

Chapter 24

Computational Fractal-Based Analysis of Brain Tumor Microvascular Networks

Antonio Di Ieva and Omar S. Al-Kadi

Abstract Brain parenchyma microvasculature is set in disarray in the presence of tumors, and malignant brain tumors are among the most vascularized neoplasms in humans. As microvessels can be easily identified in histologic specimens, quantification of microvasculature can be used alone or in combination with other histological features to increase the understanding of the dynamic behavior, diagnosis, and prognosis of brain tumors. Different brain tumors, and even subtypes of the same tumor, show specific microvascular patterns, as a kind of “microvascular fingerprint,” which is particular to each histotype. Reliable morphometric parameters are required for the qualitative and quantitative characterization of the neoplastic angio-architecture, although the lack of standardization of a technique able to quantify the microvascular patterns in an objective way has limited the “morphometric approach” in neuro-oncology.

In this chapter we focus on the importance of the computational-based morphometrics, for the objective description of the tumoral microvascular fingerprinting. By also introducing the concept of “angio-space,” which is the tumoral space occupied by the microvessels, we here present fractal analysis as the most reliable computational tool able to offer objective parameters for the description of the microvascular networks.

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The spectrum of different angioarchitectural configurations can be quantified by means of Euclidean and fractal-based parameters in a multiparametric analysis, aimed to offer surrogate biomarkers of cancer. Such parameters are here described from the methodological point of view (i.e., feature extraction) as well as from the clinical perspective (i.e., relation to underlying physiology), in order to offer new computational parameters to the clinicians with the final goal of improving diagnostic and prognostic power of patients affected by brain tumors.

Keywords Angioarchitecture • Brain tumor • Fractal dimension • Fractal analysis • Glioblastoma multiforme • Microvasculature

24.1 Introduction

The brain is among the most perfused organs of the human body, and in order to meet its high metabolic demand, it has one of the most complex vascular and microvascular systems. Different regions of the brain are characterized by specific microvascular networks [8, 22–24, 38] whose level of complexity most likely reflects the function related to that specific area [11]. Since tumors are supported by different patterns of vascularization [34], it is possible to speculate that tumorigenesis (i.e., the sequential accumulation of mutations within tissue cells) is related to the disarray of the physiological microvascular bed or even related to a programmed rearrangement of a new and specific microvascular network. The study of the neoplastic angioarchitecture seems to be of paramount importance not only for research purposes but for clinical applications as well since it has been shown that the “normalization” of the abnormal microvascular architecture of tumors by means of anti-angiogenic agents, for example, can reduce regional hypoxia and eventually increase the efficacy of therapies [32, 33]. According to this model, the imbalance between pro- and anti-angiogenic factors in a tumor disorganizes the angioarchitecture, which might be brought back from its irregular pathological state to a physiologic state and then “normalized.” Fractal analysis offers several tools for the quantification and temporal follow-up of such angiostructural changes [25, 44].

24.2 Brain Tumors and Vascularization

The latest available World Health Organization (WHO 2007¹) grading system classifies brain tumors according to histological features in a sort of benign-to-malignant gradient (from grade I to grade IV, with grade IV being the most malignant) [41].

¹At the time of the chapter writing, the WHO classification system for brain tumors published in 2016 was not available yet.

The grading has diagnostic, prognostic, and therapeutic value, considering that grade I lesions have low proliferative and infiltrative potential, and can be virtually cured by means of surgery and/or radiosurgery alone, while grade IV cancer requires multimodal approaches (i.e., surgery followed by radio-chemotherapy) and has fatal prognosis.

Malignant brain tumors are among the most vascularized tumors in humans [41]. As microvessels can be easily identified in biopsy specimens, it has been suggested that quantification of microvasculature might be used alone or in combination with other histological features to increase the understanding of the dynamic behavior and prognosis of brain tumors, as well as for diagnostic purposes. However, microvasculature is not always considered a feature related to brain tumor grading. Microvascular proliferation is considered a histopathological hallmark of glioblastoma multiforme (GBM) [41], which is the most frequent and malignant glioma (WHO grade IV), but the differences in microvasculature between grade II and III gliomas, for example, are less clear and not used for grading. Also, meningiomas of different grades, as well as meningiomas in comparison to hemangiopericytomas, clearly display different vascular patterns (see Table 24.1), but other features are still used to differentiate the different grades and/or tumors [2]. This is also related to the lack of standardization of a technique able to quantify the microvascular patterns in an objective way.

The vascularization of malignant brain tumors is the target for anti-angiogenic treatment (such as bevacizumab), providing reasoning on the importance of the different methods in quantifying and assessing the changes in tumor microvasculature. On one hand, it is true that different brain tumors show different angiogenic patterns, but on the other it seems that single histotypes, such as GBM, show a very highly heterogeneous microangioarchitecture [5, 17, 18, 45]. We here summarize the concepts and methods related to microvascular detection and image analysis for the following morphometric analyses.

24.2.1 Immunohistochemistry (IHC)

In histological specimens, microvessels can be detected by means of IHC. Several antibodies have been described for the immunodetection of the endothelium, each one with its own pros and cons, summarized in Table 24.2. The most commonly

Table 24.1 Examples of brain tumors showing peculiar microvasculatures

Different brain tumors	Same brain tumors of different grades
Gliomas, gliosarcomas, metastases, lymphomas	Gliomas: grades I to IV
Meningiomas vs. hemangiopericytomas	Meningiomas: grades I to III
Pituitary adenomas vs. normal pituitary gland	

Table 24.2 Vascular markers commonly used in immunohistochemistry

Antibody	Description	Pros	Cons
CD31	Also known as platelet-endothelial cell adhesion molecule (PECAM-1); membrane-bound glycoprotein and member of the immunoglobulin family	Stains vessels in both neoplastic and healthy tissue; sufficient sensitivity for blood vessels of all sizes	Frequent antigen loss during retrieval
CD34	Endothelial membrane-bound glycoprotein	Stains blood vessels of all sizes	Pan-endothelial: stains newly formed as well as preexisting vessels, making it ineffective for visualizing only angiogenesis. May lack sensitivity in some tissues
CD105	Endothelial membrane-bound glycoprotein; essential for angiogenesis. Also called endoglin	Stains only newly-formed blood vessels during tumorigenesis; good for visualizing angiogenesis	Ineffective as pan-endothelial marker; weak or no staining in normal tissue
Factor VIII	Glycoprotein polymer involved in platelet aggregation; comprised of von Willebrand factor and smaller antihemophilic factor	Effective for larger vessels staining	Lacks sensitivity for smaller vessels in comparison of other antibodies
Endocan	Cell-specific-molecule-1, proteoglycan secreted by endothelial cells; regulated by vascular endothelial growth factor and related to angiogenesis	Marker of small and large neoangiogenic vessels	Scattered immunostaining of vessels; generally not expressed in non-tumoral tissues

See review in Di Ieva et al. [21]

used are CD34 and CD31 (CD refers to “cluster of differentiation”), which are considered pan-endothelial markers, while CD105 and Endocan are more commonly used for newly formed vessels (i.e., in the process of angiogenesis). Factor VIII and Ulex europeus agglutinin I (UEAI) have almost fallen into disuse due to their multiple limitations. The ratios of CD34:CD105, CD34:Endocan, or CD31:Endocan immunoreactive vessels can provide the “angiogenic fraction” of the tumor [Di Ieva, unpublished results].

24.3 Morphometrics of Microvasculature

Qualitative rather than quantitative analyses have generally been used to describe the differences in the vascularization of the normal brain vs. brain tumors.

It has been suggested that vessel quantification can be added into the tumor classification system for grading as well as for prognostication. If the aim is to provide a morphometric biomarker (i.e., predicting patient prognosis, treatment response, etc.), the limitations are due to the lack of standardization of the proposed parameters. Morphometrics aims to reduce shapes to numbers, offering a reductionist approach to describe the geometrical complexity of natural objects. In pathology, morphometric analyses add a quantitative element to the qualitative description of the tissue [11], generating continuous variables, which can be used for statistical comparison.

Defining the “tumor-space” as the neoplastic volume, the “angio-space” refers to the proportion of microvessels filling such volume, not just in terms of quantity but also in terms of size, shape, and pattern of distribution [11].

Pathologists recognize distinct vascular angiogenic subtypes, described as palisades, glomeruloid vascular proliferation, vascular garlands, vascular clusters, and microvascular sprouting, and may also use semiquantitative scores to quantify the level of “clusterization” of the microvessels [20] (see Table 24.3). This seems to be relevant, especially when considering the relationship between vascular patterns and clinical outcomes, as different angiogenic protein expression associated with different angioarchitecture can respond in different ways to chemotherapy [5]. These descriptions lack standardization and some intra- and interobserver variability may be present.

Since the introduction of the use of the vessel density in 1972 [6], several other parameters have been proposed as potential “quantitators” of the microvascular system, used to quantify the angio-space (Tables 24.3 and 24.4).

We here classify these morphometric parameters in “Euclidean-based” and “fractal-based” parameters.

Table 24.3 Morphometric parameters used to quantify the angio-space

Euclidean-based parameters	Fractal-based parameters
Microvessel density (MVD)	Fractal dimension (FD), microvascular fractal dimension (mvFD)
Density/number of vessels	Local box-counting dimension
Total vascular area (TVA)	Tortuosity
Local vascular area (in the hot spot)	Lacunarity
Perimeter	Representative elementary volume (REV) length scale
Mean diameter of the microvessels	Hurst exponent
Branching count	
Major and minor axis length	
Pathologist-based analysis	Other non-Euclidean parameters
Semiquantitative scoring systems, that is, 0 (no clusters), 1 (clusters), 2 (very prominent clusters)	Compactness
	Shape factor
	Distance maps
	Microvascular structural entropy

Table 24.4 Morphometric parameters used to quantify the microvascular pattern identified in histological specimens of grade II and III gliomas

Angioarchitectural morphometric parameters	
A_{rum}	Area covered by the specimen of brain tumor on the histological section
TVA	Total vascular area, the area of the microvessels of the whole specimen
A%	Ratio between TVA and A_{rum}
MVs/mm ²	Microvascular density expressed as the mean number of microvessels per mm ²
Hot spot	1 mm ² area of the specimen having the highest local A(%)
Local A(%)	The % vascular area in the hot spot
mvFD	Microvascular fractal dimension: monofractal dimension of the microvascular pattern of the whole specimen (in a pre-determined scaling window)
loc mvFD	Local mvFD: monofractal dimension of the microvascular pattern expressed in the hot spot area (scaling window, $\epsilon_{min} = 1 \mu\text{m}$; $\epsilon_{max} = 100 \mu\text{m}$)
loc bcD	Local box-counting dimension: value of fractal dimension estimated by the box-counting method in a non-monofractal range (scaling window, $\epsilon_{min} = 1 \mu\text{m}$; $\epsilon_{max} = 1,000 \mu\text{m}$)

From Di Ieva et al. [20]

24.3.1 Euclidean-Based Parameters

One of the most used parameters is the microvessel density (MVD), based on the measure of the number of immunostained microvessels per square millimeter in the most densely vascularized area of the histological specimen (termed “hot spot”). Different techniques have been proposed for the MVD calculation, and several seminal papers have investigated its role in every human tumor (see review in [14]), including the brain and pituitary tumors (reviews in [13, 14]), but in conclusion it has been shown that it is not a valid measure for guiding and/or evaluating anti-angiogenic treatment [31], nor for distinguishing different grades of brain tumors. All the histological analyses are limited by the sampling, especially in GBMs, which show a very high histologic heterogeneity. The greatest limitation of the use of the MVD is the choice of the hot spot, due to (a) high inter- and intra-observer variability in its selection, (b) potential lack of representativity of the whole specimen, and (c) potential lack of representation of a complex 3D structure, such as the microvascular tree, in a 2D histological area [15, 18, 43].

Other Euclidean parameters can be computed on the histological specimens of brain tumors (see Table 24.3). Multiparametric analyses of combinations of such indices have shown microvascular morphometrics as a valid tool to differentiate different grades of brain tumors [18, 37, 45].

The greatest limitation of the described parameters is that Euclidean geometry can only quantify regular and smooth objects, and therefore can only offer approximations of the roughness expressed by natural objects, such as the microvascular trees. The complexity of vascular systems depends on (a) the number of vessels, (b) their size and shape, and (c) the pattern of the vessel distribution and the nonlinear, temporal, and spatial advance of the promotion, progress, mediation, and inhibition

of angiogenesis generating the complex ramified network irregularly filling the environment surrounding the tumor, which can be described by Euclidean geometry only with great approximation [28].

Fractal geometry overcomes the limits of Euclidean parameters. The fractal-based parameters are described next.

24.3.2 Image Analysis

The first step to apply any morphometric analyses on the neoplastic angio-space is the choice of the technique to visualize the microvessels and the following image analysis, which should be standardized (see Chap. 12) (Fig. 24.1).

When the target of analysis is the microvascular network, several issues have to be considered, for example, whether the analysis is performed on the basis of the space-filling area of the vessels or on skeletonized representations of the microvessels.

In the image analysis, eventual artifacts related to the segmentation of immunoreactive vessels have to be considered. For example, nonspecific immunoreactivity can limit the automatic detection of the vessels, as in the case when antigens diffuse to other compartments (e.g., CD34 antigens which immunostain cell nuclei or necrotic areas rather than endothelial cells). In such cases, a pathologist is still required to identify any artifacts, which should be eliminated from the analysis.

Other digital image acquisition calibration parameters should be standardized as well, in terms of regulating the light intensity of the microscope, maintaining the same resolution over the whole series, and using an identical threshold to extract the

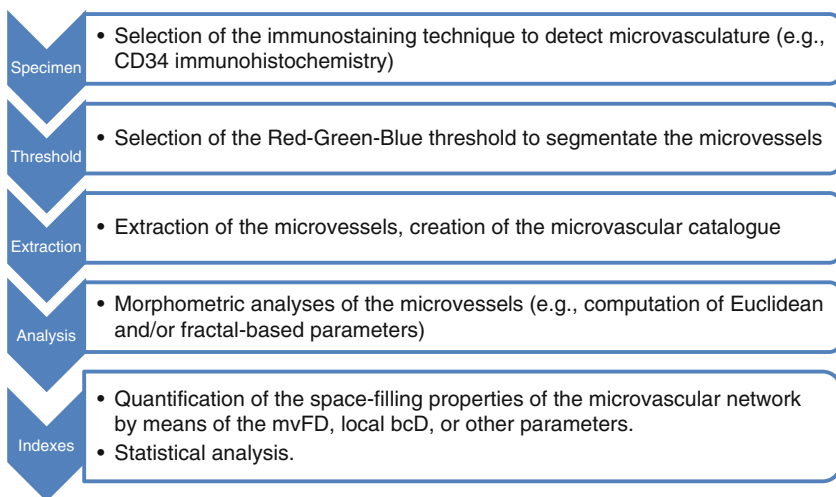


Fig. 24.1 Flowchart used for image and morphometric analyses of microvasculature in histological specimens

immunoreactive vessels. A virtual microscope can be used to digitize the entire specimen rather than few areas selected manually. The vessels can be extracted as a whole from the specimen and organized in a vascular catalog, giving a snapshot on the number and shape of the microvessels within the specimen (Fig. 24.2).

24.4 Fractal-Based Morphometric Analyses of Microvessels

Fractal geometry provides a computer-aided method to describe and quantify the roughness and geometrical complexity of the microvessels distributed within the tumor, as it has been shown in a vast variety of neoplasia, including prostate, kidney, lung, colon, skin, and brain tumors as well. By concentrating on the irregularity of tumor growth rather than individual measure of size (such as diameter and volume or other Euclidean parameters), fractal geometry is well suited to quantify the morphological features that pathologists have long used to describe malignancies in a qualitative way (i.e., wrinkled borders within the host tissue, random microvascular patterns, etc.) [4]. Several researchers have demonstrated the fractality (or “semi-fractality”) and multifractality of the vascular and microvascular trees of many tissues and organs, in physiologic as well as pathologic states, including the human brain [1, 2, 4, 8–10, 26, 27, 29, 35, 40, 44, 47].

The main aim of computational fractal-based analysis of the microvascular patterns is the quantification of parameters, which can be used as surrogate biomarkers.

24.4.1 Microvascular Fractal Dimension (*mvFD*)

The most widely used parameter in fractal geometry is the fractal dimension (FD or D_F), which describes the space-filling properties of irregularly shaped objects [3, 9, 10, 26]. FD has been shown to be the most robust estimator of vascular networks [36, 49]. Extending the use of FD to the quantification of microvessels, the microvascular fractal dimension (*mvFD*) quantifies the space-filling properties of the microvessels within a tissue; in the case of a tumor, *mvFD* quantifies the level of geometrical complexity of the microvessels embedded within the tumor. The *mvFD* adds a qualitative component to the parameterization of the angio-space, trying to answer the following question: assuming an equal number of vessels, what can be said about their shape, size, and pattern of distribution? This means that histological specimens of similar or different tissues can show the same value of MVD or total vascular area, for example, but a very different pattern of distribution, with different values of *mvFD*. Of course, it can also happen that histological tissues with same *mvFD* values differ for the Euclidean parameters, making the approaches complementary methods of quantification. Therefore,



Fig. 24.2 Example of a vascular catalog (in this case, of a glioblastoma multiforme, GBM, with magnification of the vessels in the *inset*). The catalogs give information at a glance on number and shapes of all the microvessels extracted from the entire specimen

mvFD offers useful information about the tumoral space filled by microvessels. The parameter is computer aided, and its objective measure makes it a comparable parameter among different operators. If it is important to analyze the distribution of the vessels within the tissue, by means of FD, for example, it is equally important to analyze complementary parameters, the “vascular gappiness,” that is, the avascular spaces, by means of the lacunarity or other methods, like the distance maps [44].

In a 2D histological specimen, mvFD ranges from 0 to 2. For example, a tumor with a hypothetical mvFD value of 1.95 owns a microvascular network almost filling its area, like the Peano’s curve, with a very complex geometric and highly space-filling microvascular pattern (Fig. 24.3). This microvascular pattern virtually fills the whole tissue in which it is embedded, reducing the avascular spaces to the minimum (therefore increasing the lacunarity) [3] (Fig. 24.4). In the 3D space, such

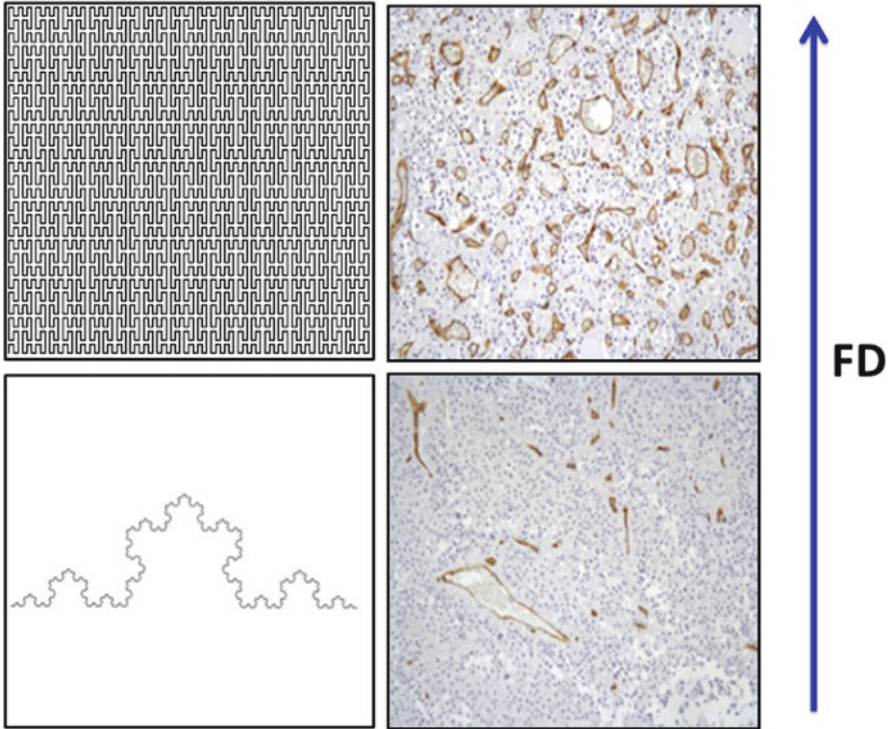


Fig. 24.3 Values of the fractal dimension (FD) on a 2D plane, comparing two fractal objects with the microvasculature (endothelial CD34 immunostaining) of a pituitary adenoma (*lower image*) with the specimen of a normal pituitary gland. The *lower image on the left* shows the Koch's curve ("snowflake"), with $FD = 1.26$. FD increases in more space-filling objects, namely, in the Peano's curve (*higher image on the left*, with $FD \approx 2$) and in the microvasculature of the normal pituitary gland, which is more homogeneously distributed within the parenchyma of the gland

as in electron microscopy reconstructions of the microvascular trees in the 3D volume, FD trends toward a maximal value of 3 [44, 47].

In several kinds of tissues, it has been shown that FD and lacunarity values differ in the physiological and pathological vascular networks [3, 19, 25]. This means that the disarray of the microvascular network, which extends from the normal tissue (e.g., the brain) to the tumor, can be quantified by means of fractal analysis, in a kind of gradient of morphometric differences between tissues. Based on the evidence that the neoplastic microvascular network is abnormal, it has been experimentally shown that the "normalization" of such disarray can be used for therapeutic purposes, as previously quoted, and the FD can be used as a surrogate biomarker for diagnosis and/or treatment follow-up [3, 25]. By means of 2D and 3D analyses, FD values of normal brain microvasculature have been found to be higher than that of neoplastic regions, meaning that the normal brain is better suited to more space-occupying microvasculature [3, 25, 47]. However, due to the

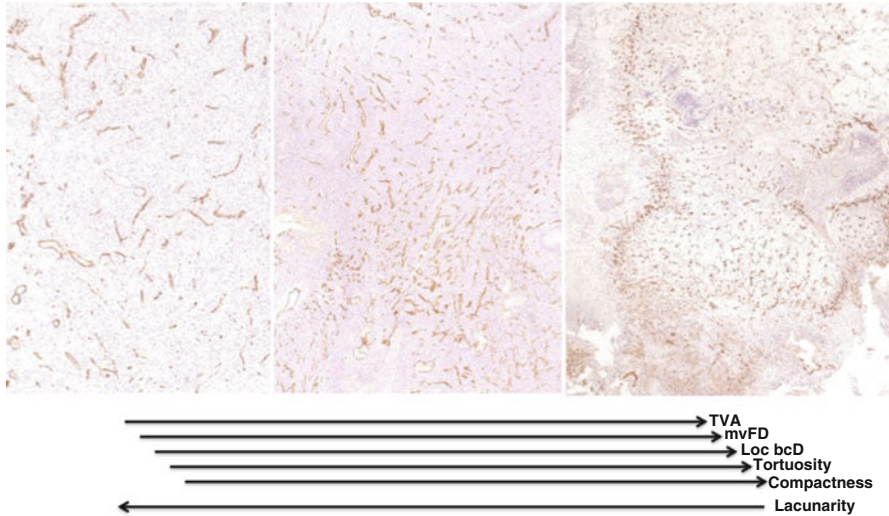


Fig. 24.4 Specimens of three cases of GBM, showing different microvasculature (microvessels detected by using antibodies raised against CD34 with brown coloration). The angio-space of each tumor can be described by means of several parameters (*TVA* total vascular area, *mvFD* microvascular fractal dimension, *loc bcD* local box-counting dimension. See Table 24.4)

use of different methods and techniques, these results are controversial. For example, in one experimental study, the vascular FD values of primate and rat brain tumors have been found to be higher than those of normal brain parenchyma, thus indicating a greater metabolic supply and bigger surface area for gas exchange within the tumors [44]. In numerical terms, fractal-based demonstration that tumoral and normal vascular networks are intrinsically different; Baish et al. introduced a complementary metric to the FD, the tortuosity, defined as the ratio between the minimum vascular path joining two points and their geometric distance from each other [3]. A greater tortuosity has been correlated with a worse response to drugs, confirming again the importance of the analysis of the angio-structure from the therapeutic perspective.

Among several reasons, including their relative simplicity and the speed to be computed, the most used methods to compute mvFD are the box-counting and the sandbox methods (see Chap. 2). These methods have been applied to whole histological specimens of brain tumors as well as on selected spots. The choice of the “hot spot” can be performed manually (i.e., the pathologist chooses the area subjectively considered to be the most vascularized and representative of the microvascular network of the tumor) or by computer-aided methods (e.g., the spot with the highest vascular area, as calculated by the computer). The first method is biased by the introduction of the operator-dependent choice of the hot spot, while the second method can be biased by the operator-dependent choice of the threshold to automatically extract the immunoreactive vessels from the specimen or from manually

erasing artifacts outside the region of interest. An ideal method should be objective and reproducible, without any operator-dependent biases. Moreover, as it often happens in pathology, the analysis can be biased by the choice of the specimen itself (the problem of sampling, which could not be representative of the whole tumor, like a sample resembling a WHO grade II glioma within a grade III tumor). To limit this problem, other parameters have been introduced.

24.4.2 Local Fractal Dimension and Local Box-Counting Dimension

In order to avoid the sampling of nonrepresentative areas of the tumors such as necrotic areas, the local fractal dimension considers FD in the hot spot. The subjective selection of the hot spot would limit the objective reproducibility of such parameters in the same way as with the MVD. In order to avoid such limitations, the hot spot can be automatically chosen, for example, by computing the vascular area for each 1 mm² area on a grid drawn on the specimen (Fig. 24.5) and extracting the spot with the highest ratio (immunoreactive surface of vessels/tumoral area). Once selected, the 1 mm² spot with the highest vascular area becomes the hot spot for the following analysis (Figs. 24.5 and 24.6). On such a spot, the local mvFD can be computed. A previous analysis showed that a monofractal behavior was found considering a two orders of magnitude window, that is, between 1 and 100 μm [20]. Assuming an average microvessel diameter between 5 and 10 μm , such a fractal window considers the smallest features of the microvessels (i.e., the roughness of the outline), as well as the agglomerate of micro-clusters of vessels (at the magnitude of 100 μm). As shown, this was considered insufficient to cover the entirety of the histopathological features of the microvessels, such as the macro-agglomerates in large clusters (like the garland-like vascular structures). A fractal window ranging between 1 μm and 1 mm, that is, covering three orders of magnitude, was considered significant to cover all of the mean histopathological characteristics of the microvessels, including the intrinsic morphologic features and their clustering properties. Applying the box-counting method in such a range, a monofractal behavior was not confirmed (see Fig. 24.6), although a similar slope of the curve was found across all the analyzed specimens. The several power laws showed that glioma microvasculature clearly demonstrates a multifractal distribution of geometrical complexity of the glioma's microvessels when observed at various scales of magnification. The slope of a straight line interpolating the points on the log-log graphs was considered as the mean value expressing the space-filling properties of the microvessels in the hot spot, and the value was indicated as the local box-counting dimension (local bcD) [20]. Both mvFD and local bcD express, in a single variable, the ability of the microvessels to fill the space in which they are embedded (i.e., the tumor), which is a characteristic intrinsically correlated to their number, size, shape, and distribution pattern.

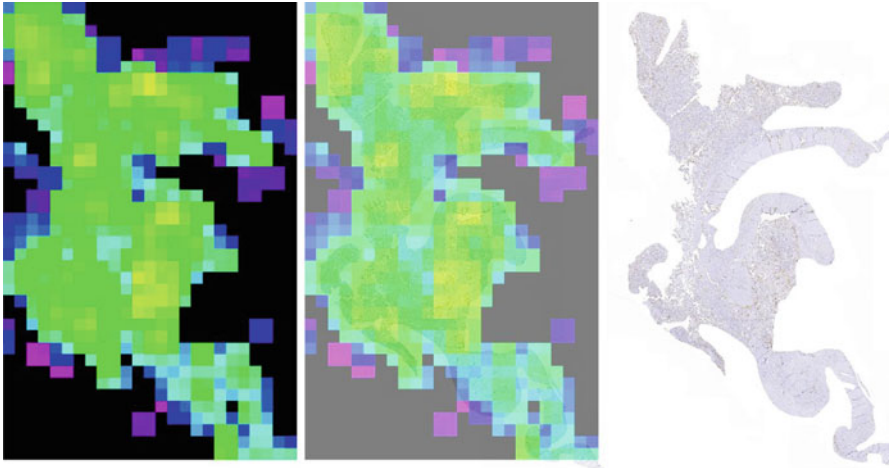


Fig. 24.5 Microvascular map for the automatic selection of the hot spot. Over-imposing a grid of 1 mm^2 boxes on the entire specimen, the vascular area can be computed on each box and a color map can be associated to the different ratios. In this example, the yellow boxes represent the regions of the specimen with the highest vascular area. Raw data show also the value of the vascular area for each box, and the one with the highest value is automatically chosen as hot spot for the following computational analyses (software by Carlo Russo)

24.5 Fractal-Based Analysis of the Angio-Space in Brain Pathology

We define the concept of “angio-space” as the set of parameters quantifying the microvasculature within a tumor or the tumoral space occupied by its own angioarchitecture (Di Ieva [19] and unpublished data). According to the speculation that different tumors, or even different subtypes of the same tumor have a specific “microvascular fingerprint” [19], it has been speculated that a gradient of geometrical representation (and then of fractal angioarchitectural parameters, including FD) exists between physiological and pathological tissues, as well as different states of the same tissues. The pituitary gland is an exemplary model: Euclidean-based approaches (i.e., vascular area and MVD) and fractal-based analyses have clearly shown that it is not only more vascularized but also exhibits a higher microvascular heterogeneity and geometrical complexity in the comparison of the pituitary adenomas [13, 16, 21]. Regarding the different subtypes of pituitary adenomas, analyses of the MVD have shown very discordant results [13], with some preliminary fractal-based analyses not showing any statistically significant differences in the angioarchitecture of micro- vs. macro-adenomas, for example [Di Ieva, unpublished results]. The pituitary gland is a simple model from which to extend the fractal analysis of the microvasculature to the study of brain tumors, by analyzing histological specimens treated with antibodies raised against the endothelium (in the whole specimen and/or hot spots), or volumetric reconstructions of histological

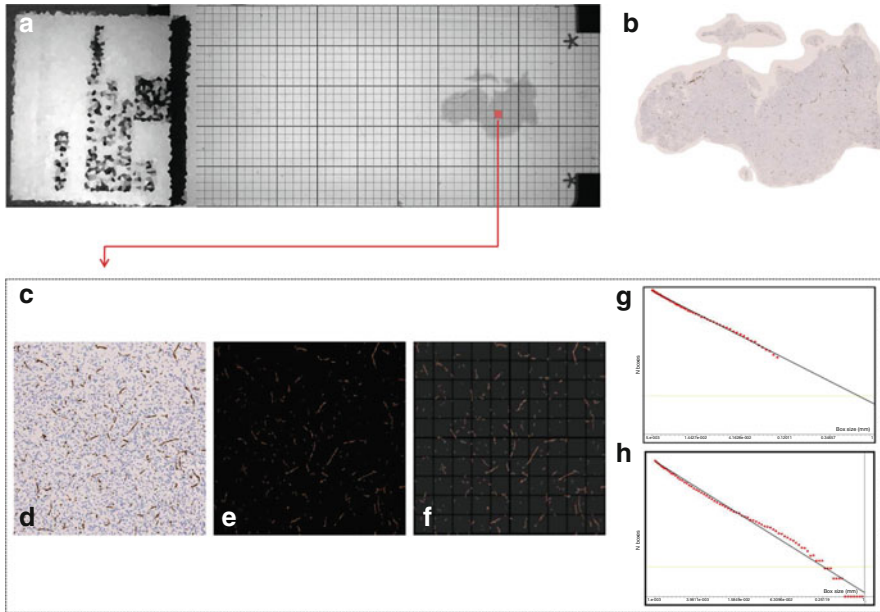


Fig. 24.6 Computer-aided technique for fractal analysis of the microvasculature in histological specimens. Estimation of the morphometric fractal parameter in the whole section and in the hot spot region of specimens of brain gliomas; detection of microvessels using antibodies raised against CD34. The extraction of the whole specimen and the segmentation of the immunoreactive vessels (**a**, **b**) allow the automatic measurement of the morphometric parameters listed in Table 24.4. Moreover, a grid of boxes measuring 1 mm to a side length is superimposed on the whole histological section. For each box, the ratio between the local vascular area and the local neoplastic area is calculated [*local A*(%)] ; the box with the highest *local A*(%) is automatically selected and extracted to be used as the “hot spot” representative of the whole specimen (**c**). (**d**) Hot spot of the specimen, automatically extracted. (**e**) Selection of the RGB threshold to automatically extract the CD34+ vessels. (**f**) Box-counting method: superimposition on the image of grids with boxes of different lengths, from a minimum size (ϵ_{min}) to a maximum size (ϵ_{max}), in the pre-selected scaling window. (**g**) A monofractal behavior was found by limiting the scaling window to a range of two orders of magnitude ($\epsilon_{min} = 1 \mu\text{m} - \epsilon_{max} = 100 \mu\text{m}$). The slope of the obtained curve is the microvascular fractal dimension (mvFD) [Axes of the log-log graph (box-counting method): X-axis=Box size (mm); Y-axis=N Boxes]. (**h**) Considering that a maximum size of the box of 100 μm does not cover the complete hot spot, a wider scaling window was also applied (Reproduced from Di Ieva et al. [20])

tissues and/or radiological images (see Chaps. 19, 20, 21, 22, and 23). If the pituitary gland shows a relatively low variability of the microvascular patterns (meaning that the vascular network is quite homogeneous) [12], malignant brain tumors show very high microvascular heterogeneity. By analyzing 114 GBM specimens, a mean mvFD value of about 1.44 (ranging 1.06–1.87) was found, with a coefficient of variation (CV, standard deviation divided by the mean) of 44% [18]. For comparison, the CV of pituitary adenomas was found less than 10% [12]. This very high

variability in the space-filling properties of microvasculature of GBM reflects the very high angioarchitectural heterogeneity shown by this malignant tumor. The microvascular networks of GBM can be represented in a very broad and continuous spectrum of possible geometric configurations; this was the first mathematical demonstration that GBM is *multiforme* (from the Latin meaning “multi-shapes”) also in relation to its microvasculature [18].

Morphometric and computational fractal-based analyses also showed that grade II gliomas have different angio-spaces in comparison to grade III gliomas (Fig. 24.7). Among several morphometric parameters, the local mvFD and especially the local bcD were shown to be the most reliable quantitative indicators of the neoplastic microvasculature, making them potential surrogate biomarkers [20]. The histopathological grading of gliomas can be challenging in some cases, especially in grade II vs. grade III gliomas, and the erroneous diagnosis can result in patients’ over- or undertreatment [48]. For this reason, computational methods, which can help in differential diagnosis, are more than welcome to be added in the pathologists’ and clinicians’ armamentarium.

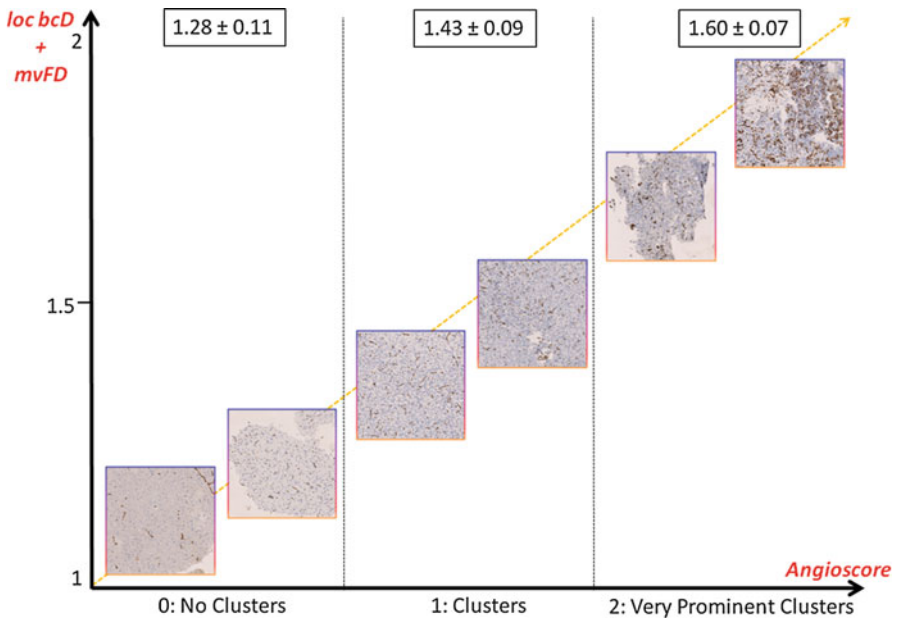


Fig. 24.7 Correlation between the fractal angiormorphometric parameters and the level of clustering of the microvessels. Schema showing the correlation between the angioscore assigned by the neuropathologist to each specimen and the fractal-based variables [local box-counting dimension (loc bcD) and microvascular fractal dimension (mvFD)]. The value reported for each angioarchitectural group is the mean value of loc bcD \pm standard deviation. The yellow line indicates the increasing grade of malignancy of the tumor. The fractal parameters assigned nearly 75% of the gliomas to the correct histological grade and angioscore (Reproduced from Di Ieva et al. [20])

24.6 Limitations

If fractal-based angioarchitectural classification of brain tumors can be useful for diagnostic and prognostic purposes, there are still some limitations to acknowledge. For a description of the limitation of image and fractal analysis, see Chaps. 1 and 12. Below, we summarize some main points. First of all, according to the basic principles of fractal geometry, it does not mean that objects owning the same mvFD, for example, show the same microvascular pattern. As previously stated, this means that fractal indexes should be analyzed as complementary to other morphometric parameters in a multiparametric analysis.

When the analysis is performed on histological specimens, the sampling problem can limit its benefit. The analysis of the whole specimen or of the computationally chosen most representative spot of the specimen tends to decrease the limitations of other analyses, like the MVD-based ones, but a histological sample still often does not represent anything more than a very small fraction of the whole tumor, and this should be taken into consideration. The estimation of a 2D representation of the microvascular tree could not reflect the real 3D complexity of the microvascular network, because 2D methods performed on histological specimens are length related rather than mass related, and this can underestimate the true 3D complexity of the tree in the space [7, 42]. This issue can be limited by using volumetric analyses and 3D reconstructions of tissues. The effect of geometrical deformation (e.g., cracks) during the process of sample preparations, and other artifacts such as nonhomogeneity of staining, may also have some impact on the accuracy of the fractal analysis. For this reason, also the sampling, immunostaining and preparation of the histological specimen should be standardized.

Moreover, the histological section represents a “snapshot” in time, a “frozen photo” of the tumor at the moment of the surgical operation, and each quantification refers to that specific moment, not taking in account the dynamic process on the temporal trend of the tumor. For this reason, the temporal changes of tumors should also be considered by means of neuroradiological and/or nuclear medicine findings over time (such as perfusion parameters or methionine uptake in PET, positron emission tomography) [17].

As previously stated, the value of FD and other fractal-based parameters is highly dependent on the image analysis parameters (and “pixel related”, according to the resolution). Several research inquiries have considered different tissues (brain cortex, tumors) in different models (humans, rats), visualized in different ways after undergoing different treatments. When comparing the several values found in the published research, a plethora of results show contrasting findings, and this could be avoided only by a standardization of techniques, materials, and methods.

The standardization ought to be done also in the analytical analysis of the fractal parameters. It is in fact known that biological fractals, such as microvascular networks, are statistically self-similar only within a specific scaling window (i.e., covering at least two orders of magnitude) [40]. The choice of different windows may give different results, and such choice should be standardized as well or at least

justified: if a window ranging 1–100 μm , for example, considers a FD that reflects the organization of the vessels in micro-clusters and their roughness, on the other hand, a range of 100–10,000 μm does not take into account the roughness of the single vessels but rather the distribution of vascular clusters on a larger scale [14]. The lack of standardization of all these elements could produce results no more comparable between different laboratories. Last but not least, microvessels cannot be real fractals, as pointed out, and the “fractalization” of such natural objects can be a mathematical idealization limited by computational manipulation. In order to avoid the several definitions expressed in the very thick related literature on fractality, quasi-fractality, pseudo-fractality, or multifractality of microvessels [27, 39, 46, 50, 51], it is here enough to say that fractal analysis, far from being an infallible and universal method, is still able to offer several reliable parameters for an objective, reproducible, and realistic description and quantification of the microvasculature in normal as well as pathological tissues.

24.7 Future Perspectives and Conclusion

As the architectural pattern of each tumor seems to be its specific microvascular fingerprinting, objective and reproducible morphometric biomarkers are required to describe and quantify the so-called angio-space. Such indexes should be (a) able to quantify what pathologists describe in a qualitative way on the histological specimens of tumors, (b) objective, that is, not affected by intra- and interobserver variability, and (c) clinically meaningful, having prognostic and/or predictive value [20]. Fractal analysis offers several parameters that are promising candidates for such purposes.

In addition, the methodological phase of testing morphometric parameters should be followed by clinically oriented research; to be tested as a potential biomarker, in fact, the morphometric fractal-based parameters should be analyzed in the same way that new oncologic therapies are introduced into clinical practice, in step-by-step successive phases of study [30]. Although the translation of the concept of “fractal microvasculature” into clinical practice (i.e., from “bench to bedside”) is still difficult to be realized [11], it should be emphasized that the research in the field of microvessel morphometrics should run alongside research focused on tumoral angiogenesis (research on molecular biomarkers, such as vascular endothelial growth factor, hypoxia-induced factor, etc.), in order to offer a holistic view on the effects of structure and function on the environment and viability of brain tumors. This means that the morphometric analyses should run alongside the biological approach. Moreover, morphometric analyses should be integrated into *in vivo* techniques (neuroimaging and nuclear medicine methods) to follow-up brain tumors and response to treatment.

In light of the speculations and findings that have been illustrated in this chapter, computational fractal-based and multiparameter morphometric analyses will ultimately find its place in the oncologist’s toolbox as a conventional diagnostic procedure.

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