

Hikmat A. Al-Ahmadie and Gopa Iyer

Immunohistochemical Markers With Diagnostic Value in Bladder Cancer

Immunohistochemistry

In the majority of cases, an accurate diagnosis of urothelial carcinoma as well as the presence and extent of invasion is achieved by regular histological examination without the need for ancillary studies such as immunohistochemistry (IHC). As discussed below, however, there are situations where IHC might be helpful. The following are some of the common scenarios. Details about new or relevant markers for urothelial differentiation will be reported following the section.

Distinction of Reactive Atypia from Urothelial Carcinoma In Situ

In flat urothelial lesions, IHC with CK20, CD44, p53, and Ki-67 may be utilized to aid in the distinction of reactive flat urothelial lesions from urothelial carcinoma in situ (CIS) [1–3]. In the normal state, CK20 (Fig. 18.1a) expression is limited to the umbrella cell layer and CD44

(Fig. 18.1b) stains predominantly the basal cell layer. Urothelial CIS is expected to express CK20 (Fig. 18.2a) in the majority of tumor cells (full thickness) with total loss of CD44 expression (Fig. 18.2b). CIS is also expected to exhibit diffuse labeling with p53 and the proliferation marker Ki-67. On the other hand, reactive urothelial lesions are expected to express CD44 with the other markers exhibiting limited expression. This pattern has been recently shown to aid in the differential diagnosis of radiation-induced atypia versus urothelial CIS [4].

It is important to note that none of these markers should be used individually to establish a malignant or benign diagnosis. Aberrant expression of these markers is well established and their interpretation must be made in the correct context. Moreover, IHC should not be used in all cases as a screening test, but rather as an adjunctive tool to aid in the histological classification of atypical flat urothelial lesions or in the de novo diagnosis of CIS where the morphologic features are questionable.

Differentiating Urothelial Carcinoma (with or Without Divergent Differentiation) from Other Carcinomas that Secondarily Involve the Urinary Bladder

Distinguishing invasive urothelial carcinoma from carcinomas secondarily involving the urinary bladder is of paramount significance and can at times be difficult due to significant morphologic overlap. Prostatic adenocarcinoma, colorectal adenocarcinoma, squamous cell carcinoma of

H. A. Al-Ahmadie (✉)
Department of Pathology, Memorial Sloan Kettering
Cancer Center, New York, NY, USA
e-mail: alahmadh@mskcc.org

G. Iyer
Department of Medicine, Memorial Sloan Kettering
Cancer Center, New York, NY, USA
e-mail: iyerg@mskcc.org

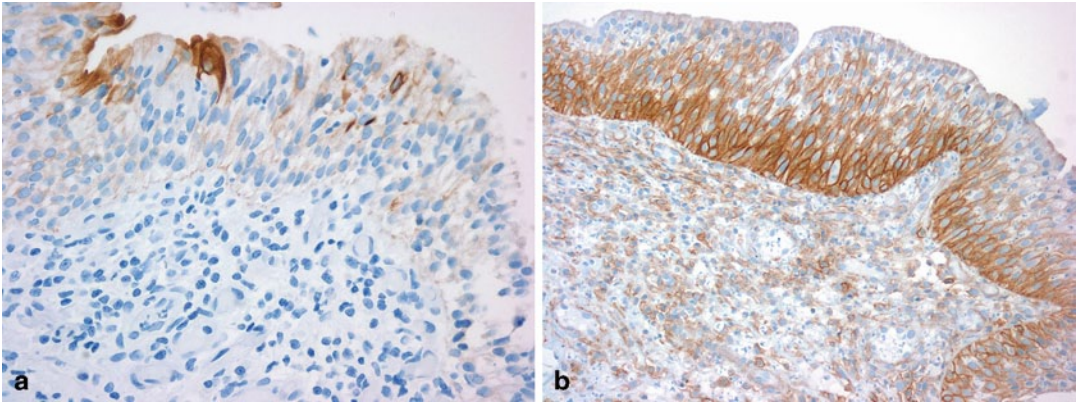


Fig. 18.1 In normal urothelium and in reactive atypia CK20 expression is limited to the umbrella cell layer (a); CD44 stains predominantly the basal cell layer, although patchy positivity can be seen in all layers (b)

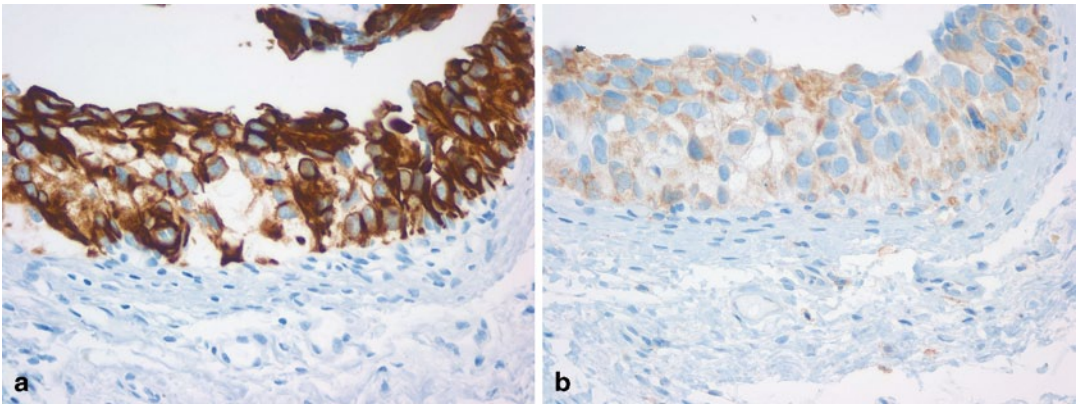


Fig. 18.2 Urothelial carcinoma in situ expresses CK20 in the majority of tumor cells (full thickness) (a), with loss of CD44 expression (b)

the uterine cervix, and to a lesser extent, carcinomas of the uterus or ovary, and rarely those from breast, lung, stomach, and skin [5, 6], can involve the bladder during their course and may present a diagnostic challenge primarily in the absence of a relevant clinical history regarding the potential primary site of origin.

It is important to keep in mind that both squamous and glandular differentiations are common findings in primary urothelial carcinoma of the bladder but the diagnostic dilemmas can arise in cases of pure adenocarcinoma or squamous cell carcinoma involving the bladder. The best way to solve any potential misdiagnosis is to think of the possibility that these tumors can present in the bladder and to explore the clinical situation

of the patient. Having a similar tumor in a site where it is more common to have tumors with the given morphology is perhaps the strongest clue favoring a metastatic origin of the bladder tumor.

Poorly Differentiated Prostatic Adenocarcinoma Versus Urothelial Carcinoma

These two entities might have morphologic overlap and the clinical management implications are significant. The history of prostatic adenocarcinoma might not be provided or might be overlooked. Additionally, some of these patients might have received treatments that affected the morphological appearance of the prostate cancer, further complicating its recognition.

Generally, a panel of markers is useful in separating the two entities in the majority of the cases. Markers that are supportive of prostatic differentiation (Fig. 18.3a) include prostate-specific antigen (PSA) (Fig. 18.3b), prostate-specific acid phosphatase (PSAP), prostate-specific membrane antigen (PSMA), P501s (Fig. 18.3c), NKX3.1 (Fig. 18.3d), and erythroblast transformation-specific-related gene (ERG); whereas, markers favoring urothelial differentiation (Fig. 18.4a) and origin include high molecular weight cytokeratin (34 β e12), CK7 (Fig. 18.4b), p63 (Fig. 18.4c), thrombomodulin, uroplakin III, and recently GATA3 (Fig. 18.4d) [7–12].

Obviously not all of these markers are needed in any individual case. It is recommended to start with a few markers with high sensitivity and

specificity and then use additional markers as needed. PSA, CK34 β e12, and p63 are very useful as a start in the majority of cases.

A unique scenario is the presence of prostatic adenocarcinoma with extensive squamous differentiation involving the bladder. This is a rare situation that occurs primarily postradiation or hormone therapy for prostate cancer. The clues to the prostatic origin of such a tumor is the clinical suspicion based on the clinical history which should prompt careful and extensive examination of the tumor to find even the slightest glandular differentiation, which would then be confirmed by any of the prostatic markers mentioned above. It is only logical to keep in mind that these prostatic markers will not be expressed in the component with pure squamous differentiation.

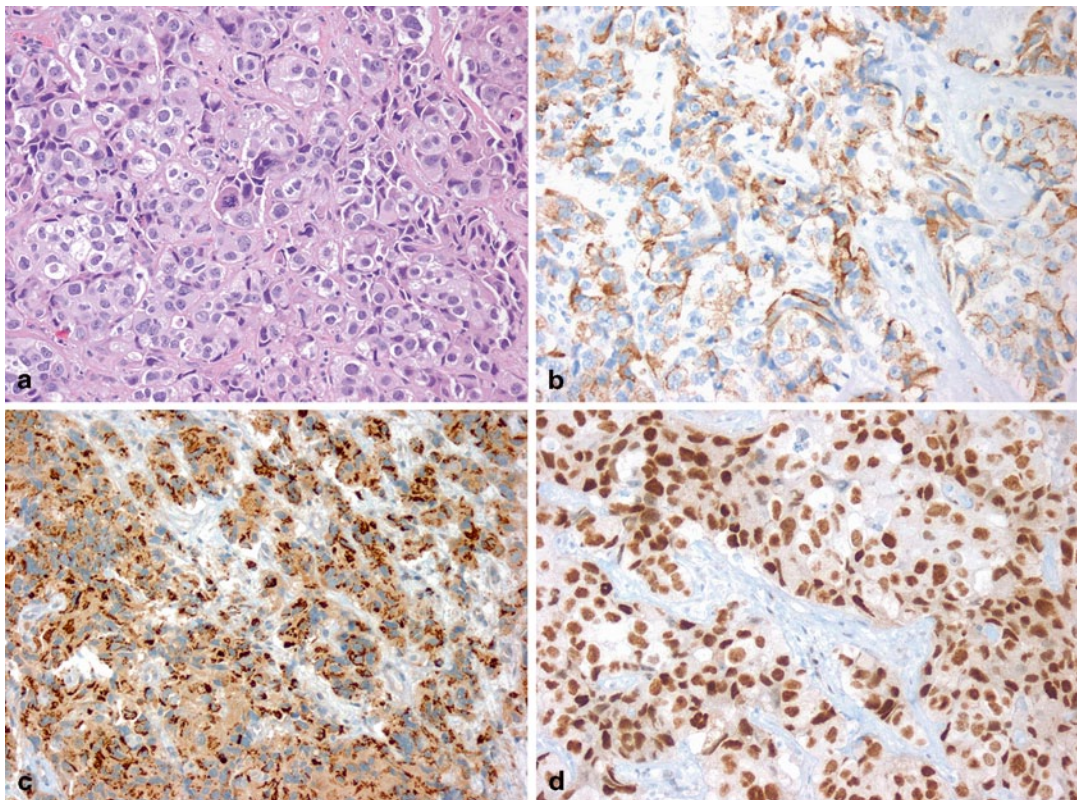


Fig. 18.3 Bladder neck tumor from a 70-year-old man with urine cytology positive for urothelial carcinoma (a). Tumor cells are positive for PSA (b), P501s (c), and NKX3.1 (d), supporting the diagnosis of prostatic adenocarcinoma

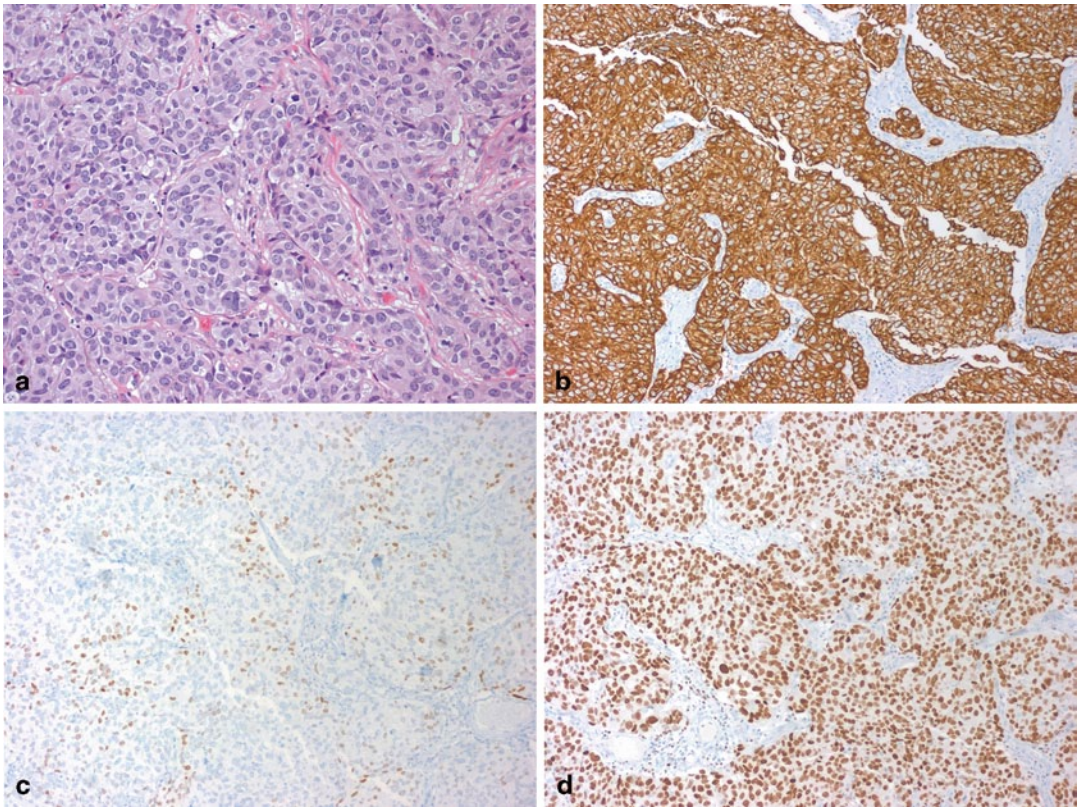


Fig. 18.4 Bladder mass from a 67-year-old man with urine cytology positive for urothelial carcinoma (a). Tumor is diffusely positive for CK7 (b), focally positive

for p63 (c), and diffusely positive for GATA3 (d), supporting the diagnosis of urothelial carcinoma

Colorectal Adenocarcinoma Involving the Bladder by Direct Extension or Metastasis Versus Primary Bladder Adenocarcinoma (Enteric Morphology)

For this differential diagnosis, the clinical history is also extremely important which should include knowledge of the presence of a prior or current tumor of the colorectal region, its grade, and stage. These tumors may even colonize the bladder mucosa giving the impression of a “precursor” or “in situ” lesion. Unfortunately, IHC currently is of limited value in this differential diagnosis as tumors with enteric phenotype will generally stain similarly regardless of the site of origin. There have been suggestions that β -catenin might be of value in this scenario

as it will not label the nuclei of primary bladder adenocarcinoma compared to those originating from the colorectal region [13–16]. While this pattern seems to be of value, nuclear localization of β -catenin was still reported in cases of primary bladder adenocarcinoma in some of these studies. Moreover, nuclear localization of β -catenin is not universal to all cases of primary colorectal adenocarcinoma, and a negative stain (i.e., only membranous and or cytoplasmic expression) will not exclude a colorectal primary. It is therefore, very important to always inquire about the clinical history of the patient for the possibility of a primary in the colorectal region when facing the diagnosis of enteric adenocarcinoma in the urinary bladder.

Squamous Cell Carcinoma of Uterine Cervix Involving the Bladder by Direct Extension or Metastasis Versus Primary Squamous Cell Carcinoma of the Bladder (or Urothelial Carcinoma with Squamous Differentiation)

Bladder involvement by squamous cell carcinoma of uterine cervical origin is admittedly rare but can still be diagnostically challenging when encountered, especially considering that squamous differentiation is rather common in urothelial carcinoma.

A number of markers have shown strong correlation with squamous neoplasms but unfortunately, these markers will not be able to point to a specific site of origin for these squamous carcinomas. Examples of such markers include desmoglein-3, MAC387, and TRIM29, which although sensitive markers for squamous phenotype, can be positive in squamous cell carcinoma of the cervix as well as that of the bladder. These markers can also be expressed in the squamous component of urothelial carcinoma with squamous differentiation and less commonly within the classical urothelial component [17, 18]. This is why these markers are not reliable as the sole means of establishing a site of origin for a tumor with squamous differentiation.

The role of the human papillomavirus (HPV) is well established in the vast majority of cervical squamous cell carcinoma for which p16 serves as a surrogate marker for the detection of HPV in these tumors [19, 20]. The expression of p16 in squamous cell carcinoma of the bladder has been shown to be less specific with little if any association with HPV infection in such setting [21]. There were, however, rare cases of true HPV-associated squamous cell carcinoma of the bladder and at least in some of them the tumors exhibited basaloid morphology and were associated with a history of neurogenic bladder or other situations that required repeated catheterization of the bladder [22, 23].

A number of *other carcinomas* may rarely involve the bladder during their course such as mammary carcinoma, endometrial or ovarian carcinoma, gastric carcinoma, etc. It is prudent to review the primary tumor alongside the metastasis. Immunostains might be ordered according to

the suspected primary tumor, particularly if the status of such markers is known in the primary site.

Markers for the Differential Diagnosis of Spindle Cell Lesions of the Bladder

Many entities exist in the bladder in which spindle cell morphology predominates and range from reactive myofibroblastic lesions to frankly malignant (sarcomatous) entities. The main categories include inflammatory myofibroblastic tumor/pseudosarcomatous myofibroblastic proliferations (IMT/PMP), sarcomatoid urothelial carcinoma, and sarcomas with spindle cell morphology (leiomyosarcoma, rhabdomyosarcoma). There is marked overlap in morphology and immunoprofile among these entities and a judicious use of IHC in the context of morphology plays an important supportive role in this differential diagnosis.

Establishing the diagnosis of IMT can be aided by the expression of ALK by IHC or the presence of *ALK* rearrangement by fluorescence in situ hybridization (FISH) or other molecular technique. Since this expression or rearrangement is not present in all cases, a negative test does not rule out the diagnosis of IMT [24]. The overall morphologic features and the expression of other markers such as smooth muscle actin and cytokeratins may help. The challenge remains to differentiate this entity from a reactive myofibroblastic proliferation, which can be exuberant in the bladder.

For sarcomatoid urothelial carcinoma, finding an unequivocal epithelial component would be the ideal scenario but when this is not feasible, the presence of epithelial differentiation by IHC might be helpful in pointing toward the diagnosis of sarcomatoid UC. This can be achieved by a number of epithelial markers such as wide spectrum cytokeratins (AE1/AE3, CAM5.2...), epithelial membrane antigen (EMA), high molecular weight keratins, and p63. GATA3 might be helpful as well but we still do not know its full functions in spindle cell lesions in general and more studies are needed to assess its value in this setting.

For true sarcomas with specific lineage or differentiation such as leiomyosarcoma and rhabdomyosarcoma, the diagnosis can be confirmed by the markers related to these entities such as actin, desmin, myogenin, etc.

The Confirmation of Urothelial Differentiation at a Metastatic Site

Generally, urothelial carcinoma presents at metastatic sites with the morphology of a poorly differentiated carcinoma without specific morphologic features. It could be particularly difficult to distinguish metastatic urothelial carcinoma from metastatic squamous cell carcinoma (or from primary squamous cell carcinoma in the example of a lung tumor). What might be helpful in pointing to an origin of a urothelial primary include (1) prior history of bladder cancer, which should warrant review of the primary tumor if available and (2) the presence of divergent differentiation (squamous, glandular, etc.).

In these settings, IHC can play a role in establishing the urothelial origin of such tumors.

Antibodies that can be used to confirm urothelial differentiation/origin include GATA3, cytokeratins 7 and 20, high molecular weight cytokeratin, p63, uroplakin III, thrombomodulin, cytokeratin CK5/6, and S100P [25–28].

It is important to keep in mind that, despite their relative specific pattern of expressions, none of these markers is by itself diagnostic of a primary urothelial carcinoma as certain degree of overlap still exists and it may take more than one marker to help in this differential.

The Role of IHC in Confirming the Presence of Lymphovascular Invasion (LVI)

LVI in urothelial carcinoma has been reported to be an independent prognostic factor for metastasis, recurrence, and survival [29–31]. Identifying LVI, however, can be complicated by the presence of peri-tumoral stromal retraction, which is a relatively common finding in invasive urothelial carcinoma that mimics LVI. This is particularly problematic within the lamina propria. As a result, assessing LVI suffers from a considerable lack of diagnostic reproducibility, which limits

its utility as a prognostic finding [32, 33]. If LVI is to retain its clinical significance, it should be reported with caution and after applying rigid criteria for its identification. In this regard, a number of endothelial/vascular IHC markers can be used to confirm the presence of LVI such as CD31, CD34, D2-40, and ERG [34, 35]. It is not recommended, however, to use these markers as a screening tool in all cases of invasive urothelial carcinoma and they should be used only in histologically equivocal cases for confirmation.

The Role of IHC in Staging of Bladder Cancer

For the majority of cases, documenting invasion in bladder cancer is not problematic by following well-established and recognized criteria [36]. In cases of ambiguity, however, such as thermal artifact, marked inflammation, or disrupted anatomy due to a prior biopsy, applying IHC may be helpful. The most commonly used markers are cytokeratins (AE1/AE3, CK7, CK8/18). An important caveat is the potential staining of stromal myofibroblasts with such epithelial markers.

Documenting tumor invasion of the muscularis propria (MP) is an important parameter in staging urothelial carcinoma, upon which major management decisions depend, such as proceeding to radical cystectomy or the administration of neoadjuvant chemotherapy. The distinction between MP and muscularis mucosae (MM), although readily achieved by light microscopy in most cases, may be challenging in some situations, such as extensive tumor infiltration of tissue fragments, post-biopsy changes that mask the normal anatomy, marked thermal artifact of tumor-bearing tissue, or hyperplastic MM. Several muscle markers have been tried in the past but were found to be of limited utility such as smooth muscle actin, desmin, and caldesmon. Recent reports have identified a new marker, smoothelin, expressed by terminally differentiated smooth muscle cells, to be differentially expressed in smooth muscle of the MP compared to that of the MM [37–41]. It should be noted, however, that other studies reported overlap of staining intensity of smoothelin between MM and MP [42]. Hence, it is still early to determine the

exact role of smoothelin as a diagnostic marker to determine tumor invasion into MP and should be used with caution.

Immunohistochemical Markers with Prognostic Value in Bladder Cancer

In papillary urothelial tumors, a number of markers have shown promising results, particularly in distinguishing between low-grade and high-grade papillary urothelial carcinoma and decreasing the interobserver variability in this category. Ki-67 and survivin were two markers that have been frequently studied and whose increased expression correlates with recurrence and progression of papillary tumors [43, 44]. Similar results were reported when the mRNA levels of survivin were measured both in urine cytology and tumor tissue [43, 45–48].

Despite the great advancement in the molecular biology of urothelial carcinoma, there has not been to date a molecular marker that outperforms a combination of established morphologic and clinical markers such as grade, histologic type, and stage, in predicting clinical outcome. This has been the case with the tumor suppressor genes p53 and Rb, which are known to be involved in urothelial neoplasia. Although they have been shown by several investigators to be accurate predictors of progression, metastasis, survival, and possibly response to systemic chemotherapy [49–53], others have challenged these results which have not been validated prospectively.

On the other hand, IHC can serve as a surrogate marker for underlying molecular aberrations that can be used in targeted therapy. In particular, alterations in receptor tyrosine kinases present promising opportunities for targeted therapy in urothelial carcinomas, such as those targeting *ERBB2* (Her2) amplifications or mutations and *FGFR3* mutations; as well as aberrations in the mTOR/Akt/PI3K pathway that are known to affect subsets of urothelial carcinoma [54–57].

GATA3 is a transcription factor of the GATA family whose functions include regulating genes involved in the luminal differentiation of breast epithelium, genes related to T-cell development, gene regulation in the development

or maintenance of skin, trophoblasts, and some endothelial cells [12, 58]. *GATA3* has been identified as an IHC marker for mammary and urothelial carcinomas in both primary and metastatic setting. It has been suggested useful in the distinction between urothelial versus prostatic adenocarcinoma and metastatic urothelial versus squamous cell carcinoma in the lung. Despite the early promising specificity and sensitivity, however, more recent studies have shown that not all non-urothelial squamous cell carcinomas or prostate cancers to be negative [12, 18, 25–27]. *GATA3* can still be of use in the workup of a neoplasm with possible urothelial origin if used with the right context and right combination with other antibodies.

Uroplakins are widely regarded as urothelium-specific proteins of terminal urothelial cell differentiation and have been reported positive in both primary and metastatic urothelial carcinoma [59–62]. Despite being specific to urothelial differentiation, they are not very sensitive as some urothelial carcinomas are not positive for these markers, which limits their practical use and requires the addition of other markers in the workup for a potential urothelial tumor.

Thrombomodulin is a surface glycoprotein involved in the regulation of intravascular coagulation that has been reported to be expressed in a variety of tumors including mesothelioma, endothelial vascular tumors, squamous carcinomas, urothelial carcinoma, and various adenocarcinomas in primary and metastatic setting [63]. The lack of specificity of this marker to urothelial differentiation limits its use in this setting. But as it has been shown in a number of studies mentioned in this section, this marker can be useful when used in combination with other markers in the workup of a potential urothelial tumor.

S100P is a member of the S100 family of proteins that was first discovered in placenta and was thus designated S100P (it is different from the S100 that is widely used in the melanocytic and nerve sheath tumors). Although it was initially identified in the placenta, expression of S100P by IHC has also been described in benign and malignant urothelial cells, pancreatic carcinoma, esophageal squamous mucosa, and breast carcinoma [11].

Urine-Based Markers for Diagnosis of Bladder Cancer

Fluorescence In Situ Hybridization (UroVysion®, Abbott Molecular, Abbott Park, IL, USA)

UroVysion® is a FISH probe set with Food and Drug Administration (FDA) approval for use in monitoring tumor recurrence and primary detection of UC in voided urine specimens from patients with gross or microscopic hematuria, but no previous history of UC. The UroVysion® test probe set contains a mixture of four fluorescent labeled DNA probes; a locus-specific probe to the 9p21 band on chromosome 9 and to the centromere of chromosomes 3, 7, and 17. The individual sensitivity of the centromeric probes for chromosome 3, 7, 17 is reported to be 73.7, 76.2, and 61.9%, respectively, while the sensitivity of homozygous 9p21 deletion for UC has been reported as 28.6% [64]. The UroVysion® test is based on combination of these probes and the sensitivity and specificity has been reported to be 72 and 83%, respectively [65]. This test, however, is not free of false positive and false negative results [66]. Inflammation may interfere with proper interpretation of the test.

The Bladder Tumor Antigen (BTA) Tests

This test is based on the detection of the human complement factor H-related protein, which is reported to be expressed only in bladder tumor cells [67, 68]. There are two types of BTA tests, one can be used in the physician's office or even in the patient's home (BTA stat), while the other has to be sent to a reference laboratory for analysis (BTA trak). The sensitivity of the BTA stat is reported to be 50% for low-grade urothelial carcinomas, which is higher than cytology. Conversely, the specificity of BTA stat is reportedly lower than cytology [69, 70]. The BTA stat test is FDA-approved for use by patients undergoing monitoring for recurrent bladder cancer.

Nuclear Matrix Protein 22 (NMP22)

This test is based on the detection of NMP22, which is a member of a family of proteins that is part of the structural framework of the nucleus and provide support for the nuclear shape. It is also involved in DNA replication, RNA transcription, and regulation of gene expression [71]. This protein is reported to have a concentration as high as 25 times in UC as compared to normal urothelial cells [71, 72]. This assay is FDA-approved for both the detection of new cancers and the follow-up of patients with a prior history of urothelial carcinoma. The reported sensitivity ranges are 34.6–100%, and 49.5–65.0%, but false positive results have been reported [73, 74].

Bladder Cancer Immunofluorescence Assay (Former Immunocyt®)

This is an immunofluorescence assay designed to improve the sensitivity of urine cytology. It employs a cocktail of three monoclonal antibodies; M344, LDQ10, and 19A211 [75]. The first two detect a mucin-like antigen, while the third one recognizes a high molecular weight glycosylated form of carcinoembryonic antigen in exfoliated tumor cells. This assay is FDA approved only for use as a surveillance test if used in conjunction with cytology. The overall sensitivity of the combined Bladder Cancer Immunofluorescence Assay and cytology is approximately 84%, which is better than either test alone. It performs better at the detection of low-grade UC [76, 77].

Telomerase

Telomeres are repetition sequences at the end of chromosomes that protect genetic stability during DNA replication. As a result of telomeric loss during each cell division, chromosomal instability and cell senescence develops. Bladder cancer cells express telomerase, which is an enzyme that regenerates telomeres at the end of each DNA replication. The detection of the ribonucleoprotein

telomerase (the telomerase subunits human telomerase RNA [hTR] and human telomerase reverse transcriptase [hTERT]) in urine samples may offer diagnostic applications as the activity of this enzyme is generally limited to malignant cells and tissues. Detection of telomerase activity is available by the TRAP-assay (telomeric repeat amplification protocol), which is a polymerase chain reaction (PCR)-based method [78, 79]. Most studies on telomerase activity in bladder cancer report good sensitivity of the tests but low specificity. Moreover, test results can be influenced by the patient's age and inflammatory conditions of the urinary system, making this assay a suboptimal test for the detection of bladder cancer [78, 80].

In a recent comprehensive review of the role of urine biomarkers in the detection and surveillance of bladder cancer, there were several markers that showed higher sensitivity compared with cytology but were less specific. Hence, they remain insufficient to replace cystoscopy approach to establish the diagnosis of urothelial carcinoma [81].

There is a need for well-designed protocols and prospective, controlled trials to provide the basis to integrate biomarkers into clinical decision making for bladder cancer detection and screening in the future.

Acknowledgement Microphotographs provided by Dr. Magi-Galluzzi

References

- McKenney JK, Desai S, Cohen C, Amin MB. Discriminatory immunohistochemical staining of urothelial carcinoma in situ and non-neoplastic urothelium: an analysis of cytokeratin 20, p53, and CD44 antigens. *Am J Surg Pathol.* 2001;25(8):1074–8.
- Kunju LP, Lee CT, Montie J, Shah RB. Utility of cytokeratin 20 and Ki-67 as markers of urothelial dysplasia. *Pathol Int.* 2005;55(5):248–54.
- Mallofre C, Castillo M, Morente V, Sole M. Immunohistochemical expression of CK20, p53, and Ki-67 as objective markers of urothelial dysplasia. *Mod Pathol.* 2003;16(3):187–91.
- Oliva E, Pinheiro NF, Heney NM, Kaufman DS, Shipley WU, Gurski C, et al. Immunohistochemistry as an adjunct in the differential diagnosis of radiation-induced atypia versus urothelial carcinoma in situ of the bladder: a study of 45 cases. *Hum Pathol.* 2013;44(5):860–6.
- Coleman JF, Hansel DE. Utility of diagnostic and prognostic markers in urothelial carcinoma of the bladder. *Adv Anat Pathol.* 2009;16(2):67–78.
- Bates AW, Baithun SI. Secondary neoplasms of the bladder are histological mimics of nontransitional cell primary tumours: clinicopathological and histological features of 282 cases. *Histopathology.* 2000;36(1):32–40.
- Genega EM, Hutchinson B, Reuter VE, Gaudin PB. Immunophenotype of high-grade prostatic adenocarcinoma and urothelial carcinoma. *Mod Pathol.* 2000;13(11):1186–91.
- Chuang AY, DeMarzo AM, Veltri RW, Sharma RB, Bieberich CJ, Epstein JI. Immunohistochemical differentiation of high-grade prostate carcinoma from urothelial carcinoma. *Am J Surg Pathol.* 2007;31(8):1246–55.
- Hameed O, Humphrey PA. Immunohistochemistry in diagnostic surgical pathology of the prostate. *Semin Diagn Pathol.* 2005;22(1):88–104.
- Liu H, Shi J, Wilkerson ML, Lin F. Immunohistochemical evaluation of GATA3 expression in tumors and normal tissues: a useful immunomarker for breast and urothelial carcinomas. *Am J Clin Pathol.* 2012;138(1):57–64.
- Higgins JP, Kaygusuz G, Wang L, Montgomery K, Mason V, Zhu SX, et al. Placental S100 (S100P) and GATA3: markers for transitional epithelium and urothelial carcinoma discovered by complementary DNA microarray. *Am J Surg Pathol.* 2007;31(5):673–80.
- Miettinen M, McCue PA, Sarlomo-Rikala M, Rys J, Czapiewski P, Wazny K, et al. GATA3: a multi-specific but potentially useful marker in surgical pathology: a systematic analysis of 2500 epithelial and nonepithelial tumors. *Am J Surg Pathol.* 2014;38(1):13–22.
- Wang HL, Lu DW, Yerian LM, Alsikafi N, Steinberg G, Hart J, et al. Immunohistochemical distinction between primary adenocarcinoma of the bladder and secondary colorectal adenocarcinoma. *Am J Surg Pathol.* 2001;25(11):1380–7.
- Paner GP, McKenney JK, Barkan GA, Yao JL, Frankel WL, Sebo TJ, et al. Immunohistochemical analysis in a morphologic spectrum of urachal epithelial neoplasms: diagnostic implications and pitfalls. *Am J Surg Pathol.* 2011;35(6):787–98.
- Roy S, Smith MA, Cieply KM, Acquafondata MB, Parwani AV. Primary bladder adenocarcinoma versus metastatic colorectal adenocarcinoma: a persisting diagnostic challenge. *Diagn Pathol.* 2012;7:151.
- Rao Q, Williamson SR, Lopez-Beltran A, Montironi R, Huang W, Eble JN, et al. Distinguishing primary adenocarcinoma of the urinary bladder from secondary involvement by colorectal adenocarcinoma: extended immunohistochemical profiles emphasizing novel markers. *Mod Pathol.* 2013;26(5):725–32.
- Huang W, Williamson SR, Rao Q, Lopez-Beltran A, Montironi R, Eble JN, et al. Novel markers of squamous differentiation in the urinary bladder. *Hum Pathol.* 2013;44(10):1989–97.

18. Gulmann C, Paner GP, Parakh RS, Hansel DE, Shen SS, Ro JY, et al. Immunohistochemical profile to distinguish urothelial from squamous differentiation in carcinomas of urothelial tract. *Hum Pathol.* 2013;44(2):164–72.
19. Dehn D, Torkko KC, Shroyer KR. Human papillomavirus testing and molecular markers of cervical dysplasia and carcinoma. *Cancer.* 2007;111(1):1–14.
20. Agoff SN, Lin P, Morigara J, Mao C, Kiviat NB, Koutsky LA. p16(INK4a) expression correlates with degree of cervical neoplasia: a comparison with Ki-67 expression and detection of high-risk HPV types. *Mod Pathol.* 2003;16(7):665–73.
21. Alexander RE, Hu Y, Kum JB, Montironi R, Lopez-Beltran A, MacLennan GT, et al. p16 expression is not associated with human papillomavirus in urinary bladder squamous cell carcinoma. *Mod Pathol.* 2012;25(11):1526–33.
22. Blochin EB, Park KJ, Tickoo SK, Reuter VE, Al-Ahmadie H. Urothelial carcinoma with prominent squamous differentiation in the setting of neurogenic bladder: role of human papillomavirus infection. *Mod Pathol.* 2012;25(11):1534–42.
23. Chapman-Fredricks JR, Cioffi-Lavina M, Accola MA, Rehrauer WM, Garcia-Buitrago MT, Gomez-Fernandez C, et al. High-risk human papillomavirus DNA detected in primary squamous cell carcinoma of urinary bladder. *Arch Pathol Lab Med.* 2013;137(8):1088–93.
24. Shanks JH, Iczkowski KA. Spindle cell lesions of the bladder and urinary tract. *Histopathology.* 2009;55(5):491–504.
25. Zhao L, Antic T, Witten D, Paner GP, Taxy JB, Husain A, et al. Is GATA3 expression maintained in regional metastases?: a study of paired primary and metastatic urothelial carcinomas. *Am J Surg Pathol.* 2013;37(12):1876–81.
26. Gruver AM, Amin MB, Luthringer DJ, Westfall D, Arora K, Farver CF, et al. Selective immunohistochemical markers to distinguish between metastatic high-grade urothelial carcinoma and primary poorly differentiated invasive squamous cell carcinoma of the lung. *Arch Pathol Lab Med.* 2012;136(11):1339–46.
27. Chang A, Amin A, Gabrielson E, Illei P, Roden RB, Sharma R, et al. Utility of GATA3 immunohistochemistry in differentiating urothelial carcinoma from prostate adenocarcinoma and squamous cell carcinomas of the uterine cervix, anus, and lung. *Am J Surg Pathol.* 2012;36(10):1472–6.
28. Parker DC, Folpe AL, Bell J, Oliva E, Young RH, Cohen C, et al. Potential utility of uroplakin III, thrombomodulin, high molecular weight cytokeratin, and cytokeratin 20 in noninvasive, invasive, and metastatic urothelial (transitional cell) carcinomas. *Am J Surg Pathol.* 2003;27(1):1–10.
29. Gondo T, Nakashima J, Ozu C, Ohno Y, Horiguchi Y, Namiki K, et al. Risk stratification of survival by lymphovascular invasion, pathological stage, and surgical margin in patients with bladder cancer treated with radical cystectomy. *Int J Clin Oncol.* 2012;17(5):456–61.
30. Lotan Y, Gupta A, Shariat SF, Palapattu GS, Vazina A, Karakiewicz PI, et al. Lymphovascular invasion is independently associated with overall survival, cause-specific survival, and local and distant recurrence in patients with negative lymph nodes at radical cystectomy. *J Clin Oncol.* 2005;23(27):6533–9.
31. Bolenz C, Herrmann E, Bastian PJ, Michel MS, Wulffing C, Tiemann A, et al. Lymphovascular invasion is an independent predictor of oncological outcomes in patients with lymph node-negative urothelial bladder cancer treated by radical cystectomy: a multicentre validation trial. *BJU Int.* 2010;106(4):493–9.
32. Algaba F. Lymphovascular invasion as a prognostic tool for advanced bladder cancer. *Curr Opin Urol.* 2006;16(5):367–71.
33. Reuter VE. Lymphovascular invasion as an independent predictor of recurrence and survival in node-negative bladder cancer remains to be proven. *J Clin Oncol.* 2005;23(27):6450–1.
34. Park K, Tomlins SA, Mudaliar KM, Chiu YL, Esgueva R, Mehra R, et al. Antibody-based detection of ERG rearrangement-positive prostate cancer. *Neoplasia.* 2010;12(7):590–8. P
35. Kahn HJ, Marks A. A new monoclonal antibody, D2–40, for detection of lymphatic invasion in primary tumors. *Lab Invest.* 2002;82(9):1255–7.
36. Epstein JI, Amin MB, Reuter VR, Mostofi FK. The world health organization/international society of urological pathology consensus classification of urothelial (transitional cell) neoplasms of the urinary bladder. Bladder consensus conference committee. *Am J Surg Pathol.* 1998;22(12):1435–48.
37. Paner GP, Shen SS, Lapetino S, Venkataraman G, Barkan GA, Quek ML, et al. Diagnostic utility of antibody to smoothelin in the distinction of muscularis propria from muscularis mucosae of the urinary bladder: a potential ancillary tool in the pathologic staging of invasive urothelial carcinoma. *Am J Surg Pathol.* 2009;33(1):91–8.
38. Council L, Hameed O. Differential expression of immunohistochemical markers in bladder smooth muscle and myofibroblasts, and the potential utility of desmin, smoothelin, and vimentin in staging of bladder carcinoma. *Mod Pathol.* 2009;22(5):639–50.
39. Paner GP, Brown JG, Lapetino S, Nese N, Gupta R, Shen SS, et al. Diagnostic use of antibody to smoothelin in the recognition of muscularis propria in transurethral resection of urinary bladder tumor (TURBT) specimens. *Am J Surg Pathol.* 2010;34(6):792–9.
40. Hansel DE, Paner GP, Nese N, Amin MB. Limited smoothelin expression within the muscularis mucosae: validation in bladder diverticula. *Hum Pathol.* 2011;42(11):1770–6.
41. Khayyata S, Dudas M, Rohan SM, Gopalan A, Fine SW, Reuter VE, et al. Distribution of smoothelin expression in the musculature of the genitourinary tract. *Lab Invest.* 2009;89:175a–a.

42. Miyamoto H, Sharma RB, Illei PB, Epstein JI. Pitfalls in the use of smoothelin to identify muscularis propria invasion by urothelial carcinoma. *Am J Surg Pathol*. 2010;34(3):418–22.
43. Chen YB, Tu JJ, Kao J, Zhou XK, Chen YT. Survivin as a useful adjunct marker for the grading of papillary urothelial carcinoma. *Arch Pathol Lab Med*. 2008;132(2):224–31.
44. Yin W, Chen N, Zhang Y, Zeng H, Chen X, He Y, et al. Survivin nuclear labeling index: a superior biomarker in superficial urothelial carcinoma of human urinary bladder. *Mod Pathol*. 2006;19(11):1487–97.
45. Karam JA, Lotan Y, Ashfaq R, Sagalowsky AI, Shariat SF. Survivin expression in patients with non-muscle-invasive urothelial cell carcinoma of the bladder. *Urology*. 2007;70(3):482–6.
46. Schultz IJ, Kiemeny LA, Karthaus HF, Witjes JA, Willems JL, Swinkels DW, et al. Survivin mRNA copy number in bladder washings predicts tumor recurrence in patients with superficial urothelial cell carcinomas. *Clin Chem*. 2004;50(8):1425–8.
47. Moussa O, Abol-Enein H, Bissada NK, Keane T, Ghoneim MA, Watson DK. Evaluation of survivin reverse transcriptase-polymerase chain reaction for noninvasive detection of bladder cancer. *J Urol*. 2006;175(6):2312–6.
48. Eissa S, Badr S, Barakat M, Zaghoul AS, Mohanad M. The diagnostic efficacy of urinary survivin and hyaluronidase mRNA as urine markers in patients with bladder cancer. *Clin Lab*. 2013;59(7–8):893–900.
49. Dalbagni G, Presti JC Jr., Reuter VE, Zhang ZF, Sarkis AS, Fair WR, et al. Molecular genetic alterations of chromosome 17 and p53 nuclear overexpression in human bladder cancer. *Diagn Mol Pathol*. 1993;2(1):4–13.
50. Fujimoto K, Yamada Y, Okajima E, Kakizoe T, Sasaki H, Sugimura T, et al. Frequent association of p53 gene mutation in invasive bladder cancer. *Cancer Res*. 1992;52(6):1393–8.
51. Sidransky D, Von Eschenbach A, Tsai YC, Jones P, Summerhayes I, Marshall F, et al. Identification of p53 gene mutations in bladder cancers and urine samples. *Science*. 1991;252(5006):706–9.
52. Cordon-Cardo C, Waringer D, Petrylak D, Dalbagni G, Fair WR, Fuks Z, et al. Altered expression of the retinoblastoma gene product: prognostic indicator in bladder cancer. *J Natl Cancer Inst*. 1992;84(16):1251–6.
53. Cairns P, Proctor AJ, Knowles MA. Loss of heterozygosity at the RB locus is frequent and correlates with muscle invasion in bladder carcinoma. *Oncogene*. 1991;6(12):2305–9.
54. Iyer G, Al-Ahmadie H, Schultz N, Hanrahan AJ, Ostrovnaya I, Balar AV, et al. Prevalence and co-occurrence of actionable genomic alterations in high-grade bladder cancer. *J Clin Oncol*. 2013;31(25):3133–40.
55. Milowsky MI, Iyer G, Regazzi AM, Al-Ahmadie H, Gerst SR, Ostrovnaya I, et al. Phase II study of everolimus in metastatic urothelial cancer. *BJU Int*. 2013;112(4):462–70.
56. Iyer G, Hanrahan AJ, Milowsky MI, Al-Ahmadie H, Scott SN, Janakiraman M, et al. Genome sequencing identifies a basis for everolimus sensitivity. *Science*. 2012;338(6104):221.
57. Al-Ahmadie HA, Iyer G, Janakiraman M, Lin O, Heguy A, Tickoo SK, et al. Somatic mutation of fibroblast growth factor receptor-3 (FGFR3) defines a distinct morphological subtype of high-grade urothelial carcinoma. *J Pathol*. 2011;224(2):270–9.
58. Chou J, Provot S, Werb Z. GATA3 in development and cancer differentiation: cells GATA have it! *J Cell Physiol*. 2010;222(1):42–9.
59. Olsburgh J, Harnden P, Weeks R, Smith B, Joyce A, Hall G, et al. Uroplakin gene expression in normal human tissues and locally advanced bladder cancer. *J Pathol*. 2003;199(1):41–9.
60. Kageyama S, Yoshiki T, Isono T, Tanaka T, Kim CJ, Yuasa T, et al. High expression of human uroplakin Ia in urinary bladder transitional cell carcinoma. *Jpn J Cancer Res*. 2002;93(5):523–31.
61. Mhaweche P, Uchida T, Pelte MF. Immunohistochemical profile of high-grade urothelial bladder carcinoma and prostate adenocarcinoma. *Hum Pathol*. 2002;33(11):1136–40.
62. Xu X, Sun TT, Gupta PK, Zhang P, Nasuti JF. Uroplakin as a marker for typing metastatic transitional cell carcinoma on fine-needle aspiration specimens. *Cancer*. 2001;93(3):216–21.
63. Ordonez NG. Thrombomodulin expression in transitional cell carcinoma. *Am J Clin Pathol*. 1998;110(3):385–90.
64. Sokolova IA, Halling KC, Jenkins RB, Burkhardt HM, Meyer RG, Seelig SA, et al. The development of a multitarget, multicolor fluorescence in situ hybridization assay for the detection of urothelial carcinoma in urine. *J Mol Diagn*. 2000;2(3):116–23.
65. Hajdinjak T. UroVysion FISH test for detecting urothelial cancers: meta-analysis of diagnostic accuracy and comparison with urinary cytology testing. *Urol Oncol*. 2008;26(6):646–51.
66. Ferra S, Denley R, Herr H, Dalbagni G, Jhanwar S, Lin O. Reflex UroVysion testing in suspicious urine cytology cases. *Cancer*. 2009;117(1):7–14.
67. Burchardt M, Burchardt T, Shabsigh A, De La Taille A, Benson MC, Sawczuk I. Current concepts in biomarker technology for bladder cancers. *Clin Chem*. 2000;46(5):595–605.
68. Heicappell R, Wettig IC, Schostak M, Muller M, Steiner U, Sauter T, et al. Quantitative detection of human complement factor H-related protein in transitional cell carcinoma of the urinary bladder. *Eur Urol*. 1999;35(1):81–7.
69. Murphy WM, Rivera-Ramirez I, Medina CA, Wright NJ, Wajzman Z. The bladder tumor antigen (BTA) test compared to voided urine cytology in the detection of bladder neoplasms. *J Urol*. 1997;158(6):2102–6.
70. Sanchez-Carbayo M, Herrero E, Megias J, Mira A, Espasa A, Chinchilla V, et al. Initial evaluation

- of the diagnostic performance of the new urinary bladder cancer antigen test as a tumor marker for transitional cell carcinoma of the bladder. *J Urol*. 1999;161(4):1110–5.
71. Berezney R, Coffey DS. Identification of a nuclear protein matrix. *Biochem Biophys Res Commun*. 1974;60(4):1410–7.
 72. Fey EG, Krochmalnic G, Penman S. The nonchromatin substructures of the nucleus: the ribonucleoprotein (RNP)-containing and RNP-depleted matrices analyzed by sequential fractionation and resinless section electron microscopy. *J Cell Biol*. 1986;102(5):1654–65.
 73. Budman LI, Kassouf W, Steinberg JR. Biomarkers for detection and surveillance of bladder cancer. *Can Urol Assoc J*. 2008;2(3):212–21.
 74. Chang YH, Wu CH, Lee YL, Huang PH, Kao YL, Shiau MY. Evaluation of nuclear matrix protein-22 as a clinical diagnostic marker for bladder cancer. *Urology*. 2004;64(4):687–92.
 75. Fradet Y, Lockhard C. Performance characteristics of a new monoclonal antibody test for bladder cancer: immunoCyt trade mark. *Can J Urol*. 1997;4(3):400–5.
 76. Tetu B, Tiguert R, Harel F, Fradet Y. ImmunoCyt/uCyt+ improves the sensitivity of urine cytology in patients followed for urothelial carcinoma. *Mod Pathol*. 2005;18(1):83–9.
 77. Sullivan PS, Nooraie F, Sanchez H, Hirschowitz S, Levin M, Rao PN, et al. Comparison of immunoCyt, UroVysion, and urine cytology in detection of recurrent urothelial carcinoma: a “split-sample” study. *Cancer*. 2009;117(3):167–73.
 78. Muller M. Telomerase: its clinical relevance in the diagnosis of bladder cancer. *Oncogene*. 2002;21(4):650–5.
 79. Kim NW, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PL, et al. Specific association of human telomerase activity with immortal cells and cancer. *Science*. 1994;266(5193):2011–5.
 80. Weikert S, Krause H, Wolff I, Christoph F, Schrader M, Emrich T, et al. Quantitative evaluation of telomerase subunits in urine as biomarkers for noninvasive detection of bladder cancer. *Int J Cancer*. 2005;117(2):274–80.
 81. Xylinas E, Kluth LA, Rieken M, Karakiewicz PI, Lotan Y, Shariat SF. Urine markers for detection and surveillance of bladder cancer. *Urol Oncol*. 2014;32(3):222–9.