Biology of Nutrition Support and Gut Access in Critically III Patients

17

Norio Sato and Rifat Latifi

Background

Nutrition support for critically ill and injured patients has undergone significant advances in recent decades as the direct result of scientific progress and our increased knowledge of the biology and biochemistry of key nutrient changes induced by injury, sepsis, and other critical illnesses, both in adults and in children. The science of nutrition support (or, more accurately, of nutrition therapy) has become more diseasebased. Also called "specialized" or "artificial" nutrition support, nutrition therapy refers to the provision of either enteral nutrition (EN) via tube feeding or total parenteral nutrition (TPN). In contrast, "standard therapy" refers to a patient's own volitional intake, without the provision of nutrition therapy [1].

Of all the advances in nutritional support, the development of Total Parenteral Nutrition by Dr. Stanley J. Dudrick and his team is the most significant event to have clearly changed the way we

Department of Primary Care and Emergency Medicine, Kyoto University Hospital, Kyoto City, Japan e-mail: drnori@kuhp.kyoto-u.ac.jp

R. Latifi, M.D., F.A.C.S. (⊠) Department of Surgery, University of Arizona Medical Center, Tucson, AZ, USA e-mail: latifi@surgery.arizona.edu think about nutrition, the way we feed the critically ill and the way we support these patients who cannot, should not, or will not be fed orally [2]. Furthermore, in the past three to four decades, the understanding of the molecular and biological effects of nutrients in maintaining homeostasis in the critically ill population has made exponential advances. Traditionally, nutrition support in the critically ill population was regarded as adjunctive care designed to provide exogenous fuels to support the patient during the stress response. This support had three main objectives: to preserve lean body mass, to maintain immune function, and to avert metabolic complications. Recently, these goals have become more focused on nutrition therapy; specifically attempting to attenuate the metabolic response to stress, prevent oxidative cellular injury, and favorably modulate the immune response.

Depending on the individual patient's metabolic needs, nutrition therapy helps ensure that key nutrient substrates are replenished, or added in larger amounts, to supplement specific deficiencies or to simply prevent further deterioration and clinical consequences. The benefit of early institution of either EN or TPN in the overall care of critically ill and injured patients has now been well-established. After a critical illness or injury, the patient's energy and overall metabolic requirements greatly increase in order to sustain increased metabolism and the process of wound repair. Given current evidence derived from

N. Sato, M.D., Ph.D.

[©] Springer Science+Business Media New York 2015

R. Latifi et al. (eds.), *Technological Advances in Surgery, Trauma and Critical Care*, DOI 10.1007/978-1-4939-2671-8_17

randomized controlled trials (RCTs), early provision of nutrition for critically ill and injured patients is now a level I recommendation [3].

Critical illness and tissue injury initiate a complex series of rapidly responding homeostatic events in an attempt to prevent ongoing tissue damage and to activate the repair process. Classically, inflammation has been recognized as the hallmark of the homeostatic response. But more recently, attention has been focused on defining the complexities of response at the cellular, metabolic, and molecular levels. There is mounting evidence regarding the multiple specific metabolic changes in critically ill and injured patients and their need for fundamental nutrients and special substrates.

The benefit of the early institution of adequate enteral or parenteral nutrition in the overall management of critically ill patients has been wellestablished. Early nutritional support has the potential to reduce disease severity, diminish complications, and decrease the intensive care unit (ICU) length of stay and favorably impact patient outcome. Recognizing the importance of the appropriate nutritional support of trauma and critically ill patients with severe and/or multiple injuries, we have implemented a standard nutritional assessment and support program that aligns with guidelines set by the SCCM, ASPEN, ESPEN, and EAST. Despite all of this, it is very difficult to extrapolate nutritional support alone as the major factor impacting the patient's outcome. Sophisticated delivery tubing systems for enteral feeding, complex pharmacological mixtures of various nutrients in the form of parenteral nutrition, and recognition of various nutrients as pharmaco-nutrients have exploded in recent years. Gut access for provision of nutrition support has undergone major changes as well. Advanced laparoscopic approaches, percutaneous endoscopic access, and fine technology of these tubing systems have been refined continuously for the last few decades. In the following pages we describe some of the key nutrients and their effect on the biology of the disease.

n-3 Fatty Acids

Pharmacological Effects of n-3 Fatty Acids

Fatty acids are categorized as either saturated or unsaturated fatty acids. Monounsaturated fatty acids have one double bond while polyunsaturated fatty acids have two or more double bonds. Depending on the number of carbon atoms, fatty acids are also referred to as "short chain" (<8 carbons), "medium chain" (8-14 carbons), or "long chain" (≥ 15 carbons) fatty acids. Unsaturated fatty acids are also distinguished by where the first double bond occurs, counting from the carbon atom at the very end. These are typically designated $\omega 3$ (n-3), $\omega 6$ (n-6), and $\omega 9$ (n-9). The role of fatty acids include important aspects such as (1) sources of energy, (2) components of cell membranes, and (3) conversion into lipid metabolites that take part in physiological activities (e.g., prostagladins). Fatty acid administration leads to their incorporation into cell membranes. Arachidonic acid, which is an n-6 series fatty acid, plays a role in the formation of cell membranes, along with the n-3 series fatty acids eicosapentaenoic acid (EPA) and docosahexanoic acid (DHA). These fatty acids are discharged from cell membranes if cell injury or phospholipase activation occurs in association with a cellular insult. Although n-3 and n-6 fatty acids are metabolized by the same enzymes (cyclooxygenase and 5-lipoxygenase), the final metabolic products are different. When inflammatory mediators such as prostaglandin (PG) E_2 , thromboxane (TX) A_2 , or leukotriene (LT) B₄ are produced from n-6 fatty acids in excessive amounts, neutrophil activation is triggered which leads to systemic inflammatory response syndrome (SIRS). PGE₃, TXA₃, LTB₅, and other similar factors are produced from n-3 fatty acids; they are known to compete with metabolites of n-6 fatty acids and bring about anti-inflammatory activity. Reports indicate that n-3 fatty acids inhibit the production of the inflammatory cytokines interleukin (IL)-1 and tumor necrosis factor (TNF) by neutrophils and reduce the production of inducible nitric oxide synthase (iNOS) by macrophages. Furthermore, it has been demonstrated that n-3 fatty acids metabolize into resolvins and protectins, which promote convergence of inflammation, and have gained particular attention for their anti-inflammatory effects. The anti-inflammatory effect of n-3 fatty acids has been corroborated by many fundamental studies.

Clinical Studies Involving n-3 Fatty Acids

n-3 fatty acids is commonly found in a number of IEN or IMN, but one formula that has received particular attention is known that $Oxepa^{(0)}$ (Abbott Labs, Abbott Park, IL), which contains EPA, DHA, γ -linoleric acid (GLA), and antioxidants, is recommended for Acute lung injury (ALI) and Adult respiratory distress syndrome (ARDS) by guidelines in Europe, the USA, Canada, and other countries [3–5].

Pontes-Aruda et al. [6] studied 165 patients with severe septicemia involving acute lung injury (ALI) and acute respiratory distress syndrome who received Oxepa® and Pulmocare® (Abbott Labs, Abbott Park, IL). They reported that Oxepa[®] significantly improved the survival rate (67.3 % vs. 47.9 %) and decreased the number of days under artificial respiration control (14.6 days vs. 22.2 days) and the number of days patients remained in the intensive care unit (ICU) (17.2 days vs. 23.4 days). Furthermore, significant improvements were seen in oxygenating function and there was a significant reduction in the occurrence of new organ dysfunction. The fact that the control group in this study received a nutritional supplement that contained 55 % n-6 polysaturated fatty acid became an issue. In a study of the effectiveness of Oxepa® that included 106 patients with early-stage septicemia, the control group was given an ordinary nutritional supplement that contained 29 % fatty ingredients [7]. In that study, Oxepa® significantly decreased the incidence of septicemia (26.2 % vs. 51.0 %), the incidence of new organ dysfunction such as cardiovascular failure (21.0 % vs. 36.2 %) or respiratory failure (24.6 % vs. 39.6 %), the number of days patients remained in the ICU (7 days vs. 13 days), and the number of days patients remained in the hospital (9 days vs. 19 days). However, there was no significant change in mortality rates. Grau-Carmona et al. [8] compared Oxepa® to an ordinary nutritional supplement that contained 30 % fatty acids in 133 patients with septicemia. In that study, Oxepa[®] did not lead to any significant differences in mortality, oxygenation function, infection outbreak rate, or organ dysfunction incidence rate. However, Oxepa® was associated with significantly fewer days in the ICU (16 days vs. 18 days). Investigators also conducted a randomized controlled trial to examine the effectiveness of enteral bolus administration of EPA and DHA to patients with ALI. Stapleton et al. performed bolus administration of EPA and DHA to 90 patients with ALI and compared them to a group that did not receive this treatment. The group treated with bolus administration experienced a significant rise in the concentrations of EPA and DHA in the blood, but no significant differences were found in proinflammatory cytokine levels in the bronchoalveolar lavage fluid. Furthermore, there were no significant differences in mortality, number of days in the hospital, number of days in the ICU, or organ dysfunction incidence. Rice et al. [9] performed bolus administration of EPA, DHA, GLA, and antioxidants in 272 patients with ALI and compared them to a group that did not receive this treatment. The group that received the bolus administration had a significantly increased concentration of EPA in the blood but no significant differences were found in proinflammatory cytokines in the blood or in LTB4 concentration. Furthermore, a significant rise in the rate of death was observed in the group that received the bolus administration. It is necessary to consider the rise in death rate observed after bolus administration of EPA, DHA, and GLA separately from the antiinflammatory effects of Oxepa® that support the guidelines.

Fat emulsions containing fish oil (EPA and DHA) have been widely administered intravenously,

primarily in Europe, and clinical studies of their effectiveness have been conducted [10]. A recent meta-analysis of critically ill patients conducted by Manzanares et al. [11] confirmed a tendency toward decreased mortality and shortened period of mechanical ventilation; however, no effect was found on the infection rate or number of days in the ICU. Pradelli et al. [12] conducted a meta-analysis of patients who underwent elective surgery and critically ill patients, and they found significant decreases in infection rates, the number of days in the ICU, and the number of days in the hospital. When Palmer et al. [13] conducted a meta-analysis of critically ill patients in a similar manner, the conclusions were somewhat different; they reported no improvements in mortality, infection rate, or number of days in the ICU in patients who used fish oil. Currently, clinical studies have not yielded consistent conclusions. Although the issue of the sale of fish oil in the USA and Japan has not yet been available, there are physicians who are awaiting the permission of usage fat emulsions containing fish oil.

Glutamine

Pharmacological Effects of Glutamine

Glutamine is synthesized from other amino acids. Therefore, it is not necessary to ingest it as a nutrient when cells are not subject to insult. However, when the body is experiencing an insult such as trauma and sepsis the demand for glutamine increases, and the amount that can be synthesized by the body cannot keep up with demand. For this reason, it is considered a conditionally essential amino acid. Supplementation with glutamine to make up for relative shortages is considered beneficial for the body. Glutamine plays a diverse range of roles and can be converted into metabolites such as ammonia, nucleic acids, glutathione, arginine, and glucosamine. When the body is experiencing an insult, glutamine acts as an energy substrate in the intestinal mucosa, lymphocytes, macrophages, and neutrophils, and administration of glutamine to the

intestinal tract is considered effective for sustaining the intestinal tract barrier function and preventing bacterial translocation. Many reports have demonstrated that glutamine increases levels of heat shock proteins (HSPs) as an antiinflammatory mechanism. Intravenous administration of glutamine after the onset of septicemia activates the transcription factor heat shock factor-1, causing expression of HSP-70 and HSP-25 and thereby reducing the occurrence of acute lung disorders. Similarly, HSP-32 (heme oxygenase-1) is also expressed to reduce apoptosis and decrease tissue damage in the intestinal tract [14]. A report indicated that administration of glutamine inhibits apoptosis following stress or injury, and depletion of glutamine triggers apoptosis, which then activates caspase-3 and -9. Intravenous administration of glutamine inhibits the activation of nuclear factor-κB in the septicemia model [15]. It also inhibits activation of the stress-related p38 mitogen-activated protein kinase and extracellular signal-related kinase, resulting in reduced TNFa, IL-6, and IL-18. Similarly, enteral administration of glutamine has been shown to reduce serum IL-6 levels, increase IL-4 production by lymphocytes, and reduce interferon- γ production [16]. In an ischemia reperfusion model where a jenunal pouch was prepared and glutamine was administered, researchers confirmed inhibited activation of activator protein-1 and enhanced activation of proliferator-activated peroxisome receptor (PPAR). PPAR has drawn attention as a transcription factor with anti-inflammatory functions that is associated with reduced iNOS, reduced accumulation of neutrophils in the jejunum, retention of adenosine triphosphate levels in the jejunum, inhibition of intestinal tract permeability, and reduced tissue damage. Another study found that syndecan 1 was involved in reducing the action of glutamine in intestinal tract ischemia reperfusion damage [17]. It has also been noted that there is potential for arginine conversion through metabolism of glutamine to produce nitric oxide (NO), which can be harmful.

Clinical Studies Related to Glutamine

Many clinical trials have supported the usefulness of glutamine administration for reducing the infection rate during insult to the body. The results of large-scale clinical studies for determining the effectiveness of glutamine administration (enteral administration or intravenous administration) have been debatable, as some papers have cast doubt on the effectiveness of glutamine. Wernerman et al. [18] conducted a clinical study involving intravenous administration of glutamine (0.28 g/kg/day). The targeted number of cases was 1,000 over a 4-year period at 9 facilities in Sweden, 1 facility in Norway, and 1 facility in Finland (11 facilities in total). However, the study did not reach the target number of subjects; instead, the study included only 413 cases. Intravenous administration of glutamine in targeted patients with Acute Physiology and Chronic Health Evaluation II scores of 10 or higher, as well as in some mild cases, confirmed differences in the ICU death rate and the 28-day death rate; however, no difference was found in the mortality over a longer period. Andrews et al. [19] conducted a clinical study involving intravenous administration of glutamine (20.2 g/day) and intravenous administration of selenium (500 µg/ day); this was known as the SIGNET trial. Analysis of 502 cases from ten ICU facilities in Scotland could not confirm that glutamine effectively reduced the infectious disease occurrence rate, number of days in the ICU, or the death rate. Heyland et al. [20] found astounding results when they conducted a large-scale clinical study of glutamine administration (intravenous administration of 0.35 g/kg/day and enteral administration of 30 g/day) along with selenium and antioxidant administration. They analyzed 1,223 critically ill patients with dysfunction of two or more organs from 40 facilities in Canada, the USA, and Europe, and found that the death rate increased with glutamine. The results of this study, known as the REDOX study, indicate that excessive amounts of glutamine must not be administered to patients with highly critical illnesses. It is important to note that this study did not completely refute the benefits of glutamine yet. Additionally

SIGNET Trial [19] and REDOX Trial [20] tested the antioxidant nutrient using selenium. There was a reduction in the infection rate after intravenous administration of selenium (500 µg/day) for 5 days or more during the SIGNET Trial. However, in the REDOX study, when intravenous selenium administration (500 g/day) and enteral administration of selenium (300 µg), zinc (2 mg), β -carotene (10 mg), Vitamin E (500 mg), and Vitamin C (1,500 mg) was compared to groups without them, there were no differences in outcomes between the two groups.

Arginine

Pharmacological Effects Related to Arginine

Similar to glutamine, arginine is a conditionally essential amino acid. Arginine is metabolized in the body primarily through two routes [21]. One route leads to ornithine and urea owing to the action of arginase, while the other route leads to NO and citrulline because of the action of nitric oxide synthase (NOS). Arginase I is found primarily in the liver, and ornithine is metabolized further into polyamine and proline to improve collagen synthesis and wound healing. Alternatively, arginase II occurs primarily outside the liver and produces creatine in the small intestine mucous membrane to store muscle energy. Arginine increases the secretion of hormones such as growth hormones, glucagon, prolactin, and somatostatin, raising immune function when insults occur. Although appropriate concentrations of ornithine are essential for sustaining immune function, a report found that the differentiation of B-cells and the number of intestinal Peyer patch cells deceased in mice with a large amount of arginase, inhibiting production of T-cells and the T-cell receptor CD3. An appropriate amount of NO is essential for bactericidal action, and the vasodilation action of NO improves microcirculation, making oxygen transport to the periphery possible. However, excessive NO production leads to peroxynitrite production and tissue damage and can cause septic shock owing to vasodilation. The three arginase I and II enzymes as well as iNOS are all induced when inflammation occurs, and arginase and NOS compete in immune cells. There is a rise in the demand for arginine and because the metabolic behavior of arginine changes according to its concentration, it is difficult to confirm or repudiate the benefits of arginine.

Trends of Clinical Studies Related to Arginine

There have not been any clinical studies of arginine administration alone; clinical studies have used arginine-containing nutritional supplements that are commercially available. Galban et al. [22] studied 176 septicemia patients and reported that the group that received a nutritional supplement with enhanced arginine had a significantly reduced death rate compared to the control nutritional supplement group (19 % vs. 32 %). Kieft et al. [23] studied 597 ICU patients and compared a group that received a nutritional supplement containing enhanced arginine to a control group. They found that there were no differences between the groups in death rate, infection rate, or the number of days in the ICU. A meta-analysis of septicemia patients and critically ill patients except those with external injuries or burns, who received arginine found no differences in mortality or infection rate [24]. However, arginine administration in patients undergoing elective surgery has been found to reduce the infection rate and shorten the number of days in the hospital. It is also economically beneficial. Arginine administration in patients with critical septicemia for whom the hemodynamics may deteriorate is considered a situation to be avoided from a theoretical perspective [25].

Pharmacological Nutrients and Clinical Studies Related to Leucine

Reducing lean body mass index is known to be associated to mortality; therefore, avoiding reduction of skeletal muscles is essential. Among

the amino acids, some have functions that contribute to important aspects of body function such as signal transmission, phosphorylation, and gene expression. Leucine is a branched amino acid (BCAA), along with valine and isoleucine. Unlike other amino acids, leucine is directly metabolized in skeletal muscles. Leucine has cell growth promoting action and has been shown to stimulate the mammalian target of rapamycin (mTOR) to promote protein synthesis in skeletal muscles, cardiac muscles, and other tissues. Protein synthesis declines with septicemia, but administration of leucine is known to affect mTOR and increase protein synthesis. Studies on BCAAs have been conducted for more than 30 years, including a clinical study conducted in the 1990s investigating the administration of BCAAs for septic patients [26]. Jimenes et al. [27] conducted a study of 80 peritonitis patients in which they compared a group treated with 22.5 % BCAAs to another group treated with 45 % BCAAs at a nitrogen quantity of 12 g/day. While there was no difference in the death rate, there was an improvement in nitrogen balance and a reduction in 3-methylhistidine levels in the urine. Garcia-de-Lorenzo et al. [28] compared three groups of septic patients for whom enteral administration of nutrients was not possible. One group received 1.5 kg/kg of protein included 23 % BCAA, one group was given 1.5 g/kg of protein included 23 % BCAAs, and the last group was given 1.1 g/kg protein included 45 % BCAAs. While there were no improvements in nitrogen balance or similar outcomes, the two groups that received higher concentration of BCAA of 45 % had reduced mortality rates. The BCAAs included a combination of isoleucine, leucine, and valine in a 0.8:1.4:1 ratio. Many studies with high quantities of leucine have indicated the effectiveness of BCAAs in patients who are experiencing an injury. With the focus on early rehabilitation of ICU patients, the expectations for leucine increase in relation to the effectiveness of hydroxymethylbutyrate (HMB), a metabolite of leucine, as adjustment of skeletal muscle metabolism (suppression of protein catabolism and promotion of protein synthesis) and sustaining and increasing muscle mass lead to patient recovery and enhanced quality of life. Although there are

metabolic behaviors in which functional amino acids are important, it is important to remember that administering amino acids in a proper balance is important for maintenance of homeostasis of the entire body.

Pharmacological Nutrients and Clinical Studies Related to Whey Protein

Protein in milk is made up of 20 % whey protein and 80 % casein. The protein component that remains in milk when coagulated is referred to as whey protein (milk whey). Milk whey consists of α -lactalbumin, β -lactalbumin, glycomacropeptide, lactoferrin, and albumin. Reports have highlighted the capabilities of whey protein, including promotion of protein synthesis, anti-inflammatory action, and antioxidative action, among others. Suppression of phospholipase A2 and cyclooxygenase 2 by α -lactalbumin has an antiinflammatory effect. While whey protein suppresses production of cytokines such as TNF and IL-6 in a rodent liver injury model, it also increases production of cytokines via neutrophils and epithelial cells to improve immune function. Lactoferrin is an important factor for natural immunity related to biophylaxis in the digestive tract and mucous membranes. Discharge of lactoferrin included in the granules of neutrophils from activated neutrophils is known to raise the lactoferrin concentration in blood, from normal levels of 0.5-1 µg/mL to 200 g/mL. Lactoferrin has antioxidant properties, suppresses cytokine production, and functions in immunoregulation. Lactoferrin is known to bond with lactoferrinlipopolysaccharide (LPS), alleviate toll-like receptor signal transmission, stabilize circulatory behavior for administration of LPS, and suppress TNF- α and IL-6 production.

Manzoni et al. studied 472 newborns weighing 1,500 g or less in neonatal ICUs at 11 facilities [29]. They compared a group that received lactoferrin to a placebo control group and found that administration of lactoferrin significantly suppressed the occurrence of tardive septicemia, significantly reduced fungal infection, and reduced mortality. A group in the USA conducted a double-blinded randomized comparison in adults with severe sepsis at 24 facilities in a phase 2 study of talactoferrin, which is a recombinant form of human lactoferrin [30]. In this study of 194 targeted patients, the 28-day death rate of the group treated with human lactoferrin was 14.4 %, which was significantly lower than the 26.9 % death rate in the control group. The death rate was also significantly lower after 6 months. This clinical study indicated the effectiveness of lactoferrin in adults.

Gut Access

The indications for EN are fairly straightforward and well established. All resuscitated critically ill and trauma patients who cannot, will not, or should not receive nutrition orally and who have a functional GI tract should receive nutrition support via GI access as early as possible. EN delivery can be via temporary nasoenteric tube placement (orogastric, nasogastric, nasoduodenal, or nasojejunal) or via more long-term techniques (gastrostomy or gastrojejunostomy, which can each be done either surgically or endoscopically). Factors to consider in terms of the individual patient include the anticipated duration of EN, aspiration risk, GI tract function, ability to tolerate the given EN technique, degree of critical illness or injury, and specific goal of EN.

As stated above, gut access has undergone major technological changes. A recent study by Tuma et al. [31] that reviewed all articles on gut access found that the most common gut access techniques included nasoenteric tube placement (nasogastric, nasoduodenal, or nasojejunal) as well as a percutaneous endoscopic gastrostomy (PEG). Other open or laparoscopic techniques such as a jejunostomy or a gastrojejunostomy are also used. Early EN continues to be preferred whenever feasible. In addition, evidence is mounting that, during the early phase of critical illness or trauma, trophic feeding has an outcome comparable to that of full-strength formulas. Most patients tolerate EN through the stomach, so post pyloric tube feeding is not needed

initially. In critically ill and trauma patients early EN through the stomach should be instituted whenever feasible. Other approaches can be used according to patient needs, available expertise, and institutional guidelines. Nonetheless, more research is needed to ensure the safe use of surgical tubes in the open abdomen.

Indications for Gut Access

The indications for EN are fairly straightforward and well-established. All critically ill and trauma patients who cannot, will not, or should not receive nutrition orally and who have a functional GI tract need gastrointestinal access, as early as possible after they are fully resuscitated for nutritional support [32–36].

Techniques for Gut Access

Various techniques to obtain gut access and deliver EN exist. These include short-term options (nasogastric, nasoduodenal, or naso-jejunal tube placement) or more long-term or even permanent options (gastrostomy or gastroje-junostomy). All of these techniques can be done either surgically or endoscopically. The decision which technique to use depends on the individual patient and on anticipated duration of nutrition support, aspiration risk, GI tract function, ability to tolerate the given placement technique, degree of critical illness or injury, and the goals of EN [37–39].

Whichever technique for gut access is used, the tube placement must be confirmed by the presence of aspirating gastric fluid or by radiologic or other techniques of verification (for example, intraoperative view). The position of the feeding tube should be confirmed at the time of insertion before intermittent feedings, and during continuous feedings; a chest or abdominal radiograph may confirm placement, and the mark on the tube at the exit site indicates that the exposed tube length has not changed [40].

Post-pyloric Versus Gastric Tubes

The debate over post-pyloric tube feeding versus gastric feeding continues to this date [41–44]. Many critically ill and trauma patients can be started with orogastric or nasogastric tube feeding; if they continue to be intubated and require prolonged tube feeding, they may be switched to post pyloric feeding. However, if patients do not tolerate gastric feeding, switching them to naso-jejunal nutrition was not associated with better outcomes [45].

While nasogastric tubes or orogastric tubes (for intubated patients) constitute the initial short-term access for many patients, this is not ideal access for long-term access. Potential complications, although they are rare, include erosion of the nasal cartilage, sinusitis, and gastroesophageal reflux which in turn can lead to aspiration, pneumonitis, or pneumonia. Furthermore, dislodgment and blocking of these tubes is not uncommon.

Percutaneous Endoscopic Gastrostomy (PEG)

Ever since Ponsky and Gauderer [46] first reported a technique for percutaneous endoscopic gastrostomy (PEG) in the early 1980s, countless patients have been provided with EN through PEG. PEG is relatively safe and it is minimally invasive when compared with open surgical procedures. The indications and contraindications for PEG placement have been reported and are established [23]. The technique, however, has been standardized and does need further elaborations except to note that there are two ways of advancing the wire: the push and the pull technique. Two methods are possible for advancing the guidewire. If the pull method is used, the tapered gastrostomy tube is attached to the guidewire outside the patient's mouth; gentle tension is applied to the guidewire from the abdominal side to introduce the gastrostomy tube and guidewire unit into the esophagus and then into the stomach [47]. If the push method is used, the gastrostomy tube is passed over the guidewire, and the endoscope is used to grasp and stabilize the end of the wire beyond the gastrostomy tube; the gastrostomy tube is then pushed, by the endoscope, through the abdominal wall into the stomach, while tension is applied to both ends of the guide wire. With both methods, the endoscope may be reintroduced to confirm optimal placement (but without undue tension) of the internal bumper against the gastric mucosa. An external bumper is then used to secure PEG placement, with care taken to avoid undue pressure on the skin. A distance of 1–2 mm between the bolster and the skin is advisable, in order to decrease ischemic necrosis and subsequent infection.

PEG Gastrojejunostomy

Another technique for patients who cannot tolerate stomach feeding is the percutaneous gastrojejunostomy (PEGJ). The J-tube is passed through the existing PEG; the suture on the distal end of the J-tube is grasped with an endoscopic clip applicator passed through the working channel of the endoscope. The gastroscope and jejunal tube are then passed, under direct visualization, through the pylorus and into the small bowel. By firing the endoscopic clip, the surgeon secures the suture loop to the small-bowel mucosa. This technique allows for easier removal of the endoscope from the small bowel and minimizes the risk for inadvertent dislodgement of the jejunal feeding tube back to the stomach.

Surgical Gastrostomy

If PEG placement is contraindicated (see above), a surgically placed gastrostomy is an option, either via open technique or laparoscopy [48]. The Stamm open technique begins with the surgeon placing inner and outer purse string sutures on the mid-anterior gastric wall then an incision is made in the central portion of the purse string.

Laparoscopic approaches can be used as a modification of the Stamm procedure [49]. With two ports, one at the umbilicus for a 30° camera and the other in the right upper quadrant for use as a working port, the surgeon grasps the stomach and elevates it to the anterior abdominal wall, then selects an appropriate location for gastrostomy tube placement. The use of T-fasteners through the anterior abdominal wall and the gastric wall ensures the stomach is secured well and reduces overall morbidity if dislodgement occurs [50]. Through a small incision, a 14-gauge needle is introduced into the stomach and a guide wire is passed into the gastric lumen. The tract is serially dilated until the gastrostomy tube can be introduced.

Laparoscopically Assisted PEG

Laparoscopically assisted PEG is a preferred procedure in obese patients, in patients that have undergone previously laparotomies, and in those when we "do not see the light" as this ensures the gastrostomy-tube placement is checked by both the endoscopist and the surgeon [51]. The peritoneal cavity is accessed in the usual fashion. We use an open technique to insert a 30-degree laparoscope. A one or two 5 mm port is inserted also under direct vision. The stomach is inflated with a gastroscope and the needle from the anterior abdominal wall is inserted into the stomach under direct vision. The rest of the procedure is similar to PEG described above. One has to remember to lower the intraperitoneal pressure to 8-10 cm of H20 in order to allow the approximation of the stomach to the posterior abdominal wall.

Surgical Jejunostomy

Surgical jejunal access for feeding is another option. Both open and laparoscopic approaches are acceptable. Which technique the surgeon selects depends on experience and the clinical situation. When the patient is undergoing laparotomy and is in need of long-term nutrition support, an open Witzel jejunostomy is the preferred technique. Although in good hands, this procedure is safe and easy to perform, numerous technical and mechanical complications have been described [52].

Summary

This chapter outlined clinical studies of representative nutrients that are relevant to antiinflammation and some of the basic techniques for gut access. Nutritional studies performed in ICUs have historically been conducted using guidelines based on relatively small-scale clinical studies or opinions based on experience and there have been clear differences and unspecified aspects in the guidelines set by various nations and societies. Large-scale clinical studies have been conducted to clarify recommendations but their results and predictions are sometimes contradictory, which adds to the confusion. While it is important to pay attention to the outcomes of clinical studies, the studies conducted in different nations at numerous facilities have provided results that were not obtained consistently with regard to the treatment details, leading to a detrimental reduction in accuracy. Clinical studies with higher accuracy are believed to have more significance even if the number of targeted patients is smaller. Cases involving distress of critically ill patients require medical treatment over longer periods so anti-inflammatory action against persistent and recurring inflammation as well as immune-enhancing action is extremely important. Therefore, we should consider administration of nutrients with renewed focus on the actions of the nutrients in the living body. While nutrition support has changed dramatically in the recent two to three decades, it is imperative that, in order to improve our practice of nutritional support for trauma and critically ill patients, we should continue to pursue further understanding of their metabolism, energy requirements and specific requirements of each nutrient, and not rely on old dogma and guide-

Specifically, we need to continue to challenge the current understanding of nutritional science, the biology of nutrients and the interaction of these nutrients with trauma, injury and critical illness, and how to further advance timely and disease-directed nutritional support. In the meantime, we need to provide nutritional support to critically ill or injured patients early and we should tailor nutritional formulas based on the content of the nutrient compositions [34].

lines established by experts' opinions and semi-

quasi data [32, 33].

References

- Latifi R. Nutritional therapy in critically ill and injured patients. Surg Clin North Am. 2011;91(3):579–93.
- Dudrick SJ, Wilmore DW, Vars HM, Rhoads JE. Long-term total parenteral nutrition with growth, development, and positive nitrogen balance. Surgery. 1968;64(1):134–42.
- McClave SA, Martindale RG, Vanek VW, et al. Guidelines for the provision and assessment of nutrition support therapy in the adult critically ill patient: Society of Critical Care Medicine (SCCM) and American Society for Parenteral and Enteral Nutrition (A.S.P.E.N.). JPEN. 2009;33:277–315.
- Kreymanna KG, Bergerb MM, Deutz NEP, et al. ESPEN guidelines on enteral nutrition: intensive care. Clin Nutr. 2006;25:210–23.
- Canadian Clinical Practice Guidelines 2014. Summary of topics and recommendations. (http:// www.criticalcarenutrition.com/)
- Pontes-Arruda A, Aragao AM, Albuquerque JD. Effects of enteral feeding with eicosapentaenoic acid, gamma-linolenic acid, and antioxidants in mechanically ventilated patients with severe sepsis and septic shock. Crit Care Med. 2006;34:2325–33.
- Pontes-Arruda A, Martins LF, de Lima SM, et al. Enteral nutrition with eicosapentaenoic acid, gammalinolenic acid and antioxidants in the early treatment of sepsis: results from a multicenter, prospective, randomized, double-blinded, controlled study: the INTERSEPT study. Crit Care (London, England). 2011;15:R144.
- Grau-Carmona T, Moran-Garcia V, Garcia-de-Lorenzo A, et al. Effect of an enteral diet enriched with eicosapentaenoic acid, gamma-linolenic acid and anti-oxidants on the outcome of mechanically ventilated, critically ill, septic patients. Clin Nutr (Edinburgh, Scotland). 2011;30:578–84.
- Rice TW, Wheeler AP, Thompson BT, deBoisblan BP, Steingrub J, Rock P. Enteral omega-3 fatty acid, gamma-linolenic acid, and antioxidant supplementation in acute lung injury. JAMA. 2011;306:1574–81.
- Singer P, Berger MM, Van den Berghe G, et al. ESPEN guidelines on parenteral nutrition: intensive care. Clin Nutr. 2009;28:387–400.
- Manzanares W, Dhaliwal R, Jurewitsch B, Stapleton RD, Jeejeebhoy KN, Heyland DK. Parenteral fish oil lipid emulsions in the critically ill: a systematic review and meta-analysis. JPEN. 2014;38(1):20–8.
- Pradelli L, Mayer K, Muscaritoli M, Heller AR. n-3 fatty acid-enriched parenteral nutrition regimens in elective surgical and ICU patients: a meta-analysis. Crit Care (London, England). 2012;16:R184.
- Palmer AJ, Ho CK, Ajibola O, Avenell A. The role of omega-3 fatty acid supplemented parenteral nutrition in critical illness in adults: a systematic review and meta-analysis. Crit Care Med. 2013;41:307–16.
- 14. Uehara K, Takahashi T, Fujii H, et al. The lower intestinal tract-specific induction of heme oxygenase-1 by

glutamine protects against endotoxemic intestinal injury. Crit Care Med. 2005;33:381–90.

- Singleton KD, Beckey VE, Wischmeyer PE. Glutamine prevents activation of Nf-kappab and stress kinase pathways, attenuates inflammatory cytokine release, and prevents acute respiratory distress syndrome (ARDS) following sepsis. Shock (Augusta, Ga). 2005;24:583–9.
- Yeh CL, Hsu CS, Yeh SL, Chen WJ. Dietary glutamine supplementation modulates Th1/Th2 cytokine and interleukin-6 expressions in septic mice. Cytokine. 2005;31:329–34.
- Peng Z, Ban K, Sen A, et al. Syndecan-1 plays a novel role in enteral glutamine's gut protective effects of the post ischemic gut: role of syndecan-1 in gut protection by glutamine. Shock. 2012;38(1):57–62.
- Wernerman J, Kirketeig T, Andersson B, et al. Scandinavian glutamine trial: a pragmatic multicentre randomised clinical trial of intensive care unit patients. Acta Anaesthesiol Scand. 2011;55:812–8.
- Andrews PJ, Avenell A, Noble DW, et al. Randomised trial of glutamine, selenium, or both, to supplement parenteral nutrition for critically ill patients. BMJ (Clin Res ed). 2011;342:d1542.
- Heyland D, Muscedere J, Wischmeyer PE, et al. A randomized trial of glutamine and antioxidants in critically ill patients. N Engl J Med. 2013;368:1489–97.
- Wu G. Amino acids: metabolism, functions, and nutrition. Amino Acids. 2009;37:1–17.
- 22. Galban C, Montejo JC, Mesejo A, et al. An immuneenhancing enteral diet reduces mortality rate and episodes of bacteremia in septic intensive care unit patients. Crit Care Med. 2000;28:643–8.
- Kieft H, Roos AN, van Drunen JD, Bindels AJ, Bindels JG, Hofman Z. Clinical outcome of immunonutrition in a heterogeneous intensive care population. Intensive Care Med. 2005;31:524–32.
- Waitzberg DL, Saito H, Plank LD, et al. Postsurgical infections are reduced with specialized nutrition support. World J Surg. 2006;30:1592–604.
- Zaloga GP, Roberts PR, Marik P. Feeding the hemodynamically unstable patient: a critical evaluation of the evidence. Nutr Clin Pract. 2003;18:285–93.
- 26. Kazi AA, Pruznak AM, Frost RA, Lang CH. Sepsisinduced alterations in protein-protein interactions within mTOR complex 1 and the modulating effect of leucine on muscle protein synthesis. Shock (Augusta, Ga). 2011;35:117–25.
- Jimenez Jimenez FJ, Ortiz Leyba C, Morales Menedez S, Barros Perez M, Munoz Garcia J. Prospective study on the efficacy of branched-chain amino acids in septic patients. JPEN. 1991;15:252–61.
- Garcia-de-Lorenzo A, Ortiz-Leyba C, Planas M, et al. Parenteral administration of different amounts of branch-chain amino acids in septic patients: clinical and metabolic aspects. Crit Care Med. 1997;25: 418–24.
- Manzoni P, Stolfi I, Messner H, et al. Bovine lactoferrin prevents invasive fungal infections in very low birth weight infants: a randomized controlled trial. Pediatrics. 2012;129:116–23.

- Manzoni P, Rinaldi M, Cattani S, et al. Bovine lactoferrin supplementation for prevention of late-onset sepsis in very low-birth-weight neonates: a randomized trial. JAMA. 2009;302:1421–8.
- Tuma M, Latifi R, El-Menyar A, Al-Thani H. Gastrointestinal tract access for enteral nutrition in critically ill and trauma patients: indications, techniques, and complications. Eur J Trauma Emerg Surg. 2013;39:235–42.
- Joseph B, Wynne JL, Dudrick SJ, Latifi R. Nutrition in trauma and critically ill patients. Eur J Emerg Surg. 2010;36(1):25–30.
- Latifi R. Editorial: nutritional support of critically injured patients: do it early and do it right. Eur Surg. 2011;43(1):5–6.
- Latifi R, Uranues S. Feed the gut, feed early and with the right stuff, but do not forget total parenteral nutrition. Eur J Trauma Emerg Surg. 2013;39:201–2.
- DeLegge M. Enteral access—the foundation of feeding. JPEN. 2001;25:58.
- 36. Burlew CC, Moore EE, Cuschieri J, Jurkovich GJ, Codner P, Nirula R, Millar D, Cohen MJ, Kutcher ME, Haan J, Macnew HG, Ochsner MG, Rowell SE, Truitt MS, Moore FO, Pieracci FM, Kaups KL, the WTA Study Group. Who should we feed? A Western Trauma Association multi-institutional study of enteral nutrition in the open abdomen after injury. J Trauma Acute Care Surg. 2012;73(6):1380–7. Discussion 1387–8.
- Gopalan A, Khanna S. Enteral nutrition delivery technique. Curr Opin Clin Nutr Metab Care. 2003;6(3): 313–7.
- Huang HH, Chang SJ, Hsu CW, et al. Severity of illness influences the efficacy of enteral feeding route on clinical outcomes in patients with critical illness. Acad Nutr Diet. 2012;112(8):1138–46.
- Rice TW. Can we stomach gastric feeding in critically ill patients with gastrointestinal intolerance? Crit Care Med. 2012;40(8):2508–9.
- 40. Metheny NA, Titler MG. Assessing placement of feeding tubes. Am J Nurs. 2001;101:36–46.
- 41. Montecalvo MA, Steger KA, Farber HW, et al. Nutritional outcome and pneumonia in critical care patients randomized to gastric versus jejunal tube feedings. Crit Care Med. 1992;20:1377–87.
- 42. Kearns PJ, Chin D, Mueller L, et al. The incidence of ventilator-associated pneumonia and success in nutrient delivery with gastric versus small intestinal feeding: a randomized clinical trial. Crit Care Med. 2000;28(6):1742–6.
- 43. ChienWei H, ShuFen S, Shoa L, et al. Duodenal versus gastric feeding in medical intensive care unit patients: a prospective, randomized, clinical study. Crit Care Med. 2009;37:1866–72.
- 44. Kulvatunyou N, Joseph B, Tang A, et al. Gut access in critically ill patients: where have we gone thus far? Eur Surg. 2011;43(1):24–9.
- 45. Davies AR, Morrison SS, Bailey MJ, et al. Multicenter, randomized controlled trial comparing early nasojejunal with nasogastric nutrition in critical illness. Crit Care Med. 2012;40(8):2342–8.

- Ponsky JL, Gauderer MW. Percutaneous endoscopic gastrostomy: a nonoperative technique for feeding gastrostomy. Gastrointest Endosc. 1981;27:9–11.
- Jane P. Technique of upper gastrointestinal endoscopy. Gastrointest Endosc Clin N Am. 1994;4:501–52.
- 48. Zollinger Jr RM, Zollinger Sr RM. Plate 10. Percutaneous endoscopic gastrostomy—PEG. In: Zollinger Jr RM, Zollinger Sr RM, editors. Zollinger's atlas of surgical operations. 9th ed. New York: McGraw-Hill; 2011.
- Zollinger Jr RM, Zollinger Sr RM. Plate 9. Gastrostomy. In: Zollinger Jr RM, Zollinger Sr RM,

editors. Zollinger's atlas of surgical operations. 9th ed. New York: McGraw-Hill; 2011.

- Duh Q, Way L. Laparoscopic gastrostomy using T-fasteners as retractors and anchors. Surg Endosc. 1993;7:60–3.
- Charlesworth P, Hallows M, van der Avoirt A. Singlecenter experience of laparoscopically assisted percutaneous endoscopic gastrostomy placement. J Laparoendosc Adv Surg Tech A. 2010;20(1):73–5.
- Tapia J, Murguia R, Garcia G. Jejunostomy: techniques, indications, and complications. World J Surg. 1999;23(6):596–602.