

# Subarachnoid Hemorrhage Pattern Predicts Acute Cerebral Blood Flow Response in the Rat



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**Abstract** There is considerable variability in the presentation of patients with acute subarachnoid hemorrhage (aSAH). Evidence suggests that a thick, diffuse clot better predicts the development of delayed cerebral ischemia and poor outcomes. In a rodent model of acute SAH, we directly measured the effects of the volume of blood injected versus the pattern of distribution of hemorrhage in the subarachnoid space on markers of early brain injury, namely, cerebral blood flow (CBF), cerebrospinal fluid (CSF) concentrations of P450 eicosanoids and catecholamines, and cortical spreading depolarizations (CSDs). There is a significant decrease in CBF, an increase in CSF biomarkers, and a trend toward increasing frequency and severity of CSDs when grouped by severity of hemorrhage but not by volume of blood injected. In severe hemorrhage grade animals, there was a progressive decrease in CBF after successive CSD events. These results suggest that the pattern of SAH (thick diffuse clots) correlates with the “clinical” severity of SAH.

**Keywords** Subarachnoid hemorrhage · P450 eicosanoids · 20-HETE · 14,15-EET · Cortical spreading depolarization · Early brain injury · Stroke glymphatic

## Introduction

Aneurysmal subarachnoid hemorrhage (aSAH) is a devastating form of stroke with an incidence of approximately 9 in 100,000 person-years [1]. This condition results in immediate mortality of approximately 22%. There is a nearly 50% mortality rate in the first month, and of those patients surviving at least 1 year, 46–60% remain dependent [2]. One major hallmark of aSAH is the marked variability in clinical presentation, radiographic appearance, and long-term outcomes across patients. Furthermore, many patients, but not all, develop delayed cerebral ischemia (DCI) that, in the past, has been attributed to large vessel vasospasm due to numerous vasoactive compounds in the subarachnoid space. Progressive iterations of radiographic grading scales have improved the prediction of DCI by accounting for diffuse thick hemorrhage [3–5]. In addition, several recent reports have shown that the *pattern* of SAH and not the *volume* of blood per se, in patients with *non-aneurysmal* SAH is correlated with the development of angiographic vasospasm and delayed infarcts [6–8].

Early brain injury (EBI), damage sustained within the first 72 h after ictus, leads to delayed pathological changes such as brain tissue hypoxia, cerebral inflammation, and blood-brain barrier disruption [9, 10]. EBI plays a primary role in the initial clinical presentation of patients with SAH and contributes to secondary injuries, including DCI and subsequent poor prognosis [10]. Consequently, multiple groups have been studying the effects of various biomarkers after aSAH. The P450 eicosanoids are vasoactive and inflammatory products of arachidonic acid metabolism that have been shown to be present in the spinal fluid of aSAH patients. These metabolites play numerous and often opposing roles. Some have been shown to be vasoconstrictive and contribute to DCI [11, 12], and others have been shown to be neuroprotective in cerebral ischemia/reperfusion injury and can reduce cerebral edema [13, 14].

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Epoxyeicosatrienoic acids (EETs) are vasodilatory, anti-inflammatory, and antithrombotic [15]. 14,15-EET may protect against the development of DCI in both human and animal studies [16, 17], whereas 20-hydroxyeicoastetraenoic acid (20-HETE) and 5-hydroxytryptamine (5-HT) are associated with early and delayed vasospasm [11, 12]. Another hallmark brain injury after SAH is the presence of cortical spreading depolarizations (CSDs) that may contribute to the development of DCI [18]. These are self-propagating waves of neuronal and glial depolarization that trigger a vascular response through neurovascular coupling. The vascular response can range from a spreading hyperemia to a pronounced oligemia and can lead to significant ischemia and cell death [19]. Under normal conditions, CSDs lead to increased cerebral metabolic demand and trigger a hemodynamic response composed of multiple opposing vasomotor influences at various stages of the CSD wave [20]. In the pathologic state after SAH CSDs lead to paradoxical hypoperfusion during the period of elevated metabolic demand which leads to tissue damage [21]. CSDs have been well-documented in animal models and human patients with severe SAH [19, 21]. We propose that a thick diffuse pattern of hemorrhage is the driver of the cascade of events that occurs after hemorrhage rather than a dose response to the spasmogens in the extravasated blood.

In this study, we directly compare the *pattern* of blood in experimental SAH [22, 23] to the *volume* of blood on cerebral blood flow, the CSF expression of P450 eicosanoids, and the development of CSDs. We find that a thick diffuse hemorrhage most closely correlates with the severity of injury after SAH.

## Materials and Methods

### Animals and Surgical Preparation

This study protocol was approved by the Institutional Animal Care and Use Committee at Oregon Health & Science University, which is in accordance with the NIH Guide for the Care and Use of Laboratory Animals. All institutional and national guidelines for the care and use of laboratory animals were followed. As previously published [22], rats were anesthetized with isoflurane, catheterized (central venous and arterial), placed in a stereotactic frame, and maintained in a “lightly anesthetized state” with i.v. Brevital. The skull was thinned for laser Doppler flowmetry and optical intrinsic signaling (OIS) and a craniotomy performed for placement of a spinal needle in the prechiasmatic cistern for injection of autologous blood into the subarachnoid space.

### Experimental Subarachnoid Hemorrhage and Recording

Freshly drawn autologous blood of varying volumes (50  $\mu$ L, 100  $\mu$ L, 200  $\mu$ L, and 250  $\mu$ L) was infused over 1 min through the spinal needle to minimize changes in intracranial pressure (ICP) [24, 25]. Control animals received an injection of 200  $\mu$ L of artificial cerebral spinal fluid (aCSF). CBF, mean arterial pressure (MAP), flick latency, and temperature were monitored during a 10-min baseline to ensure a stable anesthetic plane and for an additional 30 min after induction of SAH. OIS was recorded using a multispectral imaging system using QiOptiq Optem FUSION (QiOptiq, NY) modular lens system. A broadband light from a fiber-optic illuminator filtered through a liquid crystal tunable filter (VariSpec VIS, Perkin-Elmer, MA) was used to illuminate the thin skull preparation 3 mm right lateral to the sagittal suture and 2 mm posterior to the coronal suture, at 30° angle with respect to the optical axis of the imaging system. The reflectance from the sample was acquired by monochrome camera (Model: Flea2 IEEEEb, Point Grey Research, Inc., Canada) controlled by custom software written using LabVIEW (National Instruments, Austin, TX).

### Cerebrospinal Fluid Collection

The rat was given a bolus of 0.5 mL sodium Brevital (20 mg/mL), and the head angled ventrally 30°. A 27-gauge needle was percutaneously inserted into the cisterna magna and 100–200  $\mu$ L of CSF aspirated. CSF was centrifuged for 10 s at 10,000 RPM and then at  $-80^{\circ}$  C. Control CSF was obtained from anesthetized naïve rats in a similar fashion. All samples were analyzed for P450 eicosanoids and catecholamines by the lipidomics core on a mass spectrometer. Following CSF collection, the animals were euthanized by sodium Brevital overdose and perfusion fixed with 100 mL saline followed by 100 mL 10% formalin. The whole brain was removed carefully and images of the ventral and dorsal surfaces obtained. The hemorrhage grade was assigned as previously described [22]. Briefly, the ventral surface of the brain was partitioned into six sections. Each section was allotted a number from 0 to 3 depending on the amount of subarachnoid blood in the section: 0 no blood, 1 minimal blood, 2 moderate blood clot with recognizable arteries, and 3 blood clot obliterating all arteries. Scores were summed from all six sections. Scores were categorized as follows: 0–7 mild, 8–12 moderate, and 13–18 severe.

### Analysis

CBF was normalized to the baseline. The mean percent change, in 5-min epochs, was calculated for each group and

compared using the one-way ANOVA and post hoc Tukey test. Differences in the mean concentrations of metabolite markers in the CSF were compared using the one-way ANOVA and post hoc Tukey test. The number of cortical spreading depressions was compared with the nonparametric Kruskal-Wallis test. Statistics were performed with MATLAB and IBM SPSS v24 software.

## Results

There were a total of 31 experimental animals. Five (5) animals were given a 250  $\mu$ L injection, eight (8) animals were given a 200  $\mu$ L injection, four (4) animals were given a 100  $\mu$ L injection, and four (4) animals were given a 50  $\mu$ L injection. Ten (10) animals were given an aCSF injection. This resulted in fourteen (14) mild hemorrhages, eight (8) moderate hemorrhages, and nine (9) severe hemorrhages. There were no severe hemorrhages in the 50  $\mu$ L group (2/4 mild and 2/4 moderate). Of the other three injection groups, there were 25% (1/4), 62.5% (5/8), and 60% (3/5) severe hemorrhages in the 100  $\mu$ L, 200  $\mu$ L, and 250  $\mu$ L injection groups, respectively.

### Cerebral Blood Flow

CBF was reduced immediately after experimental SAH in all animals; however, there was a large amount of variability in the severity of the reduction. CBF categorized according to the volume of blood injected was only significantly reduced from aCSF injection in the 250  $\mu$ L group (the largest volume injected) due to the large variability in responses across groups (Fig. 1a). By contrast, there is a significant difference in relative CBF depending on the hemorrhage grade of the animal (Fig. 1b) consistent with previous reports [22]. There is a substantial decrease in CBF in the early phase after SAH injection compared to the baseline in the moderate and severe groups. The differences between mild, moderate, and severe hemorrhages are significant for the first minutes after injection ( $P < 0.05$ ), after which differences between mild and severe hemorrhages persist for 20 min post-injection ( $P < 0.05$ ).

### Cerebrospinal Fluid

The concentration of 14,15-EET was significantly elevated in all hemorrhage grades compared to naïve controls ( $P < 0.01$ ). When grouped by injected blood volume, only the largest blood volume group was statistically different

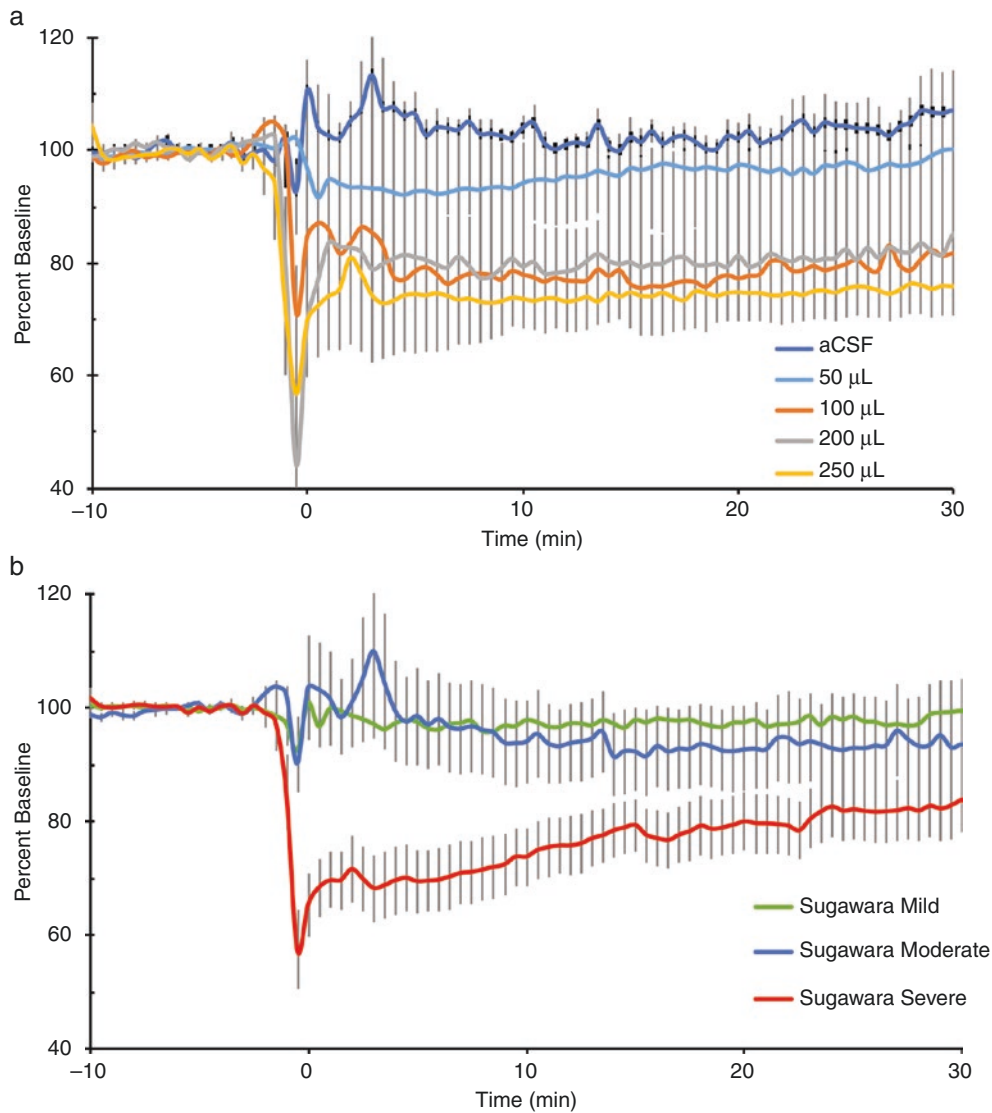
( $P < 0.05$ ) likely due to the large variability. Similarly, the mean concentrations of 20-HETE are significantly different when grouped by hemorrhage severity but not by volume injected. The mean concentration of 5-HT is significantly different in severe hemorrhage compared to control ( $P < 0.01$ ) and moderate hemorrhage ( $P < 0.05$ ), whereas there is no significance when grouped by volume injected. There was no significant difference in the CSF concentration of norepinephrine (NE) independent of grouping (Fig. 2 and Table 1).

### Cortical Spreading Depolarization

The number of CSD events were compared between hemorrhage grades as well as volume of blood injected. Of the 14 animals with mild hemorrhages, 3 experienced CSD events; of the 8 moderate hemorrhage animals, 3 experienced CSD events; and of the 9 severe hemorrhage animals, 5 experienced CSD events. There was a trend toward increased frequency of CSD events in severe hemorrhages, but it did not quite reach significance ( $P = 0.064$ ). In animals with multiple CSD events, subsequent CSD events appeared different between mild and severe hemorrhage grades (Fig. 3). In the mild hemorrhage animal, the initial CSD event was a brief period of oligemia followed by hyperemia. In subsequent CSD events, oligemia diminished or was absent and the hyperemic response increased. In the moderate hemorrhage animal, there is a more gradual shift from oligemia to hyperemia. In the severe hemorrhage animal, successive CSD events lead to progressively worsening oligemia and a loss of the hyperemic component (Fig. 3).

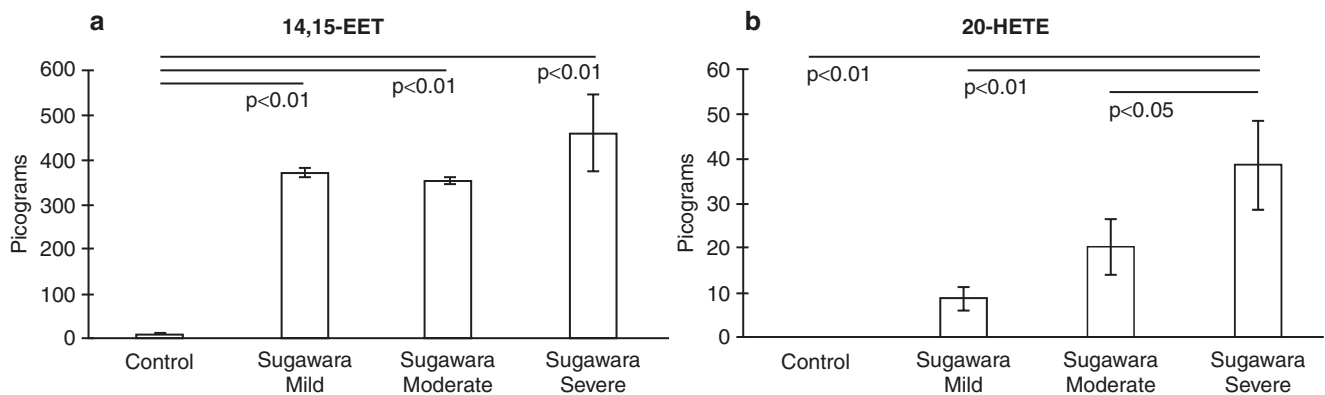
## Discussion

In this study we examined the correlation between the volume of blood injected in the subarachnoid space and the distribution or pattern of SAH on markers of early brain injury: cerebral perfusion, CSF levels of the P450 eicosanoids and catecholamines (14,15-EET, 20-HETE, 5-HT, and NE), as well as the development of CSDs and their effects on CBF. We found that an acute decrease in cerebral perfusion was quite variable across animals if grouped by the volume of blood injected. By contrast, Sugawara grade correlated closely with the severity of cerebral perfusion deficits and levels of the vasoactive metabolites 20-HETE, 5-HT, and 14–15-EET. There was a trend toward increasing frequency of CSD events in high-grade hemorrhages, and these CSD profiles demonstrated worsening oligemia compared to those associated with low-grade hemorrhages. These changes



**Fig. 1** rCBF after induced SAH (a) When grouped by volume of blood injected, only the 250  $\mu\text{L}$  group demonstrated a significant difference from controls (aCSF injection) and was not different from other injected blood volume groups. (b) When grouped by pattern of clot in subarachnoid space (Sugawara grade), there is a substantial decrease in rCBF

immediately after blood injection in the moderate and severe groups compared to the baseline. Mild, moderate, and severe hemorrhages are significantly different from each other until 5 min after injection ( $P < 0.05$ ), after which mild and severe hemorrhages remain significantly different until 20 min after the injection ( $P < 0.05$ )



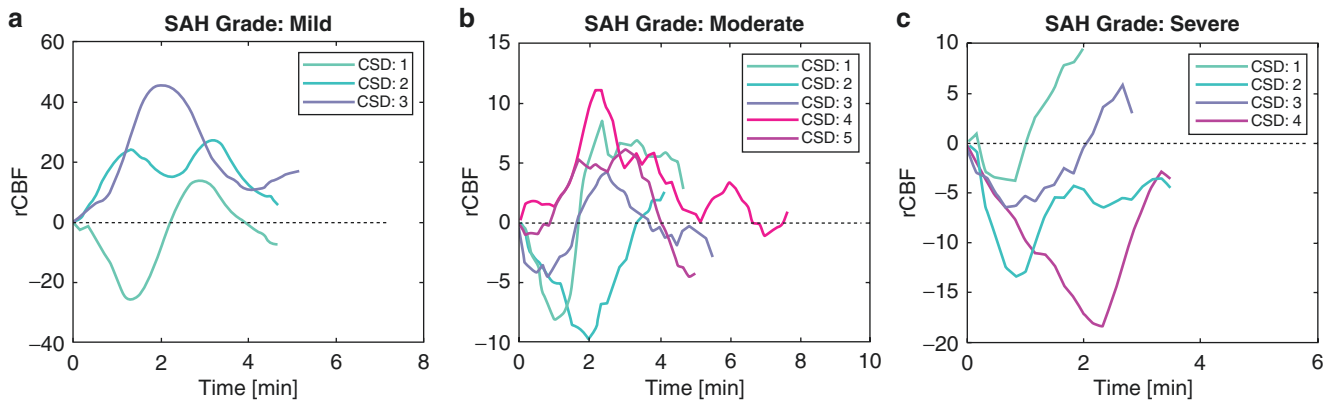
**Fig. 2** (a) CSF concentrations of 14,15-EET increased in all hemorrhage grades after experimental hemorrhage. (b) CSF concentrations of 20-HETE increased proportionally with increasing severity of hemorrhage grade

**Table 1** Eicosanoid concentrations by hemorrhage grade and volume injected blood

Metabolite	Hemorrhage Grade				Injection Volume		
	Control	Sugawara Mild	Sugawara Moderate	Sugawara Severe	aCSF	50&100 $\mu$ L	200&250 $\mu$ L
14,15-EET	6.75 $\pm$ 3.94	371.5 $\pm$ 11.44*	354.4 $\pm$ 6.66*	460.8 $\pm$ 87.5*	396.1 $\pm$ 45.77	440.7 $\pm$ 97.80	458.3 $\pm$ 121.65°
20-HETE	0	8.7 $\pm$ 2.52	20.2 $\pm$ 6.31	38.6 $\pm$ 10.20*#‡	1.5 $\pm$ 1.45	19.9 $\pm$ 7.55	22.1 $\pm$ 9.10
5-HT	0.09 $\pm$ 0.02	18 $\pm$ 5.70	11.6 $\pm$ 5.23	44.4 $\pm$ 10.91**	13.1 $\pm$ 4.78	19.6 $\pm$ 8.34	28.0 $\pm$ 8.86
NE	0.58 $\pm$ 0.03	1.80 $\pm$ 1.27	3.43 $\pm$ 2.63	0.97 $\pm$ 0.21	3.24 $\pm$ 1.99	0.46 $\pm$ 0.23	0.53 $\pm$ 0.20

Units are pg/100  $\mu$ L for the eicosanoids (EETs, HETEs, and DHETs) and units are ng/mL for NE

\* $P < 0.01$  when compared to Control; ° $P < 0.05$  when compared to Control; #  $P < 0.05$  when compared to Moderate; ‡  $P < 0.01$  when compared to Mild



**Fig. 3** rCBF changes in successive CSD events. (a) An animal with mild-grade hemorrhage and three spontaneous CSD events. Initial oligemia of the first CSD is absent in subsequent CSD events. (b) An animal with moderate-grade hemorrhage and five spontaneous CSD events. Initial oligemia persists for four CSD events but begins to diminish after the second event and is absent in the fifth event. The late hyperemic phase increases in magnitude with each successive wave. In

high-grade hemorrhage, there is an initial oligemia with a mild compensatory hyperemia that progresses to severe, uncompensated oligemia. (c) An animal with severe-grade hemorrhage and four CSD events. Initial oligemia increases with each successive event, while the compensatory hyperemia diminishes and is eventually absent in the third and fourth waves

likely contribute to early brain injury and prime the brain for DCI [26].

Acute global ischemia is well-known to accompany high clinical grade hemorrhages. Patients have poor neurologic function, cerebral edema, and at times early infarcts [27, 28]. In our study, animals with large perfusion deficits had significant increases in the CSF concentration of 20-HETE, 14,15-EET, and 5-HT. Previous work has shown that an increase in 5-HT stimulates the synthesis and/or release of 20-HETE, which is a powerful vasoconstrictor [11] that contributes to vasospasm and poor outcomes in animal models and human studies [17, 29, 30]. 14,15-EET, on the other hand, is a potent vasodilator [17] and has been shown to be neuroprotective in ischemic stroke [31, 32], reduce perivascular inflammation [14], and decrease DCI [17, 33] after SAH. Different genetic polymorphisms of soluble epoxide hydrolase (sEH), an enzyme that metabolizes EETs into inactive secondary products, have been shown to correlate with neurologic outcomes after SAH [14, 16, 33]. Human polymorphisms with decreased sEH activity or animal models of knockout sEH demon-

strated improved neurologic outcomes. Both 14,15-EET and 20-HETE are elevated in the CSF of patients with poor clinical grade [17, 30, 33]. Consistent with these clinical studies, we demonstrate elevated 20-HETE and 14,15-EET with severe hemorrhage grades. Interestingly, 14,15-EET is also elevated in low-grade hemorrhages suggesting that an increase in 14,15-EET is a compensatory response to SAH to maintain cerebral blood flow and that it is overwhelmed by the increased vasoconstrictive tone secondary to 20-HETE and 5-HT release in high-grade hemorrhages [17, 34].

Though previous work has reported elevated CSF epinephrine levels in human subjects at a higher risk for early death or disability [34], the CSF concentration of NE was not significantly different regardless of animals grouped by hemorrhage grade or volume of blood injected. The predictive value of elevated NE and epinephrine was weak with low sensitivity and specificity in the human study, and our finding is consistent given the small sample size. It has also been shown that plasma concentrations of catecholamines can be increased after SAH [35]. Our data did not demonstrate con-

sistently elevated catecholamines in the CSF in high-grade hemorrhages, which suggests that catecholamine release associated with SAH is peripheral rather than central. Interestingly, we did not see a close relationship between CSF marker and volume of blood injected which would have been expected if CSF levels of metabolites were due to their presence in the peripheral blood used for SAH. In our model, ICP is controlled which may impact the degree of plasma catecholamines in the CSF sample due to a diminished Cushing's response.

Consistent with the findings of reduced CBF and increased CSF levels of 20-HETE, there is a trend toward increasing frequency of CSD events with increasing hemorrhage grade. Increased 20-HETE reduces CBF through vasoconstriction and has also been shown to be released at the level of the capillaries after CSD events [36] potentially exacerbating underlying ischemia. Interestingly, the CSD events in an animal with severe hemorrhage and multiple CSD events demonstrated increasing oligemia with each successive event (Fig. 3). By contrast, in animals with mild and moderate hemorrhages, the successive CSD events demonstrated hyperemic responses suggesting intact compensatory systems. Without these systems, animals with severe hemorrhages are at an increased risk of global ischemia and potential infarction due to decoupling of the neurovascular unit and an inability to compensate for the elevated metabolic demand [37].

The risk of DCI is also likely related to the diffuse distribution of blood through the subarachnoid space. Previous work has demonstrated the importance of a paravascular, glymphatic, waste clearance mechanism in the brain which is disrupted by blood in the subarachnoid space [38–40]. Tissue plasminogen activator (tPA) injected into the intraventricular space improves glymphatic perfusion and restores CSF flow [41–43] and cortical perfusion [43] after experimental SAH. In addition, parenchymal CSF flow is severely impaired secondary to perivascular blood clots which supports a localized effect of blood and its contribution to DCI [39]. Our model and data support the idea that the diffuse, thick distribution of subarachnoid blood impairs paravascular, glymphatic CSF flow and increases the incidence of EBI and DCI.

## Conclusion

In conclusion, this study demonstrates no significant correlation between the volume of blood injected into the subarachnoid space and the severity of induced SAH as measured by the modified Sugawara grade. When grouped by hemorrhage grade, there is a significant decreased initial post SAH CBF; increased CSF concentrations of 20-HETE, 14,15-EET, and 5-HT; and a trend toward increased CSD events in high

Sugawara grade experimental SAH. Our model confirms that the diffuse, thick pattern of SAH leads to increased early brain injury and therefore a likely increased risk of DCI.

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**Conflict of Interest:** The authors declare that they have no conflict of interest.

## References

1. de Rooij NK, Linn FHH, van der Plas JA, Algra A, Rinkel GJE. Incidence of subarachnoid haemorrhage: a systematic review with emphasis on region, age, gender and time trends. *J Neurol Neurosurg Psychiatry*. 2007;78(12):1365–72.
2. Al-Shahi R. Subarachnoid haemorrhage. *BMJ*. 2006;333(7561):235–40.
3. Dengler NF, Diesing D, Sarrafzadeh A, Wolf S, Vajkoczy P. The Barrow neurological institute scale revisited: predictive capabilities for cerebral infarction and clinical outcome in patients with aneurysmal subarachnoid hemorrhage. *Neurosurgery*. 2017;81(2):341–9.
4. Fisher CM, Kistler JP, Davis JM. Relation of cerebral vasospasm to subarachnoid hemorrhage visualized by computerized tomographic scanning. *Neurosurgery*. 1980;6(1):1–9.
5. Frontera JA, Claassen J, Schmidt JM, Wartenberg KE, Temes R, Connolly ES, MacDonald RL, Mayer SA. Prediction of symptomatic vasospasm after subarachnoid hemorrhage: the modified fisher scale. *Neurosurgery*. 2006;59(1):21–7. discussion 21–27.
6. Konczalla J, Kashefiolasi S, Brawanski N, Bruder M, Gessler F, Senft C, Berkefeld J, Seifert V, Tritt S. Cerebral vasospasm-dependent and cerebral vasospasm-independent cerebral infarctions predict outcome after nonaneurysmal subarachnoid hemorrhage: a single-center series with 250 patients. *World Neurosurg*. 2017;106:861–869.e4.
7. Raya A, Zipfel GJ, Diringer MN, Dacey RG, Derdeyn CP, Rich KM, Chicoine MR, Dhar R. Pattern not volume of bleeding predicts angiographic vasospasm in nonaneurysmal subarachnoid hemorrhage. *Stroke*. 2014;45(1):265–7.
8. Walcott BP, Stapleton CJ, Koch MJ, Ogilvy CS. Diffuse patterns of nonaneurysmal subarachnoid hemorrhage originating from the Basal cisterns have predictable vasospasm rates similar to aneurysmal subarachnoid hemorrhage. *J Stroke Cerebrovasc Dis*. 2015;24(4):795–801.
9. Al-Mufti F, Amuluru K, Smith B, Damodara N, El-Ghanem M, Singh IP, Dangayach N, Gandhi CD. Emerging markers of early brain injury and delayed cerebral ischemia in aneurysmal subarachnoid hemorrhage. *World Neurosurg*. 2017;107:148–59.
10. Sehba FA, Hou J, Pluta RM, Zhang JH. The importance of early brain injury after subarachnoid hemorrhage. *Prog Neurobiol*. 2012;97(1):14–37.
11. Cambj-Sapunar L, Yu M, Harder DR, Roman RJ. Contribution of 5-hydroxytryptamine 1b receptors and 20-hydroxyeicosatetraenoic acid to fall in cerebral blood flow after subarachnoid hemorrhage. *Stroke*. 2003;34(5):1269–75.
12. Roman RJ, Renic M, Dunn KMJ, Takeuchi K, Haccin-Bey L. Evidence that 20-HETE contributes to the development of acute and delayed cerebral vasospasm. *Neurol Res*. 2006;28(7):738–49.
13. Qu Y-Y, Yuan M-Y, Liu Y, Xiao X-J, Zhu Y-L. The protective effect of epoxyeicosatrienoic acids on cerebral ischemia/reperfusion

- injury is associated with PI3K/AKT pathway and ATP-sensitive potassium channels. *Neurochem Res.* 2015;40(1):1–14.
14. Siler DA, Berlow YA, Kukino A, Davis CM, Nelson JW, Grafe MR, Ono H, Cetas JS, Pike M, Alkayed NJ. Soluble epoxide hydrolase in hydrocephalus, cerebral edema, and vascular inflammation after subarachnoid hemorrhage. *Stroke.* 2015;46(7):1916–22.
  15. Sudhahar V, Shaw S, Imig JD. Epoxyeicosatrienoic acid analogs and vascular function. *Curr Med Chem.* 2010;17(12):1181–90.
  16. Martini RP, Ward J, Siler D, Eastman JM, Nelson J, Borkar R, Alkayed N, Dogan A, Cetas J. Genetic variation in soluble epoxide hydrolase is associated with outcome after aneurysmal subarachnoid hemorrhage. *J Neurosurg.* 2014;121(6):1359–66.
  17. Siler DA, Martini RP, Ward JP, et al. Protective role of P450 epoxyeicosanoids in subarachnoid hemorrhage. *Neurocrit Care.* 2015;22(2):306–19.
  18. Sánchez-Porras R, Zheng Z, Santos E, Schöll M, Unterberg AW, Sakowitz OW. The role of spreading depolarization in subarachnoid hemorrhage. *Eur J Neurol.* 2013;20(8):1121–7.
  19. Dreier JP, Drenkhahn C, Woitzik J, et al. Spreading ischemia after aneurysmal subarachnoid hemorrhage. *Acta Neurochir Suppl.* 2013;115:125–9.
  20. Ayata C, Lauritzen M. Spreading depression, spreading depolarizations, and the cerebral vasculature. *Physiol Rev.* 2015;95(3):953–93.
  21. Hamming AM, Wermer MJ, Umesh Rudrapatna S, et al. Spreading depolarizations increase delayed brain injury in a rat model of subarachnoid hemorrhage. *J Cereb Blood Flow Metab.* 2016;36(7):1224–31.
  22. Cetas JS, McFarlane R, Kronfeld K, Smitasin P, Liu JJ, Raskin JS. Brainstem opioidergic system is involved in early response to experimental SAH. *Transl Stroke Res.* 2015;6(2):140–7.
  23. Sugawara T, Ayer R, Jadhav V, Zhang JH. A new grading system evaluating bleeding scale in filament perforation subarachnoid hemorrhage rat model. *J Neurosci Methods.* 2008;167(2):327–34.
  24. Prunell GF, Mathiesen T, Diemer NH, Svendgaard N-A. Experimental subarachnoid hemorrhage: subarachnoid blood volume, mortality rate, neuronal death, cerebral blood flow, and perfusion pressure in three different rat models. *Neurosurgery.* 2003;52(1):165–76.
  25. Prunell GF, Mathiesen T, Svendgaard N-A. A new experimental model in rats for study of the pathophysiology of subarachnoid hemorrhage. *Neuroreport.* 2002;13(18):2553.
  26. Fujii M, Yan J, Rolland WB, Soejima Y, Caner B, Zhang JH. Early brain injury, an evolving frontier in subarachnoid hemorrhage research. *Transl Stroke Res.* 2013;4(4):432–46.
  27. Balbi M, Koide M, Wellman GC, Plesnila N. Inversion of neurovascular coupling after subarachnoid hemorrhage in vivo. *J Cereb Blood Flow Metab.* 2017;37(11):3625–34.
  28. Wang J, Alotaibi NM, Akbar MA, Ayling OGS, Ibrahim GM, Macdonald RL. Loss of consciousness at onset of aneurysmal subarachnoid hemorrhage is associated with functional outcomes in good-grade patients. *World Neurosurg.* 2017;98:308–13.
  29. Crago EA, Thampatty BP, Sherwood PR, Kuo C-WJ, Bender C, Balzer J, Horowitz M, Poloyac SM. CSF 20-HETE is associated with delayed cerebral ischemia and poor outcomes after aneurysmal subarachnoid hemorrhage. *Stroke J Cereb Circ.* 2011;42(7):1872–7.
  30. Donnelly MK, Crago EA, Conley YP, Balzer JR, Ren D, Ducruet AF, Kochanek PM, Sherwood PR, Poloyac SM. 20-HETE is associated with unfavorable outcomes in subarachnoid hemorrhage patients. *J Cereb Blood Flow Metab.* 2015;35(9):1515–22.
  31. Geng H-X, Li R-P, Li Y-G, Wang X-Q, Zhang L, Deng J-B, Wang L, Deng J-X. 14,15-EET suppresses neuronal apoptosis in ischemia-reperfusion through the mitochondrial pathway. *Neurochem Res.* 2017;42(10):2841–9.
  32. Yuan L, Liu J, Dong R, Zhu J, Tao C, Zheng R, Zhu S. 14,15-epoxyeicosatrienoic acid promotes production of brain derived neurotrophic factor from astrocytes and exerts neuroprotective effects during ischaemic injury. *Neuropathol Appl Neurobiol.* 2016;42(7):607–20.
  33. Donnelly MK, Conley YP, Crago EA, Ren D, Sherwood PR, Balzer JR, Poloyac SM. Genetic markers in the EET metabolic pathway are associated with outcomes in patients with aneurysmal subarachnoid hemorrhage. *J Cereb Blood Flow Metab.* 2015;35(2):267–76.
  34. Moussouttas M, Huynh TT, Khoury J, Lai EW, Dombrowski K, Pello S, Pacak K. Cerebrospinal fluid catecholamine levels as predictors of outcome in subarachnoid hemorrhage. *Cerebrovasc Dis.* 2012;33(2):173–81.
  35. Ogura T, Satoh A, Ooigawa H, et al. Characteristics and prognostic value of acute catecholamine surge in patients with aneurysmal subarachnoid hemorrhage. *Neurol Res.* 2012;34(5):484–90.
  36. Hall CN, Reynell C, Gesslein B, Hamilton NB, Mishra A, Sutherland BA, O'Farrell FM, Buchan AM, Lauritzen M, Attwell D. Capillary pericytes regulate cerebral blood flow in health and disease. *Nature.* 2014;508(7494):55–60.
  37. Piilgaard H, Lauritzen M. Persistent increase in oxygen consumption and impaired neurovascular coupling after spreading depression in rat neocortex. *J Cereb Blood Flow Metab.* 2009;29(9):1517–27.
  38. Bacyinski A, Xu M, Wang W, Hu J. The paravascular pathway for brain waste clearance: current understanding, significance and controversy. *Front Neuroanat.* 2017;11:101. <https://doi.org/10.3389/fnana.2017.00101>.
  39. Goulay R, Flament J, Gauberti M, et al. Subarachnoid hemorrhage severely impairs brain parenchymal cerebrospinal fluid circulation in nonhuman primate. *Stroke.* 2017;48(8):2301–5.
  40. Iliff JJ, Nedergaard M. Is there a cerebral lymphatic system? *Stroke.* 2013;44(6 suppl 1):S93–5.
  41. Gaberel T, Gakuba C, Goulay R, De Lizarrondo SM, Hanouz J-L, Emery E, Touze E, Vivien D, Gauberti M. Impaired glymphatic perfusion after strokes revealed by contrast-enhanced MRI. *Stroke.* 2014;45(10):3092–6.
  42. Luo C, Yao X, Li J, et al. Paravascular pathways contribute to vasculitis and neuroinflammation after subarachnoid hemorrhage independently of glymphatic control. *Cell Death Dis.* 2016;7(3):e2160.
  43. Siler DA, Gonzalez JA, Wang RK, Cetas JS, Alkayed NJ. Intracisternal administration of tissue plasminogen activator improves cerebrospinal fluid flow and cortical perfusion after subarachnoid hemorrhage in mice. *Transl Stroke Res.* 2014;5(2):227–37.