

Chapter 8

Multiple Sclerosis and Microbes

8.1 Introduction

Multiple sclerosis [MS] is a chronic demyelinating, immune-mediated disorder of the brain and spinal cord of unknown etiology. Although MS is not considered a common disease as such, it is not a rare disorder and it is estimated to be the second commonest cause of neurological disability after traumatic injuries. There is a marked geographical variation in the incidence of the disease, more prevalent in temperate regions and rare in tropical and subtropical countries. In the United States MS affects about 35,000 persons and worldwide 2.5 million people are afflicted [1]. In most temperate regions of the world [North America, Northern Europe, Southern Australia, and South New Zealand] the prevalence of MS is 0.1–0.2 % of the population, whereas in the tropics and Middle East the prevalence is 10- to 20-fold less [1]. The age of onset of MS is typically between 20 and 40 years, about 5–10 % occurs in children less than 18 years, and women are affected threefold more common than men.

Based on the geographical variation it has been proposed that sunlight exposure is protective through the production of vitamin D. Low serum vitamin D levels are common in the population of temperate zones, and prospective studies show that vitamin D deficiency is associated with a greater risk of MS and for relapses [1]. The clinical and pathological extent of MS is very variable and heterogenous, and the course is often characterized by spontaneous relapses and remissions early in the disease.

8.2 Pathobiology of Multiple Sclerosis

Initially in the early stages of MS, patchy inflammation with focal lymphocytic infiltration is the primary pathological feature in the brain, which leads to damage of the myelin and axons [2]. The inflammation is often transient with remyelination and recovery of neurological dysfunction early in the course. Over time the predominant

pathological changes consist of widespread microglial activation with extensive chronic neurodegeneration and plaques of demyelination [2]. Normally myelin is produced by mature oligodendrocytes which are adjacent to axons of the white matter tracts in the central nervous system [CNS].

It is believed that MS is initiated by some environmental factor that stimulates autoreactive lymphocytes in the CNS. Although MS is considered an autoimmune disease, transfer of antibodies directed against self-antigens [identified in MS] failed to cause MS-like disease in animals [3]. Despite that, T-cells reactive to myelin components, especially myelin basic protein [MBP], are activated in MS patients but not in controls [4]. There is some evidence that uncontrolled autoreactive lymphocytes may induce inflammation [predominantly by perivascular CD8+ cells] to cause neuronal damage due to dysfunction of regulatory lymphocytes and regulatory mechanisms in the CNS of MS patients [5]. Failure of regulatory lymphocytes to suppress autoreactive T-cells appears to be related to overexpression of β -arrestin1, which is a promoter of naïve and activated CD4+ T-cell survival [6]. Previous animal models of experimental allergic encephalomyelitis had supported a critical role of Th1-type γ secreting cells [3], but recent studies indicate that the inflammation in MS is driven by T-lymphocyte subtype that secretes interleukin [IL]-17 under IL-23 control [7].

Myelin proteins may not be the only target of autoreactive lymphocytes, and there is evidence that antibodies against neurofascin may mediate axonal injury in MS [8]; and autoimmune response against α B-crystalline prevents counter-regulatory suppression of inflammation [9]. Cortical biopsies of brain lesions in early MS have revealed perivascular inflammation with CD3+ and CD8+ T cells in the majority of cases [which were highly inflammatory], and 27 % of cortical plaques also contained B-cells [10]. In this study [10] cortical demyelination was present in 40 % of patients and 66 % of the lesions contained foamy macrophages, and all had activated microglia, indicating ongoing demyelination.

8.2.1 Pathogenesis of Multiple Sclerosis

The pathogenic mechanisms of MS are complex and involve multiple genetic and environmental factors. Epidemiological studies have implicated increased risk of MS by gender, sex hormones, ethnic origin, geographical location/latitude/distance from the equator, smoking, viral exposure, and vitamin D status [2, 11–13]. In family studies, first-degree relatives showed a 20- to 40-fold increased risk for MS, and identical twins display 300-fold increased risk over the general population [14]. Genetic studies have reported that genes in the major histocompatibility complex [HLA] region are associated with MS [15]. The primary association was with DRB1 gene in African Americans and individuals of European descent [16, 17]. Further genome-wide association studies have reported about 50 genes associated with MS [18]. However, MS concordance rate in monozygotic twins is only approximately 30 %

and this suggests that environmental factors have a major influence on genetic trait. This would be consistent with epistatic interaction, where two or more independent factors promote disease only when combined [19]. In a large genome-wide association study, DRB1 risk alleles had the strongest association with MS, and HLA-A gene variation had a protective effect [18]. A multitude of genes encoding cytokine pathways and immune related mechanisms were overrepresented, particularly those implicated in T-helper cell differentiation, and acting on cell surface receptors. This study also implicated genes encoding pathways for vitamin D function and targets for therapies for MS, such as VCAM1 [natalizumab] and IL2RA [daclizumab] [18]. Several of the genes are also associated with other autoimmune diseases [IL2RA and IL7RA], and these pathways are involved in regulation of autoimmunity in animal models [20, 21]. It has also been suggested that several genetic variants [IL7RA, IL2RA, MGAT1, and CTLA-4] lead to dysregulation of N-glycosylation that cause pathogenesis in MS [22]. Faulty N-glycosylation of cytotoxic T lymphocyte antigen 4 [CTLA-4] and T-cell receptor [TCR] generates T-cell hyperactivity and promotes autoimmunity in mice, which induces a spontaneous MS-like disease [23, 24].

It has also been proposed that environmental factors may regulate disease manifestation by modulating the epigenome in MS to promote changes in the immune system and brain [25]. These epigenetic mechanisms include DNA methylation, regulation of noncoding miRNAs, and post-translational modification of histone that can be affected by smoking, diet, exercise, and possible previous infection.

8.2.2 Hypovitaminosis D in MS

Geographical regions of the world with limited sunshine for 4–6 months of the year, beyond the 40th parallels North or South, are those with the highest prevalence of MS [26]. Several epidemiological studies have found a vitamin D insufficiency in the great majority of MS patients, including in the early stages [27]. Vitamin D receptors are present in numerous tissues and cells, including circulating immunity cells [lymphocytes, macrophages, and monocytes], brain [microglia], intestine, bone, kidney, gonads, breast, pancreas, and cardiovascular tissues. Besides its classic role in calcium homeostasis and related metabolic functions, vitamin D and its active metabolite 1,25-dihydroxy vitamin D have other important functions: anti-inflammatory, anti-infective, immunomodulatory, antiproliferative, and neurotransmitter, which may be involved to prevent many autoimmune diseases, including MS [27]. The major action of vitamin D that may be important in the pathogenesis of MS is through immunomodulation. Hypovitaminosis D may affect cell proliferation of CD4+ T cells, the proportion and function of regulatory T lymphocytes [28–30]. Thus, in the final global analysis, the current data is supportive of hypovitaminosis D as a risk factor for MS, but acting in combination with other environmental factors in the genetically predisposed individuals.

8.3 Role of Microbes in Multiple Sclerosis

Viruses have been suggested to play a role in the development of MS since the early 1990s, and this debate still continues today [31]. There are several lines of evidence that link viral infections with MS and these include: (1) epidemiological evidence which consistently demonstrate increased risk of MS with some past infections; (2) the CD8+ lymphocyte infiltration in MS lesions is consistent with a viral-immune response; (3) and the CSF oligoclonal IgG bands typically present in MS are also present in CNS viral infections. Moreover, many naturally occurring demyelinating CNS diseases of animals and humans are of known viral origin. This topic of virus-induced demyelination was previously reviewed in 2003 [32]. There are two primary mechanisms by which viruses produce demyelinating CNS disease. The first of these is by an autoimmune process, as exemplified by postinfectious [also postvaccination] encephalomyelitis, which is preceded by a viral infection [i.e., an exanthema] with no evidence of direct invasion of the CNS [33]. For some viral demyelinating diseases there is direct CNS invasion and replication of the agent, with neuronal and axonal pathology. A very rare but well described example is subacute sclerosing panencephalitis due to chronic measles infection of the brain at an early age, <2 years old [33]. In this condition the measles virus can be recovered from the brain and histology demonstrates cytoplasmic and nuclear inclusion bodies, with signs of astrocytes and microglia activation and neuronal loss. Measles is also one of several viruses that can present more acutely with postinfectious encephalomyelitis.

In postinfectious encephalomyelitis the hypersensitivity reaction may occur both against viral and host antigen. In a study of measles postinfectious encephalitis, immune response was demonstrated to MBP with early destruction of myelin in about 50 % [34]. Other viral infections associated with demyelination include JC-virus in progressive multifocal leucoencephalopathy, seen mainly in immunosuppression, human immunodeficiency virus [HIV] itself in subacute HIV encephalopathy, and the human T-cell lymphotropic virus type 1 [HTLV-1]-associated myelopathy affecting the spinal cord [32]. There are also several naturally occurring animal viruses that produce demyelinating CNS disease, some of which are used as animal models to study the biology of MS. These include Theiler's virus, neurotropic strain of mouse hepatitis virus, and Semliki Forest virus [32].

Exposure to microbes in early childhood has also been proposed to influence the development of autoimmune disorders such as MS, the hygiene hypothesis. This could explain the geographical differences in incidence of MS, greater in developed nations of the temperate zones with advanced hygienic communities compared to poorer countries in tropical and subtropical regions with substandard hygiene, but lower rates of MS. There are also marked differences in the incidence of MS in persons migrating from one country to another in which the rates are different. In Israel, MS is common in immigrants from Europe and rare among immigrants from Africa or Asia, whereas in native born Israelis of African or Asian descent have similar rates of MS as the European migrants [35]. These differences cannot be

explained by sunlight exposure or genetic factors. Similar trends have been reported in US migrant studies with a large number of MS cases [$>5,000$] [36]. It is postulated that multiple infectious exposures [even carriage of parasites in the gut] could reduce the risk of MS by modulating the immunity toward helper T cells [Th]2 and regulatory T cells, with attenuation of the proinflammatory Th-1 cellular immunity [37, 38]. Decreased antigenic stimulation from low childhood infections and reduced microbial exposure burden may result in decreased levels of regulatory cytokines, IL-10, and transforming growth factor B[TGF-B], which are produced by CD 25+ T cells and other regulatory T-cells, to downregulate both Th-1 and Th-2 mediated immune responses [39]. Intestinal helminth, which induces predominantly Th-2 response, has been reported to produce a beneficial effect in patients with MS [40, 41].

8.3.1 Specific Microbes

The role of microbes in autoimmune disorders is complex, some infections can trigger autoimmune responses and others may prevent these reactions [37]. This could be related to the microorganism itself, host genetic trait, age of onset, and epistatic effect with other environmental factors.

8.3.2 Epstein–Barr Virus

Epstein–Barr virus [EBV] is strongly implicated as playing an important role in the pathogenesis of MS. EBV is a human herpesvirus that infects B-cells in nearly 95 % of the population and persists latently in the memory B-cell pool for life. It was proposed more than 50 years ago that MS may be caused by infection that is harmless to the host and confers protective immunity when acquired in early childhood, but become pathogenic later in life [42]. This could apply to EBV infection and the pattern of infection between resource endowed and resource deprived countries of the world could explain the discrepancy in geographic distribution of MS. Almost all children are infected with EBV at an early age in developing countries of tropical and subtropical regions of the world, with a seropositivity of >90 % by 4 years of age, whereas in Europe and the United States only 30–40 % are infected at the same age [42]. The prevalence of age-related EBV infection is also increased with lower socioeconomic status and overcrowding. Infection in later years is more commonly manifested by symptomatic disease and pathology, such as clinical infectious mononucleosis, peak incidence 15–25 years of age in developed countries, whereas infection at a very young age is largely asymptomatic [43]. The epidemiology of MS in developed countries is strikingly similar to that of infectious mononucleosis with respect to age of onset [44]. MS risk in EBV-negative adults is extremely low but is increased in those with previous infectious mononucleosis. There is a 20-fold increased risk of MS in adolescents

and young adults with a history of infectious mononucleosis compared to those who are EBV negative, even for similar childhood hygienic environment [45]. In a meta-analysis of 13 case-control studies comparing MS patients and matched controls, 99.5 % of MS patients were EBV seropositive compared to 94 % of controls, but the risk of MS in EBV-seronegative subjects was extremely low, odds ratio 0.6, highly significant [46]. The mean interval between primary EBV infection and onset of MS [during the vulnerable age of 15–40 years] is estimated to be 5.6 years [47]. A clinical history of infectious mononucleosis increases the risk of MS more than twofold with a relative risk of 2.3 [48].

A few prospective studies have reported that elevated IgG antibodies to EBV nuclear antigen-1 [EBNA1] were found to increase the risk of MS [49–51]. In a study involving US military personnel before onset of MS, high serum titers to EBNA1 increased the risk 36-fold for developing the disease later [51]. Seroepidemiological studies of EBV in children with MS and matched controls have shown a similar pattern as in adults but somewhat less robust. EBV-seropositivity rate in MS children varies from 86 to 94 %, compared to 64–72 % in age-matched controls [52–54]. It can be argued that EBV infection is not essential for development of MS, as 14 % of children with this disorder were EBV seronegative [53]. However, in a recent meta-analysis of 22 adult and 3 pediatric studies on the risk of development of MS in EBV seronegative individuals, it was concluded that EBV appears to be present in almost 100 % of MS patients [55].

The increased synthesis of antibodies in the CSF as reflected by the corrected Antibody Index [56], also support a role of EBV in the pathogenesis of MS [57–59]. CSF oligoclonal IgG bands in MS patients predominantly consist of antibodies against EBV proteins, EBNA1 and BRRF2 [58]. However, increase in CSF antibodies to other viruses [measles and rubella] has also been reported [56, 57]. It is proposed that EBV infection of the CNS could stimulate dominant anti-EBV antibody response and promote synthesis of other viral antibodies by infecting B-cells [60].

A key issue in determining the pathogenic role of viruses in MS is the presence of the microorganism in the CNS and demyelinating plaques. Various studies have examined brain tissue samples for EBV by different methods, such as in situ hybridization, immunohistochemistry, and PCR, in subjects with MS with conflicting results. Serafini et al. [61] previously reported that B-cells and plasma cells of 21 of 22 MS brain sections had detectable EBV by in situ hybridization and immunohistochemical stains. However, Willis et al. [62] detected EBV in only 2 of 24 MS brains by real-time [RT] PCR and none by in situ hybridization nor immunohistochemistry. A subsequent focused workshop was held in Vienna in 2011 to review the data of EBV in MS brain [63]. Overall most studies using PCR for EBV detection from brain sections or CSF failed to detect the virus, except on rare occasions. The data on detection of EBV by in situ hybridization and immunohistochemistry were more mixed with varying results [63]. Hence, unequivocal proof that EBV exist in the brain lesions of MS patients, compared to EBV-related tumors, is still lacking.

The detection of EBV DNA in the blood of MS patients and matched controls by PCR methods also had been reported. In one study from Spain, EBV was detected in the blood of 70/75 [93.3 %] MS patients versus 123/186 [66.1 %] of controls,

$p < 0.001$ [64]. Moreover, dual infection with types 1 and 2 EBV was detected in 63 [90 %] of MS patients and only 37 [30 %] of controls, $p < 0.001$. In contrast, in a nested study from Australia with 215 MS cases and 216 controls, detectable EBV in blood was similar [55.8 and 50.5 %], and there was no difference between the two groups in EBV DNA load [65]. However, similar to other reports, past history of infectious mononucleosis, high anti-EBV titers, and HLA-DR B1 status had additive risk for MS.

8.3.3 Human Herpesvirus-6 in Multiple Sclerosis

Human herpesvirus-6 [HHV-6], which causes roseola in early childhood, infects >90 % of the population and remain latent probably in lymphocytes. The virus has been associated with meningoencephalitis in immunocompromised hosts on occasion [66] and also has been associated with MS. There is evidence that there is cross reactivity with MBP and HHV-6 in MS patients, which could activate autoimmune reactivity through molecular mimicry [67]. Phosphorylation of HHV-6 protein U24 may confound signaling and other pathways normally utilized by phosphorylated MBP that could precipitate the pathological process in MS [68]. Some studies have reported evidence of HHV-6 reactivation with MS activity but others have not. In one study measuring viral mRNA in peripheral blood mononuclear cells [PBMC] by RT-PCR, and plasma IgG and IgM antibodies, the prevalence of HHV-6 active infection was significantly higher in MS patients than other neurological diseases and in blood donors [69]. Moreover, there was correlation with reactivation of HHV-6 and with relapsing and progressive MS. In another study with 1 year of follow-up, serum samples were analyzed by quantitative PCR to assess HHV-6 prevalence and viral load. Among 63 patients with relapsing–remitting MS only 19.1 % of samples in relapse had active infection compared to 7.9 % of samples in remission [70]. HHV-6 DNA was found in 16 of 64 [25.4 %] MS patients at least once but in none of 63 healthy blood donors, $p = 0.04$ [71]. In another report from the same group of investigators, only 16 % of 105 MS patients had active HHV-6 infection versus 0 of 49 healthy controls, but the viral load was higher during attacks than during remission, $p = 0.04$ [71]. A subsequent study of 57 MS patients and 57 controls followed for a year also found that reactivation or new infection with HHV-6 variant A was related to relapse of symptoms, with a prevalence of 80.7 % in MS cases and 29.8 % in controls [72]. These investigators also assessed the effect of beta-interferon [IFN- β] treatment on HHV-6 viral load in MS subjects. Treatment with IFN- β was given to 105 patients and 84 were untreated; the viral load of HHV-6 was twice as high in untreated than treated cases in relapse [73]. IFN- β treatment reduced HHV-6 viral burden in patients in relapse but not in remission.

Other investigators have reported discrepant results of the presence of HHV-6 in MS. In the Finnish twin study of 17 MS twin pairs, serum and CSF were analyzed for HHV-6 DNA by PCR and for IgG and IgM antibodies [74]. The prevalence of antibodies was similar between twins with MS and healthy twin siblings, 88 and 86 %;

and there were no detectable antibodies in any CSF sample and no HHV-6 DNA was found in serum or CSF. Other negative studies for HHV-6 DNA in serum or CSF in MS patients have been reported by several groups [75–77].

Several studies have been performed to detect the presence of HHV-6 genome or antigen in brain lesions of MS patients with varying results. HHV-6 genome was reported in acute brain lesions of all five MS cases [78] and in 58 % of established plaques [79]. However, HHV-6 viral mRNA can be detected in both MS plaques and normal appearing white matter, although at higher levels than normal control samples [80]. In another report HHV-6 DNA was detected at similar rates, 41 and 44 %, and quantity in MS and control samples [81].

A previous systematic review of the association of HHV-6 and MS was reported in 2010. Overall, 25 of 61 [41 %] studies showed a significant positive correlation but only 15 of the studies were considered of high [A] quality [82]. Thus, the role of HHV-6 in the pathogenesis of MS remains unclear and correlation with disease activity is not robust.

8.3.4 Human Retrovirus in Multiple Sclerosis

Human endogenous retrovirus [HERV] genetic elements comprise about 1–8 % of the human genome and are believed to be remnants of ancestral infections of exogenous retroviruses during our evolution [83, 84]. HERVs are divided into specific families and may occur in up to 1,000 copies distributed throughout the human genome and inherited by a Mendelian pattern [85]. Although not replication competent, HERV genes may be intact and encode functional proteins [86]. HERV have been implicated in carcinogenesis and autoimmune diseases in both animals and humans [84]. In the late 1980s a novel retrovirus element was isolated from cells derived from CSF of a MS patient and was named MS-associated retrovirus [MSRV], and later was incorporated in the HERV-W family [87]. Although HERV elements are considered normal constituents of the human genome which are rarely expressed in cells, activation in cell culture to develop viral bodies [and possibly in humans] may be precipitated by environmental and endogenous stress. Activation could be an epiphenomenon after flares of inflammatory cytokines, but there is some evidence that specific HERVs may act as auto-, super-, or neoantigen that could enhance inflammation or induce autoimmune reactions [88]. In a humanized SCID mouse model, MSRV retroviral particles injection caused acute neuropathological changes with multifocal brain hemorrhages, mediated by the expression of inflammatory cytokines through T-cell stimulation [89].

Viral RNA from HERVs has been detected by reverse-transcriptase [RT]-PCR in blood and brain of MS patients, but not exclusively [88]. It has been postulated that herpesviruses may activate [transactivation] HERV-W particles and enhance immunopathological reactions in MS [90, 91]. In a recent in vitro study, EBV activated the potentially neuropathogenic HERV-W/MSRV/syncytin-1 in cells derived from blood and brain [92]. The authors proposed a model that include EBV as initial

trigger of future MS, and years later interaction of HERV-W/MSRV/syncytin-1 contributes to MS pathogenicity, paralleling the observed relationship of EBV infection in MS patients. Several studies have found elements of HERV-W family in blood, CSF, and brain lesions of MS subjects and significantly less in controls [93–95]. HERV particles in CSF of early MS cases, followed for 10 years, were associated with greater risk of disability and progression of disease in a small number of subjects [96]. Also B-cells and monocytes from patients with active MS exhibit increased surface expression and high antibody reactivities in sera to HERV epitopes, more than stable MS cases and controls [97].

However, other studies have failed to confirm the association of MS and HERV elements. In a study of 92 CSF samples, 48 from MS patients and 23 from other inflammatory neurological diseases, and 21 from patients with non-inflammatory CNS diseases no HERV sequences were found in any sample [98]. Analysis of humoral and cellular immune responses against MSRV/HERV-W antigens in 50 MS cases and 59 controls, in another study failed to detect any appreciable immune responses [99]. The majority of HERVs are present in 100 % of healthy humans and the paucity of functional genes argues against a causative role in disease. However, recently a new class of polymorphic HERVs has been described with widespread differences in geographic and racial distribution that could explain the geographic variation in MS distribution, if implicated in disease pathogenesis. A subtype, HERV-K113 is present in 0–28 % of humans and could be a disease causing HERV [100]. In a large family-based study, genomic DNA samples from 951 MS patients and 1,902 unaffected parents were tested for the presence of HERV-K113 allele by PCR [101]. HERV-K113 provirus was detected in only 70 of 951 [7.3 %] MS patients and 6.5 % in the parents, which did not support a role in MS. To cloud the issue further, another member of the HERV-K family has been reported to be associated with MS in a large study population. HERV-K18 is considered an EBV-associated superantigen and is a plausible candidate to influence the genetic susceptibility to MS. In a nested case–control study of 207 MS cases and 403 matched controls, with analysis replicated in 909 MS patients and 339 controls, risk of MS was threefold higher in individuals with HERV-K18 env alleles [102].

8.4 Conclusion

Microbes, especially viruses that remain latent in the host for life, are biological plausible triggers or key factors in the development and pathogenesis of MS. This maybe through molecular mimicry, alteration of the immune response to other antigens, and by genetic influence [through HERV elements], acting in a manner similar to gene variation such as with single nucleotide polymorphism. However, it is difficult to explain the discrepancies noted with different viruses in various studies and their findings on the relationship with MS. The most likely explanation is the difference in methods of detection used, which are not standardized or commercialized but locally developed without independent validation.

The present data indicate that the timing of primary EBV infection at a certain age or period in life [teenage to young adulthood] in those genetically susceptible plays an important role in the development of MS. The exact mechanism of this relationship remains elusive. The hypothesis that genetic influence of ancestral endogenous retrovirus interaction with latent EBV is an attractive paradigm but remain unproven.

8.5 Future Direction

Further studies on the role of microbes in MS need to concentrate on the most attractive theory with the best data available, in order to expend valuable resources on a large, prospective cohort with observation over several years. This would preferably be implemented by an international collection of interested investigators, with different interest and expertise, using standardized or validated methods of investigation, repeated at intervals over the years from blood, CSF, and brain samples where feasible. Such a study would be best performed in subjects with strong family history or evidence of genetic predisposition to MS.

References

1. Hauser SL, Goodin DS. Multiple sclerosis and demyelinating diseases. In: Longo DL, Fauci AS, Kasper DL, Jameson JL, Loscalzo J, editors. *Harrison's principles of internal medicine*. 18th ed. New York, NY: Mc Graw Hill Medical; 2011. p. 3395–409.
2. Compston A, Coles A. Multiple sclerosis. *Lancet*. 2008;372:502–17.
3. Weiner HL. Multiple sclerosis is an inflammatory T-cell mediated autoimmune disease. *Arch Neurol*. 2004;61:1613–5.
4. Zhang J, Markovic-Plese S, Lacet B, Raus J, Weiner HL, Hafler DA. Increased frequency of interleukin-responsive T cells, specific for myelin basic protein and proteolipid protein, in peripheral blood and cerebrospinal fluid of patients with multiple sclerosis. *J Exp Med*. 1994;17:9973–84.
5. Viglietta V, Baecher-Allen C, Weiner HL, Hafler DA. Loss of functional suppression by CD+CD25+ regulatory T-cells in patients with multiple sclerosis. *J Exp Med*. 2004;199: 9973–84.
6. Shi Y, Feng Y, Kang J, et al. Critical regulation of CD4+ T cell survival and autoimmunity by beta-arrestin 1. *Nat Immunol*. 2007;8:817–24.
7. Langrish CL, Chen Y, Blumenschein WM, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med*. 2005;201:233–40.
8. Mathey EK, Derfuss T, Storch MK, et al. Neurofascin as a novel target for autoantibody-mediated axonal injury. *J Exp Med*. 2007;204:2363–72.
9. Ousman SS, Tomooka BH, van Noort JM, et al. Protective and therapeutic role for alpha B-crystallin in autoimmune demyelination. *Nature*. 2007;448:474–9.
10. Lucchinetti CF, Popescu FG, Bunyun RF, et al. Inflammatory cortical demyelination in early multiple sclerosis. *N Engl J Med*. 2011;365:2188–97.
11. Hafler DA, Compston A, Sawcer S, et al. Risk alleles for multiple sclerosis identified by a genome-wide study. *N Engl J Med*. 2007;375:851–62.

12. Smolders J, Damorseux J, Menheere P, Hupperts R. Vitamin D is an immune modulator in multiple sclerosis, a review. *J Neuroimmunol.* 2008;194:7–17.
13. Kurtzle JF. Epidemiological evidence for multiple sclerosis as an infection. *Clin Microbiol Rev.* 1993;6:382–427.
14. Ebers GC, Bulman DE, Sadovnic AD, et al. A population-based study of multiple sclerosis in twins. *N Engl J Med.* 1986;315:1638–42.
15. Lincoln MR, Montpetit A, Cader MZ, et al. A prominent role for the HLA class II region in association of the MHC region with multiple sclerosis. *Nat Genet.* 2005;37:1108–12.
16. Oksenberg JR, Barcellos LF, Cree BA, et al. Mapping multiple sclerosis susceptibility to the HLA-DR locus in African Americans. *Am J Hum Genet.* 2004;74:160–7.
17. Yeo TW, De Jager PL, Gregory SG, et al. A second major histocompatibility complex susceptibility locus for multiple sclerosis. *Ann Neurol.* 2007;11:228–36.
18. Sawcer S, Hellenthal G, Primvien M, et al. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature.* 2011;476:214–9.
19. Culverhouse R, Suarez BK, Lin J, Reich T. A perspective on epistasis: limits of models displaying no main effect. *Am J Hum Genet.* 2002;70:461–71.
20. Malek TR. The biology of interleukin-2. *Annu Rev Immunol.* 2008;26:453–79.
21. Peschon JJ, Morrissey PJ, Grabstein KH, et al. Early lymphocyte expansion is severely impaired in interleukin 7 receptor-deficient mice. *J Exp Med.* 1994;180:1955–60.
22. Grigorian A, Mkhikian H, Li CF, Newton BL, Zhou RW, Demetriou M. Pathogenesis of multiple sclerosis via environmental and genetic dysregulation of N-glycosylation. *Semin Immunopathol.* 2012;34:415–24.
23. Demetriou M, Granovsky M, Quaggin S, Dennis JW. Negative regulation of T-cell activation and autoimmunity by Mgats N-glycosylation. *Nature.* 2001;409:733–9.
24. Lee SU, Grigorian A, Pawling J, et al. N-glycan processing deficiency promotes spontaneous inflammatory demyelination and neurodegeneration. *J Biol Chem.* 2007;262:33725–34.
25. Huynh JL, Casaccia P. Epigenetic mechanisms in multiple sclerosis: implications for pathogenesis and treatment. *Lancet Neurol.* 2013;12:195–206.
26. Goodin DS. The causal cascade to multiple sclerosis: a model for pathogenesis. *PLoS One.* 2009;4:e4565.
27. Pierrot-Deseilligny C, Sourberbielle JC. Is Hypovitaminosis D one of the environmental risk factors for multiple sclerosis? *Brain.* 2010;133:1869–88.
28. Correale J, Ysrraelit MC, Gaitan MI. Immunomodulatory aspects of vitamin D in multiple sclerosis. *Brain.* 2009;132:1146–60.
29. Royal 3rd W, Mia Y, Li H, Nauton K. Peripheral blood regulatory T cell measurements correlate with serum vitamin D levels in patients with multiple sclerosis. *J Neuroimmunol.* 2009; 213:135–41.
30. Smolders J, Thewissen M, Peclen E, et al. Vitamin D status is positively correlated with regulatory T cell function in patients with multiple sclerosis. *PLoS One.* 2009;4(8):e6635.
31. Kurtzke JF. Epidemiologic evidence for multiple sclerosis as an infection. *Clin Microbiol Rev.* 1993;6:382–427.
32. Fazakerleg JK, Walker R. Virus demyelination. *J Neurovirol.* 2003;9:148–64.
33. Johnson RT. Viral infections of the nervous system. 2nd ed. Philadelphia, PA: Lippincott-Raven; 1998.
34. Johnson RT, Griffin D, Hirsch R, et al. Measles encephalomyelitis: clinical and immunological studies. *N Engl J Med.* 1984;310:137–41.
35. Leibowitz U, Kahana E, Atter M. The changing frequency of multiple sclerosis in Israel. *Arch Neurol.* 1973;29:107–10.
36. Gale CR, Martyn CN. Migrant studies in multiple sclerosis. *Prog Neurobiol.* 1995;47:425–8.
37. Bach JF. The effect of infections on susceptibility to autoimmune and allergy diseases. *N Engl J Med.* 2002;347:911–20.
38. Sewell DL, Reinke EK, Hopgan LH, Sondor M, Fabry Z. Immunoregulation of CNS autoimmunity by helminth and mycobacterial infections. *Immunol Lett.* 2002;82:101–10.

39. Weiss ST. Eat dirt- the hygiene hypothesis and allergic diseases. *N Engl J Med.* 2002; 347:930–1.
40. Correale J, Farez M. Association between parasitic infection and immune responses in multiple sclerosis. *Ann Neurol.* 2007;61:97–108.
41. Flemming J, Fabry Z. The hygiene hypothesis and multiple sclerosis. *Ann Neurol.* 2007; 61:85–9.
42. Poskanzer DC, Schapira K, Miller H. Multiple sclerosis and poliomyelitis. *Lancet.* 1963; 2:917–21.
43. Evans AS. Epidemiology of Epstein-Barr virus infection and disease. In: Nahmias AJ, Dowelle WK, Schinazi RF, editors. *The human herpesviruses. An interdisciplinary perspective.* New York, NY: Elsevier North Holland Inc.; 1981. p. 172–83.
44. Warner HB, Carp RI. Multiple sclerosis and Epstein-Barr virus. *Lancet.* 1981;2:1290.
45. Ascherio A, Munger KL. 99th Dahleem Conference on infection, inflammation and chronic inflammatory disorders: Epstein-Barr virus and multiple sclerosis. *Clin Exp Immunol.* 2010; 160:120.
46. Ascherio A, Munger KL. Environmental risk factors for multiple sclerosis: Part 1. The role of infection. *Ann Neurol.* 2007;61:288–99.
47. Levin LI, Munger KL, O'Reilly EJ, Falk KJ, Ascherio A. Primary infection with the Epstein-Barr virus and risk of multiple sclerosis. *Ann Neurol.* 2010;67:824–30.
48. Thacker EL, Miraezi F, Ascherio A. Infectious mononucleosis and risk for multiple sclerosis: a meta-analysis. *Ann Neurol.* 2006;59:499–503.
49. Ascherio A, Munger KL, Lennette ET, Spiegelman D, Hernan MA, Olwk MJ, et al. Epstein-Barr virus antibodies and risk of multiple sclerosis: a prospective study. *JAMA.* 2001; 286:3083–8.
50. Sundstrom P, Juto P, Waddell G, Hallmans G, Suenningsson A, Nystrom L, et al. An altered immune response to Epstein-Barr virus in multiple sclerosis: a prospective study. *Neurology.* 2004;62:2277–82.
51. Levin LJ, Munger KL, Ruberstone MV, Peck CA, Lennette ET, Spiegelman D, et al. Temporal relationship between elevation of Epstein-Barr virus antibody titers and initial onset of neurological symptoms in multiple sclerosis. *JAMA.* 2005;293:2496–500.
52. Pohl D, Krone B, Rostasy K, Brunner E, Lehnert M, et al. High seroprevalence of Epstein-Barr virus in children with multiple sclerosis. *Neurology.* 2006;67:2063–5.
53. Banwell B, Krupp L, Kennedy J, Tellier R, Tenebaum S, Ness J, et al. Clinical features and viral serologies in children with multiple sclerosis: a multinational observational study. *Lancet Neurol.* 2007;6:773–81.
54. Lunemann JD, Huppke O, Roberts S, Bruck W, Gartner J, Mung C. Broadened and elevated humoral immune response to EBNA1 in pediatric multiple sclerosis. *Neurology.* 2008;71: 1033–5.
55. Pakpoor J, Disanto G, Gerber JG, Dobson R, Meier UC, Giovannoni G, Ramagopalan SV. The risk of developing multiple sclerosis in individuals seronegative for Epstein-Barr virus: a meta-analysis. *Mult Scler.* 2013;19:162–6.
56. Reiber H, Lange P. Quantification of virus-specific antibodies in cerebrospinal fluid and serum: sensitive and specific detection of antibody synthesis in brain. *Clin Chem.* 1991; 37:1153–60.
57. Rand KH, Houck H, Denslow ND, Heilman KM. Epstein-Barr virus nuclear antigen-1 [EBNA-1] associated oligoclonal bands in patients with multiple sclerosis. *J Neurol Sci.* 2000;173:32–9.
58. Cepok S, Zhou D, Srivastava R, Nessler S, Bussow K, et al. Identification of Epstein-Barr virus proteins as putative targets of the immune response in multiple sclerosis. *J Clin Invest.* 2005;115:1352.
59. Jaqueiry E, Jilek S, Schlupe M, et al. Intrathecal immune responses to EBV in early MS. *Eur J Immunol.* 2010;40:878–87.
60. Pender MP. Infection of autoreactive B-lymphocytes with EBV, causing chronic autoimmune diseases. *Trends Immunol.* 2003;24:584–8.
61. Serafini B, Rosicarelli B, Franciotta D, et al. Dysregulated Epstein Barr virus infection in the multiple sclerosis. *J Exp Med.* 2007;204:2899–912.

62. Willis SN, Stadelmann C, Rodig SJ, et al. Epstein-Barr virus infection is not a characteristic feature of multiple sclerosis brain. *Brain*. 2009;132:3318–28.
63. Lassmann H, Niedobite KG, Aloisi F, Middelborg JM, the Neuropro MiSe Working Group. Epstein-Barr virus in the multiple sclerosis brain: a controversial issue—report on a focused work-shop in the Centre for Brain Research of the Medical University of Vienna, Austria. *Brain*. 2011;134:2772–86.
64. Santon A, Cristobal E, Aparicio M, Royvela A, Villar CA, Alvarez-Cermeno JC. High frequency of co-infection by Epstein-Barr virus types 1 and 2 in patients with multiple sclerosis. *Mult Scler*. 2011;17:1295–300.
65. Lucas RM, Ponsonby AL, Dear K, et al. Current and past Epstein-Barr virus infection in risk of initial CNS demyelination. *Neurology*. 2011;77:371–9.
66. Krug LT, Teo CG, Tanaka-Taya K, Inoue N. Newly identified human herpesviruses: HHV-6, HHV-7. In: Fong IW, Alibek K, editors. *New and evolving Infections of the 21st Century*. New York, NY: Springer; 2007. p. 195–276.
67. Tejada-Simon MV, Zang YC, Hong J, Rivera VM, Zhang JZ. Cross-reactivity with myelin basic protein and human herpesvirus-6 in multiple sclerosis. *Ann Neurol*. 2003;53:189–97.
68. Tait AR, Straus SK. Phosphorylation of U2 from human herpes virus type 6 [HHV-6] and its potential role in mimicking myelin basic protein [MBP] in multiple sclerosis. *FEBS Lett*. 2008;582:2685–8.
69. Chapenko S, Millers A, Nora Z, Logina I, Kubaine R, Murovska M. Correlation between HHV-6 reactivation and multiple sclerosis disease activity. *J Med Virol*. 2003;69:111–7.
70. Alvarez-Lafuente R, Garcia-Montojo M, De las Heras V, Bartolome M, Arroyo R. Clinical parameters and HHV-6 active replication in relapsing-remitting multiple sclerosis patients. *J Clin Virol*. 2006;37 suppl 1:S24–6.
71. Alvarez-Lafuente R, De las Heras V, Bartolome M, Picazo JJ, Arroyo R. Relapsing remitting multiple sclerosis and human herpesvirus 6 active infection. *Arch Neurol*. 2004;61:1523–7.
72. Alvarez-Lafuente R, De las Heras V, Bartolome M, Garcia-Montojo M, Arroyo R. Human herpesvirus 6 and multiple sclerosis: a one year follow-up study. *Brain Pathol*. 2006;16:20–7.
73. Alvarez-Lafuente R, De las Heras V, Bartolome M, Picazo JJ, Arroyo R. Beta-interferon treatment reduces human herpesvirus 6 viral load in multiple sclerosis relapses but not in remission. *Eur J Neurol*. 2004;52:87–91.
74. Kuusisto H, Hyoty H, Kares S, Kinnunen E, Elovaara I. Human herpes virus 6 and multiple sclerosis: a Finnish twin study. *Mult Scler*. 2008;14:54–8.
75. Frnciotta D, Bestetti A, Sala S, et al. Broad screening for human herpesviridae DNA in multiple sclerosis cerebrospinal fluid and serum. *Acta Neurol Belg*. 2009;109:277–82.
76. Aheram M, El-Omar A, Baho Y, Lubad MA. Association between human herpesvirus 6 and occurrence of multiple sclerosis among Jordanian patients. *Acta Neurol Scand*. 2009;120:430–5.
77. Mancuso R, Hernis A, Cavarretta R, et al. Detection of viral DNA sequences in the cerebrospinal fluid of patients with multiple sclerosis. *J Med Virol*. 2010;82:1051–7.
78. Goodman AD, Mack DJ, Powers JM, Baker JV, Blumberg BM. Human herpesvirus 6 genome and antigen in acute multiple sclerosis lesions. *J Infect Dis*. 2003;187:1365–76.
79. Cermelli C, Berti R, Soldan SS, Mayne M. High frequency of human herpesvirus 6 DNA in multiple sclerosis plaques isolated by laser microdissection. *J Infect Dis*. 2003;187:1377–87.
80. Opsahl ML, Kennedy PG. Early and late HHV-6 gene transcripts in multiple sclerosis lesions and normal appearing white matter. *Brain*. 2005;128:516–27.
81. Tuke PW, Hawke S, Griffiths PD, Clark DA. Distribution and quantification of human herpesvirus 6 in multiple sclerosis and control brains. *Mult Scler*. 2004;10:355–9.
82. Voumvourakis KI, Kitsos DK, Tsiodras S, Petrikos G, Stamboulis E. Human herpesvirus 6 as a trigger of multiple sclerosis. *Mayo Clin Proc*. 2010;85:1023–30.
83. Willer A, Soussele S, Gimbel W, et al. Two groups of endogenous MMTV related retroviral env transcripts expressed in human tissues. *Virus Genes*. 1997;15:123–33.
84. Dolei A. Endogenous retroviruses and human disease. *Expet Rev Clin Immunol*. 2006;2:149–67.
85. Tristem M. Identification and characterization of novel endogenous retrovirus families by phylogenetic screening of the human genome mapping project database. *J Virol*. 2000;74:3715–30.

86. Harris JM, Haynes R, Mc Intosh EM. A consensus sequence for a functional human endogenous retrovirus K [HERV-K] dUTPase. *Biochem Cell Biol.* 1997;75:143–51.
87. Perron H, Geng C, Laurent A, Mouriquard C, Pellat J, Perret J, Seigneurin JM. Leptomeningeal cell line from multiple sclerosis with reverse transcriptase activity and viral particles. *Res Virol.* 1989;140:551–61.
88. Clausen J. Endogenous retroviruses and MS: using ERVs as disease marker. *Int MS J.* 2003;10:22–8.
89. Firouzi R, Rolland A, Michel M, et al. Multiple sclerosis-associated retrovirus particles cause T lymphocyte-dependent death with brain hemorrhage in humanized SCID mice model. *J Neurovirol.* 2003;9:79–93.
90. Brudek T, Christensen T, Hansen HJ, Bobecka J, Mollar-Larsen A. Simultaneous presence of endogenous retrovirus and herpes virus antigens has profound effect on cell-mediated immune responses: implication for multiple sclerosis. *AIDS Res Hum Retrovir.* 2004; 20: 415–23.
91. Christensen T. Association of human endogenous retroviruses with multiple sclerosis and possible interaction with herpes viruses. *Rev Med Virol.* 2005;15:179–211.
92. Mameli G, Poddighe L, Mei A, Sotgiu S, Sera C, Manetti R, Dolei A. Expression and activation by Epstein-Barr virus of human endogenous retroviruses W in blood cells and astrocytes: inference for multiple sclerosis. *PLoS One.* 2012;7:e44991.
93. Dolei A, Perron H. Multiple sclerosis-associated retrovirus and its HERV-W endogenous family: a biological interface between virology, genetics, and immunology in human physiology and disease. *J Neurovirol.* 2009;15:4–13.
94. Laska MJ, Brudek T, Nissen KK, Christensen T, Moller-Larsen A, Petersen T, Nexø BA. Expression of HERV-Fc1, a human endogenous retrovirus, is increased in patients with active multiple sclerosis. *J Virol.* 2012;86:3713–22.
95. Perron H, Germe R, Bernard C, et al. Human endogenous retrovirus type W envelope expression in blood and brain cells provide new insights into multiple sclerosis disease. *Mult Scler.* 2012;18:1721–36.
96. Sotgiu S, Mameli G, Serra C, Zarbo IR, Arru G, Dolei A. Multiple sclerosis-associated retrovirus and progressive disability of multiple sclerosis. *Mult Scler.* 2010;16:1248–51.
97. Brudek T, Christensen T, Aagaard L, Petersen T, Hansen MJ, Mollar-Larsen A. B cells and monocytes from patients with active multiple sclerosis exhibit increased surface expression of both HERV-H Env and HERV-W Env, accompanied by increased seroreactivity. *Retrovirology.* 2009;6:104.
98. Alvarez-Lafuente R, Garcio-Montojo M, De las Heras V, Dominguez-Mozo MI, Bartolme M, Benito-Martin MS, Arroyo R. Herpesviruses and human endogenous retroviral sequences in the cerebrospinal fluid of multiple sclerosis. *Mult Scler.* 2008;14:595–601.
99. Ruprech K, Groven F, Sauter M, Best B, Rieckmann P, Mueller-Lantzsch N. Lack of immune responses against multiple sclerosis-associated retrovirus/human endogenous retrovirus W in patients with multiple sclerosis. *J Neurol.* 2008;14:143–51.
100. Moyes DL, Martin A, Sawcer S, Temperton W, Worthington J, Griffiths DJ, Venables PJ. The distribution of the endogenous retrovirus HERV-K113 and HERV-K115 in health and disease. *Genomics.* 2005;86:337–41.
101. Moyes DL, Goris A, Ban M, Compston A, Griffiths DJ, Sawcer S, Venables PJ. HERV-K113 is not associated with multiple sclerosis in a large family-based study. *AIDS Res Hum Retrovir.* 2008;24:363–5.
102. Tai AK, O'Reilly EJ, Alroy KA, Simon KC, Munger KL, Huber BT. Human endogenous retrovirus-K18 Env as a risk factor in multiple sclerosis. *Mult Scler.* 2008;14:1175–80.