## Chapter 9 Newer Pulmonary Function Tests

Graham L. Hall and Paul D. Robinson

**Abstract** The measurement of lung function is an integral component of respiratory medicine. In the past 10–15 years there has been significant progress in the development of newer lung function tests such that there are now standardized guidelines and commercially available equipment for some of these techniques. This chapter focuses on the forced oscillation technique, the interrupter technique and the multiple breath washout test and their application in preschool and school aged children and the potential role of these tests in the diagnosis and management of children with respiratory disease. A primary advantage of these tests is the relatively minimal level of cooperation that is required to obtained acceptable measurements thus making them ideally suited for use in children as young as 2–3 years of age. This creates opportunities to introduce objective measurements of respiratory function at a significantly younger age than previously possible. The aim of this chapter is to provide the reader with an overview of each of these tests and to summarize the evidence that these tests can be used to monitor changes in clinical status.

**Keywords** Multiple breath washout • Lung clearance index • Forced oscillation technique • Interrupter technique • Respiratory physiology • Lung function testing • Children

Discipline of Paediatrics and Child Health, Faculty of Medicine, The Children's Hospital at Westmead Clinical School, University of Sydney, Sydney, NSW, Australia

The Woolcock Institute of Medical Research, Sydney, NSW, Australia e-mail: paul.robinson1@health.nsw.gov.au

G.L. Hall, Ph.D., F.R.A.N.Z.S.R.S. (⊠) Telethon Kids Institute, University of Western Australia, PO Box 855, West Perth, WA 6872, Australia

Respiratory Medicine, Princess Margaret Hospital for Children, Perth, WA, Australia e-mail: graham.hall@telethonkids.org.au

P.D. Robinson, M.B.Ch.B., F.R.A.C.P., Ph.D. Respiratory Medicine, The Children's Hospital at Westmead, Locked Bag 4001, Sydney, NSW 2145, Australia

#### Introduction

The objective measurement of respiratory function has been an integral component of respiratory medicine for decades. The most commonly used lung function tests in the pediatric respiratory function laboratory will include spirometry, static lung volumes (usually by plethysmography), and the gas transfer of carbon monoxide. As highlighted elsewhere in this book a disadvantage of these lung function tests is the difficulty in using them in infants and young children primarily due to the active cooperation required to achieve acceptable and repeatable outcomes with these tests. In the past 10–15 years there has been significant progress in the development of newer lung function tests from techniques employed in a limited number of specialized research centers with research prototypes to tests able to be performed in a standard pediatric respiratory function laboratory with commercially available equipment. These tests include the raised volume rapid thoracic compression technique and body plethysmography for use in infants; the forced oscillation technique (FOT), the interrupter technique, specific airway resistance and the multiple breath washout (MBW) technique suitable for use in both infants and young children. This chapter will review FOT, the MBW test and the interrupter technique and their application in preschool and school aged children and the potential role of these tests in the diagnosis and management of children with respiratory disease. Readers interested in infant lung function tests are directed to Chap. 9.

These newer lung function tests offer new ways to understand, diagnose, and manage respiratory disease in children. A primary advantage of these tests is the relatively minimal level of cooperation that is required to obtain acceptable measurements thus making them ideally suited for use in children as young as 2–3 years of age. This creates opportunities to introduce objective measurements of respiratory function at a significantly younger age than previously possible. However, there are limitations with each of these tests and their role in the clinical management of individual patients is not clear.

The aim of this chapter is to provide the reader with an overview of each lung function test, including a summary of test protocols; definitions of acceptable and repeatable test outcomes; currently available reference data; the current knowledge of a clinically meaningful difference for each of the tests; and the evidence that these tests can be used to monitor changes in clinical status.

#### **Forced Oscillation Technique**

The forced oscillation technique (FOT) was first described by Dubois and colleagues in 1956 [1]. Since that initial description the technique has been applied extensively in preclinical animal models (as reviewed by Sly et al. [2]), infants (for example [3–7]), and preschool and school aged children as well as adults of all ages (as reviewed in [8, 9]). Oscillatory equipment has advanced from discrete single center research prototypes to being commercially available for both animal based research and human research and clinical testing.

The FOT uses the application of a forcing signal to the respiratory system, most commonly at the mouth, to quantify respiratory system mechanics. At the most basic level FOT is similar to that of a ventilator applied sine wave to an intubated patient and the use of the resultant pressures, flows and volumes to derive respiratory system resistance (Rrs) and compliance (being the inverse of respiratory system elastance (Ers)). In practice the majority of research and commercial oscillatory equipment apply a forcing signal that covers a range of frequencies with the response to this signal being the respiratory system impedance (Zrs). The respiratory system impedance is comprised of the Rrs and the respiratory reactance (Xrs) across the range of frequencies being measured. The Rrs represents the resistive elements of the respiratory system, including the airways, lung, and chest wall; however, the airway resistance represents the largest component of Rrs. The respiratory reactance includes both the elastic properties of the lung at lower frequencies and the inertive properties of the airways at higher frequencies. These components are opposite in sign and the frequency at which the elastic and inertive properties are equal (but opposite) is the resonant frequency (Fres) which occurs at the point that Xrs equals zero [10]. Readers seeking detailed information on technical aspects of the technique are directed to comprehensive reviews [9, 10].

#### The Forced Oscillation Technique Methodology

There are commercially available FOT systems and the most commonly reported are the CareFusion Impulse oscillation system (IOS) and the Cosmed Quark I2M. For the purposes of this chapter the generic term FOT is used to mean all forced oscillation systems and individual commercial systems are only identified if there are equipment specific issues.

The FOT is usually applied in awake individuals but has also been reported in anesthetized children [11] and has been combined with continuous positive airway pressure ventilation [12]. As the focus of this review is the application of the FOT in the pediatric setting we have limited the methodological description below to that used in preschool and school aged children. There are no current international standards for the measurement of forced oscillatory mechanics similar to those used for other lung function tests [13–15]. However, guidelines for the use of FOT in clinical practice [9] and in preschool children [8] are available and readers are encouraged to consult these for full details.

Generally, FOT measurements are performed in a sitting upright position with the subject's head in the midline. Children maintain normal tidal breathing through a mouthpiece (usually incorporating a bacterial filter) while wearing a nose clip. To minimize the impact of shunting in the highly compliant extra-thoracic airways, the cheeks and floor of the mouth need to be firmly supported [16, 17]. In young children this is preferably performed by a staff member to maximize test quality (as seen in Fig. 9.1), but can be performed by a parent. In older children this can be performed by the children themselves.



**Fig. 9.1** Forced oscillation measurement in a 5-year-old child. Children should be seated upright with their head in the midline and neutral position. The child breathes through a mouthpiece incorporating a bacterial filter with a nose clip in place. Firm support of the cheeks and roof of the mouth is important to minimize the pressure loss of the oscillatory signal

#### Acceptability and Repeatability of FOT Measurements

Acceptable measurements should be free of artifact including leak, swallowing, mouth movements, talking and other noises and obstruction of the mouthpiece with the child's tongue. These criteria can be assessed through visual inspection of the Zrs spectra and the individual pressure and flow recordings. In young children additional feedback from the staff member supporting the cheeks and the floor of the mouth can be very helpful. A minimum of three to five acceptable measurements should be obtained and the average and standard deviation (SD) of all acceptable measurements are reported [8, 9]. The most commonly reported FOT outcomes are the Rrs and Xrs at individual specific frequencies and reported as Rrsf and Xrsf (for example Rrs and Xrs at 8 Hz are denoted as Rrs8 and Xrs8), the resonant frequency (Fres) and the area under the reactance curve (AX; defined as the area under the Xrs curve from a defined frequency to the resonant frequency) [18]. Figure 9.2 illustrates these commonly reported outcomes in a healthy child and a child with cystic fibrosis (CF).

To date there is insufficient evidence to allow the definition of intra-test repeatability that can be used to state that a test session is repeatable. The mean within-test coefficient of variability (CV: defined as the standard deviation (SD)/mean and expressed as a percentage) has been reported as ranging between 5 and 10 % for Rrs and up to 20 % for Xrs [19–22]. Until such time as definitive criteria for repeatability are available users should retain all acceptable measurements and exercise caution when deciding to exclude apparently acceptable data based on repeatability criteria alone.



**Fig. 9.2** Respiratory impedance spectra from a healthy child (shown in *blue*) and a child with cystic fibrosis (shown in *red*) of similar age and heights. The respiratory resistance (Rrs: *solid line*) tends to be increased and show negative frequency dependence at lower frequencies while respiratory reactance (Xrs: *dashed lines*) is decreased (more negative) in children with lung disease. Commonly reported FOT outcomes include the Rrs and Xrs at a specific frequency (shown here as Rrs and Xrs at 8 Hz: Rrs8 and Xrs8, respectively). The resonant frequency (Fres) is the frequency at which Xrs is equal to zero and represents the point at which the respiratory system recoil (or compliance) and inertance (reflecting the properties of the central airways) are balance. The area under the reactance curve (AUX) is the sum of the Xrs curve from the lowest frequency measured to the resonant frequency

The between test (inter-test) coefficient of repeatability (twice the SD of the difference between two measurements) in healthy children ranges between 1.1 and 2.6 hPa s/L for Rrs and equates to a relative change of 12-30 % [23–25] with similar short- and long term repeatability reported in children with lung disease [19, 20, 26]. The repeatability of Xrs is reported as absolute value due to the proximity of Xrs values to zero and ranges from 1.2 to 2.0 hPa s/L [19, 20, 23–26].

#### Reference Ranges in Preschool and School Aged Children Using Forced Oscillations

There is a range of reference equations available for use in preschool and school aged children and to date the majority of these studies are in Caucasian children [24, 27–29] although reference data in children from Mexico [30], Iran [31], Korea [32], and Vietnam [33] are now available. It is not clear what impact ethnicity will have

on reported FOT outcomes and this area deserves attention. Not all studies reporting reference equations have reported all of the FOT outcomes outlined above. It is important to note that the frequency range of the FOT outcomes reported by the CareFusion IOS is different to that of the Cosmed Quark I2M and as such reference equations are currently equipment specific. Users should carefully review the available studies for suitability relating to local equipment and patient populations.

# The Role of the Forced Oscillation Technique in Clinical Practice

The use of the FOT in the management of individual children with lung disease remains unclear with only limited information on its potential use. The primary reasons for this shortage of studies assessing the clinical role of the FOT are likely to be the historical lack of standardized methodological guidelines and availability of commercial equipment. The clinical utility of the FOT in young children with recurrent wheeze and/or asthma, CF and in those children born preterm with or without bronchopulmonary dysplasia (BPD) has recently been comprehensively reviewed as part of an ATS workshop report on optimal lung function tests in young children [34], while the role of the FOT in older children with lung disease remains to be formally assessed.

The change in FOT outcomes following bronchodilator inhalation considered to be clinically relevant is reasonably consistent across multiple studies. A clinically relevant bronchodilator response is defined using the 5th/95th centiles of the response to bronchodilators in healthy populations. These have been reported to be between -33 and -42 % for Rrs, 61 and 70 % for Xrs and approximately 80 % for AX, irrespective of the dose of the salbutamol [25, 27, 33, 35–37]. The ability of the FOT to assist in the diagnosis and/or management of asthmatic and/or wheezy children either before or after bronchodilator inhalation remains unclear. Some studies demonstrated no differences in the baseline lung function or bronchodilator responsiveness between healthy and asthmatic children [22, 35–37]. In contrast, other studies have reported that FOT may provide some benefit in the identification of children with asthma [38–41]. One study from Shi et al. [42] suggest that FOT outcomes may predict loss of asthma control and therefore may have a role in the management of children with asthma and further work in this area is needed.

The FOT has been used to assess lung function in children with CF; however, the majority of these studies have been cross-sectional with only limited assessment of pulmonary infection or inflammation and therefore the ability of FOT to assist in the clinical management of younger children with CF is unclear. Brennan et al. assessed the relationship of respiratory mechanics derived from the low-frequency FOT [43] in a group of infants and young children with CF at the time of an annual bronchoal-veolar lavage and reported increased pulmonary inflammation, but not infection, was associated with increased respiratory resistance. However, this variation of FOT is not easily transferred to a clinical setting. Most studies using commercially

available FOT equipment suggest that the FOT is not sensitive to the early CF lung disease either cross-sectionally or longitudinally [26, 44–48]. In contrast, increased Rrs and decreased Xrs have been reported in young children with CF in the presence of respiratory symptoms [19].

The primary impact of preterm birth and BPD on the respiratory system is likely to be the peripheral lung with altered alveolar structure [49]. As such the FOT should be a particularly suitable test for use in infants and children following preterm birth, particularly those born very preterm (<32 weeks gestational age). Despite this the FOT has not been used widely in infants and children born preterm. Using the low-frequency FOT Pillow et al. demonstrated that oscillatory mechanics can be measured in preterm infants around term [3]. Studies using commercially available equipment have demonstrated that FOT outcomes are abnormal in both preschool and school aged preterm children, both with and without BPD and that these differences are more pronounced in measures of respiratory reactance than resistance suggesting that the FOT is sensitive to the altered peripheral lung pathophysiology evident in these children [20, 50, 51]. There are limited reports of the FOT being applied in children with upper airway dysfunction and further research in this patient group is required [52–54].

#### **Future Work and Conclusions**

The measurement of respiratory system impedance has the potential to provide a great deal of information on a variety of conditions during the early years of life; however, further work is required if the FOT is yet to reach its full clinical potential. In the short term it is important to identify which FOT outcomes are most sensitive to each specific pediatric lung disease with the knowledge that the pathophysiological mechanisms of many respiratory diseases exhibit strong peripheral lung involvement during early life. In the longer term it is important to gain an understanding of how respiratory mechanics alters longitudinally during development and what kind of deviation from this path requires intervention.

#### The Multiple Breath Washout Technique

Inert gas washout was first described over 60 years ago following the advent of fast responding gas analysers [55, 56], but it was not until the development of personal computers that breath-by-breath analysis became feasible and enabled MBW analysis techniques as we know them today. The choice of a suitable inert gas for MBW testing should consider if the inert tracer gas is safe for patients to inhale and does not participate in gas exchange or dissolve significantly in the blood or other tissues. Inert gases may either be resident within the lung (i.e., present within room air, e.g., Nitrogen ( $N_2$ ) or Argon) or non-resident (e.g., Sulfur hexafluoride, SF<sub>6</sub>, or Helium).

Inert gas washout tests allow the distribution of ventilation to be assessed. Ventilation within the lung is determined by the structure of the respiratory airway tree and the gas exchange unit or alveoli. Gas transport and mixing by convection (i.e., bulk flow) predominates in the conducting airways. In the lung periphery, bulk flow is minimal and gas transport by molecular diffusion dominates. In the region of the entry of the acinus, the relative contributions of convection and diffusion to gas mixing are equal, generating a "diffusion-convection front" in the healthy adult lung [57]. Pathological processes affecting the dimensions of these peripheral airways in a *heterogeneous*, or patchy, manner, affect the distribution of ventilation. It is this *unevenness* of ventilation that is detected by tests such as the MBW technique. A typical MBW test is performed over a series of tidal breaths, requiring minimal cooperation or coordination, offering feasibility across the *entire* pediatric age range. Improved sensitivity to detect lung disease, in comparison to conventional spirometry, across a number of important pediatric lung diseases [58–61].

#### The Technique Methodology

The MBW examines the pattern by which an inert gas is washed out of the lungs during tidal breathing. Standardization guidelines for equipment validation, MBW test performance and subsequent analysis have recently been developed for use in all age groups and are covered in more detail elsewhere [62]. Inert gas washout is based on accurate measurement of respiratory flow and inert gas concentration signals, which need to be correctly aligned in time prior to subsequent analyses. The majority of equipment in publications prior to the early to mid-2000s has been custom-made and based on a variety of flow measurement devices and inert gas analysers. In recent years commercial systems have been developed, based on SF<sub>6</sub> measurement for the infant age range (Exhalyzer D, ECO Medics AG, Switzerland) or on N<sub>2</sub> measurement suitable for preschoolers and above (Exhalyzer D, ECO Medics AG, Switzerland; Easyone Pro, ndd Medical Technologies, Switzerland). Recent advances in equipment validation [63] appear to have provided robust clinical devices suitable for widespread use.

For the patient, minimal cooperation and coordination are required: an adequate mouthpiece/facemask seal must be maintained to prevent leaks, and a regular breathing pattern needs to be maintained. A facemask is used in infants and may be used for preschool children due to difficulty maintaining an adequate mouthpiece seal. A regular breathing pattern is achieved in infants by performing the test during natural sleep or under sedation (in the supine position), or in older children, sitting upright and using distraction with an interesting video [62].

In N<sub>2</sub>-based MBW the nitrogen is washed out of the lung by switching the patient into breathing 100 % O<sub>2</sub>. Non-resident inert gases require an additional wash-in phase during which the inhaled inert gas concentration (typically 4 % SF<sub>6</sub> or 4 % He with 21 % O<sub>2</sub> and the balance N<sub>2</sub>) equilibrates within the lung, before being washed out by breathing room air. Initial MBW studies demonstrated feasibility using N<sub>2</sub> as the inert gas [64–66], but inhalation of pure O<sub>2</sub> in infants was subsequently shown to alter normal tidal breathing patterns [67], with potential effects on subsequent calculated indices. As a result interest in non-resident inert marker gases increased, with strong feasibility subsequently demonstrated for SF<sub>6</sub>based MBW [58, 68–70]. The clinical utility of the MBW may be hampered by the test duration which may be up to an hour in some children. Recent efforts to shorten the MBW protocol suggest the test duration can be shortened [71, 72]. Feasibility within the busy clinical environment and in more remote settings has also recently been demonstrated [73, 74].

#### Available MBW Indices

Each MBW test is conventionally performed until the end-tidal inert gas concentration reaches 1/40 of its starting concentration (i.e., approximately 2 % for N<sub>2</sub> MBW and 0.1 % for SF<sub>6</sub>/He MBW at the concentrations mentioned prior) (Fig. 9.3). Three technically acceptable tests should be the target of each testing session. In addition to providing information about ventilation distribution, lung volume data (functional residual capacity, FRC), trapped gas and measures of the volume of the conducting airways (Fowler [75] and Langley [76] airways dead space) can also be determined. These ventilation inhomogeneity parameters can be grouped into those reflecting the presence of overall global abnormalities, such as the Lung Clearance Index (LCI) [77] and moment ratios [78], and those based on more sophisticated phase III slope analysis providing additional information about the location of any abnormality [79].



**Fig. 9.3** Real time tracings from a MBW test. Tidal volume (*upper panel*) and inert gas SF<sub>6</sub> concentration (*lower panel*) are displayed. The washout phase of the MBW test commences after equilibration of SF<sub>6</sub> concentration within the lungs to approximately 4 % and continues until SF<sub>6</sub> concentration has decreased to below 1/40th or 0.1 %

LCI is the simplest parameter to calculate, the easiest for physicians and patients/ parents to understand conceptually, and the most popular index reported in the recent pediatric literature. It represents the number of lung volume turnovers (TO, or FRCs) required to clear the lungs of the inert marker gas to 1/40 of the starting concentration. Moment ratios (calculated using an approach termed "moment analysis") are more complicated and describe the degree of skewing of the washout curve, with increased skewing representing increased release of inert gas at a later stage of the washout [78]. Calculation of moment ratios are described in more detail elsewhere [80]. The proposed advantage of moment analysis is that it may compensate for the impact of variation in respiratory rate and tidal volume during the washout [65, 81].

Phase III slope analysis from MBW is the main index reported in the adult literature, and has been developed from theoretical [79], experimental [82, 83], and adult lung modelling data [83–85], to separate ventilation inhomogeneity arising from conductive airway zones (termed convection-dependent inhomogeneity or CDI) and within more distal acinar zones (where diffusion and convection interact to generate inhomogeneity, termed diffusion-convection-dependent inhomogeneity or DCDI). The CDI and DCDI can be expressed as the clinical indices,  $S_{\text{cond}}$  and  $S_{\text{acin}}$ , respectively [86]. The original method for calculation proposed in adults required a strict breathing protocol (tidal volumes 1.0-1.3 L and breathing rate of 10-12 breaths/ min), to try to avoid variation in pre-inspiratory lung volume, inspired and expired volumes and flow [57, 87-94] affecting the magnitude of the phase III slope. Adaptation of this method for pediatric testing, where a structured breathing protocol is not feasible <16 years, has required the incorporation of a tidal volume compensation for phase III slope values [62]. The use of the strict adult breathing protocol in children has been shown to significantly affect MBW outcomes [45]. Phase III slope parameters do not appear as robust as LCI to variations in breathing pattern, which compromises feasibility to a degree in the preschool age range, whilst the low parenchymal-to-airway volume ratio encountered in infants makes identification and accurate assessment of a phase III slope from tidal breath expirograms challenging. These indices remain exploratory in the pediatric range at present and have not demonstrated the same degree of clinical utility to date as LCI.

#### Acceptability and Repeatability of MBW Measurements

The majority of the data presented in this section is based on SF<sub>6</sub>-based MBW, whereas the recent availability of commercial N<sub>2</sub>-based equipment is starting to generate equivalent data. Strong feasibility exists across all age ranges: in infants 76–90 % during natural sleep (aged <6 weeks) [95, 96] and 87 % in sedated infants [70], compared to 80 % in preschoolers (ranging from 50 % of 2–3-year-olds to 87 % of 5–6-year-olds) [58]. Within-session variability ranges from 3 to 8 % across studies in preschoolers and above, with between-session variability approximately 5 % [97], equating to a statistically significant change of ±1 TO for LCI. A collaborative multiple center approach has led to publication of recent SF<sub>6</sub> mass

spectrometer based normative data for the entire pediatric age range [98]. Recently preliminary data describing normative  $N_2$ -based LCI data has also been published from 7 to 70 years of age [99]. Both these sets of normative data illustrate the higher LCI values encountered at the extremes of age (e.g., infancy) but show largely stable values in between, making LCI an attractive longitudinal tool.

#### The Role of LCI in Clinical Practice

LCI is now strongly supported as a research tool in clinical studies in CF, due to a large body of evidence supporting potential clinical utility in this disease group [51]. The heterogeneous distribution of CF lung disease has been nicely illustrated by recent imaging studies [100], and includes the peripheral airways [101].

Abnormal LCI values are frequently present in CF subjects, from infancy onwards, and disparity between health and disease appears to increase over time, based on this cross-sectional data. Improved sensitivity of LCI to detect lung disease despite normal spirometry exists across a number of studies from the preschool age range onwards [58, 60, 69, 102]. Strong correlation between LCI and high resolution computed tomography (HRCT) measured structural lung damage has also been described in several studies, from infancy [103–106]. Prognostic value is also starting to emerge, with preschool LCI values predicting those subjects with abnormal spirometry at school age [107]. LCI improves with treatment intervention [108–110], and its potential to detect significant changes despite small cohort sizes is exciting for future intervention studies. However, heterogeneity of response encountered in established disease [80] may limit utility in more severe subjects.

As in CF, the heterogeneous distribution of the disease process in asthma, involving the *entire* airway tree, not just the central airways, has been demonstrated in imaging and pathology studies [111–113], predicting potential utility for MBW. Pediatric asthmatics have increased global ventilation inhomogeneity (LCI and moment ratio values), compared to controls [65, 114]. Phase III analysis describes a predominant conducting airways pattern of abnormality (elevated  $S_{cond}$ ) [115]. Improved sensitivity to detect disease involvement, in comparison to spirometry, is suggested by both pathological gas trapping in childhood and adolescent asthmatics [116], and elevated LCI values in well controlled childhood asthmatics [61], despite normal baseline spirometry in both groups. Whether this represents established airway remodelling is unclear. Increased ventilation inhomogeneity has also been suggested as an important mechanism for airway hyperresponsiveness in adult asthmatics [113, 117].

In younger preschool recurrent wheeze phenotypes, MBW also appears to differentiate between multi-trigger wheeze and viral-induced wheeze phenotypes (increased LCI and  $S_{cond}$  values) [118]. Establishing a predictive value to detect those who subsequently develop classical asthma will require longitudinal studies. Utility in challenge testing is suggested by the ability to detect a marked peripheral airways response, missed by spirometry, on challenge testing [119], but the time-consuming nature of the test compromises routine clinical utility in this setting. The same applies for assessment of bronchodilator response. Response to treatment with therapy targeting the peripheral airways (e.g., fine particle inhaled corticosteroid) has been demonstrated in adults [120], but corresponding pediatric data is lacking.

Discussion of clinical utility of MBW in infants with BPD illustrates the importance of disease distribution when assessing potential utility. "Old" BPD crosssectional studies describe elevated gas trapping (as assessed by  $FRC_{pleth}$  and  $FRC_{N2}$ ) and LCI (or moment ratios) in BPD infants during the newborn period, in comparison to healthy controls [81, 121]. However, recent larger multicenter studies, of "new" BPD, show little difference between groups [95, 96]. This probably reflects both improved study design, adequately correcting for important confounders such as prematurity and intrauterine growth, and the more diffuse nature of "new" BPD, generating less marked ventilation unevenness.

#### **Future Work and Conclusions**

The availability of robust commercial equipment and improved standardization of the MBW technique represent essential steps if exciting potential utility suggested by research studies to date is to translate into feasibility in the busy clinical laboratory. Strongest utility is currently suggested for CF, but may also exist for asthma and other disease groups. Efforts in the future to shorten test duration are important and evaluation of current commercial equipment is ongoing. Longitudinal studies will provide a clearer idea of true utility.

#### The Interrupter Technique

The interrupter technique allows the measurement of the resistance of the respiratory system, including the airway tree, lung tissue and chest wall. The primary advantage of the technique is that it requires minimal cooperation from the individual being tested and hence is of particular interest in infants and young children. There have been a number of studies conducted in sedated and unsedated sleeping infants [122–128] and spontaneously breathing children as young as 2 years [24, 129–133]. Historically, there was little standardization of the technique. The availability of commercial equipment and the release of guidelines for its use in preschool children [8] should lead to an improved understanding of its role in the clinical management of young children [34].

The classical description of the interrupter technique involves the rapid occlusion of the airway opening and the measurement of the flow immediately preceding the interruption and the changes in airway opening pressure (Pao) following the interruption. The interrupter resistance (Rint) is derived from the change in Pao by the flow. An alternative approach derives the Rint using the change in Pao and the flow immediately after the interruption [134]. The outcomes derived from these two approaches differ and cannot be used interchangeably [135]. The classical approach is the most commonly used and unless specified otherwise is the technique for which details are provided in this chapter.

#### The Interrupter Technique Methodology

Commercial equipment for both the classical and the alternative methods of measuring the Rint are available. The interrupter technique assumes that following the interruption to airflow the alveolar pressure rapidly equilibrates with the airway opening and that the change in Pao reflects the pressure drop across the whole airway tree and therefore equates to the alveolar pressure [128, 136, 137]. Following occlusion two distinct phases are seen in the Pao trace (Fig. 9.4). There is an initial rapid rise in Pao that reflects the resistive drop across the airway tree and a component of the resistive properties of the lung and chest wall. This is followed by a slower rise to a plateau and reflects the stress relaxation of the respiratory tissues.

The Rint is influenced by the closure time of the occlusion valve [128], the compliance of the upper airways [128], the type of patient interface (face mask or mouthpiece) used [126, 138, 139] and the method of determining the change in Pao following the interruption [126, 140, 141]. The most commonly used approach for the derivation of the Pao is to use the linear back extrapolation of Pao between 30 and 70 ms following the interruption [8].

In infants Rint is measured with the infant in a supine position during quiet sleep. The facemask is placed over the nose and mouth of the infant and supported firmly to minimize pressure loss across the cheeks and floor of the mouth [126]. The use of equipment designed for older children and not adapted for infants is not recommended, with low success rates and poor repeatability of Rint reported [122, 123].



Recommendations for the measurement of Rint in young children are available and in the absence of detailed guidelines in older children should be used for all measurements of interrupter resistance in cooperative children [8]. In preschool and school aged children measurements are made with the child seated and looking directly ahead while breathing through a mouthpiece and with a nose clip in place and the cheeks firmly supported during measurements. The airway is occluded during expiration with the interrupter valve for a period of 100 ms at a flow equating to the peak tidal expiratory flow. A minimum of ten interruptions should be obtained with at least five acceptable measurements retained.

#### Acceptability and Repeatability of Interrupter Technique Measurements

Measurements should show a smooth increase in airway opening pressure to a peak and be free of leak or unusual changes in pressure and/or flow at the time of the interruption. The median of all acceptable measurements is reported. The withintest repeatability of Rint in healthy infants is dependent on the analysis technique [126] and the mean (or median) coefficient of variation has been reported to range 16.6–19.3 % [124–126] The data available for the between test repeatability of Rint in infants is scarce with Fuchs et al. reporting the mean between test difference in 22 unsedated 6 week old infants to be 1.4 hPa s/L with the limits of agreement being –21.2 and 23.9 hPa s/L [124].

In contrast there are many studies that have assessed the within- and between-test repeatability of Rint in preschool and school aged children and these are extensively reviewed elsewhere [8, 34]. The coefficient of variability of a single testing session ranges between 10 and 12 % of the median Rint value [133, 142–145]. The short term (15–30 min) and longer term (2 weeks to up to 3 months) repeatability of the Rint has been assessed in healthy children and in children with lung disease. The coefficient of repeatability (equating to two times the standard deviation of the measurement) ranges from 1.7 to 4.4 hPa s/L [132, 146, 147].

### Reference Ranges for Interrupter Resistance in Infants, Preschool and School Aged Children

Fuchs et al. [124] reported upper limits of normal for Rint in unsedated infants aged 5–7 weeks of age. To date only one study reported reference ranges for the Rint in sedated infants over a broader age range [125]. Measurements of Rint in healthy preschool and school aged children have been reported in Caucasian children [24, 132, 133, 143, 148] as well as children of differing ethnic backgrounds [125, 148–150]. Collated reference values for the classical technique in children aged 3–13 years have been developed and offer the most robust references ranges currently available [151].

#### The Role of the Interrupter Technique in Clinical Practice

The potential role for measurements of Rint in infants has not been assessed. The lack of appropriate commercially available equipment and standardized methodology guidelines in this age group limit the ability of health professionals and researchers to assess the ability of Rint to assist in the diagnosis and/or management of infants with lung disease.

The clinical role of Rint in preschool children has been extensively reviewed by the ATS workshop report on optimal lung function tests in young children [34]. While studies reporting measurements of Rint in older, school aged children show responsible agreement with other clinically available lung function tests [144] its primary role is likely to be in younger children unable to perform these tests accurately.

The limits of agreement for the determination of a clinically relevant bronchodilator response in Rint obtained with the classical technique are similar and are reported as a decrease in Rint of >2.5 hPa s/L or >32–35 % of baseline [143, 152, 153].

The majority of studies using the interrupter technique have been in children with a history of recurrent wheeze or cough and asthma. The majority of studies have reported that children with asthma have increased Rint compared to healthy controls; however, the proportion of asthmatic children with Rint outside of the normal range varies significantly and these differences are likely to relate to subject selection (community versus hospital/clinic asthmatics) and current asthma treatments [36, 131, 133, 144, 145, 152]. Children with asthma or recurrent wheeze tend to have larger decreases in Rint following bronchodilator inhalation when compared to healthy children, and studies have reported sensitivity and specificity ranges from 24 to 76 % and 70 to 92 %, respectively to distinguish between asthmatic and healthy children [36, 148, 152]. However, each of these studies expressed the bronchodilator responses differently making direct comparisons difficult. A number of investigators have assessed the Rint in placebo controlled clinical trials of asthma medications [131, 154-156] with significant improvements in Rint seen in some [131, 155] but not all trials [154, 156], and provide early evidence that Rint may be a useful clinical trial outcome measure in young children with asthma.

Studies measuring Rint in children with CF are primarily cross-sectional [26, 142, 157–160] with only two studies tracking Rint longitudinally [26, 160]. While children with CF tend to have increased Rint when compared to healthy children, there is a large overlap and in the longitudinal studies Rint was not altered with changing clinical status. The sensitivity of Rint to predict pulmonary infection and/ or exacerbations or to track the effectiveness of treatments in children with CF has not been assessed.

There are only limited data using the interrupter technique in children born preterm. Vrijlandt et al. reported increased Rint in preterm children with and without BPD when compared to healthy controls. However, there were no significant differences in the baseline Rint or response to bronchodilators between the preterm groups [50]. In a group of preterm children attending tertiary respiratory clinics Kairamkonda and coworkers reported preterm children with a history of BPD had significantly increased Rint compared to preterm children without BPD [161]. As described previously the likely anatomical site of altered physiology in children born preterm will be in the peripheral lung and the emerging evidence that respiratory reactance from the FOT (a measure of peripheral lung function) has increased sensitivity compared to respiratory resistance suggests that the clinical utility of the interrupter technique in preterm children may be limited.

#### **Future Work and Conclusions**

Further development of the interrupter technique in infants is required including the most appropriate triggering flow, interruption duration and airway opening pressure analysis method. In preschool children the data available to date suggest the interrupter technique is going to be most informative in children with a history of recurrent wheeze and in the assessment of airway responsiveness including both bronchodilator responsiveness and inhaled bronchial challenge tests. Future studies should focus on longitudinal measurement of Rint and prospectively collected clinical information to provide a better understanding of the sensitivity of Rint to changing clinical status.

#### Summary

The advent of newer pulmonary function tests and in particular the availability of commercial equipment and standardized guidelines has provided renewed interest in the objective measurement of lung function in pediatric lung disease. The ability of these tests to allow the quantification of lung function from as young as 2 years of age provides a window of opportunity for pediatric health professionals and researchers alike to accurately characterize the natural history of pediatric lung disease and in the diagnosis and management of children with a range of respiratory disorders.

The current status for each of these tests is similar as is the way forward. Robust reference ranges across a range of ethnicities are required. Longitudinal studies incorporating some or all of these techniques are required to allow appropriate conclusions to be drawn on the relative merits of each test within the context of the disease pathophysiology and the determination of the minimal meaningful clinical difference to assist in the management of our patients with lung disease.

#### References

- 1. Dubois AB, et al. Oscillation mechanics of lungs and chest in man. J Appl Physiol. 1956;8:587–94.
- Sly PD, et al. Measuring lung function in murine models of pulmonary disease. Drug Discov Today Dis Models. 2004;1:337–43.

#### 9 Newer Pulmonary Function Tests

- 3. Pillow JJ, et al. Partitioning of airway and parenchymal mechanics in unsedated newborn infants. Pediatr Res. 2005;58:1210–5.
- 4. Wohl ME, et al. Resistance of the total respiratory system in healthy infants and infants with bronchiolitis. Pediatrics. 1969;43:495–509.
- 5. Frey U, et al. High-frequency respiratory impedance measured by forced-oscillation technique in infants. Am J Respir Crit Care Med. 1998;158:363–70.
- Hall GL, et al. Altered respiratory tissue mechanics in asymptomatic wheezy infants. Am J Respir Crit Care Med. 2001;164:1387–91.
- Sly PD, et al. Measurement of low-frequency respiratory impedance in infants. Am J Respir Crit Care Med. 1996;154:161–6.
- Beydon N, et al. An official American Thoracic Society/European Respiratory Society statement: pulmonary function testing in preschool children. Am J Respir Crit Care Med. 2007;175:1304–45.
- 9. Oostveen E, et al. The forced oscillation technique in clinical practice: methodology, recommendations and future developments. Eur Respir J. 2003;22:1026–41.
- 10. Bates JHT, et al. Oscillation mechanics of the respiratory system. Compr Physiol. 2011;1:1233–72.
- 11. Petak F, et al. Airway and tissue mechanics in anesthetized paralyzed children. Pediatr Pulmonol. 2003;35:169–76.
- 12. Farre R, et al. A system to generate simultaneous forced oscillation and continuous positive airway pressure. Eur Respir J. 1997;10:1349–53.
- 13. Macintyre N, et al. Standardisation of the single-breath determination of carbon monoxide uptake in the lung. Eur Respir J. 2005;26:720–35.
- 14. Miller MR, et al. Standardisation of spirometry. Eur Respir J. 2005;26:319-38.
- 15. Wanger J, et al. Standardisation of the measurement of lung volumes. Eur Respir J. 2005;26:511–22.
- 16. Cauberghs M, Van de Woestijne KP. Effect of upper airway shunt and series properties on respiratory impedance measurements. J Appl Physiol. 1989;66:2274–9.
- Peslin R, et al. Respiratory impedance measured with head generator to minimize upper airway shunt. J Appl Physiol. 1985;59:1790–5.
- Goldman MD, et al. Clinical applications of forced oscillation to assess peripheral airway function. Respir Physiol Neurobiol. 2005;148:179–94.
- Gangell CL, et al. Respiratory impedance in children with cystic fibrosis using forced oscillations in clinic. Eur Respir J. 2007;30:892–7.
- 20. Udomittipong K, et al. Forced oscillations in the clinical setting in young children with neonatal lung disease. Eur Respir J. 2008;31:1292–9.
- Lall CA, et al. Airway resistance variability and response to bronchodilator in children with asthma. Eur Respir J. 2007;30:260–8.
- Harrison J, et al. Lung function in preschool children with a history of wheezing measured by forced oscillation and plethysmographic specific airway resistance. Pediatr Pulmonol. 2010;45:1049–56.
- Hall GL, et al. Respiratory function in healthy young children using forced oscillations. Thorax. 2007;62:521–6.
- Klug B, Bisgaard H. Specific airway resistance, interrupter resistance, and respiratory impedance in healthy children aged 2–7 years. Pediatr Pulmonol. 1998;25:322–31.
- Malmberg LP, et al. Determinants of respiratory system input impedance and bronchodilator response in healthy Finnish preschool children. Clin Physiol Funct Imaging. 2002;22: 64–71.
- Nielsen KG, et al. Serial lung function and responsiveness in cystic fibrosis during early childhood. Am J Respir Crit Care Med. 2004;169:1209–16.
- Calogero C, et al. Respiratory impedance and bronchodilator responsiveness in healthy children aged 2–13 years. Pediatr Pulmonol. 2013;48(7):707–15.
- Dencker M, et al. Reference values for respiratory system impedance by using impulse oscillometry in children aged 2–11 years. Clin Physiol Funct Imaging. 2006;26:247–50.

- 29. Frei J, et al. Impulse oscillometry: reference values in children 100 to 150 cm in height and 3 to 10 years of age. Chest. 2005;128:1266–73.
- Shackleton C, et al. Reference ranges for Mexican preschool-aged children using the forced oscillation technique. Arch Bronconeumol. 2013;49:326–9.
- Amra B, et al. Respiratory resistance by impulse oscillometry in healthy Iranian children aged 5–19 years. Iran J Allergy Asthma Immunol. 2008;7:25–9.
- 32. Lee JY, et al. Reference values of impulse oscillometry and its utility in the diagnosis of asthma in young Korean children. J Asthma. 2012;49:811–6.
- Vu LT, et al. Respiratory impedance and response to salbutamol in healthy Vietnamese children. Pediatr Pulmonol. 2008;43:1013–9.
- 34. Rosenfeld M, et al. An Official American Thoracic Society Workshop report: optimal lung function tests for monitoring cystic fibrosis, bronchopulmonary dysplasia and recurrent wheezing in children <6 years of age. Ann Am Thorac Soc. 2013;10:S1–11.
- Hellinckx J, et al. Bronchodilator response in 3–6.5 years old healthy and stable asthmatic children. Eur Respir J. 1998;12:438–43.
- 36. Nielsen KG, Bisgaard H. Discriminative capacity of bronchodilator response measured with three different lung function techniques in asthmatic and healthy children aged 2 to 5 years. Am J Respir Crit Care Med. 2001;164:554–9.
- Thamrin C, et al. Assessment of bronchodilator responsiveness in preschool children using forced oscillations. Thorax. 2007;62:814–9.
- 38. Komarow HD, et al. A study of the use of impulse oscillometry in the evaluation of children with asthma: analysis of lung parameters, order effect, and utility compared with spirometry. Pediatr Pulmonol. 2012;47:18–26.
- 39. Oostveen E, et al. Lung function and bronchodilator response in 4-year-old children with different wheezing phenotypes. Eur Respir J. 2010;35:865–72.
- 40. Shin YH, et al. Oscillometric and spirometric bronchodilator response in preschool children with and without asthma. Can Respir J. 2012;19:273–7.
- 41. Vu LT, et al. Respiratory impedance and response to salbutamol in asthmatic Vietnamese children. Pediatr Pulmonol. 2010;45:380–6.
- 42. Shi Y, et al. Peripheral airway impairment measured by oscillometry predicts loss of asthma control in children. J Allergy Clin Immunol. 2013;131:718–23.
- 43. Brennan S, et al. Correlation of forced oscillation technique in preschool children with cystic fibrosis with pulmonary inflammation. Thorax. 2005;60:159–63.
- 44. Ren CL, et al. Analysis of the associations between lung function and clinical features in preschool children with cystic fibrosis. Pediatr Pulmonol. 2012;47:574–81.
- 45. Yammine S, et al. Impact of different breathing protocols on multiple-breath washout outcomes in children. J Cyst Fibros. 2014;13:190–7.
- 46. Mazurek HK, et al. Specificity and sensitivity of respiratory impedance in assessing reversibility of airway obstruction in children. Chest. 1995;107:996–1002.
- 47. Lebecque P, Stanescu D. Respiratory resistance by the forced oscillation technique in asthmatic children and cystic fibrosis patients. Eur Respir J. 1997;10:891–5.
- 48. Kerby GS, et al. Lung function distinguishes preschool children with CF from healthy controls in a multi-center setting. Pediatr Pulmonol. 2012;47:597–605.
- Jobe AH, Bancalari E. Bronchopulmonary dysplasia. Am J Respir Crit Care Med. 2001;163:1723–9.
- 50. Vrijlandt EJ, et al. Respiratory health in prematurely born preschool children with and without bronchopulmonary dysplasia. J Pediatr. 2007;150:256–61.
- 51. Kent L, et al. Lung clearance index: evidence for use in clinical trials in cystic fibrosis. J Cyst Fibros. 2014;13:123–38.
- 52. Harrison J, et al. Lung function in children with repaired tracheo-oesophageal fistula using the forced oscillation technique. Pediatr Pulmonol. 2010;45:1057–63.
- Hoijer U, et al. The ability of noninvasive methods to detect and quantify laryngeal obstruction. Eur Respir J. 1991;4:109–14.
- 54. Rigau J, et al. Oscillometric assessment of airway obstruction in a mechanical model of vocal cord dysfunction. J Biomech. 2004;37:37–43.

- 55. Siri WE. A mass spectroscope for analysis in the low mass range. Rev Sci Instrum. 1947;18:540.
- Lilly JC. Mixing of gases within respiratory system with a new type nitrogen meter. Am J Physiol. 1950;161:342–51.
- 57. Paiva M. Gas transport in the human lung. J Appl Physiol. 1973;35:401-10.
- Aurora P, et al. Multiple-breath washout as a marker of lung disease in preschool children with cystic fibrosis. Am J Respir Crit Care Med. 2005;171:249–56.
- 59. Fuchs SI, et al. A novel sidestream ultrasonic flow sensor for multiple breath washout in children. Pediatr Pulmonol. 2008;43:731–8.
- Gustafsson PM, et al. Evaluation of ventilation maldistribution as an early indicator of lung disease in children with cystic fibrosis. Eur Respir J. 2003;22:972–9.
- 61. Macleod KA, et al. Ventilation heterogeneity in children with well controlled asthma with normal spirometry indicates residual airways disease. Thorax. 2009;64:33–7.
- Robinson PD, et al. Consensus statement for inert gas washout measurement using multipleand single-breath tests. Eur Respir J. 2013;41:507–22.
- 63. Singer F, et al. A realistic validation study of a new nitrogen multiple-breath washout system. PLoS One. 2012;7:e36083.
- 64. Wall MA. Moment analysis of multibreath nitrogen washout in young children. J Appl Physiol. 1985;59:274–9.
- 65. Kraemer R, Meister B. Fast real-time moment-ratio analysis of multibreath nitrogen washout in children. J Appl Physiol. 1985;59:1137–44.
- Hjalmarson O, Sandberg K. Abnormal lung function in healthy preterm infants. Am J Respir Crit Care Med. 2002;165:83–7.
- 67. Schibler A, et al. Moment ratio analysis of multiple breath nitrogen washout in infants with lung disease. Eur Respir J. 2000;15:1094–101.
- 68. Schibler A, et al. Measurement of lung volume and ventilation distribution with an ultrasonic flow meter in healthy infants. Eur Respir J. 2002;20:912–8.
- 69. Aurora P, et al. Multiple breath inert gas washout as a measure of ventilation distribution in children with cystic fibrosis. Thorax. 2004;59:1068–73.
- 70. Lum S, et al. Early detection of cystic fibrosis lung disease: multiple-breath washout versus raised volume tests. Thorax. 2007;62:341–7.
- Robinson PD, et al. Abbreviated multi-breath washout for calculation of lung clearance index. Pediatr Pulmonol. 2013;48:336–43.
- 72. Yammine S, et al. Multiple-breath washout measurements can be significantly shortened in children. Thorax. 2013;68:586–7.
- Gray DM, Willemse L, Alberts A, Simpson S, Sly PD, Hall GL, Zar HJ. Lung function in African infants: a pilot study. Pediatr Pulmonol. DOI 10.1002/ppul.22965. 2013.
- Singer F, et al. Practicability of nitrogen multiple-breath washout measurements in a pediatric cystic fibrosis outpatient setting. Pediatr Pulmonol. 2013;48:739–46.
- Fowler WS. Lung function studies; the respiratory dead space. Am J Physiol. 1948;154:405–16.
- 76. Langley F, et al. Ventilatory consequences of unilateral pulmonary artery occlusion. Les Colloques des l'Institut National de la Sante' et de la Recherche Me'dicale. 1975;51: 209–12.
- Bouhuys A. Pulmonary nitrogen clearance in relation to age in healthy males. J Appl Physiol. 1963;18:297–300.
- 78. Saidel GM, et al. Moment analysis of multibreath lung washout. J Appl Physiol. 1975;38:328–34.
- Paiva M. Two new pulmonary functional indexes suggested by a simple mathematical model. Respiration. 1975;32:389–403.
- Robinson PD, et al. Inert gas washout: theoretical background and clinical utility in respiratory disease. Respiration. 2009;78:339–55.
- Shao H, et al. Impaired gas mixing and low lung volume in preterm infants with mild chronic lung disease. Pediatr Res. 1998;43:536–41.

- 82. Paiva M, et al. Slope of phase III in multibreath nitrogen washout and washin. Bull Eur Physiopathol Respir. 1982;18:273–80.
- 83. Verbanck S, Paiva M. Model simulations of gas mixing and ventilation distribution in the human lung. J Appl Physiol. 1990;69:2269–79.
- Butrieue B, et al. A human acinar structure for simulation of realistic alveolar plateau slopes. J Appl Physiol. 2000;89:1859–67.
- 85. Paiva M, Engel LA. Model analysis of gas distribution within human lung acinus. J Appl Physiol Respir Environ Exerc Physiol. 1984;56:418–25.
- Verbanck S, et al. Ventilation distribution during histamine provocation. J Appl Physiol. 1997;83:1907–16.
- 87. Cumming G, et al. The influence of gaseous diffusion on the alveolar plateau at different lung volumes. Respir Physiol. 1967;2:386–98.
- Fowler WS. Lung function studies; uneven pulmonary ventilation in normal subjects and in patients with pulmonary disease. J Appl Physiol. 1949;2:283–99.
- Jones JG. The effect of preinspiratory lung volume on the result of the single breath O<sub>2</sub> test. Respir Physiol. 1967;2:375–85.
- Crawford AB, et al. Effect of airway closure on ventilation distribution. J Appl Physiol. 1989;66:2511–5.
- 91. Crawford AB, et al. Effect of lung volume on ventilation distribution. J Appl Physiol. 1989;66:2502–10.
- 92. Lacquet LM, van Muylem A. He and SF6 single-breath expiration curves. Comparison with the paiva-engel model. Bull Eur Physiopathol Respir. 1982;18:239–46.
- 93. Crawford AB, et al. Effect of tidal volume on ventilation maldistribution. Respir Physiol. 1986;66:11–25.
- 94. Gronkvist M, et al. Effects of body posture and tidal volume on inter- and intraregional ventilation distribution in healthy men. J Appl Physiol. 2002;92:634–42.
- 95. Hulskamp G, et al. Association of prematurity, lung disease and body size with lung volume and ventilation inhomogeneity in unsedated neonates: a multicentre study. Thorax. 2009;64: 240–5.
- 96. Latzin P, et al. Lung volume, breathing pattern and ventilation inhomogeneity in preterm and term infants. PLoS One. 2009;4:e4635.
- 97. Fuchs SI, et al. Lung clearance index: normal values, repeatability, and reproducibility in healthy children and adolescents. Pediatr Pulmonol. 2009;44:1180–5.
- 98. Lum S, et al. Age and height dependence of lung clearance index and functional residual capacity. Eur Respir J. 2013;41(6):1371–7.
- 99. Houltz B, et al. Tidal  $N_2$  washout ventilation inhomogeneity indices in a reference population aged 7–70 years. Eur Respir J. 2012;40:694s.
- van Beek EJ, et al. Assessment of lung disease in children with cystic fibrosis using hyperpolarized 3-Helium MRI: comparison with Shwachman score, Chrispin-Norman score and spirometry. Eur Radiol. 2007;17:1018–24.
- Hamutcu R, et al. Clinical findings and lung pathology in children with cystic fibrosis. Am J Respir Crit Care Med. 2002;165:1172–5.
- 102. Horsley AR, et al. Lung clearance index is a sensitive, repeatable and practical measure of airways disease in adults with cystic fibrosis. Thorax. 2008;63:135–40.
- 103. Gustafsson PM, et al. Multiple-breath inert gas washout and spirometry versus structural lung disease in cystic fibrosis. Thorax. 2008;63:129–34.
- 104. Owens CM, et al. Lung clearance index and HRCT are complementary markers of lung abnormalities in young children with CF. Thorax. 2011;66:481–8.
- 105. Ellemunter H, et al. Sensitivity of lung clearance index and chest computed tomography in early CF lung disease. Respir Med. 2010;104:1834–42.
- 106. Hall GL, et al. Air trapping on chest CT is associated with worse ventilation distribution in infants with cystic fibrosis diagnosed following newborn screening. PLoS One. 2011;6:e23932.
- 107. Aurora P, et al. Lung clearance index at 4 years predicts subsequent lung function in children with cystic fibrosis. Am J Respir Crit Care Med. 2011;183:752–8.

- 108. Robinson PD, et al. Using index of ventilation to assess response to treatment for acute pulmonary exacerbation in children with cystic fibrosis. Pediatr Pulmonol. 2009;44:733–42.
- Amin R, et al. Hypertonic saline improves the LCI in paediatric CF patients with normal lung function. Thorax. 2010;65:379–83.
- Amin R, et al. The effect of dornase alfa on ventilation inhomogeneity in patients with cystic fibrosis. Eur Respir J. 2011;37:806–12.
- 111. Hamid Q, et al. Inflammation of small airways in asthma. J Allergy Clin Immunol. 1997;100:44-51.
- 112. Carroll N, et al. The structure of large and small airways in nonfatal and fatal asthma. Am Rev Respir Dis. 1993;147:405–10.
- 113. Venegas JG, et al. Self-organized patchiness in asthma as a prelude to catastrophic shifts. Nature. 2005;434:777–82.
- 114. Saniie J, et al. Real-time moment analysis of pulmonary nitrogen washout. J Appl Physiol. 1979;46:1184–90.
- Gustafsson PM, et al. Peripheral airway involvement in asthma assessed by single-breath SF6 and He washout. Eur Respir J. 2003;21:1033–9.
- 116. Gustafsson PM, et al. Pneumotachographic nitrogen washout method for measurement of the volume of trapped gas in the lungs. Pediatr Pulmonol. 1994;17:258–68.
- 117. Downie SR, et al. Ventilation heterogeneity is a major determinant of airway hyperresponsiveness in asthma, independent of airway inflammation. Thorax. 2007;62:684–9.
- 118. Sonnappa S, et al. Symptom-pattern phenotype and pulmonary function in preschool wheezers. J Allergy Clin Immunol. 2010;126:519–26.
- 119. Aljassim F, et al. A whisper from the silent lung zone. Pediatr Pulmonol. 2009;44:829-32.
- Verbanck S, et al. The functional benefit of anti-inflammatory aerosols in the lung periphery. J Allergy Clin Immunol. 2006;118:340–6.
- 121. Wauer RR, et al. Assessment of functional residual capacity using nitrogen washout and plethysmographic techniques in infants with and without bronchopulmonary dysplasia. Intensive Care Med. 1998;24:469–75.
- Adams AM, et al. Measurement and repeatability of interrupter resistance in unsedated newborn infants. Pediatr Pulmonol. 2009;44:1168–73.
- 123. Chavasse RJ, et al. Comparison of resistance measured by the interrupter technique and by passive mechanics in sedated infants. Eur Respir J. 2001;18:330–4.
- 124. Fuchs O, et al. Normative data for lung function and exhaled nitric oxide in unsedated healthy infants. Eur Respir J. 2011;37:1208–16.
- 125. Gochicoa LG, et al. Reference values for airway resistance in newborns, infants and preschoolers from a Latin American population. Respirology. 2012;17:667–73.
- Hall GL, et al. Evaluation of the interrupter technique in healthy, unsedated infants. Eur Respir J. 2001;18:982–8.
- 127. Lanteri CJ, Sly PD. Changes in respiratory mechanics with age. J Appl Physiol. 1993;74:369–78.
- 128. Sly PD, Bates JH. Computer analysis of physical factors affecting the use of the interrupter technique in infants. Pediatr Pulmonol. 1988;4:219–24.
- 129. Bridge PD, et al. Measurement of airway resistance using the interrupter technique in preschool children in the ambulatory setting. Eur Respir J. 1999;13:792–6.
- 130. McKenzie SA, et al. Airway resistance and atopy in preschool children with wheeze and cough. Eur Respir J. 2000;15:833–8.
- 131. Nielsen KG, Bisgaard H. The effect of inhaled budesonide on symptoms, lung function, and cold air and methacholine responsiveness in 2- to 5-year-old asthmatic children. Am J Respir Crit Care Med. 2000;162:1500–6.
- 132. Lombardi E, et al. Reference values of interrupter respiratory resistance in healthy preschool white children. Thorax. 2001;56:691–5.
- 133. Merkus PJ, et al. Interrupter resistance in preschool children: measurement characteristics and reference values. Am J Respir Crit Care Med. 2001;163:1350–5.
- Bisgaard H, Klug B. Lung function measurement in awake young children. Eur Respir J. 1995;8:2067–75.

- 135. Oswald-Mammosser M, et al. The opening interrupter technique for respiratory resistance measurements in children. Respirology. 2010;15:1104–10.
- 136. Bates JH, et al. A theoretical analysis of interrupter technique for measuring respiratory mechanics. J Appl Physiol. 1988;64:2204–14.
- Bates JH, et al. Interrupter resistance elucidated by alveolar pressure measurement in openchest normal dogs. J Appl Physiol. 1988;65:408–14.
- 138. Oswald-Mammosser M, et al. Measurements of respiratory system resistance by the interrupter technique in healthy and asthmatic children. Pediatr Pulmonol. 1997;24:78–85.
- 139. Thamrin C, Frey U. Effect of bacterial filter on measurement of interrupter resistance in preschool and school-aged children. Pediatr Pulmonol. 2008;43:781–7.
- 140. Phagoo SB, et al. Accuracy and sensitivity of the interrupter technique for measuring the response to bronchial challenge in normal subjects. Eur Respir J. 1993;6:996–1003.
- 141. Phagoo SB, et al. Evaluation of the interrupter technique for measuring change in airway resistance in 5-year-old asthmatic children. Pediatr Pulmonol. 1995;20:387–95.
- 142. Beydon N, et al. Pulmonary function tests in preschool children with cystic fibrosis. Am J Respir Crit Care Med. 2002;166:1099–104.
- 143. Beydon N, et al. Pre/postbronchodilator interrupter resistance values in healthy young children. Am J Respir Crit Care Med. 2002;165:1388–94.
- 144. Beydon N, et al. Baseline and post-bronchodilator interrupter resistance and spirometry in asthmatic children. Pediatr Pulmonol. 2012;47:987–93.
- 145. Beydon N, et al. Interrupter resistance short-term repeatability and bronchodilator response in preschool children. Respir Med. 2007;101:2482–7.
- 146. Beelen RM, et al. Short and long term variability of the interrupter technique under field and standardised conditions in 3–6 year old children. Thorax. 2003;58:761–4.
- 147. Chan EY, et al. Repeatability of airway resistance measurements made using the interrupter technique. Thorax. 2003;58:344–7.
- 148. McKenzie SA, et al. Airway resistance measured by the interrupter technique: normative data for 2–10 year olds of three ethnicities. Arch Dis Child. 2002;87:248–51.
- 149. Li AM, et al. Interrupter respiratory resistance in healthy Chinese preschool children. Chest. 2009;136:554–60.
- Rech VV, et al. Airway resistance in children measured using the interrupter technique: reference values. J Bras Pneumol. 2008;34:796–803.
- 151. Merkus PJ, et al. Reference ranges for interrupter resistance technique: the Asthma UK Initiative. Eur Respir J. 2010;36:157–63.
- 152. Beydon N, et al. Pulmonary function tests in preschool children with asthma. Am J Respir Crit Care Med. 2003;168:640–4.
- 153. Mele L, et al. Assessment and validation of bronchodilation using the interrupter technique in preschool children. Pediatr Pulmonol. 2010;45:633–8.
- 154. Kooi EM, et al. Fluticasone or Montelukast for preschool children with asthma-like symptoms: randomized controlled trial. Pulm Pharmacol Ther. 2008;21:798–804.
- 155. Pao CS, McKenzie SA. Randomized controlled trial of fluticasone in preschool children with intermittent wheeze. Am J Respir Crit Care Med. 2002;166:945–9.
- 156. Schokker S, et al. Inhaled corticosteroids for recurrent respiratory symptoms in preschool children in general practice: randomized controlled trial. Pulm Pharmacol Ther. 2008;21:88–97.
- 157. Carter ER, et al. Evaluation of the interrupter technique for the use of assessing airway obstruction in children. Pediatr Pulmonol. 1994;17:211–7.
- 158. Davies PL, et al. The interrupter technique to assess airway responsiveness in children with cystic fibrosis. Pediatr Pulmonol. 2007;42:23–8.
- 159. Oswald-Mammosser M, et al. Interrupter technique versus plethysmography for measurement of respiratory resistance in children with asthma or cystic fibrosis. Pediatr Pulmonol. 2000;29:213–20.
- Terheggen-Lagro SW, et al. Radiological and functional changes over 3 years in young children with cystic fibrosis. Eur Respir J. 2007;30:279–85.
- 161. Kairamkonda VR, et al. Lung function measurement in prematurely born preschool children with and without chronic lung disease. J Perinatol. 2008;28:199–204.