# Chapter 3 Relevance of Angiogenesis in Neuroendocrine Tumors

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Abstract While traditional cytotoxic drugs have shown limited efficacy in neuroendocrine tumors (NETs), their biological features have been characterized and can be exploited therapeutically. Their most prominent trait is an extraordinary vascularization in low-grade NETs and a hypoxia-dependent angiogenesis in high-grade NETs, which is associated with a significant expression of many pro-angiogenic molecules. Therefore, several antiangiogenic compounds have been tested in these malignancies, and among these, sunitinib has demonstrated activity in pancreatic NET patients by dually targeting the vascular endothelial growth factor receptor (VEGFR) and platelet-derived growth factor receptor (PDGFR) pathways. In spite of these efficacious clinical results, apparent resistance to antiangiogenic therapies has been described in NET animal models and in clinical trials. Therefore, overcoming antiangiogenic resistance is a crucial step in the subsequent development of antiangiogenic therapies. Several strategies have been postulated to fight resistance, but preclinical studies and clinical trials will investigate and address these therapeutic approaches in the coming years in order to overcome resistance of antiangiogenic therapies in NETs.

Keywords Neuroendocrine tumor · Angiogenesis · Sunitinib · Molecular target

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## Introduction

Neuroendocrine tumors (NETs) are rare malignancies, but their incidence and prevalence has increased in the last decades [1]. This type of tumors comprises a heterogeneous family with a wide and complex spectrum of clinical behavior. The limited effectiveness of traditional DNA-damaging agents has led to the exploration of new targeted drugs based on the molecular features of these tumors, in order to improve their systemic treatment.

NETs have a number of biological features that can be exploited therapeutically, such as an extraordinary tumor vascularization with high expression of several pro-angiogenic molecules, being vascular endothelial growth factor (VEGF) the major mediator of tumor angiogenesis due to its properties of potent endothelial cell mitogen and vascular permeability-inducing agent. The presence of VEGF may be required to maintain the differentiated state of capillary vessels in the hypervascular tumors [2, 3]. Indeed, most NETs are hypervascular, as it is characteristic of normal endocrine glands, which have a dense vascular network that facilitates hormone secretion and dumping to the bloodstream. Specifically, NETs show a microvascular density ranging from 10- to 20-fold higher than in typical carcinomas. However, many studies have shown that in pancreatic NETs, microvascular density is higher in benign, low-grade tumors than in malignant, high-grade tumors [4]. Furthermore, these studies have demonstrated that intratumoral vessel density is associated with a good prognosis and prolonged survival [5], which is completely the opposite of other digestive epithelial tumors and most carcinomas in general. Thus, an intriguing characteristic of NETs is their physiologically derived high vessel density in low-grade tumors that is diminished over tumor progression and aggressiveness. Nevertheless, high-grade NETs typically show hypoxic areas and upregulation of hypoxia-response transcription factors (HIFs) and genes of cellular response to hypoxia (carbonic anhydrase IX, glucose transporters, etc.) [6]. The upregulation of HIFs involves the induction of various pro-angiogenic factors as hypoxia is a mechanism that induces angiogenic responses. Thus, while vessel density is lower in high-grade NETs, they show a very robust pro-angiogenic response that is clearly observed by increased endothelial proliferation and vascular overgrowth.

The high vascularization of NETs has its molecular base on the specific repertoire of secreted molecules from neuroendocrine cells. Indeed, neuroendocrine cells physiologically express a high level of pro-angiogenic molecules, particularly in the pancreas, but also in peptidergic endocrine cells, which constitutively synthesize several members of the VEGF family [7]. Consistently, NETs also typically express a variety of pro-angiogenic cytokines and growth factors, including vascular endothelial growth factors (VEGF-A, VEGF-C), fibroblast growth factors (FGFs), ephrins, or angiopoietins, among others. For example, neuroendocrine tumors (NETs) and their derived cell lines demonstrate a high capacity to synthesize and secrete high levels of several VEGF family members [2]. Overall, NETs not only show a dense vascular structure, but also have an angiogenic capacity that is characteristic of vessel-dependent tumors and thus evidencing a strong rationale for the use of antiangiogenic therapies in this type of malignancies. Therefore, several antiangiogenic compounds are currently undergoing clinical evaluation in NETs, either as monotherapy or in combination with chemotherapy or other targeted drugs. We will mention the biology of each of these mechanisms of angiogenesis and discuss the clinical data that are available to date.

## Early Days, "Early" Antiangiogenic Drugs

Experimental evidence of the sensitivity of NETs to antiangiogenic drugs is based on preclinical studies in animal models, where promising results were described in the mid- and late 1990s with strikingly efficacious effects ranging from tumor stabilization to tumor regression depending on the model used. In particular, several antiangiogenic drugs have been evaluated in a transgenic mouse model of insulinoma, the RIP-Tag2, developed by Douglas Hanahan [8]. In this transgenic mouse model, an angiogenic switch occurs in premalignant lesions followed by a persistent angiogenesis during progression to expansive solid tumors and invasive carcinomas. For this reason, different angiogenesis inhibitors, such as the naturally occurring antiangiogenic molecules angiostatin and endostatin, were tested at distinct stages of disease progression. The different antiangiogenic treatments have proved to prevent the antiangiogenic switch in premalignant lesions, intervene in the rapid expansion of small tumors, or induce the regression of large end-stage cancers. Thus, antiangiogenic drugs may prove most efficacious when they are targeted to specific stages of cancer [9]. Early studies with the aminopeptidase inhibitor TNP-470, minocycline, and interferon- $\alpha/\beta$  demonstrated an antiangiogenic effect together with an effective tumor growth impairment [10].

These preclinical results are associated with the clinical use of thalidomide in NETs. Thalidomide is an orally bioavailable immunomodulatory drug with antiangiogenic properties due to its capacity to inhibit tumor necrosis factor-alpha (TNF- $\alpha$ ) production and also VEGF and basic fibroblast growth factor (bFGF) pathways. The first small clinical study (n = 18) in NETs with thalidomide in monotherapy did not show objective responses [11]. Nevertheless, the combination of thalidomide and temozolomide was evaluated in another phase II study with a radiological response rate of 45 % in pancreatic NETs and 7 % in carcinoid tumors, respectively. However, a high incidence of grade 3–4 of lymphopenia was reported and 10 % of the patients had opportunistic infections [12].

The use of endostatin in the clinic has also demonstrated some benefit. *Endo-statin* is a 20-kDa proteolytic fragment of collagen XVIII with antiangiogenic and antitumor activity in preclinical studies (Fig. 3.1). The antiangiogenic function of endostatin has been well documented during the past decade. However, the exact mechanism that endostatin executes its antiangiogenic functions remains elusive.

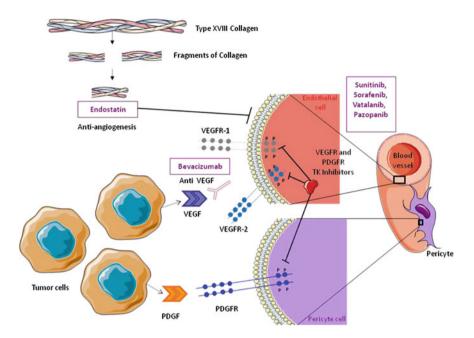


Fig. 3.1 Target pathways for antiangiogenic therapy in NETs. Image depicts the cellular and molecular components that drive angiogenesis in NETs (tumor cells, endothelial cells, pericytes, and extracellular matrix). Furthermore, in order to block the main pro-angiogenic pathways (VEGF/VEGFR and PDGF/PDGFR), different drugs such as endogenous inhibitors (endostatin), antibodies (bevacizumab) or small molecule inhibitors (sunitinib, sorafenib, vatalanib, and pazopanib) that can target vascular or perivascular cells have been developed

Both preclinical and human phase I studies of recombinant human endostatin (rhEndostatin) indicated activity in NETs. However, the phase II study performed in 40 patients with advanced NETs showed a high rate of stable disease (80 %) but did not result in significant tumor regression. The toxicity was minimal [13].

# The VEGF/VEGFR Axis

The key mediator of angiogenesis is the VEGF, and VEGF signaling inhibition has been shown to result in significant tumor growth delay in a wide range of animal models [14]. The inhibition of VEGF signaling not only arrests endothelial cells (ECs) proliferation and prevents vessel growth, but also induces regression of existing vessels by increasing EC death. VEGF inhibitors also suppress the mobilization of endothelial progenitor cells (EPCs) from the bone marrow and improve cytotoxic drug delivery by normalizing the chaotic and abnormal architecture of tumor vessels and reducing vascular permeability. Consistently, several antiangiogenic therapies targeting the VEGF/VEGFR2/KDR signaling axis have shown to be effective in mouse models of NETs. In particular, a monoclonal antibody that blocks VEGF-A ligand (AF-493-NA) and a blocking antibody of the VEGFR2 (DC101) has been tested in the RIP-Tag2 mouse model of insulinoma with consistent antiangiogenic effects in microvessel density, endothelial cell proliferation, and antitumor activity with increased apoptosis [3, 15]. Bevacizumab, a humanized monoclonal antibody that recognizes and blocks VEGF (Fig. 3.1), failed to inhibit growth NETs cells in vitro, but reduced their angiogenic potential by blocking the cells' ability to stimulate endothelial cell tube formation and proliferation and impaired tumor growth in animals [16].

Clinically, the activity of bevacizumab in NETs was tested in a randomized phase II study [17]. Forty-four patients on stable doses of octreotide were randomly assigned to 18 weeks of treatment with bevacizumab or PEG interferon alfa-2b. At disease progression (DP) or at the end of 18 weeks (whichever occurred earlier), patients received bevacizumab plus PEG interferon until progression. In the bevacizumab arm, four patients (18 %) achieved confirmed partial response (PR), 17 patients (77 %) had stable disease (SD), and one patient (5 %) had PD. No objective responses were observed in PEG interferon arm. Progression-free survival (PFS) rates after 18 weeks of monotherapy were 95 % in bevacizumab arm versus 68 % on the PEG interferon arm. Bevacizumab therapy also resulted in a significant reduction of tumor blood flow measured by functional CT scans.

A larger randomized phase III in patients with unresectable metastatic or locally advanced carcinoid tumors comparing depot octreotide acetate and interferon alfa-2b versus depot octreotide acetate and bevacizumab is being conducted since 2007 (SWOG S0518, clinicaltrials.gov NCT00569127). The results of this study are awaited in the near future.

Bevacizumab has also been tested in combination with cytotoxic drugs. Kulke et al. explored the efficacy and safety of the combination of bevacizumab plus temozolomide in a small phase II trial [18]. The combination showed an objective response rate of 24 % in pancreatic NETs but 0 % in carcinoid tumors. A phase II study of capecitabine, oxaliplatin, and bevacizumab for metastatic or unresectable NETs was reported in 2010 ASCO Annual Meeting. PR was observed in 7 pts (23 %), SD in 22 pts (71 %), and PD in 2 pts (6 %). Of the patients who achieved a PR, 6 had pancreatic NETs [19]. The combination with FOLFOX (oxaliplatin, leucovorin, and 5-fluorouracil) has also been tested with similar results [20]. Recently, a new phase II trial has been reported using the combination of bevacizumab with capecitabine in 49 patients with intestinal NETs. Nine (18.4 %) PR and 34 (69.4 %) SD were observed [21]. Another phase II trial testing bevacizumab plus traditional chemotherapy 5-FU/streptozotocin achieved an encouraging 55 % of PR with an acceptable toxicity profile [22]. Further phase III trials are warranted to establish the efficacy of adding bevacizumab to chemotherapy in NETs.

# Other Vascular Players: PDGFR Axis and the Pericytes

Not only vascular cells are important for angiogenesis, but also the periendothelial support cells of the microvasculature or pericytes have shown to be relevant targets for effective antiangiogenesis. These cells mediate the stabilization of the vessels based on the synthesis of new basement membrane and tight association with endothelial cells; thus, endothelial cells can induce pericyte recruitment to protect themselves from death consequent to the lack of the crucial tumor-derived survival signals conveyed by VEGF [23]. Molecularly, a specific cross talk between endothelial cells and pericytes that implicates VEGF and PDGF is key for the vascular formation and maintenance and creates a crucial therapeutic opportunity that has been exploited [24]. For its supportive cooperative function aiding the endothelial cell stabilization and function. PDGFR inhibition has been developed in the context of dual inhibition of VEGFR and PDGFR [9]. Indeed, experimental studies with the RIP-Tag2 transgenic mouse model demonstrate a significant synergy when both endothelial cells and pericytes are dually blocked with VEGFR and PDGFR small molecule inhibitors such as sunitinib, which elicits detachment of pericytes and disruption of tumor vascularity in multiple stages in tumorigenesis, most notably in the often-intractable late-stage solid tumor [25, 26]. Although these positive results of the dual targeting of VEGFR and PDGFR, undesirable effects could emerge because a severe reduction or lack of pericyte coverage may disrupt the integrity of the vasculature, enabling tumor cells to transit into the circulatory system, thereby facilitating metastasis [23, 27].

On the clinical side, PDGFRs have been characterized in human pancreatic NET samples. PDGFR- $\alpha$  and PDGFR- $\beta$  are commonly expressed both on tumor cells and tumor stroma [28]. The clinical approach to dually inhibit both VEGFR and PDGFR in NETs has been developed using several small molecule compounds such as sunitinib, sorafenib, vatalanib, and pazopanib (Fig. 3.1).

Sunitinib is the only antiangiogenic drug tested in a randomized phase III placebo-controlled trial [29] in patients with progressive well-differentiated pancreatic NETs, which is statistically positive in progression-free survival (11.4 months in sunitinib arm vs. 5.5 months in placebo arm). Sunitinib 37.5 mg/ day was administered orally in a continuous schedule. The objective response rate was 9.3 % in the sunitinib group versus 0 % in the placebo group. This study was the first positive phase III trial with antiangiogenic drugs in the field and has changed the daily clinical practice in NETs. In a previous phase II study, 107 patients (41 carcinoid tumors and 66 pancreatic NETs) with documented disease progression were treated with repeated six-week cycles of sunitinib 50 mg/day, four weeks on and two weeks off. The overall objective response rate was 16.7 % in pancreatic NETs and 2.4 % in carcinoid tumors [30].

Sorafenib is an orally active, multikinase inhibitor with selectivity for the VEGFR-2, VEGFR-3, PDGFR- $\beta$ , FLT3, c-kit, RET and RAF kinases. Sorafenib monotherapy has been evaluated in a phase II trial in 93 patients with NETs. The overall response rate was 10 % in both pancreatic and carcinoid NETs [31].

Vatalanib inhibits all known VEGFRs, with particular selectivity for VEGFR-2. At higher concentrations, vatalanib also inhibits PDGFR- $\beta$  and c-kit. Two phase II studies were reported in 2008 in NETs, but both showed no significant radiological responses [32, 33]. Finally, pazopanib, another potent inhibitor of VEGFR, PDFGR- $\alpha/\beta$ , and c-kit, has been tested in 33 patients, most of them previously treated with mTOR inhibitors or other antiangiogénica drugs, with a 6 % of PR 79 % SD. This trial may introduce the concept of treatment sequencing with novel targeted agents in NETs [34]. Pazopanib has also been tested in combination with octreotide LAR in pancreatic NETs with 17 % of PR and a PFS of 11.7 months [35]. Axitinib, another potent inhibitor of VEGFR 1-3, PDFGR- $\beta$ , and c-kit, is also been tested in this field.

#### Antiangiogenic Resistance

Clinical results using antiangiogenic drugs demonstrate only moderate gains in time to progression, and scarce benefits in overall survival, despite the long-term treatment. Why are there such modest and short-lasted benefits of antiangiogenic therapies in the clinic? The initial hypothesis was that antiangiogenesis therapy would not induce resistance (it would be "resistant to resistance") because it targeted endothelial cells instead of the tumor cell itself [36]. Nevertheless, clinical and experimental evidence indicates that a vascular regrowth in tumors is present after reversal of VEGF inhibition [37]. In some cases, there is a period of benefit followed by progression and mortality that reflects an adaptive response by tumors. Tumors can manifest an "evasive response" by upregulating alternative proangiogenic signals (such as ephrins or angiopoietins), recruiting pro-angiogenic inflammatory cells or pericytes, accentuating invasiveness of tumor cells into local tissue to co-opt normal vasculature, and increasing metastatic seeding and tumor cell growth in lymph nodes and distant organs. By contrast, patients for whom there is no tangible benefit at the beginning of the therapy indicate that an intrinsic resistance to angiogenesis inhibitors exists [38].

VEGF inhibition produces vascular trimming and hypoxia, which leads to upregulation of multiple pro-angiogenic molecules, including VEGFs, FGFs, and angiopoietins, which can contribute to eventual resistance [3, 38]. Tumor hypoxia could select for tumor populations able to grow in low oxygen environments [39, 40] and/or provide alternate compensatory pro-angiogenic pathways to allow persistent neovascularization [41] Furthermore, studies in the RIP-Tag2 model have described progression of NETs in course of antiangiogenic therapies targeting the VEGF/VEGFR signaling axis. Thus, genetic or pharmacological potent angiogenesis inhibition can alter the natural history of tumors by triggering resistance to therapy and increasing invasion and lymphatic or distant metastasis [42, 43]. Similar results have been observed in other models [44].

Acquired resistance can also be developed due to rapid vascular remodeling of tumor-associated vessels as a consequence of antiangiogenic therapy. The mature

Antiangiogenic drug	PDGFR	VEGF	VEGFR	FGFR	FLT-3	HIF-alfa
Sunitinib						
Bevacizumab						
Sorafenib						
Pazopanib						
Dovitinib						
Vatalanib						
Axitinib						
Brivanib						

Table 3.1 Multi-target inhibitory profile of antiangiogenic drugs to address resistance

remodeled vessels are resistant to antiangiogenic drugs, which usually target relatively immature vessels [45, 46].

Strategies to overcome this resistance mechanism are warranted. Based on preclinical data, several authors have proposed some strategies to overcome the antiangiogenic resistance that are based in combinatorial targeting of the VEGF pathway with other "escape" pathways that could be used for resistance (Table 3.1). In particular, some of these strategies, such as dual-targeted therapies, have been tested in xenografts [47]. The combination of bevacizumab and HIF-1 or Sp1 inhibitors may increase the therapeutic efficacy of antiangiogenic treatment [48, 49]. In another study, Allen et al. [50] suggest that co-targeting of VEGF and FGF signaling pathways can improve efficacy and overcome adaptive resistance to VEGF inhibition in the RIP-Tag2 model of pancreatic NETs. They tested the dual-FGFR/VEGFR tyrosine kinase inhibitor brivanib in both first and second line following the failure of anti-VEGFR2 antibody (DC101) or sorafenib showing promising results in overcoming resistance to VEGF-selective therapy.

On the clinical side, some phase II studies have tested the combination of antiangiogenic drugs. 2-Methoxyestradiol (2ME2) administered in combination with bevacizumab has been evaluated in a prospective study in thirty-one patients with metastatic carcinoid tumors [51]. No confirmed radiological responses by RECIST were observed. However, 68 % of the radiologically evaluable patients experienced at least some degree of tumor reduction, and the median PFS time was 11.3 months. The results of a study [52] with the combination of sorafenib and bevacizumab were reported in 2011 ASCO Annual Meeting. The overall response ratio was 9.8 %, and the disease control rate at 6 months was 95.1 %. Median progression-free survival was 12.4 months. The most common grade 3-4 toxicities were hand-foot syndrome and asthenia, which occurred in 20.5 % and 15.9 % of patients, respectively. Another trial has tested the combination of bevacizumab and everolimus in NETs. Addition of everolimus to bevacizumab was associated with further decrease in tumor blood flow (15 %; p = 0.02) than bevacizumab alone. By intention-to-treat (ITT) analyses, there were 26 % of PR and 27 % of SD. The median PFS was 14.4 months [53]. Recently, preliminary results of another phase II trial with the combination of bevacizumab and temsirolimus, another mTOR inhibitor, have been reported. Confirmed PR was documented in 11 of the first 25 (44 %) evaluable patients [54].

On the other hand, the identification of biomarkers for response or resistance to a particular antiangiogenic regimen is imperative in order to monitor the efficacy of antiangiogenic therapy. A study in the RIP-Tag2 model of pancreatic NETs described that tumors refractory to therapy following long-term treatment with a vascular endothelial growth factor receptor-2 blocking antibody contained blood vessels with a prolific investment of pericytes expressing  $\alpha$ -smooth muscle actin. This is a response of resistant tumors to the therapy, which is impairing neovascularization and/or eliciting vascular regression, and in order to maintain a core of preexisting blood vessels alive and functional, they increase the amount of pericytes [23]. Further studies are warranted to validate the occurrence of pericytes expressing  $\alpha$ -smooth muscle actin as a biomarker for tumors refractory to therapy [55].

#### **A** Perspective

Morphological, histological, and molecular features of NETs strongly support the notion that angiogenesis is a promising target in these malignancies. Indeed, several antiangiogenic drugs have been clinically validated, and two of those have been recently approved and are being incorporated in the daily clinical practice of pancreatic NETs. Nevertheless, not all patients respond to these therapies, demonstrating upfront refractoriness to therapy or intrinsic resistance. This patient population has to be carefully studied and detected in the future to find the most appropriate patient selection marker or characteristic in order to effectively treat these refractory patients. On the other hand, antiangiogenic drugs demonstrate clinical efficacy in many NETs patients, but these clinical benefits are overshadowed by apparent acquired resistance to antiangiogenic therapies emerging in NETs. Therefore, overcoming antiangiogenic resistance is a crucial step in the future development of antiangiogenic therapies. Several strategies have been postulated to fight resistance, including multi-pathway inhibitors or multi-combination of antiangiogenic drugs that target different pathways that can revert resistance caused by the upregulation of alternative pro-angiogenic signaling molecules, the recruitment of vascular progenitor cells or pericytes to the forming blood vessels, and also in order to fight against the increased capabilities for invasion without angiogenesis observed in some animal models. In this sense, clinical studies that investigate and address these approaches in the coming years are warranted.

Nevertheless, preclinical data in the RIP-Tag2 model indicate that many of these mechanisms of resistance show reversibility after antiangiogenic therapy has been stopped (Pàez-Ribes and Casanovas, unpublished observations). This confirms that these forms of resistance may reflect adaptations to therapy rather than irreversibly acquired capabilities and thus suggest that switching to a non-angiogenic drug in these resistant patients could revert their angiogenesis dependence and resensitize these patients to antiangiogenic drugs. Following this hypothesis, sequential

treatment with an antiangiogenic drug followed by a non-antiangiogenic drug (i.e., another targeted therapy or chemotherapy) could resensitize patients to another antiangiogenic drug as a third line of treatment. Obviously, many studies are warranted to unravel the preclinical basis and clinical potential of this hypothetical sequential treatment and to finally determine its clinical benefit for NETs patients.

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### References

- Yao JC, Hassan M, Phan A, Dagohoy C, Leary C, Mares JE, Abdalla EK, Fleming JB, Vauthey JN, Rashid A, Evans DB (2008) One hundred years after "carcinoid": epidemiology of and prognostic factors for neuroendocrine tumors in 35,825 cases in the United States. J Clin Oncol 26:3063–3072
- Terris B, Scoazec JY, Rubbia L, Bregeaud L, Pepper MS, Ruszniewski P, Belghiti J, Fléjou J, Degott C (1998) Expression of vascular endothelial growth factor in digestive neuroendocrine tumours. Histopathology 32:133–138
- Casanovas O, Hicklin DJ, Bergers G, Hanahan D (2005) Drug resistance by evasion of antiangiogenic targeting of VEGF signaling in late-stage pancreatic islet tumors. Cancer Cell 8:299–309
- 4. Couvelard A, O'Toole D, Turley H, Leek R, Sauvanet A, Degott C, Ruszniewski P, Belghiti J, Harris AL, Gatter K, Pezzella F (2005) Microvascular density and hypoxia-inducible factor pathway in pancreatic endocrine tumours: negative correlation of microvascular density and VEGF expression with tumour progression. Br J Cancer 92(1):94–101
- Marion-Audibert AM, Barel C, Gouysse G, Dumortier J, Pilleul F, Pourreyron C, Hervieu V, Poncet G, Lombard-Bohas C, Chayvialle JA, Partensky C, Scoazec JY (2003) Low microvessel density is an unfavorable histoprognostic factor in pancreatic endocrine tumors. Gastroenterology 125:1094–1104
- Couvelard A, Deschamps L, Rebours V, Sauvanet A, Gatter K, Pezzella F, Ruszniewski P, Bedossa P (2008) Overexpression of the oxygen sensors PHD-1, PHD-2, PHD-3, and FIH Is associated with tumor aggressiveness in pancreatic endocrine tumors. Clin Cancer Res 14:6634–6639
- Konstantinova I, Lammert E (2004) Microvascular development: learning from pancreatic islets. BioEssays 26:1069–1075
- 8. Hanahan D (1985) Heritable formation of pancreatic beta-cell tumours in transgenic mice expressing recombinant insulin/simian virus 40 oncogenes. Nature 315:115–122
- 9. Bergers G, Javaherian K, Lo KM, Folkman J, Hanahan D (1999) Effects of angiogenesis inhibitors on multistage carcinogenesis in mice. Science 284:808–812
- 10. Parangi S, Dietrich W, Christofori G, Holmgren L, Grosfeld J, Folkman J, Hanahan D et al (1995) Tumor suppressor loci on mouse chromosomes 9 and 16 are lost at distinct stages of tumorigenesis in a transgenic model of islet cell carcinoma. Cancer Res 55:6071–6076
- 11. Varker KA, Campbell J, Shah MH (2008) Phase II study of thalidomide in patients with metastatic carcinoid and islet cell tumors. Cancer Chemother Pharmacol 61:661–668
- Kulke MH, Stuart K, Enzinger PC, Ryan DP, Clark JW, Muzikansky A, Vincitore M, Michelini A, Fuchs CS (2006) Phase II study of temozolomide and thalidomide in patients with metastatic neuroendocrine tumors. J Clin Oncol 24(3):401–406

- 3 Relevance of Angiogenesis in Neuroendocrine Tumors
- Kulke MH, Bergsland EK, Ryan DP, Enzinger PC, Lynch TJ, Zhu AX, Meyerhardt JA, Heymach JV, Fogler WE, Sidor C, Michelini A, Kinsella K, Venook AP, Fuchs CS (2006) Phase II study of recombinant human endostatin in patients with advanced neuroendocrine tumors. J Clin Oncol 24(22):3555–3561
- 14. Kim KJ, Li B, Winer J, Armanini M, Gillett N, Phillips HS, Ferrara N (1993) Inhibition of vascular endothelial growth factor-induced angiogenesis suppresses tumour growth in vivo. Nature 362:841–844
- 15. Sennino B, Ishiguro-Oonuma T, Wei Y, Naylor RM, Williamson CW, Bhagwandin V, Tabruyn SP, You WK, Chapman HA, Christensen JG, Aftab DT, McDonald DM (2012) Suppression of tumor invasion and metastasis by concurrent inhibition of c-Met and VEGF signaling in pancreatic neuroendocrine tumors. Cancer Discov 2(3):270–287
- 16. Zhang J, Jia Z, Li Q, Wang L, Rashid A, Zhu Z, Evans DB, Vauthey JN, Xie K, Yao JC (2007) Elevated expression of vascular endothelial growth factor correlates with increased angiogenesis and decreased progression-free survival among patients with low-grade neuroendocrine tumors. Cancer 109(8):1478–1486
- 17. Yao JC, Phan A, Hoff PM, Chen HX, Charnsangavej C, Yeung SC, Hess K, Ng C, Abbruzzese JL, Ajani JA (2008) Targeting vascular endothelial growth factor in advanced carcinoid tumor: a random assignment phase II study of depot octreotide with bevacizumab and pegylated interferon alpha-2b. J Clin Oncol 26:1316–1323
- Kulke MH, Stuart K, Earle CC, Bhargava P, Clark JW, Enzinger PC, Meyerhardt JA, Attawia M, Lawrence C, Fuchs CS (2006) A phase II study of temozolomide and bevacizumab in patients with advanced neuroendocrine tumors. J Clin Oncol 24(18S):4044 (June 20 Supplement)
- Kunz PL, Kuo T, Zahn JM, Kaiser HL, Norton JA, Visser BC, Longacre TA, Ford JM, Balise RR, Fisher GA (2010) A phase II study of capecitabine, oxaliplatin, and bevacizumab for metastatic or unresectable neuroendocrine tumors. J Clin Oncol 28(15) (suppl; abstr 4104)
- 20. Venook AP, Ko AH, Tempero MA, Uy J, Weber T, Korn M, Bergsland EK (2008) Phase II trial of FOLFOX plus bevacizumab in advanced, progressive neuroendocrine tumors. J Clin Oncol 26:abstr
- 21. Mitry E, Walter T, Baudin E, Kurtz JE, Ruszniewski P, Dominguez S, Bengrine-Lefevre L, Cadiot G, Kraemer S, Ducreux M (2012) Efficacy and safety of bevacizumab combined with capecitabine in progressive, metastatic well-differentiated digestive endocrine tumors (BETTER study). J Clin Oncol 30 (suppl; abstr 4071)
- 22. Ducreux M, Seitz JF, Smith D, O'Toole D, Lepère C, Bitoun L, Mitry E (2012) Efficacy and safety of bevacizumab combined with chemotherapy in the treatment of patients with metastatic well-differentiated duodeno-pancreatic endocrine tumors (BETTER study). J Clin Oncol 30 (suppl; abstr 4036)
- Bergers G, Hanahan D (2008) Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer 8(8):592–603
- Franco M, Roswall P, Cortez E, Hanahan D, Pietras K (2011) Pericytes promote endothelial cell survival through induction of autocrine VEGF-A signaling and Bcl-w expression. Blood 118(10):2906–2917
- Bergers G, Song S, Meyer-Morse N et al (2003) Benefits of targeting both pericytes and endothelial cells in the tumor vasculature with kinase inhibitors. J Clin Invest 111:1287–1295
- 26. Pietras K, Hanahan D (2005) A multitargeted, metronomic, and maximum-tolerated dose "chemo-switch" regimen is antiangiogenic, producing objective responses and survival benefit in a mouse model of cancer. J Clin Oncol 23:939–952
- Xian X, Håkansson J, Ståhlberg A, Lindblom P, Betsholtz C, Gerhardt H, Semb H (2006) Pericytes limit tumor cell metastasis. J Clin Invest 116(3):642–651
- Fjallskog ML, Lejonklou MH, Oberg KE, Eriksson BK, Janson ET (2003) Expression of molecular targets for tyrosine kinase receptor antagonists in malignant endocrine pancreatic tumors. Clin Cancer Res 9:1469–1473

- 29. Raymond E, Dahan L, Raoul JL, Bang YJ, Borbath I, Lombard-Bohas C, Valle J, Metrakos P, Smith D, Vinik A, Chen JS, Horsch D, Hammel P, Wiedenmann B, Van Cutsem E, Patyna S, Lu DR, Blanckmeister C, Chao R, Ruszniewski P (2011) Sunitinib malate for the treatment of pancreatic neuroendocrine tumors. N Engl J Med 364:501–513
- Kulke MH, Lenz HJ, Meropol NJ, Posey J, Ryan DP, Picus J, Bergsland E, Stuart K, Tye L, Huang X, Li JZ, Baum CM, Fuchs CS (2008) Activity of sunitinib in patients with advanced neuroendocrine tumors. J Clin Oncol 26(20):3403–3410
- 31. Hobday TJ, Rubin J, Holen K, et al (2007) MC044 h, a phase II trial of sorafenib in patients (pts) with metastatic neuroendocrine tumors (NET): a Phase II Consortium (P2C) study. J Clin Oncol 25:abstr
- 32. Pavel ME, Bartel C, Heuck F, Neumann F, Tiling N, Pape UF, Plöckinger U, Wiedenmann B (2008) Open-label, non-randomized, multicenter phase II study evaluating the angiogenesis inhibitor PTK787/ZK222584 (PTK/ZK) in patients with advanced neuroendocrine carcinomas (NEC). J Clin Oncol 26:abstr
- Anthony L, Chester M, Michael S, O'Dorisio TM, O'Dorisio MS (2008) Phase II open-label clinical trial of vatalanib (PTK/ZK) in patients with progressive neuroendocrine cancer. J Clin Oncol (May 20 suppl; abstr 14624)
- 34. Grande E, Castellano D, Garcia-Carbonero R, Teule A, Duran I, Fuster J, Sevilla I, Escudero P, Sastre J, Capdevila J (2012) PAZONET: Results of a phase II trial of pazopanib as a sequencing treatment in progressive metastatic neuroendocrine tumors (NETs) patients (pts), on behalf of the Spanish taskforce for NETs (GETNE) NCT01280201. J Clin Oncol 30 (suppl; abstr 4119)
- 35. Phan AT, Yao JC, Fogelman DR, Hess KR, Ng CS, Bullock SA, Malinowski P, Regan E, Kulke M (2010) A prospective, multi-institutional phase II study of GW786034 (pazopanib) and depot octreotide (sandostatin LAR) in advanced low-grade neuroendocrine carcinoma (LGNEC). J Clin Oncol 28(7) (suppl; abstr 4001)
- Boehm T, Folkman J, Browder T, O'Reilly MS (1997) Antiangiogenic therapy of experimental cancer does not induce acquired drug resistance. Nature 390:404–407
- 37. Mancuso MR, Davis R, Norberg SM, O'Brien S, Sennino B, Nakahara T, Yao VJ, Inai T, Brooks P, Freimark B, Shalinsky DR, Hu-Lowe DD, McDonald DM (2006) Rapid vascular regrowth in tumors after reversal of VEGF inhibition. J Clin Invest 116(10):2610–2621
- Bergers G, Hanahan D (2008) Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer 8(8):592–603
- Hirota K, Semenza GL (2006) Regulation of angiogenesis by hypoxia-inducible factor 1. Crit Rev Oncol Hematol 59(1):15–26
- 40. Yu JL, Rak JW, Coomber BL, Hicklin DJ, Kerbel RS (2002) Effect of p53 status on tumor response to antiangiogenic therapy. Science 295(5559):1526–1528
- Rapisarda A, Melillo G (2009) Role of the hypoxic tumor microenvironment in the resistance to anti-angiogenic therapies. Drug Resist Updat 12(3):74–80
- 42. Pàez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Viñals F, Inoue M, Bergers G, Hanahan D, Casanovas O (2009) Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. Cancer Cell 15(3):220–231
- 43. Takeda T, Okuyama H, Nishizawa Y, Tomita S, Inoue M (2012) Hypoxia inducible factor-1α is necessary for invasive phenotype in Vegf-deleted islet cell tumors. Sci Rep 2:494
- 44. Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG, Kerbel RS (2009) Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. Cancer Cell 15(3):232–239
- 45. Glade Bender J, Cooney EM, Kandel JJ, Yamashiro DJ (2004) Vascular remodeling and clinical resistance to antiangiogenic cancer therapy. Drug Resist Updat 7, 289–300
- 46. Benjamin LE, Golijanin D, Itin A, Pode D, Keshet E (1999) Selective ablation of immature blood vessels in established human tumors follows vascular endothelial growth factor withdrawal. J Clin Invest 103(2):159–165
- 47. Yao JC, Phan A (2011) Overcoming antiangiogenic resistance. Clin Cancer Res 17(16):5217–5219

- 3 Relevance of Angiogenesis in Neuroendocrine Tumors
- 48. Rapisarda A, Hollingshead M, Uranchimeg B, Bonomi CA, Borgel SD, Carter JP et al (2009) Increased antitumor activity of bevacizumab in combination with hypoxia inducible factor-1 inhibition. Mol Cancer Ther 8:1867–1877
- 49. Jia Z, Zhang J, Wei D, Wang L, Yuan P, Le X et al (2007) Molecular basis of the synergistic antiangiogenic activity of bevacizumab and mithramycin. Cancer Res 67:4878–4885
- 50. Allen E, Walters IB, Hanahan D (2011) Brivanib, a Dual FGF/VEGF inhibitor, is active both first and second line against mouse pancreatic neuroendocrine tumors developing adaptive/ evasive resistance to VEGF inhibition. Clin Cancer Res 17:5299–5310
- 51. Kulke MH, Chan JA, Meyerhardt JA, Zhu AX, Abrams TA, Blaszkowsky LS, Regan E, Sidor C, Fuchs CS (2011) A prospective phase II study of 2-methoxyestradiol administered in combination with bevacizumab in patients with metastatic carcinoid tumors. Cancer Chemother Pharmacol 68:293–300
- 52. Castellano DE, Capdevila J, Salazar R, Sastre J, Alonso V, Llanos M, Garcia-Carbonero R, Abad A, Sevilla I, Duran I (2011) Sorafenib and bevacizumab combination targeted therapy in advanced neuroendocrine tumor: a phase II study of the Spanish Neuroendocrine Tumor Group (GETNE0801). J Clin Oncol 29 (suppl; abstr 4113)
- 53. Yao JC, Phan AT, Fogleman D, Ng CS, Jacobs CB, Dagohoy CD, Leary C, Hess KR (2010) Randomized run-in study of bevacizumab (B) and everolimus (E) in low- to intermediategrade neuroendocrine tumors (LGNETs) using perfusion CT as functional biomarker. J Clin Oncol 28(15) (suppl; abstr 4002)
- 54. Hobday TJ, Qin R, Reidy DL, Moore MJ, Strosberg JR, Kaubisch A, Shah M, Kindler HL, Lenz HJ, Chen HX, Erlichman C (2012) Multicenter phase II trial of temsirolimus (TEM) and bevacizumab (BEV) in pancreatic neuroendocrine tumor (PNET). J Clin Oncol 30 (suppl 4; abstr 260)
- 55. Franco M, Pàez-Ribes M, Cortez E, Casanovas O, Pietras K (2011) Use of a mouse model of pancreatic neuroendocrine tumors to find pericyte biomarkers of resistance to anti-angiogenic therapy. Horm Metab Res 43(12):884–889 Epub 2011 Sep 29