic fungal strains like *Fusarium solani* (Kusari et al. 2011) and *Aspergillus* sp. (Pu et al. 2013); after subsequent subculturing the loss of the biosynthetic ability was observed. Recently, Venugopalan and Srivastava (2015) highlighted the increased production of CPT by endophytic fungi *F. solani* isolated from *Camptotheca acuminata*.

α-Viridin (29), β-viridin (30), and adenosyl 9a-D-arabinofuranoside (31) were reported from an endophytic *Trichoderma* sp. cultured from the stems of the medicinal plant *Centaurea stoebe*. 29–31 shows promising cytotoxic and antitumor effects on different cancer cell lines such as HUVEC, K-562, and HeLa. β-Viridin shows GI₅₀ value of 0.5 and 0.25 µg/mL against HUVEC and K-562 cell lines, respectively, whereas α-viridin shows GI₅₀ of 0.02 and 0.01 µg/mL, respectively. α-Viridin and β-viridin show potent cytotoxicity with CC₅₀ (HeLa) of 29.09 and 11.43 µg/mL, respectively. A novel α-pyrone derivative, together with four reported metabolites, was isolated from *Pestalotiopsis microspora*, an endophytic fungus cultured from the stem of *Taxus chinensis* on the solid-state media. Both antimicrobial and anticancer activities were evaluated, but this compound did not have any significant activity (Li et al. 2015b).

Homoharringtonine (HHT) (**60**) is a natural alkaloid obtained from the bark of *Cephalotaxus hainanensis* and is widely used for treating human chronic myeloid leukemia. Due to slow growing and scarcity of the plant, there is a need for some other alternative that can fulfill the rising requirement of the drug in the market. An endophytic fungus *Alternaria tenuissima* isolated from this plant was found to produce the compound homoharringtonine; this supports the theory that endophytes during its coevolution with plants had gained the ability to synthesize the same product as host plant. The extract of endophytic fungi exhibited strong antiproliferative activity against K562, NB4, and HL-60 cancer cell lines with the IC₅₀ values 67.25 ± 4.26 , 65.02 ± 4.75 , and $99.23 \pm 4.26 \mu g/mL$, respectively. Activity may be due to additional compounds in the crude extract that shows synergetic effect. Screening of HHT production was carried out using HPLC analysis and comparing the mass with that of standard (Hu et al. 2016).

Pyrrocidines A (61) is a natural metabolite of alkaloid class and reported from endophytic fungi (*Cylindrocarpon* sp. and *Acremonium zeae*), and it is well known for its antimicrobial activity. This compound is also capable of inducing shrinkage of nuclear DNA and its breakage into small fragments and activation of caspase activity in HL60 cancer cell lines. Its action of inducing apoptosis is different from other clinically approved drugs in the market, and it is due to the presence of unique α,β -unsaturated carbonyl moiety in the molecule (Uesugi et al. 2016).

In one study four metabolites, altiloxin A, enamidin, eremofortin F (62), and mycoepoxydiene (63), were reported from the *Sabicea cinerea*, an endophytic fungus closely related to *Diaporthe pseudomangiferae* obtained from the leaves of a plant of Rubiaceae family. Mycoepoxydiene and altiloxin A are well-known metabolites, whereas enamidin and eremofortin F were reported first time in the literature. Mycoepoxydiene showed promising cytotoxic activity with IC₅₀ values of 7.5, 17.7, and 15.8 μ M against KB, MDA-MB-435, and MRC5 cancer cell lines, respectively. Altiloxin A and enamidin showed no effect (IC₅₀ > 30 μ M) on all tested cell lines, and eremofortin F was cytotoxic on KB and MRC5 cells (IC₅₀ = 13.9 and 12.2 μ M, respectively) (Mandavid et al. 2015).

An endophytic fungus *Penicillium citrinum* isolated from the pericarp surface of *Garcinia mangostana* was reported to produce GKK1032B (**64**), citrinin, and ergosterol. Citrinin is a nephrotoxin mycotoxin that is repeatedly synthesized by fungus *P. citrinum* (Malmstrøm et al. 2000). GKK1032B an antitumor antibiotic was first reported from unidentified species of *Penicillium* in 2001 along with two more metabolites GKK1032A1 and GKK1032A2 (Koizumi et al. 2001). Pastre et al. in 2007 reported reisolation of the compound from an endophytic fungi *Penicillium* sp. of *Melia azedarach* and *Murraya paniculata*. This was the third time to report isolation of GKK1032B from *Penicillium citrinum*. Since this class of metabolites has not been isolated from any other species of *Penicillium*, it was concluded that the

two unidentified *Penicillium* strains in the previous papers can be *P. citrinum* (Qader et al. 2015).

Similarly, new fungal metabolites, penicilliumolide A, penicilliumolide B, penicilliumolide C, and penicilliumolide D, a derivative of PR-toxin, along with three reported compounds, TMC-264 (65), PR-toxin (66), and a sesquiterpene, were isolated from *Penicillium chermesinum*, an endophytic fungus of a mangrove tree, *Heritiera littoralis*. The TMC-264 and PR-toxin showed effective as well as selective cytotoxicity against some cancer cell lines. TMC-264 also showed similar cytotoxic activity as of standard doxorubicin against T47D and MDA-MB231 cancer cell lines, and its cytotoxicity against HepG2 cell line was higher than etoposide. Derivatives of TMC-264 was probably because of the presence of β -chlorosubstituted α , β -unsaturated ketone in its structure (Darsih et al. 2015).

A new depsipeptide (PM181110) (67) was isolated from *Phomopsis glabrae*, an endophytic fungus cultured from the leaves of *Pongamia pinnata*. The compound was tested for its cytotoxicity against a panel of 40 human tumor cell lines out of which it showed activity in a concentration-dependent manner against all cell lines of bladder, colon, gastric, head and neck, liver, lung (non-small-cell lung carcinoma), mammary, ovarian, pancreatic, prostate, renal, and uterus cancer, as well as melanoma, pleural mesothelioma, and sarcoma. Pancreatic cancer cell line PAXF 546L (IC₅₀, 0.016 μ M) and the lung cancer cell line LXFA 526 L (IC₅₀, 0.021 μ M) were the most sensitive cell lines. The compound also exhibited ex vivo efficiency toward all human tumor xenografts (mean IC₅₀ 0.245 μ M) (Verekar et al. 2014).

In one study, two new norsesquiterpenes citicoline A and spiroacaciicolide A, a new nor-chamigrane 3-*epi*-steperoxide A (**68**), along with three known compounds steperoxide A (**69**), merulin B (**70**), and merulin C (**71**), were reported from an endophytic fungus *Pseudolagarobasidium acaciicola*, cultured from a mangrove tree *Bruguiera gymnorrhiza*. The compounds **68–71** showed cytotoxic activity toward many cancer cell lines. Among them compounds **70** and **71** displayed promising cytotoxic activity against MOLT-3, HuCCA-1, A549, HepG2, HL-60, MDA-MB-231, T47D, and HeLa cancer cell lines with IC₅₀ in the ranges of 0.68–3.71 and 0.67–5.25 µg/mL, respectively. Compound **70** exhibited weak cytotoxic activity with an IC₅₀ range of 11.94–49.08 µg/mL, whereas compound **71** exhibited the most potent cytotoxic activity against HL-60 cancer cells, with an IC₅₀ value of 0.08 µg/mL, and in addition displayed IC₅₀ value in the range of 0.19–3.75 µg/mL against other tested cell lines (Wibowo et al. 2014).

Versicoumarin A (72) was reported from an endophytic fungus *Aspergillus* versicolor cultured from the rhizomes of Paris marmorata plant. This compound exhibited strong cytotoxicities against A549 and MCF7cancer cells with IC₅₀ values of 3.8 and 4.0 mM, respectively (Ye et al. 2014).

Two naphthoquinones were isolated from *Dendryphionnanum*, an endophytic fungus of *Ficus religiosa*. This was the first report for the isolation of naphthoquinone compounds from this fungus. Naphthoquinone antibiotic herbarin (73) exhibited potent inhibition of TNF- α and IL-6 cytokine production in human mononuclear

cell line (THP-1) induced with LPS with IC₅₀ of 0.60 \pm 0.100 μ M and 0.06 \pm 0.009 μ M, respectively (Mishra et al. 2013).

Recently, Brefeldin A (74) produced from *Penicillium* sp., an endophytic fungus isolated from the healthy root of *Panax notoginseng*, showed strong anticancer activity against 293 (human renal epithelial cell line), HepG2, Huh (human hepatocellular carcinoma cell line), and KB (human oral epidermoid carcinoma cell line) with ID_{50} value of 0.45 ± 0.008 , 0.0024 ± 0.002 , 0.035 ± 0.005 , and $0.06 \pm 0.009 \,\mu$ M, respectively (Xie et al. 2017).

One new compound xylarione A (**75**) along with (-) 5-methylmellein (**76**) has been reported from an endophytic fungus *Xylaria psidii* cultured from the leaves of a well-known traditional medicinal plant, *Aegle marmelos*, also called "Bael" in Hindi. Both the compounds were active against pancreatic cancer cell line (MIA-Pa-Ca-2) and exhibited cytotoxicity with IC₅₀ values of 16.0 and 19.0 μ M, respectively. Compounds were capable of cell arrest at the sub-G1 phase, exhibited different signs of apoptotic mechanism, and also cause loss of MMP in a dosedependent manner when analyzed using flow cytometry. This result concludes that these metabolites can be modified for making its derivatives and for making more potent drugs (Arora et al. 2016).

One new cytochalasin, named jammosporin A (77), and four known compounds 19,20-epoxycytochalasin D, cytochalasin D, 19,20-epoxycytochalasin C, and cytochalasin C, were isolated from the endophytic fungus *Rosellinia sanctae-cruciana*, cultured from the leaves of medicinal plant *Albizia lebbeck*. Compound 77 showed considerable cytotoxic potential against the human leukemia cancer cell line (MOLT-4) with IC₅₀ values of 20.0 μ M. This was the first report for the isolation of this class of secondary metabolites in *Rosellinia sanctae-cruciana* fungus from this plant (Sharma et al. 2018).

As discussed in the earlier section about the compounds 4-hydroxymellein (**38**) and 6-hydroxymellein (**39**) having antifungal activity, both the compounds also exhibited cytotoxic activity toward MCF-7 cancer cell lines with ED_{50} value of 6.1 and 8.3 µg/mL, respectively (Elkhayat and Goda 2017). Similarly, the compound **35** at concentration of 500 nM exhibits 50% cell death in HeLa and B16F10 cells and 25% in MDA-MB-231 cells (Taware et al. 2014).

6.2.4 Antioxidant Compounds from Endophytic Fungi

Antioxidants of natural origin are the compounds that are commonly found in medicinal plants, fruits, and vegetables. However, it is reported that a large number of secondary metabolites from fungal endophytes are having potent antioxidant activity, and some of the recent compounds (Fig. 6.6) are listed in Table 6.4.

Recently, two antioxidant compounds, terreic acid (78) and 6-methylsalicylic acid (79), were reported from *Pestalotiopsis* sp. EST 02, an endophytic fungus of plant *Elaeocarpus sylvestris*. Both the compounds have shown potent DPPH radical



Fig. 6.6 Structures of antioxidant compounds (78-81) from endophytic fungi

 Table 6.4
 Antioxidant compounds derived from endophytic fungi

Compounds	Endophytic fungi	Host plant	References
Terreic acid (78) and 6-methylsalicylic acid (79)	Pestalotiopsis sp.	Elaeocarpus sylvestris	Prihantini and Tachibana (2017)
Anthraquinone quinizarin (80)	Epicoccum nigrum	Entada abyssinica	Dzoyem et al. (2017)
Calbistrin H (81)	<i>Dothideomycete</i> sp.	-	Hewage et al. (2014)

scavenging activity with IC₅₀ value of 0.22 ± 0.02 mmol/L and 3.87 ± 0.27 mmol/L, respectively. The compounds also showed good activities from the reducing power and β -carotene bleaching assays (Prihantini and Tachibana 2017).

The four compounds beauvericin, parahydroxybenzaldehyde, indole-3-carboxylic acid, and quinizarin were isolated from *Epicoccum nigrum*, an endophytic fungus of *Entada abyssinica*. Anthraquinone quinizarin (**80**) exhibited excellent scavenging activity having IC₅₀ values of 10.86 and 11.36 μ g/mL in the ABTS and DPPH assays, respectively (Dzoyem et al. 2017).

Calbistrin H (81) was reported from an endophytic fungus *Dothideomycete* sp. CRI7 on PDB medium using one strain many compounds (OSMAC) approach. It shows radical scavenging potential with an IC₅₀ value of 21.7 μ M (Hewage et al. 2014).

6.2.5 Antidiabetic Compounds from Endophytic Fungi

Diabetes mellitus is a metabolic disease caused by a defect in secretion of insulin and its action. Since diabetes prevalence has been rising, urge to find natural products of antidiabetic potential. The recent findings have been listed in Table 6.5 and represented in Fig. 6.7.

An endophytic fungus *Dendryphion nanum* (Nees) S. Hughes was isolated from the leaves of *Ficus religiosa* from which a naphthoquinone compound herbarin (73)

	Endophytic		
Compound	fungi	Host plant	References
Naphthoquinone (herbarin) (73)	Dendryphion nanum	Ficus religiosa	Mishra et al. (2013)
(<i>S</i>)-(+)-2- <i>cis</i> -4- <i>trans</i> -abscisic acid (82), 7'-hydroxy-abscisic acid (83), and 4-des-hydroxyl altersolanol A (84)	Nigrospora oryzae	Combretum dolichopetalum	Uzor et al. (2017)
Thielavins A (85), J (86), and K (87)	MEXU 27095	Hintonia latiflora	Rivera-Chávez et al. (2013)

Table 6.5 Antidiabetic compounds derived from endophytic fungi





(S)-(+)-2-*cis*-4-*trans*abscisic acid (82)





OH OH



4-des-hydroxyl altersolanol A (84)

Thielavins A (85)

Thielavins J (86) Thielavins K (87)

Fig. 6.7 Structures of antidiabetic compounds (82-87) from endophytic fungi

was isolated. The herbarin was found to induce the glucose uptake in rat skeleton muscle with IC_{50} of $0.80 \pm 0.090 \,\mu$ M in the presence of insulin (Mishra et al. 2013).

A peptide was obtained via semi-prep HPLC from an endophytic fungus *Aspergillus awamori* of *Acacia nilotica*. The peptide was further resolved using SDS-PAGE to determine its molecular weight, which was found to be of 22 kDa size. The peptide resulted in a mixed type of inhibition against both α -amylase and α -glucosidase and showed IC₅₀ values of 3.75 and 5.625 µg/mL, respectively (Singh and Kaur 2016).

Three metabolites (*S*)-(+)-2-*cis*-4-*trans*-abscisic acid (82), 7'-hydroxy-abscisic acid (83), and 4-des-hydroxyl altersolanol A (84) were isolated from *Nigrospora oryzae*, an endophytic fungus of *Combretum dolichopetalum*. The crude extract of the fungus reduced the fasting blood sugar by 30.79% in 3 h; simultaneously the compounds also showed reduction of 46.37% (3 h), 44.96% (9 h), and 43.70% (9 h), respectively. Comparable activity of extract and compounds was found with that of standard glibenclamide (reduction of 48.72% in 9 h) (Uzor et al. 2017).

MEXU 27095, an endophytic fungus, was isolated from the Mexican medicinal plant *Hintonia latiflora*, and the bioassay-guided fractionation of the bioactive organic extract resulted in the separation of three tridepsides, namely, thielavins A, J, and K (**85–87**). Compounds **85–87** were capable of inhibiting α -glucosidase (α -GHY) *Saccharomyces cerevisiae* in a concentration-dependent manner and exhibited IC₅₀ values of 23.8, 15.8, and 22.1 μ M, respectively. Inhibitory action of the compounds was higher than that of the standard acarbose (IC₅₀ = 545 μ M), used as a positive control for the analysis (Rivera-Chávez et al. 2013).

6.2.6 Antileishmaniasis Compounds from Endophytic Fungi

Leishmaniasis is a serious health problem worldwide, mainly in tropical and subtropical areas where the protozoan has developed resistance to current available drugs in the market. In the absence of effective vaccines, still now chemotherapy plays an important role in fighting this disease. Therefore, the search for novel, effective, and safe drugs is essential for the treatment, control, and prevention of leishmaniasis. Natural products derived from various endophytic fungi associated with medicinal plants have shown promise as a potential source for antiprotozoal drugs (Table 6.6) (Fig. 6.8).

Terrenolide S (88), a novel derivative of butenolide, along with six already reported compounds, (22E,24R)-stigmasta-5,7,22-trien-3- β -ol (89), stigmast-4-ene-3-one (90), stigmasta-4,6,8(14),22-tetraen-3-one, terretonin A, terretonin, and buty-rolactone VI, had recently been isolated from *Aspergillus terreus*, an endophytic fungus cultured from the roots of *Carthamus lanatus*. Compounds 88–90 showed antileishmaniasis against *Leishmania donovani* and exhibited IC₅₀ values of 11.24, 15.32, and 27.27 μ M, respectively (Elkhayat et al. 2016).

Compound	Endophytic fungi	Host plant	References
Purpureone (17)	Purpureocillium lilacinum	Rauvolfia macrophylla	Lenta et al. (2016)
Alternariol 9-methyl ether (18)	Alternaria sp.	Salvia miltiorrhiza	Lou et al. (2016)
Terrenolide S (88), $(22E,24R)$ -stigmasta- 5,7,22-trien-3- β -ol (89) and stigmast-4-ene- 3-one (90)	Aspergillus terreus	Carthamus lanatus	Elkhayat et al. (2016)
(6-S)-3-(1,3-dihydroxypropyl)-6-(2- methylpropyl)piperazine-2,5-dione (91) and (6-R)-3-(1,3-dihydroxypropyl)-6-(2- methylpropyl)piperazine-2,5-dione (92)	Trichosporum sp.	Trigonella foenum- graecum	Metwaly et al. (2015)

 Table 6.6
 Antileishmaniasis compounds derived from endophytic fungi



Terrenolide S (88)

(22*E*,24*R*)-stigmasta-5,7,22-trien-3-β-ol **(89)**



(6-R)-3-(1,3-dihydroxy propyl)-6-(2-methylpropyl) piperazine-2,5-dione (91)



(6-S)-3-(1,3-dihydroxy propyl)-6-(2-methylpropyl) piperazine-2,5-dione (92)

Fig. 6.8 Structures of antileishmaniasis compounds (88-92) from endophytic fungi

Two new diketopiperazine alkaloid isomers (6-S)-3-(1,3-dihydroxypropyl)-6-(2methylpropyl)piperazine-2,5-dione (**91**) and (6-R)-3-(1,3-dihydroxypropyl)-6-(2methylpropyl)piperazine-2,5-dione (**92**) were isolated from an endophytic fungus *Trichosporum* sp. cultured from the seeds of *Trigonella foenum-graecum*. Three chiral centers in both these compounds were not identified due to very low in amount. The compounds **91–92** exhibited moderate antileishmanial activities with IC₅₀ values of 96.3 and 82.5 μ g/mL, respectively (Metwaly et al. 2015).

Enniatins (ENs) are a group of antibiotics having six-membered cyclic depsipeptides formed by the combination of three molecules of D- α -hydroxyisovaleric acid and three N-methyl-L-amino acids commonly reported in the literature being produced by *Fusarium* sp. Six enniatins, namely, ENs A, A1, B, B1, B2, and Q, were reported via LCMS from the methanolic extract of *Fusarium tricinctum*, an endophytic fungus cultured from the fruits of *Hordeum sativum* cultivated on rice medium. The methanolic extract of this fungus exhibited mild antibacterial and antileishmanial activity against *L. donovani* ATTC 39930D with an IC₅₀ value of 16.96 µg/mL. Moreover the extract at 100 µg/mL was also capable of inhibiting the activity of thioredoxin reductase enzyme present in *Plasmodium falciparum* by a factor of 95% (Zaher et al. 2015).

The antibacterial compound **17** also displayed excellent antileishmanial activity against *L. donovani* with an IC₅₀ value of 0.63 µg/mL (0.87 µm) and with good selectivity (SI = 49.5) against the L6 cell line (Lenta et al. 2016). Similarly, the compound **18** inhibited spore germination in *Magnaporthe oryzae* with an IC₅₀ value of 87.18 µg/mL. The compound also exhibited antinematodal activity against *Caenorhabditis elegans* and *Bursaphelenchus xylophilus* with an IC₅₀ value of 74.62 µg/mL and 98.17 µg/mL, respectively (Lou et al. 2016).

6.2.7 Antiviral Compounds from Endophytic Fungi

These days the attack of various viruses causing dreadful effects on human lives is very common. In view of evolving new strains of dreadful viruses, there is a requirement of the search for cheaper, safer, and more robust drug alternative from natural sources like endophytic fungi. Few antiviral compounds have been reported from various endophytic fungi (Fig. 6.9), as listed in Table 6.7.

Three novel isocoumarins, versicoumarins A–C, with four reported isocoumarins, were characterized from an endophytic fungus *Aspergillus versicolor* cultured from *Paris marmorata* plant rhizomes. Versicoumarin A (**72**) exhibited significant anti-TMV activity with 28.6%, inhibition that was comparable to that of positive standard ningnanmycin (31.5%) (Ye et al. 2014). Brefeldin A (**74**) reported from *Penicillium* sp., which is discussed as anticancer compound, also exhibited potent antiviral activity against HCV (hepatitis C virus) and HBV (hepatitis B virus) with an ID₅₀ value of 0.022 and >0.008 μ M, respectively (Xie et al. 2017).

The fungus *Pestalotiopsis thea*, isolated from *Fagara zanthoxyloides*, yielded three known compounds chloroisosulochrin (**93**), ficipyrone A, and pestheic acid. Compound **93** exhibited potent inhibition with an IC₅₀ value of $4.22 \pm 1.03 \mu$ M comparable with that of the standard ribavirin, having IC₅₀ value of $4.91 \pm 1.85 \mu$ M. The rest of the compounds exhibited mild inhibition with IC₅₀ values in the range of 45.00 ± 0.98 – $259.23 \pm 2.36 \mu$ M (Uzor et al. 2016).

The fungus *Periconia* sp. F-31, isolated from *Annona muricata*, was explored and yielded pericoannosin A (94), periconiasin D, periconiasin E, and periconiasin F (95). Compounds 94 and 95 showed considerable anti-HIV activity with IC₅₀ of 69.6 and 29.2 μ M, respectively (Zhang et al. 2015).









Chloroisosulochrin (93)

Pericoannosin A (94)

Periconiasin D (95)

Altertoxin V (96)





Altertoxin II (99)

Fig. 6.9 Structures of antiviral compounds (93-99) from endophytic fungi

Altertoxin II (98)

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Compounds	Endophytic fungi	Host plant	References
Versicoumarin A (72)	Aspergillus versicolor	Paris marmorata	Ye et al. (2014)
Brefeldin A (74)	Penicillium sp.	Panax notoginseng	Xie et al. (2017)
Chloroisosulochrin (93)	Pestalotiopsis thea	Fagara zanthoxyloides	Uzor et al. (2016)
Pericoannosin A (94)	Periconia sp.	Annona muricata	Zhang et al. (2015)
Pericoannosin D (95)			
Altertoxin V (96)	Alternaria tenuissima	Quercus emoryi	Bashyal et al. (2014)
Altertoxin I (97)			
Altertoxin II (98)			
Altertoxin III (99)			

 Table 6.7 Antiviral compounds derived from endophytic fungi

During the bioprospection of endophytic fungi of Sonoran Desert plants, *Alternaria tenuissima* was isolated from *Quercus emoryi*. In screening of natural product extract library and bioactivity-guided fractionation, altertoxin V (96), altertoxin I (97), altertoxin II (98), altertoxin III (99), and altertoxin IV were isolated and characterized. Compounds 96, 97, 98, and 99 have completely inhibited the replication of the HIV-1 virus at a very lower concentration with 0.50, 2.20, 0.30, and 1.50 μ M, respectively (Bashyal et al. 2014).

6.3 Conclusion

Endophytes stay in the internal tissues of the host plants in symbiotic relationship. They are considered to be a rich source for the production of diverse bioactive metabolites and responsible to cause physiological modification in the host which results in the production of compounds required for tolerating biotic and abiotic stresses by plant. The extensive ability of endophytic fungi to synthesize products of pharmaceutical importance led the researchers and scientists to do the sampling from unique niches so that structurally diverse compounds can be isolated. In cases the endophyte gains the ability to produce either the same or different compounds from the host, the production of the same metabolite as of host plant is due to the long coevolution with their host. In the past two decades, researchers have focused on studying fungal diversity, better understanding of their relationship with the host, isolation of novel metabolites from fungus, and different ways to increase the yield of isolated bioactive compounds using genetic engineering and media engineering, improving culture conditions, and using one strain many compounds (OSMAC) approach. Endophytic fungi are considered as a boon for the society due to their ability to produce diverse metabolites of broad applications in various fields like agriculture, horticulture, medical, and industries. Instantly in favor of mankind, there is a need for the hunt of novel and diverse compounds to combat the upcoming challenges of drug resistance among pathogens.

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References

- Abdou R, Abdelhady MI (2015) Anticancer endophytic metabolites of the medicinal plant *Centaurea stoebe*. World J Pharm Pharm Sci 4:220–230
- Arora D, Sharma N, Singamaneni V, Sharma V, Kushwaha M, Abrol V, Guru S, Sharma S, Gupta AP, Bhushan S (2016) Isolation and characterization of bioactive metabolites from *Xylaria psidii*, an endophytic fungus of the medicinal plant *Aegle marmelos* and their role in mito-chondrial dependent apoptosis against pancreatic cancer cells. Phytomedicine 23:1312–1320. https://doi.org/10.1016/j.phymed.2016.07.004
- Bascom-Slack CA, Ma C, Moore E, Babbs B, Fenn K, Greene JS, Hann BD, Keehner J, Kelley-Swift EG, Kembaiyan V, Lee SJ, Li P, Light DY, Lin EH, Schorn MA, Vekhter D, Boulanger LA, Hess WM, Vargas PN, Strobel GA, Strobel SA (2009) Multiple, novel biologically active endophytic actinomycetes isolated from upper Amazonian rain forests. Microb Ecol 58:374– 383. https://doi.org/10.1007/s00248-009-9494-z
- Bashyal BP, Wellensiek BP, Ramakrishnan R, Faeth SH, Ahmad N, Gunatilaka AA (2014) Altertoxins with potent anti-HIV activity from *Alternaria tenuissima* QUE1Se, a fungal endophyte of *Quercus emoryi*. Bioorg Med Chem 22:6112–6116. https://doi.org/10.1016/j. bmc.2014.08.039
- Brader G, Compant S, Mitter B, Trognitz F, Sessitsch A (2014) Metabolic potential of endophytic bacteria. Curr Opin Biotechnol 27:30–37. https://doi.org/10.1016/j.copbio.2013.09.012
- Chapla VM, Zeraik ML, Leptokarydis IH, Silva GH, Bolzani VS, Young MC, Pfenning LH, Araújo AR (2014) Antifungal compounds produced by *Collectorichum gloeosporioides*, an endophytic fungus from *Michelia champaca*. Molecules 19:19243–19252. https://doi.org/10.3390/ molecules191119243
- Dame ZT, Silima B, Gryzenhout M, van Ree T (2016) Bioactive compounds from the endophytic fungus *Fusarium proliferatum*. Nat Prod Res 30:1301–1304. https://doi.org/10.1080/1478641 9.2015.1053089
- Darsih C, Prachyawarakorn V, Wiyakrutta S, Mahidol C, Ruchirawat S, Kittakoop P (2015) Cytotoxic metabolites from the endophytic fungus *Penicillium chermesinum*: discovery of a cysteine-targeted Michael acceptor as a pharmacophore for fragment-based drug discovery, bioconjugation and click reactions. RSC Adv 5:70595–70603. https://doi.org/10.1039/ C5RA13735G
- de Felício R, Pavao GB, de Oliveira ALL, Erbert C, Conti R, Pupo MT, Furtado NAJC, Ferreira EG, Lotufo LVC, Young MCM, Yokoya NS, Debonsi HM (2015) Antibacterial, antifungal and cytotoxic activities exhibited by endophytic fungi from the Brazilian marine red alga *Bostrychia tenella* (Ceramiales). Rev Bras Farmacogn 25:641–650. https://doi.org/10.1016/j. bjp.2015.08.003
- Deshidi R, Devari S, Kushwaha M, Gupta AP, Sharma R, Chib R, Khan IA, Jaglan S, Shah BA (2017) Isolation and quantification of alternariols from endophytic fungus, *Alternaria alternata*: LC-ESI-MS/MS analysis. Chem Select 2:364–368. https://doi.org/10.1002/slct.201601649
- Dudeja S, Giri R, Saini R, Suneja MP, Kothe E (2012) Interaction of endophytic microbes with legumes. J Basic Microbiol 52:248–260. https://doi.org/10.1002/jobm.201100063
- Dzoyem JP, Melong R, Tsamo AT, Maffo T, Kapche DGWF, Ngadjui BT, McGaw LJ, Eloff JN (2017) Cytotoxicity, antioxidant and antibacterial activity of four compounds produced by an endophytic fungus *Epicoccum nigrum* associated with *Entada abyssinica*. Rev Rev Bras Farmacogn 27:251–253. https://doi.org/10.1016/j.bjp.2016.08.0110102-695X

- Elkhayat ES, Goda AM (2017) Antifungal and cytotoxic constituents from the endophytic fungus *Penicillium* sp. Bull Fac Pharm Cairo Univ 55:85–89. https://doi.org/10.1016/j. bfopcu.2017.03.001
- Elkhayat ES, Ibrahim SR, Mohamed GA, Ross SA (2016) Terrenolide S, a new antileishmanial butenolide from the endophytic fungus Aspergillus terreus. Nat Prod Res 30:814–820. https:// doi.org/10.1080/14786419.2015.1072711
- Gao H, Li G, Lou H-X (2018) Structural diversity and biological activities of novel secondary metabolites from endophytes. Molecules 23:646. https://doi.org/10.3390/molecules23030646
- Gasong BT, Tjandrawinata RR (2016) Production of secondary metabolite E2. 2 from *Phaleria macrocarpa* endophytic fungus. Asian Pac J Trop Biomed 6:881–885. https://doi.org/10.1016/j. apjtb.2016.01.005
- Goutam J, Sharma G, Tiwari VK, Mishra A, Kharwar RN, Ramaraj V, Koch B (2017) Isolation and characterization of "Terrein" an antimicrobial and antitumor compound from endophytic fungus Aspergillus terreus (JAS-2) associated from Achyranthes aspera Varanasi, India. Front Microbiol 18(1334). https://doi.org/10.3389/fmicb.2017.01334
- Gunatilaka AL (2006) Natural products from plant-associated microorganisms: distribution, structural diversity, bioactivity, and implications of their occurrence. J Nat Prod 69:509. https://doi. org/10.1021/np058128n
- Hani M, Eman H (2015) Anticancer compounds from *Chaetomium globosum*. Biochem Anal Biochem 4(1). https://doi.org/10.4172/2161-1009.1000174
- Haque MA et al (2005) Isolation of bioactive secondary metabolites from the endophytic fungus of *Ocimum basilicum*. J Pharm Sci 4:127–130
- Hewage RT, Aree T, Mahidol C, Ruchirawat S, Kittakoop P (2014) One strain-many compounds (OSMAC) method for production of polyketides, azaphilones, and an isochromanone using the endophytic fungus *Dothideomycete* sp. Phytochemistry 108:87–94. https://doi.org/10.1016/j. phytochem.2014.09.013
- Hsiao Y, Chang H-S, Ta-Wei L, Sung-Yuan H, Gwo-Fang Y, Ming-Jen C, Chen I-S (2016) Secondary metabolites and bioactivity of the endophytic fungus *Phomopsis theicola* from Taiwanese endemic plant. Rec Nat Prod 10:189
- Hu X, Li W, Yuan M, Li C, Liu S, Jiang C, Wu Y, Cai K, Liu Y (2016) Homoharringtonine production by endophytic fungus isolated from *Cephalotaxus hainanensis* Li. World J Microbiol Biotechnol 32:1–9. https://doi.org/10.1007/s11274-016-2073-9
- Hussain H et al (2014) Antimicrobial constituents from three endophytic fungi. Asian Pac J Trop Med 7:S224–S227. https://doi.org/10.1016/S1995-7645(14)60236-4
- Jouda J-B, Mbazoa CD, Douala-Meli C, Sarkar P, Bag PK, Wandji J (2016) Antibacterial and cytotoxic cytochalasins from the endophytic fungus *Phomopsis sp.* harbored in *Garcinia kola* (Heckel) nut. BMC Complement Altern Med 16:462. https://doi.org/10.1186/ s12906-016-1454-9
- Kajula M, Ward JM, Turpeinen A, Tejesvi MV, Hokkanen J, Tolonen A, Häkkänen H, Picart P, Ihalainen J, Sahl HG, Pirttila AM, Mattila S (2016) Bridged epipolythiodiketopiperazines from *Penicillium raciborskii*, an endophytic fungus of *Rhododendron tomentosum* Harmaja. J Nat Prod 79:685–690. https://doi.org/10.1021/np500822k
- Khan MIH, Sohrab MH, Rony SR, Tareq FS, Hasan CM, Mazid MA (2016) Cytotoxic and antibacterial naphthoquinones from an endophytic fungus, *Cladosporium sp.* Toxicol Rep 3:861– 865. https://doi.org/10.1016/j.toxrep.2016.10.005
- Koizumi F, Hasegawa A, Ando K, Ogawa T, Hara M (2001) Jpn Kokai Tokkyo Koho. JP 2001147574 A2 200109
- Kudo T, Matsushima K, Itoh T, Sasaki J, Suzuki K (1998) Description of four new species of the genus Kineosporia: *Kineosporia succinea* sp. nov., *Kineosporia rhizophila* sp. nov., *Kineosporia mikuniensis* sp. nov. and *Kineosporia rhamnosa* sp. nov., isolated from plant samples, and amended description of the genus Kineosporia. Int J Syst Bacteriol 48:1245–1255. https://doi.org/10.1099/00207713-48-4-1245
- Kumala S, Yuliani KD, Simanjuntak P (2015) Antimicrobial activity of secondary metabolites produced by endophytic fungi isolated from stems of jati tree (*tectona grandis lf*). Int J Pharm Sci Res 6:2349. https://doi.org/10.13040/IJPSR.0975-8232.6(6).2349-53

- Kuriakose GC, Palem PP, Jayabaskaran C (2016) Fungal vincristine from *Eutypella spp*-CrP14 isolated from *Catharanthus roseus* induces apoptosis in human squamous carcinoma cell line-A431. BMC Complement Altern Med 16:302. https://doi.org/10.1186/s12906-016-1299-2
- Kusari S, Zuhlke S, Spiteller M (2011) Effect of artificial reconstitution of the interaction between the plant *Camptotheca acuminata* and the fungal endophyte *Fusarium solani* on camptothecin biosynthesis. J Nat Prod 74:764–775. https://doi.org/10.1021/np1008398
- Lenta BN, Ngatchou J, Frese M, Ladoh-Yemeda F, Voundi S, Nardella F, Michalek C, Wibberg D, Ngouela S, Tsamo E, Kaiser M, Kaliowski J, Sewald N (2016) Purpureone, an antileishmanial ergochrome from the endophytic fungus *Purpureocillium lilacinum*. Z NaturforschB 71:1159–1167. https://doi.org/10.1515/znb-2016-0128
- Li X, Guo Z, Deng Z, Yang J, Zou K (2015a) A new [alpha]-pyrone derivative from endophytic fungus Pestalotiopsis microspora. Rec Nat Prod 9:503. https://doi.org/10.1007/s12010-014-1422-0
- Li Y, Yang J, Zhou X, Zhao W, Jian Z (2015b) Isolation and identification of a 10-deacetyl baccatin-III-producing endophyte from *Taxus wallichiana*. Appl Biochem Biotechnol 175:2224–2231. https://doi.org/10.1007/s12010-014-1422-0
- Li CS, Ren G, Yang BJ, Miklossy G, Turkson J, Fei P, Ding Y, Walker LA, Cao S (2016) Meroterpenoids with antiproliferative activity from a Hawaiian-plant associated fungus *Peyronellaea coffeae-arabicae* FT238. Org Lett 18:2335–2338. https://doi.org/10.1021/acs. orglett.6b00685
- Liang Z, Zhang J, Zhang X, Li J, Zhang X, Zhao C (2015) Endophytic fungus from *Sinopodophyllum emodi* (Wall.) Ying that produces Podophyllotoxin. J Chrom Sci 54:175–178. https://doi.org/10.1093/chromsci/bmv124
- Liu XD, Xu Y (2008) A novel raw starch digesting α-amylase from a newly isolated Bacillussp. YX-1: pur charact. Bioresour Technol 99:4315–4320. https://doi.org/10.1016/j.biortech.2007.08.040
- Liu K, Ding X, Deng B, Chen W (2010) 10-Hydroxycamptothecin produced by a new endophytic *Xylaria sp.*, M20, from *Camptotheca acuminata*. Biotechnol Lett 32:689–693. https://doi. org/10.1007/s10529-010-0201-4
- Lou J, Yu R, Wang X, Mao Z, Fu L, Liu Y, Zhou L (2016) Alternariol 9-methyl ether from the endophytic fungus *Alternaria sp.* Samif01 and its bioactivities. Braz J Microbiol 47:96–101. https://doi.org/10.1016/j.bjm.2015.11.004
- Malmstrøm J, Christophersen C, Frisvad JC (2000) Secondary metabolites characteristic of *Penicillium citrinum, Penicillium steckii* and related species. Phytochemistry 54:301–309. https://doi.org/10.1016/S0031-9422(00)00106-0
- Mandavid H, Rodrigues AM, Espindola LS, Eparvier V, Stien D (2015) Secondary metabolites isolated from the amazonian endophytic fungus *Diaporthe sp.* SNB-GSS10. J Nat Prod 78:1735– 1739. https://doi.org/10.1021/np501029s
- Metwaly AM, Ghoneim MM, Musa A (2015) Two new antileishmanial diketopiperazine alkaloids from the endophytic fungus *Trichosporum* sp. Der Pharma Chem 7:322–327
- Mishra P, Verekar S, Kulkarni-Almeida A, Roy S (2013) Anti-inflammatory and anti-diabetic naphthaquinones from an endophytic fungus *Dendryphion nanum* (Nees) S. Hughes. Indian J Chem 52B:565–567
- Musavi SF, Dhavale A, Balakrishnan RM (2015) Optimization and kinetic modeling of cellassociated camptothecin production from an endophytic *Fusarium oxysporum* NFX06. Prep Biochem Biotechnol 45:158–172. https://doi.org/10.1016/j.biortech.2014.12.106
- Nair DN, Padmavathy S (2014) Impact of endophytic microorganisms on plants, environment and humans. Sci World J 2014:250693. https://doi.org/10.1155/2014/250693
- Naureen H, Asker MMS, Shaaban M (2015) Structural elucidation and bioactivity studies of secondary metabolites from endophytic Aspergillus niger. Indian J Appl Res 5:76–83. https://doi. org/10.15373/2249555X
- Page M, Landry N, Boissinot M, Helie M-C, Harvey M, Gagne M (2000) Bacterial mass production of taxanes and paclitaxel. US Patent, US6030818 A, 29 feb 2000
- Pastre R, Marinho AM, Rodrigues-Filho E, Souza AQ, Pereira JO (2007) Diversity of polyketides produced by *Penicillium species* isolated from Melia azedarach and Murraya paniculata. Quim Nova 30:1867–1871. https://doi.org/10.1590/S0100-40422007000800013

- Pereira CB, de Oliveira DM, Hughes AFS, Kohlhoff M, LA Vieira M, Martins Vaz AB, Ferreira MC, Carvalho CR, Rosa LH, Rosa CA, Alves TM, Zani CL, Johann S, Cota BB (2015) Endophytic fungal compounds active against *Cryptococcus neoformans* and *C. gattii*. J Antibiot 68:436. https://doi.org/10.1038/ja.2015.11
- Pimentel MR, Molina G, Dionísio AP, Maróstica Junior MR, Pastore GM (2011) The use of endophytes to obtain bioactive compounds and their application in biotransformation process. Biotechnol Res Int 2011:1–11. https://doi.org/10.4061/2011/576286
- Prihantini AI, Tachibana S (2017) Antioxidant compounds produced by *Pseudocercospora sp.* ESL 02, an endophytic fungus isolated from *Elaeocarpus sylvestris*. Asian Pac J Trop Biomed 7:110–115. https://doi.org/10.1016/j.apjtb.2016.11.020
- Pu X, Qu X, Chen F, Bao J, Zhang G, Luo Y (2013) Camptothecin-producing endophytic fungus Trichoderma atroviride LY357: isolation, identification, and fermentation conditions optimization for camptothecin production. Appl Microbiol Biotechnol 97:9365–9375. https://doi. org/10.1007/s00253-013-5163-8
- Qader M, Kumar N, Jayasinghe L, Fujimoto Y (2015) Production of antitumor antibiotic GKK1032B by *Penicillium citrinum*, an endophytic fungus isolated from *Garcinia mangostana* fruits. Med Aromat Plants 5:225. https://doi.org/10.4172/2167-0412.100022
- Qin S, Wang H-B, Chen H-H, Zhang Y-Q, Jiang C-L, Xu L-H, Li W-J (2008) Glycomyces endophyticus sp. nov., an endophytic actinomycete isolated from the root of Carex baccans Nees. Int J Syst Evol Microbiol 58:2525–2528. https://doi.org/10.1099/ijs.0.2008/000398-0
- Qin S, Chen H-H, Klenk H-P, Kim C-J, Xu L-H, Li W-J (2010) Saccharopolyspora gloriosae sp. nov., an endophytic actinomycete isolated from the stem of Gloriosa superba L. Int J Syst Evol Microbiol 60:1147–1151. https://doi.org/10.1099/ijs.0.015792-0
- Rathod D, Dar M, Gade A, Rai M (2014) Griseofulvin producing endophytic *Nigrospora oryzae* from Indian *Emblica officinalis* Gaertn: a new report. Austin J Biotechnol Bioeng 1:5
- Ratnaweera PB, de Silva ED, Williams DE, Andersen RJ (2015) Antimicrobial activities of endophytic fungi obtained from the arid zone invasive plant *Opuntia dillenii* and the isolation of equisetin, from endophytic *Fusarium* sp. BMC Complement Altern Med 15:220. https://doi. org/10.1186/s12906-015-0722-4
- Rivera-Chávez J, González-Andrade M, del Carmen González M, Glenn AE, Mata R (2013) Thielavins A, J and K: α-glucosidase inhibitors from MEXU 27095, an endophytic fungus from *Hintonia latiflora*. Phytochemistry 94:198–205. https://doi.org/10.1016/j. phytochem.2013.05.021
- Shaaban M, Nasr H, Hassan AZ, Asker MS (2013) Bioactive secondary metabolites from endophytic Aspergillus fumigatus: structural elucidation and bioactivity studies. Rev Latinoam Quim 41:50–60
- Sharma N, Kushwaha M, Arora D, Jain S, Singamaneni V, Sharma S, Shankar R, Bhushan S, Gupta P, Jaglan S (2018) New cytochalasin from *Rosellinia sanctae-cruciana*, an endophytic fungus of *Albizia lebbeck*. J Appl Microbiol 125(1):111–120. https://doi.org/10.1111/jam.13764
- Shentu X, Zhan X, Ma Z, Yu X, Zhang C (2014) Antifungal activity of metabolites of the endophytic fungus *Trichoderma brevicompactum* from garlic. Braz J Microbiol 45:248–254. https:// doi.org/10.1590/S1517-83822014005000036
- Shimizu M, Furumai T, Igarashi Y, Onaka H, Nishimura T, Yoshida R, Kunoh H (2001) Association of induced disease resistance of *rhododendron* seedlings with inoculation of *Streptomyces* sp. R-5 and treatment with actinomycin D and amphotericin B to the tissue-culture medium. J Antibiot 54:501–505. https://doi.org/10.7164/antibiotics.54.501
- Silva MO, Kawai K, Hosoe T, Takaki GC, Gusmão NB, Fukushima K (2013) Viriditoxin, an antibacterial substance produced by mangrove endophytic fungus *Paecilomyces variotii*. Microbial pathogens and strategies for combating them: science, technology and education (A Méndez-Vilas, Ed):85–100
- Singh B, Kaur A (2016) Antidiabetic potential of a peptide isolated from an endophytic *Aspergillus awamori*. J Appl Microbiol 120:301–311. https://doi.org/10.1111/jam.12998

- Sousa J, Aguilar Pérez M, Arnold A, Rios N, Coley P, Kursar T, Cubilla Rios L (2016) Chemical constituents and their antibacterial activity from the tropical endophytic fungus *Diaporthe* sp. F2934. J Appl Microbiol 120:1501–1508. https://doi.org/10.1111/jam.13132
- Stierle A, Strobel G, Stierle D (1993) Taxol and taxane production by *Taxomyces andreanae*, an endophytic fungus of Pacific yew. Science 260:214–216. https://doi.org/10.1126/science.8097061
- Strobel G, Yang X, Sears J, Kramer R, Sidhu RS, Hess W (1996) Taxol from *Pestalotiopsis microspora*, an endophytic fungus of Taxus wallichiana. Microbiology 142:435–440. https://doi.org/10.1099/13500872-142-2-435
- Tan RX, Zou WX (2001) Endophytes: a rich source of functional metabolites. Nat Prod Rep 18:448–459. https://doi.org/10.1039/B100918O
- Taware R, Abnave P, Patil D, Rajamohananan PR, Raja R, Soundararajan G, Kundu GC, Ahmad A (2014) Isolation, purification and characterization of Trichothecinol-A produced by endophytic fungus *Trichothecium* sp. and its antifungal, anticancer and antimetastatic activities. Sustain Chem Process 2:8. https://doi.org/10.1186/2043-7129-2-8
- Tayung K, Barik B, Jha D, Deka D (2011) Identification and characterization of antimicrobial metabolite from an endophytic fungus, *Fusarium solani* isolated from bark of Himalayan yew. Mycosphere 2:203–213
- Uesugi S, Fujisawa N, Yoshida J, Watanabe M, Dan S, Yamori T, Shiono Y, Kimura K (2016) Pyrrocidine A, a metabolite of endophytic fungi, has a potent apoptosis-inducing activity against HL60 cells through caspase activation via the Michael addition. J Antibiot 69:133. https://doi.org/10.1038/ja.2015.103
- Uzor PF, Odimegwu DC, Ebrahim W, Osadebe PO, Nwodo NJ, Okoye FB, Liu Z, Proksch P (2016) Anti-respiratory syncytial virus compounds from two endophytic fungi isolated from nigerian medicinal plants. Drug Res 66:527–531. https://doi.org/10.1055/s-0042-111008
- Uzor PF, Osadebe PO, Nwodo NJ (2017) Antidiabetic activity of extract and compounds from an endophytic fungus *Nigrospora oryzae*. Drug Res 67:308–311. https://doi. org/10.1055/s-0042-122777
- Venugopalan A, Srivastava S (2015) Enhanced camptothecin production by ethanol addition in the suspension culture of the endophyte, *Fusarium solani*. Bioresour Technol 188:251–257. https://doi.org/10.1016/j.biortech.2014.12.106
- Verekar SA, Mishra PD, Sreekumar ES, Deshmukh SK, Fiebig H-H, Kelter G, Maier A (2014) Anticancer activity of new depsipeptide compound isolated from an endophytic fungus. J Antibiot 67:697. https://doi.org/10.1038/ja.2014.58
- Wang X, Radwan MM, Taráwneh AH, Gao J, Wedge DE, Rosa LH, Cutler HG, Cutler SJ (2013) Antifungal activity against plant pathogens of metabolites from the endophytic fungus *Cladosporium cladosporioides*. J Agric Food Chem 61:4551. https://doi.org/10.1021/ jf400212y
- Wang W-X, Kusari S, Laatsch H, Golz C, Kusari P, Strohmann C, Kayser O, Spiteller M (2016) Antibacterial azaphilones from an endophytic fungus, *Colletotrichum* sp. BS4. J Nat Prod 79:704–710. https://doi.org/10.1021/acs.jnatprod.5b00436
- Wani MC, Taylor HL (1971) Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemia and antitumor agent from [the stem bark of] *Taxus brevifolia*. J Am Chem Soc 93:2325–2327. https://doi.org/10.1021/ja00738a045
- Wibowo M, Prachyawarakorn V, Aree T, Wiyakrutta S, Mahidol C, Ruchirawat S, Kittakoop P (2014) Tricyclic and spirobicyclic norsesquiterpenes from the endophytic fungus *Pseudolagarobasidium acaciicola*. Eur J Org Chem 2014:3976–3980. https://doi.org/10.1002/ ejoc.201402262
- Xie J, Wu Y-Y, Zhang T-Y, Zhang M-Y, Zhu W-W, Gullen EA, Wang Z-J, Cheng Y-C, Zhang Y-X (2017) New and bioactive natural products from an endophyte of *Panax notoginseng*. RSC Adv 7:38100–38109. https://doi.org/10.1039/c7ra07060h
- Ye Y-q, Xia C-F, Yang J-X Qin Y, Zhou M, Gao X-M, Du G, Yang H-Y, Li X-M, Hu Q-F (2014) Isocoumarins from the fermentation products of an endophytic fungus of *Aspergillus versi*color. Phytochem Lett 10:215–218. https://doi.org/10.1016/j.phytol.2014.09.016

- Yin OCJ, Ibrahim D, Lee CC (2015) Bioactive compounds from *Aspergillus terreus* MP15, an endophytic fungus isolated from *Swietenia Macrophylla* leaf. Malay J Med Biol Res 2:262–272. https://doi:10.18034/mjmbr
- Zaher AM, Makboul MA, Moharram AM, Tekwani BL, Calderón AI (2015) A new enniatin antibiotic from the endophyte *Fusarium tricinctum Corda*. J Antibiot 68:197. https://doi.org/10.1038/ja.2014.129
- Zhang D, Tao X, Chen R, Liu J, Li L, Fang X, Yu L, Dai J (2015) Pericoannosin A, a polyketide synthase nonribosomal peptide synthetase hybrid metabolite with new carbon skeleton from the endophytic fungus *Periconia* sp. Org Lett 17:4304–4307. https://doi.org/10.1021/acs. orglett.5b02123
- Zhao J, Zhou L, Wang J, Shan T, Zhong L, Liu X, Gao X (2010) Endophytic fungi for producing bioactive compounds originally from their host plants. Curr Res Technol Educ Trop Appl Microbiol Microb Biotechnol 1:567–576
- Zuck D (1998) Drugs prototypes and their exploitation. Anaesthesia 53:103–104. https://doi. org/10.1111/j.1365-2044.1998.0345b.x