

Chapter 15

Hepatic Encephalopathy: A Primary Neurogliopathy

Sharon DeMorrow and Roger F. Butterworth

Abstract Hepatic encephalopathy (HE) and brain edema are serious central nervous complications of both acute and chronic liver failure. HE starts as a neuropsychiatric syndrome beginning with changes in personality and sleep disturbances progressing through cognitive dysfunction and motor symptoms to stupor and coma. In acute liver failure (ALF) progression may be very rapid, sometimes of the order of days (Butterworth and Vaquero, *The liver; biology and pathobiology*, pp. 600–617, 2009). Since the appearance of overt signs of encephalopathy in liver failure generally heralds a poor prognosis with a significant impact on clinical management options, on liver transplant outcome and on health-related quality of life, rational effective therapies are urgently required. Such therapies require knowledge of the precise mechanisms implicated in the pathogenesis of HE.

Keywords Hepatic encephalopathy · Liver failure · Cirrhosis · Alzheimer type 2 astrocytosis · Brain edema · Microglial activation · Minocycline · Chemokines

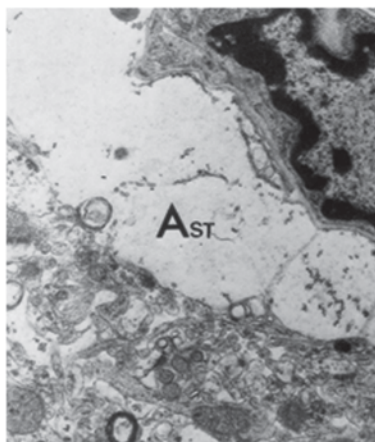
15.1 Neuroglial Pathology in Liver Failure

Pathologically, HE is characterized primarily by neuroglial alterations, the nature and extent of which relate to the type of liver failure (acute or chronic) and the severity of the liver injury. Terminal stages of ALF frequently result in brain herniation caused by intracranial hypertension resulting from brain edema and electron microscopic studies reveal that the brain edema in ALF is primarily cytotoxic (due to cell swelling) rather than vasogenic (due to breakdown of the blood-brain barrier) in nature (Kato et al. 1992). The cell manifesting the most severe swelling in ALF is the astrocyte (Fig. 15.1).

R. F. Butterworth (✉)
Neuroscience Research Unit, Hopital St-Luc (CHUM), 1058 St-Denis,
Montreal, QC H2X 3J4, Canada
e-mail: rogerbutterworthuk@yahoo.com

S. DeMorrow
Department of Internal Medicine, Texas A&M Health Science Center College of Medicine,
1901 South 1st Street, Temple, TX 76504, USA
e-mail: demorrow@medicine.tamhsc.edu

Fig. 15.1 Acute hyperammonemia resulting from liver failure results in astrocyte swelling. A representative electron micrograph showing swelling of a perivascular astrocyte (Ast) from a patient with acute liver failure who died of brain herniation. (Figure adapted from Felipo and Butterworth (2002) and reproduced with permission from Elsevier)



In contrast to ALF, end-stage chronic liver failure (cirrhosis) results in characteristic pathological changes to the astrocyte; these changes are known as Alzheimer type 2 astrocytosis where the cell nuclei take on a characteristic shape and pattern consisting of marked pallor and swelling with a prominent nucleolus, margination of the chromatin pattern and glycogen deposition (Fig. 15.2). The nuclei may take on an array of shapes and forms depending upon the brain structure examined. These forms range from the classical spherical shape in the cerebral cortex to irregular lobulated forms in some basal ganglia structures; the appearance of multiplets has been reported to suggest an element of hyperplasia (Norenberg 1987).

The occurrence of multiple episodes of grade 4 (or stage IV) HE (coma) and/or a single prolonged period of coma in a patient with end-stage chronic liver failure may result in a condition known as “acquired non-Wilsonian hepatocerebral degeneration”, the neuropathological features of which include Alzheimer type 2 astrocytosis. In addition, varying degrees of neuronal cell loss occur in cerebral cortex, cerebellum and basal ganglia structures (Butterworth 2007). A related condition known as “Parkinsonism in cirrhosis” appears to result from manganese deposition in substantia nigra of these patients (Butterworth 2013b).

Evidence of discrete alterations of the blood-brain barrier in HE have been proposed based upon studies of barrier permeability in an animal model of ALF (Nguyen 2010) but these findings were not confirmed by others (Bemour et al. 2010a). Swelling of cerebrovascular endothelial cells has been demonstrated using electron microscopy in material from ALF patients (Kato et al. 1992) again suggesting that, if barrier changes do occur in HE, they are discrete in nature.

Microglial activation and its role in the pathogenesis of HE was first described in the brains of animals with ALF resulting from hepatic devascularisation (Jiang et al. 2006). Subsequent studies in this model revealed that activation of microglia was accompanied by grade 4 HE (coma) and brain edema as well as by increased expression of genes coding for pro-inflammatory cytokines in brain (Jiang et al. 2009a, b)

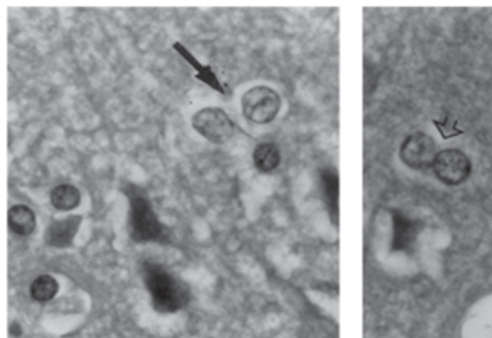


Fig. 15.2 Chronic hyperammonemia resulting from liver cirrhosis results in Alzheimer type II astrocytes. Alzheimer type II cells (*arrow, left-hand panel*) show enlarged nuclei and margination of chromatin in this hematoxylin and eosin stained section of frontal cortex from a 51-year-old cirrhotic patient who died in hepatic coma. Doublets suggestive of a proliferative response are frequently seen (*right-hand panel, arrow head*). (Figure adapted from Felipo and Butterworth (2002) and reproduced with permission from Elsevier)

(Fig. 15.3). Microglial activation has subsequently been confirmed using various techniques and cellular markers in an animal model of ALF resulting from toxic liver injury (McMillin et al. 2012b; Rangroo Thrane et al. 2012). Activation of microglia has also been reported in material from a patient with ALF resulting from viral hepatitis (Butterworth 2011), in a material from cirrhotic patients who died in hepatic coma (Zemtsova et al. 2011) and increased signals in positron emission tomography (PET) studies in HE patients was attributed to microglial activation (Cagnin et al. 2006). Microglial activation has also been described in an animal model of biliary cirrhosis (D’Mello et al. 2009).

The subject of activation of microglia in HE and its relationship to pro-inflammatory mechanisms and the role of cytokines/chemokines in the pathogenesis of the CNS complications of liver failure are covered in-depth in a later section of this chapter.

15.2 Abnormalities of Neuroglial Function in Liver Failure

In addition to frank neuroglial pathology, both acute and chronic liver failure result in a wide range of alterations of neuroglial function including deficits in neuroglial cell volume regulation and neuroglial-neuronal metabolic trafficking of key intermediates as well as neuroinflammatory changes. Some of these changes appear to relate to exposure of neuroglia *in situ* to ammonia and manganese, two substances known to accumulate to toxic concentrations in brain in liver failure.

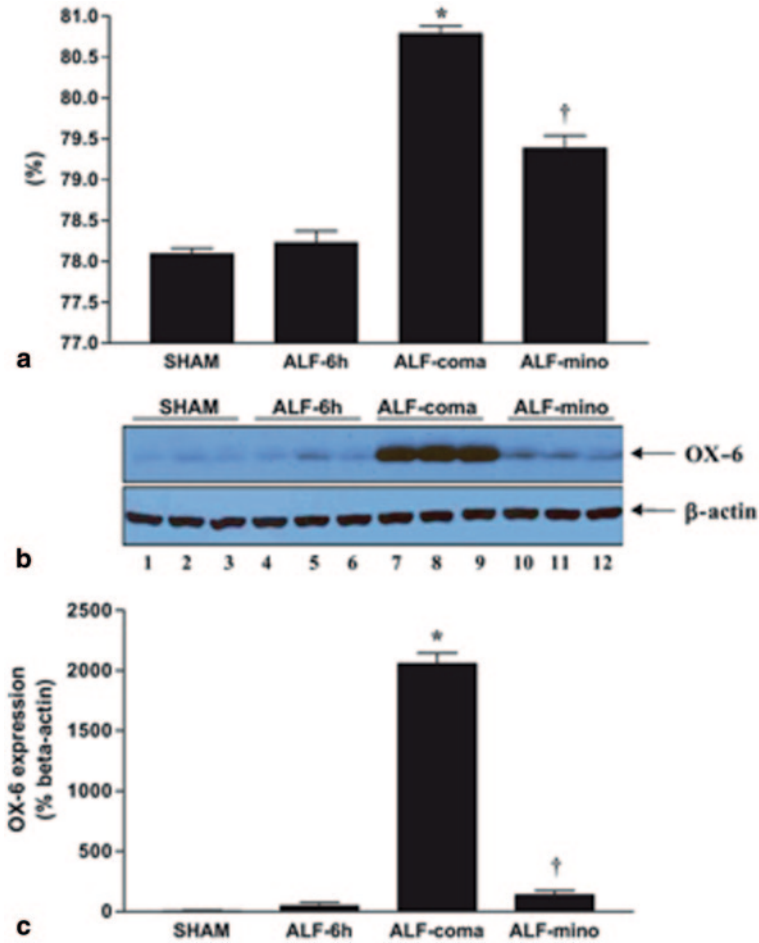


Fig. 15.3 Western blot analysis of OX-6 expression in acute liver failure (ALF) rats reveals that inhibition of microglial activation by minocycline treatment is correlated with attenuation of brain edema. **a** Percentage of brain water content of cerebral cortex from sham-operated controls (sham), ALF rats 6 h post- hepatic artery ligation/HAL (ALF-6 h), ALF rats at coma stage of encephalopathy (ALF-coma) and in ALF rats treated with minocycline (ALF-mino); **b** OX-6 protein expression in cerebral cortex from sham-operated controls (lanes 1–3), ALF rats 6 h post-HAL (lanes 4–6), ALF rats at coma stage of encephalopathy (lanes 7–9) and in ALF rats treated with minocycline (lanes 10–12); **c** Histogram representation of OX-6 expression in the various treatment groups. Data represent mean ± SEM of n=10 animals per treatment group. Significant differences indicated by *p<0.01 versus sham-operated controls and ALF-6 h; †p<0.001 versus ALF-coma (ANOVA with post hoc Tukey’s test). (Figure reproduced from Jiang et al 2009a, with permission from Wiley)

15.2.1 Neuroglial Cell Volume Regulation

In ALF, it is generally considered that failure of cell volume regulation in astroglia underpins the phenomenon of cytotoxic brain edema that so frequently leads to increased intracranial pressure, brain herniation and patient death. Various mechanisms have been proposed to explain this failure of neuroglial cell volume regulation in ALF that include as described in two subheadings below.

15.2.1.1 Osmotic Effects of Increased Intracellular Glutamine

Being devoid of a functional urea cycle, brain relies upon glutamine synthesis for the removal of excess ammonia and the enzyme responsible, glutamine synthetase (GS) has a predominantly, if not exclusively, astroglial localization. Brain and cerebrospinal fluid (CSF) ammonia and glutamine concentrations correlate well with severity of HE in both ALF (Clemmesen et al. 1999) and in end-stage cirrhosis (Laubenberger et al. 1997). Based upon studies in cultured cortical astrocytes exposed to ammonia as well as in animal models of pure hyperammonemia (in the absence of liver failure), it has been proposed that the accumulation of glutamine in brain in liver failure leads to an osmotic gradient that contributes to cell swelling. This hypothesis is supported by a report describing a protective effect of the GS inhibitor methionine sulfoximine on cell swelling and brain edema in hyperammonemic animals (Brusilow et al. 2010). However, although inhibition of glutamine synthesis was beneficial under these conditions, the use of ¹³C-nuclear magnetic resonance spectroscopy failed to show a correlation between *de novo* synthesis of glutamine in brain with either the severity of HE or with the presence of brain edema in animals with ALF (Chatauret et al. 2003) suggesting the presence of additional/alternative factors.

15.2.1.2 Neuroglial Proteins Involved in Cell Volume Regulation

Alterations in expression of genes coding for key neuroglial proteins with suggested roles in cell volume regulation have been reported in experimental models of HE.

Aquaporin-4 (AQP-4) is highly expressed in astroglia, particularly in the end-feet that ensheath brain capillaries where it mediates transmembrane movement of water. An important role for the protein has been suggested to contribute to the pathogenesis of brain edema in a range of clinical situations. In relation to the cerebral complications of liver failure, increased AQP-4 expression was reported in cultured cortical astrocytes exposed to millimolar concentrations of ammonia that led to significant cell swelling (Rama Rao et al. 2003). However, results in animal models of liver failure have so far given conflicting results; whereas HE and brain edema were accompanied by increased concentrations of AQP-4 in one model of ALF (Eefsen et al. 2010), no such changes were reported in a subsequent study (Wright et al. 2010).

In contrast to the equivocal nature of AQP-4 in relation to HE, progressive decreases in expression of the astroglial marker protein, glial fibrillary acidic protein (GFAP) have consistently been reported in both experimental animal models of liver failure and in patient material. In rats with ALF resulting from hepatic devascularisation, GFAP expression was decreased as a function of the increase in brain water (Belanger et al. 2002). Selective decreases of GFAP were previously reported in several brain structures from cirrhotic patients with HE; these structures included cerebral cortex, thalamus and basal ganglia (Sobel et al. 1981) whereas no such changes were subsequently reported in cerebellum of similar patients (Kril et al. 1997). There is evidence to suggest a role of ammonia in the pathogenesis of the loss of GFAP in liver failure, a proposal that is based upon the report of a loss of expression of GFAP in cultured astrocytes exposed to ammonia which resulted in significant cell swelling and destabilization of the GFAP molecule (Neary et al. 1994). It was proposed that loss of expression of GFAP, given its importance as a cytoskeletal protein in astroglia, could lead to altered visco-elastic properties of the cell thus favouring cell swelling (Belanger et al. 2002).

Significant increases in expression of the astroglial 45 kDa isoform of the facilitative glucose transporter GLUT-1 were reported in an animal model of ALF (Belanger et al. 2006) and, as for GFAP, a role for exposure to ammonia was suggested. In addition to its established role in the transport of glucose, it has been proposed that GLUT-1 acts as a water channel leading to the suggestion that up-regulation of the protein in ALF may contribute to (or result from) the appearance of brain edema (Belanger et al. 2006).

One report described significant losses in expression of Kir 4.1, an inwardly-rectifying potassium channel expressed in astroglial endfeet that may have a function, in part, in cell volume regulation, has been described in a rat model of ALF (Obara-Michlewska et al. 2011).

15.2.2 Neuroglial Amino Acid Transporter Proteins

High affinity transport of amino acids is a major function of astroglia. Such transport mechanisms exist to maintain the supply of key intermediates required for cellular energy metabolism, synthetic processes and for the termination of action of neurotransmitters such as glutamate and γ -aminobutyric acid (GABA) that form the basis of the so-called “glial-neuronal metabolic interactions”. Astroglial *glutamate transporters* are essential components of the glutamate-glutamine cycle and are responsible for the removal of excess glutamate from the synaptic cleft. Expression of the sodium-dependent, high affinity glutamate transporters EAAT-1 and EAAT-2 (in human; in rodents GLAST and GLT-1, respectively) have been reported in animal models of both acute and chronic liver failure (Knecht et al. 1997; Suarez et al. 2000) resulting in increases in extracellular brain glutamate concentrations (Vaquero and Butterworth 2006). It has been suggested that limitations in the availability of glutamate in astroglia could limit

ammonia-removal capacity since glutamate is the obligate precursor for GS, and, if so, could limit the synthesis of glutamine.

Decreases in expression of the high affinity *glycine transporter* (GLYT-1) have been reported in cerebral cortical extracts from rats with ALF resulting from hepatic devascularisation (Zwingmann et al. 2002). Given the predominantly astroglial localization of GLYT-1 in cerebral cortex, it was proposed that these changes might relate to increases in availability of extracellular glycine with the potential to activate the glycine neuromodulatory site on the N-methyl-D aspartate (NMDA) subclass of glutamate receptor. NMDA receptor activation has been proposed to explain the hyperexcitability and nitrosative stress that occur in ALF (Vaquero and Butterworth 2006).

Following the cloning and characterization of high affinity *glutamine transporters* (small neutral amino acid transporters or SNATs) and given the consistent finding of increased brain glutamine in HE, a study was undertaken to assess SNATs in an animal model of ALF (Desjardins et al. 2012). Coma/edema stages of encephalopathy were accompanied by a selective decrease in expression of SNAT-5. Given the astroglial localization of SNAT-5 (Cubelos et al. 2005) it was proposed that down-regulation of transporter expression in liver failure could result in effective “trapping” of glutamine in the cell, an action that is consistent with cell swelling due to glutamine accumulation in the astrocyte as had previously been widely proposed. Moreover, since astroglial glutamine functions as immediate precursor of releasable (transmitter) glutamate, a limit upon its availability following decreased release from the astroglial cell has the potential to result in impairment of glutamatergic transmission an action that could result in excessive inhibition that is also characteristic of HE.

15.2.3 Neuroglial Translocator Protein

Translocator protein (TSPO), previously known as the “peripheral-type benzodiazepine receptor” is a mitochondrial protein responsible for the transport of cholesterol into the mitochondrion. The protein is expressed predominantly by neuroglia with both astroglia and microglia exhibiting high levels of expression in mammalian systems. Increased expression of TSPO has been reported in a wide range of hyperammonemic disorders including urea cycle enzymopathies, ALF, animals with end-to-side portacaval anastomoses and in patients with end-stage chronic liver failure (see (Ahboucha and Butterworth 2008) for review). In one study in humans using PET and the TSPO ligand 11C-PK11195, increased signals were observed in anterior cingulate cortex where the magnitude of the increased signals correlated with the degree of cognitive impairment (Cagnin et al. 2006). The origin of these increased signals was considered to relate to microglial activation. Interest in TSPO in HE relates to the well-established relationship of the protein to the GABA system. Activation of neuroglial TSPO results in increased transport of cholesterol into the mitochondrion followed by increased synthesis of so-called “neurosteroids”, one of

which, allopregnanolone is a very high affinity agonist for the neuronal GABA-A receptor. Increases in concentration of allopregnanolone were reported in autopsied brain tissue from patients who died in hepatic coma (Ahboucha and Butterworth 2005). Based upon these findings, it was proposed that the increase in “GABAergic tone” resulting from activation of TSPO sites and the subsequent increase in synthesis of allopregnanolone could also contribute to the excessive neuroinhibition in HE.

15.3 Evidence for Microglial Activation in the Pathogenesis of HE

As stated above, microglial activation has been demonstrated to be a key feature in the pathogenesis of HE due to both ALF and chronic liver cirrhosis. Clinically, indirect evidence for microglial activation has been demonstrated by an upregulation of the microglial marker Ionized calcium binding adaptor molecule 1 (Iba-1) in post mortem cortical brain tissue from patients with liver cirrhosis and HE, when compared to cirrhotic patients without HE (Zemtsova et al. 2011). In addition, data from a comprehensive gene expression profile analysis demonstrated an upregulation of markers for both the pro-inflammatory M1 and anti-inflammatory M2 microglial phenotypes, suggesting that both subpopulations of microglia may be present in patients with HE due to cirrhosis (Gorg et al. 2013). Furthermore, increased [11C]-PK11195 binding to the TSPO in patients with proven cirrhosis and minimal HE was suggested to be a reflection of microglial activation in these patients (Cagnin et al. 2006). Taken together, these clinical data indirectly support a role of microglia activation in HE.

In contrast, evidence for a direct role for microglia activation in the neurological consequences of both ALF and liver cirrhosis is more striking in many animal models of these diseases. Furthermore, in many of the models used, treatment modalities shown to inhibit microglia activation also alleviated or prevented the cognitive impairment and neurological decline observed during HE. Specific details are described below.

15.3.1 Toxic Liver Injury

A range of hepatotoxic agents have been used to uncover basic mechanisms responsible for the CNS complications of liver failure. This topic was reviewed by a panel of experts nominated by The International Society for Hepatic Encephalopathy and Nitrogen Metabolism (ISHEN) who, after careful deliberation, recommended two toxic models based upon the extent of the characterization. The two models of ALF were the azoxymethane (AOM) mouse model and the thioacetamide (TAA) rat model (Butterworth et al. 2009). The AOM mouse model of ALF exhibits many of

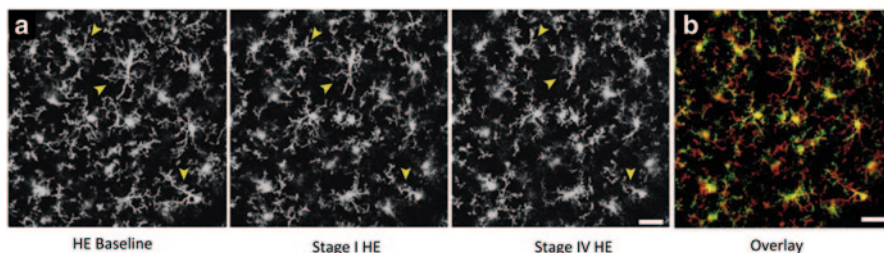


Fig. 15.4 The terminal coma stage of HE is associated with microglial activation. **a** Representative 2 photon laser scanning microscopy images from transgenic mice expressing green fluorescence protein under the control of the CX3CR1 promoter (expressed only in microglia) following AOM administration at 0 h (baseline), stage I and stage IV of HE (collapsed Z-stacks). **b** Overlay of representative images from HE at 0 h (*red*) and stage IV (*green*). *Yellow* represents the overlapping staining. During the latter stage, microglia appear more amoeboid and activated. Scale bar represents 15 μ M. (Figure adapted from Rangroo Thrane et al. (2012) and reproduced with permission from Elsevier)

the pathophysiological characteristics of human HE due to acute liver failure. These features include (1) a clear pattern of neurological behaviors starting with the prodromal phase due to liver failure, where neurological symptoms are not yet evident followed by a number of distinct phases of neurological decline that rapidly progress to stupor and coma; (2) the presence of cerebral edema; and (3) high levels of ammonia in the blood and brain. Very elegant and detailed analyses of the morphological changes in microglia and real-time analysis of microglial dysmotility after AOM have been demonstrated (Rangroo Thrane et al. 2012). Both microglia activation (as demonstrated by an amoeboid phenotype) and motility (as demonstrated by analysis of the turnover rate) were shown to be altered in the cerebral cortex at late stages of HE when severe neurological symptoms were evident, coinciding with the appearance of brain edema (Fig. 15.4). Increased OX-42/CD11b immunoreactivity was also demonstrated in the cerebral cortex of AOM-injected mice (Chastre et al. 2012), which was attenuated by treatment with the tumor necrosis factor (TNF)- α neutralizing molecule etanercept. Furthermore, there was a concomitant attenuation of AOM-induced liver injury and decreased expression of neuroinflammatory molecules in the brain after etanercept treatment (Chastre et al. 2012).

15.3.2 Ischemic Liver Failure

Ischemic liver failure, although uncommon, is encountered clinically. Experimental ALF can be induced by the performing an end-to-side portacaval anastomosis followed by hepatic artery ligation. Rats undergoing this surgery exhibit key clinical features of HE, including cerebral edema and hyperammonemia, which ultimately result in grade 4 HE (hepatic coma). An increase in the number of OX-42/CD11b positive microglia has been demonstrated in the frontal cortex, thalamus and

hippocampus starting 6 h after surgery (early stage HE) and worsening at the time of coma/edema (Jiang et al. 2009a, b), which could be alleviated by either mild hypothermia (Jiang et al. 2009b) or minocycline (Jiang et al. 2009a), with a concomitant attenuation of the progression of HE and brain edema.

15.3.3 Portal-Systemic (Bypass) Encephalopathy

In a related, more subtle model of HE induced by end-to-side portacaval shunt surgery alone, without subsequent hepatic artery ligation, rats develop mild cognitive impairment over the following 3–4 weeks. Associated with this mild form of HE, currently referred to clinically as “minimal HE”, is a change in the morphology of major histocompatibility complex class II (MHCII)-positive microglia to a more ameboid, activated phenotype (Agusti et al. 2011). Curiously, these changes were restricted to cerebellum. Chronic infusion of a p38 mitogen-activated protein kinase inhibitor reversed the morphological changes observed in microglia and prevented the cognitive impairment (Agusti et al. 2011).

15.3.4 Biliary Cirrhosis

Obstruction of the common bile duct induces a reproducible model of biliary cirrhosis in rats. Bile duct-ligated (BDL) animals have liver failure, developing jaundice, portal hypertension, portal-systemic shunting, bacterial translocation and immune system dysfunction. BDL rats are hyperammonemic but show only low-grade encephalopathy (decreased locomotor activities) (Butterworth et al. 2009). Using this model, microglia are activated predominantly in the cerebellum, with only traces of activation in the striatum and thalamus (Rodrigo et al. 2010) (Fig. 15.5a). Interestingly, no microglia activation was evident in the frontal cortex. Treatment with ibuprofen reduced the microglia activation and reversed the concomitant cognitive impairments observed (Rodrigo et al. 2010). Similarly, microglia activation has been shown after BDL in mice, as demonstrated by morphological changes in Iba-1 positive microglia (D’Mello et al. 2009). However, in contrast to the rat model, the activation of microglia was localized to the cerebral cortex rather than the cerebellum (Fig. 15.5b). The cause of these region- and species-selective changes remains unknown. As discussed in a separate section of this chapter, the activation of microglia in BDL mice is thought to subsequently recruit monocytes to the brain that contribute to the cognitive impairment observed (Kerfoot et al. 2006).

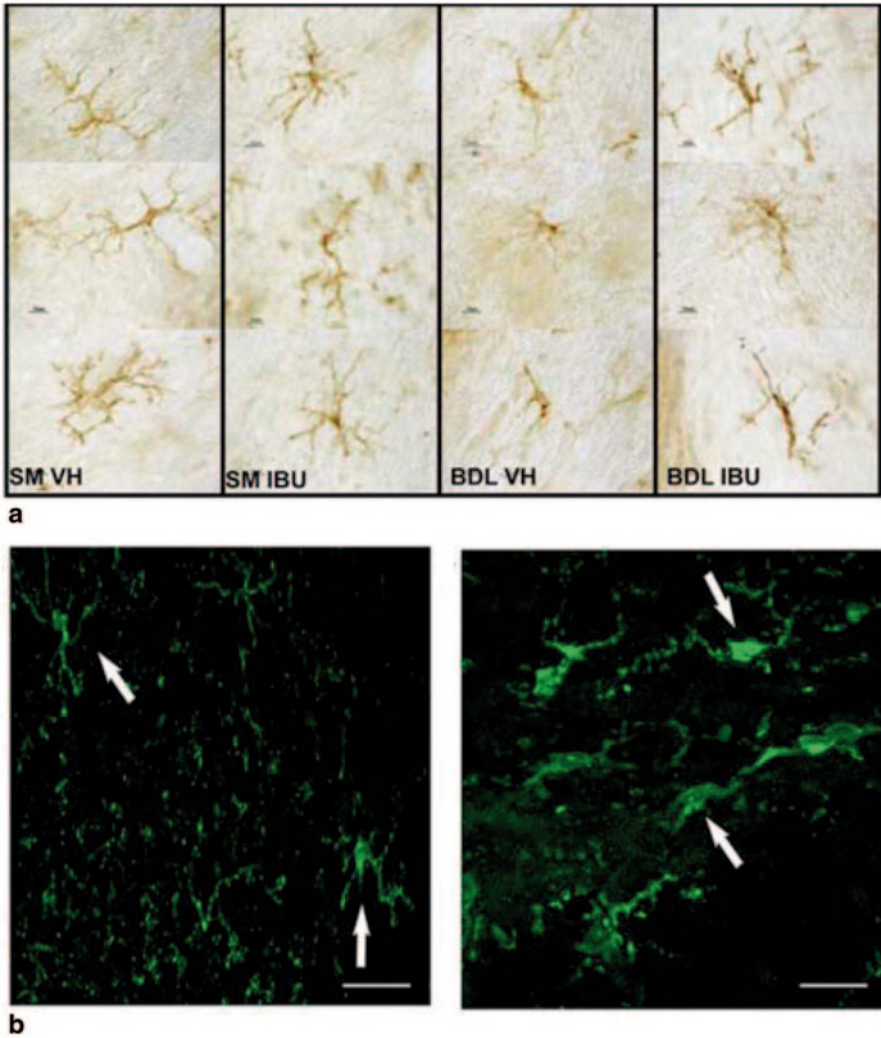


Fig. 15.5 Experimental models of biliary cirrhosis and mild HE are associated with microglial activation. **a** Rats underwent bile duct ligation (*BDL*) or sham (*SM*) surgery and were subsequently treated with either vehicle (*VH*) or Ibuprofen (*IBU*). Microglial activation was assessed using MHCII expression. Microglia in the cerebellum of *BDL* rats showed an activated, amoeboid phenotype. (Figure adapted from Rodrigo et al. (2010), and reproduced with permission from Elsevier.) **b** Immunohistochemistry staining for Iba-1-positive microglia in the cortex of *BDL* mice. Microglia in mice undergoing sham surgery (*left panel*) have resting ramified morphology and longer processes, while after *BDL*, microglia are more rounded with retracted and thicker processes indicative of microglia activation. (Figure adapted from D’Mello et al. (2009) and reproduced with permission from the Society for Neuroscience)

15.3.5 Ammonia Neurotoxicity

It has long been accepted that ammonia plays an important role in the neurological symptoms of HE. Hyperammonemia has been demonstrated in the blood and cerebral cortex at all stages of HE in both ALF and in cirrhosis. Furthermore, injection of a bolus dose of ammonium acetate into rats in the absence of liver failure results in a transient comatose state, supporting a role for ammonia in the neurological decline of HE. In contrast, rats fed an ammonium-containing diet for up to 28 days display cognitive deficits more indicative of minimal HE (Felipo et al. 1988). However, the role of ammonia toxicity in the activation of microglia and subsequent neuro-inflammatory response is not as clearly defined. Treatment of primary microglial cultures with ammonia results in microglia swelling, migration, alterations in filipodia length, and Iba-1 expression (Zemtsova et al. 2011; Lachmann et al. 2013). Similarly, increased Iba-1 immunoreactivity and expression was demonstrated in the cerebral cortex of the *in vivo* rat model of ammonia intoxication (Zemtsova et al. 2011). In contrast, injection of an acute bolus dose of ammonium acetate into mice had no effect on the morphology or turn-over rate of microglia (Rangroo Thrane et al. 2012).

Microglial activation was evident in the cerebellum of chronic ammonium-fed rats, as demonstrated by in-depth morphometric analyses of MHCII positive cells (Rodrigo et al. 2010). In support of a role for microglia activation in the cognitive deficits observed in this chronic hyperammonia model, treatment with ibuprofen reduced the microglia activation and reversed the concomitant cognitive impairments observed (Rodrigo et al. 2010).

A note of caution: It should be borne in mind that experimental preparations, either *in vitro* or *ex vivo* in which mammalian cells in culture or experimental animals with intact livers are exposed to very high doses of ammonia alone are satisfactory preparations only for the study of mechanisms related to ammonia toxicity in normal animals. Given the multiple other factors that are known to affect brain function and that are generated in liver failure such as manganese accumulation, hyponatremia, certain fatty acids and inflammation, extrapolation of findings from ammonia-treatment paradigms to the liver failure clinic should be made with extreme caution.

15.4 Neuroinflammatory Cytokines Associated with Microglial Activation in HE

It is now commonly accepted that the development and progression of HE in liver failure shares a strong relationship with neuroinflammatory processes. Indeed, in patients and in animal models of HE, systemic inflammation causes worsening of the encephalopathy, and it has been proposed that proinflammatory signals may act synergistically with ammonia toxicity to bring about the neurological complications of acute and chronic liver failure (Shawcross et al. 2004; Butterworth 2011, 2013a).

Inflammatory molecules such as TNF α , interleukin (IL)-1 β and IL-6 levels were increased in patients with ALF as well as in animal models (Wright et al. 2007; Bemeur et al. 2010b). In addition to the proinflammatory cytokines, neuroinflammation can be regulated by chemokines, or chemotactic cytokines. These molecules are involved in cell-cell communication, effecting a directional migration and activating various cell types of the immune system including microglia. A number of chemokines, such as chemokine ligand 2 (CCL2), have been shown to be upregulated in liver failure patients and in animal models of HE (D'Mello et al. 2009; McMillin et al. 2012b). A summary of our current knowledge of the inflammatory cytokines and chemokines in HE can be found in Table 15.1. Key inflammatory molecules and their reported effects on microglial activation and/or function are summarized below:

15.4.1 TNF α

TNF α is a potent proinflammatory cytokine. Circulating levels of TNF α are increased as a function of the severity of HE in both patients (Odeh 2007) and experimental animals (Jiang et al. 2009a) with liver failure. Moreover, the presence of TNF α gene polymorphisms is known to influence the clinical outcome in patients with ALF (Bernal et al. 1998). In experimental models of ALF, mice lacking the TNF receptor 1 (TNFR1) gene had a delayed onset of encephalopathy and an attenuation of brain edema (Bemeur et al. 2010b). TNF α has been shown to activate microglia in a number of experimental models of neuroinflammation (Lambertsen et al. 2012; Rubio-Perez and Morillas-Ruiz 2012). With respect to HE, as mentioned above, systemic levels of TNF α are increased in the AOM model of ALF (Chastre et al. 2012). Inhibition of TNF α signaling by systemic treatment with etanercept reduced systemic inflammation, attenuated the neurological decline and prevented microglial activation in the cerebral cortex (Chastre et al. 2012). These data support the hypothesis that peripherally-derived TNF α , at least in part, contributes to the microglial activation and subsequent neurological decline in liver failure. In support of this concept, neurological complications occurring in the BDL model of biliary cirrhosis were shown to be the consequence of monocyte recruitment in response to TNF α signaling and occurred via microglial activation. Specifically, peripheral TNF α signaling stimulates microglia to produce CCL2, which subsequently mediates monocyte recruitment into the brain (D'Mello et al. 2009). These findings were suggested to constitute a novel immune-to-brain communication pathway with the potential to result in altered neuronal excitability and neurological complications in cholestatic liver disease. A schematic diagram of this concept can be found in Fig. 15.6.

Table 15.1 Known inflammatory markers in HE

Inflammatory modulator	Model and species	Tissue assessed	Mechanism	Reference
TNF α	Hepatic devascularization in rats	Frontal cortex/serum/CSF	Specific role for TNF α not assessed; however, treatment with the anti-inflammatory minocycline or hypothermia inhibited microglia activation and cognitive deficits	Jiang et al (2009a, b)
	Bile duct resection in mice	Cerebral cortex, whole blood	Shown to be upregulated in the circulation, Thought to be responsible for microglia activation (TNFR1 knockout mice have dampened microglia response after bile duct resection)	D'Mello et al. (2009)
	Azoxymethane model of acute liver failure in mice	Frontal cortex	Onset of severe encephalopathy (coma) and brain edema was delayed in mice lacking the TNFR	Bemeur et al. (2010b)
	Portacaval shunt in rats	Cerebellum and plasma	Specific role for TNF α not assessed; however, treatment with specific p38 MAPK inhibitor inhibited microglia activation and cognitive deficits	Agusti et al. (2011)
	Azoxymethane model of acute liver failure in mice	Plasma	TNF antagonist etanercept reduced systemic inflammation, microglial activation and time to coma after AOM treatment	Chastre et al. (2012)
	Acetaminophen model of acute liver failure in mice	Liver/whole brain/serum	Used as a marker for inflammation	Villano et al. 2012
	IL-1 β	Hepatic devascularization in rats	Frontal cortex/serum/CSF	Specific role for IL-1 β not assessed; however, treatment with the anti-inflammatory minocycline or hypothermia inhibited microglia activation and cognitive deficits
Azoxymethane model of acute liver failure in mice		Frontal cortex	Onset of severe encephalopathy (coma) and brain edema was delayed in mice lacking the IL-1 receptor	Bemeur et al. (2010b)
BDL and chronic hyperammonemia in rats		Cerebellum	Specific role for IL-1 β not assessed; however, treatment with the non-steroidal anti-inflammatory drug ibuprofen inhibited microglia activation and cognitive deficits	Rodrigo et al. (2010)

Table 15.1 (continued)

Inflammatory modulator	Model and species	Tissue assessed	Mechanism	Reference
	Portacaval shunt in rats	Cerebellum	Specific role for IL-1 β not assessed; however, treatment with specific p38 MAPK inhibitor inhibited microglia activation and cognitive deficits	Agusti et al. (2011)
IL-6	Hepatic devascularization in rats	Frontalcortex/serum/CSF	Specific role for IL-6 not assessed; however, treatment with the anti-inflammatory minocycline or hypothermia inhibited microglia activation and cognitive deficits	Jiang et al (2009a, b)
	Azoxymethane model of acute liver failure in mice	Plasma and cerebral cortex	Specific role for IL-6 not directly assessed; however, treatment with the TNF antagonist etanercept reduced systemic inflammation, microglial activation and time to coma after AOM treatment	Chastre et al. (2012)
	Acetaminophen model of acute liver failure in mice	Liver/whole brain/serum	Used as a marker for inflammation	Villano et al. (2012)
IL-10	Acetaminophen model of acute liver failure in mice	Liver/whole brain/serum	Used as a marker for inflammation. Upregulation attenuated in brains from transgenic mice expressing the human Serpin B3	Villano et al. (2012)
CCL2	Bile duct resection in mice	Cerebral cortex, whole blood	CCL2 expression increased in activated microglia in response to TNF α signaling. Causes recruitment of CCR2-expressing monocytes in the brain	D'Mello et al. (2009)
	Azoxymethane model of acute liver failure in mice	Cerebral cortex	CCL2 expression increased predominantly in neurons. Correlated to increased microglia activation. Specific inhibitors of CR2 and CCR4 delays neurological decline and time to coma	McMillin et al. (2012b)
	Human autopsy samples of liver cirrhosis	Frontal cortex	CCL2 mRNA and immunoreactivity increased in patients with liver cirrhosis with HE but not in cirrhotics without HE	Bradley et al. (2013)

Table 15.1 (continued)

Inflammatory modulator	Model and species	Tissue assessed	Mechanism	Reference
TGF β	Azoxymethane model of acute liver failure in mice	Liver/serum/ cerebral cortex	TGF β is thought to be derived from the periphery. Systemic treatment with a TGF β specific neutralizing antibody attenuates the microglia activation and delays neurological decline	McMillin et al. (2012a, 2013)

AOM azoxymethane, *BDL* bile duct ligation, *CCL2* chemokine ligand 2, *CCR* chemokine receptor, *CSF* cerebrospinal fluid, *HE* hepatic encephalopathy, *IL* Interleukin, *MAPK* mitogen-activated protein kinase, *TGF β* transforming growth factor β , *TNF α* tumor necrosis factor α , *TNFR1* TNF receptor 1

15.4.2 IL-1 β

IL-1 β is a potent proinflammatory cytokine that is predominantly expressed in activated macrophages (peripherally) and microglia (centrally) (Deng et al. 2011; Lambertsen et al. 2012). In the brain, rather than regulating microglia activation, it is proposed that IL-1 β plays an important role in the downstream consequences of microglial activation on brain function, injury and repair (Lambertsen et al. 2012). Increased cortical IL-1 β expression has been demonstrated in animal models HE due to ALF or cirrhosis (Jiang et al. 2009b; Bemeur et al. 2010b; Rodrigo et al. 2010). Furthermore, mice specifically lacking the IL-1 β receptor had a delayed onset of encephalopathy and an attenuation of brain edema (Bemeur et al. 2010b). Increased expression and secretion of IL-1 β in the brain during HE may then exert deleterious effects on astroglial function. Indeed, treatment of astrocyte cultures with recombinant IL-1 β increased the expression of the astrocyte proteins, GFAP, AQP-4, hemoxygenase 1 and inducible nitric oxide synthase (Chastre et al. 2010) and led to induction of the mitochondrial permeability transition complex (Alvarez et al. 2011).

15.4.3 TGF β

The role of transforming growth factor β (TGF β) in the inflammatory response is largely context dependent. Specifically, TGF β has both anti-inflammatory and pro-inflammatory effects on various immune cells in the body, including microglial activation. Increased levels of TGF β have been demonstrated in the liver and serum in the AOM model of ALF (McMillin et al. 2012a). The authors demonstrated that peripheral TGF β inhibits the expression of the protective factor Gli1 in the cerebral cortex during both acute and chronic liver damage, and speculated that this has implications on microglial activation (McMillin et al. 2013). Systemic treatment

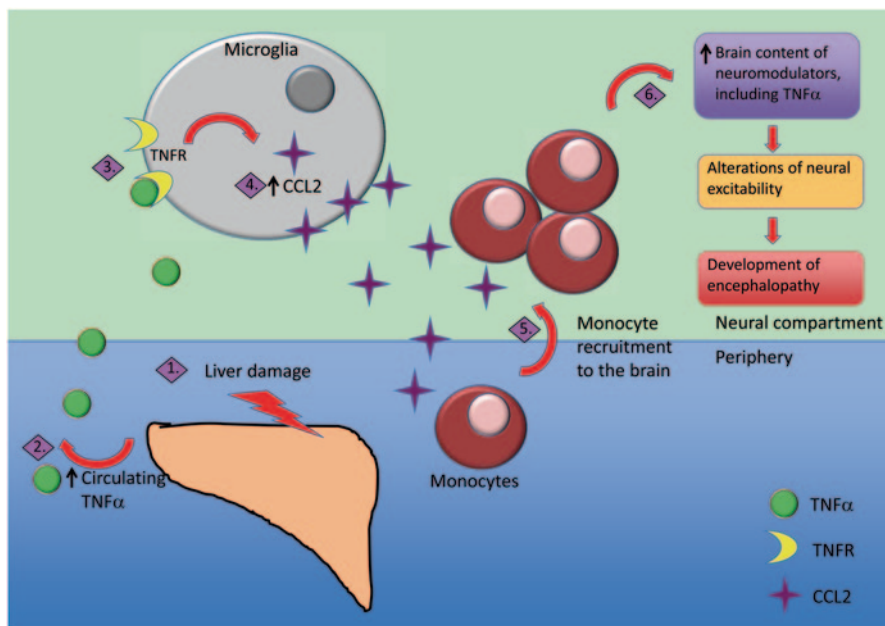


Fig. 15.6 Possible mechanism whereby systemic inflammation leads to microglial activation and monocyte recruitment into the brain as key factors in the pathogenesis of HE in liver failure. Upon damage to the liver (1) there is an increase in circulating TNF α (2). It is proposed that this increase in TNF α is then able to activate TNFR located on microglia (3) to bring about their activation. Activated microglia then produce increased amounts of the chemokine CCL2 (4), which signals for the recruitment of peripheral monocytes to the brain (5). Together, the activated microglia and increased infiltration of monocytes leads to an increased brain content of neuromodulators including TNF α , alterations in neural excitability and ultimately the development of encephalopathy (6)

of mice with a neutralizing anti-TGF β antibody delayed the neurological decline observed in AOM-induced ALF (McMillin et al. 2013), and attenuated the morphological changes in Iba-1 positive microglia (McMillin et al. 2013). However, whether TGF β is acting directly on microglia to regulate the neuroinflammatory response in these models of HE, or whether the changes in microglial activation are an indirect effect of the protective effect of anti-TGF β neutralizing antibodies remains to be established.

15.4.4 CCL2

The chemokine ligand 2 (CCL2 or monocyte chemoattractant protein-1) and its receptors CCR2 and CCR4 have been implicated in a number of neuropathologies ranging from traumatic brain injury to autoimmune disease (Yao et al. 2012). CCL2 can be produced by a number of cell types in the brain, including astrocytes and

microglia (Yao et al. 2012). Neurons have also been shown to release CCL2 after brain ischemia or lipopolysaccharide administration (Kim et al. 2012). The consequences of CCL2 expression are varied and context-dependent. For example, CCL2 has been shown to activate microglia as well as increase the infiltration of circulating macrophages (Yao et al. 2012). Furthermore, while CCL2 expression is upregulated in a number of neuropathies, the consequences are detrimental in some disease states and protective in others (Conductier et al. 2010).

CCL2 immunoreactivity has recently been demonstrated to be increased in the frontal cortex of autopsy samples from cirrhotic patients with HE but not in cirrhotic patients who died with no neurological symptoms (Bradley et al. 2013). As mentioned above, increased CCL2 expression has also been demonstrated in microglia in the cerebral cortex in the animal model of biliary cirrhosis. It is postulated that the release of this chemokine is responsible for triggering the recruitment of circulating monocytes to the affected brain region, which contributes to the subsequent neurological decline (D’Mello et al. 2009) (Fig. 15.6). Similarly, in the AOM model of HE, CCL2 mRNA and protein expression is upregulated in the cerebral cortex (McMillin et al. 2012b). However, in this model CCL2 immunoreactivity appeared to be predominantly neuronal (McMillin et al. 2012b). Pretreatment of mice with specific inhibitors of CCR2 and CCR4 attenuated the increase in microglia activation (demonstrated by morphological changes in Iba-1 positive microglia) and subsequently delayed the neurological decline (McMillin et al. 2012b), supporting the hypothesis that during ALF, neurons produce and secrete the chemoattractant CCL2 that may cause the recruitment, proliferation and activation of microglia. Whether CCL2 also causes a recruitment of circulating monocytes in ALF, in a similar manner to that observed in chronic liver cirrhosis is unknown.

Conclusion

Hepatic encephalopathy and brain edema are serious neurological complications of liver failure that are characterized by neuroglial pathology involving primarily swelling of astrocytes and activation of microglia. In addition to frank pathology, significant alterations of neuroglial function are encountered in liver failure. Such changes include cell volume regulatory deficiencies leading to cytotoxic brain edema and its complications, reductions in expression of astroglial amino acid transporters and the increased expression of the mitochondrial translocator protein. Activation of microglia are revealed using a range of cell-specific markers and are a predominant feature of liver failure independent of the cause of liver damage. Increases in expression of genes coding for pro-inflammatory cytokines (TNF α , IL-1b, IL-6) and chemokines (CCL2) occur in brain as a consequence of microglial activation leading to the currently well-accepted notion that neuroinflammation (inflammation of the brain *per se*) is a major feature of HE in liver failure. Possible mechanisms responsible for activation of microglia in liver failure include ammonia neurotoxicity, brain lactate accumulation and liver-brain pro-inflammatory signalling. A

series of studies in experimental liver failure demonstrate that anti-inflammatory treatments using a range of approaches including proinflammatory cytokine gene deletion, neutralizing anti-cytokine antibodies such as etanercept, CCL2 receptor antagonists and minocycline result in prevention or significant attenuation of the severity of HE and brain edema in liver failure. Translation of these interesting leads into the clinic has the potential to provide novel therapeutic opportunities for the management and treatment of HE and brain edema in liver failure.

Acknowledgements This material is the result of work supported in part with resources and the use of facilities at the Central Texas Veterans Health Care System, Temple, Texas and from The Canadian Institutes of Health Research (CIHR).

References

- Agusti A, Cauli O, Rodrigo R, Llansola M, Hernandez-Rabaza V, Felipe V (2011) p38 MAP kinase is a therapeutic target for hepatic encephalopathy in rats with portacaval shunts. *Gut* 60:1572–1579
- Ahboucha S, Butterworth RF (2005) Role of endogenous benzodiazepine ligands and their GABA-A-associated receptors in hepatic encephalopathy. *Metab Brain Dis* 20:425–437
- Ahboucha S, Butterworth RF (2008) The neurosteroid system: implication in the pathophysiology of hepatic encephalopathy. *Neurochem Int* 52:575–587
- Alvarez VM, Rama Rao KV, Brahmhatt M, Norenberg MD (2011) Interaction between cytokines and ammonia in the mitochondrial permeability transition in cultured astrocytes. *J Neurosci Res* 89:2028–2040
- Belanger M, Desjardins P, Chatauret N, Butterworth RF (2002) Loss of expression of glial fibrillary acidic protein in acute hyperammonemia. *Neurochem Int* 41:155–160
- Belanger M, Desjardins P, Chatauret N, Butterworth RF (2006) Selectively increased expression of the astrocytic/endothelial glucose transporter protein GLUT1 in acute liver failure. *Glia* 53:557–562
- Bemeur C, Chastre A, Desjardins P, Butterworth R (2010a) No changes in expression of tight junction proteins or blood-brain barrier permeability in azoxymethane-induced experimental acute liver failure. *Neurochem Int* 56:205–207
- Bemeur C, Qu H, Desjardins P, Butterworth RF (2010b) IL-1 or TNF receptor gene deletion delays onset of encephalopathy and attenuates brain edema in experimental acute liver failure. *Neurochem Int* 56:213–215
- Bernal W, Donaldson P, Underhill J, Wendon J, Williams R (1998) Tumor necrosis factor genomic polymorphism and outcome of acetaminophen (paracetamol)-induced acute liver failure. *J Hepatol* 29:53–59
- Bradley M, McMillin M, Galindo C, Frampton G, Pae H, Quinn M, DeMorrow S (2013) CCR2 and CCR4 activation via CCL2/MCP-1 contributes to neurological decline and microglia activation associated with hepatic encephalopathy pathogenesis. *Gastroenterology* 144:S1026–S1027
- Brusilow SW, Koehler RC, Traystman RJ, Cooper AJ (2010) Astrocyte glutamine synthetase: importance in hyperammonemic syndromes and potential target for therapy. *Neurotherapeutics* 7:452–470
- Butterworth R (2007) Neuronal cell death in hepatic encephalopathy. *Metab Brain Dis* 22:309–320
- Butterworth RF (2011) Hepatic encephalopathy: a central neuroinflammatory disorder? *Hepatology* 53:1372–1376

- Butterworth RF (2013a) The liver-brain axis in liver failure: neuroinflammation and encephalopathy. *Nat Rev Gastroenterol Hepatol* 10:522–528
- Butterworth RF (2013b) Parkinsonism in cirrhosis: pathogenesis and current therapeutic options. *Metab Brain Dis* 28:261–267
- Butterworth RF, Vaquero J (2009) Hepatic encephalopathy. In: Arias I, Alter H, Boyer J, Cohen D, Fausto N, Shafritz D et al (eds) *The liver; biology and pathobiology*. Wiley, Chichester, pp 600–617
- Butterworth RF, Norenberg MD, Felipe V, Ferenci P, Albrecht J, Blei AT (2009) Experimental models of hepatic encephalopathy: ISHEN guidelines. *Liver Int* 29:783–788
- Cagnin A, Taylor-Robinson SD, Forton DM, Banati RB (2006) In vivo imaging of cerebral “peripheral benzodiazepine binding sites” in patients with hepatic encephalopathy. *Gut* 55:547–553
- Chastre A, Jiang W, Desjardins P, Butterworth RF (2010) Ammonia and proinflammatory cytokines modify expression of genes coding for astrocytic proteins implicated in brain edema in acute liver failure. *Metab Brain Dis* 25:17–21
- Chastre A, Belanger M, Beauchesne E, Nguyen BN, Desjardins P, Butterworth RF (2012) Inflammatory cascades driven by tumor necrosis factor-alpha play a major role in the progression of acute liver failure and its neurological complications. *PLoS ONE* 7:e49670
- Chatauret N, Zwingmann C, Rose C, Leibfritz D, Butterworth RF (2003) Effects of hypothermia on brain glucose metabolism in acute liver failure: a H/C-nuclear magnetic resonance study. *Gastroenterology* 125:815–824
- Clemmesen JO, Larsen FS, Kondrup J, Hansen BA, Ott P (1999) Cerebral herniation in patients with acute liver failure is correlated with arterial ammonia concentration. *Hepatology* 29:648–653
- Conductier G, Blondeau N, Guyon A, Nahon JL, Rovere C (2010) The role of monocyte chemoattractant protein MCP1/CCL2 in neuroinflammatory diseases. *J Neuroimmunol* 224:93–100
- Cubelos B, Gonzalez-Gonzalez IM, Gimenez C, Zafra F (2005) Amino acid transporter SNAT5 localizes to glial cells in the rat brain. *Glia* 49:230–244
- D’Mello C, Le T, Swain MG (2009) Cerebral microglia recruit monocytes into the brain in response to tumor necrosis factor-alpha signaling during peripheral organ inflammation. *J Neurosci* 29:2089–2102
- Deng YY, Lu J, Ling EA, Kaur C (2011) Role of microglia in the process of inflammation in the hypoxic developing brain. *Front Biosci (Schol Ed)* 3:884–900
- Desjardins P, Du T, Jiang W, Peng L, Butterworth RF (2012) Pathogenesis of hepatic encephalopathy and brain edema in acute liver failure: role of glutamine redefined. *Neurochem Int* 60:690–696
- Eefsen M, Jelnes P, Schmidt LE, Vainer B, Bisgaard HC, Larsen FS (2010) Brain expression of the water channels aquaporin-1 and -4 in mice with acute liver injury, hyperammonemia and brain edema. *Metab Brain Dis* 25:315–323
- Felipo V, Butterworth RF (2002) Neurobiology of ammonia. *Prog Neurobiol* 67(4):259–279. Review
- Felipo V, Minana MD, Grisolia S (1988) Long-term ingestion of ammonium increases acetylglutamate and urea levels without affecting the amount of carbamoyl-phosphate synthase. *Eur J Biochem/FEBS* 176:567–571
- Gorg B, Bidmon HJ, Haussinger D (2013) Gene expression profiling in the cerebral cortex of patients with cirrhosis with and without hepatic encephalopathy. *Hepatology* 57:2436–2447
- Jiang W, Qu H, Desjardins P, Chatauret N, Belanger M, Butterworth R (2006) Unequivocal evidence for cytokine accumulation in brain in experimental acute liver failure. *Hepatology* 44:336A
- Jiang W, Desjardins P, Butterworth RF (2009a) Cerebral inflammation contributes to encephalopathy and brain edema in acute liver failure: protective effect of minocycline. *J Neurochem* 109:485–493

- Jiang W, Desjardins P, Butterworth RF (2009b) Direct evidence for central proinflammatory mechanisms in rats with experimental acute liver failure: protective effect of hypothermia. *J Cereb Blood Flow Metab* 29:944–952
- Kato M, Hughes RD, Keays RT, Williams R (1992) Electron microscopic study of brain capillaries in cerebral edema from fulminant hepatic failure. *Hepatology* 15:1060–1066
- Kerfoot SM, D’Mello C, Nguyen H, Ajuebor MN, Kubes P, Le T, Swain MG (2006) TNF- α -secreting monocytes are recruited into the brain of cholestatic mice. *Hepatology* 43:154–162
- Kim SE, Lee EO, Yang JH, Kang JH, Suh YH, Chong YH (2012) 15-deoxy-Delta(1)(2), (1)(4)-prostaglandin J(2) inhibits human immunodeficiency virus-1 tat-induced monocyte chemoattractant protein-1/CCL2 production by blocking the extracellular signal-regulated kinase-1/2 signaling pathway independently of peroxisome proliferator-activated receptor- γ and heme oxygenase-1 in rat hippocampal slices. *J Neurosci Res* 90:1732–1742
- Knecht K, Michalak A, Rose C, Rothstein JD, Butterworth RF (1997) Decreased glutamate transporter (GLT-1) expression in frontal cortex of rats with acute liver failure. *Neurosci Lett* 229:201–203
- Kril JJ, Flowers D, Butterworth RF (1997) Distinctive pattern of Bergmann glial pathology in human hepatic encephalopathy. *Mol chem neuropathol* 31:279–287
- Lachmann V, Gorg B, Bidmon HJ, Keitel V, Haussinger D (2013) Precipitants of hepatic encephalopathy induce rapid astrocyte swelling in an oxidative stress dependent manner. *Arch Biochem Biophys* 536:143–151
- Lambertsen KL, Biber K, Finsen B (2012) Inflammatory cytokines in experimental and human stroke. *J Cereb Blood Flow Metab* 32:1677–1698
- Laubenberger J, Haussinger D, Bayer S, Gufler H, Hennig J, Langer M (1997) Proton magnetic resonance spectroscopy of the brain in symptomatic and asymptomatic patients with liver cirrhosis. *Gastroenterology* 112:1610–1616
- McMillin M, Galindo C, Frampton G, Pae H, Quinn M, DeMorrow S (2012a) Gli1 is activated via the non-classical TGF β /SMAD3 pathway rather than the sonic hedgehog pathway in a murine model of hepatic encephalopathy. *FASEB J* 26:1110–1112
- McMillin M, Galindo C, Frampton G, Pae H, Quinn M, Bradley M, Jacobs A, DeMorrow S (2012b) Increased neuronal chemokine CCL2/MCP-1 expression is associated with hepatic encephalopathy and contributes to neurological decline. *Hepatology* 56:958A
- McMillin M, Galindo C, Frampton G, Pae H, Quinn M, DeMorrow S (2013) CCL3 is upregulated in the frontal cortex during hepatic encephalopathy and can be inhibited by the suppression of circulating TGF β 1. *J Neurochem* 125(Suppl S1):136
- Neary JT, Whitemore SR, Zhu Q, Norenberg MD (1994) Destabilization of glial fibrillary acidic protein mRNA in astrocytes by ammonia and protection by extracellular ATP. *J Neurochem* 63:2021–2027
- Nguyen JH (2010) Subtle BBB alterations in brain edema associated with acute liver failure. *Neurochem Int* 56:203–204; author reply 205–207
- Norenberg MD (1987) The role of astrocytes in hepatic encephalopathy. *Neurochem Pathol* 6:13–33
- Obara-Michlewska M, Pannicke T, Karl A, Bringmann A, Reichenbach A, Szeliga M, Hilgier W, Wrzosek A, Szweczyk A, Albrecht J (2011) Down-regulation of Kir4.1 in the cerebral cortex of rats with liver failure and in cultured astrocytes treated with glutamine: implications for astrocytic dysfunction in hepatic encephalopathy. *J Neurosci Res* 89:2018–2027
- Odeh M (2007) Pathogenesis of hepatic encephalopathy: the tumour necrosis factor- α theory. *Eur J Clin Invest* 37:291–304
- Rama Rao KV, Chen M, Simard JM, Norenberg MD (2003) Increased aquaporin-4 expression in ammonia-treated cultured astrocytes. *Neuroreport* 14:2379–2382
- Rangroo Thrane V, Thrane AS, Chang J, Alleluia V, Nagelhus EA, Nedergaard M (2012) Real-time analysis of microglial activation and motility in hepatic and hyperammonemic encephalopathy. *Neuroscience* 220:247–255

- Rodrigo R, Cauli O, Gomez-Pinedo U, Agusti A, Hernandez-Rabaza V, Garcia-Verdugo JM, Felipe V (2010) Hyperammonemia induces neuroinflammation that contributes to cognitive impairment in rats with hepatic encephalopathy. *Gastroenterology* 139:675–684
- Rubio-Perez JM, Morillas-Ruiz JM (2012) A review: inflammatory process in Alzheimer's disease, role of cytokines. *ScientificWorldJournal* 2012:756357
- Shawcross DL, Davies NA, Williams R, Jalan R (2004) Systemic inflammatory response exacerbates the neuropsychological effects of induced hyperammonemia in cirrhosis. *J Hepatol* 40:247–254
- Sobel RA, DeArmond SJ, Forno LS, Eng LF (1981) Glial fibrillary acidic protein in hepatic encephalopathy. An immunohistochemical study. *J Neuropathol Exp Neurol* 40:625–632
- Suarez I, Bodega G, Fernandez B (2000) Modulation of glutamate transporters (GLAST, GLT-1 and EAAC1) in the rat cerebellum following portocaval anastomosis. *Brain Res* 859:293–302
- Vaquero J, Butterworth RF (2006) The brain glutamate system in liver failure. *J Neurochem* 98:661–669
- Villano G, Lunardi F, Turato C, Schiff S, Tono N, Campagna F, Gatta A, Amodio P, Calabrese F, Pontissa P (2012) Increased Th1 immune response in SERPINB3 transgenic mice during acute liver failure. *Exp Biol Med* 237:1474–1482
- Wright G, Shawcross D, Olde Damink SW, Jalan R (2007) Brain cytokine flux in acute liver failure and its relationship with intracranial hypertension. *Metab Brain Dis* 22:375–388
- Wright G, Soper R, Brooks HF, Stadlbauer V, Vairappan B, Davies NA, Andreola F, Hodges S, Moss RF, Davies DC, Jalan R (2010) Role of aquaporin-4 in the development of brain oedema in liver failure. *J Hepatol* 53:91–97
- Yao HL, Gao FH, Li ZZ, Wu HX, Xu MD, Zhang Z, Dai QY (2012) Monocyte chemoattractant protein-1 mediates angiotensin II-induced vascular smooth muscle cell proliferation via SAPK/JNK and ERK1/2. *Mol Cell Biochem* 366:355–362
- Zemtsova I, Gorg B, Keitel V, Bidmon HJ, Schror K, Haussinger D (2011) Microglia activation in hepatic encephalopathy in rats and humans. *Hepatology* 54:204–215
- Zwingmann C, Desjardins P, Hazell A, Chatauret N, Michalak A, Butterworth RF (2002) Reduced expression of astrocytic glycine transporter (Glyt-1) in acute liver failure. *Metab Brain Dis* 17:263–273