

Chapter 6

Immunology of Autism

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Abstract Autism spectrum disorders (ASD) are developmental disorders characterized by behavioral deficits in verbal and nonverbal communication as well as social interactions, and are accompanied by repetitive or stereotyped behaviors and interests. Numerous studies over the last forty years have recognized altered immune responses in individuals with ASD; concurrently basic research has highlighted the myriad of neuroimmune interactions and the cross talk that occurs between nervous and immune systems. Neuroinflammation, particularly in the cerebellum, has been found in post mortem brain tissues from individuals with ASD and is characterized by the presence of profound glia activation processes. This and altered gene expression profiles indicating perturbed immune suggest a contributing role for immunological systems in the pathology of ASD. Peripheral immune abnormalities have also been found; shifts in both direction of TH1 and TH2 skewing have been reported as well as autoantibody production, increased NK cell activation, T cell responses and monocyte cell function overwhelmingly suggesting the presence of immune dysfunction in individuals with ASD. Many of these findings are associated with worsening behavioral scores, suggesting treatment of immune function could be useful in alleviating symptoms associated with ASD. Immune activation *in utero* is also associated with an increased risk of the child for having a diagnosis of ASD, where increased cytokine production in the offspring is directly linked to changes in offspring behavior. In addition to peripheral changes, brain and CSF immune variations in ASD are reported as well as an increase in gastrointestinal/mucosal dysfunction which has led to an increased interest in exploring the gut-brain-immune connections and its role in ASD. Further research in neuroimmune interactions may bring further insight and elicit new therapeutic tools for ASD.

Keywords Adaptive immune system · Autism spectrum disorders · Behavior · Cerebellum · Cytokine · Innate immune system · Immunity · Maternal immune activation · Neuroimmunology · Social interactions · T cells · Gastrointestinal

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6.1 Immune-Brain Interactions

6.1.1 Cytokines and Neurodevelopment

The immune system is involved in three main areas—surveillance of foreign antigen, fighting infections and tissue remodeling and participates in these functions in all types of tissue during development, health, disease and wound healing. The brain is no exception; many immune responses take part in shaping and maintaining normal central nervous system (CNS) function (Fig. 6.1). During the course of neurodevelopment and throughout adulthood, normal immunological processes take part in influencing appropriate neurological features and functions. Cytokines, the immune system's signaling molecules have a significant role in both of these areas. Cytokines act as both chemoattractants, guiding direction of growth and migration as well as acting as neurotrophic factors that promote survival to developing neurons in the brain and spinal cord (Deverman and Patterson 2009). Interleukin (IL)-1 β a prominent inflammatory cytokine of the immune system has been found to be involved in many different functions of the CNS. During development, IL-1 β expression is observed in the embryonic spinal cord of both chickens (stage 17-HH) and rats (embryonic day 12) (de la Mano et al. 2007). Delivery of IL-1 β on microbeads implanted near the spinal cord of chick embryos increased the number of proliferating (BrdU⁺) neuroepithelial cells in the dorsal spinal cord and led to a reduction in the ventral spinal cord. Blocking IL-1 β through anti-IL-1 β antibodies reduced BrdU incorporation in the dorsal spinal cord. (de la Mano et al. 2007). In the mammalian adult brain, cytokines including IL-1 β can act on the neural and progenitor stem cells in response to injury, disease and stress influencing proliferation and neurogenesis. (Carpentier and Palmer 2009). IL-1 β is also believed to be required for normal learning and memory processes in the hippocampus (Goshen et al. 2007). Gene expression of IL-1 β has been found to be increased in the hippocampus 24 h after contextual learning and blocking IL-1 β results in impairments in spatial memory and fear conditioning tests (Goshen et al. 2007). IL-1 receptor knockout mice were found to have a reduction in neuronal dendritic spine size which may contribute to the defects seen in memory in these mice (Goshen et al. 2009).

Other pro-inflammatory cytokines that are involved in spatial learning and memory include both IL-6 and Tumor necrosis factor (TNF)- α ; however, these cytokines also have complex roles in shaping memory and learning and have reported beneficial and detrimental effects. A recent study showed that TNF- α signaling through the NF κ B pathway lead to increased neural stem cell (NSC) proliferation. This proliferation was attributed to the activation of the IKK- β and NF κ B pathway leading to up regulation of cyclin D1 (Widera et al. 2006). How cytokines affect the CNS is context dependent and environments surrounding the release of these cytokines play an equally important role in determining the ultimate effect than any single cytokine will play (Yirmiya and Goshen 2011). Transforming growth factor (TGF)- β , an important immune regulatory cytokine, is involved in signaling required for mouse

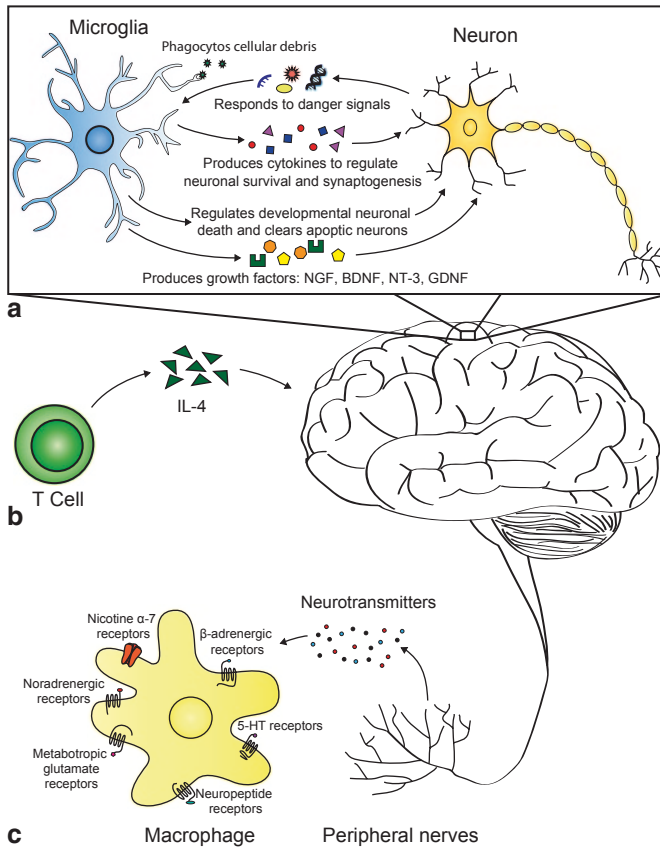


Fig. 6.1 a) Many immunological processes take part both in developing and maintaining the CNS. Microglia are the main contributor to many of these processes. As the resident phagocytic cell, microglia clear cellular debris and apoptotic neurons. They also further neural development by producing various growth factors and many cytokines that regulate neuronal survival and synaptogenesis. Microglia also respond to danger signals by producing reactive oxygen species and pro-inflammatory cytokines. Some of the cytokines that can affect the CNS include: IL-1 β , TNF- α , IL-6 and TGF- β . These cytokines not only contribute to neuronal development but also play a role in learning and memory. **b)** Lymphocytes, part of the adaptive immune system have also been found to contribute to cognitive function. More specifically T cells have been demonstrated to contribute to visuospatial learning and enhance neurogenesis in the dentate gyrus. T cells important to cognitive function are believed to be found in the meningeal spaces where they produce cytokines; specifically IL-4 has been associated with visuospatial learning. **c)** Outside the CNS, neuro-immune interactions also take place. Many immune cells including macrophages express receptors for neurotransmitters. Some of these receptors include beta adrenergic, nicotine α -7, noradrenergic, metabotropic glutamate, neuropeptide and 5-hydroxytryptophan (5-HT) receptors. Signaling through these receptors can either enhance or suppress the immune response depending on the neurotransmitter and environmental conditions. Norepinephrine, for example, generally tends to have an inhibitory effect on pro-inflammatory cytokine production in macrophages but this can be reversed if other factors such as LPS are present. *NGF* nerve growth factor, *BDNF* brain derived neurotrophic factor, *NT-3* neurotrophin-3, *GDNF*, Glial cell line-derived neurotrophic factor, *IL* interleukin, *5-HT* 5-hydroxytryptophan

mesencephalic progenitors to differentiate into tyrosine hydroxylase (TH)⁺dopaminergic neurons *in vitro* and *in vivo* (Roussa et al. 2006). When TGF β 2/TGF β 3 double knockout mice were examined it was found they have reduced numbers of TH⁺ neurons in the ventral mesencephalon, however, in the locus coeruleus TH⁺ neurons were not significantly different from controls indicating that while TGF β signaling may be important in the ventral mesencephalon it does not seem to contribute to ventral midbrain dopaminergic neuron development (Roussa et al. 2006).

6.1.2 Microglia

Microglia, the brain's resident phagocytic cells, play a central role in CNS development and maintenance through regulating developmental neuronal death and the clearing of apoptotic neurons (Wakselman et al. 2008; Takahashi et al. 2005). They also phagocytose cellular debris and respond to 'danger' signals through production of reactive oxygen species (ROS) and inflammatory cytokines (Neumann et al. 2009; Ron-Harel et al. 2011). The significance of this function is illustrated through the depletion of microglia from murine neonatal cerebellar slice cultures. This specific elimination of microglia leads to increased Purkinje cell survival due to the reduction of phagocytosis of caspase-3-expressing Purkinje cells (Marin-Teva et al. 2004). More recently *in vivo* studies have shown that microglia regulate neurogenesis in the cerebral cortex of primates, rodents and human fetal tissues (Cunningham et al. 2013). Cunningham et al. (2013) show microglia enter and colonize the cortical proliferative zones near the end of neurogenesis and phagocytose neural precursor cells. Manipulation of microglia in rats either by suppressing microglia using doxycycline or activating them using injection of LPS resulted in increased and decreased numbers of neural precursors cells, respectively, further demonstrating the critical role microglia and the immune system play in regulating neuronal numbers in early brain development.

In general, microglia activation increases expression of inflammatory cytokines and can have toxic effects on the surrounding cells; however, they are also vital for the down regulation of immune responses through autocrine feedback loops and production of anti-inflammatory cytokines (Garden and Moller 2006). Embryonic microglia are known producers of TNF- α and are key regulators of developmental apoptosis and synaptogenesis. Disrupting TNF- α signaling through use of anti-TNF- α antibodies or soluble TNF- α receptor (TNFR1) results in the decrease of AMPA-type glutamate receptors on hippocampal neurons and thereby modulate synaptic strength in these neurons (Beattie et al. 2002). TNF- α has been shown to upregulate expression of β 3 integrins that help increase synaptic strength through stabilization of AMPARs (Cingolani et al. 2008). TNF- α is therefore suggested to have a central role for the homeostatic potentiation of synaptic strength during developmental synaptic refinement. Glial cells, in response to levels of activity in hippocampal cultures, have been shown to regulate TNF- α levels (Deverman and Patterson 2009).

Macrophage colony-stimulating factor (M-CSF) a growth factor for macrophages and microglia is necessary for proper development of certain areas of the brain. *M-CSF* mutant mice containing a null mutation in the M-CSF gene were shown to have auditory and visual processing impairment with failure of the newborn pups to respond to external cues and electrophysiologic abnormalities detected by intracortical recordings of brainstem auditory evoked potentials and visual evoked potentials (Michaelson et al. 1996). The effects of M-CSF are thought to be indirectly regulated by cytokine secreting microglia (Deverman and Patterson 2009). In addition to cytokines and chemokines, microglia produce many other regulatory and trophic factors promoting neuronal survival including: nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin (NT)-3, basic fibroblast growth factor, and glial-derived neurotrophic factor (GDNF) (Garden and Moller 2006). Microglia not only influence neuronal cells but can also be influenced by neuronal activity through neurotransmitter receptors such as glutamate receptors and by astrocytes through purinergic receptors such as P2X4, P2X7, P2Y2, P2Y6 and P2Y12 (Biber et al. 2007; Hung et al. 2010; Ferrari et al. 2006).

6.1.3 Cytokines, Immune Cells and Cognitive Function

Cytokines and immune cells are supportive in brain function including neurogenesis and cognitive functioning. Lymphocytes, for example, have been found to have a supportive role in cognitive functioning (Kipnis et al. 2012). Experiments with severe combined immune deficient (SCID) mice, that do not have any T or B cells, display impairments in hippocampal dependent spatial learning and memory through analysis of the Morris water maze behavioral test (MWM) (Kipnis et al. 2004). Furthermore, nude mice (lacking only mature T cells) display similar impairments assessed by MWM which could be partially rescued with replenishment of T cells from wild-type mice, demonstrating the role of T cells more specifically in areas of visuospatial learning (Kipnis et al. 2004; Ron-Harel et al. 2008; Brynskikh et al. 2008). To provide further support of the role of T cells in learning and memory another study looked at rats that were either raised under normal environmental conditions or under enriched conditions in which neurogenesis was enhanced in the dentate gyrus. To confirm the role of T cells, mice were used in which mono-specific T cells to either myelin basic protein (auto specific) (TMBP) or ovalbumin (OVA) (non-CNS specific) (TOVA) were used. TMBP mice were found to have higher amounts of proliferating neurons compared with controls and performed better in the MWM, while TOVA had less proliferating neurons than controls (Ziv et al. 2006). To emphasize the importance of T cell/microglia interactions in neurogenesis the TMBP mice were treated with a microglia blocking drug, minocycline, which significantly decreased neurogenesis in the dentate gyrus (Ziv et al. 2006). These findings support the role of T cells mediating neurogenesis and spatial learning through possible interaction with microglia.

Although T cells are not normally found in the CNS parenchyma under normal conditions, T-cell based support of behavioral plasticity is thought to take place in the meningeal spaces (Derecki et al. 2010; Schwartz and Shechter 2010). Depletion of T cells from the meningeal spaces results in impairments of learning and memory based on MWM results. Of the meningeal T cell population, CD4⁺ IL-4⁺ T cells were found to be the most important for spatial learning and memory as IL-4 deficient mice showed defects in MWM (Derecki et al. 2010). Other studies have also used T cell manipulations to demonstrate improved learning and memory (Ron-Harel and Schwartz 2009; Ron-Harel et al. 2008). Manipulation of T cells in aged mice through bone marrow transplantation improved spatial memory in these animals compared to young animals and was increased when compared with non treated aged animals (Ron-Harel et al. 2008). It has been suggested that assessing T cell immunity in old age could be used as a predictor of potential future memory loss and enhancing T-cell immunity could benefit age associated memory loss (Ron-Harel and Schwartz 2009).

Major histocompatibility complex (MHC) molecules are important cell surface molecules that interact with T cells involved in host immunity. It was long thought that the neural cells were among the small list of cells that did not express this family of surface proteins; however, in the late 1990's it was found that not only are these important immune molecules present on neurons but they actually influence synapse plasticity (Elmer and McAllister 2012). Mice with deficient signaling of class I MHC displayed impaired synapse plasticity (Huh et al. 2000). Peptides from within the cell are presented in MHC molecules and MHC I: peptide complexes, in an immunological context are scanned by cytotoxic T cells to detect abnormal conditions such as the presence of a viral infection or tumor. In the CNS it is thought that similar roles of MHC molecules are employed in presenting peptides in order to regulate normal developmental elimination of inappropriate synaptic connections, although the mechanism remains elusive (Boulanger 2009).

Communication between the immune system and CNS is not one way. Signaling is multidirectional and information can also pass from the CNS to the immune system. Norepinephrine (NE) released from sympathetic nerve terminals can signal to macrophages through beta adrenergic receptors (Kin and Sanders 2006). In general NE seems to have inhibitory effects on pro-inflammatory cytokine production such as TNF- α , IL-1 β , and sometimes IL-6, based on data from splenic macrophages (Meltzer et al. 2004; Ignatowski et al. 1996; Nance and Sanders 2007). IL-6 production has been found to be both increased or decreased in response to NE depending on other signals and stimuli such as the presence of LPS (Nance and Sanders 2007). Other neuro-based receptors found on immune cells include noradrenergic receptors, nicotinic α -7 receptors, receptors for neuropeptides and hormones, metabotropic glutamate receptors (mGluRs) and receptors for monoamines serotonin and dopamine (Nance and Sanders 2007; Tracey 2002; Besedovsky and Rey 2007; Friedman and Irwin 1997; Pacheco et al. 2004). Signaling through these receptors can regulate and modulate immune function and may be important in response to stress and or in neuro-psychiatric disorders were imbalances in neuromodulators have been observed.

6.1.4 Neuroinflammation

While there are many beneficial roles of the immune system in CNS function, too much inflammation can be detrimental. Exposure to pathogens which activate immune responses to protect against infections can result in increased production of pro-inflammatory cytokines that contribute to sickness behavior, while both anxiety and depression have also been associated with inflammation (Irwin and Miller 2007). Pro-inflammatory cytokines including IL-1 β , TNF- α and IL-6 can act on the brain causing sickness behaviors ranging from loss of appetite, lethargy to irritability (Dantzer et al. 2008). In addition to inflammation or as a result of infection, events causing stress, injury and ageing can also induce these same inflammatory mediators (Yirmiya and Goshen 2011). In experiments where IL-1 β was injected intracerebroventricularly (i.c.v.) into the right lateral cerebral ventricle either 24 h or 1 h before training in the MWM, those rats injected for 1 h but not those injected 24 h before training showed impaired performance in spatial memory the next day (Oitzl et al. 1993) suggesting changes may be fast acting but also dose-dependent and transient. Furthermore, increased peripheral levels of IL-1 β following infection with *Legionella pneumophila* or by direct administration of IL-1 β daily also showed impaired spatial memory and learning in mice (Gibertini et al. 1995) suggesting there may be a conditioning effect with repeated prolonged exposure to cytokines. Transgenic mice that over express IL-1 β show impairments in spatial memory that are particularly restricted to hippocampal dependent memory (Hein et al. 2010; Moore et al. 2009). Introduction of LPS also increases hippocampal IL-1 levels and induce similar impairments to spatial learning and memory (Nguyen et al. 1998). However, other study designs with different regimens of IL-1 β administration did not show memory or learning impairments suggesting that the conditions and environmental factors contribute to memory and learning (Yirmiya and Goshen 2011). Additionally IL-1 β associated neuroinflammation is linked in ageing and may play a role in age associated memory loss (Krabbe et al. 2004). Caspase-1 inhibitors when administered to aged mice over time reduced hippocampal IL-1 β and helped to improve contextual memory (Gemma et al. 2005; Krabbe et al. 2004). In Alzheimer's disease increased levels of TNF- α , IL-6 and IL-1 β have been detected in the serum and cerebral spinal fluid (Akiyama et al. 2000; Shaftel et al. 2008). Activation of microglia have been found in Alzheimer and other neurological diseases such as Parkinson's disease, multiple sclerosis and acquired immune deficiency syndrome dementia complex (Kim and de Vellis 2005). As stated above, the immune system orchestrates a vital and delicate balancing act necessary for the proper development and maintenance of the CNS. When there is imbalance in either direction, increased or decreased, appropriate functions of the CNS can become impaired.

6.1.5 *Neuroinflammation in ASD*

Recent studies have suggested that neuroinflammation occurs in individuals with ASD. Inflammation in post mortem brain specimens of a wide range of individuals with ASD age 4–45 years old have been observed, specifically, the cerebellum, anterior cingulate gyrus and the midfrontal regions of the brain (Vargas et al. 2005). Neuroglial activation and presence of increased levels of inflammatory cytokines such as IFN- γ , IL-1 β , IL-6, TNF- α and chemokines CCL-2 were found in brain tissue and CSF (Li et al. 2009; Morgan et al. 2010; Vargas et al. 2005). Additionally postmortem brain samples of patients with ASD were also found to have increased levels of glial fibrillary acidic protein (GFAP) in the frontal, parietal and cerebellar cortices (Laurence and Fatemi 2005). GFAP is expressed in activated astrocytes and is also a sign of inflammation. The cerebellum in particular showed the most prominent histological changes and microglial activation in individuals with ASD. In addition, some of the cerebellar tissues from individuals with ASD, but none of the control tissues had accumulation of perivascular macrophages and monocytes and deposition of complement membrane attack complexes which suggest that the neuroinflammation seen may be primarily driven by innate immune responses (Vargas et al. 2005). Furthermore, researchers found increases in TH1 with no differences in TH2 cytokines suggesting that ASD patients have increased neuroinflammatory immune response through the TH1 pathway (Li et al. 2009). Increases in TH1 cytokines such as IFN γ were not compensated by increases in IL-10 also suggesting a failure in immune regulation (Li et al. 2009). In addition to increases in cytokines, post-mortem temporal cortex samples from ASD and general population controls were assessed for transcriptome differences and increases in expression of immune related genes were found in the ASD population (Garbett et al. 2008). In particular cytokine signaling and immune regulatory genes were altered, which included genes from the NF κ B, IL-1 τ , Toll, IL-6, Caspase, TH1/TH2 and FAS pathways. Interestingly, the ASD samples had higher variability in transcriptome differences when compared to controls (Garbett et al. 2008). Furthermore, activation of microglial cells and perivascular macrophages measured by increased MHC II expression was seen in the cortical regions, white matter and most prominently in the cerebellum of patients with autism. This microglial and astroglial activation in the cerebellum was associated with degenerating purkinje cells, granule cells, and axons (Vargas et al. 2005). Altered microglial profiles found in post mortem brain samples of ASD patients showed an increase in average microglial somal volume and increase in microglial density in white and grey matter respectively and activation ranged from severe to mild in ASD brain specimens (Morgan et al. 2010). The data also suggested that microglial activation maybe particularly prominent in younger individuals, though more samples are needed to verify this (Morgan et al. 2010).

The specific inducer of microglia activation in ASD is unknown and whether dysfunction in immune pathways leads to neuroinflammation or if CNS impairments in ASD lead to immune dysregulation, or in fact an interplay between the two systems, is yet to be fully elucidated. Both environmental and genetic risk factors

are thought to play a role in ASD. Genetic contributions to ASD were first suggested in the 1980's after investigation of co-occurrences of rare syndromes and chromosomal disorders were observed with ASD (Blomquist et al. 1985). Moreover the increased occurrences of ASD in families shown in twin and sibling studies further provided evidence for a genetic component to ASD (Kates et al. 2004; Bailey et al. 1995; Constantino and Todd 2000; Steffenburg et al. 1989; Jorde et al. 1991). Candidate gene association studies and whole-genome linkage studies have been used to identify loci of interest and assess copy number variation. Even with a long list of putative contributing genetic mutations and syndromes associated with ASD, these only account for 10–20% of cases (Abrahams and Geschwind 2008). Genetic risk factors for ASD include genes that affect both CNS and immune pathways. Immune related genes associated with ASD include: phosphoinositide-3 kinase (PI3K) pathway proteins such as MET, PTEN, TSC1 and 2, as well as MHC II, complement 4B, and macrophage inhibitory factor (MIF) (Onore et al. 2012).

6.2 Maternal Immune Activation and ASD

6.2.1 Infection During Pregnancy

In addition to genetic contributions, environmental factors are also thought to play a role in ASD. Maternal immune activation (MIA) during pregnancy is one potential environmental factor that may increase the risk for developing ASD (Patterson 2009). Studies investigating viral and bacterial infections during pregnancy have shown associations with maternal infection and increases in ASD, including in 1964 when a rubella outbreak was connected with increased cases of autism (Chess et al. 1978). Other viruses that have been linked to congenital infection and associated with ASD include the herpes viruses: herpes simplex virus, cytomegalovirus, varicella and the paramyxovirus mumps (Libbey et al. 2005). The study of data from the Danish Medical Birth Register investigated 10,133 ASD diagnoses from children born from 1980 to 2005 looking at mothers who were hospitalized during pregnancy and found evidence to support association of viral infection during the first trimester and bacterial infection during the second trimester with increased risk of the child developing ASD (Atladottir et al. 2009). It is possible that genetically predisposed or susceptible individuals who encounter a prenatal infection may develop ASD due to high levels of cytokines or initiation of autoimmune processes resulting in increased maternal inflammation which could affect the developing fetal brain (Libbey et al. 2005). Additional evidence to infer maternal immune involvement in autism is data showing increased rates of autoimmunity in families with ASD (Croen et al. 2005; Atladottir et al. 2009). In support of a role of MIA, one study showed that mid-gestational findings of increased IFN γ , IL-4 and IL-5 in maternal serum significantly increased the risk of ASD (Goines et al. 2011).

6.2.2 *Other Inflammatory Processes*

In addition to increased frequencies of autoimmunity among families with individuals with ASD some reports have also identified fetal specific autoantibodies in the mothers of children with autism (Braunschweig et al. 2008; Croen et al. 2008). IgG maternal antibodies can cross the placenta and persist for up to 6 months after birth (Heininger et al. 2006). Antibodies with autoreactivity to fetal brain proteins were found at 37 kDa and 73 kDa molecular weights in approximately 12% of mothers with an autistic child but no mothers of typically developing children or children with developmental delays other than ASD (Braunschweig et al. 2008); later another band with a molecular weight of 39 kDa was also found to be associated with ASD (Croen et al. 2008). Maternal antibodies are present in detectable levels at 18 weeks in the developing fetus and reach levels comparable to the mothers by 38 weeks of gestation (Croen et al. 2008). To further test the role these autoantibodies are playing in ASD, several studies have injected serum or purified IgG from mothers of children with ASD and mothers of controls into various animal models mid gestation. In one study pregnant mouse dams were intraperitoneally injected with purified IgG from mothers of children with autism disorders (MCAD) or from mothers of typically developing children. Injections were given daily during embryonic days 13–18, resulting in adolescent offspring from MCAD injected dams which displayed long-term behavioral differences compared with controls (Singer et al. 2009). In another study non-human primate, rhesus macaques were injected with purified IgG from mothers of children with ASD and from those of typically developed children. Animals were found to have higher amounts of stereotypical behaviors and increased motor activity than controls (Martin et al. 2008). These data suggest that dysfunction of the maternal immune system may play an active role in the pathology of some children who develop ASD.

6.2.3 *Rodent Models of MIA*

Other models that investigate the role of MIA include rodent models of immune activation of pregnant dams. IL-6 is an important cytokine involved in maternal immune influence of fetal development (Hsiao and Patterson 2011). Injecting IL-6 in the absence of other immune stimulus at embryonic day 12.5 is sufficient to cause behavioral changes in the offspring, particularly in measurements of prepulse inhibition of adult offspring (Smith et al. 2007). Other pro-inflammatory cytokines such as IL-1 β , TNF- α and IFN- γ did not cause any changes in behaviors. Likewise, injection of neutralizing anti IL-6 antibodies administered when MIA was induced prevented development of behavioral abnormalities. IL-6 knockout mice also exhibited resistance to *in utero* MIA induced behavioral changes (Smith et al. 2007). Pregnant mice infected with the human influenza virus on embryonic day 9.5 had offspring who as adults displayed behavioral defects in prepulse inhibition and acoustic startle response (Shi et al. 2003). Additionally when polyinosinic –poly-

cytidylic acid (poly I:C), a viral mimic, was injected into dams at embryonic day 12.5 the offspring had similar behavioral defects as the influenza infected offspring suggesting that the behavioral abnormalities are indeed due to activation of the maternal immune system not the virus itself (Shi et al. 2003). Recent studies with poly IC induced MIA in mice show behavioral changes in three areas relevant to those seen in ASD which include impairments in communication, social interactions and repetitive behaviors (Malkova et al. 2012; Schwartzer et al. 2013). Male offspring of MIA mice were found to produce less ultrasonic vocalizations (a murine form of communication) in different social situations compared with controls. In addition, the offspring were also found to spend less time with novel mice and more time with a novel object when compared to saline controls indicating a difference in social interactions and finally the MIA offspring displayed more repetitive behaviors as measured by time spent self grooming and time spent burying marbles compared with controls (Malkova et al. 2012). In addition to poly I:C models, the use of the bacterial component lipopolysaccharide (LPS) as a mid-gestational activator of the maternal immune system has been tested and results in behavioral changes in offspring similar to those observed using poly I:C (Patterson 2009). In the latter model neuroglial activation and increased cytokine production has been shown that likely results in permanent elevation of cytokines in the brain that affect postnatal behaviors (Patterson 2009). These models together with epidemiological data of human infection during pregnancy demonstrate that the immune status of the mother is important for the developing fetus (Patterson 2009). Since not all mothers who are infected with a pathogen have offspring with ASD it is likely that genetic background acts as a factor to enhance ASD risk. Gene—environment interactions are thought to play a major role in ASD. One study examined these interactions by testing MIA in mice heterozygous for the tuberous sclerosis 2 (*Tsc2*) gene. Offspring of dams injected with poly I:C exhibited increased asocial behavioral abnormalities more than MIA alone suggesting a double hit of genetic and environmental factors results in severe behavioral defects (Ehninger et al. 2012). In addition to alteration in fetal neurodevelopment it is also possible that MIA alters peripheral immune responses as well. In one study of MIA, a TH17 skewing of T cells were seen in poly I:C maternally exposed mice compared with controls (Mandal et al. 2011).

6.3 Systemic Immune Activation in ASD

6.3.1 *Peripheral Cytokines and Chemokines in ASD*

Immune abnormalities in ASD have been reported since 1977 (Stubbs and Crawford 1977). Since that initial report there have been a number of immune related problems described with some conflicting findings likely reflecting the heterogeneity of ASD. Elevated pro-inflammatory cytokines have been found in plasma of children with ASD aged 2–5 years old including IL-1 β , IL-6, IL-8 and IL-12p40 (Ashwood et al.

2011b). Elevated amounts of chemokines MCP-1, RANTES and eotaxin were also found in children with ASD (Ashwood et al. 2011d). In both studies these elevated inflammatory mediators were associated with more impaired or aberrant behaviors. Other reports of inflammatory cytokines found elevations of IFN γ (Singh 1996), MIF (Grigorenko et al. 2008) and platelet derived growth factor BB (PDGF-BB) in plasma of children with ASD (Kajizuka et al. 2010). Both MIF and PDGF correlated with behavioral scores as well. In addition to increases in pro-inflammatory cytokines, decreases in TGF β , a regulatory cytokine, were also found in children with ASD which were associated with worsening behavioral scores (Ashwood et al. 2008). In addition to plasma cytokine differences, there have also been reports of differences in immunoglobulin levels. One study reported that children with autism have reduced levels of plasma IgG and IgM which also correlated with increased behavioral severity (Heuer et al. 2011). Other studies have reported increases in serum proteins attributed mostly to increases in albumin; however, IgG, specifically IgG2 and IgG4 were also seen elevated in individuals with ASD and these increases in immunoglobulin correlated with behavioral abnormalities (Croonenberghs et al. 2002; Enstrom et al. 2009a). Autoantibodies to various and diverse targets have been reported in children with autism and could point to cellular damage that may be involved in increasing inflammation, revealing antigens otherwise hidden and/or epitope spreading (Onore et al. 2012).

6.3.2 *Adaptive Responses in ASD*

Adaptive immune responses in children with ASD also show increased cytokine production. Peripheral blood mononuclear cells (PBMC) isolated from the blood of children with ASD ages 2–5 were stimulated and compared to age matched controls. Unstimulated cells from children with ASD produced higher amounts of IL-8 when cultured overnight. After stimulation with phytohemagglutinin (PHA), cells from individuals with ASD produced larger quantities of GM-CSF, IL-13 and TNF- α (Ashwood et al. 2011c). A number of these increased cytokines also correlated to behavioral abnormalities. Increased production of TNF- α and IFN- γ were associated with more stereotyped behaviors. Increased impaired communications were associated with higher IFN- γ and IL-8 production. IL-12p40, a subunit of IL-12, correlated with worsening speech and increased hyperactivity (Ashwood et al. 2011c). This data suggest that perhaps an increased Th1 response may worsen behaviors. Both increases in IL-10 and IL-5 may help to improve behaviors—IL-10 increases were associated with better expressive language while increased IL-5 production correlated with improved fine motor skills. Besides increased production of cytokines, PBMC differences were also seen in T cell activation markers suggesting an altered activation of T cells which may contribute to the differences in cytokines produced (Ashwood et al. 2011c). Other studies have also looked at CD4 and CD8 T cells and have found a shift in Th1 and Th2 cytokines (Gupta et al. 1998).

Adhesion molecules play an important role in leukocyte migration and are involved in modulating immune—CNS connections via passage of T cells through epithelial barriers. Soluble adhesion molecules such as sPECAM, sL-selectin, and sP-selectin were found in lower amounts in high functioning ASD individuals when compared to controls (Iwata et al. 2008; Tsuchiya et al. 2007). Reports of improved behaviors during febrile outbreaks in children with ASD have also been described; these changes in behavior are transient and may be attributed to increased up-regulation of adhesion molecules allowing for more T cell-CNS interactions (Onore et al. 2012; Curran et al. 2007).

6.3.3 Innate Responses in ASD

Changes in innate immune responses have been described in children with ASD. Natural killer (NK) cells, normally involved in killing atypical host cells, have been found to have reduced ability to kill K562 target cells (an immortalized myelogenous leukemia cell line) in children with ASD (Warren et al. 1987; Enstrom et al. 2009b; Vojdani et al. 2008). Factors that may contribute to decreased NK cell activity may be attributed to production of lower amounts of perforin, granzyme B and IFN- γ following stimulation conditions in children with ASD (Enstrom et al. 2009b). Increased numbers of circulating monocytes have also been reported in ASD (Sweeten et al. 2003). Moreover, increased expression of activation markers on these monocytes suggest that these cells are in an activated state (Ashwood et al. 2011a). Indeed these cells have been found to have released increased inflammatory cytokines such as IL-1 β , TNF- α and IL-6 in response to TLR2 and TLR4 stimulus. Increased production of IL-6 and IL-1 β correlated with increased impairment of social behaviors in children with ASD (Enstrom et al. 2010). Monocytes under certain conditions can give rise to other myeloid cells such as dendritic cells, tissue macrophages and microglia (Djukic et al. 2006; Geissmann et al. 2010). Altered activation and responses in myeloid cells, therefore, have many implications for inflammation in both peripheral and CNS systems.

6.4 Gastrointestinal Abnormalities in ASD

6.4.1 GI Symptoms and Frequency in ASD

Recent studies have suggested that many children with ASD suffer from gastrointestinal (GI) symptoms, dysfunction and inflammation. Associations between ASD and GI symptoms were first reported in the early 1970s (Goodwin et al. 1971). Goodwin looked at 15 autistic children and found seven of them had GI issues.

Since then various other groups have reported the frequency of GI symptoms in the ASD population ranging from 17 to 86% (Erickson et al. 2005). The differences in reported GI abnormalities in children with ASD are in part due to the design of these studies with many lacking proper controls or were based on referral biases that only include children with ASD who have GI complaints. Other factors contributing to the wide range of reports include the heavily skewed amount of retrospective studies that rely on either medical records which mainly look at confirmed diagnosis of GI disease and may not necessarily include GI symptoms experienced in all patients or via a parental survey. Conversely, purely parental based surveys have the disadvantage of often involving memory recall of past GI problems whilst additional impairments in language in non-verbal children make it more difficult for parents to perceive pain in children with ASD. Differences between studies can also be attributed to non standardized GI surveys which define GI symptoms differently among the various studies (Mannion et al. 2013). Common gastrointestinal symptoms that are reported include: diarrhea, constipation, foul smelling stools, gaseousness, abdominal pain, and food regurgitation/reflux.

6.4.2 Nutrition and GI Immunity in ASD

Many of the reports of gastrointestinal symptoms in children with ASD have led some researchers to look into the role that nutritional imbalance may be playing in some of the GI symptoms reported. Studies examining the relationship between nutritional input and its effect on ASD; however, have not supported this hypothesis (Levy et al. 2007). Core features of autism such as repetitive behaviors and resistance to change may impact feeding behaviors and nutrition in children with ASD (Erickson et al. 2005). Overall these studies show that while children with ASD tend to have increased food selectivity but that selectivity does not seem to cause malnutrition and overall, nutrient intake is adequate in children with ASD (Raiten and Massaro 1986; Shearer et al. 1982; Ahearn et al. 2001; Field et al. 2003). Some reports concerning increased rates of food allergies among ASD populations have been reported (Horvath and Perman 2002). One such study seeking to address these concerns found that children with ASD had more responses to food allergens as measured by positive pin prick reactions (Lucarelli et al. 1995). In another study addressing these same concerns, children with ASD were compared to normal siblings and children with known dietary protein intolerances; elevated levels of IFN γ and TNF α were found in PBMC response to dietary proteins in both the ASD group and the known dietary intolerances group (Jyonouchi et al. 2002). Also of note in this same study a correlation between elevated IFN γ and TNF α responses with dietary proteins and elevated response to LPS was seen in children with ASD (Jyonouchi et al. 2002). This suggests that an imbalanced immune system may be playing a role in GI dysfunction.

6.4.3 *GI Immunity*

The gastrointestinal tract is the immune system's largest source of lymphoid tissue and is an important site of immune regulation (Turner and Goldsmith 2009). It is therefore conceivable that mucosal immune dysfunction could be playing a role in children with ASD who have GI symptoms. Immunohistochemical findings of children with ASD and GI problems showed an increase in CD8 T cells in duodenal and colonic samples (Torrente et al. 2002; Furlano et al. 2001), with increases also seen in $\gamma\delta$ T cells in transverse colon samples (Furlano et al. 2001). In addition, decreases in peripheral T cell numbers were reported in children with ASD who have GI symptoms (Ashwood et al. 2003) and may reflect numbers of T cells translocating to the GI mucosa in this subset of ASD individuals. Other immunohistochemical findings revealed deposition of IgG and complement C1q co-localized on the basolateral enterocyte membrane in ASD GI samples (Torrente et al. 2002; Ashwood et al. 2003) suggesting a possible autoimmune component to ASD GI dysfunction. Other studies have illustrated findings of increased number of paneth cells in children with autism and gastrointestinal symptoms (Horvath and Perman 2002; Torrente et al. 2002; Horvath et al. 1999).

Common reports of children with ASD having “leaky gut” have been noted and one study in particular looked at 21 autistic children without known GI disease and found 9 with mucosal permeability; none of the 40 normal controls had permeability issues (D'Eufemia et al. 1996). Results of this study were confirmed when another group also found increased GI permeability in children with ASD and GI problems (Horvath 2000). In humans, disruption of mucosal barriers can occur in the absence of inflammation. Thus, increased mucosal permeability does not necessarily predict inflammation. Interestingly though, findings of atypical intestinal microbiota composition in ASD including increased findings of *Clostridium* species in stool samples of children with ASD, (Finegold et al. 2010; Finegold et al. 2004; Finegold et al. 2002) may elicit mucosal immune responses and GI dysfunction in some children with ASD and GI symptoms especially if bacteria are able to move across a more permeable intestinal barrier. Moreover, a small study developed on the reports of abnormal microbiota composition in children with ASD looked at 11 children with ASD, and treated with the antibiotic vancomycin, aiming to remove deleterious microbes with the antibiotic. Findings of this study include temporarily improved behavioral symptoms (Sandler et al. 2000). Another study addressed this issue by treating children with ASD and GI issues with oral human immunoglobulin (IG). The results showed that after administering oral IG for 8 weeks, 50% of the patients had improvements in GI severity; however, benefits for the treatment were not maintained at the 30 day follow up. Behavioral improvements were also seen from baseline through the end of the 8 week treatment (Schneider et al. 2006). Other studies that assessed treatment for GI and behavioral symptoms include treatment with secretin. Erickson et al. (2005) reported findings from 11 double blind and 2 open label studies testing secretin treatments in the ASD population. While one of the

open label studies reported behavioral improvements based on parental reporting, all double blind studies showed no differences between control (placebo) groups and secretin treatment groups (Erickson et al. 2005). These studies suggest that more research is needed to address the GI/immune/brain connection. There have been some studies that try to investigate the role GI inflammation has in altering behaviors and CNS function. One group of researchers infected AKR mice with the parasite *trichuris muris* and measured GI inflammation, brain biochemistry changes and behaviors related to anxiety. The authors found that chronic GI inflammation induces anxiety-like behavior in AKR mice (Bercik et al. 2010). Increased use and development of behavioral testing in mice now make it easier to measure behavioral changes associated with various induced and natural occurring conditions in mice.

6.4.4 Behavior and GI Dysfunction

Behavioral abnormalities associated with GI symptoms in autism include studies that have found children who had sleep abnormalities were also more likely to have GI problems (Maenner et al. 2012; Mannion et al. 2013; Ming et al. 2008). Mazurek et al. (2012) looked at 2973 children enrolled in autism treatment network and found that children with ASD and GI problems had higher levels of anxiety and sensory overresponsivity. The authors also suggest that the relationship of these three symptoms could include the involvement of the hypothalamic-pituitary-adrenal (HPA) axis and amygdala based circuits. It is of note that the HPA axis also regulates immune function (Mazurek et al. 2012; Herman and Cullinan 1997). The likelihood of a connection between mucosal immune irregularities and behavior in ASD is intriguing. While it is possible that immune dysfunction in the GI tract interacts with an already abnormal brain to aggravate behavioral symptoms or whether GI problems are just another manifestation of systemic immune abnormalities are questions still to be explored. Many studies examining animal and human subjects reveal elevated peripheral cytokines are able to cause striking changes in behavior (Patterson 2009). It still remains to be seen if similar phenomenon occurs with mucosal inflammation.

6.5 Conclusion

ASD is a complex and heterogeneous spectrum of behavioral disorders made even more apparent by the wide array of genetic and environmental factors and influences that have been linked to it. This heterogeneity has also impacted studies of the immune system with many findings that are often confusing and sometimes conflicting. Overall one theme that has remained constant is that, inflammation is associated with ASD. Findings of neuroinflammation in the CSF and brain tissue, peripheral immune abnormalities on both the innate and adaptive responses and the increasing number of studies on GI dysfunction within ASD all come to a consensus

that immune dysfunction can have persistent impact on behavior. More research is needed to further elucidate the mechanisms by which the immune system affects neurological and behavioral changes and to investigate immune modulating therapeutics for ASD.

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