

Leili Mirsadraei, Michelle S. Hirsch,  
Christopher J. Kane, and Donna E. Hansel

## Mucinous Tubular and Spindle Cell Carcinoma (MTSCC)

### Introduction

Mucinous tubular and spindle cell carcinoma (MTSCC) is an uncommon, low-grade renal epithelial tumor [5, 6]. It was initially believed to derive from the collecting duct and was originally considered the well-differentiated variant of collecting duct carcinoma [6, 7]. This neoplasm has been previously reported under different names: renal cell carcinoma originating from the distal loop of Henle, renal cell carcinoma of distal nephron origin, loopoma, and low-grade collecting duct

(Bellini) carcinoma [8]. MTSCC occurs more commonly in females (female-to-male ratio of 2:1 to 3:1) and encompasses a broad age range (17–82 years) [7–9].

### Clinical Presentation

Most patients with MTSCC are asymptomatic, although some may experience flank pain and hematuria [5]. A subset of patients may present with renal stones [10]. A common clinical finding is the presence of and associated staghorn renal calculus in approximately 50 % of the cases [5]. MSTCC is often found incidentally when imaging for other indications; however, imaging modalities cannot definitively distinguish MTSCC from other RCC subtypes [11, 12]. Reported cases have been found in the subcapsular, corticomedullary, and medullary areas of the kidney, with the majority presenting as clinical stage pT1 [6, 9].

### Pathology

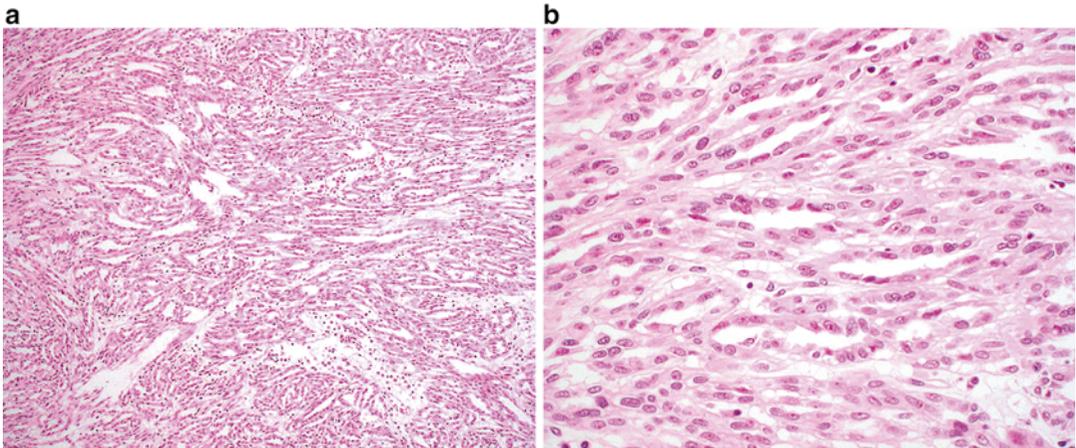
On gross evaluation, MTSCC is typically small but can vary in size, is often localized to the cortex or corticomedullary junction, and appears well defined with a tan-white homogenous cut surface [13]. Occasionally, mucin-containing areas may appear gelatinous in nature [5].

---

L. Mirsadraei, M.D. • D.E. Hansel, M.D., Ph.D. (✉)  
Department of Pathology, University of California at  
San Diego, San Diego, CA, USA  
e-mail: [lmirsadraei@ucsd.edu](mailto:lmirsadraei@ucsd.edu); [dhansel@ucsd.edu](mailto:dhansel@ucsd.edu)

M.S. Hirsch, M.D., Ph.D.  
Department of Pathology, Brigham and Women's  
Hospital, Harvard Medical School, Boston,  
MA 02115, USA  
e-mail: [mhirsch1@partners.org](mailto:mhirsch1@partners.org)

C.J. Kane, M.D., F.A.C.S.  
Department of Urology, University of California  
San Diego, San Diego, CA 92093, USA  
e-mail: [ckane@ucsd.edu](mailto:ckane@ucsd.edu)



**Fig. 9.1** Mucinous and tubular spindle cell carcinoma. (a) Inter-anastomosing cords of cells in a mucinous background are seen at low magnification. (b) Tumor cells show relatively uniform nuclei with minimal atypia

At low magnification, three distinct components are generally identifiable: a mucinous background, monomorphic spindle cells present in sheets, and cystic, elongated, and compressed tubules lined by a single layer of cuboidal cells (Fig. 9.1a). At higher magnification, the tumor cells are well differentiated and show low-grade morphology and minimal variation in cytologic appearance (Fig. 9.1b). Nuclei are commonly low grade in appearance with a round to oval shape, smooth chromatin, and occasional small nucleoli. The cytoplasm may be eosinophilic, and an admixture of inflammatory cells might be seen [5, 11]. Less common features include papillary architecture, oncocytic change, presence of clear cells, and calcifications (psammoma bodies). Rarely, sarcomatoid change has been described in association with MTSCC [14, 15]. If papillary structures are seen, the main differential diagnosis includes conventional papillary RCC; however, the presence of a mucinous background and the absence of characteristic chromosomal alterations associated with papillary RCC support the diagnosis of MTSCC.

MTSCC has been shown to harbor multiple chromosomal alterations, including 1, 4, 8, 9, 13, 14, 15, and 22 [16]. The genetic changes characteristic of papillary RCC, namely, trisomy 7 and 17 and loss of the Y chromosome, are not identified in MTSCC [17].

Most MTSCC lesions express markers that are reminiscent of distal nephron differentiation, including EMA, AE1/3, CK7, E-cadherin, and  $\alpha$ -methylacyl CoA racemase. RCC antigen is positive in the majority of cases. The proximal tubular marker CD10 is positive in only 10–15 % of MTSCC cases [18, 19]. The immunohistochemical profile of MTSCC is nonspecific and partially overlaps with papillary RCC with the absence of CD10 being the most supportive immunohistochemical biomarker for the former [6].

## Prognosis and Clinical Management

MTSCC is often indolent and surgery is curative in most cases [20–22]. A small subset of cases may recur or be associated with regional lymph node metastases [13, 16, 22, 23]. Sarcomatoid transformation is associated with an increased likelihood of distant metastases and risk of death from disease [14, 24, 25].

## Clear Cell Papillary RCC

### Introduction

Clear cell papillary RCC (also referred to by some as “clear cell tubulopapillary RCC”) is reported to

account for less than 5 % of renal epithelial neoplasms; however, this incidence is likely an underestimation due to the misclassification of this tumor as a clear cell or less frequently a papillary, RCC. Clear cell papillary RCC affects men and women equally, and the age distribution ranges from 18 to 88 years [26–28]. Clear cell papillary RCC may occur sporadically or may be associated with end-stage renal disease [29–33].

## Clinical Presentation

The majority of clear cell papillary RCC cases are discovered incidentally, although a subset of patients may present with hematuria or flank pain [27].

## Pathology

Clear cell papillary RCC is primarily a cortical lesion. On gross evaluation, these tumors are well circumscribed, encapsulated, and relatively small in size (0.6–5.5 cm) [28, 31]. Cystic change may be present. The cut surface ranges from yellow to tan to brown in appearance. These tumors are commonly unifocal, although bilateral and multifocal tumors have been described.

Microscopically, several architectural patterns may be present and include cystic and solid structures as well as branching glands [27, 31, 34]. A defining characteristic of these tumors is the presence of tubular and papillary structures lined by clear cells (Fig. 9.2a). Occasional eosinophilic luminal secretions may be seen. Tumor cells show clear cytoplasm, and the nuclei are uniform and small and lack nucleoli (Fig. 9.2b). Prominent nuclear polarization toward the luminal surface is striking in these cases [27, 31, 35]. The intervening stroma is often fibrous or hyalinized but may contain smooth muscle.

Genetic analysis shows these tumors to be distinct from clear cell RCC and papillary RCC. Specifically, clear cell papillary RCC lacks chromosome 3p deletions or *VHL* gene alteration characteristic of clear cell RCC, as well as extra copies of chromosomes 7 and 17 as is seen in

papillary RCC [30, 31, 36–39]. Some molecular differences have been reported between clear cell papillary RCC that occurs in the sporadic and the ESRD settings [31, 40, 41].

Immunohistochemistry is helpful in the diagnosis of this entity by distinguishing it from both clear cell RCC and papillary RCC. Clear cell papillary RCC shows positive immunoreactivity for CK7; carbonic anhydrase IX (CAIX), PAX2, and PAX8; and high-molecular-weight cytokeratin (34βE12) [26, 29, 42]. Immunostains for AMACR (racemase) and RCC antigen are negative, and CD10 is negative or only focally positive. In contrast, clear cell RCC is typically negative for CK7 and positive for CD10, RCC antigen, and CAIX and may show variable AMACR staining. Papillary RCC is typically positive for CK7, AMACR, and CD10; RCC antigen may be variable and CAIX should be negative.

## Prognosis and Clinical Management

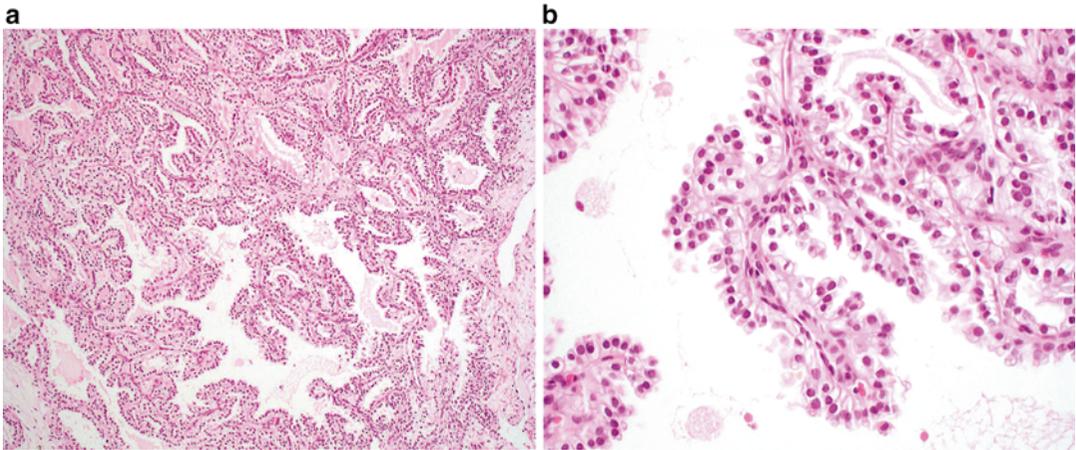
All cases reported to date in the literature have shown indolent behavior, without recurrence or metastatic spread [1, 32, 33, 42].

---

## Tubulocystic Carcinoma

### Introduction

Tubulocystic carcinoma is an uncommon form of RCC described in numerous case reports and small case series [6, 43–46]. Tubulocystic carcinoma was initially described by Amin et al. [43] in 2009; however, earlier descriptions of similar tumors date back to 1956, when it was included with Bellini (collecting duct) carcinomas, and later described as the low-grade collecting duct carcinoma [47, 48]. However, more recent studies have demonstrated that tubulocystic carcinomas are more likely related to papillary RCC instead of collecting duct carcinoma [43, 44]. Tubulocystic carcinoma affects adults over a broad age range of 29–94 years of age [49, 50]. There is a strong male predominance (male-to-female ratio of 7:1) [47, 51]. Based on



**Fig. 9.2** Clear cell papillary renal cell carcinoma. (a) A combination of papillary and nested patterns is identifiable in this example. Uniform alignment of the nuclei to

the luminal surface is evident even at low magnification. (b) Papillary fronds lined by clear cells are a characteristic feature of this lesion

immunohistochemical staining pattern, ultra-structural features, and gene expression profiling, tubulocystic carcinoma is favored to originate from either the proximal convoluted tubule or intercalated tubule [6, 43, 51].

### Clinical Presentation

Patients are often asymptomatic, although some may present with abdominal pain, distension, and hematuria. Tubulocystic carcinoma is usually solitary and often involves the left kidney [51]; however, multifocal tumors have been described in approximately 23 % of patients [44, 47]. Tubulocystic carcinoma should be considered in the differential diagnosis of a renal lesion with a cystic component on radiological examination.

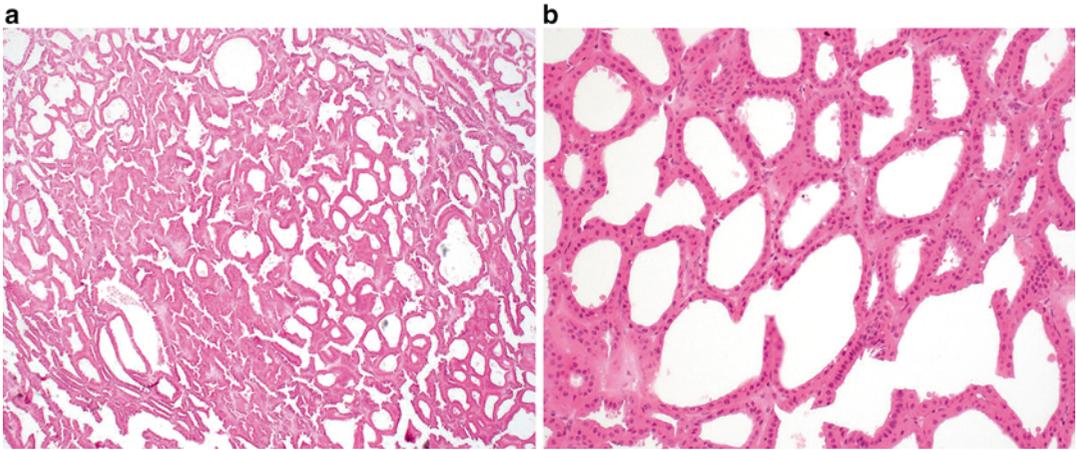
### Pathology

On gross examination, tubulocystic carcinoma is usually solitary and well circumscribed and has a tan-gray, spongy appearance; the latter is due to numerous cysts/microcysts [47]. Tubulocystic carcinoma varies in size from 0.2 to 17 cm [48, 51]. Multifocal tumors may be present, which

can include multiple distinct tubulocystic carcinomas or tubulocystic carcinoma associated with a concurrent second renal neoplasm, most frequently a papillary RCC [47, 48]. Tumors are most commonly located in the cortex, although up to a third can be present at the corticomedullary junction [6].

Microscopically, tubulocystic carcinoma is composed of many cystic structures that are small to medium in size, which are separated by thin fibrous septa (Fig. 9.3a). Solid growth is not seen. The cells lining the cysts are cuboidal, columnar, or hobnail in appearance with abundant eosinophilic or amphophilic cytoplasm which characteristically contain large nuclei with prominent red nucleoli (Fig. 9.3b) [47]. Rarely, sarcomatoid features may be seen in association with this tumor but only in the presence of solid growth (i.e., an associated component of high-grade papillary RCC) [1, 52]. On ultrastructural examination, the tumors show abundant microvilli with brush border organization suggestive of proximal convoluted tubules, with some features resembling intercalated cells of the collecting ducts [51].

Genetic analysis of these tumors report a similar profile to papillary RCC, which include gains of chromosome 7 and 17 and loss of the Y chromosome. However, classic features associated with “low-grade” papillary RCC are typically not



**Fig. 9.3** Tubulocystic renal cell carcinoma. (a) Multiple cystic structures of variable size are present at low magnification. (b) Cysts are lined by a single layer of cells with eosinophilic cytoplasm, large nuclei, and prominent nucleoli

seen in this entity; in contrast, high-grade tumors with solid and papillary growth patterns and similar nuclear and immunophenotypic features can be seen [44, 45, 53]

Immunohistochemical studies demonstrate immunoreactivity for cytokeratins 8, 18, and 19 [6, 44]. CD10 and P504S (racemase) are positive in greater than 90 % of cases. Diffuse and strong AMACR positivity, positive to variable CK7 expression, and staining for kidney-specific cadherin and PAX2 have been reported [44, 47]. Tubulocystic carcinoma is negative for high-molecular-weight cytokeratin (34BE12) [47, 48].

The main differential diagnoses include other tumors with a multiloculated gross appearance, including multiloculated clear cell RCC, cystic clear cell tubulopapillary RCC, cystic nephroma, mixed epithelial and stromal tumor, and cystic oncocytoma [47, 51, 54, 55]. Distinction between these entities is primarily based on light microscopic evaluation, rather than use of ancillary studies.

### Prognosis and Clinical Management

Whereas the majority of reported cases of tubulocystic carcinoma demonstrate indolent behavior, recurrence and metastases to regional and distant sites such as the bone and liver have been reported

in rare cases; the latter is associated with tumors that demonstrate solid and papillary growth patterns [44, 45, 47, 51, 56].

## Acquired Cystic Disease-Associated RCC

### Introduction

Cystic change occurs in the majority of patients with end-stage renal disease (ESRD) who undergo dialysis and is termed “acquired renal cystic disease” (ARCD) [5, 6]. Patients with ARCD have increased risk of developing RCC comparing to normal population [6, 57, 58], with an RCC incidence rate of 10 % in ARCD patients [5, 6, 37, 59]. A direct correlation between the duration of dialysis and RCC development in ESRD patients has been suggested in some studies [5, 6, 29, 60]. Multiple types of renal neoplasms may arise in the background of ESRD, including clear cell RCC, papillary RCC, clear cell papillary RCC, and chromophobe RCC [5, 61–63], although acquired cystic disease-associated RCC represents up to 36 % of all renal neoplasms in this setting [29]. Similar to non-ESRD RCC, acquired cystic disease-associated RCC occurs most commonly in men between the ages of 55 and 65 years [5, 30, 40, 61].

## Clinical Presentation

Most cases are found as an incidental finding on nephrectomy specimens removed for ESRD [5]. However, a subset of patients undergoing dialysis may have pain or demonstrate hematuria. In addition, imaging may identify an incidental lesion in the background of cystic renal parenchyma [6, 62].

## Pathology

On gross examination, acquired cystic disease-associated RCC is well circumscribed and tan/yellow in appearance and may show focal hemorrhage or necrosis [5, 6]. The background renal parenchyma shows multiple variably sized cysts, with the neoplasm appearing to arise in association with a cyst wall or from one of the connecting trabecular areas. Multifocal tumors occur approximately 50 % of cases, and bilateral tumors are found in up to 20 % of patients [1, 29].

At low magnification, acquired cystic disease-associated RCC shows a distinct “sieve-like” appearance (Fig. 9.4a). At higher magnification, multiple growth patterns that include acinar, alveolar, solid, cystic, and papillary regions may be identified. The tumor cells show granular, eosinophilic cytoplasm and contain large nuclei with prominent nucleoli [6]. Intratumoral calcium

oxalate crystal is a relatively specific feature in this entity, although may be lacking a minority of cases (Fig. 9.4b) [6, 29, 59]. Rhabdoid and sarcomatoid features have been reported in a small number of cases [29, 61, 64].

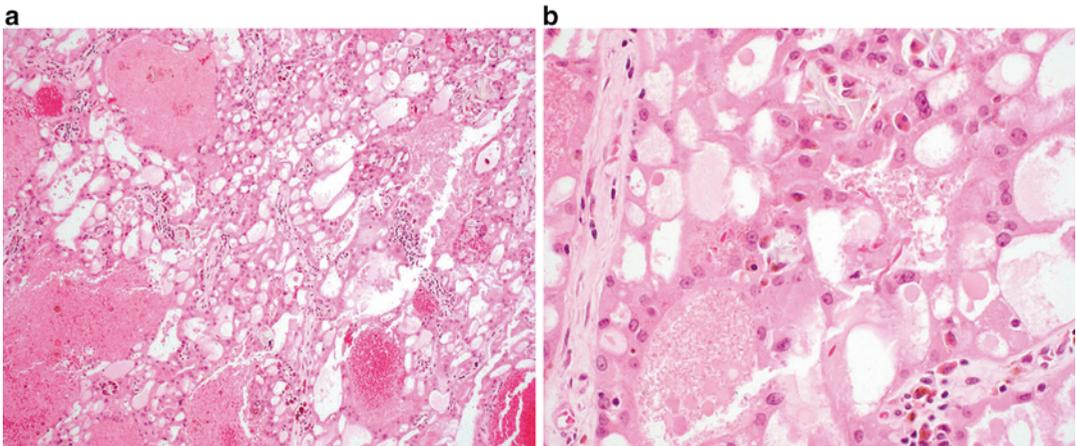
Fluorescence in situ hybridization (FISH) and comparative genomic hybridization performed on nine cases showed variable combined gains of chromosomes 3, 7, 16, 17, and Y [6, 60].

Acquired cystic disease-associated RCC shows immunoreactivity for  $\alpha$ -methylacyl-coenzyme A racemase (AMACR), CD10, RCC antigen, and glutathione S-transferase A [6, 60]. CK7 is often negative but can show focal immunoreactivity [6].

The main differential diagnosis includes papillary renal cell carcinoma. Immunostains for CK7 and chromosomal analysis can be used to distinguish these two entities in most cases, if required.

## Prognosis and Clinical Management

Most cases of acquired cystic renal disease-associated RCC typically show indolent behavior, which may reflect early detection in patients with ESRD [6, 62]. As anticipated, the minority of tumors that show rhabdoid or sarcomatoid features may behave more aggressively and be associated with metastatic spread [1, 29, 64, 65].



**Fig. 9.4** Acquired cystic disease-associated renal cell carcinoma. (a) A sieve-like pattern is evident at low magnification. (b) Crystals may be seen in a subset of cases

## Neuroblastoma-Associated RCC

### Introduction

Neuroblastoma-associated RCC is a unique entity that occurs 3–35 years after diagnosis and treatment of childhood or adult neuroblastoma [5, 6, 66, 67]. It is a rare tumor, accounting for less than 1 % of all cases of RCC and approximately 2.5 % of the cases of RCC in children [5, 11]. Males and females have an equal risk of involvement [63]. Some reports suggest a relationship between the treatment for neuroblastoma and development of neuroblastoma-associated RCC [11]. However, a small number of patients develop RCC in the absence of radiation or chemotherapy, suggesting a possible underlying germ line susceptibility for these lesions [67].

### Clinical Presentation

Neuroblastoma-associated RCC can be unilateral or bilateral [11]. Rare cases showing multifocal tumors have also been reported. No specific radiologic findings have been associated with this entity. The diagnosis may be suggested based on a history of treatment for neuroblastoma [11].

### Pathology

On gross evaluation, tumors range in size from 1 to 8 cm [5, 63]. Neuroblastoma-associated RCC is histologically heterogeneous [11]. The most common appearance is that of an oncocytic tumor that shows solid and papillary architecture and occasional vacuolization of the cytoplasm [5, 53, 63, 68]. Variable nuclear size and medium-sized nucleoli are present, and mitotic figures may be identified. One should note that other forms of RCC, including Xp11.2 translocation-associated carcinoma and classic clear cell RCC, can also occur in a subset of patients following neuroblastoma [5, 63, 69].

Due to the small number of cases described to date, the genetic profile of this tumor has not been well defined, although chromosomal altera-

tions affecting 14q31 and 20q13 have been reported [5, 70].

Immunohistochemistry shows neuroblastoma-associated RCC to be frequently positive for EMA, vimentin, and cytokeratins 8, 18, and 20 and negative for cytokeratins 7, 14, 19, 20, S100, and HMB45 [5, 63]. The main differential diagnosis includes oncocytoma and chromophobe RCC in the oncocytic subtype of neuroblastoma-associated RCC. Clinical history is critical in this differential diagnosis.

### Prognosis and Clinical Management

The oncocytic subtype of neuroblastoma-associated RCC behaves in an indolent manner. Outcomes associated with clear cell RCC and Xp11-associated RCC are likely similar to their de novo counterparts. Awareness of this entity is important to ensure close follow-up and frequent assessment of survivors of neuroblastoma in order to detect RCC early in the course of disease.

---

## Thyroid-Like Follicular Carcinoma of the Kidney (TLFCK)

### Introduction

Thyroid-like follicular carcinoma of the kidney (TLFCK) is a rare renal neoplasm. To date, only a limited number of TLFCK cases have been reported in the literature [1, 6, 71–75]. This tumor occurs over a broad range of 29–83 years, with an equal gender distribution [1, 6, 73, 75]. This entity has not yet been classified as a distinct renal neoplasm by the World Health Organization.

### Clinical Presentation

Most of these carcinomas are identified as an incidental finding on radiologic imaging, although a subset of patients present with abdominal pain, flank pain, hematuria, and relapsing urinary infection [6, 71, 75].

## Pathology

On gross examination, these tumors are well defined and encapsulated, with a solid or cystic and yellow-tan cut surface [71, 75]. Focal areas of hemorrhage or necrosis may be seen. Microscopically, TLFCCK shows variably sized follicular structures containing both microfollicular and macrofollicular patterns (Fig. 9.5a). The follicles contain pink colloid-like material and are lined by a single layer of cuboidal to columnar cells with moderate amphophilic to eosinophilic cytoplasm (Fig. 9.5b) [71, 73]. Nuclei are typically round to oval and may demonstrate nuclear grooves, similar to that seen in thyroid neoplasms. Lymphocytic infiltrates with or without reactive germinal centers can be present [71].

Given the low incidence, the genetic profile of TLFCCK has not been well defined.

The immunohistochemical profile of TLFCCK shows positive immunoreactivity for PAX8, EMA, CK7, and vimentin, with a subset of cases reported to express CD10 and PAX2 [71, 76]. The colloid-like material in TLFCCK is composed of Tamm-Horsfall glycoprotein, the most abundant protein in normal urine [71, 77]. The main differential diagnosis is metastatic follicular carcinoma of the thyroid; however, TLFCCK is negative for thyroid markers TTF-1 and thyroglobulin [6, 71]. The diagnosis of TLFCCK is strengthened by the absence of lesions in the thyroid or other locations

by imaging [6, 73]. TLFCCK should also be distinguished from thyroidization of the kidney, which is characterized by atrophic distal tubules or collecting ducts, with colloid-like hyaline casts that mimic normal thyroid parenchyma. Thyroidization is a benign, widespread, and usually bilateral process that occurs in patients with a history of chronic pyelonephritis, obstructive uropathy, or end-stage renal disease. In contrast, TLFCCK presents as a well-circumscribed tumor in a background of unremarkable kidney and is often in patients with no history of renal disease [71].

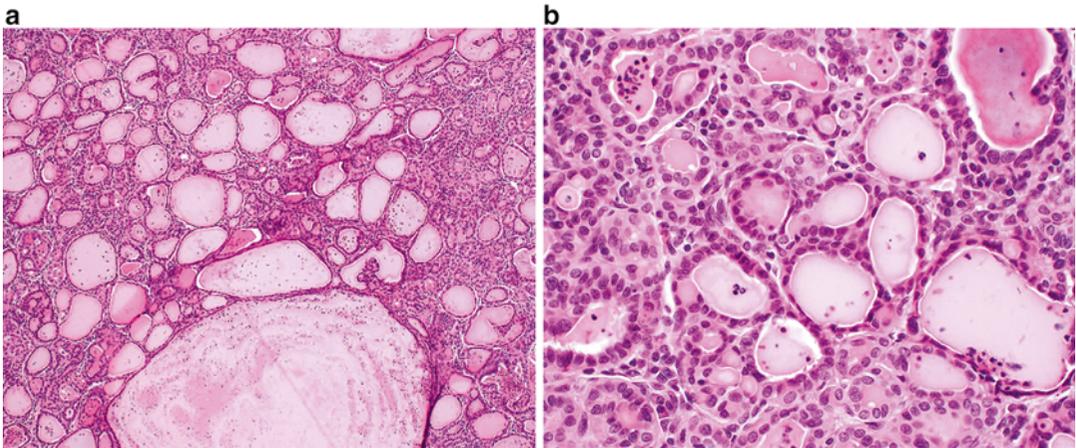
## Prognosis and Clinical Management

Although most cases of TLFCCK remain localized, one case of TLFCCK metastasized to the renal hilar lymph nodes and a second showed widespread metastases to the lungs and retroperitoneal lymph nodes [71, 73].

## Succinate Dehydrogenase-Deficient RCC

### Introduction

Succinate dehydrogenase (SDH)-deficient renal cell carcinoma is a recently described entity that is not yet widely recognized due to its morphologic



**Fig. 9.5** Thyroid-like follicular carcinoma of the kidney. (a) A combination of variably sized follicular structures filled with colloid-like material and small nests may be

seen in this entity. (b) The follicular structures are lined by a single layer of cuboidal to columnar cells with eosinophilic cytoplasm and minimal nuclear atypia

overlap with other oncocytic renal neoplasms. Nevertheless, SDH-deficient RCC has been accepted as a provisional entity in the 2013 International Society of Urological Pathology Vancouver Classification of renal tumors, as it represents a distinct renal neoplasm, defined by loss of IHC staining for the B subunit of the SDH mitochondrial complex (the entire complex consists of four subunits: A, B, C, and D) [1, 78, 79]. The tumor is rare, estimated as less than 1 % of all renal epithelial tumors [79], and it occurs in individuals with germ line mutations of the genes that encode the SDH complex [1, 78]. SDH-deficient RCC has been reported more commonly in men, with the mean age of 40 years [78, 79]. Patients with germ line mutation in a SDH subunit gene are also prone to develop paraganglioma/pheochromocytoma, gastrointestinal stromal tumor (GIST), and pituitary adenoma [1, 78–80].

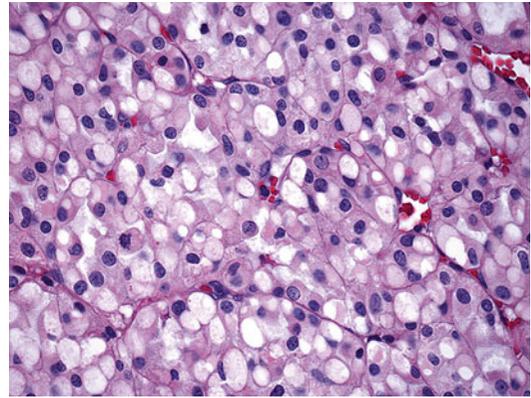
### Clinical Presentation

Due to the low incidence of this renal cancer, clinical presentation has yet not been described in detail. Some patients have reported episodic headache and palpitation associated with paraganglioma-related hypertension, with an incidental renal mass discovered on imaging during work-up [81]. Bilateral tumors have been reported [79].

### Pathology

On gross examination, this tumor is well circumscribed with a tan to red cut surface [78, 79]. Hemorrhage and microcystic change may be present [78, 79]. Tumor diameter ranges from 2.0 to 20 cm [78]. Both unifocal, multifocal, and bilateral tumors have been described [78, 79].

Microscopically, cells are arranged in solid or nested patterns that contain uniform neoplastic cells with eosinophilic, variably granular cytoplasm [1, 78, 79, 81]. The most distinctive histological feature is the presence of perinuclear cytoplasmic vacuoles and inclusion-like spaces, some of which contain pink flocculent-like material (Fig. 9.6) [78, 79, 81]. Nuclei are typically



**Fig. 9.6** SDHB-associated renal cell carcinoma. Distinctive features include perinuclear cytoplasmic vacuoles and inclusion-like spaces filled with pink secretions

round with smooth borders, finely clumped chromatin, and small or inconspicuous nucleoli [78]. Benign tubules or glomeruli are often entrapped at the edges of the tumors [81]. Stromal hemorrhage, fibrosis, and hyalinization are commonly seen [78]. Intratumoral mast cells and aggregates of lymphocytes, as well as necrosis, may be present [78]. Sarcomatoid change has been described [79].

By immunohistochemistry, the characteristic and pathognomonic finding is loss of SDHB; however, this can occur when any of the four SDH complex components (A, B, C, or D) are mutated. In SDHB-mutated RCC, the SDHA antibody will highlight the perinuclear cytoplasmic inclusions which contain the mutated mitochondria [78, 79]. Immunohistochemistry beyond loss of expression with SDH antibodies is non-specific: neoplastic cells are positive for PAX8 (confirming renal origin), EMA, and S100 and negative for RCC; labeling for CAM5.2, AE1/AE3, cytokeratin 7, CD10, and AMCR is variable, and C-KIT highlights intratumoral mast cells but is negative in the tumor cells [78, 79].

The differential diagnosis of SDH-deficient RCC includes oncocytoma, chromophobe RCC, and the “granular variant” of clear cell RCC. Loss of staining for SDHB is a definitive confirmation of the diagnosis. Also, SDH-deficient RCC commonly shows negative or focal cytokeratin reactivity [79].

## Prognosis and Clinical Management

Metastatic disease to the liver, lung, ribs, vertebrae, adrenal gland, and lymph nodes has been reported. Concurrent paragangliomas may also be observed [78, 79, 81]. Tumor progression and death from disease have been reported [78, 79, 81].

### TCEB1-Mutated (Monosomy-8) Renal Cell Carcinoma

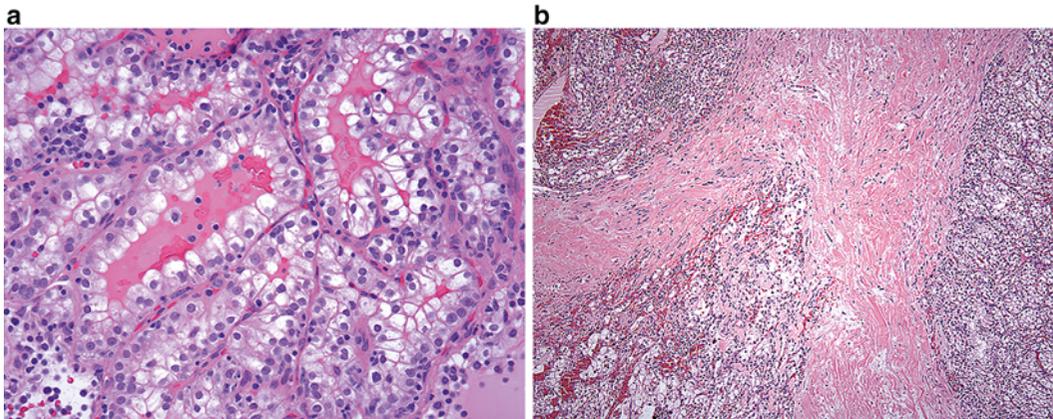
Renal cell tumor with TCEB1 mutations and loss of heterozygosity of chromosome 8 are a recently described renal tumor variant that has morphologic overlap with clear cell RCC and clear cell papillary RCC [82, 83]. This emerging renal neoplasm was identified through common mutations in the TCEB1 gene (which encodes elongin, a member of the VHL complex) and by loss of chromosome 8 with conventional G-band karyotyping and fluorescence in situ hybridization. To date, only 17 tumors have been reported [82–84]. Alterations of TCEB1 impact VHL-pathway signaling, a commonly altered pathway in clear cell renal cell carcinoma, but in a mechanism that is independent of direct VHL mutation [82, 84].

## Clinical Presentation

All but one reported TCEB1-mutated RCCs reported to date have been small (stage pT1) and therefore likely to present incidentally, although nonspecific symptoms typically associated with other renal masses (pain and hematuria) may be present.

## Pathology

Grossly, tumors are typically small, tan/yellow, and well circumscribed. A prominent thick fibrous capsule is almost invariably present. Microscopically, TCEB1-mutated tumors show nests and tubule of clear cells with occasional focal papillary structures. Cells contain abundant clear cytoplasm and pink amorphous material within tubule lumens [82, 83] (Fig. 9.7a). Nuclei are low grade but typically larger than is seen in clear cell tubulopapillary RCC; nucleoli are absent (i.e., size suggestive of Fuhrman nuclear grades 2–3, but absence of nucleoli is more in keeping with Fuhrman nuclear grade 2). There is a hint of nuclear polarization but not to the extent seen in clear cell papillary RCC. A relatively distinct and reproducible feature is the presence of thick fibromuscular bands that dissect through the mass (Fig. 9.7b).



**Fig. 9.7** TCEB1-mutated renal cell carcinoma. (a) Tubular structures lined by clear cells with abundant clear cytoplasm. *Pink amorphous material* is present in the

tubule lumen. (b) A unique feature is the presence of thick fibromuscular bands within the lesion

Immunohistochemistry shows immunoreactivity for CAIX and HIF-1-alpha, as well as CK-7; the latter distinguishes it from most clear cell RCCs. Tumor cells are negative for high-molecular-weight cytokeratin (34BE12) [82], and unlike clear cell tubulopapillary RCC, TCEB1-mutated tumors show variable staining for AMACR, CD-10, and RCC antigen [83]. Of note, the cuplike staining pattern described for CAIX in clear cell papillary RCC is not present in TCEB1-mutated RCC; instead a membranous pattern, similar to clear cell RCC, is observed.

The differential diagnosis includes both clear cell renal cell carcinoma and clear cell papillary renal cell carcinoma. Although morphological features can be very suggestive and immunohistochemistry can be supportive, definitive diagnosis requires molecular analysis to show mutations in the TCEB1 gene and/or loss of heterozygosity at the chromosomal region that contains TCEB1 (8q21.11). Clinically, the use of G-band karyotyping (fresh tissue required) or FISH analysis (possible with formalin-fixed tissue) can be used to demonstrate a loss of chromosome 8. The short arms of chromosome 3 should be intact to make a diagnosis of a TCEB1-mutated RCC.

## Prognosis and Clinical Management

All cases reported thus far have been low stage (pT1), except one pT3a case. During a median 48-month follow-up period, no patient was reported to develop metastatic disease. However, further evaluation on a larger number of tumors with extended follow-up is needed to determine outcomes relative to conventional clear cell RCC.

---

## Renal Carcinoid Tumor

### Introduction

Less than 1 % of carcinoid tumors arise in the genitourinary system, with the testes and ovaries being the most common locations, followed by

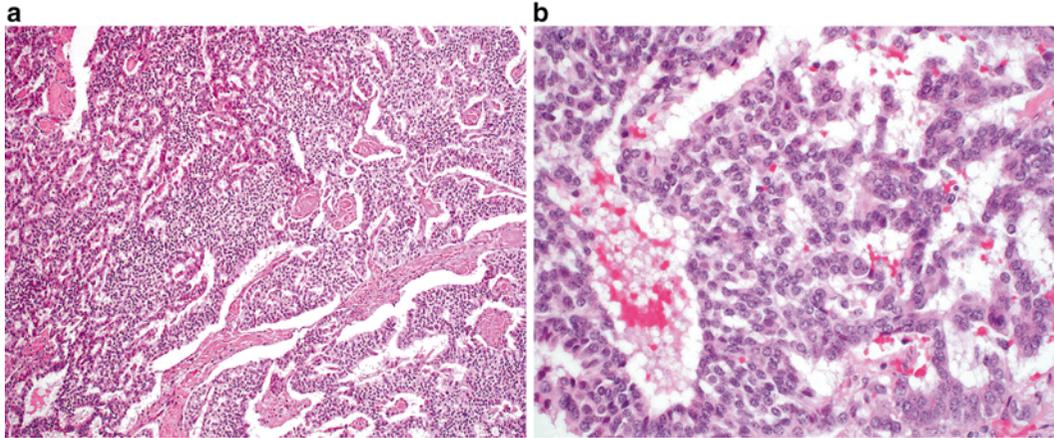
the kidneys [85]. Primary renal carcinoid tumor of the kidney is very rare, and less than 100 cases are reported in the literature [5, 86]. Age of diagnosis is broad, ranging from 13 to 79 years [86, 87]. There is an equal male-to-female distribution [5]. Intrinsic neuroendocrine cells in normal kidneys have not been reported previously; however, the pathogenesis of renal carcinoid tumors is controversial [86]. Carcinoid tumors in the kidney have been suggested to occur in patients with underlying renal congenital or acquired anomalies such as horseshoe kidney and mature teratoma [86, 88, 89]. These tumors demonstrate distinct morphological features from small cell carcinoma, which have also been reported to occur in pure form in the renal cortex [90, 91] or associated with invasive urothelial carcinoma of the renal pelvis [92].

### Clinical Presentation

Patients may present with abdominal pain or fullness, gross hematuria, and evidence of carcinoid syndrome, including serotonin-related flushing, edema, and less frequently diarrhea [5, 86, 93]. The diagnosis of primary renal carcinoid tumor is incidental in 25–30 % of cases [86, 94]. Imaging studies show a heterogeneous tumor, with solid and cystic components. Calcification has been reported in ~1/4 of cases [86, 95]. The tumors show minimal enhancement by CT imaging [86, 96].

### Pathology

On gross evaluation, renal carcinoid tumor is well circumscribed and soft and shows a homogeneous white-yellow cut surface. Calcifications, hemorrhage, and cystic change may be present [5, 86]. On microscopic review, tumor cells can show ribbonlike, trabecular, rosette-like, and rarely nested patterns (Fig. 9.8a) [5, 86]. Tumor cells have a monotonous appearance. Nuclei are relatively uniform and round and contain finely



**Fig. 9.8** Renal carcinoid tumor. (a) This example shows both trabecular and nested growth patterns. (b) Nuclei show the characteristic granular chromatin pattern associated with carcinoid lesions

granular chromatin (Fig. 9.8b) [5, 86, 89]. Rarely, moderate nuclear atypia or mitotic figures can be identified.

The genetic profile of renal carcinoid tumor has not been defined due to a small number of cases studied. One study has reported heterozygosity of 3p21 [5, 97].

The immunohistochemical profile of renal carcinoid is similar to neuroendocrine tumors in other locations. Synaptophysin and chromogranin are positive in the majority of tumors [5, 89, 98]. In addition, tumor cells can show dot-like immunoreactivity for CAM5.2 and may be positive for vimentin and cytokeratin 7 [89]. Renal carcinoid tumor is negative for TTF-1, WT-1, PAX2, and PAX8 [5, 89, 98].

Renal carcinoid tumor should be differentiated from other lesions that demonstrate neuroendocrine features, including small cell carcinoma, primitive neuroectodermal tumor (PNET), pheochromocytoma, and neuroblastoma. Small cell carcinoma is distinguished by the presence of classic features that include frequent mitotic features, apoptotic debris, necrosis, and nuclear molding. Morphology and clinical information is most helpful, as CD99, and typical neuroendocrine markers will not distinguish these entities [89, 98]. S100 immunostain may be helpful when neuroblastoma

and pheochromocytoma (sustentacular cells) are in the differential diagnosis. Metastases from other primary sites (such as the gastrointestinal tract) should be clinically excluded. Of note, PAX8 expression supports the diagnosis of a metastatic carcinoid as primary renal carcinoids have been shown to be negative for PAX8, whereas a subset of GI primary carcinoid tumors are positive for polyclonal PAX8 [98–100].

## Prognosis and Clinical Management

Distant metastases to the liver, bone, and orbit have been reported [82, 85, 86, 89].

## References

1. Srigley JR, Delahunt B, Eble JN, Egevad L, Epstein JI, Grignon D, et al. The International Society of Urological Pathology (ISUP) Vancouver classification of renal neoplasia. *Am J Surg Pathol*. 2013;37(10):1469–89.
2. Eble J, Sauter G, Epstein J, et al. *Pathology and genetics of tumours of the urinary system and male genital organs*. Lyon: IARC; 2004.
3. Motzer RJ, Jonasch E, Agarwal N, Beard C, Bhayani S, Bolger GB, et al. *Kidney cancer, version 3.2015*. *J Natl Compr Canc Netw*. 2015;13(2):151–9.

4. Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, et al. Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. *N Engl J Med*. 2007;356(2):115–24.
5. Hansel DE, Zhou M. Practical renal pathology, a diagnostic approach. 1st ed. Philadelphia: Elsevier Saunders; 2013. p. 1–22.
6. Crumley SM, Divatia M, Truong L, Shen S, Ayala AG, Ro JY. Renal cell carcinoma: evolving and emerging subtypes. *World J Clin Cases*. 2013;1(9):262–75.
7. MacLennan GT, Bostwick DG. Tubulocystic carcinoma, mucinous tubular and spindle cell carcinoma, and other recently described rare renal tumors. *Clin Lab Med*. 2005;25(2):393–416.
8. MacLennan GT, Farrow GM, Bostwick DG. Low-grade collecting duct carcinoma of the kidney: report of 13 cases of low-grade mucinous tubulocystic renal carcinoma of possible collecting duct origin. *Urology*. 1997;50(5):679–84.
9. Eble JN. Mucinous tubular and spindle cell carcinoma and post-neuroblastoma carcinoma: newly recognised entities in the renal cell carcinoma family. *Pathology*. 2003;35(6):499–504.
10. Hes O, Hora M, Perez-Montiel DM, Suster S, Curík R, Sokol L, et al. Spindle and cuboidal renal cell carcinoma, a tumor having frequent association with nephrolithiasis: report of 11 cases including a case with hybrid conventional renal cell carcinoma/ spindle and cuboidal renal cell carcinoma components. *Histopathology*. 2002;41(6):549–55.
11. Prasad SR, Humphrey PA, Catena JR, Narra VR, Srigley JR, Cortez AD, et al. Common and uncommon histologic subtypes of renal cell carcinoma: imaging spectrum with pathologic correlation. *Radiographics*. 2006;26(6):1795–806.
12. Sahni VA, Hirsch MS, Sadow CA, Silverman SG. Mucinous tubular and spindle cell carcinoma of the kidney: imaging features. *Cancer Imaging*. 2012;12:66–71.
13. Ursani NA, Robertson AR, Schieman SM, Bainbridge T, Srigley JR. Mucinous tubular and spindle cell carcinoma of kidney without sarcomatoid change showing metastases to liver and retroperitoneal lymph node. *Hum Pathol*. 2011;42(3):444–8.
14. Bulimbasic S, Ljubanovic D, Sima R, Michal M, Hes O, Kuroda N, et al. Aggressive high-grade mucinous tubular and spindle cell carcinoma. *Hum Pathol*. 2009;40(6):906–7.
15. Pillay N, Ramdial PK, Cooper K, Batuule D. Mucinous tubular and spindle cell carcinoma with aggressive histomorphology – a sarcomatoid variant. *Hum Pathol*. 2008;39(6):966–9.
16. Rakozzy C, Schmahl GE, Bogner S, Störkel S. Low-grade tubular-mucinous renal neoplasms: morphologic, immunohistochemical, and genetic features. *Mod Pathol*. 2002;15(11):1162–71.
17. Cossu-Rocca P, Eble JN, Delahunt B, Zhang S, Martignoni G, Brunelli M, et al. Renal mucinous tubular and spindle carcinoma lacks the gains of chromosomes 7 and 17 and losses of chromosome Y that are prevalent in papillary renal cell carcinoma. *Mod Pathol*. 2006;19(4):488–93.
18. Ferlicot S, Allory Y, Comperat E, Mege-Lechevalier F, Dimet S, Sibony M, et al. Mucinous tubular and spindle cell carcinoma: a report of 15 cases and a review of the literature. *Virchows Arch*. 2005;447(6):978–83.
19. Paner GP, Srigley JR, Radhakrishnan A, Cohen C, Skinnider BF, Tickoo SK, et al. Immunohistochemical analysis of mucinous tubular and spindle cell carcinoma and papillary renal cell carcinoma of the kidney: significant immunophenotypic overlap warrants diagnostic caution. *Am J Surg Pathol*. 2006;30(1):13–9.
20. Kuroda N, Toi M, Hiroi M, Shuin T, Enzan H. Review of mucinous tubular and spindle-cell carcinoma of the kidney with a focus on clinical and pathobiological aspects. *Histol Histopathol*. 2005;20(1):221–4.
21. Parwani AV, Husain AN, Epstein JI, Beckwith JB, Argani P. Low-grade myxoid renal epithelial neoplasms with distal nephron differentiation. *Hum Pathol*. 2001;32(5):506–12.
22. Shen SS, Ro JY, Tamboli P, Truong LD, Zhai Q, Jung SJ, et al. Mucinous tubular and spindle cell carcinoma of kidney is probably a variant of papillary renal cell carcinoma with spindle cell features. *Ann Diagn Pathol*. 2007;11(1):13–21.
23. Thway K, du Parcq J, Larkin JM, Fisher C, Livni N. Metastatic renal mucinous tubular and spindle cell carcinoma. Atypical behavior of a rare, morphologically bland tumor. *Ann Diagn Pathol*. 2012;16(5):407–10.
24. Kuroda N, Hes O, Michal M, Nemcova J, Gal V, Yamaguchi T, et al. Mucinous tubular and spindle cell carcinoma with Fuhrman nuclear grade 3: a histological, immunohistochemical, ultrastructural and FISH study. *Histol Histopathol*. 2008;23(12):1517–23.
25. Simon RA, di Sant'agnese PA, Palapattu GS, Singer EA, Candelario GD, Huang J, et al. Mucinous tubular and spindle cell carcinoma of the kidney with sarcomatoid differentiation. *Int J Clin Exp Pathol*. 2008;1(2):180–4.
26. Williamson SR, Eble JN, Cheng L, Grignon DJ. Clear cell papillary renal cell carcinoma: differential diagnosis and extended immunohistochemical profile. *Mod Pathol*. 2013;26(5):697–708.
27. Zhou H, Zheng S, Truong LD, Ro JY, Ayala AG, Shen SS. Clear cell papillary renal cell carcinoma is the fourth most common histologic type of renal cell carcinoma in 290 consecutive nephrectomies for renal cell carcinoma. *Hum Pathol*. 2014;45(1):59–64.
28. Alexiev BA, Drachenberg CB. Clear cell papillary renal cell carcinoma: incidence, morphological features, immunohistochemical profile, and biologic behavior—a single institution study. *Pathol Res Pract*. 2014;210(4):234–41.
29. Tickoo SK, de Peralta-Venturina MN, Harik LR, Worcester HD, Salama ME, Young AN, et al.

- Spectrum of epithelial neoplasms in end-stage renal disease: an experience from 66 tumor-bearing kidneys with emphasis on histologic patterns distinct from those in sporadic adult renal neoplasia. *Am J Surg Pathol.* 2006;30:141–53.
30. Gobbo S, Eble JN, Grignon DJ, Martignoni G, MacLennan GT, Shah RB, et al. Clear cell papillary renal cell carcinoma: a distinct histopathologic and molecular genetic entity. *Am J Surg Pathol.* 2008;32:1239–45.
  31. Aydin H, Chen L, Cheng L, Vaziri S, He H, Ganapathi R, et al. Clear cell tubulopapillary renal cell carcinoma: a study of 36 distinctive low-grade epithelial tumors of the kidney. *Am J Surg Pathol.* 2010;34(11):1608–21.
  32. Aron M, Chang E, Herrera L, Hes O, Hirsch MS, Comperat E, et al. Clear cell-papillary renal cell carcinoma of the kidney not associated with end-stage renal disease: clinicopathologic correlation with expanded immunophenotypic and molecular characterization of a large cohort with emphasis on relationship with renal angiomyoadenomatous tumor. *Am J Surg Pathol.* 2015;39(7):873–88.
  33. Deml KF, Schildhaus HU, Compérat E, von Teichman A, Storz M, Schraml P, et al. Clear cell papillary renal cell carcinoma and renal angiomyoadenomatous tumor: two variants of a morphologic, immunohistochemical, and genetic distinct entity of renal cell carcinoma. *Am J Surg Pathol.* 2015;39(7):889–901.
  34. Park JH, Lee C, Suh JH, Moon KC. Clear cell papillary renal cell carcinoma: a report of 15 cases including three cases of concurrent other-type renal cell carcinomas. *Korean J Pathol.* 2012;46(6):541–7.
  35. Michal M, Hes O, Nemcova J, Sima R, Kuroda N, Bulimbasic S, et al. Renal angiomyoadenomatous tumor: morphologic, immunohistochemical, and molecular genetic study of a distinct entity. *Virchows Arch.* 2009;454(1):89–99.
  36. Fisher KE, Yin-Goen Q, Alexis D, Sirintrapun JS, Harrison W, Benjamin Issett R, et al. Gene expression profiling of clear cell papillary renal cell carcinoma: comparison with clear cell renal cell carcinoma and papillary renal cell carcinoma. *Mod Pathol.* 2014;27(2):222–30.
  37. Rohan SM, Xiao Y, Liang Y, Dudas ME, Al-Ahmadie HA, Fine SW, et al. Clear-cell papillary renal cell carcinoma: molecular and immunohistochemical analysis with emphasis on the von Hippel-Lindau gene and hypoxia-inducible factor pathway-related proteins. *Mod Pathol.* 2011;24:1207–20.
  38. Munari E, Marchionni L, Chitre A, Hayashi M, Martignoni G, Brunelli M, et al. Clear cell papillary renal cell carcinoma: micro-RNA expression profiling and comparison with clear cell renal cell carcinoma and papillary renal cell carcinoma. *Hum Pathol.* 2014;45(6):1130–8.
  39. Lawrie CH, Larrea E, Larrinaga G, Goicoechea I, Arestin M, Fernandez-Mercado M, et al. Targeted next-generation sequencing and non-coding RNA expression analysis of clear cell papillary renal cell carcinoma suggests distinct pathological mechanisms from other renal tumour subtypes. *J Pathol.* 2014;232(1):32–42.
  40. Adam J, Couturier J, Molinié V, Vieillefond A, Sibony M. Clear-cell papillary renal cell carcinoma: 24 cases of a distinct low-grade renal tumour and a comparative genomic hybridization array study of seven cases. *Histopathology.* 2011;58:1064–71.
  41. Inoue T, Matsuura K, Yoshimoto T, Nguyen LT, Tsukamoto Y, Nakada C, et al. Genomic profiling of renal cell carcinoma in patients with end-stage renal disease. *Cancer Sci.* 2012;103(3):569–76.
  42. Kuroda N, Ohe C, Kawakami F, Mikami S, Furuya M, Matsuura K, et al. Clear cell papillary renal cell carcinoma: a review. *Int J Clin Exp Pathol.* 2014;7(11):7312–8.
  43. Osunkoya AO, Young AN, Wang W, Netto GJ, Epstein JI. Comparison of gene expression profiles in tubulocystic carcinoma and collecting duct carcinoma of the kidney. *Am J Surg Pathol.* 2009;33:1103–6.
  44. Yang XJ, Zhou M, Hes O, Shen S, Li R, Lopez J, et al. Tubulocystic carcinoma of the kidney: clinicopathologic and molecular characterization. *Am J Surg Pathol.* 2008;32:177–87.
  45. Zhou M, Yang XJ, Lopez JI, Shah RB, Hes O, Shen SS, et al. Renal tubulocystic carcinoma is closely related to papillary renal cell carcinoma: implications for pathologic classification. *Am J Surg Pathol.* 2009;33:1840–9.
  46. Quiroga-Garza G, Piña-Oviedo S, Cuevas-Ocampo K, Goldfarb R, Schwartz MR, Ayala AG, et al. Synchronous clear cell renal cell carcinoma and tubulocystic carcinoma: genetic evidence of independent ontogenesis and implications of chromosomal imbalances in tumor progression. *Diagn Pathol.* 2012;7:21.
  47. Bhullar JS, Varshney N, Bhullar AK, Mittal VK. A new type of renal cancer-tubulocystic carcinoma of the kidney: a review of the literature. *Int J Surg Pathol.* 2013;22(4):297–302.
  48. Hora M, Michal M, Hes O. Cystic nephroma and mixed epithelial and stromal tumour of the kidney: opposite ends of the spectrum of the same entity? *Eur Urol.* 2008;54:1237–46.
  49. Kryvenko ON, Jorda M, Argani P, Epstein JI. Diagnostic approach to eosinophilic renal neoplasms. *Arch Pathol Lab Med.* 2014; 138(11):1531–41.
  50. Alexiev BA, Drachenberg CB. Tubulocystic carcinoma of the kidney: a histologic, immunohistochemical, and ultrastructural study. *Virchows Arch.* 2013;462(5):575–81.
  51. Amin MB, MacLennan GT, Gupta R, Grignon D, Paraf F, Vieillefond A, et al. Tubulocystic carcinoma of the kidney: clinicopathologic analysis of 31 cases of a distinctive rare subtype of renal cell carcinoma. *Am J Surg Pathol.* 2009;33:384–92.

52. Bhullar JS, Thamboo T, Esuvaranathan K. Unique case of tubulocystic carcinoma of the kidney with sarcomatoid features: a new entity. *Urology*. 2011;78(5):1071–2.
53. Chen N, Nie L, Gong J, Chen X, Xu M, Chen M, et al. Gains of chromosomes 7 and 17 in tubulocystic carcinoma of kidney: two cases with fluorescence in situ hybridisation analysis. *J Clin Pathol*. 2014;67(11):1006–9.
54. Azoulay S, Vieillefond A, Paraf F, et al. Tubulocystic carcinoma of the kidney: a new entity among renal tumors. *Virchows Arch*. 2007;451:905–9.
55. Eble JN, Bonsib SM. Extensively cystic renal neoplasms: cystic nephroma, cystic partially differential nephroblastoma, multi-locular cyst renal cell carcinoma, and cystic hamartoma of renal pelvis. *Semin Diagn Pathol*. 1998;15:2–20.
56. Al-Hussain TO, Cheng L, Zhang S, Epstein JI. Tubulocystic carcinoma of the kidney with poorly differentiated foci: a series of 3 cases with fluorescence in situ hybridization analysis. *Hum Pathol*. 2013;44(7):1406–11.
57. Denton MD, Magee CC, Ovuworie C, Mauyyedi S, Pascual M, Colvin RB, et al. Prevalence of renal cell carcinoma in patients with ESRD pre-transplantation: a pathologic analysis. *Kidney Int*. 2002;61:2201–9.
58. Hughson MD, Buchwald D, Fox M. Renal neoplasia and acquired cystic kidney disease in patients receiving longterm dialysis. *Arch Pathol Lab Med*. 1986;110:592–601.
59. Sule N, Yakupoglu U, Shen SS, Krishnan B, Yang G, Lerner S, et al. Calcium oxalate deposition in renal cell carcinoma associated with acquired cystic kidney disease: a comprehensive study. *Am J Surg Pathol*. 2005;29:443–51.
60. Pan CC, Chen YJ, Chang LC, Chang YH, Ho DM. Immunohistochemical and molecular genetic profiling of acquired cystic disease-associated renal cell carcinoma. *Histopathology*. 2009;55:145–53.
61. Kuroda N, Tamura M, Taguchi T, Tominaga A, Hes O, Michal M, et al. Sarcomatoid acquired cystic disease-associated renal cell carcinoma. *Histol Histopathol*. 2008;23:1327–31.
62. Ishikawa I, Saito Y, Asaka M, Tomosugi N, Yuri T, Watanabe M, et al. Twenty-year follow-up of acquired renal cystic disease. *Clin Nephrol*. 2003;59:153–9.
63. Medeiros LJ, Palmedo G, Krigman HR, et al. Oncocytoid renal cell carcinoma after neuroblastoma: a report of four cases of a distinct clinicopathologic entity. *Am J Surg Pathol*. 1999;23(7):772–80.
64. Kuroda N, Tamura M, Hamaguchi N, Mikami S, Pan CC, Brunelli M, et al. Acquired cystic disease-associated renal cell carcinoma with sarcomatoid change and rhabdoid features. *Ann Diagn Pathol*. 2011;15(6):462–6.
65. Bhatnagar R, Alexiev BA. Renal-cell carcinomas in end-stage kidneys: a clinicopathological study with emphasis on clear-cell papillary renal-cell carcinoma and acquired cystic kidney disease-associated carcinoma. *Int J Surg Pathol*. 2012;20(1):19–28.
66. Fleitz JM, Wootton-Gorges SL, Wyatt-Ashmead J, McGavran L, Koyle M, West DC, et al. Renal cell carcinoma in long-term survivors of advanced stage neuroblastoma in early childhood. *Pediatr Radiol*. 2003;33(8):540–5.
67. Altinok G, Kattar MM, Mohamed A, Poulik J, Grignon D, Rabah R. Pediatric renal carcinoma associated with Xp11.2 translocations/TFE3 gene fusions and clinicopathologic associations. *Pediatr Dev Pathol*. 2005;8(2):168–80.
68. Koyle MA, Hatch DA, Furness PD, et al. Long-term urological complications in survivors younger than 15 months of advanced stage abdominal neuroblastoma. *J Urol*. 2001;166(4):1455–8.
69. Hedgepeth RC, Zhou M, Ross J. Rapid development of metastatic Xp11 translocation renal cell carcinoma in a girl treated for neuroblastoma. *J Pediatr Hematol Oncol*. 2009;31(8):602–4.
70. Vogelzang NJ, Yang X, Goldman S, et al. Radiation induced renal cell cancer: a report of 4 cases and review of the literature. *J Urol*. 1998;160(6 Pt 1):1987–90.
71. Ghaouti M, Roquet L, Baron M, Pfister C, Sabourin JC. Thyroid-like follicular carcinoma of the kidney: a case report and review of the literature. *Diagn Pathol*. 2014;9:186.
72. Jung SJ, Chung JI, Park SH, Ayala AG, Ro JY. Thyroid follicular carcinoma-like tumor of kidney: a case report with morphologic, immunohistochemical, and genetic analysis. *Am J Surg Pathol*. 2006;30:411–5.
73. Amin MB, Gupta R, Ondrej H, McKenney JK, Michal M, Young AN, et al. Primary thyroidlike follicular carcinoma of the kidney: report of 6 cases of a histologically distinctive adult epithelial neoplasm. *Am J Surg Pathol*. 2009;33:393–400.
74. Khoja HA, Almutawa A, Binmahfooz A, Aslam M, Ghazi AA, Almainan S. Papillary thyroid carcinoma-like tumor of the kidney: a case report. *Int J Surg Pathol*. 2012;20:411–5.
75. Malde S, Sheikh I, Woodman I, Fish D, Bilagi P, Sheriff MK. Primary thyroid-like follicular renal cell carcinoma: an emerging entity. *Case Rep Pathol*. 2013;2013:687427.
76. Argani P, Olgac S, Tickoo SK, Goldfischer M, Moch H, Chan DY, et al. Xp11 translocation renal cell carcinoma in adults: expanded clinical, pathologic, and genetic spectrum. *Am J Surg Pathol*. 2007;31:1149–60.
77. Srigley J. Mucinous tubular and spindle cell carcinoma. In: Eble J, Sauter G, Epstein J, Sesterhenn I, editors. *Pathology and genetics of tumours of the urinary system and male genital organs*. Lyon: IARC; 2004. p. 40.
78. Williamson SR, Eble JN, Amin MB, Gupta NS, Smith SC, Sholl LM, et al. Succinate dehydrogenase-deficient renal cell carcinoma: detailed characterization

- of 11 tumors defining a unique subtype of renal cell carcinoma. *Mod Pathol.* 2015;28(1):80–94.
79. Gill AJ et al. Succinate dehydrogenase (SDH)-deficient renal carcinoma: a morphologically distinct entity: a clinicopathologic series of 36 tumors from 27 patients. *Am J Surg Pathol.* 2014;38(12):1588–602.
80. Henderson A, Douglas F, Perros P, et al. SDHB-associated renal oncocytoma suggests a broadening of the renal phenotype in hereditary paragangliomatosis. *Fam Cancer.* 2009;8:257–60.
81. Gill AJ, Pachter NS, Chou A, et al. Renal tumors associated with germline SDHB mutation show distinctive morphology. *Am J Surg Pathol.* 2011;35:1578–85.
82. Hakimi AA, Tickoo SK, Jacobsen A, Sarungbam J, Sfakianos JP, Sato Y, et al. TCEB1-mutated renal cell carcinoma: a distinct genomic and morphological subtype. *Mod Pathol.* 2015;28(6):845–53.
83. Hirsch MS, Barletta J, Gorman M, Dal Cin P. Renal cell carcinoma with monosomy 8 and CAIX expression: a distinct entity or another member of the clear cell tubulopapillary RCC/RAT family. *Mod Pathol.* 2015;28(S2):229A.
84. Sato Y, Yoshizato T, Shiraishi Y, Maekawa S, Okuno Y, Kamura T, et al. Integrated molecular analysis of clear-cell renal cell carcinoma. *Nat Genet.* 2013;45(8):860–7.
85. Parikh D, Shinder R. Primary renal carcinoid metastatic to the orbit. *Ophthal Plast Reconstr Surg.* 2015;31(2):e37–8.
86. Tanaka T, Yamamoto H, Imai A, Shingo H, Yoneyama T, Koie T, et al. A case of primary renal carcinoid tumor. *Case Rep Urol.* 2015;2015:736213.
87. Krishnan B, Truong LD, Saleh G, Sirbasku DM, Slawin KM. Horseshoe kidney is associated with an increased relative risk of primary renal carcinoid tumor. *J Urol.* 1997;157(6):2059–66.
88. McCaffrey JA, Reuter V, Herr HW, Macapinlac HA, Russo P, Motzer RJ. Carcinoid tumor of the kidney: the use of somatostatin receptor scintigraphy in diagnosis and management. *Urol Oncol.* 2000;5(3):108–11.
89. Hansel DE, Epstein JI, Berbesu E, Fine SW, Young RH, Cheville JC. Renal carcinoid tumor: a clinicopathologic study of 21 cases. *Am J Surg Pathol.* 2007;31(10):1539–44.
90. Si Q, Dancer J, Stanton ML, Tamboli P, Ro JY, Czerniak BA, et al. Small cell carcinoma of the kidney: a clinicopathologic study of 14 cases. *Hum Pathol.* 2011;42(11):1792–8.
91. Kuroda N, Imamura Y, Hamashima T, Ohe C, Mikami S, Nagashima Y, et al. Review of small cell carcinoma of the kidney with focus on clinical and pathobiological aspects. *Pol J Pathol.* 2014;65(1):15–9.
92. Miller RJ, Holmång S, Johansson SL, Lele SM. Small cell carcinoma of the renal pelvis and ureter: clinicopathologic and immunohistochemical features. *Arch Pathol Lab Med.* 2011;135(12):1565–9.
93. Raslan WF, Ro JY, Ordonez NG, et al. Primary carcinoid of the kidney. Immunohistochemical and ultrastructural studies of five patients. *Cancer.* 1993;72(9):2660–6.
94. Murali R, Kneale K, Lalak N, Delprado W. Carcinoid tumors of the urinary tract and prostate. *Arch Pathol Lab Med.* 2006;130(11):1693–706.
95. Romero FR, Rais-Bahrami S, Permpongkosol S, Fine SW, Kohanim S, Jarrett TW. Primary carcinoid tumors of the kidney. *J Urol.* 2006;176(6):2359–66.
96. Kurl S, Rytkänonen H, Farin P, Ala-Opas M, Soimakallio S. A primary carcinoid tumor of the kidney: a case report and review of the literature. *Abdom Imaging.* 1996;21(5):464–7.
97. El-Naggar AK, Troncoso P, Ordonez NG. Primary renal carcinoid tumor with molecular abnormality characteristic of conventional renal cell neoplasms. *Diagn Mol Pathol.* 1995;4(1):48–53.
98. Jeung JA, Cao D, Selli BW, Clapp WL, Oliari BR, Parwani AV, et al. Primary renal carcinoid tumors: clinicopathologic features of 9 cases with emphasis on novel immunohistochemical findings. *Hum Pathol.* 2011;42(10):1554–61.
99. Long KB, Srivastava A, Hirsch MS, Hornick JL. PAX8 expression in well-differentiated pancreatic endocrine tumors: correlation with clinicopathologic features and comparison with gastrointestinal and pulmonary carcinoid tumors. *Am J Surg Pathol.* 2010;34(5):723–9.
100. Sangoi AR, Ohgami RS, Pai RK, Beck AH, McKenney JK, Pai RK. PAX8 expression reliably distinguishes pancreatic well-differentiated neuroendocrine tumors from ileal and pulmonary well-differentiated neuroendocrine tumors and pancreatic acinar cell carcinoma. *Mod Pathol.* 2011;24(3):412–24.