
Vascular Anomalies of the Neonatal Airway

David H. Darrow and Gresham T. Richter

Introduction

In 1982, Mulliken and Glowacki proposed a biological classification of vascular anomalies [1]. These authors described two major categories of vascular anomalies based on their clinical behavior, natural history, and cellular kinetics: *vascular tumors*, which are lesions characterized by endothelial proliferation, and *vascular malformations*, which result from errors in vascular morphogenesis while exhibiting normal endothelial turnover [1–3]. The modification of this classification accepted by the International Society for the Study of Vascular Anomalies (ISSVA) in 1996 [2] is shown in Table 1.

The most common vascular anomalies of the neonatal airway are infantile hemangiomas, lymphatic malformations, and venous malformations. Infantile hemangiomas, which are among the vascular tumors, have a predilection for the subglottic airway. In contrast, lymphatic and venous vascular malformations are typically pharyngeal and supraglottic and cause obstruction in higher regions of the airway [4].

Infantile Hemangioma of the Airway

Infantile hemangiomas (IHs) are benign tumors of the vascular endothelium. The most common tumors of infancy, IHs have the unique ability to involute after a period of rapid

proliferation. Most IHs involve the skin, while only a small percentage involve the mucosa or the viscera.

IHs of the neonatal airway are relatively uncommon. Also known as “subglottic” hemangiomas, these lesions have a tendency to affect the narrowest portion of the pediatric airway, and therefore present with symptoms of stridor and cough. Growth of the tumor may result in airway compromise and severe respiratory distress.

Epidemiology

The incidence of IH in general has been estimated at 4–5 % based on retrospective and cross-sectional data [5], although other studies suggest rates of 1–3 % among newborns [6, 7], and 2.6–9.9 % among older children. [8, 9]. IHs occur more commonly among female infants at rate of about 3:1 [9, 10], and most studies report a significantly higher incidence in Caucasian infants [9–11]. The incidence of IH is increased among preterm infants, affecting 22–30 % of babies weighing less than 1 kg [10, 12]. Low birth weight has been implicated as the causal reason, with a 25 % increase in risk of IH development for every 500 g reduction in birth weight [13].

In a 1967 series, IHs of the airway accounted for 1.5 % of congenital laryngeal anomalies [14]. However, the actual incidence of IHs has not been determined in any formal study. An analysis of the 37-hospital Pediatric Health Information System (PHIS) database over the 5-year period from 2001 to 2005 found that of 2,890 admissions for a primary or secondary diagnosis of IH, 337 (12 %) underwent an airway procedure during at least one admission [15]. Thus, on average, pediatric hospitals in this cohort likely treated fewer than three symptomatic airway IHs each year. As with IH in general, airway IH of the subglottis have been reported more frequently in females, but with a 2:1 female to male preponderance [16–20]. It is unknown if the other risk factors associated with IH in general apply equally to the subset of IHs of the airway.

D.H. Darrow, M.D., D.D.S. (✉)
Department of Otolaryngology, Children’s Hospital
of The King’s Daughters, Eastern Virginia Medical School,
600 Gresham Drive, Norfolk, VA 23507, USA
e-mail: David.Darrow@chkd.org

G.T. Richter, M.D.
Arkansas Children’s Hospital, University of Arkansas Medical
School, Little Rock, AR, USA

Table 1 Classification of cutaneous vascular anomalies (Modified from issva.org/classification, updated 2014)

<i>Vascular malformations</i>
Venous
Lymphatic
Capillary
Arterial and arteriovenous
Mixed (combined)
<i>Vascular neoplasms</i>
Infantile hemangioma (IH)
Congenital hemangioma (CH)
Rapidly involuting (RICH)
Non-involuting (NICH)
Others
Kaposiform hemangioendothelioma (KHE)
Tufted angioma (TA)
Lobulated capillary hemangiomas (LCH) (pyogenic granuloma) ^a

Notes: ^aconsidered cutaneous hyperplasia

Etiology and Pathogenesis

The etiology and pathogenesis of IH remain theories that are incompletely proven. The endothelial progenitor cell theory suggests that IHs develop from clonal expansion of circulating endothelial progenitor cells (EPCs), resulting in vasculogenesis, or the de novo formation of new blood vessels. [21, 22]. The proposed stimulus for division of EPCs is a somatic mutation or abnormal signals from local tissues. The theory of placental origin suggests that fetal progenitor cells arise from disruption of the placenta during gestation or birth. This concept derived from research demonstrating that molecular markers unique to placental tissue were also present in IHs [23–25]. Clinical evidence for this theory is suggested by studies demonstrating an increased incidence of IH in association with chorionic villus sampling, placenta previa, and preeclampsia.

A unifying theory suggests that circulating EPCs find their way to certain locations that provide conditions favorable for growth into placenta-like tissues. In such tissues as the skin and liver, progenitor cells may encounter cellular signals and local tissue factors that stimulate their development. Such factors may include angiogenic and vasculogenic factors within the IH [26–28]. It is also theorized that disturbances causing placental hypoxia trigger a vascular response that increases the likelihood of IH [29–31]. Such disturbances include maternal chorionic villus sampling (CVS) or amniocentesis [[10, 32, 33], and placental anomalies such as retroplacental hematoma, infarction, and dilated vascular communications [29]. In utero hypoxia is also the most common cause of low birth weight and may explain the association seen with premature delivery.

Clinical Presentation

Although considered congenital lesions, airway IHs are not generally symptomatic at birth. However, like cutaneous IH, they typically exhibit a characteristic life cycle consisting of proliferation and involution. Proliferation occurs during early infancy, while gradual spontaneous involution or regression starts by 1 year of age [34–41]. Symptoms usually manifest during the early proliferative phase; accordingly, some airway IHs will become symptomatic during the neonatal period. 80–90 % of affected babies will present within the first 6 months with a mean age of 3.6 months at diagnosis [16, 42]. Symptoms are often present for several weeks before a definitive diagnosis is made. An intermediate period between proliferation and involution during mid-to-late infancy, often referred to as the “plateau” phase, more likely represents a period of temporary balance between individual cells that are proliferating and those undergoing involution and apoptosis [34–41]. The process of involution takes several years and varies in duration.

Most *symptomatic* airway IHs involve the subglottic larynx and, as a result, authors have traditionally referred to these lesions as “subglottic hemangiomas.” In fact, many such IHs have been demonstrated to have transglottic or paratracheal extension [43]. Nevertheless, their associated symptom complex is consistent, including biphasic stridor and barking cough, usually in the absence of dysphonia. The symptoms are often mistaken for those of infectious or inflammatory croup, especially since the symptoms typically worsen in the presence of upper respiratory illness. Furthermore, IHs respond to many of the same treatments used for croup, including racemic epinephrine and nebulized and systemic steroids. As a result, recurrent croup-like symptoms and gradual worsening of stridor during early infancy are suggestive of airway IH and should be investigated further. Swallowing is usually normal; however feeding may be affected as the infant tries to coordinate sucking with breathing.

About one half of infants diagnosed with an airway IH also will have a cutaneous IH, although only 1–2 % of children with cutaneous IHs also have airway IHs [17, 44]. Even in asymptomatic children, the presence of cutaneous IH in the “beard distribution” (parotid area, lips, chin, neck) may be a predictor of IH in the airway (Fig. 1) [45]. Such cases likely represent “segmental” IHs [43, 46].

Once the proliferative phase is completed (9–12 months of age), respiratory symptoms tend to slowly resolve. Although the lesion itself may not involute completely or even significantly for many months, growth of the child over time allows the airway to better accommodate the IH, resulting in diminished stridor and cough.

Diagnosis

Diagnosis of airway IH begins with a high index of suspicion, since many congenital and inflammatory processes of the airway can mimic IH during the early months of infancy. The presence of cutaneous lesions and the persistence or

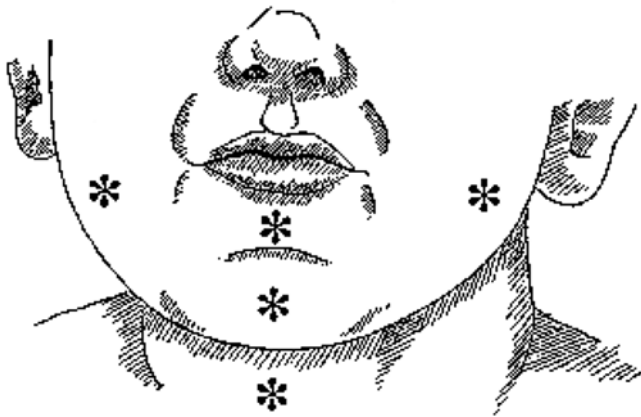


Fig. 1 Subsites of IHs in beard distribution, denoted by asterisks (Reprinted from [45], copyright © 1997, with permission from Elsevier, Ltd)

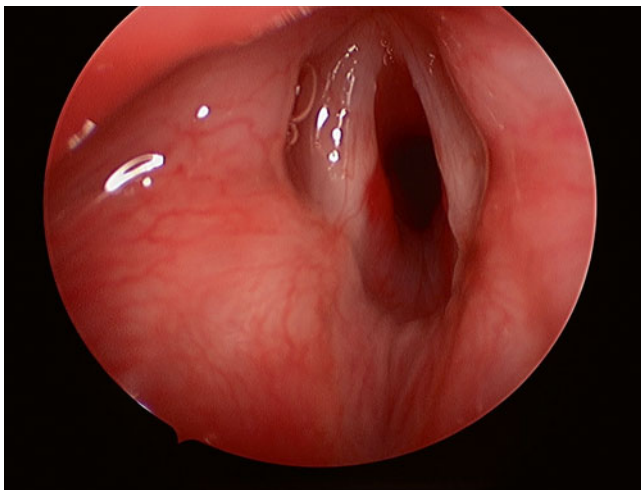


Fig. 2 Hemangioma on posterior wall of glottic and subglottis

recurrence of croup-like symptoms are important clues. When airway IH is suspected, the diagnosis may be confirmed by performing an imaging study or proceeding directly to laryngoscopy and bronchoscopy. In most cases, the uncertainty of the diagnosis or the severity of symptoms necessitates a trip to the operating room, where the pathology may be visualized directly and treated surgically if necessary. This approach avoids the risks associated with radiation exposure during infancy. Advocates of imaging, conversely, prefer to make a presumptive diagnosis based on flexible airway endoscopy, and confirm the diagnosis and assess the extent of the lesion by CT (computerized tomography) scan with angiography (“CT angio”) [47]. This approach avoids the need for general anesthesia, especially given the recent trend to treat such lesions initially with medical therapy rather than surgery.

When endoscopy is performed, symptomatic airway IHs are usually found to involve the subglottis, but bulk and blush may be present at adjacent and distant sites as well. The bulky lesion is smooth, submucosal, compressible, and pink or blue in color (Fig. 2). A left-sided predominance has been reported, but IHs may be bilateral, circumferential, or multiple. Biopsy is not usually necessary to establish the diagnosis; however when the diagnosis is in doubt, specimens from true IHs will stain positively for GLUT1 (glucose transporter protein isoform 1) [24, 48]. The most useful and widely used immunohistochemical marker for the diagnosis of IH, GLUT1 is strongly expressed by IH endothelial cells and not by other benign vascular anomalies, and has been validated for the identification of IHs of the airway [49].

On CT imaging, IHs will demonstrate an intensely staining, well-circumscribed mass with lobular architecture. MRI, although often diagnostic, is less advantageous since the duration of the study generally requires general anesthesia, often with endotracheal intubation. Fluoroscopy or anterior-posterior radiographs of the neck may demonstrate an asymmetric subglottic narrowing; however these studies do not definitively establish the diagnosis.

A staging system for airway IHs has been proposed by Perkins et al. [47]. IH stage is determined from CT angiography based on location, percentage of laryngeal airway obstruction, and estimated total volume of the IH (Table 2).

Table 2 Proposed airway IH staging system and treatment protocol

Stage ^a	Unilateral airway hemangioma	Circumferential or bilateral airway hemangioma	Percent laryngeal airway obstruction	CTA or MRI extralaryngeal hemangioma volume (mm ³)
1	Yes	No	≤50	<4,000
2	Yes	Yes	>50–90	4,001–10,000
3	No	Yes	>90	>10,000

CTA CT angiography, IH infantile hemangioma

^aStage based on lowest stage in a row with two or more positive findings

From Perkins et al. [47]

Management

Once the diagnosis of airway IH has been established, the need for and type of intervention is determined by several factors, including the degree of airway obstruction, the extent of extralaryngeal IH, the location of the patient at the time of diagnosis, the experience of the treating physician, and the preferences of the caretaker.

Since involution is the ultimate fate of nearly all infantile IHs, “watchful waiting” is reasonable in cases involving minimal symptomatology. For those patients with symptomatic IHs whose caretakers prefer no intervention for the lesion itself, tracheotomy with observation is highly successful [18]. This approach, however, requires a high level of maintenance to avoid tube occlusion, accidental decannulation, and exposure of the airway to water over a long period of time. As a result, in most cases, some sort of intervention for the IH itself is more desirable.

Pharmacotherapy

In a landmark 2008 publication by Léauté-Labrèze et al., the authors described their serendipitous observation that involution of IHs may be accelerated with the administration of propranolol [50]. At doses of 2–3 mg/kg/day used to treat cardiac complications of their IHs, two children experienced marked and rapid involution of their IHs. These results were replicated in a case series reported by the same authors [51] and in publications by a number of other investigators [52–62].

Several mechanisms have been proposed to explain the mechanism by which propranolol inhibits IH growth, including vasoconstriction due to decreased release of nitric oxide, blocking of proangiogenic signals (vascular endothelial growth factor, basic fibroblast growth factor, matrix metalloproteinases 2 and 9), and induction of apoptosis in proliferating endothelial cells [51, 62–64]. It has also been suggested that propranolol may prevent the differentiation of IH stem cells into endothelial cells or pericytes [65], or that it may hasten the differentiation of progenitor cells into adipocytes [51].

Several clinicians have reported success using propranolol in the management of airway IHs [66–76], but response is not universal [77]. The pretreatment assessment, optimal dose, and appropriate duration of therapy vary considerably in the literature. After considering contraindications such as cardiogenic shock, sinus bradycardia, hypotension, heart block greater than first-degree, heart failure, bronchial asthma, and known hypersensitivity to the drug, most clinicians will perform a cardiac evaluation, including at least an examination and a pretreatment electrocardiogram. In most series, the drug is started at a dose of 1 mg/kg/day divided two to three times a day, and then increased over several days to a week to 2–3 mg/kg/day. Heart rate and blood pressure are checked each hour for the first 2–3 h after the initial dose and with each dosage increase. The drug is administered

throughout most of the first year of life. Although protocols for propranolol initiation do appear in the otolaryngology literature [78, 79], a consensus multidisciplinary protocol has also been published [62].

Propranolol has a well-established safety profile based on years of use at higher doses for control of high blood pressure and cardiac pathology. Potential side effects and complications include sinus bradycardia, hypotension, reactive airways, hypoglycemia with secondary seizures, diarrhea, and cool extremities. Patients should be monitored for evidence of bradycardia, hypotension and/or hypoglycemia [62, 65, 80]. However, such complications are reported rarely, and it has been suggested that propranolol should become the standard for initial management of all airway IHs [81].

Although propranolol has largely supplanted systemic corticosteroids as first line pharmacotherapy, the latter are occasionally useful in refractory cases. Steroid medications inhibit growth of the lesion during the proliferative phase, losing their effectiveness once involution begins [82]. Doses of prednisolone at 2–3 mg/kg/day are generally necessary to control growth of the mass and should be maintained for 1–2 weeks before starting a 2–4 week taper. Response rates reported in the literature vary between 30 and 93 %, although there is little consistency among dosing regimens [82, 83]. Long-term management on steroids carries a significant risk of complications, including gastroesophageal reflux, gastritis, immune suppression, cushingoid changes, hyperglycemia and glycosuria, hypertension, fluid and electrolyte disturbances, and growth retardation [82]. IH patients on maintenance steroids should concomitantly receive courses of H₂ receptor blockers and trimethoprim-sulfamethoxazole as prophylaxis against gastritis and pneumocystis carinii infection, respectively. Live vaccinations should also be avoided while a child is taking high-dose steroids.

Interferon α -2A has been used with success in treating IHs, but should only be considered when all other traditional modalities fail [84–87]. Potential side effects are associated with this therapy, including fever, myalgia, transient elevation of hepatic transaminase levels, transient neutropenia, anemia, and spastic diplegia [88, 89].

Surgical Intervention

Indications for surgical management of airway IH have become few since the efficacy of propranolol was established. Surgery is a reasonable consideration when (1) the obstruction found at operative endoscopy is severe and will likely require a lengthy intubation while medical therapy is initiated, and (2) the patient remains severely symptomatic despite an adequate trial of medical therapy.

Operative intervention may include intralesional injection of corticosteroids and/or partial ablation of the IH, or complete surgical excision the portion of the lesion within the airway. In most cases, the patients will remain on

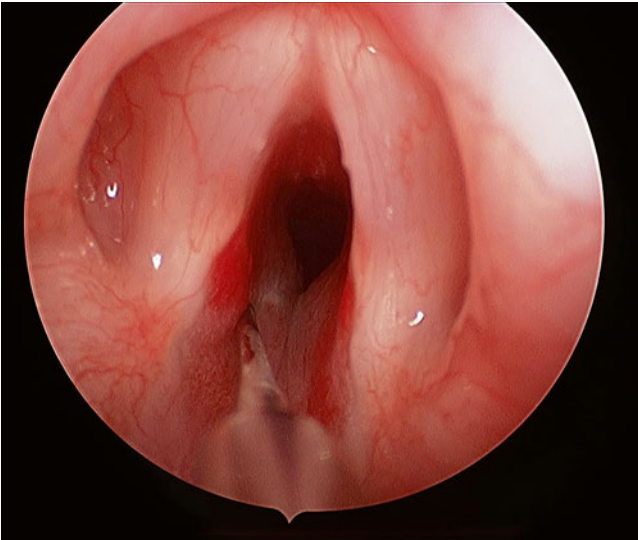


Fig. 3 Triamcinolone/betametasone steroid injection with needle in left side of subglottic hemangioma

medical therapy postoperatively to reduce the likelihood of recurrence. This is of particular importance in “segmental” airway lesions with known extension outside of the airway.

Intralesional steroids should be considered for patients whose IHs have necessitated a trip to the operating room for endoscopy or endoscopic resection. While repeated injections are usually necessary as single modality therapy [90], these medications may be effective adjuvant therapy for patients whose lesions are being observed, treated pharmacologically, or partially resected. In most cases, triamcinolone 40 mg/cm³ is administered at a dose of 3–5 mg/kg either alone or supplemented by betamethasone 6 mg/cm³ dosed at 0.5–1.0 mg/kg (Fig. 3) [82]. Total volume delivered may be limited by the size of the lesion, and care must be taken to avoid depositing the steroid medication deep enough to affect the underlying cartilage. Patients will usually require at least overnight intubation due to the increased volume of the lesion after injection. Cure rates of 77–87 % using intralesional steroids have been reported [42, 90].

Airway IHs causing focal obstruction may be addressed surgically by a subtotal endoscopic approach using a microscope or telescope, or by total excision through an open approach, dividing the thyroid and cricoid cartilages in the midline. Subtotal resection, more often than total excision, carries the risk of growth of the residual lesion during the proliferative phase, potentially resulting in additional surgical procedures unless combined with pharmacologic therapy. Endoscopic excision is usually performed using an apneic anesthesia technique, intermittently interrupting the surgery for reinsertion of the tube and ventilation of the patient. Alternatively, the procedure may be performed under spontaneous ventilation with anesthetic insufflation or Venturi jet ventilation.

The laser has been the most popular endoscopic surgical modality [42], with the carbon dioxide (CO₂) [17, 19, 44, 91, 92] potassium titanyl phosphate (KTP) [93, 94], and neodymium:yttrium-aluminum-garnet (Nd:YAG) [95, 96] lasers all demonstrating some effectiveness. All of these lasers are currently available for airway use through fiber delivery systems, however only CO₂ is used by direct beam. CO₂ lasers are preferentially absorbed by water, while KTP and Nd:YAG lasers take advantage of absorption peaks that approximate those of hemoglobin and are thought to penetrate more deeply. However, all of these lasers cause destruction by ablation rather than selective photothermolysis. As a result, in addition to the risk of recurrence, laser treatment carries a risk of subglottic stenosis of 5–25 % that is likely greatest with deeper resections and in cases of bilateral or circumferential disease [19, 44, 89, 97]. Debulking of the lesion using rotary powered instrumentation (microdebrider or “shaver”) has also been reported [98, 99]. Postoperatively, patients are observed in an intensive care setting. Some clinicians recommend face tent humidification to prevent airway obstruction due to eschar formation.

Although the first open surgical excision of a focally obstructing airway IH was reported in 1949, the procedure did not gain popularity until the 1990s, after complications of laser therapy became increasingly apparent [100–105]. Over the 15 years prior to the discovery of effects of propranolol, open resection appeared to be emerging as the intervention of choice for airway IHs. The procedure is of greatest advantage in patients with bilateral or circumferential lesions that may otherwise been at risk for postoperative stenosis, recurrence, or tracheotomy. However, open surgical excision may be more difficult in cases involving significant extra-laryngeal extension, and the procedure may potentially result in some degree of dysphonia.

After initial intubation through the obstructed portion of the airway, the lesion is approached through the anterior neck via laryngofissure. After the tube has been relocated to the inferior aspect of the incision, the IH is removed submucosally under the operating microscope. At the conclusion of the dissection, the patient is intubated; in some cases a thyroid cartilage graft may be placed to enlarge the subglottic laryngeal framework. After the neck is closed, the patient is transported to the intensive care unit where intubation is maintained for 3–7 days.

Multidisciplinary Considerations

Management of airway IH requires good communication among pediatric specialists, both in terms of diagnosis of the lesion as well as management after the diagnosis has been made. It is estimated that some 50 % of patients with airway

IHs have cutaneous lesions as well, many with problematic segmental involvement in the “beard” distribution. In addition, there is an important association of airway IH with PHACE(S) syndrome, including *Posterior fossa defects*, *Hemangiomas*, *cerebrovascular Arterial anomalies*, *Cardiovascular anomalies* including *Coarctation of the aorta*, *Eye anomalies*, and *Sternal cleft and/or Supraumbilical raphe* (OMIM 606519) [106]. In one study of 23 PHACE patients, 52 % were found to have airway IH, many of whom required treatment for stridor [107]. Another study found eight patients (42 %) meeting the criteria for PHACE among 17 with large “beard” distribution IHs and airway IHs [108]. In such cases, early consultations with pediatric dermatology and genetics are advisable.

During diagnostic endoscopy, as with all lesions of the airway, moment-to-moment communication between the otolaryngologist and the anesthesiologist is critical to successful management. Since in most cases a subglottic lesion is predictable based on symptoms, the two clinicians should have a game plan in place for intubation, management of the airway during any intervention, and emergency contingencies, and the two should agree on a plan for airway management in the immediate postoperative period.

Postoperatively, the otolaryngologist must continue to coordinate the airway management plan with the pediatric intensivists and hospitalists that will administer care to patients with airway IH while they recover from endoscopy and start on medical therapy. Such care includes the duration of intubation (if any), management of medical therapy for the IH and for reflux if indicated, and use of adjunctive racemic epinephrine and/or steroids to control exacerbations of stridor. In most cases, consultation with a pediatric cardiologist is advisable prior to starting propranolol therapy while the child is still hospitalized.

Future Considerations

There are still several important unanswered questions in the diagnosis and management of IH of the airway. One key issue in pathogenesis is the predilection of IH to involve the subglottis as opposed to other areas of the airway. As the etiology of IH in general becomes clearer, it may be possible to theorize plausible reasons for this clinical observation. It is also not yet clear if there is some subset of patients with a presentation of cutaneous IH that should undergo endoscopy prior to the development of symptomatic airway obstruction. Finally, in terms of therapeutic intervention, propranolol therapy is a relatively new tool in the management of airway IH, and additional research will be required to determine optimal administration, including dosage, frequency, and duration of therapy.

Vascular Malformations of the Neonatal Airway

Vascular malformations are congenital disorders of vasculogenesis with ectatic vascular networks and the propensity for relentless expansion and recruitment of new vessels. Unlike hemangiomas, vascular malformations do not experience a cycle of growth and involution. They are present at birth and grow commensurate with the child until periods of rapid expansion lead to functional and aesthetic compromise. Trauma, infection, and hormonal fluctuation have been associated with acute growth of these vascular anomalies [109, 110].

Vascular malformations may be composed of any blood vessel type and they are named accordingly, including capillary malformations (port wine stains), venous malformations, lymphatic malformations, and arteriovenous malformations. The most common vascular malformations affecting the neonatal airway are lymphatic and venous malformations [111–113]. Relative to subglottic hemangiomas and other disorders of the newborn airway, these malformations are rare. Although they may occur in isolation, neonatal vascular malformations of the airway are more commonly associated with diffuse cervicofacial lesions [114, 115]. Diffuse involvement of the head and neck will commonly affect the aerodigestive tract mucosa to include the oral cavity, tongue, hypopharynx and larynx (Fig. 4) [113]. Vascular malformations typically experience slow growth but airway involvement may necessitate urgent intervention, especially in the setting of acute or chronic obstruction.

Lymphatic Malformations

Epidemiology

Lymphatic malformations are believed to occur in 1 in 500 live births and comprise about 12 % of vascular cases referred to a vascular center [116]. Within the head and neck, lymphatic malformations occur at a rate of between 1 in 2,000 and 1 in 4,000 [117]. Fifty percent of cases present after birth and 80–90 % by 2 years of age. There does not appear to be a gender or race predilection for LMs.

Etiology and Pathogenesis

Lymphatic malformations (LMs) are slow flow vascular malformations composed of dilated ectatic lymphatic channels. They are thought to arise from disruptions in embryonic lymphangiogenesis but their etiology and pathogenesis remain unclear. Lymphatic malformations are present at

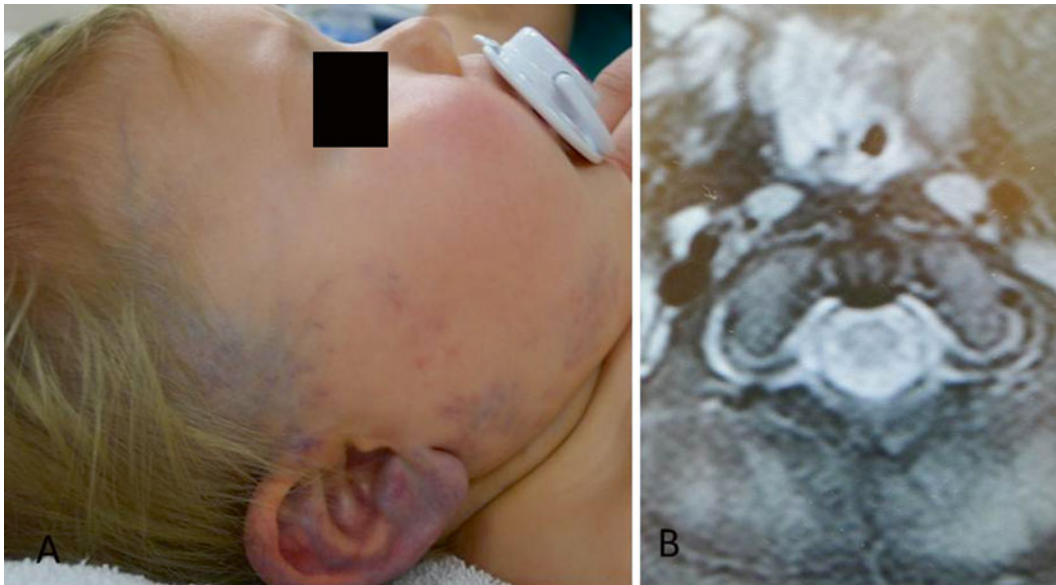


Fig. 4 Infant with venous malformation of facial soft tissue and larynx. The malformation is characterized by blue cutaneous staining (left) and enhancing lesion of supraglottis on MRI (right)

birth but may not be detectable until later in life. The majority of LMs occur in the head and neck, making them the second most common congenital mass found at these sites [118, 119]. Exacerbated by infection, cervicofacial LMs may expand in infancy or early childhood when the rate of ear and upper respiratory infections are at their peak. Diffuse lesions are more likely to progress than those that are localized. If left untreated, the likelihood of progression is significantly higher in adolescence than childhood [120]. The earlier intervention may thus be warranted in many cases.

Clinical Presentation

Based upon their response to treatment, LMs have been categorized by the size of the fluid filled lymphatic cysts present. In particular, cysts greater than 1–2 cm in diameter are considered macrocysts, while microcysts are those 1–2 cm in diameter or less [121]. While LMs may be predominantly macrocystic or microcystic, most LMs of the head and neck region are mixed lesions. Venous malformations may also be present within extensive cervicofacial LMs.

Macrocystic and microcystic LMs have different clinical courses. Macrocystic LMs grow by accumulated lymphatic fluid and expansion of the cysts into areas of least resistance. They have a tendency to encompass and compress surrounding architecture. In the neonate, they can affect the larynx and trachea by direct or indirect compression of these structures. Stridor, grunting, or symptoms consistent with obstructive sleep apnea may be present when the larynx, trachea, or pharynx are involved, respectively. With rapid expansion or

internal bleeding, macrocystic LMs may cause acute compromise of vital structures. Chronic expansion in the head and neck frequently leads to upper aerodigestive symptoms such as dysphagia and gradual airway obstruction, especially during recumbence.

Unlike macrocystic LMs, microcystic LMs are infiltrative in nature. In this condition, small microcystic vesicles penetrate soft tissue including skin, muscle, and mucosa [122, 123]. Non-descript soft tissue swelling and the emergence of weeping vesicles in the skin or mucosa often note its presence. Isolated disease commonly occurs in the tongue, upper lip, and larynx (Fig. 5). When associated with mixed cervicofacial lesions, microcysts in the tongue and larynx can cause acute life-threatening airway obstruction upon delivery, shortly after birth, or during periods of acute infection and swelling (Fig. 6) [124].

Isolated microcystic LMs rarely cause respiratory obstruction in the newborn period. However, in the presence of massive cervicofacial disease of mixed or microcystic LM, rapid airway compromise may occur, especially during periods of acute edema [124]. Upper airway obstruction is exacerbated by infiltrating disease of the tongue and floor of mouth (Fig. 6) [123, 125–127]. Laryngeal involvement will exacerbate symptoms.

Diagnosis

Airway endoscopy with office-based flexible laryngoscopy, followed by rigid microlaryngoscopy and bronchoscopy, will provide best evidence of the extent of obstruction and its

effect on the airway. Distinct margins of macrocystic LMs may also be detected on physical exam and on magnetic resonance imaging (MRI), the diagnostic study of choice for LMs. An MRI of the head and neck will demonstrate the depth of disease and differentiate macrocystic LM from more common conditions of the pediatric head and neck. Hyperintense lesion on T2 weighted images and air-fluid levels within the lesion are hallmark MRI findings for macrocystic LM [128]. Frequently mixed lesions are discovered. Microcystic and macrocystic lymphatic malformations may coexist with deep neck space venous malformations which will also be evident on MRI.



Fig. 5 Isolated lymphatic malformation of the supraglottic larynx. Lymphatic vesicles are seen on the left pharyngeal wall and vallecula

Management and Multidisciplinary Considerations

Except in rare cases, LMs ultimately need intervention to control symptoms associated with their accumulated lifetime growth. Timing of intervention for LMs in infants is dependent upon the acuity of symptoms and dictated by the presence, or risk of, acute and chronic upper aerodigestive dysfunction or pain due to inflammation. Acute airway obstruction, sleep apnea, and dysphagia are the most common reasons for intervention in these patients [113, 118, 126]. Sclerotherapy and surgery constitute the therapeutic approach to lymphatic malformations and is typically conducted in a multidisciplinary fashion. However, symptom control may be best achieved acutely with oral steroids and antibiotics that reduce the inflammation, lymph accumulation, and expansion of LMs.

Macrocystic Disease

A small percentage of isolated macrocystic LMs have been reported to spontaneously resolve [129, 130]. Because these isolated lesions usually occur in the neck, they rarely cause significant dysfunction in the newborn and can be observed into early childhood before treatment is considered.

Most macrocystic LMs compressing the neonatal airway require treatment. In acute situations, needle aspiration of the responsible cyst or cysts may alleviate the airway compromise until more definitive management with either surgical excision or sclerotherapy is possible. Sclerotherapy is the direct injection of agents that cause epithelial damage and



Fig. 6 Cervicofacial microcystic lymphatic malformation with multiple levels of airway obstruction leading to early tracheostomy

subsequent disruption of the LM channels. Various materials have been employed with equal success including doxycycline, ethanol, sodium tetradodecyl, and OK-432, an immunostimulant derived from Group A streptococci [131–135]. With these therapies, dissolution occurs as the result of intravascular injury and luminal absorption by the body. Multiple treatment sessions, under radiographic fluoroscopy, may be required for significant reduction in volume using this technique. Postoperative edema with potentially worsening compression may occur following sclerotherapy. Expectant airway management with intubation or intensive care observation is necessary when treating compressive lesions of the neonatal airway. Risk to adjacent nerves ranges from 5 to 10 % with temporary dysfunction being more common than complete injury [134].

Surgical excision can provide immediate relief of airway obstruction caused by macrocystic cervical LMs. The risk of injury to important neurovascular structures in the neck is real but fortunately rare in experienced hands. Coexistent venous malformations may be found during surgical excision of deep neck LMs. Recurrence, persistent lymphatic drainage, and postoperative scarring often occur following excision [136]. These risks are minimized by removing as much disease as possible, placing perioperative drains, and hiding incisions in natural creases of the neck.

Outcomes of surgical excision are not affected by age with successful management of head and neck macrocystic LMs in the newborn. Small recurrent LMs or seromas that occur following excision are best managed with symptom-directed sclerotherapy. Routine use of intraoperative intravenous dexamethasone and a short course of postoperative systemic steroids are recommended to prevent acute and dramatic lymphatic drainage and soft tissue edema that occurs following any intervention for LMs.

When venous malformations are found in mixed LMs, patients need to be screened for local intravascular coagulation. Elevated D-dimers and low fibrinogen levels indicate a coagulation disturbance and may be found in these patients prior to intervention. Perioperative management with low-molecular weight heparin can reduce the risk of intraoperative disseminated intravascular coagulation and perioperative thrombi in patients when these coagulation factors are abnormal.

Congenital high airway resistance syndrome (CHAOS) may occur in the presence of complex cervicofacial LMs at birth. If detected during ultrasonography and confirmed by fetal MRI, massive LMs that are expected to obstruct the newborn airway at delivery can be managed with ex utero intrapartum treatment, also known as the EXIT procedure [124]. This is a specialized surgical delivery of the newborn by a multidisciplinary team to manage potentially immediate life-threatening conditions at birth [137]. The newborn is delivered by cesarean section with the umbilical cord and



Fig. 7 Macrocystic lymphatic malformation of neck and chest in an infant. The malformation extends externally, but the airway is not compromised

placenta attached with the goal to deliver oxygen as urgent interventions are performed. The team has 1–1.5 h to stabilize the airway and establish adequate respiration to the baby. This may require intubation or tracheostomy. Fortunately, most infants with macrocystic LMs do not require an EXIT procedure as expansion of the lesion is primarily external and not towards the airway (Fig. 7). If necessary, intubation can be performed by an airway specialist. The larynges of such patients are typically easy to access. Bilateral mixed or microcystic LMs may be more difficult when other compressive components are present.

Microcystic Disease

The infiltrative nature and protracted course of microcystic LM make them particularly difficult to manage, especially when the upper aerodigestive tract is involved. In these patients urgent tracheostomy may be required as an infant that is thereby followed by periodic and staged treatments. Management of the neck, tongue, pharynx, and larynx is often necessary to ultimately relieve chronic obstruction. Unfortunately, microcystic disease is poorly responsive to sclerotherapy, due to the limited retention of sclerosing agents in the small cysts. Excision requires desiccation of normal tissue since this condition integrates into surrounding structures. Unfortunately, high rates of recurrence (90–100 %) are also experienced following excision of microcystic LM [125, 138].

Nonetheless, the tongue may be addressed with local ablative therapy. Infiltrative disease of the deep musculature of the tongue is best controlled by excision or coblation, which can reduce significant bulk and relieve obstruction.

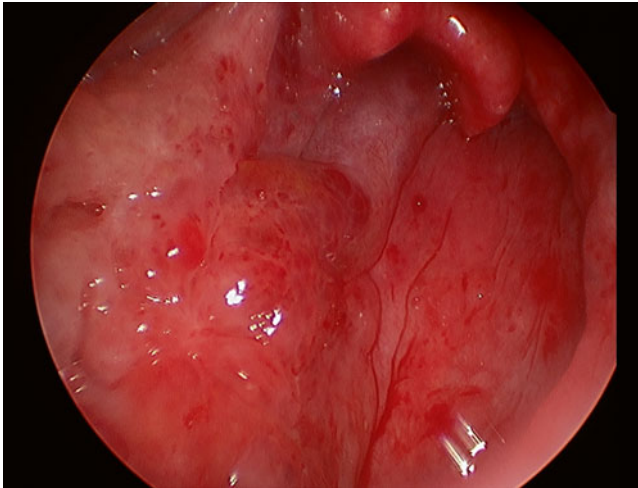


Fig. 8 Posterior and lateral pharyngeal involvement of microcystic lymphatic malformation

Early studies using intralesional bleomycin and doxycycline therapy in the management microcystic lymphatic disease have demonstrated promising results [138, 139]. Direct injections of bleomycin within the involved tissue have led to disease control and symptom improvement in some studies [139]. The risks of these agents are equivalent to those with other sclerotherapy tools, including nerve injury, pain, incomplete response, and the need for numerous treatments. There is theoretically an added risk of pulmonary fibrosis when using bleomycin, but this does not likely occur at the low doses typically used. The impact of these agents on laryngeal disease has not been closely examined.

Microcystic LM of the neonatal trachea is unlikely. However, early disease involving the larynx may be found [140–142]. This is often associated with adjacent pharyngeal or hypopharyngeal involvement (Fig. 8). Stabilization of life-threatening airway symptoms in this setting may be necessary as primary treatment with a tracheotomy [140]. A staging system for laryngeal disease demonstrates the frequent need of tracheotomy in these patients [143].

Surgical management of microcystic laryngeal disease must be performed in a staged fashion. The carbon dioxide (CO₂) laser may be used to ablate weeping or bleeding mucosal vesicles of the oral cavity, tongue, or pharynx. However, precise excision of involved laryngeal tissue is performed using the CO₂ laser in cutting mode [143, 144]. When employed, the laser is directed to the larynx to remove unwanted and obstructive microcystic disease. Unilateral treatment with management of the contralateral side at a later date will prevent unwanted scarring and worsening obstruction. This mucosa-sparing approach with partial excision of involved supraglottic disease is a complicated procedure and should be performed judiciously by experienced

otolaryngologists. Temporary intubation may be required postoperatively. Again perioperative steroids will reduce significant edema associated with interventions directed at LMs.

Future Considerations

The management of microcystic lymphatic malformations is particularly difficult. The deep tissue penetration, small vesicular cysts, and high recurrence rates of microcystic LMs limit the therapeutic benefit of surgery and sclerotherapy. Pharmacologic interventions have recently been explored for these complicated lesions with limited benefit. Rapamycin, an mTOR inhibitor, and sildenafil, a phosphodiesterase inhibitor, in particular have been tried with some early reported success. Prospective studies are necessary to determine if these tools can be truly useful in the search for a cure of microcystic disease. Nonetheless, genetic and molecular profiling may help identify new pharmacologic targets in the future.

Venous Malformations

Epidemiology

Due to their rarity, the actual incidence of venous malformations is unclear. However, based upon the referral pattern to a prominent vascular anomalies center, slow-flow vascular malformations have an incidence of about 1 in 10,000 people (0.01 % of population) [144]. There exists no race predilection for venous malformation but recent publications suggest a female predominance over males of 1.5:1 [145]. They are sporadic in 93 % of affected patients and usually unifocal. Approximately 7 % of cases of venous malformations are multifocal and associated with inherited forms comprised of cutaneomucosal venous malformations (VMCM).

Etiology and Pathogenesis

Venous malformations (VMs) are rare, low flow, slowly progressive vascular malformations. They are primarily composed of small to medium sized ectatic venous channels that coalesce to form a vascular mass of poorly draining veins. With time, gradual accumulation of blood, loss of vascular elasticity, and vascular expansion lead to growth of involved soft tissue. The etiology of VMs remains unclear although familial patterns and genetic links have been reported in patients with multifocal disease. TIE 2 mutations causing autophosphorylation of an endothelial cell specific tyrosine kinase are believed to lead to cell proliferation and cause VM to intermittently develop throughout life [144]. A second type of inherited multifocal

form is the glomovenous malformation (GVM) secondary to a glomulin gene mutation and resulting in abnormal differentiation of smooth muscle cells [144]. In familial cases of venous malformations, autosomal inheritance pattern has been observed and linked to the 9p locus [146].

Venous malformations frequently affect the head and neck. However, extremity involvement is most common (60.9 %) followed by head and neck (32.7 %) and trunk (9.8 %). The majority of venous malformations involve the integument (skin and subcutaneous tissue) but 50 % of lesions invade deeper structures including fat, muscle, bone and viscera. In the deep venous malformations, phlebectasia is the most common deep venous anomaly and is seen in about 40–50 % of patients [147, 148].

Sites commonly involved in the head and neck include the parotid gland, submandibular space, buccal space, and masseter muscle [114, 149]. Mucosal disease is the norm rather than the exception. Venous malformations of the tongue, lips, gingiva, cheek, palate, retromolar trigone, pharynx, hypopharynx, and larynx have been described and are frequently identified in early childhood [150–152]. Because of the slow expansion of VMs, they are rarely found in the newborn. When present in the airway, they may be asymptomatic in the neonatal period. Discovery during surveillance of disease elsewhere in the head and neck is common (Fig. 4).

Diagnosis and Clinical Presentation

Laryngeal involvement of VMs can lead to chronic upper airway resistance, especially during recumbence as the venous channels fill with dependency. Inspiratory stridor during eating, crying, and agitation may occur. Symptoms consistent with sleep apnea such as loud snoring, stertor, neck hyperextension, and restlessness are also common when supraglottic disease is present [149]. Acute airway obstruction is rare. Dysphagia may be present in infants with post-cricoid disease. Head and neck masses that worsen with Valsalva and dependency are consistent with deep cervical VMs. Skin involvement will appear as blue telangiectatic vessels (Fig. 4). The presence of extensive superficial or deep neck disease should prompt evaluation of the upper aerodigestive tract.

Endoscopic exam of the entire airway and MRI of the neck will provide the best evidence of mucosal involved VMs and their associated deep components. Pupuric to blue discoloration of the mucosa that expands with Valsalva or recumbence is consistent with venous malformations of the airway. Gadolinium enhancement on T1 weighted images is a common indicator of VMs on MRI. Focal hypolucencies on MRI suggest phleboliths, which are a hallmark sign of VMs compared to other vascular anomalies [128].

Management and Multidisciplinary Considerations

Early intervention for venous malformation of the airway includes raising the head of the bed to prevent progressive expansion of the venous channels during sleep. Like lymphatic malformations, both sclerotherapy and surgical excision can be employed for deep and superficial cervicofacial venous malformations with a high degree of success. However, management of upper aerodigestive and airway venous malformations requires a different approach. Sclerotherapy may injure or slough involved mucosa leading to profound edema, bleeding, and scarring. Excision of aerodigestive lesions requires the removal of involved normal tissue and can lead to functional deficits in breathing, voice, or swallowing. For these reasons surgery and sclerotherapy are secondary modalities in the management of neonatal airway venous malformations.

An alternative and effective treatment for mucosal and airway disease is Nd:YAG laser therapy [114, 151, 152]. This technique utilizes the process of selective photothermolysis to target and injure small to medium sized veins while preserving surrounding normal tissue. The Nd:YAG laser, with a wavelength of 1,064 nm, can heat and disrupt anomalous veins while preserving the overlying mucosa. The treatment is effective up to 7 mm below the mucosal surface. Immediate reduction in the size of involved aerodigestive lesions can be observed at the time of laser treatment. The reduction from a single treatment is often sustained for several months while multiple treatments spaced in 3–6 month intervals are often required for maximal benefit and control of symptoms [153]. This approach can be particularly effective for the tongue and larynx with preservation of function. Airway complications or injury following Nd:YAG therapy have not yet been reported. Interstitial laser therapy, by placing the laser deep into the tongue musculature has also proven effective. Thus the ND:YAG laser therapy of the aerodigestive tract is a healthy alternative to excision or sclerotherapy that are at higher risk of untoward functional deficits.

Cervicofacial venous malformations that compress the airway but do not directly extend into it can be addressed with sclerotherapy, surgical excision, or a combination of these modalities. Because of slow and gradual expansion, neck disease without mucosal involvement will lead to airway obstruction predominately with recumbence and sleep. Symptoms are rarely acute or life threatening in this situation and planned procedures can be performed at appropriately spaced intervals.

Future Considerations

Due to their infiltrative nature, rarely are VMs “cured.” Management techniques of laser, surgery, and sclerotherapy

reduce disease bulk, improve symptoms, and control disease progression. As novel mutations are identified in VMs, molecular targets for pharmacotherapy may become evident and exploited. Laser therapy and sclerotherapy may then selectively destroy VMs by targeted disease specific factors while reducing risk of injury to be involved and adjacent normal tissue.

References

- Mulliken JB, Glowacki J. Hemangiomas and vascular malformations in infants and children: a classification based on endothelial characteristics. *Plast Reconstr Surg.* 1982;69:412–22.
- Enjolras O, Mulliken JB. Vascular tumors and vascular malformations (new issues). *Adv Dermatol.* 1997;13:375–423.
- Mulliken JB, Fishman SJ, Burrows PE. Vascular anomalies. *Curr Probl Surg.* 2000;37:517–84.
- Rahbar R, McGill TJ, Mulliken JB. Vascular tumors and malformations of the head and neck. In: Cummings CW, Flint PW, editors. *Otolaryngology head and neck surgery.* 4th ed. Philadelphia: Mosby; 2005. p. 4013–20.
- Kilcline C, Frieden IJ. Infantile hemangiomas: how common are they? A systematic review of the medical literature. *Pediatr Dermatol.* 2008;25:168–73.
- Pratt AG. Birthmarks in infants. *Arch Dermatol.* 1953;67:302–5.
- Jacobs AH, Walton RG. The incidence of birthmarks in the neonate. *Pediatrics.* 1976;58:218–22.
- Dickison P, Christou E, Wargon O. A prospective study of infantile hemangiomas with a focus on incidence and risk factors. *Pediatr Dermatol.* 2011;28:663–9.
- Hoorweg MJ, Smeulders MJ, Ubbink DT, van der Horst CM. The prevalence and risk factors of infantile haemangiomas: a case-control study in the Dutch population. *Paediatr Perinat Epidemiol.* 2012;26:156–62.
- Haggstrom AN, Drolet BA, Baselga E, Chamlin SL, Garzon MC, Horii KA, et al. Prospective study of infantile hemangiomas: demographic, prenatal, and perinatal characteristics. *J Pediatr.* 2007;150:291–4.
- Chiller KG, Passaro D, Frieden IJ. Hemangiomas of infancy: clinical characteristics, morphologic subtypes, and their relationship to race, ethnicity, and sex. *Arch Dermatol.* 2002;138:1567–76.
- Amir J, Metzker A, Krikler R, Reisner SH. Strawberry hemangioma in preterm infants. *Pediatr Dermatol.* 1986;3:331–2.
- Drolet BA, Swanson EA, Frieden IJ. Infantile hemangiomas: an emerging health issue linked to an increased rate of low birth weight infants. *J Pediatr.* 2008;153:712–5. e1.
- Holinger PH, Brown WT. Congenital webs, cysts, laryngoceles and other anomalies of the larynx. *Ann Otol Rhinol Laryngol.* 1967;76:744–52.
- Perkins JA, Oliyai S, Garrison MM, Manning SC, Christakis DA. Airway procedures and hemangiomas: treatment patterns and outcome in U.S. pediatric hospitals. *Int J Pediatr Otorhinolaryngol.* 2009;73:1302–7.
- Shikhani AH, Marsh BR, Jones MM, et al. Infantile subglottic hemangiomas: an update. *Ann Otol Rhinol Laryngol.* 1986; 95:336–47.
- Sherrington CA, Sim DK, Freezer NJ, Robertson CF. Subglottic haemangioma. *Arch Dis Child.* 1997;76:458–9.
- Feuerstein SS. Subglottic hemangioma in infants. *Laryngoscope.* 1973;83:466–75.
- Chatrath P, Black M, Jani P, et al. A review of the current management of infantile subglottic haemangioma, including a comparison of CO₂ laser therapy versus tracheostomy. *Int J Pediatr Otorhinolaryngol.* 2002;64:143–57.
- Hughes CA, Rezaee A, Ludemann JP, Holinger LD. Management of congenital subglottic hemangioma. *J Otolaryngol.* 1999;28: 223–8.
- Yu Y, Flint AF, Mulliken JB, Wu JK, Bischoff J. Endothelial progenitor cells in infantile hemangioma. *Blood.* 2004;103:1373–5.
- Greenberger S, Boscolo E, Adini I, Mulliken JB, Bischoff J. Corticosteroid suppression of VEGF-A in infantile hemangioma-derived stem cells. *N Engl J Med.* 2010;362:1005–13.
- North PC, Waner M, James CA, Mizeracki A, Frieden IJ, Mihm MC. Congenital nonprogressive hemangioma. A distinct clinicopathologic entity unlike infantile hemangioma. *Arch Dermatol.* 2001;137:1607–20.
- North PE, Waner M, Mizeracki A, Mihm Jr MC. GLUT1: a newly discovered immunohistochemical marker for juvenile hemangiomas. *Hum Pathol.* 2000;31:11–22.
- Leon-Villapalos J, Wolfe K, Kangesu L. GLUT1: an extra diagnostic tool to differentiate between haemangiomas and vascular malformations. *Br J Plast Surg.* 2005;58:348–52.
- Greenberger S, Adini I, Boscolo E, Mulliken JB, Bischoff J. Targeting NF- κ B in infantile hemangioma-derived stem cells reduces VEGF-A expression. *Angiogenesis.* 2010;13:327–35.
- Chang J, Most D, Bresnick S, Mehrara B, Steinbrech DS, Reinisch J, et al. Proliferative hemangiomas: analysis of cytokine gene expression and angiogenesis. *Plast Reconstr Surg.* 1999;103:1–9. discussion 10.
- Razon MJ, Kräling BM, Mulliken JB, Bischoff J. Increased apoptosis coincides with onset of involution in infantile hemangioma. *Microcirculation.* 1998;5:189–95.
- Colonna V, Resta L, Napoli A, Bonifazi E, et al. Placental hypoxia and neonatal haemangioma: clinical and histological observations. *Br J Dermatol.* 2010;162:208–9.
- Drolet BA, Frieden IJ. Characteristics of infantile hemangiomas as clues to pathogenesis: does hypoxia connect the dots? *Arch Dermatol.* 2010;146:1295–9.
- Kleinman ME, Blei F, Gurtner GC. Circulating endothelial progenitor cells and vascular anomalies. *Lymphat Res Biol.* 2005;3: 234–9.
- Bauland CG, Smit JM, Bartelink LR, Zondervan HA, Spauwen PH. Hemangioma in the newborn: increased incidence after chorionic villus sampling. *Prenat Diagn.* 2010;30:913–7.
- Bauland CG, Smit JM, Scheffers SM, Bartels RH, van den Berg P, Zeebregts CJ, et al. Similar risk for hemangiomas after amniocentesis and transabdominal chorionic villus sampling. *J Obstet Gynaecol Res.* 2012;38:371–5.
- Lister WA. Natural history of strawberry nevi. *Lancet.* 1938; 231(5991):1429–30.
- Bivings L. Spontaneous regression of angiomas in children: twenty-two years' observation covering 236 cases. *J Pediatr.* 1954;45:643–7.
- Margileth AM, Museles M. Current concepts in diagnosis and management of congenital cutaneous hemangiomas. *Pediatrics.* 1965;36:410–6.
- Moroz B. Long-term follow-up of hemangiomas in children. In: Williams HB, editor. *Symposium on vascular malformations and melanotic lesions.* St Louis: CV Mosby; 1982. p. 27–35.
- Mulliken JB. Diagnosis and natural history of hemangiomas. In: Mulliken JB, Young AE, editors. *Vascular birthmarks: hemangiomas and malformations.* Philadelphia: WB Saunders; 1988. p. 41–62.
- Esterly NB. Cutaneous hemangiomas, vascular stains and malformations, and associated syndromes. *Curr Probl Pediatr.* 1996; 26:3–39.
- Bowers RE, Graham EA, Tominson KM. The natural history of the strawberry nevus. *Arch Dermatol.* 1960;82:667–70.

41. Chang LC, Haggstrom AN, Drolet BA, Baselga E, Chamlin SL, Garzon MC, et al. Growth characteristics of infantile hemangiomas: implications for management. *Pediatrics*. 2008;122:360–7.
42. Bitar MA, Moukarbel RV, Zalzal GH. Management of congenital subglottic hemangioma: trends and success over the past 17 years. *Otolaryngol Head Neck Surg*. 2005;132:226–31.
43. Perkins JA, Duke W, Chen E, Manning S. Emerging concepts in airway infantile hemangioma assessment and management. *Otolaryngol Head Neck Surg*. 2009;141:207–12.
44. Sie KC, McGill T, Healy GB. Subglottic hemangiomas: 10 years experience with the carbon dioxide laser. *Ann Otol Rhinol Laryngol*. 1994;103:167–72.
45. Orlow S, Isakoff M, Blei F. Increased risk of symptomatic hemangiomas of the airway in association with cutaneous hemangiomas in a “beard” distribution. *J Pediatr*. 1997;131:643–6.
46. O TM, Alexander RE, Lando T, et al. Segmental hemangiomas of the upper airway. *Laryngoscope*. 2009;119:2242–7.
47. Perkins JA, Chen EY, Hoffer FA, Manning SC. Proposal for staging airway hemangiomas. *Otolaryngol Head Neck Surg*. 2009;141:516–21.
48. North PE, Waner M, Mizeracki A, Mrak RE, Nicholas R, Kincannon J, et al. A unique microvascular phenotype shared by juvenile hemangiomas and human placenta. *Arch Dermatol*. 2001;137:559–70.
49. Badi AN, Kerschner JE, North PE, Drolet BA, Messner A, Perkins JA. Histopathologic and immunophenotypic profile of subglottic hemangioma: multicenter study. *Int J Pediatr Otorhinolaryngol*. 2009;73:1187–91.
50. Léauté-Labrèze C, Dumas de la Roque E, Hubiche T, Boralevi F, Thambo JB, Taïeb A. Propranolol for severe hemangiomas of infancy. *N Engl J Med*. 2008;358:2649–51.
51. Sans V, Dumas de la Roque E, Berge J, et al. Propranolol for severe infantile hemangiomas: follow-up report. *Pediatrics*. 2009;124:e423–31.
52. Tan ST, Itinteang T, Leadbitter P. Low-dose propranolol for multiple hepatic and cutaneous hemangiomas with deranged liver function. *Pediatrics*. 2011;127:e772–6.
53. Mousa W, Kues K, Haas E, et al. Successful treatment of a large hemangioma with propranolol. *J Dtsch Dermatol Ges*. 2010;8:184–6.
54. Marsciani A, Pericoli R, Alaggio R, Brisigotti M, Vergine G. Massive response of severe infantile hepatic hemangioma to propranolol. *Pediatr Blood Cancer*. 2010;54:176.
55. Saint-Jean M, Léauté-Labrèze C, Mazereeuw-Hautier J, Bodak N, Hamel-Teillac D, Kupfer-Bessagnet I, et al. Propranolol for treatment of ulcerated infantile hemangiomas. *J Am Acad Dermatol*. 2011;64:827–32.
56. Cheng JF, Gole GA, Sullivan TJ. Propranolol in the management of periorbital infantile haemangioma. *Clin Experiment Ophthalmol*. 2010;38:547–53.
57. Li YC, McCahon E, Rowe NA, Martin PA, Wilcsek GA, Martin FJ. Successful treatment of infantile haemangiomas of the orbit with propranolol. *Clin Experiment Ophthalmol*. 2010;38:554–9.
58. Fulkerson DH, Agim NG, Al-Shamy G, Metry DW, Izaddoost SA, Jea A. Emergent medical and surgical management of mediastinal infantile hemangioma with symptomatic spinal cord compression: case report and literature review. *Childs Nerv Syst*. 2010;26:1799–805.
59. Vanlander A, Decaluwe W, Vandelanotte M, Van Geet C, Cornette L. Propranolol as a novel treatment for congenital visceral haemangioma. *Neonatology*. 2010;98:229–31.
60. Theletsane T, Redfern A, Raynham O, Harris T, Prose NS, Khumalo NP. Life threatening infantile haemangioma: a dramatic response to propranolol. *J Eur Acad Dermatol Venereol*. 2009;23:1465–6.
61. Hogeling M, Adams S, Wargon O. A randomized controlled trial of propranolol for infantile hemangiomas. *Pediatrics*. 2011;128:e259–66.
62. Drolet BA, Frommelt PC, Chamlin SL, Haggstrom A, Bauman NM, Chiu YE, et al. Initiation and use of propranolol for infantile hemangioma: report of a consensus conference. *Pediatrics*. 2013;131:128–40.
63. Greenberger S, Bischoff J. Infantile hemangioma-Mechanism(s) of drug action on a vascular tumor. *Cold Spring Harb Perspect Med*. 2011;1(1):a006460.
64. Dai Y, Hou F, Buckmiller L, et al. Decreased eNOS protein expression in involuting and propranolol-treated hemangiomas. *Arch Otolaryngol Head Neck Surg*. 2012;138:177–82.
65. Frieden IJ, Drolet BA. Propranolol for infantile hemangiomas: promise, peril, pathogenesis. *Pediatr Dermatol*. 2009;26:642–4.
66. Mistry N, Tzifa K. Use of propranolol to treat multicentric airway haemangioma. *J Laryngol Otol*. 2010;7:1–4.
67. Truong MT, Chang KW, Berk DR, Heerema-McKenney A, Bruckner AL. Propranolol for the treatment of a life-threatening subglottic and mediastinal infantile hemangioma. *J Pediatr*. 2010;156:335–8.
68. Maturro S, Hartnick C. Initial experience using propranolol as the sole treatment for infantile airway hemangiomas. *Int J Pediatr Otorhinolaryngol*. 2010;74:323–5.
69. Buckmiller LM, Munson PD, Dyamenahalli U, Dai Y, Richter GT. Propranolol for infantile hemangiomas: early experience at a tertiary vascular anomalies center. *Laryngoscope*. 2010;120:676–81.
70. Manunza F, Syed S, Laguda B, et al. Propranolol for complicated infantile haemangiomas: a case series of 30 infants. *Br J Dermatol*. 2010;162:466–8.
71. Jephson CG, Manunza F, Syed S, Mills NA, Harper J, Hartley BE. Successful treatment of isolated subglottic haemangioma with propranolol alone. *Int J Pediatr Otorhinolaryngol*. 2009;73:1821–3.
72. Buckmiller L, Dyamenahalli U, Richter GT. Propranolol for airway hemangiomas: case report of novel treatment. *Laryngoscope*. 2009;119:2051–4.
73. Le Boulanger N, Fayoux P, Teissier N, Cox A, Van Den Abbeele T, Carrabin L, et al. Propranolol in the therapeutic strategy of infantile laryngotracheal hemangioma: A preliminary retrospective study of French experience. *Int J Pediatr Otorhinolaryngol*. 2010;74:1254–7.
74. Anderson de Moreno LC, Matt BH, Montgomery G, Kim YJ. Propranolol in the treatment of upper airway hemangiomas. *Ear Nose Throat J*. 2013;92:209–14.
75. Mahadevan M, Cheng A, Barber C. Treatment of subglottic hemangiomas with propranolol: initial experience in 10 infants. *ANZ J Surg*. 2011;81:456–61.
76. Rosbe KW, Suh KY, Meyer AK, Maguiness SM, Frieden IJ. Propranolol in the management of airway infantile hemangiomas. *Arch Otolaryngol Head Neck Surg*. 2010;136:658–65.
77. Raol N, Metry D, Edmonds J, Chandy B, Sulek M, Larrier D. Propranolol for the treatment of subglottic hemangiomas. *Int J Pediatr Otorhinolaryngol*. 2011;75:1510–4.
78. Bajaj Y, Kapoor K, Ifeacho S, Jephson CG, Albert DM, Harper JJ, et al. Great Ormond Street Hospital treatment guidelines for use of propranolol in infantile isolated subglottic haemangioma. *J Laryngol Otol*. 2013;127:295–8.
79. Parikh SR, Darrow DH, Grimmer JF, Manning SC, Richter GT, Perkins JA. Propranolol use for infantile hemangiomas: American Society of Pediatric Otolaryngology Vascular Anomalies Task Force practice patterns. *JAMA Otolaryngol Head Neck Surg*. 2013;139:153–6.
80. Lawley LP, Siegfried E, Todd JL. Propranolol treatment for hemangioma of infancy: risks and recommendations. *Pediatr Dermatol*. 2009;26:610–4.
81. Denoyelle F, Garabédian EN. Propranolol may become first-line treatment in obstructive subglottic infantile hemangiomas. *Otolaryngol Head Neck Surg*. 2010;142:463–4.
82. Waner M, Suen JY. Treatment options for the management of hemangiomas. In: Waner M, Suen JY, editors. *Hemangiomas and*

- vascular malformations of the head and neck. New York: Wiley-Liss, Inc.; 1999. p. 233–62.
83. Waner M, Suen JY. Advances in the management of congenital vascular lesions of the head and neck. *Adv Otorhinolaryngol.* 1996;10:31–54.
 84. Ezekowitz RAB, Mulliken JB, Folkman J. Interferon alfa-2A therapy for life-threatening hemangiomas of infancy. *N Engl J Med.* 1992;326:1456–63.
 85. Ohlms LA, Jones DT, McGill TJ, Healy GB. Interferon alfa2A therapy for airway hemangiomas. *Ann Otol Rhinol Laryngol.* 1994;103:1–8.
 86. Bauman MN, Burke DK, Smith RJH. Treatment of massive or life-threatening hemangiomas with recombinant α 2a-interferon. *Otolaryngol Head Neck Surg.* 1997;117:99–110.
 87. MacArthur CJ, Senders CW, Katz J. The use of interferon Alfa-2A for life-threatening hemangiomas. *Arch Otolaryngol Head Neck Surg.* 1995;121:690–3.
 88. Barlow CF, Priebe CJ, Mulliken JB, et al. Spastic diplegia as a complication of interferon Alpha-2a treatment of hemangiomas of infancy. *J Pediatr.* 1998;132:527–30.
 89. Michaud AP, Bauman NM, Burke DK, Manaligod JM, Smith RJ. Spastic diplegia and other motor disturbances in infants receiving interferon-alpha. *Laryngoscope.* 2004;114:1231–6.
 90. Hoeve LJ, Kuppers GLE, Verwoerd CDA. Management of infantile subglottic hemangioma: laser vaporization, submucous resection, intubation or intralesional steroids? *Int J Pediatr Otorhinolaryngol.* 1997;42:179–86.
 91. Walker P, Cooper D, MacDonald D. Subglottic haemangioma: controversies in management. *J Paediatr Child Health.* 1999;35:392–5.
 92. Re M, Forte V, Berardi C, Mallardi V. Role of endoscopic CO₂ laser surgery in the treatment of congenital infantile subglottic hemangioma. Experience in the Department of Otolaryngology, “Sick Children Hospital”, Toronto, Canada. *Acta Otorhinolaryngol Ital.* 2003;23:175–9.
 93. Madgy D, Ahsan SF, Kest D, Stein I. The application of the Potassium-Titanyl-Phosphate (KTP) laser in the management of subglottic hemangioma. *Arch Otolaryngol Head Neck Surg.* 2001;127:47–50.
 94. Kacker A, April M, Ward RF. Use of potassium titanyl phosphate (KTP) laser in management of subglottic hemangiomas. *Int J Pediatr Otorhinolaryngol.* 2001;59:15–21.
 95. Fu CH, Lee LA, Fang TJ, Wong KS, Li HY. Endoscopic Nd:YAG laser therapy of infantile subglottic hemangioma. *Pediatr Pulmonol.* 2007;42:89–92.
 96. Azizkhan RG. Laser surgery: new applications for pediatric skin and airway lesions. *Curr Opin Pediatr.* 2003;15:243–7.
 97. Cotton RT, Tewfik T. Laryngeal stenosis following carbon dioxide laser in subglottic hemangioma. *Ann Otol Rhinol Laryngol.* 1985;94:494–7.
 98. Pransky SM, Canto C. Management of subglottic hemangioma. *Curr Opin Otolaryngol Head Neck Surg.* 2004;12:509–12.
 99. Jai H, Huang Q, Lü J, Li J, Zhang Z, Wang Z, et al. Microdebrider removal under suspension laryngoscopy: an alternative surgical technique for subglottic hemangioma. *Int J Pediatr Otorhinolaryngol.* 2013;77:1424–9.
 100. Froehlich P, Stamm D, Floret D, Morgon A. Management of subglottic hemangioma. *Clin Otolaryngol.* 1995;4:336–9.
 101. Wiatrak BJ, Reilly JS, Seid AD, Pransky SM, Castillo JV. Open surgical excision of subglottic hemangioma in children. *Int J Pediatr Otorhinolaryngol.* 1996;34:191–206.
 102. Van Den Abbeele T, Triglia J-M, Lescanne E, et al. Surgical removal of subglottic hemangiomas in children. *Laryngoscope.* 1999;109:1281–6.
 103. Vijayasekaran S, White D, Hartley BEJ, Rutter MJ, Elluru RG, Cotton RT. Open excision of subglottic hemangiomas to avoid tracheostomy. *Arch Otolaryngol Head Neck Surg.* 2006;132:159–63.
 104. Bajaj Y, Hartley BEJ, Wyatt ME, Albert DM, Bailey CM. Subglottic haemangioma in children: experience with open surgical excision. *J Laryngol Otol.* 2006;120:1033–7.
 105. O-Lee TJ, Messner A. Open excision of subglottic hemangioma with microscopic dissection. *Int J Pediatr Otorhinolaryngol.* 2007;71:1371–6.
 106. Frieden IJ, Reese V, Cohen D. PHACE syndrome: the association of posterior fossa brain malformations, hemangiomas, arterial anomalies, coarctation of the aorta and cardiac defects, and eye abnormalities. *Arch Dermatol.* 1996;132:307–11.
 107. Durr ML, Meyer AK, Huoh KC, Frieden IJ, Rosbe KW. Airway hemangiomas in PHACE syndrome. *Laryngoscope.* 2012;122:2323–9.
 108. Haggstrom AN, Skillman S, Garzon MC, Drolet BA, Holland K, Matt B, et al. Clinical spectrum and risk of PHACE syndrome in cutaneous and airway hemangiomas. *Arch Otolaryngol Head Neck Surg.* 2011;137:680–7.
 109. Duyka LJ, Fan CY, Coviello-Malle JM, Buckmiller L, Suen JY. Progesterone receptors identified in vascular malformations of the head and neck. *Otolaryngol Head Neck Surg.* 2009;141:491–5.
 110. Rahbar R, Rowley H, Perez-Atayde AR, McGill TJ, Healy GB. Delayed presentation of lymphatic malformation of the cervicofacial region: role of trauma. *Ann Otol Rhinol Laryngol.* 2002;111:828–31.
 111. Ohlms LA, Forsen J, Burrows PE. Venous malformation of the pediatric airway. *Int J Pediatr Otorhinolaryngol.* 1996;37:99–114.
 112. Thompson DM, Kasperbauer JL. Congenital cystic hygroma involving the larynx presenting as an airway emergency. *J Natl Med Assoc.* 1994;86:629–32.
 113. O TM, Rickert SM, Diallo AM, et al. Lymphatic malformations of the airway. *Otolaryngol Head Neck Surg.* 2013;149:156–60.
 114. Richter GT, Braswell L. Management of venous malformations. *Facial Plast Surg.* 2012;28:603–10.
 115. Konez O, Burrows PE, Mulliken JB. Cervicofacial venous malformations. MRI features and interventional strategies. *Interv Neuroradiol.* 2002;8:227–34.
 116. Manning SC, Perkins J. Lymphatic malformations. *Curr Opin Otolaryngol Head Neck Surg.* 2013;21:571–5.
 117. Kennedy TL, Whitaker M, Pellitteri P, Wood WE. Cystic hygroma/lymphangioma: a rational approach to management. *Laryngoscope.* 2001;111:1929–37.
 118. Padwa BL, Hayward PG, Ferraro NF, Mulliken JB. Cervicofacial lymphatic malformation: clinical course, surgical intervention, and pathogenesis of skeletal hypertrophy. *Plast Reconstr Surg.* 1995;95:951–60.
 119. Harsha WJ, Perkins JA, Lewis CW, Manning SC. Pediatric admissions and procedures for lymphatic malformations in the United States: 1997 and 2000. *Lymphat Res Biol.* 2005;3:58–65.
 120. Hassanein AH, Mulliken JB, Fishman SJ, Quatrano NA, Zurakowski D, Greene AK. Lymphatic malformation: risk of progression during childhood and adolescence. *J Craniofac Surg.* 2012;23:149–52.
 121. Smith RJ, Burke DK, Sato Y, Poust RI, Kimura K, Bauman NM. OK-432 therapy for lymphangiomas. *Arch Otolaryngol Head Neck Surg.* 1996;122:1195–9.
 122. Edwards PD, Rahbar R, Ferraro NF, Burrows PE, Mulliken JB. Lymphatic malformation of the lingual base and oral floor. *Plast Reconstr Surg.* 2005;115:1906–15.
 123. Wiegand S, Eivazi B, Zimmermann AP, et al. Microcystic lymphatic malformations of the tongue: diagnosis, classification, and treatment. *Arch Otolaryngol Head Neck Surg.* 2009;135:976–83.
 124. Stefani S, Bazzana T, Smussi C, et al. EXIT (Ex utero Intrapartum Treatment) in lymphatic malformations of the head and neck:

- discussion of three cases and proposal of an EXIT-TTP (Team Time Procedure) list. *Int J Pediatr Otorhinolaryngol.* 2012;76:20–7.
125. Lei ZM, Huang XX, Sun ZJ, Zhang WF, Zhao YF. Surgery of lymphatic malformations in oral and cervicofacial regions in children. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2007;104:338–44.
 126. Perkins JA, Manning SC, Tempero RM, Cunningham MJ, Edmonds Jr JL, Hoffer FA, et al. Lymphatic malformations: review of current treatment. *Otolaryngol Head Neck Surg.* 2010;142:795–803. 803 e791.
 127. Renton JP, Smith RJ. Current treatment paradigms in the management of lymphatic malformations. *Laryngoscope.* 2011;121:56–9.
 128. Dubois J, Alison M. Vascular anomalies: what a radiologist needs to know. *Pediatr Radiol.* 2010;40:895–905.
 129. Dasgupta R, Adams D, Elluru R, Wentzel MS, Azizkhan RG. Noninterventonal treatment of selected head and neck lymphatic malformations. *J Pediatr Surg.* 2008;43:869–73.
 130. Perkins JA, Maniglia C, Magit A, Sidhu M, Manning SC, Chen EY. Clinical and radiographic findings in children with spontaneous lymphatic malformation regression. *Otolaryngol Head Neck Surg.* 2008;138:772–7.
 131. Boardman SJ, Cochrane LA, Roebuck D, Elliott MJ, Hartley BE. Multimodality treatment of pediatric lymphatic malformations of the head and neck using surgery and sclerotherapy. *Arch Otolaryngol Head Neck Surg.* 2010;136:270–6.
 132. Nehra D, Jacobson L, Barnes P, Mallory B, Albanese CT, Sylvester KG. Doxycycline sclerotherapy as primary treatment of head and neck lymphatic malformations in children. *J Pediatr Surg.* 2008;43:451–60.
 133. Smith MC, Zimmerman MB, Burke DK, Bauman NM, Sato Y, Smith RJ. Efficacy and safety of OK-432 immunotherapy of lymphatic malformations. *Laryngoscope.* 2009;119:107–15.
 134. Cahill AM, Nijs E, Ballah D, et al. Percutaneous sclerotherapy in neonatal and infant head and neck lymphatic malformations: a single center experience. *J Pediatr Surg.* 2011;46:2083–95.
 135. Shiels 2nd WE, Kang DR, Murakami JW, Hogan MJ, Wiet GJ. Percutaneous treatment of lymphatic malformations. *Otolaryngol Head Neck Surg.* 2009;141:219–24.
 136. Chen WL, Zhang B, Wang JG, Ye HS, Zhang DM, Huang ZQ. Surgical excision of cervicofacial giant macrocystic lymphatic malformations in infants and children. *Int J Pediatr Otorhinolaryngol.* 2009;73:833–7.
 137. Otteson TD, Hackam DJ, Mandell DL. The Ex Utero Intrapartum Treatment (EXIT) procedure: new challenges. *Arch Otolaryngol Head Neck Surg.* 2006;132:686–9.
 138. Burrows PE, Mitri RK, Alomari A, et al. Percutaneous sclerotherapy of lymphatic malformations with doxycycline. *Lymphat Res Biol.* 2008;6:209–16.
 139. Bai Y, Jia J, Huang XX, Alsharif MJ, Zhao JH, Zhao YF. Sclerotherapy of microcystic lymphatic malformations in oral and facial regions. *J Oral Maxillofac Surg.* 2009;67:251–6.
 140. Sobol SE, Manoukian JJ. Acute airway obstruction from a laryngeal lymphangioma in a child. *Int J Pediatr Otorhinolaryngol.* 2001;58:255–7.
 141. Kenton A, Duncan N, Bhakta K, Fernandes CJ. Laryngeal lymphatic malformation in a newborn. *J Perinatol.* 2003;23:567–71.
 142. Chan J, Younes A, Koltai PJ. Occult supraglottic lymphatic malformation presenting as obstructive sleep apnea. *Int J Pediatr Otorhinolaryngol.* 2003;67:293–6.
 143. Berg EE, Sobol SE, Jacobs I. Laryngeal obstruction by cervical and endolaryngeal lymphatic malformations in children: proposed staging system and review of treatment. *Ann Otol Rhinol Laryngol.* 2013;122:575–81.
 144. Brouillard P, Vikkula M. Genetic causes of vascular malformations. *Hum Mol Genet.* 2007;16(Spec No. 2):R140–9.
 145. Hassanein AH, Mulliken JB, Fishman SJ, Alomari AI, Zurakowski D, Greene AK. Venous malformation: risk of progression during childhood and adolescence. *Ann Plast Surg.* 2012;68:198–201.
 146. Gallione CJ, Pasyk KA, Boon LM, et al. A gene for familial venous malformations maps to chromosome 9p in a second large kindred. *J Med Genet.* 1995;32:197. e9.
 147. Eifert S, Villavicencio JL, Kao TC, Taute BM, Rich NM. Prevalence of deep venous anomalies in congenital vascular malformations of venous predominance. *J Vasc Surg.* 2000;31(3):462–71.
 148. Gloviczki P, Stanson AW, Stickler GB, Johnson CM, Toomey BJ, Meland NB, Rooke TW, Cherry Jr KJ. Klippel-Trenaunay syndrome: the risks and benefits of vascular interventions. *Surgery.* 1991;110(3):469–79.
 149. Glade RS, Richter GT, James CA, Suen JY, Buckmiller LM. Diagnosis and management of pediatric cervicofacial venous malformations: retrospective review from a vascular anomalies center. *Laryngoscope.* 2010;120:229–35.
 150. Domp Martin A, Vikkula M, Boon LM. Venous malformation: update on aetiopathogenesis, diagnosis and management. *Phlebology.* 2010;25:224–35.
 151. Eivazi B, Wiegand S, Teymoortash A, Neff A, Werner JA. Laser treatment of mucosal venous malformations of the upper aerodigestive tract in 50 patients. *Lasers Med Sci.* 2010;25:571–6.
 152. Scherer K, Waner M. Nd:YAG lasers (1,064 nm) in the treatment of venous malformations of the face and neck: challenges and benefits. *Lasers Med Sci.* 2007;22:119–26.
 153. Glade R, Vinson K, Richter G, Suen JY, Buckmiller LM. Endoscopic management of airway venous malformations with Nd:YAG laser. *Ann Otol Rhinol Laryngol.* 2010;119:289–93.