Cutaneous Lymphoid Infiltrates in Patients Receiving Biologic Modifiers

15

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Introduction

Over the past few decades, the introduction of new biologic agents, such as tumor necrosis factor-alpha inhibitors, has resulted in potent disease-modifying effects in a variety of immunemediated diseases. There have also been major advancements in cancer chemotherapy with the recent addition of granulocyte colony-stimulating factor, interferon, kinase inhibitors, and CD20 and CD52 inhibitors for the treatment of hematologic malignancies as well as solid tumors. However, a variety of toxicities, including adverse cutaneous reactions, are seen in association with these agents. Awareness of these cutaneous toxicities and recognition of corresponding histologic features is of diagnostic and therapeutic importance.

TNF- α Inhibitors

Tumor necrosis factor- α (TNF- α) is a powerful proinflammatory cytokine that is a central player to the activation of inflammation (Parameswaran and Patial 2010). Lymphocytes, macrophages,

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endothelial cells, Langerhans cells, as well as keratinocytes, melanocytes, and many others are capable of producing TNF- α (Deng et al. 2006). It is rapidly released in response to trauma, infection, or exposure to toxins wherein it functions as the "master regulator" of proinflammatory cytokine production (Parameswaran and Patial 2010). Despite its role in the normal host defense response, TNF- α is also central to the pathogenesis of several chronic inflammatory conditions and autoimmune disorders, including rheumatoid arthritis, psoriasis, inflammatory bowel disease, and ankylosing spondylitis (Kollias et al. 1999; Moustou et al. 2009).

Biologic antibody inhibitors of TNF- α , including infliximab (Remicade, Centocor, USA), etanercept (Enbrel, Immunex, USA), adalimumab (Humira), and certolizumab pegol (Cimzia), have been used with success in the treatment of these disorders. Nevertheless, their use has been associated with class-wide adverse reactions events, including opportunistic infections (Kim and Solomon 2010), reactivation of latent tuberculosis (Keane et al. 2001), lupus-like syndrome (Williams and Cohen 2011), demyelinating disease (Omair et al. 2012), exacerbation of heart failure (Colombel et al. 2004), and lymphoma (Adams et al. 2004). Cutaneous adverse events of antitumor necrosis factor therapy include infusion and injection site reactions, psoriasiform eruptions, granulomatous dermatitis, lupus-like disorders, vasculitis, cutaneous infections, and cutaneous neoplasms (Moustou et al. 2009; Bovenschen et al. 2006; Esser et al. 2004; Deng et al. 2006) (Table 15.1).

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Table 15.1 Cutaneous reactions associated with TNF- α (Moustou et al. 2009; Deng et al. 2006; Voulgari et al. 2008; Bovenschen et al. 2006; Esser et al. 2004)

Infusion and injection site reactions	
Psoriasis and psoriasiform dermatitis	
Eczematous dermatitis	
Lupus erythematosus-like eruption	
Leukocytoclastic vasculitis	
Lichenoid dermatitis	
Granuloma annulare/interstitial granulomatous dermatitis	
Cutaneous infections	
Cutaneous lymphomas	
Multiple lentigines or eruptive melanocytic nevi	
Multiple keratoacanthomas	

Granuloma Annulare/Interstitial Granulomatous Dermatitis

TNF- α inhibitor-associated granulomatous reactions, including interstitial granulomatous dermatitis and granuloma annulare (GA), are rare cutaneous lymphoid reactions that have only recently been reported in the literature (Deng et al. 2006; Voulgari et al. 2008).

Clinical Presentation

Granulomatous adverse cutaneous reactions may occur anywhere from 1 month to over 1 year after initiating anti-TNF- α therapy and may occur after treatment with any of the aforementioned agents (Deng et al. 2006; Voulgari et al. 2008). Cases of interstitial granulomatous dermatitis presented with rapid-onset, asymptomatic to mildly pruritic, 1-4 cm macules or indurated papules or plaques, many of which are annular in morphology, that occur on the trunk, shoulders, and upper extremities (Deng et al. 2006). In the first reported series, this reaction led to the discontinuation of treatment in four of five patients (Deng et al. 2006). Of these four patients, the eruption cleared or improved within 2 months of discontinuation (Deng et al. 2006). One patient had to be maintained on therapy due to the severity of underlying rheumatoid arthritis, and the lesions persisted at 3-month follow-up (Deng et al. 2006).

TNF- α inhibitor-induced GA presented with erythematous skin eruptions covering the fingers, hands, and forearms (Voulgari et al. 2008). In the largest report in the literature, nine such cases were reported among 197 patients (incidence, 4.5 %) undergoing anti-TNF therapy for the treatment of rheumatoid arthritis (Voulgari et al. 2008). Biologic therapy was maintained on seven of nine of these cases (Voulgari et al. 2008). Lesions were treated with topical corticosteroids with resolution of eruption after 3–4 weeks (Voulgari et al. 2008).

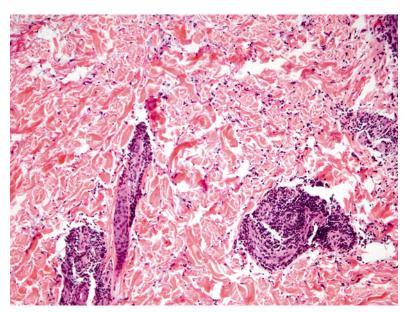
Histology Features

Histopathologic examination of TNF- α inhibitorassociated GA is characterized by palisading granulomas surrounding an area of necrobiosis and mucin deposition (Voulgari et al. 2008; Stewart et al. 2011) (Fig. 15.1). The histology of interstitial granulomatous dermatitis is characterized by a diffuse interstitial infiltrate of histiocytes and lymphocytes that palisade around partially degenerated or "piecemeal"-fragmented collagen, resulting in granulomatous foci in the mid and deep dermis (Deng et al. 2006). Eosinophils, neutrophils, and multinucleated giant cells may be present in scant numbers (Deng et al. 2006).

Pathogenesis

The precise pathomechanism underlying TNF- α inhibitor-induced GA is not known, but evidence suggests that anti-TNF therapy may induce autoreactive T cells (Sfikakis and Kollias 2003). Paradoxically, infliximab has been reported to have led to the rapid improvement of disseminated GA (Hertl et al. 2005). Chu et al. (1994) postulated that the granulomatous infiltrate in interstitial granulomatous dermatitis is a secondary reaction to dermal collagen damage induced initially by the deposition of immune complexes in dermal vessels and subsequent ischemia, followed by complement and neutrophil activation. It is known that interstitial granulomatous dermatitis can present in association with rheumatoid arthritis, lupus erythematosus, and other systemic disorders (Long et al. 1996) and may also be induced by other medications, including calcium channel blockers, lipid-lowering agents, and others (Magro et al. 1998).

Fig. 15.1 A 50-year-old female on Humira for Crohn's disease presented with a pruritic and indurated plaques and papules on her left leg. A skin biopsy shows an interstitial and perivascular infiltrate of histiocytes and lymphocytes consistent with an interstitial granulomatous dermatitis



Enbrel (Etanercept): Urticaria and Cellulitis

Urticaria and cellulitis are the most commonly reported cutaneous toxicity associated with Enbrel/etanercept, although it is not frequently biopsied.

Clinical Presentation

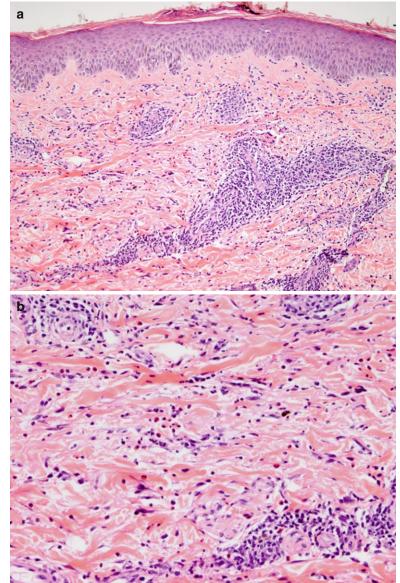
Etanercept-induced urticaria or urticaria-like reactions typically present 3-4 months after receiving twice-weekly subcutaneous etanercept injections (Skytta et al. 2000; Borrás-Blasco et al. 2009). After the inciting injection, an ervthematous, macular, papular, or plaque-like urticarial lesion with or without central clearing develops initially on the extensor surfaces of the elbows and, subsequently, on the extensor surfaces of the knees (Borrás-Blasco et al. 2009). Lesions may also appear elsewhere on the body, such as the buttocks, trunk, proximal limbs, or ears (Skytta et al. 2000; Borrás-Blasco et al. 2009). Laboratory abnormalities, including eosinophilia, are typically absent (Skytta et al. 2000). These lesions have been reported to resolve after cessation of etanercept treatment (Skytta et al. 2000) or a course of steroid (Borrás-Blasco et al. 2009).

Reactions at the injection site can be urticarial reactions (Batycka-Baran et al. 2012) or eosinophilic cellulitis-like reaction (Winfield et al. 2006). Typically within 24 h of receiving the first dose, the patient developed evanescent, pruritic, slightly indurated, and erythematous plaques surrounding the injection site. These lesions resolved upon treatment with antihistamines and prednisone (Winfield et al. 2006) or topical steroids (Batycka-Baran et al. 2012).

Histologic Features

Histopathologic examination of the etanercept injection site urticarial lesions reveals a perivascular inflammatory infiltrate composed predominantly of neutrophils without eosinophils (Batycka-Baran et al. 2012). Immunophenotypic study may reveal either CD8+ (Zeltser et al. 2001) or CD4+ (González-López et al. 2007) predominance of T lymphocytes within the dermal infiltrate. Histology of the eosinophilic cellulitis-like injection site reaction to etanercept demonstrates an unremarkable epidermis and underling perivascular and interstitial lymphocytic infiltrates with numerous eosinophils with associated edema in the dermis (Winfield et al. 2006) (Fig. 15.2). Other notable features are multiple foci of hypereosinophilic collagen surrounded by eosinophils in the dermis as well as flame figures (Winfield et al. 2006).

Fig. 15.2 A 47-year-old male with a history of psoriasis and on Enbrel treatment developed a lesion at the injection site on his right upper arm. A skin biopsy revealed an interstitial and perivascular infiltrate (**a**) of lymphocytes, histiocytes, and many eosinophils (**b**) in the dermis consistent with a hypersensitivity reaction



Pathogenesis

Batycka-Baran et al. (2012) proposed that the kinetics of the reaction, as revealed by the time course between the injection and onset of cutaneous eruption, may provide insight into the mechanism. It is thought that injection site reactions occurring rapidly after the first injection are due to an irritative mechanism and may not require discontinuation of treatment (Batycka-Baran et al. 2012). Urticarial lesions that occur within hours of the second or later injection with etanercept may reflect an immediate type I, IgE-mediated hypersensitivity reaction that would require stopping the medication and close monitoring for possible anaphylaxis (Batycka-Baran et al. 2012). Finally, reactions that occur hours after the injection and persist for several days may reflect an Arthus-like type III hypersensitivity reaction, mediated in part by the recruitment of neutrophilic granulocytes (Batycka-Baran et al. 2012).

G-CSF: Neutrophilic Dermatoses

Granulocyte colony-stimulating factor (G-CSF) is a neutrophil-specific growth factor that promotes the survival, proliferation, and maturation of neutrophil progenitors (Kaushansky 2006). This cytokine also enhances their phagocytic capacity, superoxide anion production, and bacterial killing mechanisms (Spiekermann et al. 1997). In contrast, granulocyte-macrophage colony-stimulating factor (GM-CSF) stimulates the production and activation of eosinophils, basophils, monocytes, and dendritic cells in addition to neutrophils.

G-CSF is used clinically to treat neutropenia associated with chemotherapy, myelodysplasia, or aplastic anemia as well as to reduce infections in patients with congenital, idiopathic, or cyclic neutropenia (Prendiville et al. 2001). Mild adverse effects include transient bone pain, edema, arthralgias, and myalgias as well as elevations in serum lactate dehydrogenase (LDH), alanine aminotransferase (ALT), aspartate aminotransferase (AST), and alkaline phosphatase (AP) levels (Paydas et al. 1993). It has also been found to increase the risk of treatment-related acute myelogenous leukemia, particularly in those with congenital neutropenia, wherein approximately 10 % will develop a malignant myeloid disorder (Kaushansky 2006; Dong et al. 1995).

Adverse cutaneous effects of G-CSF (Table 15.2) include diffuse eruption or reaction at injection site (Alvarez-Ruiz et al. 2004; Valks et al. 1998). Skin biopsies show an increase in the number of dermal macrophages, often enlarged

Table 15.2 Cutaneous reactions associated with G-CSF (Alvarez-Ruiz et al. 2004; Valks et al. 1998; Prendiville et al. 2001; Johnson and Grimwood 1994; Ross et al. 1991; Paydas et al. 1993; Fukutoku et al. 1994; Jain 1994; Ostlere et al. 1992)

Generalized or diffuse eruption	
Local cutaneous reaction	
Neutrophilic dermatoses	
Sweet syndrome	
Pyoderma gangrenosum	
Vasculitis	
Folliculitis	

in size (Alvarez-Ruiz et al. 2004; Valks et al. 1998). Neutrophilic dermatoses (Prendiville et al. 2001; Johnson and Grimwood 1994), including pyoderma gangrenosum (Ross et al. 1991) and Sweet syndrome (Paydas et al. 1993; Fukutoku et al. 1994), rare cases of vasculitis (Jain 1994; Couderc et al. 1995), and folliculitis (Ostlere et al. 1992) have been reported in association with G-CSF.

Sweet Syndrome Associated with G-CSF

Sweet syndrome, first documented by RD Sweet in (1964), is characterized by a pentad of findings, including fever, neutrophilia, multiple erythematous and painful cutaneous papules or nodules, dense dermal infiltrate of neutrophils, and rapid response to steroid therapy (Paydas et al. 1993). Cases of Sweet syndrome are categorized based on etiology into classical/idiopathic, malignancyassociated, and drug-induced (Cohen 2007). Classical Sweet syndrome is associated with upper respiratory tract and gastrointestinal infections, inflammatory bowel disease, and pregnancy (Cohen 2007). Malignancy-associated Sweet syndrome is most commonly caused by hematologic malignancies (especially acute myelogenous leukemia), but solid tumors of the genitourinary and gastrointestinal tracts and breast have also been associated. A wide variety of medications have been associated with drug-induced Sweet syndrome, including antibiotics such as trimethoprim-sulfamethoxazole, all-trans retinoic acid, and antineoplastic agents such as Imatinib (Cohen 2007). However, G-CSF is the most frequently and widely implicated medications in causing Sweet syndrome (White et al. 2006).

Walker and Cohen (1996) put forth the following diagnostic criteria for drug-induced Sweet syndrome: (1) abrupt onset of painful erythematous plaques or nodules, (2) histopathologic evidence of a dense neutrophilic infiltrate without evidence of leukocytoclastic vasculitis, (3) fever >38 °C, (4) temporal relationship between drug ingestion and clinical presentation or temporally related recurrence after reintroduction of drug, and (5) temporally related resolution of lesions after drug withdrawal or treatment with systemic corticosteroids. Drug-induced Sweet syndrome typically affects patients between 29 and 68 years of age (mean, 45 years) and presents anywhere from days to weeks (mean, 7.5 days) after initiation of drug therapy. It is characterized by the development of fever, arthralgias, and diffuse, erythematous, painful papules, plaques, or nodules found most commonly in the upper extremities (71 %) but may also affect the chest (50 %), head and neck (43 %), lower extremities (36 %), or oral mucous membranes (7 %) (Walker and Cohen 1996). Lesions on the face may follow a periorbital or periauricular distribution (Bidyasar et al. 2008). Cutaneous lesions in a photodistributed pattern (Walker and Cohen 1996) as well as lesions that develop clinical features of necrosis (Fukutoku et al. 1994) have also been described. Ocular manifestations, characterized by "bilateral conjunctivitis" or "conjunctival lesions," have also been documented in case reports (Bidyasar et al. 2008; Paydas et al. 1993).

Histopathologic examination of these cutaneous lesions reveals superficial edema and an underlying dense infiltrate of neutrophils in the dermis (Walker and Cohen 1996) (Fig. 15.3). Nuclear features of apoptosis such as karyorrhexis and an absence of vasculitis and pathogenic microbes are typical features (Bidyasar et al. 2008; Walker and Cohen 1996). The histologic differential diagnosis for drug-induced Sweet syndrome includes Sweet syndrome proper, abscess/cellulitis, granuloma faciale, leukemia cutis, lobular neutrophilic panniculitis, rheumatoid neutrophilic dermatitis, and pyoderma gangrenosum (Cohen et al. 2007).

Laboratory abnormalities include neutrophilia, although this is much less common with druginduced Sweet syndrome, especially that associated with G-CSF, than it is with Sweet syndrome proper (Walker and Cohen 1996). The cutaneous lesions in G-CSF associated Sweet syndrome may appear with the rise or normalization of neutrophil counts (Walker and Cohen 1996) and may disappear when they fall (van Kamp et al. 1994). Elevations in the erythrocyte sedimentation rate, anemia, and thrombocytosis have also been documented (Walker and Cohen 1996).

The mainstay of treatment for all forms of drug-induced Sweet syndrome is removal of the offending agent, after which the lesions typically resolve within 1–2 weeks (Walker and Cohen 1996). Clinical improvements in both cutaneous

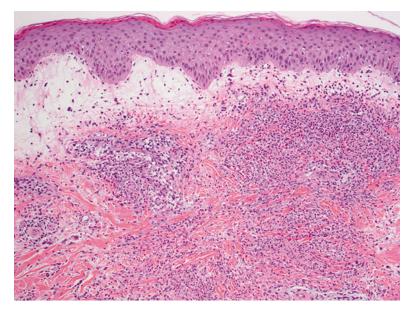


Fig. 15.3 A 51-year-old female on granulocyte colony-stimulating factor presented with erythematous plaques on her left upper arm. A skin biopsy showed marked papillary dermal edema and underlying band-like infiltrate of neutrophils consistent with Sweet syndrome

lesions and the overall condition have been achieved with low-dose systemic steroids alone (Bidyasar et al. 2008) as well as in combination with colchicine (van Kamp et al. 1994). Remission of the myeloid disorder, such as acute myeloid leukemia or myelodysplastic syndrome, also results in resolution of Sweet syndrome.

The precise etiology and pathogenic mechanism underlying all forms of Sweet syndrome is currently unknown. Elevated levels of G-CSF and interleukin-6 have been documented in a G-CSF-naïve patient with myelodysplastic syndrome-associated Sweet syndrome (Reuss-Borst et al. 1993). G-CSF, a cytokine, likely plays a direct pathogenic role, given that serum levels appear to directly correlate with dermatosis activity (Kawakami et al. 2004).

Pyoderma Gangrenosum Associated with G-CSF

The first case of biopsy-proven neutrophilic dermatosis without vasculitis was reported by Ross et al. (1991) in a 50-year-old woman who underwent G-CSF therapy for neutropenia related to chemotherapy for metastatic small cell lung cancer. Since this initial report, few additional cases of pyoderma gangrenosum associated with G-CSF and pegylated G-CSF therapy have been reported in the literature (Lewerin et al. 1997; Miall et al. 2006). G-CSFassociated pyoderma gangrenosum typically presents days to weeks after starting G-CSF therapy (Lewerin et al. 1997). It is characterized by the simultaneous onset of fever and erythematous plaques that can appear anywhere on the body, such as the face, trunk, upper, and lower extremities (Miall et al. 2006). The lesions rapidly progresses in size, swiftly develop blue or black edematous central regions of necrosis, and may appear clinically as a hemorrhagic blister, earning the name "bullous pyoderma gangrenosum" (Lewerin et al. 1997). There does not appear to be a correlation between the dosage or duration of therapy and the severity of this adverse cutaneous reaction (Ross et al. 1991).

Histopathologic examination reveals typical features of pyoderma gangrenosum characterized by a dense neutrophilic dermal infiltrate (Lewerin et al. 1997). Features of vasculitis are typically absent but have been described in association with this lesion (Miall et al. 2005; Lewerin et al. 1997). Studies for acid-fast bacilli, bacteria, and fungi are typically negative (Ross et al. 1991).

It appears that this adverse cutaneous reaction may not resolve with simple withdrawal of G-CSF (Lewerin et al. 1997). Treatment success has been reported with cyclosporin A (Lewerin et al. 1997). Others have had success with high-dose steroids in combination with dapsone (an anti-neutrophil agent) and topical tacrolimus (Miall et al. 2005). Interestingly, there was also a recent report of a case of Crohn's-associated pyoderma gangrenosum that was successfully treated with perilesional granulocyte-macrophage colony-stimulating factor (GM-CSF) (Shpiro et al. 1998).

Classic pyoderma gangrenosum is most commonly associated with inflammatory bowel disease, rheumatologic disorders, or malignancy. The pathogenesis of this condition is poorly understood but thought to be related to loss of altered neutrophil trafficking and chemotaxis, genetic variations, and dysregulation of the innate immune system (Ahronowitz et al. 2012).

Interferon: Lymphocutaneous Injection Site Reactions

Interferon is a complex family of host-derived glycoproteins produced and secreted by eukaryotic cells in response to viruses, antigens, and mitogens (Baron et al. 1991). During their study of cellular viral infections, Isaacs and Lindemann (1957) identified a protein produced by cells infected by the virus that interfered with the virus' ability to infect other cells. It is now known that interferons, of which there are three main groups (IFN- α , IFN- β , IFN- γ), have intrinsic antiviral, antimicrobial, immunomodulatory, and antiproliferative properties (Baron et al. 1991). Thus, today they are used to treat a variety of malignancies, viral infections, and inflammatory conditions, including hairy cell leukemia (IFN- α 2b), chronic hepatitis C infection (IFN- α 2b),

njection site reaction	
Local necrotizing	
Granulomatous and suppurative	
Lupus erythematosus-like	
Embolia cutis medicamentosa	
Local alopecia	
Eczematous drug eruption	
Sarcoidosis	
Leukocytoclastic vasculitis	
Alopecia	
Vitiligo	
Fixed drug eruption	
Lupus erythematosus	

 Table 15.3
 Cutaneous reactions associated with interferon

multiple sclerosis (IFN- β 1a), and chronic granulomatous disease (IFN- γ 1b).

Despite their therapeutic yield, the doses required are frequently associated with significant side effects, including flu-like symptoms, weight loss, depression, as well as neurologic toxicity (Krainick et al. 1998). Moreover, interferon is believed to precipitate autoimmune disorders such as type I diabetes, thyroid disease, and systemic lupus erythematosus and may also worsen certain granulomatous diseases (Selmi et al. 2006).

Psoriasis was one of the early cutaneous reactions (Table 15.3) developed in association with interferon treatment (Quesada and Gutterman 1986), and there have been multiple subsequent reports in the literature (Ketikoglou et al. 2005). Eczematous drug eruptions associated with interferon, which often present as ill-defined clusters of coalescing, erythematous, blanchable, pruritic papules often on the extremities and trunk, have also been described (Dereure et al. 2002; Moore et al. 2004). Sarcoidosis has been described in patients receiving interferon treatment for hepatitis C as well as in a patient receiving adjuvant interferon-alpha immunotherapy for melanoma (Fantini et al. 2009; Alonso-Perez et al. 2006).

Alopecia has been reported in up to 19 % of patients treated with combination interferon and ribavirin, including alopecia areata (Agesta et al. 2002) and alopecia universalis (Demirturk et al. 2006). Vitiligo has been reported in a number of

patients who were treated for hepatitis C with interferon (Tomasiewicz et al. 2006). Cases of cutaneous necrosis (Dalmau et al. 2005), fixed drug eruption (Tai and Tam 2005), lichenoid eruption (Pinto et al. 2003), lupus erythematosus (Fukuyama et al. 2000), indurated erythema (Detmar et al. 1989), leukocytoclastic vasculitis (Christian et al. 1997), as well as pyoderma gangrenosum (Montoto et al. 1998) have been reported in association with interferon.

Delivered through the intramuscular or subcutaneous route, interferon has also been associated with local injection site reactions in just under 5 % of patients, including local necrotizing (Krainick et al. 1998), granulomatous and suppurative (Sanders et al. 2002), lupus erythematosus-like (Arrue et al. 2007), embolia cutis medicamentosa (Koontz and Alshekhlee 2007), and local alopecia (Lang et al. 1999) lesions. The pathogenesis of these reactions remains uncertain. Multiple mechanisms have been proposed, and these, in addition to the basic clinical presentation and histopathology, are summarized by each pattern of reaction below. Whether there is a common underlying mode of pathogenesis underlying these reactions is yet to be determined, but interferon's role in modulating multiple inflammatory pathways is likely central to the explanation.

Local Cutaneous Necrotizing Lesions

Krainick et al. (1998) described seven cases of local, painful cutaneous necrotizing lesions that arose in patients undergoing intramuscular or subcutaneous interferon therapy for Philadelphia chromosome-positive chronic myelogenous leukemia (CML). They occurred without correlation to specific stage of disease, course of illness, or interferon type. The lesions arose anywhere from 3 to 108 months after initiating interferon therapy. The lesions were described as necrotizing ulcers or abscess with surrounding erythematous, painful induration. Low-grade fever was commonly present. Microbiologic culture of the lesion was most commonly sterile and ultrasound findings were consistent with cellulitis.

Histopathologic examination revealed ulceration with underlying inflamed granulation tissue with scattered eosinophils in the dermis. One case was managed with reconstructive surgery, another with local surgical debridement and topical antibiotics, while the others generally resolved with more conservative measures even while maintained on therapy. The pathogenesis of these ulcers remains uncertain, but it is believed that local interferon-mediated keratinocyte toxicity, immune complex formation, and/ or ischemia through multiple mechanisms, including interferon-mediated inhibition of angiogenesis, endothelial cell disruption, vasospasm, and intravascular thrombosis are involved (Sanders et al. 2002).

Granulomatous and Suppurative Dermatitis

Sanders et al. (2002) describe two cases wherein dusky, violaceous, multilobulated papules or indurated plaques and nodules with central ulceration developed at interferon injection sites in one patient undergoing treatment for non-remitting metastatic renal cell carcinoma (RCC) and another for metastatic melanoma. Histopathologic examination revealed suppurative and granulomatous dermatitis involving the superficial and deep dermis. The inflammatory cells were composed of neutrophils and histiocytes. Noninflammatory intravascular thrombosis was noted in the second case. Special stains for acid-fast bacilli, fungi, and bacteria were negative. Culture of lesions in one case was sterile and cultures from the second case contained surface colonization of bacteria that were not known to cause granulomas. Both patients' injection site reactions resolved with conservative management, and one was maintained on therapy.

The authors proposed that a pathogenic mechanism involving tissue ischemia and intravascular thrombi may play a role in interferon-induced granulomatous and suppurative dermatitis, citing morphologic and biochemical similarities with pyoderma gangrenosum and cutaneous Crohn's disease (Sanders et al. 2002).

Lupus Erythematosus-Like Reaction at Injection Site

Arrue et al. (2007) described five cases of cutaneous lesions at the interferon inoculation site with a lupus erythematosus-like histopathologic pattern. Three patients underwent treatment for metastatic melanoma and two patients for multiple sclerosis. The lesions generally occurred anywhere from days to years after inoculation with interferon and were described as painful erythematous nodules or exudative, infiltrated plaques that resolved spontaneously or with minimal treatment within 2-3 weeks. These patients had no history of prior lupus erythematosus, and serum studies conducted after the development of the lesions generally revealed negative antinuclear and deoxyribonucleic acid (DNA) antibodies and normal complement levels.

Histopathologic examination of biopsy specimens revealed prominent dermal mucin deposition highlighted by colloidal iron stain, perivascular and periadnexal infiltrate, and interface changes at the dermal follicular epithelium junction, while the epidermis exhibited only mild acanthosis and parakeratosis. In three of the cases, there was complete destruction of the folliculosebaceous unit, which was replaced by an inflammatory reaction or mucin pools.

While the pathogenesis of this adverse cutaneous reaction is uncertain, the authors of the case report postulate that abnormalities in platelet activation as well as the ability of interferon to transform fibroblast growth factor- β 1, a cytokine that stimulates fibroblasts and, thus, increases the dermal mucin production, may play a role (Arrue et al. 2007).

Embolia Cutis Medicamentosa

Koontz and Alshekhlee (2007) reported a case of a 55-year-old man undergoing treatment with subcutaneous IFN- β 1a who subsequently developed multiple painful necrotic skin lesions at the injection sites in various stages of healing. The diagnosis of embolia cutis medicamentosa was made based on the histologic findings of a thrombotic vasculopathy characterized by thrombotic occlusion of numerous capillaries. The authors hypothesized that patient self-administration of the medication resulted in inadvertent, deeper administration of the medication, leading to accidental arterial or periarterial injection of the agent. Their hypothesis is supported by the observation that after the patient was switched to an auto-injector, which controlled needle depth, no new lesions developed (Koontz and Alsshekhlee 2007).

Embolia cutis medicamentosa (Nicolau syndrome) was first described by Nicolau and Freudenthal in 1924 when they observed this phenomenon after an injection of bismuth salts in patients being treated for syphilis (Koontz and Alsshekhlee 2007). This adverse cutaneous effect is thought to be due to ischemic necrosis secondary to occlusion of the microvasculature (Luton et al. 2006). Proposed pathogenic mechanisms include intra-arterial, periarterial, or perivenous phenomenon wherein embolic or crystal occlusion by the injected agent, pain and sympathetically mediated vasospasm, or vascular trauma-induced thrombosis disrupts the microcirculation (Luton et al. 2006).

Alopecia

Lang et al. (1999) described three cases of transient, localized hair loss at IFN-a2b injection sites in patients undergoing treatment for chronic hepatitis C infection. The lesions were described as patches of non-scarring alopecia with or withperifollicular out erythema or papules. Histopathologic examination revealed a nonscarring alopecia with perifollicular and perivascular lymphohistiocytic infiltrate at the infundibulum and isthmus. It is believed that multiple mechanisms may be at play in this reaction, including the formation of autoantibodies directed against follicular epithelium through interferon-mediated induction of previously masked surface antigens, and direct replicative inhibition of germinative hair cells (Lang et al. 1999).

Kinase Inhibitor (Sorafenib, Sunitinib): Hand-Foot Skin Reaction

Sorafenib and sunitinib are two novel, rationally designed, orally available, multikinase inhibitors (MKIs) used in the treatment of various malignancies for their anti-angiogenic and antiproliferative properties. Initially identified as a Raf-kinase inhibitor, sorafenib has also been shown to inhibit vascular endothelial growth factor receptor (VEGFR) 2-3, platelet-derived growth factor receptor (PDGFR)-β, Fms-like tyrosine kinase 3 (Flt-3), RET, c-KIT, p38α (member of the MAPkinase family), and B-Raf (Wilhelm and Chien 2002). Sunitinib targets VEGFR 1–3, PDGFR- α , c-Kit, Flt-3, colony-stimulating factor receptor-1, and the glial cell line-derived neurotrophic factor receptor (Chow and Eckhardt 2007). Sorafenib is currently approved for the treatment of metastatic renal cell carcinoma (RCC) (Singer et al. 2012) but has also been shown to be effective in treating hepatocellular carcinoma (HCC) (Kokudo et al. 2012). Sunitinib has been approved for the treatment of gastrointestinal stromal tumor (GIST) as well as advanced RCC but has also been shown to be affective against neuroendocrine, colon, and breast cancers in phase II studies (Chow and Eckhardt 2007).

Significant cutaneous adverse events are present in up to 80 % of patients undergoing treatment with kinase inhibitors, of which the hand-foot skin reaction (HFSR) may be the most significant cutaneous toxicity. In a retrospective review of 109 patients on sorafenib for RCC or HCC and 119 patients receiving sunitinib for RCC or GIST, Lee et al. (2009) identified that HFSR was the most commonly observed cutaneous toxicity, seen in 48 % of patients on sorafenib and 36 % of patients on sunitinib. Moreover, HFSR is regarded as the most clinically significant dermatologic toxicity of sorafenib and sunitinib therapy, given the severity of symptoms and the potential for consequent chemotherapeutic dose modification (Chu et al. 2008; Lacouture et al. 2008).

Besides HFSR, a variety of other cutaneous reactions and neoplasms has been reported in

Table 15.4Cutaneous reactions associated with multiki-
nase inhibitors (sorafenib, sunitinib) (Robert et al. 2005,
2011; Smith et al. 2009; Raymond et al. 2010; Bovenschen
et al. 2006; Kong et al. 2008; Rosenbaum et al. 2008; Chu
et al. 2008; Lacouture et al. 2008)

Hand-foot skin reaction (47-83 %)	
Nonspecific rash (19–40 %)	
Hair and nail changes (27 %)	
Mucositis (20 %)	
Xerosis (16 %)	
Xerostomia (11 %)	
Multiple keratoacanthoma-type squamous cell carcinoma	
Eruptive melanocytic nevi	

association with sorafenib and sunitinib (Table 15.4) (Robert et al. 2005, 2011; Smith et al. 2009; Raymond et al. 2010; Bovenschen et al. 2006; Kong et al. 2008; Rosenbaum et al. 2008; Chu et al. 2008; Lacouture et al. 2008). Smith et al. (2009) reported 15 patients who developed keratoacanthoma-type squamous cell carcinomas associated with sorafenib treatment. Regional squamous cell carcinoma has also been recently reported to develop following limb infusion of sorafenib (Raymond et al. 2010). Rapid development of pigmented lesions has been reported in association with blistering disorders, immunodeficiency, and chemotherapeutic agents (Bovenschen et al. 2006). Kong et al. (2008) reported eruptive melanocytic nevi in two patients receiving sorafenib for treatment of metastatic RCC. Other skin changes associated with kinase inhibitors include nonspecific rash, skin discoloration, xerosis, hair color changes, alopecia, and nail changes (Rosenbaum et al. 2008).

Clinical Presentation

The hand-foot skin reaction (HFSR) is a distinct set of palmoplantar lesions in areas of frequent trauma or friction. They can develop anytime from 3 to 56 days (median, 18.4 days) after or within 4 weeks (83 %) of initiating therapy with sorafenib or sunitinib (Lee et al. 2009). These lesions are characterized by multiple, scaly, keratotic plaques on the knuckles of the dorsal hand, hyperkeratotic callus-like lesions on pressure areas of the sole, and/or yellow-colored blisters surrounded by an erythematous halo on fingers and palm (Lee et al. 2009). They are commonly associated with significant tenderness and paresthesias and result in significant alterations in quality of life (Lacouture et al. 2008).

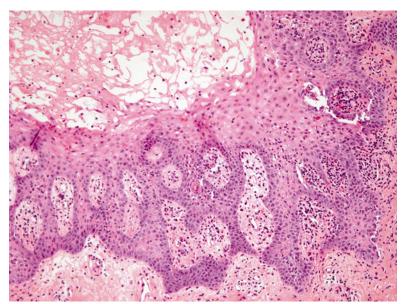
Histologic Features

Skin biopsies from seven patients with HFSR associated with sorafenib or sunitinib showed band-like foci of necrotic keratinocytes in all seven, with two resulting in blister formation (Lacouture et al. 2008). All specimens showed superficial telangiectasia, sparse perivascular lymphocytic infiltrates, and subtle dilatation and cystic degeneration of eccrine glands (Lacouture et al. 2008). Dermal eosinophils are not prominent. The extent of this epidermal change appears to correlate with the duration of therapy with the multikinase inhibitor; less than 30 days of exposure was associated with only intraepidermal changes, while longer exposure times led to changes in the outermost stratum corneum, evident as hyperkeratosis and parakeratosis (Lacouture et al. 2008). In a case series by Lee et al. (2009), similar changes were noted in the skin biopsies from six patients. The general histopathologic pattern is an interface dermatitis with some degree of epidermal necrosis or vesiculation, as noted in some cases (Fig. 15.4) (Lacouture et al. 2008; Lee et al. 2009).

Differential Diagnosis/Pathogenesis

The differential diagnosis for hand-foot skin reaction (HFSR) includes hand-foot syndrome (HFS), palmoplantar keratoderma, radiation recall dermatitis, localized epidermal necrolysis, generalized epidermal necrolysis, and acute graft-versus-host disease (Lacouture et al. 2008; Nagore et al. 2000).

Fig. 15.4 A 51-year-old woman with hepatitis C and hepatocellular carcinoma on sorafenib treatment presented with bilateral vesicles and bullae on her feet. These blisters resolved after cessation of sorafenib treatment. A skin biopsy from her left dorsal foot showed focal epidermal necrosis and necrotic keratinocyte within the epidermis. There is papillary dermal edema and a perivascular lymphocytic infiltrate in the dermis



The pathogenesis of this adverse cutaneous reaction is uncertain. The most widely accepted theory is direct toxicity of chemotherapeutic agents against acral epithelium, which may occur through inhibition of the keratinocyte c-Kit receptor (Chu et al. 2008). Other direct cytotoxic agents have been hypothesized to inflict direct keratinocyte toxicity through eccrine sweat gland excretion (Jacobi et al. 2005). Chu et al. (2008) has suggested that direct effect of sorafenib on eccrine gland receptors may also play a pathogenic role. The histologic findings of apoptotic keratinocytes with satellitosis of lymphocytes led Beard et al. (1993) to propose the host-versusaltered-host response. Despite these suggestions, sorafenib and sunitinib's inhibitory effect on vascular endothelial growth factor (VEGF) and platelet-derived growth factor at the capillary and vascular endothelial level is likely central to the explanation, given that this reaction occurs in areas exposed to repeated trauma such as the palms and soles (Lacouture et al. 2008). Alas, direct VEGF inhibitor (bevacizumab) use does appear to associate with more severe sorafenibassociated HFSR (Azad et al. 2009). However, given that HFSR incidence is rare with isolated bevacizumab use (Chu et al. 2008), sorafenib and sunitinib's inhibitory effect on multiple kinase pathways suggests that there are multiple conspiring mechanisms underlying this reaction.

Treatment

There have been no randomized trials comparing the efficacy of various agents in the treatment of hand-foot skin reaction (HFSR) to multikinase inhibitors. The literature to date supports the use of supportive podiatric measures, including a variety of moisturizers, cotton socks, shock absorbers, and soft shoes that minimize pressure points (Chu et al. 2008). Lacouture et al. (2008) obtained significant clinical improvement in treating sorafenib- and sunitinib-induced HFSR with topical urea (keratolytic that supports skin integrity) alongside tazarotene (reduces epidermal proliferation and dermal inflammation) and/ or topical fluorouracil.

CD20 Inhibitors: Urticaria and Infusion Reactions

In 1997, rituximab (Rituxan, IDEC-C2B8) became the first monoclonal antibody product approved in the United States for the treatment of a malignancy (Dillman 1999). Rituximab is a chimeric monoclonal antibody composed of human immunoglobulin G1 heavy-chain sequences and murine immunoglobulin variable regions that targets the human CD20 antigen (Maloney 2012). CD20 is a multimeric, cell-surface complex antigen that is specific to B cells (Maloney 2012). Physiologically, CD20 regulates transcellular calcium transport and is involved in the regulation of B-cell activation and proliferation (Maloney 2012). Anti-CD20 antibodies such as rituximab are thought to contribute to B-cell-specific destruction through a combination of synergistic mechanisms including complement-dependent cytotoxicity, antibodydependent cell-mediated cytotoxicity, direct antiproliferative effect, and the induction of apoptosis (Maloney 2012).

Originally approved by the Food and Drug Administration for the treatment of follicular B-cell lymphoma, rituximab is one of the most widely used anticancer drugs in the United States. It is currently used to treat an array of malignancies and several inflammatory conditions, including chronic lymphocytic leukemia (Bauer et al. 2012), B-cell lymphomas (Maloney 2012), as well as rheumatoid arthritis (Moots and Naisbett-Groet 2012), systemic lupus erythematosus (Coca and Sanz 2012), and refractory nephrotic syndrome (van Husen and Kemper 2011). Additional anti-CD20 antibodies, including ofatumumab (human antibody), tositumomab (murine antibody), and obinutuzumab (human antibody), have been developed, but their use has been less well documented (Maloney 2012).

Rituximab is typically administered by intravenous infusion (Maloney 2012). Infusionrelated reactions occur at an incidence rate of up to 14 %, as documented in clinical trials (Maloney 2012). To date, there have been three case reports of rituximab infusion-related urticarial reactions in patients undergoing treatment for follicular B-cell lymphoma, despite premedication with diphenhydramine, acetaminophen, and steroids (Heinzerling et al. 2000; Errante et al. 2006; Rey et al. 2009). This cutaneous reaction typically occurs 1 h after the first infusion and is characterized by a pruritic, erythematous to violaceous, well-demarcated, irregularly shaped plaques that develop at the site of existing or previously excised tumors (Errante et al. 2006). The lesions spontaneously regress one to two hours after stopping the infusion (Rey et al. 2009; Errante et al. 2006). Formal histopathologic descriptions of these lesions have yet to be described in the literature, as biopsies of this rapidly resolving reaction may not be readily taken.

Infusion-related localized urticaria is thought to be an epiphenomenon of malignant B-cell destruction as opposed to a true hypersensitivity reaction (Errante et al. 2006). Because it occurs within an hour after the very first infusion further supports this hypothesis (Rey et al. 2009). Van der Kolk et al. (2001) have previously demonstrated that complement activation and subsequent release of tumor necrosis factor- α , interleukin (IL)-6, and IL-8 correlate with the severity of rituximab-related side effects. As such, cytokine release may be involved in the pathomechanism of infusion-related urticaria (Errante et al. 2006). Interestingly, all three reported cases of rituximab infusion-induced urticaria have been in patients undergoing treatment for follicular B-cell lymphoma, which is known to be sensitive to rituximab (Rey et al. 2009).

Other infusion-related adverse effects include fever, chills, nausea, and headache (Maloney 2012; Errante et al. 2006). More serious adverse reactions, including cytokine-release syndrome associated with lymphopenia, thrombocytopenia, and hepatotoxicity (Winkler et al. 1999), potentially life-threatening noninfectious pulmonary toxicity (Hadjinicolaou et al. 2012), and cardiac arrhythmias and acute coronary syndromes (Lee and Kukreti 2012), have been described in the literature. A single case report of Stevens-Johnson syndrome associated with rituximab therapy has been described (Lowndes et al. 2002). Otherwise, cutaneous adverse reactions are characterized predominantly by lesion-localized urticaria.

Alemtuzumab (Campath-1H): Richter's Syndrome

Alemtuzumab (Campath-1H) is a humanized, monoclonal antibody that targets CD52, a cellsurface peptide antigen expressed on normal and malignant B and T lymphocytes (Hale et al. 1985). Approved as salvage therapy for patients with chronic lymphocytic leukemia (CLL) and cutaneous T-cell lymphoma who have failed purine analogue treatment, alemtuzumab induces more prolonged depletion of CD4 and CD8 T-cell subpopulations (Keating et al. 2002). Adverse events associated with alemtuzumab include infusion-related rigors, fever, nausea, vomiting, and rash (Keating et al. 2002). Opportunistic infections, cardiac toxicity, and neutropenia may also occur (Keating et al. 2002; Lenihan et al. 2004).

Richter's syndrome (RS) refers to the development of a second, more aggressive lymphoma in a patient with CLL (Richter 1928). The secondary lymphoma is usually a diffuse large B-cell lymphoma (DLBCL), but the development of Hodgkin's lymphoma (Jamroziak et al. 2012) and T-cell lymphoma (Lee et al. 1995) after CLL has also been described. Studies to date have estimated that Richter's syndrome occurs in anywhere from 1 to 10 % of patients with CLL (Robertson et al. 1993). Clinically, RS is characterized by sudden clinical deterioration with rapid increase in lymphadenopathy or extranodal involvement and less commonly the presence of a monoclonal gammopathy, lytic bone lesions, and multiple cytogenetic changes (Robertson et al. 1993). Molecular analyses of immunoglobulin heavy- and light-chain rearrangements suggest that the secondary lymphoma generally arises from a transformation of the underlying CLL cell (Robertson et al. 1993). However, biclonal origin in RS has been described in the literature (Tohda et al. 1990).

To date, there have been two reported cases in the literature that describe Richter's or Richter'slike transformation associated primarily with alemtuzumab therapy (Janssens et al. 2006; Faguer et al. 2007). A long-term follow-up, retrospective study of 38 patients with CLL who underwent alemtuzumab therapy found that there was an increased but statistically insignificant risk of developing Richter's syndrome in patients on alemtuzumab therapy (16 %, 6/38) compared with consecutive historical controls who underwent alternative salvage therapy (12 %, 9/75) (Karlsson et al. 2006). In their report on 39 patients who developed Richter's syndrome, Robertson et al. (1993) noted that most of those that had developed the transformation had active and more advanced disease. Thus,

whether alemtuzumab therapy poses a true risk for the development of Richter's syndrome is still to be determined. Nonetheless, a summary of the reported clinical and pathologic features and a discussion of proposed pathogenic mechanisms of alemtuzumab-induced Richter's syndrome are noted below.

Clinical Presentation/Histologic Features

Janssens et al. (2006) report the case of a patient with B-CLL who after failing treatment with chlorambucil was started on alemtuzumab. Three months after initiating alemtuzumab therapy, the patient developed a fever and rapidly increasing lactate dehydrogenase (LDH), with recurrent thrombocytopenia and autoimmune hemolytic anemia, which had previously precluded her from fludarabine. Bone marrow biopsy led to a diagnosis of large B-cell lymphoma, revealing the Richter's transformation. Molecular microbiologic studies documented a negative Epstein-Barr virus (EBV) status. Molecular analysis of the immunoglobulin heavy chain confirmed that the CLL and the RS were of different clonal origin, and further analyses revealed that, in fact, the RS cells had evolved from a clone that was present 2 years before the clinical appearance of RS.

Faguer et al. (2007) report the case of a patient with cutaneous T-cell lymphoma (CTCL) who, 2 months after initiating alemtuzumab therapy, developed high fever, night sweats, and dramatic diffuse enlargement of lymph nodes and left temporal cutaneous tumor. A month prior to this presentation, the patient had developed febrile CMV viremia, which after treatment with ganciclovir and prophylaxis with valganciclovir, became and remained negative. Biopsy disclosed a transformed CTCL with CD30+ T cells, revealing a Richter's syndrome-like phenomenon. Rising titers of EBV virus were documented at this time, but in situ hybridization with EBV-specific EBER probes on the biopsy specimen were negative. The patient was treated with a separate course of chemotherapy with no obvious clinical response.

Pathogenesis

It is known that in typical RS, acquisition of TP53 mutations and/or 17p deletions are frequent molecular events (Rossi and Gaidano 2009). However, whether these mutations are sufficient to lead to the disease remains to be determined. The authors of both case reports above are in agreement that alemtuzumab may have promoted the uncontrolled growth of a novel, Richter's clone by eradicating the initial B-CLL or CTCL efficiently and by inducing T-cell depletion with consequent impairment of immunosurveillance (Janssens et al. 2006; Faguer et al. 2007). However, the relationship between chemotherapyinduced immunosuppression and the development of RS remains a controversial issue (Cohen et al. 2002).

Previous studies have emphasized the role of circulating natural killer lymphocytes in the cytotoxic response against Sézary cells (Bouaziz et al. 2005). The transformation of CLL into diffuse large B-cell lymphoma has also been reported after rituximab (Cohen et al. 2002). A very recent case was also reported by Salihoglu et al. (2013 in press) of a CLL patient who developed diffuse large B-cell lymphoma after treatment with rituximab, alemtuzumab. and reduced-intensity conditioning allogenic stem cell transplantation. However, considering the advanced stage of disease, it is still possible that this transformation reflects the natural history of the disease process itself as opposed to being a direct result of chemotherapeutics (Cohen et al. 2002). Further study and experience with alemtuzumab may reveal further insight.

Chronic Hydroxyurea: Pseudodermatomyositis

Hydroxyurea is a cytostatic, antineoplastic agent that interrupts the formation of deoxyribonucleotides from ribonucleotides by inhibiting the enzyme ribonucleotide diphosphate reductase. It is used to treat myeloproliferative disorders such as chronic myelogenous leukemia (CML),

Table 15.5 Mucocutaneous complications of hydroxyurea (Rocamora et al. 2000; Saraceno et al. 2008; Papi et al. 1993; Stasi et al. 1992; Hagen et al. 2012; Richard et al. 1989; Senet et al. 1995; Sigal et al. 1984)

Pseudodermatomyositis	
Skin hyperpigmentation	
Nail pigmentation	
Traumatic or spontaneous ulceration	
Alopecia	
Stomatitis with ulceration	
Plantar keratoderma	
Non-melanoma skin cancer	
Multiple actinic keratoses	

essential thrombocythemia, polycythemia vera, as well as sickle cell anemia and severe, refractory psoriasis (Richard et al. 1989; Halverstam et al. 2008).

Major adverse reactions include bone marrow suppression (Oskay et al. 2002) and lifethreatening alveolitis (Dacey and Callen 2003; Senet et al. 1995) as well as an elevated risk of multiple actinic keratoses and non-melanoma skin cancers, which is thought to be related to the inhibition of DNA damage repair mechanisms (Rocamora et al. 2000; Saraceno et al. 2008; Papi et al. 1993; Stasi et al. 1992). Documented adverse cutaneous effects associated with longterm hydroxyurea therapy include xerosis, hyperpigmentation, atrophy, lower extremity ulcerations, partial alopecia, and chromonychia (Hagen et al. 2012; Richard et al. 1989) (Table 15.5), but these effects are also common with use of other chemotherapeutic agents such as cytarabine, doxorubicin, busulfan, and bleomycin (Senet et al. 1995).

Pseudodermatomyositis is a cutaneous reaction unique to long-term hydroxyurea therapy that was first described in 1975 by Kennedy et al. who documented three patients undergoing said treatment for chronic myelogenous leukemia that developed erythematous lesions, violaceous papules, and atrophy involving the dorsa of the metacarpophalangeal and interphalangeal joints. Sigal et al. (1984) later described the eruption as dermatomyositis-like but ultimately eliminated the diagnosis because of the absence of muscular abnormalities. Interestingly, this rare but characteristic cutaneous side effect has not been documented in psoriatic patients treated with hydroxyurea, but this may be related to the shorter duration of therapy (Senet et al. 1995).

Clinical Presentation

Pseudodermatomyositis typically presents after years (2–10) of hydroxyurea therapy with scaly, linear, and erythematous plaques on the dorsa of the hands (Kennedy et al. 1975) that may or may not be tender (Suehiro et al. 1998). These lesions may strongly resemble the Gottron's sign of dermatomyositis proper (Rocamora et al. 2000). Associated findings include localized atrophy, nails telangiectatic changes, scaly poikilodermatous plaques, or violaceous papules on the feet, elbows, palms, or face (Hagen et al. 2012). The violaceous and erythematous lesions, which can appear around the eyelids and elsewhere on the face, resemble the heliotrope rash of dermatomyositis (Oskay et al. 2002). However, there are typically no signs of proximal muscle weakness or muscle enzymes and electromyography abnormalities (Senet et al. 1995). Furthermore, laboratory evaluation typically reveals negative antinuclear antibody as well as normal aldolase creatine kinase levels (Dacey and Callen 2003). Unlike in dermatomyositis proper, there is little data to suggest that pseudodermatomyositis is associated with any underlying malignancy (Robinson and Reed 2011).

Histologic Features

Histopathologic examination of skin biopsies shows hyperkeratosis, epidermal atrophy, vacuolar changes in the basal layer, and perivascular lymphohistiocytic infiltrate in the superficial dermis (Bahadoran et al. 1996; Senet et al. 1995). Variations on this common theme include focal lichenoid reactions, dyskeratotic cells, dermal mucin deposition, and dermal telangiectasia (Dacey and Callen 2003). Direct immunofluorescence studies have been reportedly negative (Senet et al. 1995).

Differential Diagnosis/Pathogenesis

The histologic differential diagnosis includes dermatomyositis, poikiloderma, lichenoid dermatitis, and graft-versus-host disease (Hagen et al. 2012). The mechanism underlying this reaction is not known at this time. Chronic, cumulative cytologic and selective DNA damage to the basal layer and epidermis is likely involved and may be related to the accumulation of the drug or one of its metabolites (Richard et al. 1989; Senet et al. 1995; Dacey and Callen 2003). Given that leg ulceration is another common adverse event associated with long-term hydroxyurea use, Suehiro et al. (1998) have postulated that chronic cumulative toxicity of the drug on the dividing cells of the basal epidermal layer may play a pathogenic role.

Treatment

Pseudodermatomyositis is a mild eruption that resolves without treatment between 10 days and 18 months after discontinuation of hydroxyurea but runs a benign course even if the medication is continued (Senet et al. 1995).

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