

# Chapter 17

## Investigative Therapies in Pulmonary Arterial Hypertension

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**Abstract** Pulmonary arterial hypertension (PAH) remains a serious, life threatening disease of unclear etiology. Despite the rapid development of numerous drugs to treat the disease, no cure is presently available. New treatments for PAH that are able to reverse the abnormal pulmonary vascular remodeling that is responsible for much of this disease are badly needed. To accomplish this goal, novel therapies that target many of the dysfunctional pathways that have been identified in PAH will need to be developed. This chapter reviews many of the known alterations in gene expressions, vasoconstriction, inflammation, metabolism, and cellular proliferation that have been identified in the pathogenesis of PAH. Potential pharmacologic targets arising from these abnormalities are reviewed along with data, where available, from animal studies and small clinical trials that have attempted to treat pulmonary vascular disease through manipulation of these pathways.

**Keywords** Pulmonary arterial hypertension • Investigational therapy • Gene therapy • Cell therapy • Bone morphogenic protein receptor 2

### Abbreviations

5HT	Serotonin
ACE	Angiotensinogen-converting enzyme
ALK-1	Activin like kinase 1
AT III	Angiotensin III
AT-1	Angiotensinogen receptor-1
AT-2	Angiotensinogen receptor-2
BMPR2	Bone morphogenetic protein receptor 2
CML	Chronic myelogenous leukemia
COPD	Chronic obstructive lung disease

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DCA	Dichloroacetate
DHEA	Dehydroepiandrosterone
EIF2AK4	Eukaryotic translation initiation factor 2 alpha kinase 4
eNOS	Endothelial nitric oxide synthase
EPCs	Endothelial progenitor cells
HDAC	Histone deacetylase
HIF1 $\alpha$	Hypoxia inducible factor 1 $\alpha$
IL-1	Interleukin-1
IL-13	Interleukin-13
IL-6	Interleukin-6
KCNK3	Potassium channel subfamily K member 3
Kv	Voltage-gated potassium channels
miRNA	MicroRNA
MSC	Mesenchymal stems cells
NFAT-1	Nuclear factor of activated T-cells 1
NO	Nitric oxide
PAH	Pulmonary arterial hypertension
PASMC	Pulmonary artery smooth muscle cells
PDGF	Platelet-derived growth factor
PGI <sub>2</sub>	Prostacyclin
PH	Pulmonary hypertension
PPAR $\gamma/\beta$	Peroxisome proliferator-activated receptors $\gamma/\beta$
RAAS	Renin–angiotensin–aldosterone system
RV	Right ventricle
SERCA 2a	Sarcoendoplasmic reticulum calcium transport ATPase 2a
siRNA	Small interfering RNA
SOD2	Superoxide dismutase 2
TGF $\beta$	Transforming growth factor $\beta$
TKI	Tyrosine kinase inhibitor
TRPC	Transient receptor potential channels
TXA <sub>2</sub>	Thromboxane
VEGF	Vascular endothelial growth factor receptor
VEGFr	Vascular endothelial growth factor receptor
VIP	Vasoactive intestinal peptide

## Introduction

Despite the availability of multiple therapies specifically approved by regulatory agencies for the treatment of pulmonary arterial hypertension (PAH), the diagnosis remains indicative of a progressive, life-limiting process resulting in substantial morbidity and mortality. Identification of treatments aimed to normalize life expectancy in these patients is a significant priority and to date no single therapy for PAH

can achieve this goal. This is not at all surprising given the multifactorial nature of the pathogenesis of PAH and the clinical heterogeneity of patients classified as World Health Organization Group 1 PAH. Future effective treatments for PAH may ultimately rely on identification and development of multi-targeted strategies that go well beyond our current therapeutic targets. It is impossible to summarize all the potential therapies that are being evaluated for PAH. Focusing on important pathologic processes in PAH and investigative therapies related therein may provide an important approach to identify integrative, complementary, pluripotent therapies for future consideration [1].

## Potential Therapeutic Targets for PAH

The pathophysiology of PAH is complex with many potential targets that have been identified through various studies including genomic approaches, clinical investigations, and preclinical animal studies. There are equally numerous ways to categorize these targets. For purposes of this review, selected targets for future treatments will be considered. Additionally, most current and future treatments are aimed at reversing the abnormal structure and function of the pulmonary circulation. Enhancing right ventricle (RV) function is also an important consideration in improving patient functional status and outcome.

## Genetic and Epigenetic Targets

Identification of bone morphogenetic protein receptor 2 (BMPR2) mutations as a cause of familial PAH in 2000 heralded over 14 years of research devoted to understanding how the wide variety of mutations cause PAH. Haploinsufficiency, dominant negative effects, and abnormal signaling through downstream pathways have all been associated with PAH. Because of these variable effects of the BMPR2 mutation, the approach to restoring normal gene expression and function of BMPR2 is daunting. What is clear is that developing ways to increase expression of the normal BMPR2 allele may result in substantial benefit. Gene therapy via adenoviral gene transfer in animal studies has demonstrated benefit of increasing expression of the normal BMPR2 [2]. However, gene therapy in humans with PAH is significantly in the future. Alternative strategies, aimed at increasing BMPR2 expression and activity, have led to several new lines of clinical investigation. Recently, FK506 (tacrolimus) has been identified as a potential potent inducer of BMPR2 activity. Its effect on BMPR2 is only partially explained by inhibition of calcineurin and nuclear factor of activated T-cells 1 (NFAT-1) signaling. Tacrolimus was also able to improve endothelial cell function in cells derived from PAH patients as well as decrease pulmonary hypertension (PH) in hypoxic mice with BMPR2 haploinsufficiency and rats with monocrotaline and vascular endothelial growth factor receptor (VEGFR)/hypoxia-induced PAH [3].

Other targets aimed to increase BMPR2 expression and activity by acting as molecular chaperones and increasing transcription have been proposed. Ataluren increases ribosomal transcription of genes with stop codon mutations [4] and has been tested in several genetic diseases with mixed results. Ongoing trials in patients with cystic fibrosis are taking place now [5] and may point to a possible option in PAH patients with specific BMPR2 mutations in the future. Some mutations of BMPR2 are associated with protein folding abnormalities that do not allow BMPR2 trafficking out of the endoplasmic reticulum and several different chaperones to move the mis-folded protein to the cell surface have been studied [6].

While mutations in BMPR2 are most commonly associated with PAH, other genes such as activin-like kinase 1 (ALK-1), endoglin, potassium channel subfamily K member 3 (KCNK3), eukaryotic translation initiation factor 2 alpha kinase 4 (EIF2AK4), and voltage-gated potassium channels (Kv's) that are mutated or abnormally expressed in PAH may also benefit from the approaches above. Additionally, abnormal activity of BMPR2 signaling may also alter the expression of endogenous regulators of the lung circulation. One such example is apelin, a peptide that is implicated in endothelial and smooth muscle proliferation and vasodilation, which is reduced by mutations in BMPR2 that disrupt peroxisome proliferator-activated receptors  $\gamma/\beta$  (PPAR  $\gamma/\beta$ ) signaling, thus providing some rationale for the therapeutic potential of PPAR agonists in PAH [7–9]. PPAR agonists are also associated with attenuation of PH in animal models by modulating the levels of other vasoactive mediators such as endothelin-1 and VEGF [10].

Other epigenetic changes are also found in PAH patients. One area of increasing interest is the role of micro RNAs (miRNAs) and small interfering RNAs (siRNAs) to regulate gene expression in PAH. Transfection of RNA containing vectors to enhance or inhibit gene expression has been proposed as potential future therapies as increasing evidence supports the role of miRNA and siRNA in PAH. A recent review highlighted many of the RNAs implicated in PAH with additional RNA targets reported every year [11].

DNA modifications such as increased methylation lead to changes in expression of the gene. An example is the methylation of superoxide dismutase 2 (SOD2) by methyltransferases resulting in decreased SOD expression, changes in redox state and increased expression of pro-proliferative signals such as hypoxia inducible factor 1 $\alpha$  (HIF1 $\alpha$ ) [12, 13]. Modification of histones by acetylation in PAH has also been proposed as an important epigenetic phenomenon. Inhibition of de-acetylation by inhibitors of histone deacetylase (HDAC) causes changes in gene expression that result in inactivation of some genes and activation of others. In PAH, studies in pulmonary artery smooth muscle cells (PASMCs) and several animal models of PH have demonstrated a decrease in PASMC proliferation and inflammation associated with targeted inhibitors of HDAC [14]. Inhibitors of HDAC have also been used in studies looking at RV function in models of PAH with mixed results. In left ventricular hypertrophy, inhibitors of HDAC were beneficial, whereas in studies of different models of PAH and with different HDAC inhibitors, effects on the RV were beneficial (in compensated RV hypertrophy and monocrotaline-induced PH) but were associated with RV failure in others [15, 16].

Manipulating the expression of genes implicated in PAH is a promising area of active investigation. Hopefully, restoration of more normal gene expression and ultimately more normal cell signaling will restore normal pulmonary vascular and RV function and improve the life expectancy in PAH patients.

## Vasoconstriction and Vasoactive Targets

Enhanced pulmonary vasoconstrictive responses to a wide variety of stimuli have been identified in PAH. Indeed, all currently available PAH treatments directly target vasoconstrictive pathways.

Vasoconstriction of vascular smooth muscle requires an influx of calcium in order to activate the contractile machinery. Calcium can be released from intracellular stores or imported from the extracellular environment. Transient receptor potential channels (TRPC) regulate the release of calcium from intracellular stores. Increased activity of TRPC 6 has been identified in PAH patients and is associated with a single-nucleotide polymorphism [17]. Increased expression of TRPC 6 was also associated with enhanced proliferation of PASMCs [18]. Loss of TRPC 4 channel activity in rats is associated with improved survival in models of PAH [19]. The sarcoendoplasmic reticulum calcium transport ATPase 2a (SERCA 2a), a channel that regulates release of calcium from the sarcoplasmic reticulum, has also been implicated in PAH [20]. Modulating the influx of calcium has been a target of PAH therapies by directly inhibiting the entry of calcium through T- or L-type channels using calcium channel blockers. However, the clinical effectiveness of these alone has been modest at best and more selective regulators of calcium entry such as the channels above may provide important treatment targets [21].

Calcium influx occurs when the smooth muscle cell is depolarized. In PAH, several voltage-gated potassium channels (Kv) have been implicated. Down regulation of several potassium channels, most notable Kv1.2 and 1.5 are associated with membrane depolarization and calcium influx. Maneuvers that increase Kv expression and function (by gene transfer, dichloroacetate, etc.) reduce experimental PH by causing cellular hyperpolarization [22]. Recently, the KCNK3 gene that encodes potassium channel subfamily number 3 has been identified as a cause of heritable PAH in several families. Mutations in this channel lead to a channelopathy characterized by loss of potassium channel function and hyperpolarization of smooth muscle cells [23]. This finding definitively links abnormal potassium channel function to PAH and restoring normal channel function is a viable strategy for future consideration.

Vasoconstriction can also be achieved by a variety of vasoactive molecules that interact with cell surface receptors and cause vasoconstriction through a variety of intracellular signaling processes. Current treatments including endothelin-1 receptor blockade or increasing levels of prostacyclin-generated cAMP and nitric oxide-generated cGMP take advantage of this strategy to lessen or enhance downstream signals respectively. There are many other molecules that modulate vasoconstriction

including vasoactive intestinal peptide (VIP), thromboxane (TXA<sub>2</sub>), angiotensin III (AT III), apelin, serotonin (5HT), etc. all of which may cause pulmonary vasoconstriction through several different pathways. VIP inhibits vasoconstriction and also acts to vasodilate the circulation. Decreased VIP expression results in worse PH in animal models, and treatment with exogenous VIP attenuates this effect [24]. However, despite positive results from early clinical trials, VIP administration in PAH patients did not result in measureable improvements [25]. 5HT has long been associated with PAH and interference with the serotonin transporter has been demonstrated to decrease PH in animal models [26]. Terguride, is a 5HT transporter inhibitor, that was not found to be effective in the treatment of PAH in a preliminary phase IIa study [27], but continues to be investigated in combination with other drugs. Apelin, as described above is decreased in PAH due to mutations in BMPR2. It has many effects on the pulmonary circulation and dilates the lung circulation of animals with PH possibly through its effects on altering expression of endothelial nitric oxide synthase (eNOS) to increase nitric oxide (NO) production. Apelin administration has been associated with decreased PH in animal models [28].

Activation of Rho kinase in PAH leads to sustained contraction in PSMCs through “Ca sensitization” of the contractile apparatus, namely increased levels of phosphorylated myosin light chain [29, 30]. Inhibitors of Rho kinase, both direct and indirect, result in substantial reversal of vasoconstriction and vascular remodeling in several animal models of PH [31–36]. Several small scale human clinical trials of acute administration of the direct Rho kinase inhibitor fasudil in PAH patients demonstrated acute pulmonary vasodilatory effects [37–40]. Long-term clinical trials of direct Rho kinase inhibitors have not yet been carried out. Indirect inhibitors of Rho kinase, such as dehydroepiandrosterone (DHEA), are attractive candidate drugs that have been associated with attenuation and reversal of PH in animal models [41] and improved exercise capacity in patients with chronic obstructive lung disease (COPD) associated PH [42].

Identification, testing and production of specific inhibitors of the above mentioned agents (many of which are implicated in PAH), are daunting. A strategy that identifies important intracellular signaling hubs common to several important players in the pathogenesis of PAH may prove more effective than targeting pathways individually.

## **Inflammation and Inflammatory Mediator Targets**

Inflammation that sets in motion a series of events that promote vascular dysfunction and remodeling has long been hypothesized to be an important contributor to the development of PAH [43]. The association of PAH with autoimmune and inflammatory diseases is a significant part of this hypothesis. However, immunosuppressive therapies have had limited effectiveness in the treatment of PAH. The pro-inflammatory cytokines, interleukin-1 (IL-1) and -6 (IL-6), are increased in PAH patients and inhibition of these is associated with decreased PH in animal

models. IL-13 and signaling via transforming growth factor  $\beta$  (TGF  $\beta$ ) is additionally associated with schistosomiasis-related PAH [44]. Several animal studies and case reports have found that inhibition of IL-6 is associated with decreased PH [45]. The anti-IL-6 monoclonal antibody has been associated with decreased pulmonary vascular disease in a patient with PAH associated with connective tissue disease and may represent a future therapeutic option for some PAH patients [46]. Cell-mediated immune functions have also been implicated in PAH and may represent important therapeutic targets. Patients with human HIV and athymic rats are more susceptible to PH suggesting an important role of T-cells and chronic inflammation in PAH [47, 48]. Given the relationship to autoimmune diseases, B-cell activation may play a role in PAH suggesting the potential option for anti-B-lymphocyte antigen CD20 treatment in the future. Several case reports suggest that rituximab may be helpful in patients with PAH associated with connective tissue disease [49, 50].

## Vascular Cell Proliferation and Vessel Remodeling Targets

Excessive proliferation of cells in the walls of the pulmonary arteries and formation of the plexiform lesion is a hallmark of severe PAH. Events leading to the alteration of the normal vascular structure likely are multifactorial involving biochemical and mechanical forces that promote cell proliferation and angiogenesis. Many molecules known to be important in PAH such as NO, 5HT, TXB2, Rho kinase, and prostacyclin (PGI2), have direct effects on vascular tone, but also contribute to the proliferative phenotype. Thus, manipulation of these agents may help to decrease or reverse pulmonary vascular remodeling in addition to decreasing pulmonary vascular tone.

More direct approaches to address vascular remodeling target specific growth factor pathways and the extracellular matrix. Several important growth factors have been implicated in PAH. Platelet derived growth factor (PDGF) has been implicated in the proliferation of pulmonary vascular cells. Imatinib, a tyrosine kinase inhibitor (TKI) that inhibits the PDGF receptor, has been the most widely studied tyrosine kinase inhibitor in PAH. Imatinib was originally developed to inhibit the pro-oncogenic tyrosine kinase Bcr-abl in chronic myelogenous leukemia (CML). Given its ability to inhibit PDGF receptor and proto-oncogene c-Kit, imatinib was proposed as a potential regulator of PAH-related vascular proliferation [51, 52]. Despite initial excitement in animal models, case reports and small clinical trials [53, 54], imatinib failed to achieve regulatory approval after a large-scale clinical trial identified important concerns regarding the risk to benefit ratio of the treatment [55]. Other TKIs that target PDGF have also had some encouraging preclinical success, but may have limited clinical usefulness owing to other less beneficial effects. For example, dasitinib, a TKI with broader inhibitory properties than imatinib, also approved for treatment of CML, has been associated with the development of PAH [56].

Other potential targets are TKIs targeting epidermal growth factor receptor (erlotinib, gefitinib) and multi-kinase inhibitors (sorafenib, sunitinib), but as was seen with imatinib, significant limitations to their use lie in unacceptable adverse events and side effects and limited efficacy in animal models [57, 58]. Given these contradictory clinical effects and substantial adverse events, the future of TKIs in PAH is uncertain [59].

Modulation of extracellular matrix in PAH may also be a potential target for future treatments. Increased activity of elastases located in the pulmonary vascular wall allows for degradation of the extracellular matrix and, by inducing expression of other molecules such as tenascin and fibronectin, promotes cell proliferation and vascular remodeling [60]. Elafin, an inhibitor of serine protease, decreases and reverses severe PH in animal models [61, 62] and will likely undergo clinical testing in the near future.

## Metabolic Targets

Derangements in the energy metabolism of pulmonary artery cells in PAH are being characterized. Examination of the RV and lungs of patients and animals with experimentally induced PH demonstrate a wide array of changes in expression of genes engaged in metabolic processes [63, 64]. Several mitochondrial abnormalities have been found in pulmonary vascular cells and the RV that lead to anaerobic metabolism (glycolysis) instead of aerobic metabolism through activation of pyruvate dehydrogenase kinase. This glycolytic state then leads to decreased intracellular reactive oxygen species generation, increased HIF1 $\alpha$  expression and activity, and ultimately activation of vasoconstrictive and proliferative signaling by decreasing Kv channel expression and activity [65]. Dichloroacetate (DCA), an inhibitor of mitochondrial pyruvate dehydrogenase has demonstrated reversal or reduction of PH in animal models [66–69] and is undergoing clinical testing in PAH patients. Oxidation of fatty acids also contributes to this glycolytic shift, and several inhibitors of fatty acid oxidation, trimetazidine and ranolazine, have demonstrated potential efficacy in animal models of PAH by reversing the shift toward glycolysis [70, 71] Ranolazine is currently being evaluated in PAH patients at a single clinical center.

## Neurohormonal Targets

Much of the morbidity and mortality of PAH patients lies in the adaptation and function of the RV to the increased demands of downstream resistance. Clearly, developing effective treatments to decrease the resistance due to vasoconstriction and remodeling is the ultimate RV “specific” treatment. Short of this, improving adaptation of the RV is an important therapeutic consideration. Many of the proposed PAH treatments above additionally involve adaptations that are seen in the



RV as well as pulmonary vasculature. Recently, attention to the neurohormonal activation in PAH patients has received increasing attention. The sympathetic nervous system and renin–angiotensin–aldosterone system (RAAS) are activated in PAH [72] and represent potential targets for treatment.

Sympathetic tone is increased in patients with PAH and adrenergic receptor expression is also reported as increased in the RV of patients with PAH right ventricles [73, 74]. Activation of this system was considered adaptive to sustain cardiac output by increasing contractility, heart rate, and systemic blood pressure, and thus, manipulation of adrenergic receptors was considered unsafe in PAH patients. However, sympathetic activation has been associated with increased mortality in PAH patients [75] which has led to a number of studies of selective  $\beta$ -receptor blockade in PAH. In several preclinical models of PAH, the  $\alpha$ 1/ $\beta$ 1/ $\beta$ 2-adrenergic receptor antagonist carvedilol and  $\beta$ 1-selective receptor antagonist bisoprolol are associated with improved RV function [76–78]. A retrospective review of PAH patients revealed no difference in clinical outcomes in patients who received  $\beta$ -blockers compared to those who did not [79, 80] suggesting that the safety of  $\beta$ -blockade in PAH patients should be reconsidered [72].

Activation of RAAS is also found in patients with PAH [72, 81]. Several case series of patients with PAH hinted at efficacy of angiotensinogen converting enzyme (ACE) inhibition [82], but concerns regarding the potential for hypotension limited further investigations. More recently, approaches aimed at selective targeting of the angiotensin (AT) pathway have identified the potential therapeutic value of targeting AT receptor-1 (AT1) (using the AT-1 antagonist losartan) while keeping ACE and AT-2 function (both with purported vasodilatory effects) and signaling intact [81]. Additional support of this approach is demonstrated by the use of ACE-2 agonists to decrease pulmonary artery pressure and RV hypertrophy and dysfunction in animal models of PAH [83–85].

Aldosterone antagonism is a mainstay of left ventricular failure treatment [86], but its role in RV failure is less well understood. However, increased levels of aldosterone are found in PAH patients [87] and treatment with aldosterone antagonists is included in several recent treatment guidelines. Evidence for this recommendation is increasingly apparent as demonstrated in recent studies that identified that upregulation of aldosterone is associated with increased ET-1 and decreased NO availability which is reversed by aldosterone antagonists in animal models of PAH [88, 89]. Review of the ARIES data of the selective ET-1, a receptor antagonist ambrisentan in PAH, suggests a complimentary benefit when combined with the aldosterone antagonist spironolactone [90].

## Lung Cell-Targeted and Cell-Based Therapeutic Targets

Therapies designed to repair and remodel the pulmonary arteries in PAH may be best approached using strategies to deliver pharmacologic agents and genes directly to the abnormal lung circulation or to repopulate the pulmonary vasculature with

cells that have normal phenotypes and behaviors. Targeting molecules to the lung circulation can be achieved by a number of mechanisms including the use of chaperones, exploiting lung-specific cell surface receptors, targeting lung-specific gene expression and identification of lung-specific signaling pathways. Numerous examples of this strategy are being investigated including use of specific lung endothelial cell surface receptors, peptides that chaperone molecules to abnormal pulmonary endothelium, and RNAs that target lung-specific gene expression. By using lung-targeted approaches, the limitations of systemic delivery of therapeutics and potential systemic side effects may be limited [91].

Cell-based strategies to deliver molecules to the lung circulation or act as therapeutics directly are an attractive approach in PAH. Repair and reversal of vascular changes in PAH through delivery of pluripotent mesenchymal stem cells (MSCs) or endothelial progenitor cells (EPCs) that repopulate the vessel wall and potentially carry genes to promote repair is one potential approach [92, 93]. In PAH patients, alterations in the numbers of circulating EPCs have been observed [94]. EPCs alone or carrying specific genes, such as eNOS, have prevented and reversed PAH in several animal models of PAH [95]. Autologous EPCs in a small trial of PAH patients resulted in improvement in exercise capacity and hemodynamics [96]. A trial of autologous EPCs overexpressing eNOS in PAH patients is underway.

## Summary

Identification of new treatments that will normalize life expectancy for patient with PAH will certainly be a complex and challenging endeavor is likely to involve the targeting of multiple pathways. It is unlikely, that any single agent exists that is capable of completely reversing or preventing disease progression in PAH, but careful assessment of the molecular underpinnings of PAH, followed by the rational development of complementary multi-targeted approaches, may represent the best hope for the future.

## References

1. Morrell NW, Archer SL, Defelice A, Evans S, Fiszman M, Martin T, Saulnier M, Rabinovitch M, Schermuly R, Stewart D, Truebel H, Walker G, Stenmark KR. Anticipated classes of new medications and molecular targets for pulmonary arterial hypertension. *Pulm Circ.* 2013;3: 226–44.
2. Reynolds AM, Holmes MD, Danilov SM, Reynolds PN. Targeted gene delivery of BMPR2 attenuates pulmonary hypertension. *Eur Respir J.* 2012;39:329–43.
3. Spiekerkoetter E, Tian X, Cai J, Hopper RK, Sudheendra D, Li CG, El-Bizri N, Sawada H, Haghghat R, Chan R, Haghghat L, de Jesus Perez V, Wang L, Reddy S, Zhao M, Bernstein D, Solow-Cordero DE, Beachy PA, Wandless TJ, Ten Dijke P, Rabinovitch M. FK506 activates BMPR2, rescues endothelial dysfunction, and reverses pulmonary hypertension. *J Clin Invest.* 2013;123:3600–13.

4. Peltz SW, Morsy M, Welch EM, Jacobson A. Ataluren as an agent for therapeutic nonsense suppression. *Annu Rev Med.* 2013;64:407–25.
5. Kerem E, Konstan MW, De Boeck K, Accurso FJ, Sermet-Gaudelus I, Wilschanski M, Elborn JS, Melotti P, Bronsveld I, Fajac I, Malfroot A, Rosenbluth DB, Walker PA, McColley SA, Knoop C, Quattrucci S, Rietschel E, Zeitlin PL, Barth J, Elfring GL, Welch EM, Branstrom A, Spiegel RJ, Peltz SW, Ajayi T, Rowe SM. Ataluren for the treatment of nonsense-mutation cystic fibrosis: a randomised, double-blind, placebo-controlled phase 3 trial. *Lancet Respir Med.* 2014;2:539–47.
6. Frump AL, Lowery JW, Hamid R, Austin ED, de Caestecker M. Abnormal trafficking of endogenously expressed BMPR2 mutant allelic products in patients with heritable pulmonary arterial hypertension. *PLoS One.* 2013;8:e80319.
7. Poirier O, Ciumas M, Eyries M, Montagne K, Nadaud S, Soubrier F. Inhibition of apelin expression by BMP signaling in endothelial cells. *Am J Physiol Cell Physiol.* 2012;303:C1139–45.
8. Alastalo TP, Li M, Perez Vde J, Pham D, Sawada H, Wang JK, Koskenvuo M, Wang L, Freeman BA, Chang HY, Rabinovitch M. Disruption of PPARgamma/beta-catenin-mediated regulation of apelin impairs BMP-induced mouse and human pulmonary arterial EC survival. *J Clin Invest.* 2011;121:3735–46.
9. Hansmann G, de Jesus Perez VA, Alastalo TP, Alvira CM, Guignabert C, Bekker JM, Schellong S, Urashima T, Wang L, Morrell NW, Rabinovitch M. An antiproliferative BMP-2/PPARgamma/apoE axis in human and murine SMCs and its role in pulmonary hypertension. *J Clin Invest.* 2008;118:1846–57.
10. Kim EK, Lee JH, Oh YM, Lee YS, Lee SD. Rosiglitazone attenuates hypoxia-induced pulmonary arterial hypertension in rats. *Respirology (Carlton, Vic).* 2010;15:659–68.
11. Yuan K, Orcholski M, Tian X, Liao X, de Jesus Perez VA. MicroRNAs: promising therapeutic targets for the treatment of pulmonary arterial hypertension. *Expert Opin Ther Targets.* 2013;17:557–64.
12. Archer SL, Marsboom G, Kim GH, Zhang HJ, Toth PT, Svensson EC, Dyck JR, Gombert-Maitland M, Thebaud B, Husain AN, Cipriani N, Rehman J. Epigenetic attenuation of mitochondrial superoxide dismutase 2 in pulmonary arterial hypertension: a basis for excessive cell proliferation and a new therapeutic target. *Circulation.* 2010;121:2661–71.
13. Kim GH, Ryan JJ, Marsboom G, Archer SL. Epigenetic mechanisms of pulmonary hypertension. *Pulm Circ.* 2011;1:347–56.
14. Saco TV, Parthasarathy PT, Cho Y, Lockey RF, Kolliputi N. Role of epigenetics in pulmonary hypertension. *Am J Physiol Cell Physiol.* 2014;306:C1101–5.
15. Bogaard HJ, Mizuno S, Hussaini AA, Toldo S, Abbate A, Kraskauskas D, Kasper M, Natarajan R, Voelkel NF. Suppression of histone deacetylases worsens right ventricular dysfunction after pulmonary artery banding in rats. *Am J Respir Crit Care Med.* 2011;183:1402–10.
16. Cho YK, Eom GH, Kee HJ, Kim HS, Choi WY, Nam KI, Ma JS, Kook H. Sodium valproate, a histone deacetylase inhibitor, but not captopril, prevents right ventricular hypertrophy in rats. *Circ J.* 2010;74:760–70.
17. Yu Y, Keller SH, Remillard CV, Safrina O, Nicholson A, Zhang SL, Jiang W, Vangala N, Landsberg JW, Wang JY, Thistlethwaite PA, Channick RN, Robbins IM, Loyd JE, Ghofrani HA, Grimminger F, Schermuly RT, Cahalan MD, Rubin LJ, Yuan JX. A functional single-nucleotide polymorphism in the TRPC6 gene promoter associated with idiopathic pulmonary arterial hypertension. *Circulation.* 2009;119:2313–22.
18. Yu Y, Fantozzi I, Remillard CV, Landsberg JW, Kunichika N, Platoshyn O, Tigno DD, Thistlethwaite PA, Rubin LJ, Yuan JX. Enhanced expression of transient receptor potential channels in idiopathic pulmonary arterial hypertension. *Proc Natl Acad Sci U S A.* 2004;101:13861–6.
19. Alzoubi A, Almalouf P, Toba M, O'Neill K, Qian X, Francis M, Taylor MS, Alexeyev M, McMurtry IF, Oka M, Stevens T. TRPC4 inactivation confers a survival benefit in severe pulmonary arterial hypertension. *Am J Pathol.* 2013;183:1779–88.

20. Makanga M, Dewachter C, Maruyama H, Vuckovic A, Rondelet B, Naeije R, Dewachter L. Downregulated bone morphogenetic protein signaling in nitrofen-induced congenital diaphragmatic hernia. *Pediatr Surg Int*. 2013;29:823–34.
21. Kuhr FK, Smith KA, Song MY, Levitan I, Yuan JX. New mechanisms of pulmonary arterial hypertension: role of Ca(2+)-signaling. *Am J Physiol Heart Circ Physiol*. 2012;302:H1546–62.
22. Park WS, Firth AL, Han J, Ko EA. Patho-, physiological roles of voltage-dependent K<sup>+</sup> channels in pulmonary arterial smooth muscle cells. *J Smooth Muscle Res*. 2010;46:89–105.
23. Ma L, Roman-Campos D, Austin ED, Eyries M, Sampson KS, Soubrier F, Germain M, Tregouet DA, Borczuk A, Rosenzweig EB, Girerd B, Montani D, Humbert M, Loyd JE, Kass RS, Chung WK. A novel channelopathy in pulmonary arterial hypertension. *N Engl J Med*. 2013;369:351–61.
24. Said SI, Hamidi SA. Pharmacogenomics in pulmonary arterial hypertension: toward a mechanistic, target-based approach to therapy. *Pulm Circ*. 2011;1:383–8.
25. Galiè N, Boonstra A, Ewert R, et al. Effects of inhaled aviptadil (vasoactive intestinal peptide) in patients with pulmonary arterial hypertension (PAH). *Am J Respir Crit Care Med*. 2010;181:A2516.
26. Dumitrascu R, Kulcke C, Konigshoff M, Kouri F, Yang X, Morrell N, Ghofrani HA, Weissmann N, Reiter R, Seeger W, Grimminger F, Eickelberg O, Schermuly RT, Pullamsetti SS. Terguride ameliorates monocrotaline-induced pulmonary hypertension in rats. *Eur Respir J*. 2011;37:1104–18.
27. Ghofrani HA, Al-Hiti H, Vonk Noordegraaf A, et al. Proof-of-concept study to investigate the efficacy, hemodynamics and tolerability of terguride vs. placebo in subjects with pulmonary arterial hypertension: results of a double blind, randomised, prospective phase IIa study. *Am J Respir Crit Care Med*. 2012;185:A2496.
28. Andersen CU, Hilberg O, Mellekjaer S, Nielsen-Kudsk JE, Simonsen U. Apelin and pulmonary hypertension. *Pulm Circ*. 2011;1:334–46.
29. McMurtry IF, Abe K, Ota H, Fagan KA, Oka M. Rho kinase-mediated vasoconstriction in pulmonary hypertension. *Adv Exp Med Biol*. 2010;661:299–308.
30. Oka M, Fagan KA, Jones PL, McMurtry IF. Therapeutic potential of RhoA/Rho kinase inhibitors in pulmonary hypertension. *Br J Pharmacol*. 2008;155:444–54.
31. Fagan KA, Oka M, Bauer NR, Gebb SA, Ivy DD, Morris KG, McMurtry IF. Attenuation of acute hypoxic pulmonary vasoconstriction and hypoxic pulmonary hypertension in mice by inhibition of Rho-kinase. *Am J Physiol Lung Cell Mol Physiol*. 2004;287:L656–64.
32. Nagaoka T, Fagan KA, Gebb SA, Morris KG, Suzuki T, Shimokawa H, McMurtry IF, Oka M. Inhaled Rho kinase inhibitors are potent and selective vasodilators in rat pulmonary hypertension. *Am J Respir Crit Care Med*. 2005;171:494–9.
33. Nagaoka T, Gebb SA, Karoor V, Homma N, Morris KG, McMurtry IF, Oka M. Involvement of RhoA/Rho kinase signaling in pulmonary hypertension of the fawn-hooded rat. *J Appl Physiol* (Bethesda, Md: 1985). 2006;100:996–1002.
34. Nagaoka T, Morio Y, Casanova N, Bauer N, Gebb S, McMurtry I, Oka M. Rho/Rho kinase signaling mediates increased basal pulmonary vascular tone in chronically hypoxic rats. *Am J Physiol Lung Cell Mol Physiol*. 2004;287:L665–72.
35. Oka M, Homma N, McMurtry IF. Rho kinase-mediated vasoconstriction in rat models of pulmonary hypertension. *Methods Enzymol*. 2008;439:191–204.
36. Oka M, Homma N, Taraseviciene-Stewart L, Morris KG, Kraskauskas D, Burns N, Voelkel NF, McMurtry IF. Rho kinase-mediated vasoconstriction is important in severe occlusive pulmonary arterial hypertension in rats. *Circ Res*. 2007;100:923–9.
37. Fujita H, Fukumoto Y, Saji K, Sugimura K, Demachi J, Nawata J, Shimokawa H. Acute vasodilator effects of inhaled fasudil, a specific Rho-kinase inhibitor, in patients with pulmonary arterial hypertension. *Heart Vessels*. 2010;25:144–9.
38. Fukumoto Y, Matoba T, Ito A, Tanaka H, Kishi T, Hayashidani S, Abe K, Takeshita A, Shimokawa H. Acute vasodilator effects of a Rho-kinase inhibitor, fasudil, in patients with severe pulmonary hypertension. *Heart (British Cardiac Society)*. 2005;91:391–2.

39. Fukumoto Y, Yamada N, Matsubara H, Mizoguchi M, Uchino K, Yao A, Kihara Y, Kawano M, Watanabe H, Takeda Y, Adachi T, Osanai S, Tanabe N, Inoue T, Kubo A, Ota Y, Fukuda K, Nakano T, Shimokawa H. Double-blind, placebo-controlled clinical trial with a rho-kinase inhibitor in pulmonary arterial hypertension. *Circ J*. 2013;77:2619–25.
40. Ishikura K, Yamada N, Ito M, Ota S, Nakamura M, Isaka N, Nakano T. Beneficial acute effects of rho-kinase inhibitor in patients with pulmonary arterial hypertension. *Circ J*. 2006;70:174–8.
41. Alzoubi A, Toba M, Abe K, O'Neill KD, Rocic P, Fagan KA, McMurtry IF, Oka M. Dehydroepiandrosterone restores right ventricular structure and function in rats with severe pulmonary arterial hypertension. *Am J Physiol Heart Circ Physiol*. 2013;304:H1708–18.
42. Dumas de La Roque E, Savineau JP, Metivier AC, Billes MA, Kraemer JP, Doutreleau S, Jougon J, Marthan R, Moore N, Fayon M, Baulieu EE, Dromer C. Dehydroepiandrosterone (DHEA) improves pulmonary hypertension in chronic obstructive pulmonary disease (COPD): a pilot study. *Ann Endocrinol*. 2012;73:20–5.
43. El Chami H, Hassoun PM. Immune and inflammatory mechanisms in pulmonary arterial hypertension. *Prog Cardiovasc Dis*. 2012;55:218–28.
44. Graham BB, Chabon J, Gebreab L, Poole J, Debella E, Davis L, Tanaka T, Sanders L, Dropcho N, Bandeira A, Vandivier RW, Champion HC, Butrous G, Wang XJ, Wynn TA, Tuder RM. Transforming growth factor-beta signaling promotes pulmonary hypertension caused by *Schistosoma mansoni*. *Circulation*. 2013;128:1354–64.
45. Groth A, Vrugt B, Brock M, Speich R, Ulrich S, Huber LC. Inflammatory cytokines in pulmonary hypertension. *Respir Res*. 2014;15:47.
46. Furuya Y, Satoh T, Kuwana M. Interleukin-6 as a potential therapeutic target for pulmonary arterial hypertension. *Int J Rheumatol*. 2010;2010:720305.
47. Taraseviciene-Stewart L, Nicolls MR, Kraskauskas D, Scerbavicius R, Burns N, Cool C, Wood K, Parr JE, Boackle SA, Voelkel NF. Absence of T cells confers increased pulmonary arterial hypertension and vascular remodeling. *Am J Respir Crit Care Med*. 2007;175:1280–9.
48. Tcherakian C, Couderc LJ, Humbert M, Godot V, Sitbon O, Devillier P. Inflammatory mechanisms in HIV-associated pulmonary arterial hypertension. *Semin Respir Crit Care Med*. 2013;34:645–53.
49. Hennigan S, Channick RN, Silverman GJ. Rituximab treatment of pulmonary arterial hypertension associated with systemic lupus erythematosus: a case report. *Lupus*. 2008;17:754–6.
50. Padilla-Ibarra J, Sanchez-Ortiz A, Sandoval-Castro C, Ramos-Remus C. Rituximab treatment for pulmonary arterial hypertension in adult-onset Still's disease. *Clin Exp Rheumatol*. 2013;31:657–8.
51. Perros F, Montani D, Dorfmüller P, Durand-Gasselín I, Tcherakian C, Le Pavec J, Mazmanian M, Fadel E, Mussot S, Mercier O, Herve P, Emilie D, Eddahibi S, Simonneau G, Souza R, Humbert M. Platelet-derived growth factor expression and function in idiopathic pulmonary arterial hypertension. *Am J Respir Crit Care Med*. 2008;178:81–8.
52. Nakamura K, Akagi S, Ogawa A, Kusano KF, Matsubara H, Miura D, Fuke S, Nishii N, Nagase S, Kohno K, Morita H, Oto T, Yamanaka R, Otsuka F, Miura A, Yutani C, Ohe T, Ito H. Pro-apoptotic effects of imatinib on PDGF-stimulated pulmonary artery smooth muscle cells from patients with idiopathic pulmonary arterial hypertension. *Int J Cardiol*. 2012;159:100–6.
53. Schermuly RT, Dony E, Ghofrani HA, Pullamsetti S, Savai R, Roth M, Sydykov A, Lai YJ, Weissmann N, Seeger W, Grimminger F. Reversal of experimental pulmonary hypertension by PDGF inhibition. *J Clin Invest*. 2005;115:2811–21.
54. Ghofrani HA, Seeger W, Grimminger F. Imatinib for the treatment of pulmonary arterial hypertension. *N Engl J Med*. 2005;353:1412–3.
55. Hoeper MM, Barst RJ, Bourge RC, Feldman J, Frost AE, Galie N, Gomez-Sanchez MA, Grimminger F, Grunig E, Hassoun PM, Morrell NW, Peacock AJ, Satoh T, Simonneau G, Tapson VF, Torres F, Lawrence D, Quinn DA, Ghofrani HA. Imatinib mesylate as add-on therapy for pulmonary arterial hypertension: results of the randomized IMPRES study. *Circulation*. 2013;127:1128–38.

56. Montani D, Bergot E, Gunther S, Savale L, Bergeron A, Bourdin A, Bouvaist H, Canuet M, Pison C, Macro M, Poubreau P, Girerd B, Natali D, Guignabert C, Perros F, O'Callaghan DS, Jais X, Tubert-Bitter P, Zalcman G, Sitbon O, Simonneau G, Humbert M. Pulmonary arterial hypertension in patients treated by dasatinib. *Circulation*. 2012;125:2128–37.
57. Dahal BK, Cornitescu T, Tretyn A, Pullamsetti SS, Kosanovic D, Dumitrascu R, Ghofrani HA, Weissmann N, Voswinckel R, Banat GA, Seeger W, Grimminger F, Schermuly RT. Role of epidermal growth factor inhibition in experimental pulmonary hypertension. *Am J Respir Crit Care Med*. 2010;181:158–67.
58. Gombert-Maitland M, Maitland ML, Barst RJ, Sugeng L, Coslet S, Perrino TJ, Bond L, Lacouture ME, Archer SL, Ratain MJ. A dosing/cross-development study of the multikinase inhibitor sorafenib in patients with pulmonary arterial hypertension. *Clin Pharmacol Ther*. 2010;87:303–10.
59. Godinas L, Guignabert C, Seferian A, Perros F, Bergot E, Sibille Y, Humbert M, Montani D. Tyrosine kinase inhibitors in pulmonary arterial hypertension: a double-edge sword? *Semin Respir Crit Care Med*. 2013;34:714–24.
60. Cowan KN, Jones PL, Rabinovitch M. Elastase and matrix metalloproteinase inhibitors induce regression, and tenascin-C antisense prevents progression, of vascular disease. *J Clin Invest*. 2000;105:21–34.
61. Kim YM, Haghghat L, Spiekerkoetter E, Sawada H, Alvira CM, Wang L, Acharya S, Rodriguez-Colon G, Orton A, Zhao M, Rabinovitch M. Neutrophil elastase is produced by pulmonary artery smooth muscle cells and is linked to neointimal lesions. *Am J Pathol*. 2011;179:1560–72.
62. Zaidi SH, You XM, Ciura S, Husain M, Rabinovitch M. Overexpression of the serine elastase inhibitor elafin protects transgenic mice from hypoxic pulmonary hypertension. *Circulation*. 2002;105:516–21.
63. Gomez-Arroyo J, Mizuno S, Szczepanek K, Van Tassell B, Natarajan R, dos Remedios CG, Drake JI, Farkas L, Kraskauskas D, Wijesinghe DS, Chalfant CE, Bigbee J, Abbate A, Lesnefsky EJ, Bogaard HJ, Voelkel NF. Metabolic gene remodeling and mitochondrial dysfunction in failing right ventricular hypertrophy secondary to pulmonary arterial hypertension. *Circ Heart Fail*. 2013;6:136–44.
64. Fessel JP, Hamid R, Wittmann BM, Robinson LJ, Blackwell T, Tada Y, Tanabe N, Tatsumi K, Hemnes AR, West JD. Metabolomic analysis of bone morphogenetic protein receptor type 2 mutations in human pulmonary endothelium reveals widespread metabolic reprogramming. *Pulm Circ*. 2012;2:201–13.
65. Archer SL, Fang YH, Ryan JJ, Piao L. Metabolism and bioenergetics in the right ventricle and pulmonary vasculature in pulmonary hypertension. *Pulm Circ*. 2013;3:144–52.
66. Guignabert C, Tu L, Izikki M, Dewachter L, Zadigue P, Humbert M, Adnot S, Fadel E, Eddahibi S. Dichloroacetate treatment partially regresses established pulmonary hypertension in mice with SM22alpha-targeted overexpression of the serotonin transporter. *FASEB J*. 2009;23:4135–47.
67. Li B, Yan J, Shen Y, Liu Y, Ma Z. Dichloroacetate prevents but not reverses the formation of neointimal lesions in a rat model of severe pulmonary arterial hypertension. *Mol Med Rep*. 2014;10:2144–52.
68. McMurtry MS, Bonnet S, Wu X, Dyck JR, Haromy A, Hashimoto K, Michelakis ED. Dichloroacetate prevents and reverses pulmonary hypertension by inducing pulmonary artery smooth muscle cell apoptosis. *Circ Res*. 2004;95:830–40.
69. Piao L, Sidhu VK, Fang YH, Ryan JJ, Parikh KS, Hong Z, Toth PT, Morrow E, Kutty S, Lopaschuk GD, Archer SL. FOXO1-mediated upregulation of pyruvate dehydrogenase kinase-4 (PDK4) decreases glucose oxidation and impairs right ventricular function in pulmonary hypertension: therapeutic benefits of dichloroacetate. *J Mol Med (Berlin, Germany)*. 2013;91:333–46.
70. Fang YH, Piao L, Hong Z, Toth PT, Marsboom G, Bache-Wiig P, Rehman J, Archer SL. Therapeutic inhibition of fatty acid oxidation in right ventricular hypertrophy: exploiting Randle's cycle. *J Mol Med (Berlin, Germany)*. 2012;90:31–43.

71. Sutendra G, Bonnet S, Rochefort G, Haromy A, Folmes KD, Lopaschuk GD, Dyck JR, Michelakis ED. Fatty acid oxidation and malonyl-CoA decarboxylase in the vascular remodeling of pulmonary hypertension. *Sci Transl Med.* 2010;2:4458.
72. de Man FS, Handoko ML, Guignabert C, Bogaard HJ, Vonk-Noordegraaf A. Neurohormonal axis in patients with pulmonary arterial hypertension: friend or foe? *Am J Respir Crit Care Med.* 2013;187:14–9.
73. Mak S, Witte KK, Al-Hesayen A, Granton JJ, Parker JD. Cardiac sympathetic activation in patients with pulmonary arterial hypertension. *Am J Physiol Regul Integr Comp Physiol.* 2012;302:R1153–7.
74. Bristow MR, Minobe W, Rasmussen R, Larrabee P, Skerl L, Klein JW, Anderson FL, Murray J, Mestroni L, Karwande SV, et al. Beta-adrenergic neuroeffector abnormalities in the failing human heart are produced by local rather than systemic mechanisms. *J Clin Invest.* 1992;89:803–15.
75. Ciarka A, Doan V, Velez-Roa S, Naeije R, van de Borne P. Prognostic significance of sympathetic nervous system activation in pulmonary arterial hypertension. *Am J Respir Crit Care Med.* 2010;181:1269–75.
76. Bogaard HJ, Natarajan R, Mizuno S, Abbate A, Chang PJ, Chau VQ, Hoke NN, Kraskauskas D, Kasper M, Salloum FN, Voelkel NF. Adrenergic receptor blockade reverses right heart remodeling and dysfunction in pulmonary hypertensive rats. *Am J Respir Crit Care Med.* 2010;182:652–60.
77. de Man FS, Handoko ML, van Ballegoij JJ, Schalij I, Bogaards SJ, Postmus PE, van der Velden J, Westerhof N, Paulus WJ, Vonk-Noordegraaf A. Bisoprolol delays progression towards right heart failure in experimental pulmonary hypertension. *Circ Heart Fail.* 2012;5:97–105.
78. Drake JJ, Gomez-Arroyo J, Dumur CI, Kraskauskas D, Natarajan R, Bogaard HJ, Fawcett P, Voelkel NF. Chronic carvedilol treatment partially reverses the right ventricular failure transcriptional profile in experimental pulmonary hypertension. *Physiol Genomics.* 2013;45:449–61.
79. So PP, Davies RA, Chandy G, Stewart D, Beanlands RS, Haddad H, Pugliese C, Mielniczuk LM. Usefulness of beta-blocker therapy and outcomes in patients with pulmonary arterial hypertension. *Am J Cardiol.* 2012;109:1504–9.
80. Thenappan T, Roy SS, Duval S, Glassner-Kolmin C, Gomberg-Maitland M. Beta-blocker therapy is not associated with adverse outcomes in patients with pulmonary arterial hypertension: a propensity score analysis. *Circulation.* 2014. doi:[10.1161/CIRCHEARTFAILURE.114.001429](https://doi.org/10.1161/CIRCHEARTFAILURE.114.001429).
81. de Man FS, Tu L, Handoko ML, Rain S, Ruiter G, Francois C, Schalij I, Dorfmueller P, Simonneau G, Fadel E, Perros F, Boonstra A, Postmus PE, van der Velden J, Vonk-Noordegraaf A, Humbert M, Eddahibi S, Guignabert C. Dysregulated renin-angiotensin-aldosterone system contributes to pulmonary arterial hypertension. *Am J Respir Crit Care Med.* 2012;186:780–9.
82. Alpert MA, Pressly TA, Mukerji V, Lambert CR, Mukerji B. Short- and long-term hemodynamic effects of captopril in patients with pulmonary hypertension and selected connective tissue disease. *Chest.* 1992;102:1407–12.
83. Hampl V, Herget J, Bíbová J, Baňasová A, Husková Z, Vaňourková Z, Jířchová Š, Kujal P, Vernerová Z, Sadowski J, Červenka L. Intrapulmonary activation of the angiotensin-converting enzyme type 2/angiotensin 1-7/G-protein-coupled Mas receptor axis attenuates pulmonary hypertension in Ren-2 transgenic rats exposed to chronic hypoxia. *Physiol Res.* 2015; 64:25–38.
84. Li G, Liu Y, Zhu Y, Liu A, Xu Y, Li X, Li Z, Su J, Sun L. ACE2 activation confers endothelial protection and attenuates neointimal lesions in prevention of severe pulmonary arterial hypertension in rats. *Lung.* 2013;191:327–36.
85. Shenoy V, Ferreira AJ, Qi Y, Fraga-Silva RA, Diez-Freire C, Dooies A, Jun JY, Sriramula S, Mariappan N, Pourang D, Venugopal CS, Francis J, Reudelhuber T, Santos RA, Patel JM, Raizada MK, Katovich MJ. The angiotensin-converting enzyme 2/angiogenesis-(1-7)/Mas axis confers cardiopulmonary protection against lung fibrosis and pulmonary hypertension. *Am J Respir Crit Care Med.* 2010;182:1065–72.

86. D'Elia E, Krum H. Mineralcorticoid antagonists in heart failure. *Heart Fail Clin.* 2014;10:559–64.
87. Maron BA, Opotowsky AR, Landzberg MJ, Loscalzo J, Waxman AB, Leopold JA. Plasma aldosterone levels are elevated in patients with pulmonary arterial hypertension in the absence of left ventricular heart failure: a pilot study. *Eur J Heart Fail.* 2013;15:277–83.
88. Maron BA, Zhang YY, White K, Chan SY, Handy DE, Mahoney CE, Loscalzo J, Leopold JA. Aldosterone inactivates the endothelin-B receptor via a cysteinyl thiol redox switch to decrease pulmonary endothelial nitric oxide levels and modulate pulmonary arterial hypertension. *Circulation.* 2012;126:963–74.
89. Maron BA, Leopold JA. The role of the renin-angiotensin-aldosterone system in the pathobiology of pulmonary arterial hypertension (2013 Grover Conference series). *Pulm Circ.* 2014;4:200–10.
90. Maron BA, Waxman AB, Opotowsky AR, Gillies H, Blair C, Aghamohammadzadeh R, Loscalzo J, Leopold JA. Effectiveness of spironolactone plus ambrisentan for treatment of pulmonary arterial hypertension (from the [ARIES] study 1 and 2 trials). *Am J Cardiol.* 2013;112:720–5.
91. Urakami T, Jarvinen TA, Toba M, Sawada J, Ambalavanan N, Mann D, McMurtry I, Oka M, Ruoslahti E, Komatsu M. Peptide-directed highly selective targeting of pulmonary arterial hypertension. *Am J Pathol.* 2011;178:2489–95.
92. Suen CM, Mei SH, Kugathasan L, Stewart DJ. Targeted delivery of genes to endothelial cells and cell- and gene-based therapy in pulmonary vascular diseases. *Compr Physiol.* 2013;3:1749–79.
93. Yang JX, Pan YY, Zhao YY, Wang XX. Endothelial progenitor cell-based therapy for pulmonary arterial hypertension. *Cell Transplant.* 2013;22:1325–36.
94. Asosingh K, Aldred MA, Vasanji A, Drazba J, Sharp J, Farver C, Comhair SA, Xu W, Licina L, Huang L, Anand-Apte B, Yoder MC, Tudor RM, Erzurum SC. Circulating angiogenic precursors in idiopathic pulmonary arterial hypertension. *Am J Pathol.* 2008;172:615–27.
95. Zhao YD, Courtman DW, Deng Y, Kugathasan L, Zhang Q, Stewart DJ. Rescue of monocrotaline-induced pulmonary arterial hypertension using bone marrow-derived endothelial-like progenitor cells: efficacy of combined cell and eNOS gene therapy in established disease. *Circ Res.* 2005;96:442–50.
96. Wang XX, Zhang FR, Shang YP, Zhu JH, Xie XD, Tao QM, Zhu JH, Chen JZ. Transplantation of autologous endothelial progenitor cells may be beneficial in patients with idiopathic pulmonary arterial hypertension: a pilot randomized controlled trial. *J Am Coll Cardiol.* 2007;49:1566–71.