# **13 MRI for Detection and Evaluation of Hepatocellular Carcinoma**

# *Donald G. Mitchell, MD, FACR*

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#### **ABSTRACT**

MRI is a useful method of imaging the cirrhotic liver, including for detection and evaluation of hepatocellular carcinoma (HCC), both for its initial diagnosis and following its response to management. In this chapter, we discuss features which allow distinction of HCC from other lesions in the cirrhotic liver, such as regenerative nodules, confluent fibrosis, and benign enhancing pseudonodules. One major strength of MRI is its use of multiple pulse sequences, analogous to the use of various stains for histopathology. Pulse sequences with unique value for characterizing focal liver lesions include T1-weighted, T2-weighted, lipid-sensitive, and multiphasic contrast-enhanced images. Features that facilitate diagnosis of HCC include its shape, capsule, internal nodularity, signal intensity, and sequential

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pattern of dynamic contrast enhancement. It is particularly important that radiologists and clinicians reach understanding on terminology for expressing confidence that a given focal lesion is HCC or benign, so that reported findings are most useful for guiding management decisions. A suggested framework for categorizing this confidence is provided.

#### **Key Words:** MRI; liver; hepatocellular carcinoma

Evaluation and management of patients with cirrhosis present many challenges, one of which is the reliable detection of hepatocellular carcinoma (HCC) at a stage when treatment can improve the length and quality of a patient's life. As with other cancers, the potential value of imaging for initial detection depends on many factors, which are the following:

- 1. Is there a population of high-risk individuals who can be identified for screening by imaging?
- 2. Is imaging capable of detecting the malignancy earlier than clinical or laboratory methods?
- 3. Is there an effective method for treating the malignancy at the stage when it is most likely to be detected?
- 4. Do the benefits of early detection and treatment compare favorably with the financial and other costs of the imaging screening program?

In the case of HCC, the answers to all of the above questions are yes. Patients with cirrhosis, especially of viral etiology, are at high risk for developing HCC *[\(1](#page-12-2)*–*[4\)](#page-12-3)*. Magnetic resonance imaging (MRI), computed tomography (CT), and ultrasound can all detect HCC, often before α-fetoprotein and other nonimaging signs allow its diagnosis *[\(5](#page-12-4)*–*[9\)](#page-13-0)*. Imaging can be used judiciously to diagnose small HCC, obviating biopsy when imaging diagnostic signs are particularly compelling *[\(10\)](#page-13-1)*. HCC can be locally treated by many methods, often improved when used in combinations, including chemoembolization, radioembolization, chemical ablations (e.g., ethanol or acetic acid), and RF ablation *[\(4](#page-12-3)*, *[9](#page-13-0)*, *[11](#page-13-2)*–*[17\)](#page-13-3)*. The success of these methods might possibly be further augmented when combined with systemic therapy, such as with agents that target VEGF receptors and tumor-induced angiogenesis *[\(18](#page-13-4)*–*[20\)](#page-13-5)*. If the local and systemic treatments mentioned above can prevent or prolong the interval before HCC spreads to extrahepatic sites, liver transplant can be used to cure the patient of HCC and prevent recurrence *[\(11](#page-13-2)*, *[21](#page-13-6)*–*[25\)](#page-13-7)*. Therefore, HCC presents itself as a particularly valuable opportunity for imaging to improve the lives of patients at risk for this malignancy *[\(2\)](#page-12-5)*.

In this chapter, I will make some general comments about the challenges that must be addressed to detect HCC within a cirrhotic liver. I will provide a

framework to reduce some confusion regarding terminology that may appear in the imaging literature and in clinical imaging reports. I will then discuss several of the features that help distinguish HCC from other focal findings in cirrhotic livers. Finally, based upon limited literature and some perspective gained from clinical experience, I will offer some suggestions about how the use of MRI for detecting hepatocellular cancer may proceed during the next few years.

# <span id="page-2-0"></span>**1. FOCAL IMAGING FINDINGS IN CIRRHOTIC LIVER**

As cirrhosis develops and progresses, the remaining liver parenchyma consists of regenerative nodules of variable size, surrounded by fibrous septations. The first step in evaluating images of a cirrhotic liver is the recognition that the tissue between the fibrotic septations, that is, the regenerative nodules, should generally resemble healthy hepatocellular parenchyma. The abnormal appearance of a cirrhotic liver is caused by alterations in shape due to the combination of scarring, atrophy of some portions, and hypertrophy of others, as well as abnormal signal imparted by the presence of fibrosis and inflammation.

Once a focal part of the liver is noted that appears different compared with the surrounding liver, the next task is to determine whether this tissue is more or less abnormal than the remaining liver parenchyma *[\(26\)](#page-13-8)*. For example, a relatively sparred area within a severely diseased liver can resemble a mass, when in fact the focal finding is less diseased than the surrounding tissue. The challenge here, before even considering whether there is evidence of malignancy, is to categorize the following benign tissues:

- 1. *Regenerative nodules*. In fact, the entire cirrhotic liver consists of regenerative nodules. Therefore, any distinct nodule that looks different from the background liver should arouse at least a modest level of suspicion.
- 2. Confluent fibrosis and severely damaged liver, containing few if any hepatocytes, will look distinctly different from healthy liver parenchyma. Confluent fibrosis is darker on T1-weighted images and brighter on T2-weighted images, features shared by most malignancies, including some HCCs. Confluent fibrosis is therefore best distinguished from HCC by its shape, which is geographic rather round, and by retraction of liver shape, rather than expansion *[\(27\)](#page-14-0)*.
- 3. Benign enhancing pseudonodules are the most common problem leading to false-positive diagnosis and frequent follow-up imaging examinations. As discussed toward the end of this chapter, subcentimeter

foci of transient enhancement are extremely common in a cirrhotic liver and are usually benign *[\(28](#page-14-1)*–*[32\)](#page-14-2)*. Frequent short-term follow-up of these common benign findings therefore threatens to dramatically increase the overall cost of an imaging screening program, and should be minimized to whatever extent possible *[\(33\)](#page-14-3)*.

- 4. Hyperplastic nodule has only scant description in the literature *[\(34](#page-14-4)*–*[39\)](#page-14-5)*, although it is probably a common cause of false-positive MRI. Like the more common regenerative nodule, a hyperplastic nodule is composed entirely of benign liver tissue. In fact, there is often minimal distinction between these two entities in the pathologic literature, due to their absence of dysplastic or neoplastic cellular features *[\(40\)](#page-14-6)*. The main distinction between these two nodular entities is their blood supply, which causes dramatic differences on contrast-enhanced imaging studies but may have little or no effect on their light microscopic appearance. Hyperplastic nodules are thought to arise as a response to alterations in portal venous perfusion, giving rise to nodular hypertrophic tissue with vascular supply entirely from hepatic arteries, without meaningful contribution from portal veins *[\(41\)](#page-14-7)*. Hyperplastic nodules are most common in the setting of Budd-Chiari syndrome but can arise in any scenario where portal venous perfusion is abnormal, including in patients with cirrhosis. In the setting of an otherwise normal liver, these nodules are termed focal nodular hyperplasia (FNH). In fact, in patients with Budd-Chiari syndrome or cirrhosis, the term "FNHlike nodule" has been used *[\(35](#page-14-8)*–*[37\)](#page-14-9)*. This is an unnecessarily redundant term, so the more generic and simpler term hyperplastic nodule is preferable. Hyperplastic nodules are considered entirely benign, without premalignant nature, and should not be confused with dysplastic nodule.
- 5. Dysplastic nodule is a borderline lesion, with atypical cellular features different from those of regenerative or hyperplastic nodules but not meeting criteria for overt malignancy *[\(40](#page-14-6)*, *[42](#page-14-10)*, *[43\)](#page-14-11)*. They are considered premalignant, and foci of HCC may develop within them. Dysplastic nodules can be visible on imaging studies, although their features overlap those of some regenerative nodules and some HCCs. Therefore, dysplastic nodule can be included in the differential diagnosis of a nodule in a cirrhotic liver, but at this point, dysplastic nodule is not a specific diagnosis that can be offered by imaging.

HCC is the subject of this entire book and need not be defined here. Rather, in the next section I will describe imaging features of HCC and indicate how these may be used to distinguish them from other nodules and focal findings in the cirrhotic liver.

## **2. MRI: PULSE SEQUENCES AND GENERAL CONSIDERATIONS**

<span id="page-4-0"></span>Like other methods of imaging, MRI can depict hepatic and abdominal anatomy. However, MRI also offers a more robust and comprehensive set of tools for characterizing tissue. It is therefore customary for most MRI exams, particularly hepatic MRI, to include multiple pulse sequences repetitively interrogating the same tissue, often in identical image planes. In this respect it is analogous to light microscopy, where the same histologic structure is repeatedly evaluated using different stains, each designed to highlight a particular tissue component of interest. Most MRI examinations will include the following.

#### *2.1. Survey Images*

These typically include coronal images but may also include sagittal and transverse images. They provide a brief survey of the abdomen in 1 minute or less and help determine the region of the abdomen to be included in the remainder of the examinations. On occasion, the position of the patient and local receiver coils may need to be changed to best optimize the signals received.

#### *2.2. T1-Weighted Images with Lipid and Iron Sensitivity*

T1 is a characteristic of tissues, whereby short T1 leads to high signal intensity (bright on the images) on T1-weighted images. In order of increasing T1 (decreasing brightness on T1-weighted images) are adipose tissue, liver parenchyma, most other tissues including malignancies, and simple cysts. On basic T1-weighted images, adipose tissue is therefore bright, liver medium, and simple cysts dark. HCC has variable appearance and therefore can be dark, intermediate, or bright on T1-weighted images *[\(44](#page-14-12)*, *[45\)](#page-15-0)*.

Inherent differences between the protons in water and the protons in lipid can be exploited, in various ways, to separate the signals from water vs. lipid protons. It is now routine to obtain T1-weighted images as a pair of images, based on two consecutive echoes *[\(46](#page-15-1)*–*[48\)](#page-15-2)*. One of these is "in-phase," where the signals of water and lipid protons add together. The other is "opposed-phase," whereby water protons and most protons from lipid interfere destructively, so that points in the image that contain water and fat, such as fatty liver parenchyma, show up as darker compared with inphase images. These two paired images, obtained at exactly the same time and place, can either be visually compared or be postprocessed to generate difference images. It is also standard, at some point in the examination, to obtain T1-weighted images where lipid protons are selectively suppressed, generating "fat-suppressed T1-weighted images."

#### *2.3. T2-Weighted Images*

These images accentuate differences in the T2 between different tissues. Like T1, T2 is characteristic of tissues. T1 and T2 commonly, but not always, parallel each other. For example, both simple cysts and cerebral spinal fluid have extremely long T1 and long T2, and are therefore bright on T2-weighted images. Liver is dark on most T2-weighted images, whereas moderately to poorly differentiated HCC is usually brighter on these images, similar to spleen. Images can be made more T2 weighted by lengthening the echo time (TE). It is common to obtain two different sets of T2-weighted images, one with moderate T2 weighting to show liver tumors and enlarged lymph nodes, and one with heavy T2 weighting to show fluid as much brighter than solid tissue. In fact, extremely heavily T2 weighted images are commonly obtained to accentuate biliary and pancreatic ducts to form magnetic resonance cholangiopancreatography (MRCP) images. Heavily T2-weighted images are helpful for distinguishing benign cysts and hemangiomas from solid tissue, including HCCs.

#### *2.4. Dynamic Multiphasic Contrast-Enhanced Images*

These images are routine and considered essential for sensitive detection of HCC. As a minimum, four separate sets of T1-weighted images, usually with fat suppressed 3D thin-slice technique, are obtained. These included unenhanced images, images obtained during the first pass of contrast material through arteries (arterial phase), images obtained about 20 seconds after the arterial phase (blood pool or venous phase), and images obtained three or more minutes after contrast material has been allowed to equilibrate throughout the vascular and interstitial spaces (delayed or extracellular phase images).

Most HCCs will be bright on arterial phase images due to their predominant supply by arterial rather than portal venous perfusion, and most will be less intense blood pool or delayed phase images (probably because of less fibrosis in HCC compared with background liver parenchyma).

There is a new class of gadolinium contrast agent that has partial hepatobiliary excretion, including gadobenate dimeglumine and gadoxetic acid disodium *[\(49](#page-15-3)*–*[51\)](#page-15-4)*. These agents have weak binding to serum proteins, approximately doubling their effect on MR images at a given dose. An additional advantage of these agents is increased enhancement of liver tissue compared with most tumors during delayed phase imaging, after contrast agent has been primarily cleared from blood.

There are some additional images that are included in some protocols because of their potential to provide additive value or confirmation of information from other sequences, but are not necessarily routine.

*Diffusion weighted images* utilize microscopic water motion to highlight differences between tissues *[\(52](#page-15-5)*–*[57\)](#page-15-6)*. Generally, malignant tumors have restricted water motion compared with many benign tissues. *MR spectroscopy* allows detailed analysis of chemical differences depending on molecular structure, either of protons or other nuclei, but usually with much lower spatial resolution *[\(58\)](#page-15-7)*. At the present time, neither of these techniques should be considered routine or essential for detecting HCC.

*Bright-blood images* can be used to demonstrate blood vessels, using either motion-compensated techniques to show the water in blood or use the motion of the blood to show patent vessels. These images are often included if the contrast-enhanced images are technically inadequate due to motion or other artifacts, or if gadolinium contrast agent is not given.

*Perfusion imaging.* Advances in MRI hardware and pulse sequence design as well as image postprocessing can extend the value of dynamic contrast enhancement so that images are repeated at more rapid intervals. As a first step, early and late arterial phase images can be obtained during one breath hold. Further increases in speed are also possible, and signal intensities at various phases can be measured and applied to various perfusion algorithms to further characterize tissue. The broad class of perfusion imaging has been used to characterize properties of angiogenesis. It is possible that this method of image analysis may prove useful for characterizing response to new treatments such as VEGF antagonists *[\(18](#page-13-4)*, *[57](#page-15-6)*, *[59](#page-15-8)*–*[62\)](#page-15-9)*.

*Particulate contrast agents.* This class of contrast agent, usually consisting of iron oxide particles that accumulate avidly in Kupffer cells and other cells of the reticuloendothelial system, can darken the surrounding liver and thereby improve the visibility of HCC on appropriate MR images *[\(11](#page-13-2)*, *[21](#page-13-6)*, *[24](#page-13-9)*, *[25](#page-13-7)*, *[63](#page-15-10)*–*[65\)](#page-16-0)*. The most successful use of iron oxide contrast agents is in "double contrast MRI" when combined with gadolinium contrast agents *[\(66](#page-16-1)*–*[71\)](#page-16-2)*. The increased cost of using two contrast agents has prevented adoption of this technique at most centers.

#### **3. CHARACTERISTICS OF HCC**

<span id="page-6-0"></span>On *T1-weighted MR images*, HCCs can be dark, intermediate, or bright relative to background liver parenchyma. In spite of this extreme variability, T1-weighted images are still useful. For example, hemangiomas, cysts, and most other malignancies are more consistently dark on T1-weighted images, so intermediate or high signal helps to exclude these alternative diagnoses. Comparison of in-phase and opposed-phase images allows detection of even small quantities of lipid, a common finding in HCCs but not present in liver masses that are not derived from hepatocytes, such as hemangiomas, metastases, or cholangiocarcinoma. High signal intensity on both in-phase and opposed-phase images indicates hepatocellular tissue with copper *[\(72](#page-16-3)*–*[74\)](#page-16-4)*. These nodules may be HCC, dysplastic nodule, or other liver tissues with cholestasis.

*T2-weighted images* are often useful for depicting malignant liver tumors as brighter than background liver, although many HCCs have low signal intensity or be invisible on T2-weighted images *[\(75\)](#page-16-5)*. The main value of T2 weighted images for evaluating suspected HCC is their specificity. A solid round mass in a cirrhotic liver with high signal intensity on T2-weighted images is usually HCC.

The *shape* of a focal liver abnormality is quite helpful. HCCs are usually round, ovoid, or lobulated. HCCs can produce geographic abnormalities after they invade portal veins and disseminate by a portal venous spread.

A *capsule or pseudocapsule* is a common finding surrounding hepatocellular cancer. A capsule appearance is generally not seen with other focal liver lesions such as dysplastic nodule, hydroplastic nodule, or adenoma.

*Internal nodularity* (mosaic appearance) is a characteristic of HCC caused by variable dedifferentiation of foci within a dysplastic or a neoplastic mass *[\(76\)](#page-16-6)*. Benign entities such as dysplastic or hyperplastic nodule or liver regeneration have simpler texture, without internal nodularity. At an early stage, a "nodule-in-nodule" configuration results from focal dedifferentiation to HCC within a dysplastic nodule *[\(42](#page-14-10)*, *[77](#page-16-7)*–*[79\)](#page-16-8)*. A similar appearance can result from focal further dedifferentiation into less differentiated carcinoma within a well-differentiated carcinoma. Therefore, one or more nodules within a focal mass are a strong characteristic of HCC. These focal dedifferentiated nodules will usually have higher signal on T2-weighted images, lower signal on T1-weighted images, and more arterial vascularity. They will also tend to have rounder shape, as they exert mass effect on the less rapidly growing more differentiated remainder of the tumor.

The *dynamic contrast-enhanced series* is the single most important part of an MRI examination for HCC. The characteristics of the dynamic contrast MRI series are in many respects mimicked by dynamic multiphasic CT. One important advantage of MRI over CT is the complimentary value afforded by the additional MRI pulse sequences, which do not have analogous CT counterparts. Additionally, MRI spares the patient repeated exposures to ionizing radiation and iodinated contrast material.

The arterial phase images are the single most sensitive series for detecting HCC. However, there are caveats. While this series may be more sensitive than any single other series, there are indeed HCCs that may be visible only on other pulse sequences, not on arterial phase images *[\(31](#page-14-13)*, *[80\)](#page-16-9)*. Additionally, benign nodules are often seen as hyperintense on arterial phase MR images. In fact, more than 90% of small nodules seen only on arterial phase images are benign *[\(29](#page-14-14)*, *[30](#page-14-15)*, *[81\)](#page-16-10)*. The specificity of dynamic contrast-enhanced series is improved greatly if the nodule is visible on at least one additional series. Most commonly, this will be a "washout appearance," whereby a nodule that is hyperintense on arterial phase images is hypointense on blood pool (portal venous) phase or delayed phase images. Additionally, a nodule that is visible on an unenhanced image and then shows increased enhancement relative to liver during the arterial phase is more likely to be malignant than a nodule that is visible only on arterial phase images *[\(31\)](#page-14-13)*.

Three illustrative cases are provided in Figs. [1,](#page-8-0) [2](#page-9-1) and [3.](#page-10-0)



<span id="page-8-0"></span>**Fig. 1.** HCC with many typical MRI features. **A**. T2-weighted image shows HCC as high signal intensity (*arrow*). **B**. T1-weighted image in-phase (water plus fat) shows that most of HCC has similar intensity to remainder of liver, other than increased signal of anterior crescentic portion (*arrows*). **C**. T1-weighted image opposed-phase (water–fat cancellation) shows that HCC lost signal relative to other tissues, indicating lipid content. The anterior crescentic portion (*arrows*) has highest fat content and has therefore lost the most signal. **D**. T1-weighted fat-suppressed image shows the HCC as less signal than the remainder of liver. **E**. As in D, immediately after intravenous injection of gadolinium contrast agent. Hypervascular nodules within the HCC show strong enhancement (*arrows*). **F**. As in E, about 1 minute later. The HCC is now less intense than liver, with a multinodular appearance.



**Fig. 2.** HCC following chemoembolization, with small remaining viable portion. **A**. Unenhanced CT shows embolic material within HCC. **B**. Contrast-enhanced CT does not show any visible enhancement of tumor. **C**. Unenhanced MRI shows that HCC is of similar intensity to liver. **D**. Arterial phase MRI shows viable hypervascular tissue at the periphery of HCC (*arrows*).

### <span id="page-9-1"></span>**4. REPORTING FINDINGS SUSPICIOUS FOR HCC**

<span id="page-9-0"></span>The success of screening for HCC depends on detecting the tumor while it can still be treated, without resulting in a frequency of false-negative results that could undermine funding or compliance. Thus far, there is no sufficient data to determine whether repeated imaging at 6-month intervals is superior to annual imaging *[\(82](#page-17-0)*, *[83\)](#page-17-1)*. Our approach has been to attempt confident noninvasive diagnosis with high accuracy while minimizing the frequency of "overdiagnosis" of benign enhancing lesions as HCC. A recent study at our center confirmed that small HCCs that were initially diagnosed as probably benign did not progress to untreatable HCC if a patient adhered to an annual surveillance program *[\(33\)](#page-14-3)*. To maximize the utility of an MRI-based screening program for HCC among high-risk individuals, we recommend use of the following overall categories for reporting suspicion of HCC.



<span id="page-10-0"></span>**Fig. 3.** HCC visible by MRI but not three-phase CT. **A**–**C**. CT images prior to and during arterial and venous phases of contrast enhancement. **D**. T2-weighted MR image shows hyperintense HCC. **E**. T1-weighted MR image shows hypointense HCC. **F**. As in E, immediately after gadolinium contrast agent administration shows moderately hypervascular HCC. **G**. As in E, about 1 minute after gadolinium contrast agent administration does not show the HCC, similar to CT.

#### *4.1. No Nodule with High Suspicion of HCC*

These are patients with hepatitis C or other clinical condition that renders them of high risk for developing HCC. This is not changed if low probability lesions, such as subcentimeter foci of transient enhancement, are present. These patients should have repeated examinations at regular intervals, although we are not aware of any data to establish whether 6 month or 12-month intervals are preferable. Although a small minority of subcentimeter transiently enhancing foci may indeed be HCC, well over 90% are benign. If each of these low probability foci triggers a shortterm follow-up examination, the overall cost of the screening program may increase geometrically. It is also likely that increasing the frequency of short-term follow-ups may adversely affect overall execution and compliance with the screening program. We therefore recommend that "overcalling" tiny enhancing foci be minimized, provided that these patients are still imaged with a frequency of at least one MRI examination per year *[\(33\)](#page-14-3)*.

#### *4.2. Indeterminate Nodule*

These are usually nodules larger than 1 cm, or other imaging characteristic to generate more than a low probability level of confidence. A diameter of greater than 1 cm is important for two separate reasons. Benign enhancing nodules are usually less than 1 cm in diameter, so larger size by itself raises the possibility of HCC. Additionally, the danger of "under-calling" lesions larger than 1 cm is that tumor doubling will have a more adverse affect if the nodule is already greater than 1 cm. The goal of a screening program is to detect a nodule while it still can be treated optimally. As a mass exceeds 2 cm and becomes progressively larger, the possibility of unsuccessful treatment increases.

An indeterminate nodule will usually trigger a short-term follow-up. The recommended interval will depend on the level of concern regarding rapid interval growth. Typically, the interval recommended will be between 6 and 12 weeks. Alternatively, an ultrasound with potential biopsy may be recommended. If a nodule is visible by sonography as a distinct hypoechoic nodule, this increases the likelihood that it is HCC. Ultrasound may then be used to guide biopsy, if its location renders it accessible. It must be recognized, however, that guided biopsy may be false negative due either to sampling error or to occasional similarity between welldifferentiated HCC and benign liver tissue. Therefore, negative results of a biopsy of indeterminate nodule should still trigger short-term imaging follow-up.

#### *4.3. Probable HCC*

These will be distinct nodules which are visible on more than one pulse sequence. Their distinction vs. the next category of risk will depend largely on the expertise and experience of the interpreting radiologists, as well as the quality of the MRI examination.

# *4.4. HCC*

For these lesions, the characteristics of HCC are sufficiently clear that there is no reasonable doubt as to the diagnosis. It is becoming standard practice that a confident diagnosis from a reliable radiologist can be used to direct management decisions regarding HCC, in the absence of tissue diagnosis. In some instances, biopsy or documented rising α-fetal protein levels might be insisted upon, such as to list for transplantation, if the nodule is less than 2 cm diameter.

#### **5. CONCLUDING STATEMENTS**

<span id="page-12-0"></span>Hopefully, the above discussions will help to improve communications between the various physicians involved in managing patients with HCC with regard to their diagnosis by MRI. As official criteria for assigning priority for liver transplant evolve, standards for reporting measurements of size and number may change. Regardless, it is important that all those involved in interpreting MR images and generating their reports are fully cognizant of the affects of these reports on patients' categories for prioritization.

#### **REFERENCES**

- <span id="page-12-1"></span>1. DiBisceglie AM, Thompson J, Smith-Wilkaitis N, Brunt EM, Bacon BR. Combination of interferon and ribavirin in chronic hepatitis C: re-treatment of nonresponders to interferon. Hepatology 2001; 33:704–707.
- <span id="page-12-2"></span>2. Arguedas MR, Chen VK, Eloubeidi MA, Fallon MB. Screening for hepatocellular carcinoma in patients with hepatitis C cirrhosis: a cost-utility analysis. Am J Gastroenterol 2003; 98:679–690.
- <span id="page-12-5"></span>3. Kim AI, Saab S. Treatment of hepatitis C. Am J Med 2005; 118:808–815.
- 4. Bruix J, Hessheimer AJ, Forner A, Boix L, Vilana R, Llovet JM. New aspects of diagnosis and therapy of hepatocellular carcinoma. Oncogene 2006; 25:3848–3856.
- <span id="page-12-3"></span>5. Snowberger N, Chinnakotla S, Lepe RM, et al. Alpha fetoprotein, ultrasound, computerized tomography and magnetic resonance imaging for detection of hepatocellular carcinoma in patients with advanced cirrhosis. Aliment Pharmacol Ther 2007; 26:1187–1194.
- <span id="page-12-4"></span>6. de\_Ledinghen V, Laharie D, Lecesne R, et al. Detection of nodules in liver cirrhosis: spiral computed tomography or magnetic resonance imaging? A prospective study of 88 nodules in 34 patients. Eur J Gastroenterol Hepatol 2002; 14:159–165.
- 7. Chalasani N, Horlander JC, Sr., Said A, et al. Screening for hepatocellular carcinoma in patients with advanced cirrhosis. Am J Gastroenterol 1999; 94:2988–2993.
- 8. Bruix J, Sherman M. Management of hepatocellular carcinoma. Hepatology 2005; 42:1208–1236.
- 9. Kudo M, Okanoue T. Management of hepatocellular carcinoma in Japan: consensusbased clinical practice manual proposed by the Japan Society of Hepatology. Oncology 2007; 72 Suppl 1:2–15.
- <span id="page-13-0"></span>10. Forner A, Vilana R, Ayuso C, et al. Diagnosis of hepatic nodules 20 mm or smaller in cirrhosis: Prospective validation of the noninvasive diagnostic criteria for hepatocellular carcinoma. Hepatology 2008; 47:97–104.
- <span id="page-13-1"></span>11. Llovet JM, Mas X, Aponte JJ, et al. Cost effectiveness of adjuvant therapy for hepatocellular carcinoma during the waiting list for liver transplantation. Gut 2002; 50:123–128.
- <span id="page-13-2"></span>12. Mazzaferro V, Battiston C, Perrone S, et al. Radiofrequency ablation of small hepatocellular carcinoma in cirrhotic patients awaiting liver transplantation: a prospective study. Ann Surg 2004; 240:900–909.
- 13. Ikai I, Arii S, Ichida T, et al. Report of the 16th follow-up survey of primary liver cancer. Hepatol Res 2005.
- 14. Crocetti L, Lencioni R. Thermal ablation of hepatocellular carcinoma. Cancer Imaging 2008; 8:19–26.
- 15. Takayama T, Makuuchi M, Kojiro M, et al. Early hepatocellular carcinoma: pathology, imaging, and therapy. Ann Surg Oncol 2008; 15:972–978.
- 16. Yamakado K, Nakatsuka A, Takaki H, et al. Early-stage hepatocellular carcinoma: radiofrequency ablation combined with chemoembolization versus hepatectomy. Radiology 2008; 247:260–266.
- 17. Cheng BQ, Jia CQ, Liu CT, et al. Chemoembolization combined with radiofrequency ablation for patients with hepatocellular carcinoma larger than 3 cm: a randomized controlled trial. Jama 2008; 299:1669–1677.
- <span id="page-13-3"></span>18. Wang J, Chen LT, Tsang YM, Liu TW, Shih TT. Dynamic contrast-enhanced MRI analysis of perfusion changes in advanced hepatocellular carcinoma treated with an antiangiogenic agent: a preliminary study. AJR Am J Roentgenol 2004; 183:713–719.
- <span id="page-13-4"></span>19. Zhu AX, Abou-Alfa GK. Expanding the treatment options for hepatocellular carcinoma: combining transarterial chemoembolization with radiofrequency ablation. Jama 2008; 299:1716–1718.
- 20. Hakime A, Hines-Peralta A, Peddi H, et al. Combination of radiofrequency ablation with antiangiogenic therapy for tumor ablation efficacy: study in mice. Radiology 2007; 244:464–470.
- <span id="page-13-5"></span>21. Pal S, Pande GK. Current status of surgery and transplantation in the management of hepatocellular carcinoma: an overview. J Hepatobiliary Pancreat Surg 2001; 8: 323–336.
- <span id="page-13-6"></span>22. Ikai I, Arii S, Kojiro M, et al. Reevaluation of prognostic factors for survival after liver resection in patients with hepatocellular carcinoma in a Japanese nationwide survey. Cancer 2004; 101:796–802.
- 23. Llovet JM, Schwartz M, Fuster J, Bruix J. Expanded criteria for hepatocellular carcinoma through down-staging prior to liver transplantation: not yet there. Semin Liver Dis 2006; 26:248–253.
- 24. Burrel M, Llovet JM, Ayuso C, et al. MRI angiography is superior to helical CT for detection of HCC prior to liver transplantation: an explant correlation. Hepatology 2003; 38:1034–1042.
- <span id="page-13-9"></span>25. Zhao H, Yao JL, Wang Y, Zhou KR. Detection of small hepatocellular carcinoma: comparison of dynamic enhancement magnetic resonance imaging and multiphase multirowdetector helical CT scanning. World J Gastroenterol 2007; 13:1252–1256.
- <span id="page-13-8"></span><span id="page-13-7"></span>26. Mitchell DG. Focal manifestations of diffuse liver disease at MR imaging. Radiology 1992; 185:1–11.
- 27. Ohtomo K, Baron RL, Dodd GD, 3rd, Federle MP, Ohtomo Y, Confer SR. Confluent hepatic fibrosis in advanced cirrhosis: evaluation with MR imaging. Radiology 1993; 189:871–874.
- <span id="page-14-0"></span>28. Ito K, Choji T, Fujita F, Matsumoto T, Nakada T, Nakanishi T. Early-enhancing pseudolesion in medial segment of left hepatic lobe detected with multisection dynamic MR. Radiology 1993; 187:695–699.
- <span id="page-14-1"></span>29. Jeong YY, Mitchell DG, Kamishima T. Small (<20 mm) enhancing hepatic nodules seen on arterial phase MR imaging of the cirrhotic liver: clinical implications. Am J Roentgenol 2002; 178:1327–1334.
- <span id="page-14-14"></span>30. Ito K, Fujita T, Shimizu A, et al. Multiarterial phase dynamic MRI of small early enhancing hepatic lesions in cirrhosis or chronic hepatitis: differentiating between hypervascular hepatocellular carcinomas and pseudolesions. Am J Roentgenol 2004; 183:699–705.
- <span id="page-14-15"></span>31. Marrero JA, Hussain HK, Nghiem HV, Umar R, Fontana RJ, Lok AS. Improving the prediction of hepatocellular carcinoma in cirrhotic patients with an arterially-enhancing liver mass. Liver Transplantation: Official Publication of The American Association for the Study of Liver Diseases and the International Liver Transplantation Society 2005; 11:281–289.
- <span id="page-14-13"></span>32. Shimizu A, Ito K, Koike S, Fujita T, Shimizu K, Matsunaga N. Cirrhosis or chronic hepatitis: evaluation of small  $( $or=2$ -cm) early-enhancing hepatic lesions with serial$ contrast-enhanced dynamic MR imaging. Radiology 2003; 226:550–555.
- <span id="page-14-2"></span>33. Choi D, Mitchell DG, Verma SK, et al. Hepatocellular carcinoma with indeterminate or false-negative findings at initial MR imaging: effect on eligibility for curative treatment initial observations. Radiology 2007; 244:776–783.
- <span id="page-14-3"></span>34. Kageyama F, Kobayashi Y, Kawasaki T, et al. An unusual hyperplastic hepatocellular nodule in a patient with hepatitis C virus-related liver cirrhosis. Am J Gastroenterol 1998; 93:2588–2593.
- <span id="page-14-4"></span>35. Takahashi S, Miyanishi K, Takada K, et al. Case report of a focal nodular hyperplasialike nodule present in cirrhotic liver. Hepatol Res 2008; 38:521–528.
- <span id="page-14-8"></span>36. Lee YH, Kim SH, Cho MY, Shim KY, Kim MS. Focal nodular hyperplasia-like nodules in alcoholic liver cirrhosis: radiologic–pathologic correlation. AJR Am J Roentgenol 2007; 188:W459–463.
- 37. Nakashima O, Kurogi M, Yamaguchi R, et al. Unique hypervascular nodules in alcoholic liver cirrhosis: identical to focal nodular hyperplasia-like nodules? J Hepatol 2004; 41:992–998.
- <span id="page-14-9"></span>38. Soyer P, Lacheheb D, Caudron C, Levesque M. MRI of adenomatous hyperplastic nodules of the liver in Budd-Chiari syndrome. J Comput Assist Tomogr 1992; 17: 86–89.
- 39. Siegelman ES, Outwater EK, Furth EE, Rubin R. MR imaging of hepatic nodular regenerative hyperplasia. J Mag Res Imag 1995; 5:730–732.
- <span id="page-14-5"></span>40. Wanless IR, et al. Terminology of nodular hepatocellular lesions. International Working Party. Hepatology (Baltimore, Md.) 1995; 22:983–993.
- <span id="page-14-6"></span>41. Wanless IR, Mawdsley C, Adams R. On the pathogenesis of focal nodular hyperplasia of the liver. Hepatology 1985; 5:1194–1200.
- <span id="page-14-7"></span>42. Wu TT, Boitnott J. Dysplastic nodules: a new term for premalignant hepatic nodular lesions. Radiology 1996; 201:21–22.
- <span id="page-14-10"></span>43. Krinsky GA, Lee VS, Theise ND, et al. Hepatocellular carcinoma and dysplastic nodules in patients with cirrhosis: prospective diagnosis with MR imaging and explantation correlation. Radiology 2001; 219:445–454.
- <span id="page-14-12"></span><span id="page-14-11"></span>44. Mitchell DG, Palazzo J, Hann HW, Rifkin MD, Burk DL, Jr., Rubin R. Hepatocellular tumors with high signal on T1-weighted MR images: chemical shift MR imaging and histologic correlation. J Comput Assist Tomogr 1991; 15:762–769.
- 45. Shimizu A, Ito K, Sasaki K, et al. Small hyperintense hepatic lesions on T1-weighted images in patients with cirrhosis: evaluation with serial MRI and imaging features for clinical benignity. Magn Reson Imaging 2007.
- <span id="page-15-0"></span>46. Siegelman ES. MR imaging of diffuse liver disease. Hepatic fat and iron. Magn Reson Imag Clin N Am 1997; 5:347–365.
- <span id="page-15-1"></span>47. Mitchell DG, Kim I, Chang TS, et al. Fatty liver. Chemical shift phase-difference and suppression magnetic resonance imaging techniques in animals, phantoms, and humans. Invest Radiol 1991; 26:1041–1052.
- 48. Rinella ME, McCarthy R, Thakrar K, et al. Dual-echo, chemical shift gradient-echo magnetic resonance imaging to quantify hepatic steatosis: Implications for living liver donation. Liver Transplantation: Official Publication of the American Association for the Study of Liver Diseases and the International Liver Transplantation Society 2003; 9:851–856.
- <span id="page-15-2"></span>49. Runge VM. A comparison of two MR hepatobiliary gadolinium chelates: Gd-BOPTA and Gd-EOB-DTPA. J Comput Assist Tomogr 1998; 22:643–650.
- <span id="page-15-3"></span>50. Vogl TJ, Stupavsky A, Pegios W, et al. Hepatocellular carcinoma: evaluation with dynamic and static gadobenate dimeglumine-enhanced MR imaging and histopathologic correlation. Radiology 1997; 205:721–728.
- 51. Giovagnoni A, Paci E. Liver III: Gadolinium-based hepatobiliary contrast agents (Gd-EOB-DTPA and Gd-BOPTA/Dimeg). MRI Clin NA 1996; 4:61–72.
- <span id="page-15-4"></span>52. Okada Y, Ohtomo K, Kiryu S, Sasaki Y. Breath-hold T2-weighted MRI of hepatic tumors: value of echo planar imaging with diffusion-sensitizing gradient. J Comput Assist Tomogr 1998; 22:364–371.
- <span id="page-15-5"></span>53. Koinuma M, Ohashi I, Hanafusa K, Shibuya H. Apparent diffusion coefficient measurements with diffusion-weighted magnetic resonance imaging for evaluation of hepatic fibrosis. J Magn Reson Imag 2005; 22:80–85.
- 54. Boulanger Y, Amara M, Lepanto L, et al. Diffusion-weighted MR imaging of the liver of hepatitis C patients. NMR Biomed 2003; 16:132–136.
- 55. Namimoto T, Yamashita Y, Sumi S, Tang Y, Takahashi M. Focal liver masses: Characterization with diffusion-weighted echo-planar MR imaging. Radiology 1997; 204:739– 744.
- 56. Xu H, Li X, Xie JX, Yang ZH, Wang B. Diffusion-weighted magnetic resonance imaging of focal hepatic nodules in an experimental hepatocellular carcinoma rat model. Acad Radiol 2007; 14:279–286.
- 57. Taouli B, Losada M, Holland A, Krinsky G. Magnetic resonance imaging of hepatocellular carcinoma. Gastroenterology 2004; 127:S144–152.
- <span id="page-15-6"></span>58. Taylor-Robinson SD. Applications of magnetic resonance spectroscopy to chronic liver disease. Clin Med 2001; 1:54–60.
- <span id="page-15-7"></span>59. Pandharipande PV, Krinsky GA, Rusinek H, Lee VS. Perfusion imaging of the liver: current challenges and future goals. Radiology 2005; 234:661–673.
- <span id="page-15-8"></span>60. Delorme S, Knopp MV. Non-invasive vascular imaging: assessing tumour vascularity. Eur Radiol 1998; 8:517–527.
- 61. Sahani DV, Holalkere NS, Mueller PR, Zhu AX. Advanced hepatocellular carcinoma: CT perfusion of liver and tumor tissue – initial experience. Radiology 2007; 243:736– 743.
- 62. Miyazaki K, Collins DJ, Walker-Samuel S, et al. Quantitative mapping of hepatic perfusion index using MR imaging: a potential reproducible tool for assessing tumour response to treatment with the antiangiogenic compound BIBF 1120, a potent triple angiokinase inhibitor. Eur Radiol 2008.
- <span id="page-15-10"></span><span id="page-15-9"></span>63. Stark DD, Weissleder R, Elizondo G, et al. Superparamagnetic iron oxide: clinical application as a contrast agent for MR imaging of the liver. Radiology 1988; 168:297–301.
- 64. Yamamoto H, Yamashita Y, Yoshimatsu S, et al. Hepatocellular carcinoma in cirrhotic livers: detection with unenhanced and iron oxide-enhanced MR imaging. Radiology 1995; 195:106–112.
- 65. Imai Y, Murakami T, Hori M, et al. Hypervascular hepatocellular carcinoma: Combined dynamic MDCT and SPIO-enhanced MRI versus combined CTHA and CTAP. Hepatol Res 2008; 38:147–158.
- <span id="page-16-0"></span>66. Hanna RF, Kased N, Kwan SW, et al. Double-contrast MRI for accurate staging of hepatocellular carcinoma in patients with cirrhosis. AJR Am J Roentgenol 2008; 190: 47–57.
- <span id="page-16-1"></span>67. Semelka RC, Lee JK, Worawattanakul S, Noone TC, Patt RH, Ascher SM. Sequential use of ferumoxide particles and gadolinium chelate for the evaluation of focal liver lesions on MRI. J Magnet Reson Imag 1998; 8:670–674.
- 68. Ward J, Guthrie JA, Scott DJ, et al. Hepatocellular carcinoma in the cirrhotic liver: double-contrast MR imaging for diagnosis. Radiology 2000; 216:154–162.
- 69. Ward J, Robinson PJ. How to detect hepatocellular carcinoma in cirrhosis. Eur Radiol 2002; 12:2258–2272.
- 70. Bolog N, Pfammatter T, Mullhaupt B, Andreisek G, Weishaupt D. Double-contrast magnetic resonance imaging of hepatocellular carcinoma after transarterial chemoembolization. Abdom Imag 2007; 33:313–323.
- 71. Kim YK, Kwak HS, Han YM, Kim CS. Usefulness of combining sequentially acquired gadobenate dimeglumine-enhanced magnetic resonance imaging and resovist-enhanced magnetic resonance imaging for the detection of hepatocellular carcinoma: comparison with computed tomography hepatic arteriography and computed tomography arterioportography using 16-slice multidetector computed tomography. J Comput Assist Tomogr 2007; 31:702–711.
- <span id="page-16-2"></span>72. Ebara M, Watanabe S, Kita K, et al. MR imaging of small hepatocellular carcinoma: Effect of intratumoral copper content on signal intensity. Radiology 1991; 180: 617–621.
- <span id="page-16-3"></span>73. Kitagawa K, Matsui O, Kadoya M, et al. Hepatocellular carcinomas with excessive copper accumulation: CT and MR findings. Radiology 1991; 180:623–628.
- 74. Matsuzaki K, Sano N, Hashiguchi N, Yoshida S, Nishitani H. Influence of copper on MRI of hepatocellular carcinoma. J Magn Reson Imaging 1997; 7:478–481.
- <span id="page-16-4"></span>75. Hussain HK, Syed I, Nghiem HV, et al. T2-weighted MR imaging in the assessment of cirrhotic liver. Radiology 2004; 230:637–644.
- <span id="page-16-5"></span>76. Choi BI, Lee GK, Kim ST, et al. Mosaic pattern of encapsulated hepatocellular carcinoma: correlation of magnetic resonance imaging and pathology. Gastrointest Radiol 1990; 15:238–240.
- <span id="page-16-6"></span>77. Terada T, Nakanuma Y. Iron-negative foci in siderotic macroregenerative nodules in human cirrhotic liver. Arch Pathol Lab Med 1989; 113:916–920.
- <span id="page-16-7"></span>78. Mitchell DG, Rubin R, Siegelman ES, Burk DL, Jr., Rifkin MD. Hepatocellular carcinoma within siderotic regenerative nodules: appearance as a nodule within a nodule on MR images. Radiology 1991; 178:101–103.
- 79. Sadek AG, Mitchell DG, Siegelman ES, Outwater EK, Matteucci T, Hann HW. Early hepatocellular carcinoma that develops within macroregenerative nodules: growth rate depicted at serial MR imaging. Radiology 1995; 195:753–756.
- <span id="page-16-8"></span>80. Horigome H, Nomura T, Saso K, Itoh M, Joh T, Ohara H. Limitations of imaging diagnosis for small hepatocellular carcinoma: comparison with histological findings. Journal of Gastroenterology and Hepatology 1999; 14:559–565.
- <span id="page-16-10"></span><span id="page-16-9"></span>81. Holland AE, Hecht EM, Hahn WY, et al. Importance of small ( $\lt$  or  $=$  20-mm) enhancing lesions seen only during the hepatic arterial phase at MR imaging of the cirrhotic liver: evaluation and comparison with whole explanted liver. Radiology 2005; 237:938–944.
- 82. Trevisani F, De NS, Rapaccini G, et al. Semiannual and annual surveillance of cirrhotic patients for hepatocellular carcinoma: effects on cancer stage and patient survival (Italian experience). Am J Gastroenterol 2002; 97:734–744.
- <span id="page-17-1"></span><span id="page-17-0"></span>83. Santagostino E, Colombo M, Rivi M, et al. A 6-month versus a 12-month surveillance for hepatocellular carcinoma in 559 hemophiliacs infected with the hepatitis C virus. Blood 2003; 102:78–82.