Chapter 3 The Evolving Role of Mammalian Target of Rapamycin (mTOR) Inhibitors in Renal Cell Carcinoma

Carlyn C. Tan, Robert A. Figlin, and Andrew E. Hendifar

Abstract Over the last decade, treatment for metastatic renal cell carcinoma (mRCC) has dramatically improved. Previously limited to minimally effective immunotherapies such as interleukin-2 and interferon-alfa, the management of mRCC has been transformed by targeted therapies including two mammalian target of rapamycin (mTOR) inhibitors, four multi-targeted tyrosine kinase inhibitors, and one antivascular endothelial growth factor (VEGF) monoclonal antibody. Overall survival in the advanced disease setting has improved to over 2 years. Current available treatments have provided a framework on which to build the next generation of medications. Numerous novel inhibitors targeting various components of the mTOR pathway are currently being developed with many showing promising antitumor activity. The future success of mRCC treatment will likely involve a combination of agents targeting multiple pathways involved in cellular proliferation, migration, and angiogenesis. In addition, the development of genetic, immunologic, and other predictive biomarkers will allow for better patient selection and rational combination.

3.1 Introduction

Renal cell carcinoma (RCC) is the most common form of kidney cancer [1, 2]. Annually, there are approximately 209,000 new cases and 102,000 associated deaths worldwide with incidence rising by 2 % each year [3–7]. RCC is the seventh leading malignancy in men and the ninth most common malignancy in women [1, 3].

C.C. Tan

R.A. Figlin, MD, FACP (🖂)

A.E. Hendifar Hematology and Oncology, Cedars-Sinai Medical Center, LA, USA e-mail: andrew.hendifar@cshs.org

© Springer-Verlag France 2016 M. Mita et al. (eds.), *mTOR Inhibition for Cancer Therapy: Past, Present and Future*, DOI 10.1007/978-2-8178-0492-7_3

Hematology and Oncology, Fox Chase Cancer Center, Philadelphia, PA

Hematology and Oncology, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA e-mail: robert.figlin@cshs.org

Approximately 25–30 % of patients have metastatic renal cell carcinoma (mRCC) at the time of diagnosis [2, 5, 6, 8]. Twenty to 50 % of patients develop advanced disease within 1–3 years after surgery [4, 8]. Patients with mRCC at the time of diagnosis have an estimated 5-year survival rate of approximately 10 %, underscoring the need for improved treatment strategies [9, 10].

RCC is generally resistant to traditional chemotherapy and radiotherapy. In the past, improvements in overall survival were observed with interleukin 2 (IL-2) and interferon-alfa (IFN- α) [11–14]. Treatment with high-dose IL-2 demonstrated antitumor activity with durable complete responses in 7–10 % of patients [9, 12, 13]. IFN- α also led to a modest improvement in clinical outcome compared to supportive drugs such as medroxyprogesterone [14]. However, the small clinical benefit of IL-2 and IFN- α is achieved at the expense of significant toxicities.

Improved understanding of the pathogenesis of RCC has led to the development of a number of novel targeted therapies. Many of these new drugs control tumor growth by altering angiogenic pathways. In 2005, sorafenib was the first vascular endothelial growth factor receptor/platelet-derived growth factor receptor (VEGFR/ PDGFR)-targeted tyrosine kinase inhibitor (TKI) to be approved by the United States Food and Drug Administration (FDA) for improved progression-free survival (PFS) and overall survival (OS) in patients with advanced clear cell RCC resistant to standard therapy [15, 16]. Other approved TKIs include sunitinib, pazopanib, and axitinib [17-19]. Both sunitinib and pazopanib are National Comprehensive Cancer Network (NCCN) category 1 options for first-line therapy in patients with relapsed or medically unresectable predominantly clear cell stage IV RCC [20]. Axitinib is the newest TKI, approved in 2012 for patients with advanced RCC who had failed/progressed on one prior systemic therapy based on results of the AXIS trial [19]. An anti-VEGF monoclonal antibody, bevacizumab in combination with IFN- α , is also another first-line treatment option in patients with advanced clear cell RCC after demonstrating improved PFS and response in comparison with IFN- α plus placebo [20–24].

Advances in our understanding of signaling pathways in RCC have led to the development of a second mechanistic class. The mammalian target of rapamycin (mTOR) pathway is critical to cellular processes such as proliferation, growth, metabolism, and angiogenesis, which prompted the development and exploration of mTOR inhibitors for cancer therapy. Many of these agents, including temsirolimus and everolimus, inhibit only mTOR complex 1 (mTORC1), one of the two mTOR complexes that control cellular growth in response to environmental signals. Temsirolimus, a parenteral formulation, received FDA approval in 2007 for the treatment of advanced RCC after demonstrating improved PFS, OS, and response in comparison with IFN- α [25]. Everolimus, an orally active agent, was approved in 2009 for the treatment of patients with advanced RCC after failure of treatment with sunitinib or sorafenib [26]. These mTORC1 inhibitors have demonstrated survival benefits for patients with mRCC and have validated the importance of the phosphoinositide 3-kinase (PI3K)/Akt/mTOR pathway in the pathogenesis of RCC. This chapter will review the evolution of mTOR inhibitors in the field of renal cell carcinoma and future targets for therapy.

3.2 The Role of mTOR in Renal Cell Carcinoma

mTOR is a serine/threonine kinase involved in the PI3K/Akt signaling pathway that regulates cell growth and metabolism in response to environmental factors. The PI3K/Akt/mTOR pathway is dysregulated in many cancers and plays a critical role in RCC tumorigenesis [27–29]. In RCC tumors, activation of this pathway has been shown to correlate with aggressive behavior and poor prognostic features [27, 30, 31].

By integrating input from growth factors such as VEGF, insulin growth factor (IGF), and endothelial growth factor (EGF), hormones, and nutrients, mTOR activates protein synthesis and contributes to numerous critical cellular functions, including protein synthesis/degradation and angiogenesis. The mTOR response to growth factors and nutrients is directly controlled by PI3K/Akt. Growth factors activate PI3K through various receptor tyrosine kinases. PI3K subsequently stimulates activation of Akt, which leads to phosphorylation of tuberous sclerosis tumor complex 2 (TSC2) and the inactivation of the TSC1-2 complex, a key regulator of mTOR [32, 33]. Inactivation of the TSC1-2 complex then leads to activation of mTOR. Overactive mTOR signaling can occur through various mechanisms, including overexpression or activation of growth factor receptors (VEGFR, IGF-1R, EGFR) or decreased expression of *TCS1/2*, *PTEN*, or von Hippel-Lindau (*VHL*) tumor suppressor genes [34, 35].

Structurally mTOR exists as two distinct protein complexes, mTORC1 and mTORC2. mTORC1 is involved in rapamycin-sensitive control of cell growth and is activated by Akt through inhibition of TSC2 and by regulation of cellular energy. mTORC1 stimulates protein synthesis through the phosphorylation of p70 ribosomal protein S6 kinase (p70S6K) and 4E-binding-protein 1 (4E-BP1). Activated p70S6K phosphorylates the 40S ribosomal protein S6 kinase which causes promotion of mRNA translation, stimulation of protein synthesis, and entrance into the G1 phase of the cell cycle [36]. mTORC1 mediates the downstream inhibitory phosphorylation of 4E-BP1. 4E-BP1 is subsequently unable to inactivate the translation/initiation factor, eukaryotic translation initiation factor 4E (eIF4E). As a result, active eIF4E is able to associate with eIF4G to form an active eIF4F complex, a key component of protein synthesis. eIF4F complex is particularly important for the translation of 5'capped mRNA, including *VEGF*, *cyclin D*, *c-Myc*, and *survivin* [37, 38].

mTORC2 is involved in rapamycin-insensitive control of cell growth. Both mTORC1 and mTORC2 increase hypoxia-inducible factor (HIF-1 α) gene expression at the level of mRNA translation and protein stabilization [39, 40]. mTORC2, on the other hand, also controls expression of HIF-2 α [41, 42]. HIF-1 α and HIF-2 α activate transcription of genes that regulate angiogenesis, proliferation, and invasion as well as factors important for responding to hypoxic and stressful conditions, such as VEGF and glycolytic enzymes [39, 40]. Upregulation of these factors is critical to the pathogenesis of RCC.

Most mRCC tumors exhibit dysregulated genes that alter or depend on mTOR activity for their pathology [43]. Up to 60 % of sporadic clear cell RCC contain altera-

tion of the *VHL* tumor suppressor gene [44]. The primary function of VHL protein is to target HIF- α for degradation. *VHL* mutation or gene silencing leads to increased HIF levels and is considered to have a critical role in tumorigenesis [45]. Activated mTOR activity increases synthesis of HIF, whereas inhibition of mTOR has been shown to reduce expression of HIF in mouse xenograft models [45]. This suggests that mTOR plays a critical role in RCC pathogenesis. In addition, *PTEN* gene expression has been shown to be downmodulated in a large percentage of RCC tumors [46]. Lack of *PTEN* expression leads to increased activity of the PI3K/Akt/mTOR pathway and is a prognostic indicator of poor survival in mRCC patients [47]. A tissue microarray-based analysis of the mTOR pathway in RCC has shown predictive and prognostic relevance [30]. Specifically, the expression of p70S6K was significantly higher in mRCC patients and was found to be a strong predictor of survival in localized and metastatic RCC [30]. Although the baseline activity of the mTOR pathway is associated with RCC pathogenesis and poor prognostic features of RCC tumors.

3.3 Development of Novel mTORC1-Targeted Therapies

mTORC1 inhibitors are structural derivatives of the macrocyclic lactone rapamycin or sirolimus. Found to have fungicidal, immunosuppressive, and antiproliferative properties, sirolimus was initially approved in 1999 as an immunosuppressant for solid organ transplants. Preclinical data showed promising results in tumor cell models; phase I and II trials showed that sirolimus reduced the size of angiomyolipomas in patients with tuberous sclerosis complex and lymphangioleiomyomatosis [48, 49]. Temsirolimus was the first mTORC1 inhibitor approved in 2007 for the treatment of advanced RCC. Everolimus was initially developed in the organ transplant setting but was approved in 2009 for the treatment of advanced RCC patients who had failed treatment with sunitinib or sorafenib.

Temsirolimus and everolimus inhibit mTOR by binding to the cytosolic protein FKBP-12. The resulting protein-drug complex inhibits mTOR through allosteric binding to the FKBP12-rapamycin binding domain adjacent to the catalytic site of mTOR [50, 51]. The protein-drug complex is only able to bind mTORC1 and is unable to inhibit mTORC2. The inhibition of mTORC1 pathway prevents protein synthesis, cellular growth and proliferation, and angiogenesis, thereby arresting the cells in the G1 phase of the cell cycle.

3.3.1 Temsirolimus

Temsirolimus was first identified to have antitumor activity by the Developmental Therapeutic Branch of the National Cancer Institute [52]. It is an inactive soluble ester with low oral bioavailability. As an intravenous (IV) formulation, temsirolimus acts as a prodrug that is metabolized to the active compound sirolimus. Temsirolimus exploits the antitumor properties of sirolimus with improved pharmacokinetics. In preclinical models, temsirolimus exhibited antitumor activity by normalizing p70S6K activity and reducing proliferation of murine xenografts in a variety of cancers, including glioma, rhabdomyosarcoma, medulloblastoma, and prostate and breast cancer [53–56]. A phase I study in patients with advanced solid tumors identified weekly temsirolimus 25, 75, and 250 mg IV as appropriate doses for further clinical testing [57]. Dose-limiting toxicities (DLTs) included thrombocytopenia, acneiform rash, stomatitis, and mucositis which all resolved after discontinuation of therapy [57]. In this study of 24 patients, confirmed partial responses were observed in two patients with mRCC and breast cancer. Of note, the patient with mRCC had documented progression with prior IL-2 and IFN- α therapy [57].

A phase II study enrolled 111 patients with advanced refractory RCC who were treated with temsirolimus 25, 75, and 250 mg IV weekly [58]. Antitumor activity was observed in all dosing levels, and treatment was generally well tolerated [58]. Since no major differences were observed in terms of toxicity or measurable efficacy between the three dosing levels, a 25 mg weekly dosage was selected for further evaluation. A multicenter Global Advanced Renal Cell Carcinoma (ARCC) phase III study randomized 626 treatment-naïve patients identified to have poor-risk features to one of three arms: (1) temsirolimus 25 mg IV weekly, (2) temsirolimus 15 mg IV weekly plus IFN- α 6 million units three times weekly, or (3) IFN- α 3 million units with increase to 18 million units subcutaneously three times weekly [25]. Poor-risk features are defined in Table 3.1. This study demonstrated that temsirolimus 25 mg IV weekly prolonged PFS and OS compared to IFN-α (3.8 months vs 1.9 months for PFS; 10.9 months vs 7.3 months for OS, respectively) [25]. Based on these results, IV temsirolimus was approved in 2007 for patients with advanced RCC. Guidelines recommend temsirolimus as first-line treatment for mRCC patients with poor-risk features [20, 22–24].

3.3.2 Everolimus

Prior to reports of antitumor activity, everolimus was studied extensively in the setting of cardiac and renal transplantation. Antitumor effects were initially demonstrated in a rat pancreatic tumor model [59]. A single dose of everolimus was shown

Table 3.1 Poor-risk criteria [20, 25]	1. Serum LDH>1.5 times the upper limit of normal
	2. Hemoglobin level < lowest limit of normal
	3. Corrected serum calcium level >10 mg/dL (2.5 mmol/L)
	4. Interval of less than 1 year from initial diagnosis of RCC to start of systemic therapy
	5. Karnofsky performance score \leq 70
	6. ≥2 sites of organ metastases

to block phosphorylation of 4E-BP1 and inactivate S6K1 in human peripheral blood mononuclear cells [59]. Everolimus is orally bioavailable with no active metabolites. A phase I dose escalation study demonstrated that everolimus was well tolerated at doses up to 70 mg weekly and 10 mg daily [60]. DLTs included hyperglycemia, stomatitis, and fatigue [60]. Partial responses were observed in 4 patients, and 12 patients remained progression-free for ≥ 6 months, including 5 of 10 patients with RCC [60]. Other phase I pharmacokinetic/pharmacodynamics studies showed that continuous daily dosing with everolimus 10 mg resulted in a more sustained targeted inhibition of mTOR than that achieved with a weekly dosage schedule [61, 62]. As a result, a daily dose of 10 mg was selected for further trials with everolimus.

A phase II study involving patients with mRCC, who had received at most one prior therapy other than an mTOR inhibitor, demonstrated the antitumor activity of everolimus 10 mg daily with reported median PFS and OS of 11.2 months and 22.1 months, respectively [63]. The pivotal phase III RECORD-1 trial examined the role of everolimus in patients with clear cell mRCC who had received prior sorafenib and/or sunitinib. This international study demonstrated that everolimus 10 mg daily resulted in a median PFS of 4.9 months compared to 1.9 months with placebo [26, 64]. Pharmacodynamic modeling of tumor growth in the RECORD-1 patient population showed that compared to placebo, everolimus 5 and 10 mg daily significantly slowed growth of mRCC target lesions, nontarget lesions, and new metastases; the 10 mg daily dosing was more effective than 5 mg daily in reducing growth of target lesions [65]. Based on results from the RECORD-1 study, oral everolimus was approved in the USA for patients with mRCC who had failed treatment with sunitinib or sorafenib and in Europe for patients who progressed on or after treatment with VEGF-targeted therapy [20, 22–24]. Although everolimus is well established as a second-line agent, its role as a first-line option is currently under investigation. The RECORD-3 trial is a phase II study investigating first-line everolimus followed by sunitinib versus standard sequence. Preliminary data demonstrated that PFS noninferiority was not achieved with first-line everolimus when compared with sunitinib, supporting the current standard treatment paradigm [66].

3.4 Safety Considerations with mTORC-1 Inhibitors in Renal Cell Carcinoma

mTORC1 inhibitors are commonly associated with disorders of metabolism, noninfectious pneumonitis and stomatitis. Hyperglycemia and hypercholesterolemia are common although the severity is generally mild. Noninfectious pneumonitis has been recognized as a class effect of mTORC1 inhibitors. A follow-up study of patients treated with temsirolimus in the ARCC trial identified four cases of pneumonitis with one patient progressing from grade 3 to 5 toxicity [67]. The RECORD-1 trial reported that 14 % of patients treated with everolimus developed noninfectious pneumonitis [64]. Among ten patients who developed grade 3 noninfectious pneumonitis, eight had clinical resolution with steroid therapy. A review of these cases suggests that noninfectious pneumonitis can be managed effectively with early recognition and prompt intervention [68]. The use of imaging studies to monitor patients can be particularly challenging since radiographic abnormalities are seen in a higher percentage of patients receiving mTORC1 inhibitors compared to placebo in the absence of symptoms or a clinical diagnosis of pneumonitis [63, 67, 69]. Patients receiving mTORC1 inhibitors should be monitored closely for signs and symptoms of respiratory illness. Mild stomatitis and rash occurred in more than 20 % of patients in both the ARCC and RECORD-1 trials [25, 64]. These toxicities are manageable with standard supportive measures.

3.5 Limitations of mTORC1-Targeted Therapy

Although mTORC1 inhibitors produce clinically meaningful responses with improved PFS and OS, these responses are short-lived, and rarely do these therapies induce complete responses. None of the current available mTORC1 inhibitors have been able to induce sustained disease remission. Many patients initially respond but eventually relapse usually due to the development of resistance after a median of 6-15 months of treatment. These acquired mechanisms of resistance to mTORC1 inhibitors lead to reestablishment of tumor vasculature [70, 71]. They are thought to be facilitated through activation of alternative or compensatory pathways that lead to upregulation of various factors that promote cell growth and survival, including HIF. Potential mechanisms include transient and partial inhibition of 4E-BP1 and loss of negative feedback loops that are normally induced when mTORC1/p70S6K is active. The phosphorylation of 4E-BP1 has been shown to be less responsive to rapalogs than that of p70S6K. Although rapamycin inhibits the functions of p70S6K and 4E-BP1 in the short term, prolonged treatment renders mTORC1 to be rapamycin-resistant toward 4E-BP1 resulting in reinitiation of cap-dependent translation of mRNAs despite continued mTORC1 inhibition [72]. Findings by Choo et al. also suggest that catalytic inhibitors of mTOR, including a dual PI3K and mTOR inhibitor, were more effective than rapamycin in dephosphorylating 4E-BP1, supporting their clinical promise [72].

Recent data suggest loss of negative feedback loops from inhibition of mTORC1 leads to compensatory activation of PI3K and Akt which drives resistance via upregulation of mTORC2 [73]. Activation of S6K through mTORC1 phosphorylation results in phosphorylation of rictor, which prevents mTORC2 activation [74, 75]. If mTORC1/S6K is inhibited, the negative feedback is lost leading to derepression of mTORC2 and mTORC2-mediated phosphorylation and activation of Akt [76]. Activation of mTORC2 also leads to upregulation of HIF-2 α which has been argued to be the more relevant HIF with respect to the development and progression of RCC. HIF-2 α activation has been shown to strongly suppress E-cadherin expression, allowing for increased cell motility [77]. E-cadherin loss is frequently associated with tumor progression and metastasis [78]. These findings highlight the potential therapeutic advantage of simultaneous inhibition of mTORC1 and mTORC2 in preventing tumor cell proliferation, growth, invasion, and metastasis.

Another potential mechanism of resistance is the loss of a negative feedback loop that normally prevents upstream overstimulation of insulin receptor substrate 1 (IRS1)/PI3K/Akt signaling [79]. mTORC1 activation of S6K causes destabilization of IRS1-2 which uncouples IGF-1 from the PI3K/Akt pathway. Normally, IGF-1 binds IGFR which in turn phosphorylates substrates IRS1-2 which then relays the activation to PI3K. mTORC1/S6K inhibition results in the loss of this feedback loop and leads to the upregulation of IRS1 protein and activation of the PI3K/Akt cascade [80]. PI3K/Akt signaling activates an array of kinases that promote cell growth and survival. This prosurvival effect occurs through various pathways including negative regulation of factors that promote expression of death genes, positive regulation of proapoptotic proteins, and regulation of the cell cycle [81].

3.6 Future Directions and Novel Therapies

Because of their suspected roles in resistance to mTORC1 inhibitors, PI3K, Akt, and mTORC2 are potential targets for the development of novel therapies for various malignancies, including mRCC. Consistent with their proposed roles in the development of resistance and pathogenesis of mRCC, a microarray analysis of RCC tissue specimens showed that high PI3K and mTOR expression levels corresponded with late-stage, high-grade tumors and were prognostic factors for decreased survival [82]. A number of PI3K, mTORC1/2, and Akt inhibitors have been developed and have demonstrated promising results in RCC cell lines and xenograft models. This section will focus on these novel targeted agents that have been evaluated in RCC (Table 3.2).

3.6.1 mTORC1/2 Inhibitors

Novel mTORC1/2 inhibitors bind directly to the adenosine triphosphate (ATP)binding domain of mTOR, resulting in the inhibition of both mTORC1 and mTORC2 (Table 3.2) [83, 87, 88, 92]. These mTOR kinase inhibitors prevent the rebound activation of PI3K/Akt cascade as seen with rapalogs. An mTORC1/2 inhibitor can also prevent HIF-2 α suppression of E-cadherin expression and result in restored cell-cell adhesion to prevent tumor cell motility and migration [77]. INK128/ MLN0128 is a highly potent, orally active mTOR kinase ATP-competitive inhibitor that is currently being investigated in RCC cell lines [83]. Preclinical data suggest that it has antitumor and antimetastatic activity in prostate cancer models as well as synergistic activity with TKI lapatinib in breast cancer models refractory to anti-HER2 therapy [84, 85]. INK128/MLN0128 has been shown to inhibit downstream substrates of mTOR, phosphorylation of Akt, and tumor cell proliferation as well as induce G1 cell cycle arrest [86]. INK128/MLN0128 demonstrated antitumor

			Phase of		
Agent	Target	Formulation	development	Drug company	Reference
INK128/MLN0128	mTORC1/2	Oral	Phase I	Intellikine	[83–86]
WYE-125132	mTORC1/2	Oral	Preclinical	Wyeth	[87]
AZD8055	mTORC1/2	Oral	Phase I	AstraZeneca	[88–91]
Ku0063794	mTORC1/2	Intravenous	Preclinical	Kudos Pharmaceuticals	[92]
NVP-BEZ235	PI3K/mTORC1/2	Oral	Phase I/II	Novartis	[93-100]
SF1126	PI3K/mTORC1/2	Oral	Phase I	Semaphore Pharmaceuticals	[101-103]
BKM120	PI3K	Oral	Phase I/II	Novartis	[104–106]
Perifosine	Akt	Oral	Phase II	Keryx Biopharmaceuticals	[107-110]
MK2206	Akt	Oral	Phase II	Merck	[111-116]

 Table 3.2
 Novel agents targeting PI3K/Akt and mTOR pathways in development

activity in RCC mouse models which was further enhanced in combination with sorafenib or bevacizumab. The combination resulted in a sustained regression of the tumor through inhibition of tumor cell proliferation by INK128/MLN0128 and angiogenesis by sorafenib/bevacizumab [83]. These findings suggest that combination therapy may be an option for maximizing therapeutic benefits of novel agents in the treatment of mRCC.

WYE-125132 is a pyrazolopyrimidine molecule that acts as an orally active, highly potent, ATP-competitive and specific mTOR kinase inhibitor. It has demonstrated antitumor activity in RCC cell lines and mouse models resulting in strong G1 phase arrest and tumor growth suppression [87]. Combination of WYE-125132 and bevacizumab caused dramatic tumor regression of large A498 tumors [87]. Unlike rapalogs, WYE-125132 was able to disrupt cap-dependent translation initiation eIF4F complex; after treatment with the molecule, there was a drastic increase in the inhibitory binding of 4E-BP1 to eIF4E with almost complete loss of eIF4G. WYE-125132 also strongly inhibited hypoxia-induced accumulation of HIF-1 α and HIF-2 α [87].

AZD8055 is a third potent, orally active, highly selective mTORC1/2 inhibitor. Preclinical data show that it is better at inhibiting phosphorylation of 4E-BP1 than rapamycin, resulting in significant inhibition of cap-dependent translation [88, 89]. It was also able to inhibit Akt in MCF-7 breast carcinoma cells where rapamycin treatment resulted in rebound activation of Akt [88, 89]. Chresta et al. demonstrated that AZD8055 potently inhibits cellular proliferation and induces autophagy in vitro with H838 and A549 cells. In vivo, AZD8055 induced significant tumor growth inhibition and regression in a variety of human tumor types, including breast, lung, colon, prostate, and uterine xenograft models [89]. Recent data suggest that AZD8055 has significant antitumor activity against clear cell RCC cell lines UOK-139 and UOK-140 [90]. AZD8055 is currently undergoing clinical evaluation in phase I trials. Naing et al. reported a maximum tolerated dose of 90 mg PO BID. DLTs included grade 3 transaminitis (increased alanine aminotransferase 22 %, increased aspartate aminotransferase 22 %) and fatigue (16 %) [91]. Transaminitis was reversible in all patients, except for one with liver metastases. AZD8055 was overall well tolerated, but no complete or partial responses were observed [91].

Ku0063794 is another highly specific small molecular inhibitor of mTOR kinase. It has been shown to inhibit phosphorylation of S6K and 4E-BP1 as well as Akt phosphorylation [92]. Ku0063794 has been compared with temsirolimus in preclinical RCC models. It was found to be more effective than temsirolimus in decreasing viability and growth of RCC cell lines in vitro by inducing cell cycle arrest and autophagy, but not apoptosis [92]. However, in xenograft models, there was no difference in the inhibition of tumor growth by Ku0063794 or temsirolimus [92]. A potential explanation is that temsirolimus has additional effects on tumor microenvironment, including decreasing tumor angiogenesis. VEGF and PDGF expression was lower in cells treated with temsirolimus than in cells treated with Ku0063794 [92]. This observation suggests that mTORC1/2 inhibitors may provide better tumor suppression and regression in combination with anti-angiogenic agents.

3.6.2 PI3K/mTOR Inhibitors

Because the catalytic domain of mTOR and p110a subunit of PI3K is structurally similar, multiple agents have been developed to have dual inhibitory activity against PI3K and mTORC1/2 (Table 3.2) [82, 93]. These ATP-competitive, pan-selective inhibitors of PI3K and mTOR have demonstrated impressive antitumor activity in a wide range of tumor models. NVP-BEZ235 is a potent orally available imidazoquinoline dual PI3K/mTOR inhibitor. It reversibly inhibits class 1 PI3K activity by binding to its ATP-binding domain [94]. It also directly binds to the mTOR ATPbinding domain and inhibits its catalytic activity. In preclinical studies, NVP-BEZ235 has been shown to inhibit PI3K and mTOR activity resulting in tumor growth suppression in numerous human tumor models, including glioblastoma, multiple myeloma, and prostate, breast, and pancreatic carcinoma [95-97]. A comparison of NVP-BEZ235 and rapamycin activity in RCC xenografts revealed that NVP-BEZ235 is significantly more effective at downmodulating cyclin D, survivin, and HIF-2 α than rapamycin. It was also more effective at inhibiting tumor growth both in vitro and in vivo through antiproliferative and proapoptotic effects [93]. A study with RCC cell lines 786-O and Caki-1 demonstrated that the combination of NVP-BEZ235 and sorafenib had greater antitumor activity through reduction of tumor cell growth and increasing apoptosis than either agents alone [98]. This finding suggests that dual PI3K/mTOR inhibitor in combination with an anti-angiogenic agent may result in enhanced synergistic antitumor activity. A phase I clinical trial with advanced solid tumors showed that BEZ235 is generally well tolerated with a favorable safety profile [99]. The most commonly reported adverse events included nausea, vomiting, diarrhea, fatigue/asthenia, and anorexia. Available pharmacodynamics and efficacy data also showed that NVP-BEZ235 is active, especially in patients with PI3K pathway dysregulated tumors [99]. Another phase I study with a new formulation of NVP-BEZ235 using a solid dispersion system (SDS) sachet included three RCC patients and showed that this specific formulation was well tolerated [100]. Common adverse events included nausea, vomiting, diarrhea, and fatigue/asthenia. The SDS sachet formulation of NVP-BEZ235 has been chosen for further evaluation in phase II clinical trials [100].

Another pan-PI3K/mTORC inhibitor SF1126 is a prodrug of LY294002 administered intravenously. The active LY294002 has significant antitumor and antiangiogenic activities in vivo, but is not a drug candidate due to insolubility and short half-life. To increase solubility and bioavailability, LY294002 is conjugated to RGD (Arg-Gly-Asp) peptide via a cleavable linker to form SF1126. In preclinical models, SF1126 exhibited both antitumor and anti-angiogenic activities [101]. In a 786-O RCC xenograft model, SF1126 demonstrated 50–90 % tumor inhibition or regression of tumor volume [101]. It has also been shown to significantly suppress signaling pathways downstream of PI3K, including Akt, and eliminate hypoxia-induced stabilization of HIF-2 α [102]. A phase I clinical trial found that SF1126 is generally well tolerated [103]. Grade 3 DLTs included peripheral edema, increased alkaline phosphatase, diarrhea, weakness, hypoglycemia, urticaria/pruritus, anemia, hypokalemia, and hypersensitivity [103]. Common adverse events included nausea, fatigue, vomiting, diarrhea, pyrexia, chills, pruritus, anemia, anorexia, and head-ache [103]. Stable disease was the best response observed with mean duration of 21 weeks (range of 8–84 weeks); 2 of the 3 RCC patients had stable disease at 14 and 84 weeks [103].

3.6.3 PI3K Inhibitors

In addition to mTORC1/2 inhibitors and dual PI3K/mTOR kinase inhibitors, PI3Kselective inhibitors are currently under investigation (Table 3.2). BKM120 is an oral pyrimidine-derived pan-PI3K inhibitor with specific and potent activity against class I PI3Ks [104, 105]. In preclinical studies, BKM120 demonstrated a strong antiproliferative effect and induced apoptosis in vitro on various human cancer cell lines [105]. In vivo, BKM120 had significant antitumor activity in U87MG glioblastoma and A2780 ovarian xenograft models [105, 106]. A phase I study showed that BKM120 is well tolerated with median treatment duration of 7.5 weeks and showed antitumor activity in 28 of 66 patients, including 2 patients with partial response and 26 with stable disease [104]. Adverse events included decreased appetite, rash, diarrhea, nausea, fatigue, hyperglycemia, anxiety, depression, and mucositis [104]. BKM120 is currently being tested in a number of clinical trials, including a phase I study in combination with bevacizumab in patients with mRCC who had failed prior systemic therapies.

3.6.4 Akt Inhibitors

Because of Akt's critical role in cellular survival and tumorigenesis, Akt inhibitors have been developed with promising results (Table 3.2). Perifosine is a synthetic, substituted heterocyclic alkylphospholipid with the ability to inhibit Akt activity [107]. It inhibits Akt activation by interfering with the interaction between the pleckstrin homology domain of Akt and phosphatidylinositol phosphate (PIP3) [107]. This interference precludes Akt's translocation to the plasma membrane where activation would have occurred through phosphorylation by pyruvate dehydrogenase kinase, isozyme 1 (PDK1). Fu et al. showed that perifosine induced autophagy and inhibited assembly of the mTOR complexes by promoting degradation of Akt, mTOR, rictor, raptor, p70S6K, and 4E-BP1 [108]. A phase I trial showed that perifosine was well tolerated with nausea, vomiting, diarrhea, and fatigue as the most commonly observed toxicities [109]. A phase II trial assessed the efficacy and safety of perifosine in patients with advanced RCC who had failed previous VEGF-targeted therapy. It demonstrated modest activity in patients with advanced RCC, but this activity was not superior to currently available second-line agents [110]. Further studies are needed on the possibility of combination therapy with perifosine for RCC.

MK-2206 is a potent orally active allosteric Akt inhibitor. It has nanomolar potency against purified recombinant human Akt1 (half maximal inhibitory concentration [IC₅₀], 5 nmol/L) and Akt2 enzymes (IC50, 12 nmol/L) but lower potency against human Akt3 (IC50, 65 nmol/L). MK-2206 inhibits phosphorylation at Thr308 and Ser473 of AKT and demonstrates greater than 100-fold selectivity of Akt against more than 200 other kinases [111]. It has in vitro and in vivo antitumor activity as a single agent and enhances preclinical activity of conventional cytotoxic chemotherapy and other targeted therapies [112, 113]. Hirai et al. demonstrated that MK-2206 synergistically inhibited cell proliferation in combination with molecular targeted agents, such as erlotinib and lapatinib as well as with standard cytotoxic agents, including doxorubicin, gemcitabine, 5-fluorouracil, docetaxel, and carboplatin in lung NCI-H460 and ovarian A2780 cells [113]. In vivo, the addition of MK-2206 exerted significantly more potent antitumor activity than each agent in the monotherapy setting [113]. A phase I clinical trial involving 33 patients with advanced solid tumors, including patients with RCC, showed that MK-2206 was well tolerated [114]. DLTs included skin rash and stomatitis. The maximum tolerated dose was established at 60 mg. Drug-related toxicities included skin rash (51.5 %), nausea (36.4 %), pruritus (24.2 %), hyperglycemia (21.2 %), and diarrhea (21.2 %) [114]. Another phase I study investigated the maximum tolerated dose, DLTs, PK,

advanced solid tumors, including patients with RCC, showed that MK-2206 was well tolerated [114]. DLTs included skin rash and stomatitis. The maximum tolerated dose was established at 60 mg. Drug-related toxicities included skin rash (51.5 %), nausea (36.4 %), pruritus (24.2 %), hyperglycemia (21.2 %), and diarrhea (21.2 %) [114]. Another phase I study investigated the maximum tolerated dose, DLTs, PK, and efficacy of MK-2206 in combination with targeted and cytotoxic agents in patients with advanced solid tumors, including patients with RCC [115]. MK-2206 with carboplatin/paclitaxel, docetaxel, or erlotinib was found to be well tolerated. DLTs included skin rash, febrile neutropenia, tinnitus, and stomatitis. Common adverse events included fatigue (68 %), nausea (49 %), rash (47 %), diarrhea (44 %), anorexia (44 %), alopecia (40 %), vomiting (36 %), stomatitis (32 %), and hyperglycemia (25 %) [115]. A recent phase II clinical trial compared MK-2206 with everolimus in patients with VEGF inhibitor refractory mRCC [116]. MK-2206 was held in three patients due to grade 3 rash, and one patient had to come off study for the rash. Median PFS for MK-2206 was 3.65 months and 7.43 months for everolimus. Two patients in the MK-2206 group demonstrated dramatic responses with greater than 50 % disease regression and PFS of 8 and 6 months. Jonasch et al. showed that monotherapy with MK-2206 was not superior to everolimus, but a dramatic response to MK-2206 was seen in a subset of patients [116]. Further translational studies analyzing genotype-phenotype correlations may help explain this observation and identify biomarkers to allow for patient selection and rational drug combination.

3.7 Conclusion

Over the past decade, the treatment of mRCC has been revolutionized by the advent of targeted therapies, specifically agents that target the VEGF and mTOR pathways. These agents have improved PFS and OS of patients with mRCC. However, they have not been able to induce long-term remission, and many patients relapse due to the evolution of resistance. Studies are investigating the interplay between RCC and

its microenvironment and analyzing novel mechanisms driving tumorigenesis and proliferation. One potential mechanism of resistance is thought to involve activation of proangiogenic transcription factor HIF through compensatory mTORC2 and PI3K/Akt signaling. Therefore, numerous inhibitors targeting mTORC1/2, PI3K, and Akt are currently being developed with many showing promising preclinical antitumor activity in RCC cell lines and xenograft models. The future success of mRCC treatment will likely involve a combination of agents targeting multiple pathways, including VEGFR, PI3K, and mTORC1/2 and the application of various biomarkers to allow for patient selection and rational combination.

References

- 1. Cohen HT, McGovern FJ. Renal-cell carcinoma. N Engl J Med. 2005;353(23):2477-90.
- Motzer RJ, Bander NH, Nanus DM. Renal-cell carcinoma. N Engl J Med. 1996; 335(12):865–75.
- 3. Rini BI, Campbell SC, Escudier B. Renal cell carcinoma. Lancet. 2009;373(9669):1119-32.
- 4. Motzer RJ, Agarwal N, Beard C, Bhayani S, Bolger GB, Carducci MA, et al. Kidney cancer. J Natl Compr Canc Netw. 2011;9(9):960–77.
- Gupta K, Miller JD, Li JZ, Russell MW, Charbonneau C. Epidemiologic and socioeconomic burden of metastatic renal cell carcinoma (mRCC): a literature review. Cancer Treat Rev. 2008;34(3):193–205.
- Hock LM, Lynch J, Balaji KC. Increasing incidence of all stages of kidney cancer in the last 2 decades in the United States: an analysis of surveillance, epidemiology and end results program data. J Urol. 2002;167(1):57–60.
- Hollingsworth JM, Miller DC, Daignault S, Hollenbeck BK. Rising incidence of small renal masses: a need to reassess treatment effect. J Natl Cancer Inst. 2006;98(18):1331–4.
- Flanigan RC, Campbell SC, Clark JI, Picken MM. Metastatic renal cell carcinoma. Curr Treat Options Oncol. 2003;4(5):385–90.
- Klapper JA, Downey SG, Smith FO, Yang JC, Hughes MS, Kammula US, et al. High-dose interleukin-2 for the treatment of metastatic renal cell carcinoma: a retrospective analysis of response and survival in patients treated in the surgery branch at the National Cancer Institute between 1986 and 2006. Cancer. 2008;113(2):293–301.
- Howlader N, Noone AM, Krapcho M, Garshell J, Neyman N, Altekruse SF, Kosary CL, et al. SEER cancer statistics review, 1975–2010. Bethesda: National Cancer Institute; 2013.
- Motzer RJ, Bacik J, Murphy BA, Russo P, Mazumdar M. Interferon-alfa as a comparative treatment for clinical trials of new therapies against advanced renal cell carcinoma. J Clin Oncol. 2002;20(1):289–96.
- 12. McDermott DF, Regan MM, Clark JI, Flaherty LE, Weiss GR, Logan TF, et al. Randomized phase III trial of high-dose interleukin-2 versus subcutaneous interleukin-2 and interferon in patients with metastatic renal cell carcinoma. J Clin Oncol. 2005;23(1):133–41.
- Fyfe G, Fisher RI, Rosenberg SA, Sznol M, Parkinson DR, Louie AC. Results of treatment of 255 patients with metastatic renal cell carcinoma who received high-dose recombinant interleukin-2 therapy. J Clin Oncol. 1995;13(3):688–96.
- Interferon-alpha and survival in metastatic renal carcinoma: early results of a randomised controlled trial. Medical Research Council Renal Cancer Collaborators. Lancet. 1999;353(9146):14–7.
- Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Siebels M, et al. Sorafenib in advanced clear-cell renal-cell carcinoma. N Engl J Med. 2007;356(2):125–34.

- 16. Escudier B, Eisen T, Stadler WM, Szczylik C, Oudard S, Staehler M, et al. Sorafenib for treatment of renal cell carcinoma: final efficacy and safety results of the phase III treatment approaches in renal cancer global evaluation trial. J Clin Oncol. 2009;27(20):3312–8.
- Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Oudard S, et al. Overall survival and updated results for sunitinib compared with interferon alfa in patients with metastatic renal cell carcinoma. J Clin Oncol. 2009;27(22):3584–90.
- Sternberg CN, Davis ID, Mardiak J, Szczylik C, Lee E, Wagstaff J, et al. Pazopanib in locally advanced or metastatic renal cell carcinoma: results of a randomized phase III trial. J Clin Oncol. 2010;28(6):1061–8.
- Rini BI, Escudier B, Tomczak P, Kaprin A, Szczylik C, Hutson TE, et al. Comparative effectiveness of axitinib versus sorafenib in advanced renal cell carcinoma (AXIS): a randomised phase 3 trial. Lancet. 2011;378(9807):1931–9.
- National Comprehensive Cancer Network. NCCN clinical practice guidelines in oncology: kidney cancer, version 2.2011. Fort Washington: National Comprehensive Cancer Network, Inc.; 2011.
- Escudier B, Bellmunt J, Négrier S, Bajetta E, Melichar B, Bracarda S, et al. Phase III trial of bevacizumab plus interferon alfa-2a in patients with metastatic renal cell carcinoma (AVOREN): final analysis of overall survival. J Clin Oncol. 2010;28(13):2144–50.
- 22. Escudier B, Kataja V, Group EGW. Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2010;21 Suppl 5:v137–9.
- 23. de Reijke TM, Bellmunt J, van Poppel H, Marreaud S, Aapro M. EORTC-GU group expert opinion on metastatic renal cell cancer. Eur J Cancer. 2009;45(5):765–73.
- 24. Ljungberg B, Cowan NC, Hanbury DC, Hora M, Kuczyk MA, Merseburger AS, et al. EAU guidelines on renal cell carcinoma: the 2010 update. Eur Urol. 2010;58(3):398–406.
- Hudes G, Carducci M, Tomczak P, Dutcher J, Figlin R, Kapoor A, et al. Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. N Engl J Med. 2007;356(22):2271–81.
- Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, et al. Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. Lancet (England). 2008;372:449–56.
- 27. Husseinzadeh HD, Garcia JA. Therapeutic rationale for mTOR inhibition in advanced renal cell carcinoma. Curr Clin Pharmacol. 2011;6(3):214–21.
- Sourbier C, Lindner V, Lang H, Agouni A, Schordan E, Danilin S, et al. The phosphoinositide 3-kinase/Akt pathway: a new target in human renal cell carcinoma therapy. Cancer Res. 2006;66(10):5130–42.
- 29. Altomare DA, Testa JR. Perturbations of the AKT signaling pathway in human cancer. Oncogene. 2005;24(50):7455–64.
- Pantuck AJ, Seligson DB, Klatte T, Yu H, Leppert JT, Moore L, et al. Prognostic relevance of the mTOR pathway in renal cell carcinoma: implications for molecular patient selection for targeted therapy. Cancer. 2007;109(11):2257–67.
- 31. Darwish OM, Kapur P, Youssef RF, Bagrodia A, Belsante M, Alhalabi F, et al. Cumulative number of altered biomarkers in mammalian target of rapamycin pathway is an independent predictor of outcome in patients with clear cell renal cell carcinoma. Urology. 2013;81(3):581–6.
- 32. Fingar DC, Blenis J. Target of rapamycin (TOR): an integrator of nutrient and growth factor signals and coordinator of cell growth and cell cycle progression. Oncogene. 2004;23(18):3151–71.
- 33. Potter CJ, Pedraza LG, Xu T. Akt regulates growth by directly phosphorylating Tsc2. Nat Cell Biol. 2002;4(9):658–65.
- Meric-Bernstam F, Gonzalez-Angulo AM. Targeting the mTOR signaling network for cancer therapy. J Clin Oncol. 2009;27(13):2278–87.
- Brugarolas J. Renal-cell carcinoma–molecular pathways and therapies. N Engl J Med. 2007;356(2):185–7.

- 36. Averous J, Proud CG. When translation meets transformation: the mTOR story. Oncogene. 2006;25(48):6423–35.
- Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. Nat Rev Mol Cell Biol. 2009;10(5):307–18.
- Graff JR, Konicek BW, Carter JH, Marcusson EG. Targeting the eukaryotic translation initiation factor 4E for cancer therapy. Cancer Res. 2008;68(3):631–4.
- Hudson CC, Liu M, Chiang GG, Otterness DM, Loomis DC, Kaper F, et al. Regulation of hypoxia-inducible factor 1alpha expression and function by the mammalian target of rapamycin. Mol Cell Biol. 2002;22(20):7004–14.
- 40. Turner KJ, Moore JW, Jones A, Taylor CF, Cuthbert-Heavens D, Han C, et al. Expression of hypoxia-inducible factors in human renal cancer: relationship to angiogenesis and to the von Hippel-Lindau gene mutation. Cancer Res. 2002;62(10):2957–61.
- Toschi A, Lee E, Gadir N, Ohh M, Foster DA. Differential dependence of hypoxia-inducible factors 1 alpha and 2 alpha on mTORC1 and mTORC2. J Biol Chem. 2008;283(50):34495–9.
- Linehan WM, Srinivasan R, Schmidt LS. The genetic basis of kidney cancer: a metabolic disease. Nat Rev Urol. 2010;7(5):277–85.
- 43. Cho D, Signoretti S, Regan M, Mier JW, Atkins MB. The role of mammalian target of rapamycin inhibitors in the treatment of advanced renal cancer. Clin Cancer Res. 2007;13(2 Pt 2):758s–63s.
- 44. Kim WY, Kaelin WG. Role of VHL gene mutation in human cancer. J Clin Oncol. 2004;22(24):4991–5004.
- 45. Thomas GV, Tran C, Mellinghoff IK, Welsbie DS, Chan E, Fueger B, et al. Hypoxia-inducible factor determines sensitivity to inhibitors of mTOR in kidney cancer. Nat Med. 2006;12(1):122–7.
- Brenner W, Färber G, Herget T, Lehr HA, Hengstler JG, Thüroff JW. Loss of tumor suppressor protein PTEN during renal carcinogenesis. Int J Cancer. 2002;99(1):53–7.
- Velickovic M, Delahunt B, McIver B, Grebe SK. Intragenic PTEN/MMAC1 loss of heterozygosity in conventional (clear-cell) renal cell carcinoma is associated with poor patient prognosis. Mod Pathol. 2002;15(5):479–85.
- Bissler JJ, McCormack FX, Young LR, Elwing JM, Chuck G, Leonard JM, et al. Sirolimus for angiomyolipoma in tuberous sclerosis complex or lymphangioleiomyomatosis. N Engl J Med. 2008;358(2):140–51.
- 49. Davies DM, de Vries PJ, Johnson SR, McCartney DL, Cox JA, Serra AL, et al. Sirolimus therapy for angiomyolipoma in tuberous sclerosis and sporadic lymphangioleiomyomatosis: a phase 2 trial. Clin Cancer Res. 2011;17(12):4071–81.
- 50. Choi J, Chen J, Schreiber SL, Clardy J. Structure of the FKBP12-rapamycin complex interacting with the binding domain of human FRAP. Science. 1996;273(5272):239–42.
- Liang J, Choi J, Clardy J. Refined structure of the FKBP12-rapamycin-FRB ternary complex at 2.2 A resolution. Acta Crystallogr D Biol Crystallogr. 1999;55(Pt 4):736–44.
- Dancey J, Sausville EA. Issues and progress with protein kinase inhibitors for cancer treatment. Nat Rev Drug Discov. 2003;2(4):296–313.
- 53. Grünwald V, DeGraffenried L, Russel D, Friedrichs WE, Ray RB, Hidalgo M. Inhibitors of mTOR reverse doxorubicin resistance conferred by PTEN status in prostate cancer cells. Cancer Res. 2002;62(21):6141–5.
- 54. Dudkin L, Dilling MB, Cheshire PJ, Harwood FC, Hollingshead M, Arbuck SG, et al. Biochemical correlates of mTOR inhibition by the rapamycin ester CCI-779 and tumor growth inhibition. Clin Cancer Res. 2001;7(6):1758–64.
- 55. Yu K, Toral-Barza L, Discafani C, Zhang WG, Skotnicki J, Frost P, et al. mTOR, a novel target in breast cancer: the effect of CCI-779, an mTOR inhibitor, in preclinical models of breast cancer. Endocr Relat Cancer. 2001;8(3):249–58.
- 56. Geoerger B, Kerr K, Tang CB, Fung KM, Powell B, Sutton LN, et al. Antitumor activity of the rapamycin analog CCI-779 in human primitive neuroectodermal tumor/medulloblastoma models as single agent and in combination chemotherapy. Cancer Res. 2001;61(4):1527–32.

- Raymond E, Alexandre J, Faivre S, Vera K, Materman E, Boni J, et al. Safety and pharmacokinetics of escalated doses of weekly intravenous infusion of CCI-779, a novel mTOR inhibitor, in patients with cancer. J Clin Oncol. 2004;22(12):2336–47.
- Atkins MB, Hidalgo M, Stadler WM, Logan TF, Dutcher JP, Hudes GR, et al. Randomized phase II study of multiple dose levels of CCI-779, a novel mammalian target of rapamycin kinase inhibitor, in patients with advanced refractory renal cell carcinoma. J Clin Oncol. 2004;22(5):909–18.
- 59. Boulay A, Zumstein-Mecker S, Stephan C, Beuvink I, Zilbermann F, Haller R, et al. Antitumor efficacy of intermittent treatment schedules with the rapamycin derivative RAD001 correlates with prolonged inactivation of ribosomal protein S6 kinase 1 in peripheral blood mononuclear cells. Cancer Res. 2004;64(1):252–61.
- 60. O'Donnell A, Faivre S, Burris HA, Rea D, Papadimitrakopoulou V, Shand N, et al. Phase I pharmacokinetic and pharmacodynamic study of the oral mammalian target of rapamycin inhibitor everolimus in patients with advanced solid tumors. J Clin Oncol. 2008;26(10):1588–95.
- 61. Tanaka C, O'Reilly T, Kovarik JM, Shand N, Hazell K, Judson I, et al. Identifying optimal biologic doses of everolimus (RAD001) in patients with cancer based on the modeling of preclinical and clinical pharmacokinetic and pharmacodynamic data. J Clin Oncol. 2008;26(10):1596–602.
- 62. Tabernero J, Rojo F, Calvo E, Burris H, Judson I, Hazell K, et al. Dose- and scheduledependent inhibition of the mammalian target of rapamycin pathway with everolimus: a phase I tumor pharmacodynamic study in patients with advanced solid tumors. J Clin Oncol. 2008;26(10):1603–10.
- 63. Amato RJ, Jac J, Giessinger S, Saxena S, Willis JP. A phase 2 study with a daily regimen of the oral mTOR inhibitor RAD001 (everolimus) in patients with metastatic clear cell renal cell cancer. Cancer. 2009;115(11):2438–46.
- 64. Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S, et al. Phase 3 trial of everolimus for metastatic renal cell carcinoma: final results and analysis of prognostic factors. Cancer. 2010;116(18):4256–65.
- 65. Stein A, Wang W, Carter AA, Chiparus O, Hollaender N, Kim H, et al. Dynamic tumor modeling of the dose-response relationship for everolimus in metastatic renal cell carcinoma using data from the phase 3 RECORD-1 trial. BMC Cancer. 2012;12:311.
- 66. Motzer R, Barrios C, Kim T, Falcon S, Cosgriff T, Harker W, et al. RECORD-3: phase II randomized trial comparing sequential first-line everolimus and second-line sunitinib versus first-line sunitinib and second-line everolimus in patients with metastatic renal cell carcinoma. ASCO Annu Meet. 2013;Abstract 4504.
- Bellmunt J, Szczylik C, Feingold J, Strahs A, Berkenblit A. Temsirolimus safety profile and management of toxic effects in patients with advanced renal cell carcinoma and poor prognostic features. Ann Oncol. 2008;19(8):1387–92.
- White DA, Camus P, Endo M, Escudier B, Calvo E, Akaza H, et al. Noninfectious pneumonitis after everolimus therapy for advanced renal cell carcinoma. Am J Respir Crit Care Med. 2010;182(3):396–403.
- Duran I, Siu LL, Oza AM, Chung TB, Sturgeon J, Townsley CA, et al. Characterisation of the lung toxicity of the cell cycle inhibitor temsirolimus. Eur J Cancer. 2006;42(12):1875–80.
- 70. Schor-Bardach R, Alsop DC, Pedrosa I, Solazzo SA, Wang X, Marquis RP, et al. Does arterial spin-labeling MR imaging-measured tumor perfusion correlate with renal cell cancer response to antiangiogenic therapy in a mouse model? Radiology. 2009;251(3):731–42.
- Rini BI, Atkins MB. Resistance to targeted therapy in renal-cell carcinoma. Lancet Oncol. 2009;10(10):992–1000.
- 72. Choo AY, Yoon SO, Kim SG, Roux PP, Blenis J. Rapamycin differentially inhibits S6Ks and 4E-BP1 to mediate cell-type-specific repression of mRNA translation. Proc Natl Acad Sci U S A. 2008;105(45):17414–9.
- 73. O'Reilly KE, Rojo F, She QB, Solit D, Mills GB, Smith D, et al. mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates Akt. Cancer Res. 2006;66(3):1500–8.

- Julien LA, Carriere A, Moreau J, Roux PP. mTORC1-activated S6K1 phosphorylates Rictor on threonine 1135 and regulates mTORC2 signaling. Mol Cell Biol. 2010;30(4):908–21.
- Dibble CC, Asara JM, Manning BD. Characterization of Rictor phosphorylation sites reveals direct regulation of mTOR complex 2 by S6K1. Mol Cell Biol. 2009;29(21):5657–70.
- Sarbassov DD, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/ PKB by the rictor-mTOR complex. Science. 2005;307(5712):1098–101.
- 77. Maru S, Ishigaki Y, Shinohara N, Takata T, Tomosugi N, Nonomura K. Inhibition of mTORC2 but not mTORC1 up-regulates E-cadherin expression and inhibits cell motility by blocking HIF-2α expression in human renal cell carcinoma. J Urol. 2013;189(5):1921–9.
- Thiery JP. Epithelial-mesenchymal transitions in tumour progression. Nat Rev Cancer. 2002;2(6):442–54.
- Wan X, Harkavy B, Shen N, Grohar P, Helman LJ. Rapamycin induces feedback activation of Akt signaling through an IGF-1R-dependent mechanism. Oncogene. 2007;26(13):1932–40.
- 80. Shi Y, Yan H, Frost P, Gera J, Lichtenstein A. Mammalian target of rapamycin inhibitors activate the AKT kinase in multiple myeloma cells by up-regulating the insulin-like growth factor receptor/insulin receptor substrate-1/phosphatidylinositol 3-kinase cascade. Mol Cancer Ther. 2005;4(10):1533–40.
- 81. Cantley LC. The phosphoinositide 3-kinase pathway. Science. 2002;296(5573):1655-7.
- 82. Elfiky AA, Aziz SA, Conrad PJ, Siddiqui S, Hackl W, Maira M, et al. Characterization and targeting of phosphatidylinositol-3 kinase (PI3K) and mammalian target of rapamycin (mTOR) in renal cell cancer. J Transl Med. 2011;9:133.
- Wang S, Jessen K, Kessler L, et al. INK128, a novel TORC1/2 inhibitor with potent oral antitumor activity in preclinical models of renal cancer. Am Assoc Cancer Res Congress. 2011;Abstract 4486.
- Hsieh AC, Liu Y, Edlind MP, Ingolia NT, Janes MR, Sher A, et al. The translational landscape of mTOR signalling steers cancer initiation and metastasis. Nature. 2012;485(7396):55–61.
- 85. García-García C, Ibrahim YH, Serra V, Calvo MT, Guzmán M, Grueso J, et al. Dual mTORC1/2 and HER2 blockade results in antitumor activity in preclinical models of breast cancer resistant to anti-HER2 therapy. Clin Cancer Res. 2012;18(9):2603–12.
- Jessen K, Wang S, Kessler L, Guo X, Kucharski J, Stauton J, et al. INK128 is a potent and selective TORC1/2 inhibitor with broad oral antitumor activity. Mol Cancer Ther. 2009;8(suppl):B148.
- Yu K, Shi C, Toral-Barza L, Lucas J, Shor B, Kim JE, et al. Beyond rapalog therapy: preclinical pharmacology and antitumor activity of WYE-125132, an ATP-competitive and specific inhibitor of mTORC1 and mTORC2. Cancer Res. 2010;70(2):621–31.
- 88. Pike KG, Malagu K, Hummersone MG, Menear KA, Duggan HM, Gomez S, et al. Optimization of potent and selective dual mTORC1 and mTORC2 inhibitors: the discovery of AZD8055 and AZD2014. Bioorg Med Chem Lett. 2013;23(5):1212–6.
- 89. Chresta CM, Davies BR, Hickson I, Harding T, Cosulich S, Critchlow SE, et al. AZD8055 is a potent, selective, and orally bioavailable ATP-competitive mammalian target of rapamycin kinase inhibitor with in vitro and in vivo antitumor activity. Cancer Res. 2010;70(1):288–98.
- 90. Gupta GN, Lin KY, Sourbier C, et al. Preclinical efficacy of AZD8055, an ATP-competitive mammalian target of rapamycin (mTOR) kinase inhibitor, in vitro in clear cell renal cell carcinoma (RCC). Am Assoc Cancer Res Congress. 2011;Abstract 645.
- Naing A, Aghajanian C, Raymond E, Olmos D, Schwartz G, Oelmann E, et al. Safety, tolerability, pharmacokinetics and pharmacodynamics of AZD8055 in advanced solid tumours and lymphoma. Br J Cancer. 2012;107(7):1093–9.
- 92. Zhang H, Berel D, Wang Y, Li P, Bhowmick NA, Figlin RA, et al. A comparison of Ku0063794, a dual mTORC1 and mTORC2 inhibitor, and temsirolimus in preclinical renal cell carcinoma models. PLoS One. 2013;8(1):e54918.
- 93. Cho DC, Cohen MB, Panka DJ, Collins M, Ghebremichael M, Atkins MB, et al. The efficacy of the novel dual PI3-kinase/mTOR inhibitor NVP-BEZ235 compared with rapamycin in renal cell carcinoma. Clin Cancer Res. 2010;16(14):3628–38.

- 94. Maira SM, Stauffer F, Brueggen J, Furet P, Schnell C, Fritsch C, et al. Identification and characterization of NVP-BEZ235, a new orally available dual phosphatidylinositol 3-kinase/ mammalian target of rapamycin inhibitor with potent in vivo antitumor activity. Mol Cancer Ther. 2008;7(7):1851–63.
- 95. Serra V, Markman B, Scaltriti M, Eichhorn PJ, Valero V, Guzman M, et al. NVP-BEZ235, a dual PI3K/mTOR inhibitor, prevents PI3K signaling and inhibits the growth of cancer cells with activating PI3K mutations. Cancer Res. 2008;68(19):8022–30.
- Cao P, Maira SM, García-Echeverría C, Hedley DW. Activity of a novel, dual PI3-kinase/ mTor inhibitor NVP-BEZ235 against primary human pancreatic cancers grown as orthotopic xenografts. Br J Cancer. 2009;100(8):1267–76.
- 97. Baumann P, Mandl-Weber S, Oduncu F, Schmidmaier R. The novel orally bioavailable inhibitor of phosphoinositol-3-kinase and mammalian target of rapamycin, NVP-BEZ235, inhibits growth and proliferation in multiple myeloma. Exp Cell Res. 2009;315(3):485–97.
- Roulin D, Waselle L, Dormond-Meuwly A, Dufour M, Demartines N, Dormond O. Targeting renal cell carcinoma with NVP-BEZ235, a dual PI3K/mTOR inhibitor, in combination with sorafenib. Mol Cancer. 2011;10:90.
- 99. Burris H, Rodon J, Sharma S, Herbst R, Tabernero J, Infante J, et al. First-in-human phase I study of the oral PI3K inhibitor BEZ235 in patients (pts) with advanced solid tumors. J Clin Oncol. 2010;28(15s):suppl;abstr 3005.
- 100. Peyton J, Rodon Ahnert J, Burris H, Britten C, Chen L, Tabernero J, et al. A dose-escalation study with the novel formulation of the oral pan-class I PI3K inhibitor BEZ235, solid dispersion system (SDS) sachet, in patients with advanced solid tumors. J Clin Oncol. 2011;29:suppl;abstr 3066.
- 101. Garlich JR, De P, Dey N, Su JD, Peng X, Miller A, et al. A vascular targeted pan phosphoinositide 3-kinase inhibitor prodrug, SF1126, with antitumor and antiangiogenic activity. Cancer Res. 2008;68(1):206–15.
- 102. Peng Q, De P, Dey N, et al. Preclinical studies of a pan PI3K inhibitor (SF1126) in renal cell carcinoma. Am Assoc Cancer Res Congress. 2007;Abstract 2378.
- 103. Mahadevan D, Chiorean EG, Harris WB, Von Hoff DD, Stejskal-Barnett A, Qi W, et al. Phase I pharmacokinetic and pharmacodynamic study of the pan-PI3K/mTORC vascular targeted pro-drug SF1126 in patients with advanced solid tumours and B-cell malignancies. Eur J Cancer. 2012;48(18):3319–27.
- 104. Bendell JC, Rodon J, Burris HA, de Jonge M, Verweij J, Birle D, et al. Phase I, doseescalation study of BKM120, an oral pan-Class I PI3K inhibitor, in patients with advanced solid tumors. J Clin Oncol. 2012;30(3):282–90.
- 105. Voliva CF, Pecchi S, Burger M, et al. Biological characterization of NVP-BKM120, a novel inhibitor of phosphoinositide 3-kinase in Phase I/II clinical trials. Cancer Res. 2010;70(8 suppl 1):4498.
- 106. Burger MT, Pecchi S, Wagman A, Ni ZJ, Knapp M, Hendrickson T, et al. Identification of NVP-BKM120 as a potent, selective, orally bioavailable class I PI3 kinase inhibitor for treating cancer. ACS Med Chem Lett. 2011;2(10):774–9.
- 107. Kondapaka SB, Singh SS, Dasmahapatra GP, Sausville EA, Roy KK. Perifosine, a novel alkylphospholipid, inhibits protein kinase B activation. Mol Cancer Ther. 2003;2(11):1093–103.
- 108. Fu L, Kim YA, Wang X, Wu X, Yue P, Lonial S, et al. Perifosine inhibits mammalian target of rapamycin signaling through facilitating degradation of major components in the mTOR axis and induces autophagy. Cancer Res. 2009;69(23):8967–76.
- 109. Crul M, Rosing H, de Klerk GJ, Dubbelman R, Traiser M, Reichert S, et al. Phase I and pharmacological study of daily oral administration of perifosine (D-21266) in patients with advanced solid tumours. Eur J Cancer. 2002;38(12):1615–21.
- 110. Cho DC, Hutson TE, Samlowski W, Sportelli P, Somer B, Richards P, et al. Two phase 2 trials of the novel Akt inhibitor perifosine in patients with advanced renal cell carcinoma after progression on vascular endothelial growth factor-targeted therapy. Cancer. 2012;118(24):6055–62.

- 111. Yan L. MK-2206: a potent oral allosteric AKT inhibitor. AACR Annu Meet. 2009:Abstract Number: DDT01-1.
- 112. Lu W, Defeo-Jones D, Davis L. In vitro and in vivo antitumor activities of MK-2206, a new allosteric Akt inhibitor. American Association for Cancer Research. 2009:Abstr 3714.
- 113. Hirai H, Sootome H, Nakatsuru Y, Miyama K, Taguchi S, Tsujioka K, et al. MK-2206, an allosteric Akt inhibitor, enhances antitumor efficacy by standard chemotherapeutic agents or molecular targeted drugs in vitro and in vivo. Mol Cancer Ther. 2010;9(7):1956–67.
- 114. Yap TA, Yan L, Patnaik A, Fearen I, Olmos D, Papadopoulos K, et al. First-in-man clinical trial of the oral pan-AKT inhibitor MK-2206 in patients with advanced solid tumors. J Clin Oncol. 2011;29(35):4688–95.
- 115. Molife LR, Yan L, Vitfell-Rasmussen J, Zernhelt AM, Sullivan DM, Cassier PA, et al. Phase 1 trial of the oral AKT inhibitor MK-2206 plus carboplatin/paclitaxel, docetaxel, or erlotinib in patients with advanced solid tumors. J Hematol Oncol. 2014;7(1):1.
- 116. Jonasch E, Corn PG, Pagliaro LC, et al. Randomized phase II CTEP study of MK2206 versus everolimus in VEGF inhibitor refractory renal cell carcinoma patients. J Clin Oncol. 2013;31 suppl; abstr 4517.