Chapter 15 The Pulmonary Vasculature in Chronic Obstructive Lung Disease

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15.1 Chronic Obstructive Pulmonary Disease (COPD) and its Association with Pulmonary Hypertension (PH)

Chronic obstructive lung disease, or chronic obstructive pulmonary disease (COPD), a collective term for chronic bronchitis and pulmonary emphysema, is one of the major causes of death worldwide. The World Health Organization (WHO) predicts that by 2030, COPD will rank as the third greatest cause of death worldwide. It is characterized by progressive, poorly reversible airflow limitation associated with an abnormal chronic inflammatory response in the lung. COPD is also viewed as a systemic disease, involving skeletal muscle wasting, diaphragmatic dysfunction, and systemic inflammation [[1](#page-19-0)]. In industrialized countries, tobacco smoke (80–90%) and air pollution are the major triggers for the development of COPD, whereas in developing countries, exposure to biomass smoke, especially during cooking, is most relevant [[2\]](#page-19-1).

On the one hand, COPD is viewed as a small airway disease, with destruction of the elastic architecture of the lung leading to enlargement of distal airspaces [[3](#page-19-2)] associated with chronic inflammation of the airways [[4](#page-19-3), [5\]](#page-19-4). Influx of inflammatory cells, imbalance of proteases/antiproteases, increased oxidative stress with the con-

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comitant rise in the number of apoptotic cells, and decreased proliferation are likely important upstream events [[4](#page-19-3), [6](#page-19-5), [7\]](#page-19-6).

On the other hand, there is increasing evidence that COPD is also a vascular disease in which cigarette smoke may directly affect the pulmonary vasculature, leading to vascular remodeling, pulmonary hypertension (PH), and finally to cor pulmonale [[8](#page-19-7)–[10](#page-19-8)]. The mean pulmonary artery pressure (mPAP) in PH related to COPD usually ranges between 25 and 35 mmHg, with nearly normal cardiac output [[11](#page-19-9)]. However, mPAPs higher than 40 mmHg are quite common in patients with severe COPD, especially after at least one previous incident of acute respiratory failure [[11](#page-19-9), [12](#page-19-10)]. Furthermore, the occurrence of PH in COPD is of prognostic relevance because the presence of PH clearly increases mortality [[13](#page-19-11), [14\]](#page-19-12).

The exact prevalence of PH in patients with mild or moderate COPD is not known because of the lack of extensive epidemiologic studies [[2](#page-19-1)]. According to the current literature, 30–70% of patients with mild or moderate COPD also suffer from PH [[15\]](#page-19-13). There are several reasons for the high variation in the rate of observed PH. The necessary right heart catheterization for detection of PH was/is not routinely used in patients with COPD. In addition, the respective studies used different definitions for the onset of PH and varied in study design (measuring at rest/exercise, severity of COPD/PH, methods to measure PH). Some studies have investigated PH in mild, moderate, or severe COPD, with frequencies of PH of approximately 16–44, 43–56, and 59–84%, respectively [[2](#page-19-1), [16](#page-19-14), [17](#page-19-15)].

PH is a common complication in advanced COPD [[18](#page-19-16)] and can occur as a consequence of the hypoxia associated with COPD. However, several publications showed that vascular alterations can occur before alveolar destruction is detectable [[19](#page-19-17)–[22](#page-20-0)], indicating that cor pulmonale and late-stage PH are not necessarily secondary to hypoxia in patients with COPD. For instance, in 2003 Santos and colleagues showed that pulmonary vascular remodeling can be seen in smokers who are not yet suffering from COPD [[23](#page-20-1)].

Thus, although PH can occur prior to lung emphysema, its relevance for the pathogenesis of COPD is not yet fully understood. Against this background, in this chapter we summarize the current findings of vascular alterations and pathobiology in COPD.

15.2 PH as a Cause for Right Ventricular Failure in COPD

The progression of PH in COPD is usually slow and pulmonary artery mean pressure (mPAP) can remain stable over periods of 3–12 years [[24](#page-20-2)–[26](#page-20-3)]. In a study following 93 patients during a mean period of 90 months, the average change in mPAP was only+0.5 mmHg/year, independent of the presence of initial PH (defined by mPAP >20 mmHg) [[26](#page-20-3)]. Another study investigating the development of PH in COPD over time (initial mPAP <20 mmHg) showed that only 33/121 developed PH after 6.8 ± 2.9 years [[27](#page-20-4)]. Nevertheless, approximately 30% of patients with severe COPD exhibited a remarkable worsening of mPAP during follow-up [[26](#page-20-3)]; these patients were characterized by a progressive worsening of PaO_2 and $PaCO_2$ (partial oxygen/carbon dioxide pressure) during the time course, and there was a significant correlation between the changes in PaO_2 and mPAP [\[25](#page-20-5), [26](#page-20-3)]. Typically, the development of right heart failure (RHF) in patients with chronic respiratory disease is accompanied with the preceding occurrence of PH. The severity of PH and the development of RHF correlate. PH increases the workload of the right ventricle, leading to right ventricular enlargement (hypertrophy plus dilatation), which can result in right ventricular dysfunction. The RHF is usually accompanied by peripheral edema and can be observed in some COPD patients [[28](#page-20-6), [29](#page-20-7)]. Peripheral edema is frequently observed in advanced COPD patients and is considered to reflect RHF, but the possible occurrence of RHF in these patients may simply indicate the presence of secondary hyperaldosteronism induced by functional renal insufficiency [[30\]](#page-20-8).

The role of pressure overload in the development of RHF in these patients has also been debated. The comparison of COPD patients with and without clinical (edema) and hemodynamic signs of RHF led to the conclusion that RHF was probably due to causes other than PH [[31](#page-20-9)]. In stable conditions, right ventricular contractility, measured by the end-systolic pressure–volume relation, is near normal in COPD patients with PH, but has been found to be decreased during severe exacerbations with marked peripheral edema [[31](#page-20-9)]. Hence, many patients with advanced COPD will never develop RHF. At least some patients experience sequences of RHF during exacerbations associated with a worsening of PH [[32](#page-20-10)]. The level of mPAP is a good prognostic indicator in COPD [[32](#page-20-10), [33](#page-20-11)] and in other categories of chronic respiratory disease, such as idiopathic pulmonary fibrosis [[33](#page-20-11)] (the higher the degree of PH the worse the prognosis). It has been shown that life expectancy is worse in patients with PH compared with patients without PH [[32](#page-20-10)], and is particularly poor in patients with a severe degree of PH [[33](#page-20-11), [34](#page-20-12)]. The 5-year survival rate of COPD patients with PH (mPAP >20 mmHg) is approximately 50% [[26](#page-20-3), [33\]](#page-20-11). Long-term oxygen therapy (LTOT) significantly improves the survival of markedly hypoxemic COPD patients also suffering from PH. Accordingly, it can be expected that the prognosis of PH will improve with such a therapy in these patients. This hypothesis can be explained by the fact that mPAP is a good marker of both the duration and severity of alveolar hypoxia in these patients, assuming hypoxia is the causing factor for the PH [[14](#page-19-12)].

15.3 Vascular Alterations

15.3.1 The Process of Remodeling

Vascular changes can occur in smokers without COPD, can precede the development of emphysema in animal models, and can be seen in COPD patients. The respective vessels, especially in end-stage COPD, have thickened walls or are even occluded as a result of remodeling. Vascular remodeling is characterized by thickening of the arterial wall by which the vascular lumen and the internal diameter are reduced, leading to increased resistance and higher intravascular pressure. Of interest, although apparent in vessels of different sizes, muscular arteries and arterioles (small vessels with a diameter $\langle 500 \mu m \rangle$ are predominantly affected [\[35](#page-20-13), [36\]](#page-20-14). Pulmonary vascular remodeling has been observed at different degrees of disease severity. Indeed, this phenomenon could not only be seen in patients with mild and severe COPD but also in heavy smokers with normal lung function [[37](#page-20-15)]. The major event is hyperplasia of the intima [[35,](#page-20-13) [36](#page-20-14)], but the other vessel wall layers, the media and the adventitia, are also involved [[38](#page-20-16)]. Intimal hyperplasia results from the proliferation of α -smooth muscle actin- and vimentin-positive cells (corresponding to smooth muscle cells [SMC] associated with elastic and collagen fiber deposition) [[23](#page-20-1)]. Interestingly, some SMCs in the intima express vimentin but not desmin filaments [[23](#page-20-1)]. The presence of these filaments facilitates the discrimination between differentiated and nondifferentiated SMCs [[39](#page-20-17), [40](#page-20-18)]. "Accordingly, vimentin-positive, desmin-negative SMCs represent a subpopulation of less differentiated SMCs that may possess synthetic capacity and take part in an ongoing process of vascular remodeling" [[9](#page-19-18)]. Although not fully understood, the occurrence of these not fully differentiated cells could be explained by (i) differentiation from resident precursor cells; (ii) attraction and differentiation of circulating bone-marrow-derived progenitor cells; (iii) dedifferentiation of mature SMCs from the media which migrate to the intima [[41](#page-20-19)]; and (iv) transdifferentiation of endothelial cells (ECs) to SMCs by endothelial-to-mesenchymal transition [[42](#page-20-20)]. The underlying mechanisms are still not fully resolved and have to be deciphered in future. Interestingly, "bone-marrowderived progenitor vascular cells might exert a dual, opposite effect, contributing to vascular repair through differentiation into ECs or to vessel remodeling through differentiation into SMC". [[42](#page-20-20)].

The respective pathologic picture can be explained by remodeling and angiogenesis induced by inflammation that is related to chronic infection, hypoxia, repeated stretching of hyperinflated lungs, and toxicity of cigarette smoke [[43](#page-20-21), [44\]](#page-21-0).

Chronic hypoxia at high altitudes can cause PH, but the associated PH is completely reversible a few weeks after return to sea level [[11](#page-19-9)]. This finding may be explained by the observation that PH caused by high altitude primarily induces medial hypertrophy. The involvement of all vessel layers in PH associated with COPD likely explains why PH in COPD is often not reversible by supplemental oxygen, neither acutely [[45\]](#page-21-1) nor chronically [[46](#page-21-2)].

15.3.2 Mechanistic Insights into the Remodeling Process

Hypoxemia/hypoxia, chronic inflammation, oxidative and nitrosative stress, endothelial dysfunction, apoptosis, and altered proliferation have all been suggested as factors in the pathogenesis of airway and vascular remodeling [[6](#page-19-5), [9](#page-19-18), [47](#page-21-3)–[50\]](#page-21-4). Classically, hypoxia has been considered to be the major pathogenic factor for the development of PH in COPD. However, recent studies have shown that pulmonary vascular remodeling and endothelial dysfunction can occur in animal models of lung emphysema [[19](#page-19-17), [20](#page-19-19)], in patients with mild COPD not suffering from hypoxemia, and in smokers with normal lung function [[23](#page-20-1), [35,](#page-20-13) [51](#page-21-5)]. In addition, LTOT is not able to reverse PH in many COPD patients [\[52](#page-21-6)]. Nevertheless, there are many studies showing that hypoxia plays a role in COPD, at least in severe forms of the disease. Thus, mechanistic aspects related to hypoxia will be discussed. Furthermore, below we give an overview of the most relevant molecules and associated phenomena in the context of COPD and vascular remodeling/PH.

15.3.3 Hypoxia as a Causing Factor for Vascular Remodeling

In chronic respiratory diseases, PH is a result of increased vascular resistance (PVR), whereas cardiac output is often normal. The factors leading to the increase in PVR are manifold [[28](#page-20-6), [53](#page-21-7)] but hypoxia secondary to COPD is thought to be a predominant factor [[28](#page-20-6), [54](#page-21-8), [55\]](#page-21-9). In terms of hypoxia, two mechanisms should be considered: acute hypoxia causing pulmonary vasoconstriction, and chronic hypoxia leading to structural vascular changes (remodeling) over time. Acute hypoxia induces a rise of PVR and mPAP which is part of hypoxic pulmonary vasoconstriction (HPV). Chronic alveolar hypoxia causes precapillary PH in healthy people living in high altitude, similar to that observed in COPD patients. Additionally, the morphological changes of the pulmonary vascular bed (remodeling) are comparable with those of COPD patients with PH (muscularization of pulmonary arterioles, intima thickening in muscular pulmonary arteries and arterioles) [\[56](#page-21-10)], although under hypoxic conditions the media is mainly involved in the remodeling.

With regard to hypoxia, it makes sense to distinguish between generalized hypoxia (e.g. seen in high altitude) due to the low partial pressure of oxygen, and localized, regional alveolar hypoxia [[49](#page-21-11)]. Generalized hypoxia has both systemic and organ-specific effects [\[57](#page-21-12), [58\]](#page-21-13). It results in peripheral vasodilation, general pulmonary vasoconstriction, and activation of a sympathico-adrenergic stress response to increase cardiac output [[59\]](#page-21-14). Moreover, activation of bone marrow and erythropoietin-stimulated red-cell production occurs. Increased cardiac output and polycythemia are adaptive responses to improve blood and oxygen delivery to the hypoxic tissues. In contrast, local alveolar hypoxia occurs by regional lack of alveolar ventilation, e.g. caused by airway obstruction or even in pulmonary edema. HPV here adapts the local blood perfusion to the alveolar ventilation situation, preventing hypoxemia.

Hypoxia occurring within tumors is critical for tumorigenesis caused by the elevated demand for blood and nutrients of the rapidly growing cancer cells that are supplied by structurally abnormal tumor vessels. A similar impairment of the microenvironment oxygenation can occur in the lung because of parenchyma destruction and compression or loss of small vessels. In tumors and in the damaged and remodeled lung, the systemic arterial blood partial pressure of oxygen is usually, normal whereas the tissue microenvironment can be hypoxic.

Fig. 15.1 Selection of possible hypoxia-dependent mechanisms contributing to pulmonary vascular remodeling in chronic obstructive pulmonary disease-pulmonary hypertension *(COPD-PH)*. Cigarette smoke-induced airway obstruction and emphysema can result in hypoxia. Hypoxia, amongst others, activates hypoxia-inducible factor *(HIF)-1α*, which can trigger pathways associated with inflammation, the recruitment of bone-marrow-derived cells, and alterations in proliferation/apoptosis balance of vascular endothelial and smooth muscle cells *(SMCs)*. Increased proliferation of SMCs causes narrowing of the vessels, resulting in pulmonary hypertension. All important acronyms of the molecules are explained in the text

Discovery of the oxygen-dependent transcription factor 'hypoxia-inducible factor-1 α' (HIF-1 α) [[60](#page-21-15)] and subsequent discovery of the closely related HIF-2 α was a breakthrough for the understanding of the molecular response of cells to hypoxia (Fig. [15.1,](#page-5-0) [15.2\)](#page-6-0). HIF controls a variety of hypoxia-dependent genes that are involved in protection from hypoxia or pulmonary vascular remodeling, such as erythropoietin, glucose transporters, vascular endothelial growth factor (VEGF), endothelin-1 (ET-1) and nitric oxide (NO) synthases. HIF-1 α and HIF-2 α can also link hypoxia and inflammation [[61](#page-21-16)–[65\]](#page-21-17). It has been shown that within 24 h of hypoxia, inflammatory cells accumulate in the lungs, as measured by increased activity of myeloperoxidase. Subsequently, investigations at the molecular level showed that hypoxia-induced mitogenic factor (HIMF; also known as FIZZ1) is released by lung macrophages during stimulation with hypoxia. The proinflammatory and proangiogenic effects of HIMF are mediated by VEGF and its receptor VEGFR2 [[66\]](#page-21-18). HIMF is also able to recruit bone-marrow-derived mesenchymal cells to the lung

Fig. 15.2 Hypoxia-induced ion channel-mediated increase in proliferation, contraction, and decrease of apoptosis of smooth muscle cells *(SMCs)* contributing to pulmonary vascular remodeling. *K_v* and TASK-1 channels are downregulated and are less active after chronic hypoxia, leading to accumulation of *K+* within the cell (mediating apoptosis resistance) and membrane potential depolarization of the SMCs. This depolarization causes opening of voltage-operated Ca^{2+} channels *(VOCC)*, especially L-type channels, which mediate *Ca2+* entry. Hypoxia-dependent reactive oxygen species *(ROS)* regulation derived from nicotinamide adenine dinucleotide phosphate *(NADPH)* oxidases and/or mitochondria is suggested to inhibit the *K+* channels, although it is unclear whether an increase or decrease of *ROS* occurs in hypoxia. Transient receptor potential channel *(TRPC)*-mediated Ca^{2+} or Na^{+} influx (speculatively by gating K⁺ channels) was also shown to be essential for the intracellular Ca^{2+} increase in at least acute hypoxia. Ca^{2+} release from mitochondria and sarcoplasmic reticulum *(SR)* was shown to additionally increase *Ca2+* within the cell. Thus, mediated contraction and proliferation of the SMCs can contribute to vascular remodeling. *Colored arrows* depict either activation *(green)* or inhibition *(red)*

in mice [[67](#page-22-0)]. Interestingly, hypoxia can also induce toll-like receptors (TLR) 2 and 6 via HIF-1α [[68](#page-22-1)], which in turn can activate nuclear factor-κB (NF-κB), another proinflammatory transcription factor [[69](#page-22-2), [70](#page-22-3)]. The excess of chemotactic factors such as VEGF, HIMF, leukotriene B4 (LTB4), and others induced by hypoxia in the

lung can mobilize bone-marrow-derived cells (including mast cells, mesenchymal precursor cells, megakaryocytes, dendritic cells) and their chemotaxis to the lung. Consequently, hypoxic/hypoxemic inflammation is a systemic response [[49\]](#page-21-11).

There are many cytokines mediating inflammation but, in the context of PH, special attention was given to HIF-1 α -induced interleukin (IL)-6 because its messenger RNA (mRNA), as well as protein levels, were upregulated by chronic hypoxia in mice. Although IL-6-deficient mice showed less inflammation under hypoxic conditions, chronic hypoxic pulmonary hypertension (CHPH) was, however, only marginally reduced [[71](#page-22-4)]. In contrast, IL-6 overexpression, specifically in the lungs, showed enhanced muscularization of small pulmonary vessels and PH under normoxic conditions, indicating that IL-6 may contribute to vascular remodeling in lungs. Of interest, overexpression of IL-6 under hypoxia resulted in severe PH and vascular remodeling up to obliteration [[72](#page-22-5)]. Taken together, IL-6 is upregulated after chronic hypoxia in mouse lungs, but does not seem to be essential for CHPH development. Moreover, the combination of excessive IL-6 and chronic hypoxia seems to change the mode of vascular remodeling towards angioproliferation [[49\]](#page-21-11). Based on recent results, there are currently two theories put forth to explain this observation. The first is that IL-6 is able to exert its effects through modulation of 'bone morphogenetic protein receptor type 2' (BMPR2) through a signal transducer and activator of transcription 3 **(**STAT3)-microRNA cluster 17/92 pathway [[73\]](#page-22-6). BMPR2 is a member of the transforming growth factor (TGF)-β superfamily of growth factor receptors. It is expressed ubiquitously and is a member of many different signaling pathways, including pERK, JNK, Akt/PI3K [[74](#page-22-7), [75](#page-22-8)], pSmad1/5 [[76](#page-22-9)], and p-p38MAPK [[77](#page-22-10)]. Mutations in the BMPR2 gene are shown to be linked to PH development [[78](#page-22-11)], and, moreover, BMPR2 expression is reduced in the pulmonary vasculature in primary PH patients [[79](#page-22-12)]. The other prevailing theory is based on the observation that IL-6 can influence the balance between apoptosis and proliferation in pulmonary arterial SMCs (PASMC) and ECs (PAECs), leading to vascular remodeling. Overexpression of IL-6 induces the angioproliferative growth factor VEGF and intracellular extracellular signal-regulated kinase (ERK), resulting in increased proliferation. Simultaneously, IL-6 downregulates the growth inhibitor TGFβ and proapoptotic mitogen-activated protein kinases (MAPKs) [JNK1, p38MAPK], and upregulates the inhibitors of apoptosis Bcl-2 and survivin, leading to decreased apoptosis [[72](#page-22-5)]. In conclusion, inflammatory cytokines, especially IL-6, can trigger vascular remodeling by influencing signaling pathways which lead to more proliferation and less apoptosis of PASMCs and PAECs (Fig. [15.1](#page-5-0)).

Furthermore, HIF-1 α can induce PASMC depolarization by reduction of K⁺ channel expression and activity that is associated with intracellular K^+ accumulation, and increase intracellular Ca^{2+} concentration and pH by upregulating transient receptor potential channels (TRPC) and $\text{Na}^+\text{/H}^+$ exchanger [[80](#page-22-13)–[83](#page-22-14)], (Fig. [15.2](#page-6-0)). It has been shown that the currents of voltage-gated K^+ channels (Kv) are decreased under chronic hypoxic conditions [[84](#page-22-15)–[86](#page-23-0)], most likely mediated by reactive oxygen species (ROS) derived from mitochondria [[87](#page-23-1)–[91](#page-23-2)] and/or nicotinamide adenine dinucleotide phosphate (NADPH) oxidases, such as NOX4 [[92](#page-23-3)], and can be associated with an influx of calcium via TRPC [[93](#page-23-4)]. With regard to ROS in mediating hypoxic

responses, there is current controversy as to whether they are up- or downregulated during hypoxia [[94](#page-23-5), [95](#page-23-6)].

Moreover, the voltage-independent two-pore-domain K^+ channel, TWIK-related acid-sensitive K^+ channel (TASK)-1 is also inhibited by hypoxia, leading to membrane depolarization and calcium entry through L-type channels [[96](#page-23-7), [97\]](#page-23-8). Interestingly, it was recently shown that the inhibition of TASK-1 is also mediated by an ET-1-dependent mechanism [[98](#page-23-9)]. In addition to such channels, L-type Ca^{2+} channels are shown to be a major cellular Ca^{2+} entry pathway [[99](#page-23-10), [100](#page-23-11)], (Fig. [15.2](#page-6-0)).

All these ion alterations are associated with more contractile, apoptosis-resistant, proliferative, and migratory PASMCs under hypoxic conditions [[80](#page-22-13)–[83](#page-22-14), [100\]](#page-23-11). The role of HIF-2 $α$ in this context is still not clear.

Another recently discovered key player for the development of hypoxia-induced but also nonhypoxia-induced PH, is Fhl1, a protein known to be involved in muscle development (Fig. [15.1\)](#page-5-0). It could be demonstrated that HIF-1 α , as well as HIF-2 α , induced Fhl-1 expression not only in different animal models of PH but also in human patients with idiopathic pulmonary arterial (PA) hypertension [[101\]](#page-23-12). The increase in Fhl-1 causes elevated proliferation and migration of PASMCs, contributing to vascular remodeling [[101](#page-23-12)].

15.3.4 Hypoxia-Independent Mechanisms Leading to the Development of COPD and PH

Hypoxia has long been thought to be the primary driving force for the development of PH in COPD supported by studies showing a close relationship between mPAP and/or pulmonary resistance and alveolar hypoxia [[12](#page-19-10), [102](#page-23-13), [103](#page-23-14)]. However, evidence that the causal factors can be hypoxia-independent and more complex is severalfold. (i) It has been shown that oxygen therapy is not able to fully reverse PH in COPD [[46](#page-21-2), [52](#page-21-6)]. (ii) Histological investigation in lungs from COPD patients with PH revealed involvement of all vessel layers characterized by prominent intimal thickening, medial hypertrophy, and muscularization of small arterioles [[36](#page-20-14), [104](#page-23-15)], whereas hypoxia-induced vascular remodeling is mainly restricted to the vessel media. This finding is supported by the fact that these pulmonary vascular alterations also occurred in nonhypoxic patients with mild airflow obstruction and smokers without any parenchymal disorder, suggesting that vascular changes may be driven by mechanisms independent of hypoxia/hypoxemia [[23](#page-20-1)]. (iii) Consistent with the findings described in humans, a recent study demonstrated that in mice chronically exposed to tobacco smoke, pulmonary vascular remodeling and PH preceded the development of emphysema, and these changes were independent of hypoxia; there was neither hypoxia in the smoking chamber nor hypoxemia in the mice [[20\]](#page-19-19). In addition, this study showed that the expression of genes involved in important pathways associated with PH and COPD, such as apoptosis, proliferation, oxidative stress, extracellular matrix, and inflammation, followed a completely different pattern compared with that observed in chronic hypoxia-induced vascular remodeling, indicating that the smoke- and hypoxia-driven mechanisms were different [[20](#page-19-19)]. The

combination of cigarette smoke and hypoxia can have synergistic effects in terms of affecting the vasculature. Guinea pigs exposed to cigarette smoke and subsequently to hypoxia showed a stronger elevation of pulmonary artery pressure and a more pronounced remodeling in small vessels compared with exposure to only one of these stimuli [[105](#page-24-0)].

Additional animal studies reported a direct effect of cigarette smoke on the parenchyma and vasculature. Cigarette smoke can increase the expression of vasoactive mediators in pulmonary arteries [[106](#page-24-1)], affecting the gene expression in pulmonary arteries [[107](#page-24-2)]. The exposure of guinea pigs to chronic cigarette smoke induced emphysema, which was associated with a reduced lung capillary density [[108](#page-24-3)]. It was reported that cigarette smoke extract (CSE) can induce ET-1 in pulmonary artery ECs [[109](#page-24-4)] and reduce prostacyclin synthase expression [[110](#page-24-5)]. Moreover, CSE is able to induce the production of superoxide in ECs, which in turn reacts with NO to peroxynitrite [[111](#page-24-6)]. This radical can inactivate VEGFR2 signaling [[112](#page-24-7)], which is important for EC maintenance and growth. Of interest, CSE-induced EC apoptosis via p53 [[113](#page-24-8)] can be prevented by the phosphodiesterase (PDE)-5 inhibitor sildenafil [[114](#page-24-9)], which leads to increased levels of cyclic guanosine monophosphate (cGMP), an important second messenger. This finding suggests that the cGMP pathway is involved in this context and decreased after smoke exposure.

15.4 Impairment of the Endothelium: Endothelial Dysfunction

New advances regarding the pathogenesis of PH in COPD suggest that an endothelium-derived vasoconstrictor-dilator imbalance caused by endothelial dysfunction associated with decreased expression or uncoupling of endothelial NO synthase (eNOS) could be a driving force for PH development (Fig. [15.3](#page-10-0)). In this context, expression of VEGF and serotonin transporters seem to be increased [[37](#page-20-15), [38](#page-20-16), [115](#page-24-10), [116](#page-24-11)]. ECs are important for the regulation of vascular homeostasis [[117](#page-24-12)]. They reduce the vascular tone [[118](#page-24-13)] and regulate pulmonary vessel adaptation to increases in blood flow [[119](#page-24-14)] and hypoxia [[120](#page-24-15), [121](#page-24-16)]. Endothelial dysfunction has been reported in patients with end-stage COPD who had undergone lung transplantation [\[115\]](#page-24-10), as well as in patients with mild-to-moderate COPD [[37](#page-20-15)]. The endothelial function is influenced by the expression of vasoreactive mediators controlling vasoconstriction (also proproliferative for SMCs) or vasodilation (also antiproliferative for SMCs). On the one hand, it has been shown that the vasoconstrictive protein ET-1 was increased in patients with primary and secondary PH, including COPD patients [[122](#page-24-17)] and, on the other hand, vasodilative mediators such as eNOS [[123](#page-25-0), [124](#page-25-1)] and prostacyclin synthase (PGI2-S) [[110](#page-24-5)] were shown to be downregulated in pulmonary arteries of patients with severe COPD. CSE and/or acrolein (a potent αβ-unsaturated aldehyde contained in cigarette smoke) can decrease the expression of PGI2-S in human pulmonary artery ECs (HPAECs) [[110](#page-24-5)], supporting the hypothesis that its downregulation in COPD patients arises directly from the cigarette smoke ingredients. After exposure to cigarette smoke for 8 months, mice developed

Fig. 15.3 Endothelial dysfunction as a causing factor for the development of vascular remodeling. Cigarette smoke and inflammatory mediators can cause endothelial dysfunction which is triggered by **a** a disbalance of vasodilative and vasoconstrictive molecules towards an excess of vasoconstrictors, and **b** damage/dysregulation of *EC* signaling. Additionally, vascular progenitor cells *(VPCs)* are attracted to the damaged endothelium. Such *VPCs* can either contribute to repair by differentiation into *ECs* or to remodeling by differentiation into *SMCs*. Furthermore, an endothelial-to-mesenchymal transition *(EMT)* may occur, resulting in an SMC phenotype. Vasoconstriction and altered endothelial cell signaling are stimuli for smooth muscle cells *(SMCs)* to proliferate, resulting in vascular remodeling, increased pulmonary artery pressure and, finally, pulmonary hypertension

emphysema and PH, as well as concomitant downregulation of eNOS in lungs and vessels, supporting the observation in humans [[20](#page-19-19)]. Of interest, eNOS-deficient mice developed emphysema and PH upon exposure to cigarette smoke, whereas inducible NO synthase (iNOS)-deficient mice did not [[20\]](#page-19-19).

15.5 Inflammatory Cells

Systemic inflammation is a known phenomenon in COPD [[125,](#page-25-2) [126](#page-25-3)]. Thus, inflammation could also contribute to the pathogenesis of PH in patients with COPD. In fact, the degree of pulmonary vascular remodeling correlates with the amount of inflammatory cells seen in small airways [[14](#page-19-12), [51\]](#page-21-5). Compared with nonsmokers, COPD patients show increased numbers of inflammatory cells invading the adventitia of pulmonary muscular arteries [[127\]](#page-25-4). These cells predominantly consist of activated T lymphocytes, especially CD8+T cells [[115](#page-24-10), [127](#page-25-4)], which are increased in the arterial adventitia of smokers with normal lung function. The ratio of CD4+/CD8+ is reduced compared with nonsmokers, and is comparable to the situation in patients with mild-to-moderate COPD [[127\]](#page-25-4). The correlation of IL-6 (an inflammatory mediator) expression and elevations in mPAP supports a possible role of inflammation in the pathogenesis of PH (in COPD) [[128](#page-25-5)].

15.6 Oxidative and Nitrosative Stress and the Influence in Vascular (Patho-)Physiology

Evidences for oxidative stress in COPD patients are numerous [[7](#page-19-6), [129](#page-25-6)]. Oxidative stress is the result of an imbalance between oxidants and antioxidants in favor of oxidants. Cigarette smoke itself contains high concentrations of ROS [[130](#page-25-7)]. Elevated concentrations of $H₂O₂$ and 8-isoprostane (oxidative stress markers) can be found in exhaled breath condensate of smokers and ex-smokers, as well as during exacerbations [[131](#page-25-8)–[133](#page-25-9)]. ROS can negatively influence the function of antiproteases, such as α 1-antitrypsin and SLPI. This negative influence leads to a protease/antiprotease imbalance accelerating the degradation of elastin in the lung parenchyma, resulting in emphysema [[134](#page-25-10)]. In addition, ROS play an important role in the vasculature. Traditionally, macrophages have been considered as the major ROS source, even in vessel walls [[135](#page-25-11)]. However, several studies showed that all vascular cells (endothelial, smooth muscle, adventitial cells) produce ROS in different amounts, depending on the stimuli to modulate cellular function [[136](#page-25-12)].

ROS, for example produced by macrophages, are normally involved in the elimination of pathogens and as such are critical to the organism. ROS and reactive nitrogen species (RNS) are able to cause protein modification and DNA damage [[137](#page-25-13)]. Indeed, NO, generated by NO synthases, reacts with superoxide $(O_2 \rightarrow)$ to form the potent oxidant peroxynitrite (ONOO) [[138\]](#page-25-14). This peroxynitrite can react with tyrosine residues from proteins to form nitrotyrosine, shown to be increased in COPD patients [[47](#page-21-3), [48](#page-21-19)]. In the vasculature, ROS play an important physiological role, participating as second messengers in endothelial function, in smooth muscle and EC growth and survival, and in the process of remodeling of the vessel wall [[139](#page-25-15)]. Under pathophysiological conditions, these responses are uncontrolled and imbalanced, respectively [[140](#page-25-16)–[142](#page-25-17)]. The main vascular ROS is the superoxide anion, which can inactivate NO, as mentioned above, by generating peroxynitrite. Since NO is an important vasodilator, the reaction with ROS influences the vascular tone [[143](#page-26-0), [144](#page-26-1)]. Superoxide can be converted to H_2O_2 by superoxide dismutase (SOD), which is a more stable ROS. The depletion of H_2O_2 is carried out by catalase and glutathione peroxidase producing H_2O . Hydrogen peroxide and other peroxides regulate growth-related signaling in vascular SMCs and inflammatory responses in vascular lesions [141, [145](#page-26-2)]. ROS can activate several vascular signaling cascades,

such as ERKs and MAPKs, which are important in cell growth, proliferation, apoptosis, and differentiation. Furthermore, receptor and nonreceptor tyrosine kinases (shown to be involved in cardiovascular remodeling and damage), and protein tyrosine phosphatases and transcription factors such as NF-κB and AP-1, which induce vascular inflammation [[146](#page-26-3), [147](#page-26-4)], are affected by ROS. Several studies reported the role of ROS in growth processes contributing to vascular remodeling and injury. Angiotensin II can lead to NADPH oxidase-generated superoxide production, mediated by the angiotensin type I (ATI) receptor, which is converted to H_2O_2 by SOD acting as a second messenger that results in hypertrophy or hyperplasia of vascular SMCs [[148](#page-26-5), [149](#page-26-6)]. This angiotensin II-induced reaction can be inhibited by the flavoprotein inhibitor diphenyleneiodonium (DPI) [[148](#page-26-5)], catalase [[149](#page-26-6)], and knockdown of p22^{phox} [[150\]](#page-26-7), supporting the involvement of NADPH oxidases in the vasculature.

ROS are also able to promote vascular remodeling by increasing deposition of extracellular matrix proteins. Collagen and elastic fibers can be degraded by proteinases, especially matrix metalloproteinases (MMPs). These MMPs are secreted by macrophages and vascular SMCs in an inactive form [[151](#page-26-8)]. ROS, for example peroxynitrite, activate MMP2 and 9 in human SMCs, leading to degradation of the basement membrane and elastin [\[152\]](#page-26-9). Use of a hypertension model of aldosterone-induced systemic oxidative stress revealed that ET-1-associated processes lead to vascular remodeling [\[153](#page-26-10)]. Redox-sensitive inflammatory processes can also contribute to vascular remodeling. Expression of the inflammation-related intracellular adhesion molecule-1 (ICAM-1) is elevated in the aorta from aldosterone-treated rats [\[153\]](#page-26-10). Furthermore, Liu et al. [[154](#page-26-11)] reported that angiotensin II-induced oxidative stress caused tissue hypertrophy which was associated with increased ICAM-1 expression. Of interest, NADPH oxidases were involved in this context, producing ROS [[154\]](#page-26-11). It is of note that not only the vascular but also the phagocytic NADPH oxidase is involved in superoxide production in cardiovascular diseases because monocytes and lymphocytes can infiltrate cardiovascu-lar tissues [[135](#page-25-11)].

Taken together, elevated occurrence of superoxide, H_2O_2 , and reduced NO bioavailability by reaction with superoxide forming peroxynitrite can contribute fundamentally to vascular remodeling and emphysema development.

15.7 Recent Advances in Molecular Mechanisms of COPD Associated with Vascular Remodeling/PH

15.7.1 Inducible Nitric Oxide Synthase as a Key Player for the Development of Cigarette Smoke-Induced PH and Emphysema

A recent study identified the major NO source in the context of smoke-induced emphysema and PH in mice [[20](#page-19-19)]. Cigarette smoke led to an upregulation of the iNOS mRNA and protein expression, predominantly in small pulmonary vessels, and was associated with increased NO generation. Interestingly, eNOS was downregulated when the disease was established, in mice as well as in COPD patients. It is most probable that the vasodilative NO effect did not appear because of the simultaneous abundance of ROS from both external (cigarette smoke) and internal sources. This study suggested that the subsequent formation of peroxynitrite, most probably increased in this context, had proapoptotic effects (as well as antiproliferative effects) on alveolar epithelial cells type II (AECII) and ECs, promoting emphysema development, vessel loss, and vascular remodeling. Moreover, the level of nitrotyrosine was increased in those mice as well as in smokers without COPD and patients with severe COPD. It is likely that the downregulation of eNOS was associated with uncoupling of this enzyme, leading to the switch of NO to superoxide production, which increased the oxidative stress. Mice deficient in iNOS but not eNOS were protected from vascular remodeling, PH, and emphysema. Treatment with an iNOS inhibitor (L-NIL) could not only prevent disease development but also promote lung regeneration in mice previously exposed to cigarette smoke for 8 months.

Moreover, it has been shown that the development of PH was mediated by iNOScarrying bone-marrow-derived cells, whereas emphysema development was dependent on iNOS in nonbone-marrow-derived cells [[20](#page-19-19)] (Fig. [15.4](#page-14-0)). This phenomenon shows that PH and emphysema can develop independently of each other, and could also explain the discrepancy that not all COPD patients also suffer from PH, if such results are transferable to the human situation.

Quite recently, it could be demonstrated in mice and guinea pigs that stimulation of the soluble guanylate cyclase (sGC) could not only prevent the cigarette smoke-induced development of vascular remodeling but also emphysema [[155](#page-26-12)]. The sGC is an enzyme that uses iNOS-generated NO to produce cGMP from guanosine triphosphate (GTP). cGMP acts as a second messenger, amongst others mediating vasodilation, which can decrease the vascular pressure. The combination of the iNOS-related findings [[20](#page-19-19)] with this study demonstrates the important role of the NO–sGC–cGMP axis for the physiology and pathophysiology of the pulmonary vasculature. It has already been suggested that dysregulation of this axis contributes to pulmonary diseases and PH [[156–](#page-26-13)[158\]](#page-26-14). In line with previous findings [[20](#page-19-19)], prevention from vascular remodeling was associated with prevention from emphysema, although causality was not investigated in these studies.

15.7.2 Reactive Oxygen Species as a Trigger for Vascular Remodeling

Although cigarette smoke itself produces ROS, causing oxidative stress, the additional internal ROS sources produced by the organism are still not fully resolved. Depending on the circumstances, several sources such as NADPH oxidases, mitochondria, xanthine oxidase, cyclooxygenases, lipooxygenases, and uncoupled eNOS come into consideration. As mentioned before, uncoupled eNOS can contribute to oxidative stress in terms of COPD [\[159](#page-26-15)–[163](#page-27-0)]. Based on experimental and clinical

Fig. 15.4 Identification of inducible nitric oxide synthase *(iNOS)* as an essential factor for the development of cigarette smoke-induced emphysema and pulmonary hypertension *(PH)* in mice. Cigarette smoke-mediated upregulation of *iNOS* leads to excessive *NO* production. The formation of peroxynitrite, resulting from the reaction of *NO* with superoxide, was suggested to mediate emphysema and PH development. Superoxide can derive from cigarette smoke itself and/or from uncoupled endothelial *NOS (eNOS)*, nicotinamide adenine dinucleotide phosphate *(NADPH)* oxidases, xanthine oxidases, cyclo- and lipooxidases, and mitochondria. Of interest, *iNOS* generated by nonbone-marrow-derived cells *(NBMDC)*, possibly vascular cells, leads to lung destruction, resulting in emphysema, whereas elevated *iNOS* expression in bone-marrow-derived cells *(BMDC)* causes vascular remodeling. Treatment with the specific *iNOS* inhibitor L-NIL prevents or even reverses pathological alterations

studies, NADPH oxidases are suggested to be the predominant superoxide-producing enzymes in the context of oxidative stress in cardiovascular diseases [[164\]](#page-27-1). The classical NADPH oxidase is expressed in phagocytes; it produces O_2 - as a defense mechanism against bacterial infections [[135\]](#page-25-11). The intrinsic enzyme consists

of membrane-bound catalytic subunits that produce the ROS and cytosolic subunits which regulate the function of the enzyme. In phagocytes $gp91^{phox} (=NOX2; 'phox')$ $=$ phagocytic oxidase) is associated with p22^{phox}, being essential for its function [\[165](#page-27-2)], and both are located in the membrane. They form a complex with the cyto-solic subunits p40^{phox}, p47^{phox}, and p67^{phox}, as well as the G-protein Rac2 [[166](#page-27-3)]. All these subunits, as well as the other membrane-bound enzymes NOX1, NOX4, and NOX5, and homologs to p47^{phox} (NOXO1) and p67^{phox} (NOXA1), are expressed in vascular cells. NOX5 seems to be active without any regulatory subunit. NOX4 is constitutively active, is associated with $p22^{pbox}$, and its function appears to be independent of regulatory subunits, in contrast to NOX1 and NOX2, which need to be activated [[167](#page-27-4)]. The activity of NOX1 is predominantly regulated by NOXO1 and NOXA1 [[168](#page-27-5)]. Activation by $p47^{phox}$ and $p67^{phox}$ is also possible but is less effective than involvement of NOXO1 and NOXA1 [[168](#page-27-5), [169](#page-27-6)]. In the case of NOX2, p47phox is phosphorylated upon cell stimulation, followed by complex formation with the other cytosolic subunits, mediating migration to the membrane-bound subunit. Electrons are then transferred from the substrate NADPH to O_2 , resulting in superoxide $(O_2^{\bullet -})$ production [[170\]](#page-27-7).

 In addition to inflammatory processes often related to immune defense or cigarette smoking, NADPH oxidases can be stimulated by many factors in cardiopulmonary vascular diseases. Vasoactive agonists such as angiotensin II, ET-1, and tumor necrosis factor-α (TNFα) can regulate NADPH oxidases in vascular cells [\[153](#page-26-10), [154](#page-26-11), [171](#page-27-8)]. Even genetic factors might be involved in NADPH oxidase-dependent superoxide production. For instance, polymorphisms in the promoter region of the gene encoding p22^{phox} have been identified. Such polymorphisms can increase the activity of the promoter in vascular SMCs [[172](#page-27-9)], which were shown to be associated with essential hypertension [[173](#page-27-10)] and decreased NO bioavailability [[174\]](#page-27-11).

In terms of remodeling, ROS can influence several intracellular signaling cascades, e.g. activation of ERKs and MAPKs, which affect cell growth and differentiation; protein tyrosine phosphatases and transcription factors, for instance NFκB and AP-1, inducing proinflammatory genes associated with hypertension and atherosclerosis; and receptor and nonreceptor tyrosine kinases, which have been shown to be involved in cardiovascular remodeling and vascular damage [[146\]](#page-26-3).

Because of the fact that monocytes and lymphocytes are able to infiltrate cardiovascular tissues and pulmonary vessels, it is important to note that in cardiovascular diseases, such as COPD, not only vascular NADPH oxidase can contribute to ROS formation, and hence to vascular remodeling, but phagocytic oxidase can also contribute [[175](#page-27-12)].

15.7.3 Neprilysin Downregulation Provokes Pulmonary Vascular Remodeling

A recent study demonstrated that neprilysin (neutral endopeptidase [NEP]) might be an important factor for the susceptibility of humans to pulmonary vascular remodeling in response to smoke inhalation and hypoxia [[176](#page-27-13)] (Fig. [15.5](#page-16-0)). NEP is a trans-

Fig. 15.5 Scheme of proposed neprolysin *(NEP)*-dependent mechanisms leading to vascular remodeling. Extracellular stimuli, such as hypoxia and cigarette smoke, activate pathways in vascular cells, causing downregulation of *NEP* expression and/or activity which was seen in patients with COPD associated with pulmonary hypertension *(PH)*. Stimuli can have direct negative effects on *NEP*, and also indirectly by *1)* increasing reactive oxygen species *(ROS)* and platelet-derived growth factor *(PDGF)*; and *2)* activation of other pathways, in part receptor-mediated. *NEP* downregulation leads to elevated proliferation, migration, inflammation, angiogenesis, and vasoconstriction mediated, amongst others, by depicted molecules, causing vascular remodeling. The *ROS* effect on *NEP* can be inhibited by the superoxide dismutase *(SOD)* mimetic tiron. *Colored boxes* indicate involvement in the respective pathway; *red arrows*representinhibition/downregulation; *green arrows*represent activation/upregulation. *ET-1* endothelin-1, *FGF-2* fibroblast growth factor, *AT-II* angiotension-II, *PASMC*pulmonary artery smooth muscle cells, *EC*endothelial cells, *FB*fibroblasts

membrane zinc peptidase that is widely expressed, including in PASMCs, PAECs, and fibroblasts [[177](#page-27-14)]. Its activity and expression is decreased by cigarette smoke [[178](#page-27-15)], hypoxia [[179](#page-27-16), [180](#page-27-17)], and ROS [[181](#page-28-0)]; it is particularly important in vascular SMCs in terms of remodeling [[176](#page-27-13)]. A genetic depletion of NEP in mice resulted in increased severity of PH associated with stronger proliferation of PASMCs compared with wild-type mice. This finding suggests a protective role of NEP against PH, partly by suppressing proliferation and migration of PASMCs [[180](#page-27-17)].

 Interestingly, Wick et al. [[176](#page-27-13)] showed that NEP expression was decreased in lungs from COPD patients with PH and non-COPD PH patients, especially in the

distal vasculature where prominent remodeling occurs. NEP is involved in many peptidase-dependent (e.g. degradation of vasoactive neuropeptides) and -independent (e.g. interaction of signaling molecules with NEP's intracellular cytosolic subunit) signaling pathways [[182](#page-28-1), [183](#page-28-2)], but its endopeptidase activity-dependent pathways influencing vascular remodeling are not yet well understood. Wick et al. postulated that increased proliferation/migration of dedifferentiated SMCs or myofibroblasts may promote pulmonary vascular remodeling and PH if NEP is less active or downregulated, perhaps mediated by platelet-derived growth factor (PDGF) whose expression is inversely correlated to NEP. Recent data from Karoor et al. support this assumption. They showed that PDGF receptor (PDGFR) signaling was constitutively active in NEP−/− cells and lungs; this effect could be attenuated by the endothelin A (ETA) receptor antagonist ambrisentan [[184](#page-28-3)]. Additionally, a loss of NEP could have inflammatory, angiogenic, and vasoconstrictive effects on vascular cells. The anti-inflammatory action of NEP could be explained by NEP-dependent degradation of the proinflammatory substrates substance P and bradikinin. The decrease in NEP after cigarette smoke and hypoxia may also enhance the angiogenic effect of fibroblast growth factor (FGF)-2 [[182](#page-28-1)], as well as the proproliferative and vasoconstrictive properties of ET-1 [\[185](#page-28-4)] and bombesin-like peptides [[186](#page-28-5)], all being substrates of NEP.

Recently, it was demonstrated in PASMCs that FGF-2 and ET-1 synergize with PDGF in increasing phosphorylation of Src kinase (amongst others activating PDG-FR) and PDGFR, which promoted migration and proliferation of the cells [[184\]](#page-28-3). The phosphatase PTEN (phosphatase and tensin homolog) also plays an important role in vascular biology. The loss of PTEN results in PH [[187](#page-28-6)]. It is inactivated by phosphorylation (mediated by Src and PDGFR) and downregulated in NEPdeficient PASMCs. This downregulation could be rescued by NEP overexpression in NEP null cells or by downregulation of Src or PDGFR by small interfering RNA (siRNA). These observations suggest that NEP-dependent mechanisms protect against the inactivation of PTEN [[184](#page-28-3)]. Moreover, NEP can be inactivated by ROS, as shown by decreased activity in the presence of H_2O_2 and improved NEP activity when an antioxidant, the SOD mimetic tiron, was present [[176](#page-27-13)].

Of interest, early studies suggest that inhibition of NEP could be beneficial for the treatment of PH [[188](#page-28-7), [189](#page-28-8)]. This proposal is based on the fact that NEP can inactivate atrial/brain natriuretic peptides (ANP/BNP), which promote vasodilation by increasing cGMP mediated via natriuretic peptide receptor-A (NPR-A) [[190\]](#page-28-9). cGMP-dependent protein kinase (PKG), cGMP binding PDEs, and cyclic nucleotide-gated ion channels bind cGMP, in which PKG seems to be the main mediator of cGMP signals [[191](#page-28-10)–[193](#page-28-11)]. Binding of ANP/BNP-induced cGMP activates PKG, leading to the catalytic transfer of phosphate from ATP to target proteins. These phosphorylated proteins translate the extracellular stimuli into specific biological outputs [[194](#page-28-12)], such as vasodilation.

It has been shown that NEP antagonists, alone and in combination with angiotensin-converting enzyme (ACE) and endothelin-converting enzyme (ECE) inhibitors, were able to improve cardiac function, decrease systemic blood pressure, and limit cardiac hypertrophy [\[195](#page-28-13)–[198](#page-28-14)]. Nevertheless, side effects were observed if single NEP inhibitors or dual inhibitors (NEP/ACE or NEP/ECE) were used. Nowadays, even triple vasopeptidase inhibitors (NEP/ACE/ECE) are under investigation, with promising preliminary results showing fewer side effects; in particular, the increase in ET-1 can be reversed by simultaneous application of an ECE inhibitor [[196\]](#page-28-15). These combination therapies and the different mechanisms of NEP, ACE, and ECE inhibition have been extensively reviewed by Daull et al. [[196\]](#page-28-15).

However, recent observations support the possibility that NEP could be protective against PH [[176](#page-27-13), [180](#page-27-17), [199](#page-28-16)]. One explanation for the discrepancy between the beneficial cardiac effects and harmful pulmonary effects of NEP inhibition is the well-known phenomenon that the pulmonary and systemic circulations usually respond to hypoxia (a major stimulus for PH) in opposite ways: pulmonary vessels contract to redirect blood flow to better oxygenated areas of the lung, whereas systemic vessels dilate to increase the flow of oxygenated blood to areas of tissue hypoxia or ischemia [[180\]](#page-27-17).

In conclusion, in terms of the lung, it is suggested to increase NEP to treat PH, whereas cardiac NEP inhibition could be used for treatment of hypertrophy and improvement of cardiac function. This dichotomy clearly demonstrates that a possible treatment must be dependent not only on the target itself but also on the localization of the target. These data should be kept in mind when considering NEP-related drug therapies.

15.8 Conclusions

The mechanisms of COPD and PH in COPD are still far from being fully understood. Studies primarily from the last decade have shown that vascular remodeling and PH can occur in COPD, not only in severe cases but also in mild-to-moderate forms of the disease, or even in smokers without airflow limitations. Investigations of the molecular mechanisms of COPD and the occurrence of pulmonary vascular remodeling established that COPD associated with PH and pulmonary vascular remodeling is a complicated multifactorial disease involving hypoxia-related and hypoxia-unrelated mechanisms, inflammation, and endothelial dysfunction. Recent advances have changed the long-standing view of the pathobiology of COPD. In the past, vascular alterations (vascular remodeling and PH) have been seen to be secondary events occurring after destruction of the parenchyma, predominantly caused by hypoxia/hypoxemia. However, recent studies have demonstrated that such vascular abnormalities can be early events in COPD, preceding airflow limitations and emphysema, and can be independent of hypoxia. It has been shown, at least in mice, that PH and lung emphysema triggered by tobacco smoke can occur independently, and suggest that vascular molecular alterations can be a trigger for lung emphysema development.

Further elucidation of the contribution of pulmonary vascular changes to COPD development may help to identify new therapeutic concepts for this disease.

References

- 1. Agusti AG, Noguera A, Sauleda J, Sala E, Pons J, Busquets X. Systemic effects of chronic obstructive pulmonary disease. Eur Respir J. 2003;21(2):347–60.
- 2. Sertogullarindan B, Gumrukcuoglu HA, Sezgi C, Akil MA. Frequency of pulmonary hypertension in patients with COPD due to biomass smoke and tobacco smoke. Int J Med Sci. 2012;9(6):406–12.
- 3. Black PN, Ching PS, Beaumont B, Ranasinghe S, Taylor G, Merrilees MJ. Changes in elastic fibres in the small airways and alveoli in COPD. Eur Respir J. 2008;31(5):998–1004.
- 4. Barnes PJ. Chronic obstructive pulmonary disease. N Engl J Med. 2000;343(4):269–80.
- 5. Sharafkhaneh A, Hanania NA, Kim V. Pathogenesis of emphysema: from the bench to the bedside. Proc Am Thorac Soc. 2008;5(4):475–7.
- 6. Churg A, Cosio M, Wright JL. Mechanisms of cigarette smoke-induced COPD: insights from animal models. Am J Physiol Lung Cell Mol Physiol. 2008;294(4):L612–31.
- 7. Repine JE, Bast A, Lankhorst I. Oxidative stress in chronic obstructive pulmonary disease. Oxidative Stress Study Group. Am J Respir Crit Care Med. 1997;156(2 Pt 1):341–57.
- 8. Barbera JA, Peinado VI, Santos S. Pulmonary hypertension in chronic obstructive pulmonary disease. Eur Respir J. 2003;21(5):892–905.
- 9. Peinado VI, Pizarro S, Barbera JA. Pulmonary vascular involvement in COPD. Chest. 2008;134(4):808–14.
- 10. Wright JL, Levy RD, Churg A. Pulmonary hypertension in chronic obstructive pulmonary disease: current theories of pathogenesis and their implications for treatment. Thorax. 2005;60(7):605–9.
- 11. Naeije R. Pulmonary hypertension and right heart failure in chronic obstructive pulmonary disease. Proc Am Thorac Soc. 2005;2(1):20–2.
- 12. Scharf SM, Iqbal M, Keller C, Criner G, Lee S, Fessler HE. Hemodynamic characterization of patients with severe emphysema. Am J Respir Crit Care Med. 2002;166(3):314–22.
- 13. Cuttica MJ, Kalhan R, Shlobin OA, Ahmad S, Gladwin M, Machado RF, Barnett SD, Nathan SD. Categorization and impact of pulmonary hypertension in patients with advanced COPD. Respir Med. 2010;104(12):1877–82.
- 14. Oswald-Mammosser M, Weitzenblum E, Quoix E, Moser G, Chaouat A, Charpentier C, Kessler R. Prognostic factors in COPD patients receiving long-term oxygen therapy. Importance of pulmonary artery pressure. Chest. 1995;107(5):1193–8.
- 15. Minai OA, Chaouat A, Adnot S. Pulmonary hypertension in COPD: epidemiology, significance, and management: pulmonary vascular disease: the global perspective. Chest. 2010;137(6 Suppl):39S-51S.
- 16. Blanco I, Gimeno E, Munoz PA, Pizarro S, Gistau C, Rodriguez-Roisin R, Roca J, Barbera JA. Hemodynamic and gas exchange effects of sildenafil in patients with chronic obstructive pulmonary disease and pulmonary hypertension. Am J Respir Crit Care Med. 2010;181(3):270–8.
- 17. Pynnaert C, Lamotte M, Naeije R. Aerobic exercise capacity in COPD patients with and without pulmonary hypertension. Respir Med. 2010;104(1):121–6.
- 18. Orr R, Smith LJ, Cuttica MJ. Pulmonary hypertension in advanced chronic obstructive pulmonary disease. Curr Opin Pulm Med. 2012;18(2):138–43.
- 19. Ferrer E, Peinado VI, Diez M, Carrasco JL, Musri MM, Martinez A, Rodriguez-Roisin R, Barbera JA. Effects of cigarette smoke on endothelial function of pulmonary arteries in the guinea pig. Respir Res. 2009;10:76.
- 20. Seimetz M, Parajuli N, Pichl A, Veit F, Kwapiszewska G, Weisel FC, Milger K, Egemnazarov B, Turowska A, Fuchs B, et al. Inducible NOS inhibition reverses tobacco-smoke-induced emphysema and pulmonary hypertension in mice. Cell. 2011;147(2):293–305.
- 21. Wright JL, Churg A. Cigarette smoke causes physiologic and morphologic changes of emphysema in the guinea pig. Am Rev Respir Dis. 1990;142(6 Pt 1):1422–8.
- 15 The Pulmonary Vasculature in Chronic Obstructive Lung Disease 371
- 22. Wright JL, Churg A. Effect of long-term cigarette smoke exposure on pulmonary vascular structure and function in the guinea pig. Exp Lung Res. 1991;17(6):997–1009.
- 23. Santos S, Peinado VI, Ramirez J, Melgosa T, Roca J, Rodriguez-Roisin R, Barbera JA. Characterization of pulmonary vascular remodelling in smokers and patients with mild COPD. Eur Respir J. 2002;19(4):632–8.
- 24. Schrijen F, Uffholtz H, Polu JM, Poincelot F. Pulmonary and systemic hemodynamic evolution in chronic bronchitis. Am Rev Respir Dis. 1978;117(1):25–31.
- 25. Weitzenblum E, Loiseau A, Hirth C, Mirhom R, Rasaholinjanahary J. Course of pulmonary hemodynamics in patients with chronic obstructive pulmonary disease. Chest. 1979;75(6):656–62.
- 26. Weitzenblum E, Sautegeau A, Ehrhart M, Mammosser M, Hirth C, Roegel E. Long-term course of pulmonary arterial pressure in chronic obstructive pulmonary disease. Am Rev Respir Dis. 1984;130(6):993–8.
- 27. Kessler R, Faller M, Weitzenblum E, Chaouat A, Aykut A, Ducolone A, Ehrhart M, Oswald-Mammosser M. "Natural history" of pulmonary hypertension in a series of 131 patients with chronic obstructive lung disease. Am J Respir Crit Care Med. 2001;164(2):219–24.
- 28. MacNee W. Pathophysiology of cor pulmonale in chronic obstructive pulmonary disease. Part One. Am J Respir Crit Care Med. 1994;150(3):833–52.
- 29. Richens JM, Howard P. Oedema in cor pulmonale. Clin Sci (Lond). 1982;62(3):255–9.
- 30. Farber MO, Weinberger MH, Robertson GL, Fineberg NS, Manfredi F. Hormonal abnormalities affecting sodium and water balance in acute respiratory failure due to chronic obstructive lung disease. Chest. 1984;85(1):49–54.
- 31. MacNee W, Wathen CG, Flenley DC, Muir AD. The effects of controlled oxygen therapy on ventricular function in patients with stable and decompensated cor pulmonale. Am Rev Respir Dis. 1988;137(6):1289–95.
- 32. Weitzenblum E, Apprill M, Oswald M, Chaouat A, Imbs JL. Pulmonary hemodynamics in patients with chronic obstructive pulmonary disease before and during an episode of peripheral edema. Chest. 1994;105(5):1377–82.
- 33. Bishop JM, Cross KW. Physiological variables and mortality in patients with various categories of chronic respiratory disease. Bull Eur Physiopathol Respir. 1984;20(6):495–500.
- 34. Chaouat A, Bugnet AS, Kadaoui N, Schott R, Enache I, Ducolone A, Ehrhart M, Kessler R, Weitzenblum E. Severe pulmonary hypertension and chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2005;172(2):189–94.
- 35. Barbera JA, Riverola A, Roca J, Ramirez J, Wagner PD, Ros D, Wiggs BR, Rodriguez-Roisin R. Pulmonary vascular abnormalities and ventilation-perfusion relationships in mild chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1994;149(2 Pt 1):423–9.
- 36. Magee F, Wright JL, Wiggs BR, Pare PD, Hogg JC. Pulmonary vascular structure and function in chronic obstructive pulmonary disease. Thorax. 1988;43(3):183–9.
- 37. Peinado VI, Barbera JA, Ramirez J, Gomez FP, Roca J, Jover L, Gimferrer JM, Rodriguez-Roisin R. Endothelial dysfunction in pulmonary arteries of patients with mild COPD. Am J Physiol. 1998;274(6 Pt 1):L908–13.
- 38. Eddahibi S, Chaouat A, Morrell N, Fadel E, Fuhrman C, Bugnet AS, Dartevelle P, Housset B, Hamon M, Weitzenblum E, et al. Polymorphism of the serotonin transporter gene and pulmonary hypertension in chronic obstructive pulmonary disease. Circulation. 2003;108(15):1839–44.
- 39. Owens GK. Regulation of differentiation of vascular smooth muscle cells. Physiol Rev. 1995;75(3):487–517.
- 40. van der Loop FT, Gabbiani G, Kohnen G, Ramaekers FC, van Eys GJ. Differentiation of smooth muscle cells in human blood vessels as defined by smoothelin, a novel marker for the contractile phenotype. Arterioscler Thromb Vasc Biol. 1997;17(4):665–71.
- 41. Rabinovitch M. Elastase and the pathobiology of unexplained pulmonary hypertension. Chest. 1998;114(3 Suppl):213S-24S.
- 42. Barbera JA. Mechanisms of development of chronic obstructive pulmonary disease-associated pulmonary hypertension. Pulm Circ. 2013;3(1):160–4.
- 43. Naeije R, Barbera JA. Pulmonary hypertension associated with COPD. Crit Care. 2001;5(6):286–9.
- 44. Wilkinson M, Langhorne CA, Heath D, Barer GR, Howard P. A pathophysiological study of 10 cases of hypoxic cor pulmonale. Q J Med. 1988;66(249):65–85.
- 45. Lejeune P, Mols P, Naeije R, Hallemans R, Melot C. Acute hemodynamic effects of controlled oxygen therapy in decompensated chronic obstructive pulmonary disease. Crit Care Med. 1984;12(12):1032–5.
- 46. Timms RM, Khaja FU, Williams GW. Hemodynamic response to oxygen therapy in chronic obstructive pulmonary disease. Ann Intern Med. 1985;102(1):29–36.
- 47. Ricciardolo FL, Sterk PJ, Gaston B, Folkerts G. Nitric oxide in health and disease of the respiratory system. Physiol Rev. 2004;84(3):731–65.
- 48. Tsoumakidou M, Tzanakis N, Chrysofakis G, Siafakas NM. Nitrosative stress, heme oxygenase-1 expression and airway inflammation during severe exacerbations of COPD. Chest. 2005;127(6):1911–8.
- 49. Voelkel NF, Mizuno S, Bogaard HJ. The role of hypoxia in pulmonary vascular diseases: a perspective. Am J Physiol Lung Cell Mol Physiol. 2013;304(7):L457–65.
- 50. Stockley RA, Mannino D, Barnes PJ. Burden and pathogenesis of chronic obstructive pulmonary disease. Proc Am Thorac Soc. 2009;6(6):524–6.
- 51. Peinado VI, Barbera JA, Abate P, Ramirez J, Roca J, Santos S, Rodriguez-Roisin R. Inflammatory reaction in pulmonary muscular arteries of patients with mild chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1999;159(5 Pt 1):1605–11.
- 52. Weitzenblum E, Sautegeau A, Ehrhart M, Mammosser M, Pelletier A. Long-term oxygen therapy can reverse the progression of pulmonary hypertension in patients with chronic obstructive pulmonary disease. Am Rev Respir Dis. 1985;131(4):493–8.
- 53. Chaouat A, Naeije R, Weitzenblum E. Pulmonary hypertension in COPD. Eur Respir J. 2008;32(5):1371–85.
- 54. Fishman AP. Hypoxia on the pulmonary circulation. How and where it acts. Circ Res. 1976;38(4):221–31.
- 55. Fishman AP. State of the art: chronic cor pulmonale. Am Rev Respir Dis. 1976;114(4):775– 94.
- 56. Weitzenblum E, Chaouat A. Cor pulmonale. Chron Respir Dis. 2009;6(3):177–85.
- 57. Chao J, Viets Z, Donham P, Wood JG, Gonzalez NC. Dexamethasone blocks the systemic inflammation of alveolar hypoxia at several sites in the inflammatory cascade. Am J Physiol Heart Circ Physiol. 2012;303(2):H168–77.
- 58. Chao J, Wood JG, Blanco VG, Gonzalez NC. The systemic inflammation of alveolar hypoxia is initiated by alveolar macrophage-borne mediator(s). Am J Respir Cell Mol Biol. 2009;41(5):573–82.
- 59. Prabhakar NR, Kumar GK, Peng YJ. Sympatho-adrenal activation by chronic intermittent hypoxia. J Appl Physiol. 2012;113(8):1304–10.
- 60. Wang GL, Semenza GL. Characterization of hypoxia-inducible factor 1 and regulation of DNA binding activity by hypoxia. J Biol Chem. 1993;268(29):21513–8.
- 61. Archiniegas E, Neves CY, Candelle D, Cardier JE. Thrombin and its protease-activated receptor-1 (PAR1) participate in the endothelial-mesenchymal transdifferentiation process. DNA Cell Biol. 2004;23(12):815–25.
- 62. Frohlich S, Boylan J, McLoughlin P. Hypoxia-induced inflammation in the lung: a potential therapeutic target in acute lung injury? Am J Respir Cell Mol Biol. 2013;48(3):271–9.
- 63. Nicolls MR, Voelkel NF. Hypoxia and the lung: beyond hypoxic vasoconstriction. Antioxid Redox Signal. 2007;9(6):741–3.
- 64. Rius J, Guma M, Schachtrup C, Akassoglou K, Zinkernagel AS, Nizet V, Johnson RS, Haddad GG, Karin M. NF-kappaB links innate immunity to the hypoxic response through transcriptional regulation of HIF-1alpha. Nature. 2008;453(7196):807–11.
- 65. Taylor CT. Interdependent roles for hypoxia inducible factor and nuclear factor-kappaB in hypoxic inflammation. J Physiol. 2008;586(Pt 17):4055–9.
- 66. Yamaji-Kegan K, Su Q, Angelini DJ, Champion HC, Johns RA. Hypoxia-induced mitogenic factor has proangiogenic and proinflammatory effects in the lung via VEGF and VEGF receptor-2. Am J Physiol Lung Cell Mol Physiol. 2006;291(6):L1159–68.
- 67. Angelini DJ, Su Q, Kolosova IA, Fan C, Skinner JT, Yamaji-Kegan K, Collector M, Sharkis SJ, Johns RA. Hypoxia-induced mitogenic factor (HIMF/FIZZ1/RELM alpha) recruits bone marrow-derived cells to the murine pulmonary vasculature. PLoS One. 2010;5(6):e11251.
- 68. Kuhlicke J, Frick JS, Morote-Garcia JC, Rosenberger P, Eltzschig HK. Hypoxia inducible factor (HIF)-1 coordinates induction of Toll-like receptors TLR2 and TLR6 during hypoxia. PLoS One. 2007;2(12):e1364.
- 69. van Uden P, Kenneth NS, Rocha S. Regulation of hypoxia-inducible factor-1alpha by NFkappaB. Biochem J. 2008;412(3):477–84.
- 70. van Uden P, Kenneth NS, Webster R, Muller HA, Mudie S, Rocha S. Evolutionary conserved regulation of HIF-1beta by NF-kappaB. PLoS Genet. 2011;7(1):e1001285.
- 71. Savale L, Tu L, Rideau D, Izziki M, Maitre B, Adnot S, Eddahibi S. Impact of interleukin-6 on hypoxia-induced pulmonary hypertension and lung inflammation in mice. Respir Res. 2009;10:6.
- 72. Steiner MK, Syrkina OL, Kolliputi N, Mark EJ, Hales CA, Waxman AB. Interleukin-6 overexpression induces pulmonary hypertension. Circ Res. 2009;104(2):236–44, (28p following 244).
- 73. Brock M, Trenkmann M, Gay RE, Michel BA, Gay S, Fischler M, Ulrich S, Speich R, Huber LC. Interleukin-6 modulates the expression of the bone morphogenic protein receptor type II through a novel STAT3-microRNA cluster 17/92 pathway. Circ Res. 2009;104(10):1184–91.
- 74. Grijelmo C, Rodrigue C, Svrcek M, Bruyneel E, Hendrix A, de Wever O, Gespach C. Proinvasive activity of BMP-7 through SMAD4/src-independent and ERK/Rac/JNK-dependent signaling pathways in colon cancer cells. Cell Signal. 2007;19(8):1722–32.
- 75. Tian Q, He XC, Hood L, Li L. Bridging the BMP and Wnt pathways by PI3 kinase/Akt and 14-3-3zeta. Cell Cycle. 2005;4(2):215–6.
- 76. Rudarakanchana N, Flanagan JA, Chen H, Upton PD, Machado R, Patel D, Trembath RC, Morrell NW. Functional analysis of bone morphogenetic protein type II receptor mutations underlying primary pulmonary hypertension. Hum Mol Genet. 2002;11(13):1517–25.
- 77. Nohe A, Hassel S, Ehrlich M, Neubauer F, Sebald W, Henis YI, Knaus P. The mode of bone morphogenetic protein (BMP) receptor oligomerization determines different BMP-2 signaling pathways. J Biol Chem. 2002;277(7):5330–8.
- 78. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. J Clin Invest. 2012;122(12):4306–13.
- 79. Atkinson C, Stewart S, Upton PD, Machado R, Thomson JR, Trembath RC, Morrell NW. Primary pulmonary hypertension is associated with reduced pulmonary vascular expression of type II bone morphogenetic protein receptor. Circulation. 2002;105(14):1672–8.
- 80. Shimoda LA, Fallon M, Pisarcik S, Wang J, Semenza GL. HIF-1 regulates hypoxic induction of NHE1 expression and alkalinization of intracellular pH in pulmonary arterial myocytes. Am J Physiol Lung Cell Mol Physiol. 2006;291(5):L941–9.
- 81. Shimoda LA, Manalo DJ, Sham JS, Semenza GL, Sylvester JT. Partial HIF-1alpha deficiency impairs pulmonary arterial myocyte electrophysiological responses to hypoxia. Am J Physiol Lung Cell Mol Physiol. 2001;281(1):L202–8.
- 82. Wang J, Weigand L, Lu W, Sylvester JT, Semenza GL, Shimoda LA. Hypoxia inducible factor 1 mediates hypoxia-induced TRPC expression and elevated intracellular Ca^{2+} in pulmonary arterial smooth muscle cells. Circ Res. 2006;98(12):1528–37.
- 83. Whitman EM, Pisarcik S, Luke T, Fallon M, Wang J, Sylvester JT, Semenza GL, Shimoda LA. Endothelin-1 mediates hypoxia-induced inhibition of voltage-gated K^+ channel expression in pulmonary arterial myocytes. Am J Physiol Lung Cell Mol Physiol. 2008;294(2):L309–18.
- 84. Moudgil R, Michelakis ED, Archer SL. The role of k^+ channels in determining pulmonary vascular tone, oxygen sensing, cell proliferation, and apoptosis: implications in hypoxic pulmonary vasoconstriction and pulmonary arterial hypertension. Microcirculation. 2006;13(8):615–32.
- 85. Remillard CV, Yuan JX. High altitude pulmonary hypertension: role of K^+ and Ca^{2+} channels. High Alt Med Biol. 2005;6(2):133–46.
- 86. Smirnov SV, Robertson TP, Ward JP, Aaronson PI. Chronic hypoxia is associated with reduced delayed rectifier K+ current in rat pulmonary artery muscle cells. Am J Physiol. 1994;266(1 Pt 2):H365–70.
- 87. Grimminger F, Weissmann N, Spriestersbach R, Becker E, Rosseau S, Seeger W. Effects of NADPH oxidase inhibitors on hypoxic vasoconstriction in buffer-perfused rabbit lungs. Am J Physiol. 1995;268(5 Pt 1):L747–52.
- 88. Michelakis ED, Hampl V, Nsair A, Wu X, Harry G, Haromy A, Gurtu R, Archer SL. Diversity in mitochondrial function explains differences in vascular oxygen sensing. Circ Res. 2002;90(12):1307–15.
- 89. Waypa GB, Schumacker PT. Hypoxic pulmonary vasoconstriction: redox events in oxygen sensing. J Appl Physiol. 2005;98(1):404–14.
- 90. Weissmann N, Tadic A, Hanze J, Rose F, Winterhalder S, Nollen M, Schermuly RT, Ghofrani HA, Seeger W, Grimminger F. Hypoxic vasoconstriction in intact lungs: a role for NADPH oxidase-derived H(2)O(2)? Am J Physiol Lung Cell Mol Physiol. 2000;279(4):L683–90.
- 91. Weissmann N, Zeller S, Schafer RU, Turowski C, Ay M, Quanz K, Ghofrani HA, Schermuly RT, Fink L, Seeger W, et al. Impact of mitochondria and NADPH oxidases on acute and sustained hypoxic pulmonary vasoconstriction. Am J Respir Cell Mol Biol. 2006;34(4):505–13.
- 92. Mittal M, Gu XQ, Pak O, Pamenter ME, Haag D, Fuchs DB, Schermuly RT, Ghofrani HA, Brandes RP, Seeger W, et al. Hypoxia induces Kv channel current inhibition by increased NADPH oxidase-derived reactive oxygen species. Free Radic Biol Med. 2012;52(6):1033– 42.
- 93. Landsberg JW, Yuan JX. Calcium and TRP channels in pulmonary vascular smooth muscle cell proliferation. News Physiol Sci. 2004;19:44–50.
- 94. Reeve HL, Michelakis E, Nelson DP, Weir EK, Archer SL. Alterations in a redox oxygen sensing mechanism in chronic hypoxia. J Appl Physiol. 2001;90(6):2249–56.
- 95. Fuchs B, Sommer N, Dietrich A, Schermuly RT, Ghofrani HA, Grimminger F, Seeger W, Gudermann T, Weissmann N. Redox signaling and reactive oxygen species in hypoxic pulmonary vasoconstriction. Respir Physiol Neurobiol. 2010;174(3):282–91.
- 96. Olschewski A, Li Y, Tang B, Hanze J, Eul B, Bohle RM, Wilhelm J, Morty RE, Brau ME, Weir EK, et al. Impact of TASK-1 in human pulmonary artery smooth muscle cells. Circ Res. 2006;98(8):1072–80.
- 97. Weir EK, Olschewski A. Role of ion channels in acute and chronic responses of the pulmonary vasculature to hypoxia. Cardiovasc Res. 2006;71(4):630–41.
- 98. Tang B, Li Y, Nagaraj C, Morty RE, Gabor S, Stacher E, Voswinckel R, Weissmann N, Leithner K, Olschewski H, et al. Endothelin-1 inhibits background two-pore domain channel TASK-1 in primary human pulmonary artery smooth muscle cells. Am J Respir Cell Mol Biol. 2009;41(4):476–83.
- 99. Herrera GM, Walker BR. Involvement of L-type calcium channels in hypoxic relaxation of vascular smooth muscle. J Vasc Res. 1998;35(4):265–73.
- 100. Shimoda LA, Polak J. Hypoxia. 4. Hypoxia and ion channel function. Am J Physiol Cell Physiol. 2011;300(5):C951–67.
- 101. Kwapiszewska G, Wygrecka M, Marsh LM, Schmitt S, Trosser R, Wilhelm J, Helmus K, Eul B, Zakrzewicz A, Ghofrani HA, et al. Fhl-1, a new key protein in pulmonary hypertension. Circulation. 2008;118(11):1183–94.
- 102. Sims MW, Margolis DJ, Localio AR, Panettieri RA, Kawut SM, Christie JD. Impact of pulmonary artery pressure on exercise function in severe COPD. Chest. 2009;136(2):412–9.
- 103. Thabut G, Dauriat G, Stern JB, Logeart D, Levy A, Marrash-Chahla R, Mal H. Pulmonary hemodynamics in advanced COPD candidates for lung volume reduction surgery or lung transplantation. Chest. 2005;127(5):1531–6.
- 104. Wright JL, Petty T, Thurlbeck WM. Analysis of the structure of the muscular pulmonary arteries in patients with pulmonary hypertension and COPD: National Institutes of Health nocturnal oxygen therapy trial. Lung. 1992;170(2):109–24.
- 105. Ferrer E, Peinado VI, Castaneda J, Prieto-Lloret J, Olea E, Gonzalez-Martin MC, Vega-Agapito MV, Diez M, Dominguez-Fandos D, Obeso A, et al. Effects of cigarette smoke and hypoxia on pulmonary circulation in the guinea pig. Eur Respir J. 2011;38(3):617–27.
- 106. Wright JL, Tai H, Churg A. Cigarette smoke induces persisting increases of vasoactive mediators in pulmonary arteries. Am J Respir Cell Mol Biol. 2004;31(5):501–9.
- 107. Wright JL, Tai H, Dai J, Churg A. Cigarette smoke induces rapid changes in gene expression in pulmonary arteries. Lab Invest. 2002;82(10):1391–8.
- 108. Yamato H, Sun JP, Churg A, Wright JL. Cigarette smoke-induced emphysema in guinea pigs is associated with diffusely decreased capillary density and capillary narrowing. Lab Invest. 1996;75(2):211–9.
- 109. Lee SD, Lee DS, Chun YG, Shim TS, Lim CM, Koh Y, Kim WS, Kim DS, Kim WD. Cigarette smoke extract induces endothelin-1 via protein kinase C in pulmonary artery endothelial cells. Am J Physiol Lung Cell Mol Physiol. 2001;281(2):L403–11.
- 110. Nana-Sinkam SP, Lee JD, Sotto-Santiago S, Stearman RS, Keith RL, Choudhury Q, Cool C, Parr J, Moore MD, Bull TM, et al. Prostacyclin prevents pulmonary endothelial cell apoptosis induced by cigarette smoke. Am J Respir Crit Care Med. 2007;175(7):676–85.
- 111. Peluffo G, Calcerrada P, Piacenza L, Pizzano N, Radi R. Superoxide-mediated inactivation of nitric oxide and peroxynitrite formation by tobacco smoke in vascular endothelium: studies in cultured cells and smokers. Am J Physiol Heart Circ Physiol. 2009;296(6):H1781–92.
- 112. Edirisinghe I, Arunachalam G, Wong C, Yao H, Rahman A, Phipps RP, Jin ZG, Rahman I. Cigarette-smoke-induced oxidative/nitrosative stress impairs VEGF- and fluid-shearstress-mediated signaling in endothelial cells. Antioxid Redox Signal. 2010;12(12):1355– 69.
- 113. Damico R, Simms T, Kim BS, Tekeste Z, Amankwan H, Damarla M, Hassoun PM. p53 mediates cigarette smoke-induced apoptosis of pulmonary endothelial cells: inhibitory effects of macrophage migration inhibitor factor. Am J Respir Cell Mol Biol. 2011;44(3):323–32.
- 114. Milara J, Juan G, Ortiz JL, Guijarro R, Losada M, Serrano A, Morcillo EJ, Cortijo J. Cigarette smoke-induced pulmonary endothelial dysfunction is partially suppressed by sildenafil. Eur J Pharm Sci. 2010;39(5):363–72.
- 115. Dinh-Xuan AT, Higenbottam TW, Clelland CA, Pepke-Zaba J, Cremona G, Butt AY, Large SR, Wells FC, Wallwork J. Impairment of endothelium-dependent pulmonary-artery relaxation in chronic obstructive lung disease. N Engl J Med. 1991;324(22):1539–47.
- 116. Santos S, Peinado VI, Ramirez J, Morales-Blanhir J, Bastos R, Roca J, Rodriguez-Roisin R, Barbera JA. Enhanced expression of vascular endothelial growth factor in pulmonary arteries of smokers and patients with moderate chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2003;167(9):1250–6.
- 117. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature. 1980;288(5789):373–6.
- 118. Stamler JS, Loh E, Roddy MA, Currie KE, Creager MA. Nitric oxide regulates basal systemic and pulmonary vascular resistance in healthy humans. Circulation. 1994;89(5):2035– 40.
- 119. Cremona G, Wood AM, Hall LW, Bower EA, Higenbottam T. Effect of inhibitors of nitric oxide release and action on vascular tone in isolated lungs of pig, sheep, dog and man. J Physiol. 1994;481 (Pt 1):185–95.
- 120. Barbera JA, Roger N, Roca J, Rovira I, Higenbottam TW, Rodriguez-Roisin R. Worsening of pulmonary gas exchange with nitric oxide inhalation in chronic obstructive pulmonary disease. Lancet. 1996;347(8999):436–40.
- 121. Greenberg B, Kishiyama S. Endothelium-dependent and -independent responses to severe hypoxia in rat pulmonary artery. Am J Physiol. 1993;265(5 Pt 2):H1712–20.
- 122. Giaid A, Yanagisawa M, Langleben D, Michel RP, Levy R, Shennib H, Kimura S, Masaki T, Duguid WP, Stewart DJ. Expression of endothelin-1 in the lungs of patients with pulmonary hypertension. N Engl J Med. 1993;328(24):1732–9.
- 123. Barbera JA, Peinado VI, Santos S, Ramirez J, Roca J, Rodriguez-Roisin R. Reduced expression of endothelial nitric oxide synthase in pulmonary arteries of smokers. Am J Respir Crit Care Med. 2001;164(4):709–13.
- 124. Giaid A, Saleh D. Reduced expression of endothelial nitric oxide synthase in the lungs of patients with pulmonary hypertension. N Engl J Med. 1995;333(4):214–21.
- 125. Gan WQ, Man SF, Senthilselvan A, Sin DD. Association between chronic obstructive pulmonary disease and systemic inflammation: a systematic review and a meta-analysis. Thorax. 2004;59(7):574–80.
- 126. Walter RE, Wilk JB, Larson MG, Vasan RS, Keaney JF Jr, Lipinska I, O'Connor GT, Benjamin EJ. Systemic inflammation and COPD: the Framingham Heart Study. Chest. 2008;133(1):19–25.
- 127. Saetta M, Baraldo S, Corbino L, Turato G, Braccioni F, Rea F, Cavallesco G, Tropeano G, Mapp CE, Maestrelli P, et al. CD^{8+} ve cells in the lungs of smokers with chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1999;160(2):711–7.
- 128. Chaouat A, Savale L, Chouaid C, Tu L, Sztrymf B, Canuet M, Maitre B, Housset B, Brandt C, Le Corvoisier P, et al. Role for interleukin-6 in COPD-related pulmonary hypertension. Chest. 2009;136(3):678–87.
- 129. MacNee W. Oxidative stress and lung inflammation in airways disease. Eur J Pharmacol. 2001;429(1–3):195–207.
- 130. Pryor WA, Stone K. Oxidants in cigarette smoke. Radicals, hydrogen peroxide, peroxynitrate, and peroxynitrite. Ann N Y Acad Sci. 1993;686:12–27.
- 131. Dekhuijzen PN, Aben KK, Dekker I, Aarts LP, Wielders PL, van Herwaarden CL, Bast A. Increased exhalation of hydrogen peroxide in patients with stable and unstable chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1996;154(3 Pt 1):813–6.
- 132. Montuschi P, Collins JV, Ciabattoni G, Lazzeri N, Corradi M, Kharitonov SA, Barnes PJ. Exhaled 8-isoprostane as an in vivo biomarker of lung oxidative stress in patients with COPD and healthy smokers. Am J Respir Crit Care Med. 2000;162(3 Pt 1):1175–7.
- 133. Nowak D, Kasielski M, Antczak A, Pietras T, Bialasiewicz P. Increased content of thiobarbituric acid-reactive substances and hydrogen peroxide in the expired breath condensate of patients with stable chronic obstructive pulmonary disease: no significant effect of cigarette smoking. Respir Med. 1999;93(6):389–96.
- 134. Taggart C, Cervantes-Laurean D, Kim G, McElvaney NG, Wehr N, Moss J, Levine RL. Oxidation of either methionine 351 or methionine 358 in alpha 1-antitrypsin causes loss of anti-neutrophil elastase activity. J Biol Chem. 2000;275(35):27258–65.
- 135. Cathcart MK. Regulation of superoxide anion production by NADPH oxidase in monocytes/macrophages: contributions to atherosclerosis. Arterioscler Thromb Vasc Biol. 2004;24(1):23–8.
- 136. Griendling KK, Sorescu D, Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. Circ Res. 2000;86(5):494–501.
- 137. Wink DA, Mitchell JB. Chemical biology of nitric oxide: Insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide. Free Radic Biol Med. 1998;25(4– 5):434–56.
- 138. Szabo C, Ischiropoulos H, Radi R. Peroxynitrite: biochemistry, pathophysiology and development of therapeutics. Nat Rev Drug Discov. 2007;6(8):662–80.
- 139. Fortuno A, San Jose G, Moreno MU, Diez J, Zalba G. Oxidative stress and vascular remodelling. Exp Physiol. 2005;90(4):457–62.
- 140. Griendling KK, Harrison DG. Dual role of reactive oxygen species in vascular growth. Circ Res. 1999;85(6):562–3.
- 141. Irani K. Oxidant signaling in vascular cell growth, death, and survival: a review of the roles of reactive oxygen species in smooth muscle and endothelial cell mitogenic and apoptotic signaling. Circ Res. 2000;87(3):179–83.
- 142. Taniyama Y, Griendling KK. Reactive oxygen species in the vasculature: molecular and cellular mechanisms. Hypertension. 2003;42(6):1075–81.
- 143. Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. Circ Res. 2000;87(10):840–4.
- 144. Kojda G, Harrison D. Interactions between NO and reactive oxygen species: pathophysiological importance in atherosclerosis, hypertension, diabetes and heart failure. Cardiovasc Res. 1999;43(3):562–71.
- 145. Li PF, Dietz R, von Harsdorf R. Differential effect of hydrogen peroxide and superoxide anion on apoptosis and proliferation of vascular smooth muscle cells. Circulation. 1997;96(10):3602–9.
- 146. Cai H. NAD(P)H oxidase-dependent self-propagation of hydrogen peroxide and vascular disease. Circ Res. 2005;96(8):818–22.
- 147. Touyz RM, Schiffrin EL. Increased generation of superoxide by angiotensin II in smooth muscle cells from resistance arteries of hypertensive patients: role of phospholipase Ddependent NAD(P)H oxidase-sensitive pathways. J Hypertens. 2001;19(7):1245–54.
- 148. Griendling KK, Minieri CA, Ollerenshaw JD, Alexander RW. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. Circ Res. 1994;74(6):1141–8.
- 149. Zafari AM, Ushio-Fukai M, Akers M, Yin Q, Shah A, Harrison DG, Taylor WR, Griendling KK. Role of NADH/NADPH oxidase-derived H2O2 in angiotensin II-induced vascular hypertrophy. Hypertension. 1998;32(3):488–95.
- 150. Ushio-Fukai M, Zafari AM, Fukui T, Ishizaka N, Griendling KK. p22^{phox} is a critical component of the superoxide-generating NADH/NADPH oxidase system and regulates angiotensin II-induced hypertrophy in vascular smooth muscle cells. J Biol Chem. 1996;271(38):23317–21.
- 151. Galis ZS, Khatri JJ. Matrix metalloproteinases in vascular remodeling and atherogenesis: the good, the bad, and the ugly. Circ Res. 2002;90(3):251–62.
- 152. Rajagopalan S, Meng XP, Ramasamy S, Harrison DG, Galis ZS. Reactive oxygen species produced by macrophage-derived foam cells regulate the activity of vascular matrix metalloproteinases in vitro. Implications for atherosclerotic plaque stability. J Clin Invest. 1996;98(11):2572–9.
- 153. Pu Q, Neves MF, Virdis A, Touyz RM, Schiffrin EL. Endothelin antagonism on aldosterone-induced oxidative stress and vascular remodeling. Hypertension. 2003;42(1):49–55.
- 154. Liu J, Yang F, Yang XP, Jankowski M, Pagano PJ. NAD(P)H oxidase mediates angiotensin II-induced vascular macrophage infiltration and medial hypertrophy. Arterioscler Thromb Vasc Biol. 2003;23(5):776–82.
- 155. Weissmann N, Lobo B, Pichl A, Parajuli N, Seimetz M, Puig-Pey R, Ferrer E, Peinado VI, Dominguez-Fandos D, Fysikopoulos A, et al. Stimulation of soluble guanylate cyclase prevents cigarette smoke-induced pulmonary hypertension and emphysema. Am J Respir Crit Care Med. 2014;189(11):1359–73.
- 156. Dumitrascu R, Weissmann N, Ghofrani HA, Dony E, Beuerlein K, Schmidt H, Stasch JP, Gnoth MJ, Seeger W, Grimminger F, et al. Activation of soluble guanylate cyclase reverses experimental pulmonary hypertension and vascular remodeling. Circulation. 2006;113(2):286–95.
- 157. Evgenov OV, Pacher P, Schmidt PM, Hasko G, Schmidt HH, Stasch JP. NO-independent stimulators and activators of soluble guanylate cyclase: discovery and therapeutic potential. Nat Rev Drug Discov. 2006;5(9):755–68.
- 158. Li H, Forstermann U. Nitric oxide in the pathogenesis of vascular disease. J Pathol. 2000;190(3):244–54.
- 159. Arif E, Ahsan A, Vibhuti A, Rajput C, Deepak D, Athar M, Singh B, Pasha MA. Endothelial nitric oxide synthase gene variants contribute to oxidative stress in COPD. Biochem Biophys Res Commun. 2007;361(1):182–8.
- 160. Forstermann U. Oxidative stress in vascular disease: causes, defense mechanisms and potential therapies. Nat Clin Pract Cardiovasc Med. 2008;5(6):338–49.
- 161. Hogg JC. Pathophysiology of airflow limitation in chronic obstructive pulmonary disease. Lancet. 2004;364(9435):709–21.
- 162. Kharitonov SA. NOS:molecular mechanisms, clinical aspects, therapeutic and monitoring approaches. Curr Drug Targets Inflamm Allergy. 2005;4(2):141–9.
- 163. Kharitonov SA, Barnes PJ. Nitric oxide, nitrotyrosine, and nitric oxide modulators in asthma and chronic obstructive pulmonary disease. Curr Allergy Asthma Rep. 2003;3(2):121– 9.
- 164. Mueller CF, Laude K, McNally JS, Harrison DG. ATVB in focus: redox mechanisms in blood vessels. Arterioscler Thromb Vasc Biol. 2005;25(2):274–8.
- 165. Lassegue B, Clempus RE. Vascular NAD(P)H oxidases: specific features, expression, and regulation. Am J Physiol Regul Integr Comp Physiol. 2003;285(2):R277–97.
- 166. Cross AR, Segal AW. The NADPH oxidase of professional phagocytes–prototype of the NOX electron transport chain systems. Biochim Biophys Acta. 2004;1657(1):1–22.
- 167. Selemidis S, Sobey CG, Wingler K, Schmidt HH, Drummond GR. NADPH oxidases in the vasculature: molecular features, roles in disease and pharmacological inhibition. Pharmacol Ther. 2008;120(3):254–91.
- 168. Geiszt M, Lekstrom K, Witta J, Leto TL. Proteins homologous to $p47^{\text{phox}}$ and $p67^{\text{phox}}$ support superoxide production by NAD(P)H oxidase 1 in colon epithelial cells. J Biol Chem. 2003;278(22):20006–12.
- 169. Ago T, Kuribayashi F, Hiroaki H, Takeya R, Ito T, Kohda D, Sumimoto H. Phosphorylation of p47^{phox} directs phox homology domain from SH3 domain toward phosphoinositides, leading to phagocyte NADPH oxidase activation. Proc Natl Acad Sci U S A. 2003;100(8):4474–9.
- 170. Brandes RP, Kreuzer J. Vascular NADPH oxidases: molecular mechanisms of activation. Cardiovasc Res. 2005;65(1):16–27.
- 171. De Keulenaer GW, Alexander RW, Ushio-Fukai M, Ishizaka N, Griendling KK. Tumour necrosis factor alpha activates a p22^{phox}-based NADH oxidase in vascular smooth muscle. Biochem J. 1998;329(Pt 3):653–7.
- 172. Zalba G, San Jose G, Beaumont FJ, Fortuno MA, Fortuno A, Diez J. Polymorphisms and promoter overactivity of the p22(phox) gene in vascular smooth muscle cells from spontaneously hypertensive rats. Circ Res. 2001;88(2):217–22.
- 173. Moreno MU, San Jose G, Orbe J, Paramo JA, Beloqui O, Diez J, Zalba G. Preliminary characterisation of the promoter of the human p22(phox) gene: identification of a new polymorphism associated with hypertension. FEBS Lett. 2003;542(1–3):27–31.
- 174. San Jose G, Moreno MU, Olivan S, Beloqui O, Fortuno A, Diez J, Zalba G. Functional effect of the p22^{phox}—930 A/G polymorphism on p22^{phox} expression and NADPH oxidase activity in hypertension. Hypertension. 2004;44(2):163–9.
- 175. Fortuno A, Olivan S, Beloqui O, San Jose G, Moreno MU, Diez J, Zalba G. Association of increased phagocytic NADPH oxidase-dependent superoxide production with diminished nitric oxide generation in essential hypertension. J Hypertens. 2004;22(11):2169–75.
- 176. Wick MJ, Buesing EJ, Wehling CA, Loomis ZL, Cool CD, Zamora MR, Miller YE, Colgan SP, Hersh LB, Voelkel NF, et al. Decreased neprilysin and pulmonary vascular remodeling in chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2011;183(3):330– 40.
- 177. Sunday ME, Hua J, Torday JS, Reyes B, Shipp MA. CD10/neutral endopeptidase 24.11 in developing human fetal lung. Patterns of expression and modulation of peptide-mediated proliferation. J Clin Invest. 1992;90(6):2517–25.
- 178. Dusser DJ, Djokic TD, Borson DB, Nadel JA. Cigarette smoke induces bronchoconstrictor hyperresponsiveness to substance P and inactivates airway neutral endopeptidase in the guinea pig. Possible role of free radicals. J Clin Invest. 1989;84(3):900–6.
- 179. Carpenter TC, Stenmark KR. Hypoxia decreases lung neprilysin expression and increases pulmonary vascular leak. Am J Physiol Lung Cell Mol Physiol. 2001;281(4):L941–8.
- 180. Dempsey EC, Wick MJ, Karoor V, Barr EJ, Tallman DW, Wehling CA, Walchak SJ, Laudi S, Le M, Oka M, et al. Neprilysin null mice develop exaggerated pulmonary vascular remodeling in response to chronic hypoxia. Am J Pathol. 2009;174(3):782–96.
- 181. Shinall H, Song ES, Hersh LB. Susceptibility of amyloid beta peptide degrading enzymes to oxidative damage: a potential Alzheimer's disease spiral. Biochemistry. 2005;44(46):15345–50.
- 182. Papandreou CN, Nanus DM. Is methylation the key to CD10 loss? J Pediatr Hematol Oncol. 2010;32(1):2–3.
- 183. Sumitomo M, Shen R, Nanus DM. Involvement of neutral endopeptidase in neoplastic progression. Biochim Biophys Acta. 2005;1751(1):52–9.
- 184. Karoor V, Oka M, Walchak SJ, Hersh LB, Miller YE, Dempsey EC. Neprilysin regulates pulmonary artery smooth muscle cell phenotype through a platelet-derived growth factor receptor-dependent mechanism. Hypertension. 2013;61(4):921–30.
- 185. Lee SH, Channick RN. Endothelin antagonism in pulmonary arterial hypertension. Semin Respir Crit Care Med. 2005;26(4):402–8.
- 186. Jensen RT, Battey JF, Spindel ER, Benya RV. International Union of Pharmacology. LX-VIII. Mammalian bombesin receptors: nomenclature, distribution, pharmacology, signaling, and functions in normal and disease states. Pharmacol Rev. 2008;60(1):1–42.
- 187. Nemenoff RA, Simpson PA, Furgeson SB, Kaplan-Albuquerque N, Crossno J, Garl PJ, Cooper J, Weiser-Evans MC. Targeted deletion of PTEN in smooth muscle cells results in vascular remodeling and recruitment of progenitor cells through induction of stromal cellderived factor-1alpha. Circ Res. 2008;102(9):1036–45.
- 188. Klinger JR, Petit RD, Warburton RR, Wrenn DS, Arnal F, Hill NS. Neutral endopeptidase inhibition attenuates development of hypoxic pulmonary hypertension in rats. J Appl Physiol. 1993;75(4):1615–23.
- 189. Thompson JS, Morice AH. Neutral endopeptidase inhibitors and the pulmonary circulation. Gen Pharmacol. 1996;27(4):581–5.
- 190. Casserly B, Klinger JR. Brain natriuretic peptide in pulmonary arterial hypertension: biomarker and potential therapeutic agent. Drug Des Devel Ther. 2009;3:269–87.
- 191. Feil R, Lohmann SM, de Jonge H, Walter U, Hofmann F. Cyclic GMP-dependent protein kinases and the cardiovascular system: insights from genetically modified mice. Circ Res. 2003;93(10):907–16.
- 192. Lucas KA, Pitari GM, Kazerounian S, Ruiz-Stewart I, Park J, Schulz S, Chepenik KP, Waldman SA. Guanylyl cyclases and signaling by cyclic GMP. Pharmacol Rev. 2000;52(3):375– 414.
- 193. Silberbach M, Roberts CT Jr. Natriuretic peptide signalling: molecular and cellular pathways to growth regulation. Cell Signal. 2001;13(4):221–31.
- 194. Hamet P, Tremblay J, Pang SC, Skuherska R, Schiffrin EL, Garcia R, Cantin M, Genest J, Palmour R, Ervin FR, et al. Cyclic GMP as mediator and biological marker of atrial natriuretic factor. J Hypertens Suppl. 1986;4(2):49–56.
- 195. Corti R, Burnett JC Jr, Rouleau JL, Ruschitzka F, Luscher TF. Vasopeptidase inhibitors: a new therapeutic concept in cardiovascular disease? Circulation. 2001;104(15):1856–62.
- 196. Daull P, Jeng AY, Battistini B. Towards triple vasopeptidase inhibitors for the treatment of cardiovascular diseases. J Cardiovasc Pharmacol. 2007;50(3):247–56.
- 197. Maki T, Nasa Y, Yamaguchi F, Yoshida H, Mori M, Takada T, Horikawa E, Okano K, Takeo S. Long-term treatment with neutral endopeptidase inhibitor improves cardiac function and reduces natriuretic peptides in rats with chronic heart failure. Cardiovasc Res. 2001;51(3):608–17.
- 198. Xu J, Carretero OA, Liu YH, Yang F, Shesely EG, Oja-Tebbe N, Yang XP. Dual inhibition of ACE and NEP provides greater cardioprotection in mice with heart failure. J Card Fail. 2004;10(1):83–9.
- 199. Sumitomo M, Iwase A, Zheng R, Navarro D, Kaminetzky D, Shen R, Georgescu MM, Nanus DM. Synergy in tumor suppression by direct interaction of neutral endopeptidase with PTEN. Cancer Cell. 2004;5(1):67–78.