Chapter 10 Bacterial Neuroactive Compounds Produced by Psychobiotics

Rebecca Wall, John F. Cryan, R. Paul Ross, Gerald F. Fitzgerald, Timothy G. Dinan, and Catherine Stanton

Abstract We recently coined the phrase 'psychobiotics' to describe an emerging class of probiotics of relevance to psychiatry [Dinan et al., Biol Psychiatry 2013;74 (10):720–726]. Such "mind-altering" probiotics may act via their ability to produce various biologically active compounds, such as peptides and mediators normally associated with mammalian neurotransmission. Several molecules with neuroactive functions such as gamma-aminobutyric acid (GABA), serotonin, catecholamines and acetylcholine have been reported to be microbially-derived, many of which have been isolated from bacteria within the human gut. Secreted neurotransmitters from bacteria in the intestinal lumen may induce epithelial cells to release molecules that in turn modulate neural signalling within the enteric nervous system and consequently signal brain function and behaviour of the host. Consequently, neurochemical containing/producing probiotic bacteria may be viewed as delivery vehicles for neuroactive compounds and as such, probiotic bacteria may possibly have the potential as a therapeutic strategy in the prevention and/or treatment of certain neurological and neurophysiological conditions.

e-mail: Catherine.stanton@teagasc.ie

J.F. Cryan (\boxtimes) Department of Anatomy and Neuroscience, University College Cork, Cork, Ireland e-mail: j.cryan@ucc.ie

G.F. Fitzgerald Microbiology and Alimentary Pharmabiotic Centre, University College Cork, Cork, Ireland

T.G. Dinan University College Cork, Cork, Ireland

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R. Wall • R.P. Ross • C. Stanton (\boxtimes)

Alimentary Pharmabiotic Centre, Teagasc Moorepark Food Research Centre, Fermoy, Cork, Ireland

Abbreviations

Introduction

We recently coined the phrase 'psychobiotics' to describe an emerging class of probiotics of relevance to psychiatry [[1\]](#page-12-0). Such "mind-altering" probiotics may act via their ability to produce various biologically active compounds, such as peptides and mediators normally associated with mammalian neurotransmission. Several molecules with neuroactive functions such as gamma-aminobutyric acid (GABA), serotonin, catecholamines and acetylcholine have been reported to be microbiallyderived, many of which have been isolated from bacteria within the human gut. Secreted neurotransmitters from bacteria in the intestinal lumen may induce epithelial cells to release molecules that in turn modulate neural signalling within the enteric nervous system and consequently signal brain function and behaviour of the host. Consequently, neurochemical containing/producing probiotic bacteria may be viewed as delivery vehicles for neuroactive compounds and as such, probiotic bacteria may possibly have the potential as a therapeutic strategy in the prevention and/or treatment of certain neurological and neurophysiological conditions.

In recent years, interdisciplinary investigation has revealed strong evidence of the existence of a bidirectional signalling between the intestine and the brain, the so called "brain-gut axis". This communication system integrates neural, hormonal and immunological signalling between the gut and the brain and is critical to maintain homeostasis [\[2](#page-12-0)]. More recently, however, this axis concept was expanded to the "microbiota-gut-brain axis", when it became clear that not only the intestinal tract itself but also its 100 trillion microbial inhabitants can affect the functioning of the central nervous system (CNS) and consequently mood and behaviour [\[3](#page-12-0), [4](#page-12-0)]. The

brain communicates with the enteric microbiota directly by releasing signalling molecules into the gut lumen, and indirectly by altering gastric motility, secretion and intestinal permeability [\[5](#page-12-0)]. Equally, the enteric microbiota can communicate with the host via epithelial cells, receptor-mediated signalling, and stimulation of cells of the lamina propria [\[6](#page-12-0)]. Changes in the composition of the gut microbiota may lead to deterioration in gastrointestinal, neuroendocrine, or immune pathways and relationships, which in turn could lead to alterations in brain-gut interactions and consequently result in disease [\[7](#page-12-0)].

Recently, the microbial endocrinology-based theory was introduced which claimed that probiotics (i.e. live microorganisms that, when ingested in adequate amounts, exerts a health benefit on the host [[8\]](#page-12-0)) function as pharmacological agents and hence act as drug delivery vehicles due to their ability to synthesize neuroactive compounds [\[9](#page-12-0)]. As such, probiotics may affect the brain in a direct manner by producing neurotransmitters and neuromodulators and may therefore have the potential to act as a novel treatment for neuropsychiatric diseases. The delivery of neurochemicals by probiotics may either be in the amount already contained in the bacterium at time of ingestion or what is actively produced by the bacterium once inside the gastrointestinal tract (GIT).

It is well recognized that some bacteria within the human GIT have the capacity to produce many neurotransmitters and neuromodulators. For example, Lactobacillus spp. and Bifidobacterium spp. have been reported to produce GABA; Escherichia spp., and Bacillus spp. have been reported to produce norepinephrine; Streptococcus spp., Escherichia spp. and Enterococcus spp. have been reported to produce serotonin; Bacillus spp. have been reported to produce dopamine, and Lactobacillus spp. have been reported to produce acetylcholine and histamine [[10–](#page-12-0) [14\]](#page-12-0). It is possible that the secreted neurotransmitters from bacteria in the intestinal lumen may induce epithelial cells to release molecules that in turn modulate neural signalling within the enteric nervous system, or act directly on primary afferent axons [\[15](#page-12-0)]. Other bacterially-produced metabolites with proven neuroactive functions include short chain fatty acids (SCFAs) and long chain fatty acids such as conjugated linoleic acid (CLA). Table [10.1](#page-3-0) provides a list of a range of neuroactive chemicals isolated from bacteria within the human gut (it should be noted that this is representative of neuroactives isolated, but is not a complete and comprehensive list). The production of these metabolites and the aforementioned neuroactives by bacteria naturally inhabiting the human gut will be discussed in this chapter.

Bacterial Metabolites

The microbiome has a capability to produce a spectrum of neuroactive compounds, and although we are still in an early stage of exploring its capacity, there is an expanding volume of evidence supporting the role of our intestinal inhabitants as being factories for neurochemicals. Studies comparing germ-free (GF) animals (lacking gut microbiota) with conventional animals (with a normal gut microbiota)

Genus	Neurochemical References	
Lactobacillus, Bifidobacterium	GABA	[10]
Streptococcus, Escherichia, Enterococcus, Lactococcus,	Serotonin	[17, 49]
Lactobacillus		
Escherichia, Bacillus	Norepinephrine $[14, 49]$	
Escherichia, Bacillus, Lactococcus, Lactobacillus,	Dopamine	[14, 17, 49]
<i>Streptococcus</i>		
Lactobacillus, Bacillus	Acetylcholine	[12, 13, 59, 61]
Lactobacillus, Lactococcus, Streptococcus, Enterococcus	Histamine	[11, 66, 67]

Table 10.1 Representative list of neurochemicals isolated from bacteria within the human gut

have demonstrated that the commensal microbiota influence monoamine levels in specific brain regions of the host brain $[3, 16]$ $[3, 16]$ $[3, 16]$ $[3, 16]$ $[3, 16]$. Neurochemicals that have been isolated from gut bacteria include GABA, noradrenaline, serotonin, dopamine and acetylcholine [\[10](#page-12-0), [17,](#page-12-0) [18\]](#page-12-0), which may directly affect the brain. The use of probiotic bacteria that can deliver neurochemicals has further been suggested as a novel treatment for neuropsychiatric diseases [[9\]](#page-12-0). Other bacterial metabolites with neuroactive functions include SCFAs such as propionate and long chain fatty acids such as CLA [\[19](#page-12-0)[–22\]](#page-13-0).

It is not yet clear as to why certain bacteria harbour the genes responsible for the production of neuroactive molecules. It has been proposed that late horizontal gene transfer can explain the existence of genes encoding many of the enzymes involved in the synthetic and metabolic pathways of catecholamines, acetylcholine, and GABA from bacteria. This concept is concordant with increasing evidence that signalling molecules of quorum-sensing systems, used by bacteria to communicate and coordinate their actions [[23\]](#page-13-0) can also bind to mammalian receptors and directly influence the host [[24,](#page-13-0) [25](#page-13-0)]. Neurotransmitters that are produced by the host can furthermore influence the function of members of the microbiota. As an example, the QseC sensor kinase, present in Escherichia coli O157:H7 is a bacterial receptor for host-derived epinephrine/norepinephrine which triggers the transcription of virulence genes in bacteria, a response which can be blocked by adrenergic antagonists [[26\]](#page-13-0).

Gamma-Aminobutyric Acid

Gamma-aminobutyric acid (GABA) is a major inhibitory neurotransmitter in the brain regulating many physiological and psychological processes, and dysfunctions in GABA signalling have been linked to anxiety and depression [\[27](#page-13-0)]. We have recently demonstrated that human intestinally derived strains of lactobacilli and bifidobacteria produce GABA from monosodium glutamate (MSG) in culture [\[10](#page-12-0)] and it has been suggested that microbially produced GABA may have an effect on the brain-gut axis [[28\]](#page-13-0). The production of GABA by commensal bacteria occurs via the same biosynthetic pathway as in neuronal tissue involving conversion of glutamate by the action of the enzyme glutamate decarboxylase (GAD) and vitamin co-factor pyridoxal phosphate [[29\]](#page-13-0). The GABA-producing capability held by some bacterial strains is thought to protect the organism from the acidic environment encountered in the stomach, since its synthesis involves proton exchange for the uptake of glutamate [[30\]](#page-13-0). Many studies have reported the presence of a gad gene in lactic acid bacteria (LAB) [[29,](#page-13-0) [31](#page-13-0), [32](#page-13-0)] and given the heightened interest in the physiological effects associated with GABA, many GABA-enriched fermented food products, using dairy starter cultures with GABA-producing capabilities, have been developed in the past 10 years [[31](#page-13-0), [33](#page-13-0), [34](#page-13-0)]. The levels of GABA that can be achieved in vitro by probiotic organisms are quite large. For example, in the production of fermented foodstuffs, such as Japanese funa-sushi and Chinese traditional paocai, which uses lactobacilli as starter cultures, GABA levels in the millimolar range have been detected in the final products [[35,](#page-13-0) [36\]](#page-13-0).

The prevalence of GABA-producing lactobacilli and bifidobacteria in the human GIT is not as widespread as it is among food-derived LAB. We screened 91 strains of human-derived lactobacilli and bifidobacteria for their ability to produce GABA from MSG, and found that five strains had the ability [[10\]](#page-12-0), with Lactobacillus brevis and Bifidobacterium dentium being the most efficient GABA producers. L. brevis DPC6108 was the most efficient of the strains tested, and it retained the capability to produce GABA in the presence of other gut-derived bacteria (in faecal fermentations). A recent study also demonstrated that GABA production in black soybean milk by L. brevis FPA3709 and its administration to rats resulted in an antidepressant effect similar to that of fluoxetine (a common antidepressant drug) but without the side effects of lost appetite and decreased weight [\[37](#page-13-0)]. Interestingly, neuronal cells have been shown to respond to nanomolar concentrations of GABA [[38\]](#page-13-0). At the level of gene expression, ingestion of the Lactobacillus strain, Lactobacillus *rhamnosus* (JB-1), altered the mRNA expression of both $GABA_A$ and $GABA_B$ receptors. These receptors are implicated in anxiety and depression, and are widely expressed in key brain regions responsible for maintaining normal fear and mood responses [[39\]](#page-13-0).

A number of further potential health benefits of GABA have been described, including induction of hypotension, diuretic effects, and tranquilizer effects [\[40](#page-14-0), [41\]](#page-14-0). Furthermore, GABA has a receptor-mediated role in a number of immunological (i.e. down-regulation of cytokine released by proinflammatory cells release) and intestinal neurophysiological (i.e. secretion of neuropeptides by intrinsic and extrinsic intestinal nerve fibers) processes [[38](#page-13-0), [42](#page-14-0), [43\]](#page-14-0). Given the broad health benefits associated with GABA, the use of a GABA-secreting bacterium, acting on dietary glutamate, could have potential in both neuropsychiatric diseases and in inflammatory conditions such as inflammatory bowel disease (IBD), however, in vivo studies are required to ascertain whether the host would benefit from microbially- produced GABA.

Serotonin and 5-HT Precursors

Serotonin (5-hydroxytryptamine, 5-HT) is a metabolite of the essential amino acid tryptophan and plays an important role in the regulation of a number of bodily functions, including mood. Today, the vast majority of antidepressant drugs lead to increases in the levels of serotonin in the brain. Serotonin is ubiquitously distributed in nature and has been found in some plants (fruits and nuts) and in both vertebrates and invertebrates animals [\[44](#page-14-0)]. Some studies also indicate that bacteria can synthesize serotonin and/or induce its production by the host. Wikoff et al. [[45\]](#page-14-0) utilized a metabolomics-based approach to study the metabolic products of the microbiome in mice which may impact health, and unexpectedly found that serotonin plasma levels were nearly threefold higher in conventional mice compared with GF mice, whereas plasma concentrations of tryptophan was 40 % lower in conventional animals than in their GF counterparts. The authors postulated that the increased plasma serotonin levels observed could indirectly result from an as yet undefined host microbe interaction [\[45](#page-14-0)]. Furthermore, increased serotonin turnover and altered levels of related metabolites in the striatum [[3\]](#page-12-0) and hippocampus [\[46](#page-14-0)] of GF mice have been reported. The levels of serotonin in the cortex and hippocampus were also significantly reduced in GF mice [\[4](#page-12-0)], suggesting a role for the microbiota in maintaining serotonin levels. Rats that were given *Bifidobacterium infantis* for 14 days had increased concentrations of the serotonin precursor tryptophan in plasma, suggesting that commensal bacteria have the ability to influence tryptophan metabolism [[47\]](#page-14-0). This effect on tryptophan metabolism may be mediated by the impact of the microbiota on the expression of indoleamine-2,3-dioxygenase, a key enzyme in the physiologically dominant kynurenine pathway of tryptophan degradation $[48]$ $[48]$. Moreover, Özogul $[17]$ $[17]$ investigated the influences of LAB on biogenic amine formation by foodborne pathogens using single and mix cultures. All the LAB species used in their study, including *Lactococcus lactis* subsp. *cremoris* (MG 1363), L. lactis subsp. lactis (IL1403), Lactobacillus plantarum (FI8595) and Streptococcus thermophilus (NCFB2392) produced serotonin to some extent [\[17](#page-12-0)]. Shishov et al. [[49\]](#page-14-0) also demonstrated that E. coli K-12 was capable of producing serotonin at nanomolar concentrations in culture.

Catecholamines

Catecholamines such as dopamine and norepinephrine (also known as noradrenaline) are the major neurotransmitters that mediate a variety of the CNS functions, such as motor control, cognition, memory processing, emotion and endocrine regulation [\[50](#page-14-0)]. Dysfunctions in catecholamine neurotransmission are implicated in some neurological and neuropsychiatrical disorders, including Parkinson's dis-ease [\[51](#page-14-0)], Alzheimer's disease [\[52](#page-14-0)] and major depressive disorders [\[53](#page-14-0)].

Both norepinephrine and dopamine were identified in bacteria in a study by Tsavkelova et al. [[14\]](#page-12-0). Dopamine, in concentrations from 0.45 to 2.13 mmol/L was found in the biomass of Bacillus cereus, B. mycoides, B. subtilis, Proteus vulgaris, Serratia marcescens, S. aureus, and E. coli, and norepinephrine was detected (0.21– 1.87 mmol/L) in B. mycoides, B. subtilis, P. vulgaris, and S. marcescens. Moreover, it was demonstrated that bacteria, particularly B . *subtilis* may release norepinephrine and dopamine out of the cell and perhaps in this way might participate in intercellular microbe–microbe and microbe–host communications [\[14](#page-12-0)]. Shishov et al. $[49]$ $[49]$ used the E. coli K-12 strain to investigate the production of catecholamines in vitro and demonstrated that this E. coli strain can produce dopamine and norepinephrine and also their precursor, DOPA, in culture. The culture fluid of E. coli contained micromolar concentrations of DOPA and nanomolar concentrations of dopamine and norepinephrine $[49]$ $[49]$. Moreover, Ozogul $[17]$ $[17]$ demonstrated the production of dopamine by some LAB species in culture, namely L. lactis subsp. cremoris (MG 1363), L. lactis subsp. lactis (IL1403), L. plantarum (FI8595) and S. thermophilus (NCFB2392). A recent study by Asano et al. [\[54](#page-14-0)] demonstrated that bacteria which constitute the normal microbiome in mice are capable of the in vivo production of large quantities of norepinephrine. Furthermore, adoptive transfer of the microbiome of mice that could produce norepinephrine in vivo to GF mice resulted in the in vivo elaboration of norepinephrine within the murine GIT [\[54](#page-14-0)]. Interestingly, in many cases, the content of catecholamines found in bacteria is higher than in human blood, for example concentrations of norepinephrine in human blood are found to be 0.04 mmol/L [\[55](#page-14-0)].

Acetylcholine

Acetylcholine is a well-known neurotransmitter in the central and peripheral nervous systems that plays a critical role in cognitive function, particularly in memory and learning. It is synthesized by choline acetyltransferase in the CNS and by both choline acetyltransferase and carnitine acetyltransferase in the peripheral system [[56,](#page-14-0) [57\]](#page-14-0). Acetylcholine has also been identified in non-neuronal tissues, including gastrointestinal, respiratory and urogenital epithelial cells [\[58](#page-14-0)]. In addition, acetylcholine has been found to be a component of bacteria and its production was discovered in a strain of L. plantarum [[13,](#page-12-0) [59,](#page-14-0) [60](#page-14-0)]. Cell free enzyme(s) participating in acetylcholine synthesis were also found in L. plantarum [\[59](#page-14-0)]. Horiuchi et al. [\[61](#page-15-0)] tested three different bacterial strains for acetylcholine content and synthesis, E. coli JCM 5491, Staphylococcus aureus JCM 2151 and Bacillus subtilis PCI 219. Among these, a substantial amount of acetylcholine was detected in B. subtilis (55.7 pmol/ 10^{10} colony forming units), while much smaller amounts were found in $E.$ coli (2.22 pmol) and $S.$ aureus (0.39 pmol). Although acetylcholine synthesis was detected in all bacterial samples, the levels were low and the authors suggested that an acyltransferase other than choline

acetyltransferase and carnitine acetyltransferase was responsible for acetylcholine synthesis in these bacteria [\[61](#page-15-0)].

Histamine

Histamine acts as a modulatory neurotransmitter in the mammalian brain and has an important role in the maintenance of wakefulness, while dysfunction in the histaminergic system has been linked to narcolepsy [[62\]](#page-15-0). Moreover, behavioural studies suggest that the histaminergic system in the brain has important roles in cognitive function [[63\]](#page-15-0). Levels of histamine are decreased in the hippocampus, temporal cortex and hypothalamus of patients with Alzheimer's disease, suggesting that histaminergic neurons undergo degeneration and contribute to cognitive decline in this disorder [[64\]](#page-15-0).

Histamine is produced via histidine decarboxylase (HDC) of L-histidine, an essential amino acid for humans that is present in many dietary foods [[65\]](#page-15-0). Some fermentative bacteria, including strains of Lactobacillus, Lactococcus, Streptococcus, Pediococcus and Enterococcus have been reported to possess the HDC-gene and to produce histamine at different levels [\[66](#page-15-0), [67\]](#page-15-0). However, the production of histamine by certain bacterial strains has caused alarm as a health risk in food and as a marker of food spoilage. Ingestion of food containing high concentrations of histamine has been linked with headaches, vomiting and hypertension [[68\]](#page-15-0). Nonetheless, histamine production by the probiotic strain Lactobacillus reuteri (ATCC PTA 6475) was recently reported, resulting in a suppression of human proinflammatory tumor necrosis factor (TNF) production [\[11](#page-12-0)]. Moreover, histidine supplementation increased the expression of HDC-genes and the production of histamine by L. reuteri. In addition to the role of histamine in immunomodulation observed in the study by Thomas et al. [[11\]](#page-12-0), luminal production of histamine by L. reuteri 6475 may influence signalling in the enteric nervous system. However, in vivo studies are needed to gain a better understanding of the role of bacterially-produced histamine in the gut and that such production does not induce negative side-effects.

Indole-3-Propionic Acid

Indole-3-propionic acid (IPA) is a deamination product of tryptophan and is found in plasma and cerebrospinal fluid $[69, 70]$ $[69, 70]$ $[69, 70]$. IPA has been shown to be a powerful antioxidant [[71](#page-15-0)] and has been considered as a possible treatment for Alzheimer's disease [[72](#page-15-0)] due to its ability to protect neurons and neuroblastoma cells against oxidative damage and death [\[73](#page-15-0), [74](#page-15-0)].

Wikoff et al. [\[45](#page-14-0)] demonstrated that the production of IPA was completely dependent on the presence of gut microbiota and could also be established by colonizing germ-free mice with the bacterium Clostridium sporogenes. An earlier study by Jellet et al. [[75\]](#page-15-0) also demonstrated that IPA was present in the spent bacterial media of C. sporogenes and that addition of the precursor tryptophan to the media greatly enhanced the formation of IPA. In addition, IPA was shown to be produced in vitro when human large intestinal contents were incubated with tryptophan and indolelactate [[76\]](#page-15-0).

Short-Chain Fatty Acids

Short chain fatty acids (SCFA) are the major products of the bacterial fermentation of carbohydrates and proteins in the GIT $[19, 77]$ $[19, 77]$ $[19, 77]$ $[19, 77]$. The main compounds are acetic, propionic and n-butyric acids, occurring roughly in molar ratios of 60:20:20 in the colon [[78\]](#page-15-0). Through their absorption and metabolism, the host is able to salvage energy from foodstuffs, particularly resistant starch and fibers that are not digested in the upper part of the GIT. The main site for SCFA production and absorption is the proximal large intestine, where the fermentation of undigested food by colonic bacteria occurs at high rates. Bacteria that produce SCFA include, but are not limited to, Bacteroides, Bifidobacterium, Propionibacterium, Eubacterium, Lactobacillus, Clostridium, Roseburia and Prevotella [[19\]](#page-12-0). SCFA have a multiplicity of effects in the body, and affect epithelial cell transport and metabolism, epithelial cell growth and differentiation, and hepatic control of lipid and carbohydrates, while providing energy sources for muscles and kidneys, as well as the heart and brain [\[79](#page-15-0)]. Epithelial cells in the distal colon derive 60–70 % of their energy requirements from bacterial fermentation products [\[80](#page-15-0)]. SCFA also act as signalling molecules. Propionate, acetate, and to a lesser extent butyrate are ligands for at least two G protein-coupled receptors (GPCRs), Gpr41 and Gpr43, which are broadly expressed in the distal small intestine, colon and adipocytes [\[81](#page-15-0), [82\]](#page-15-0). SCFA interaction with Gpr43 can profoundly affect inflammatory responses. For example, mice treated with oral acetate showed a substantial decrease in inflammation. This protection was mediated by acetate binding to Gpr43, because acetate had no effect in Gpr43-deficient mice. Furthermore, it was shown that Gpr43 exhibited enhanced expression in neutrophils and eosinophils, suggesting that SCFA-Gpr43 signalling is one of the molecular pathways whereby commensal bacteria regulate immune and inflammatory responses [\[83](#page-16-0)]. Gpr43 is also induced during adipocyte differentiation and exhibits increased levels during high-fat feeding in rodents, suggesting that Gpr43 may also affect adipocyte function [\[84](#page-16-0)]. Hong et al. [\[85](#page-16-0)] demonstrated that acetate and propionate act on lipid accumulation and inhibition of lipolysis mainly through Gpr43 [\[85](#page-16-0)]. Gpr41 has been shown to be implicated in microbiota-dependent regulation of host adiposity and leptin production [\[86](#page-16-0)].

SCFA can cross the blood-brain barrier and enter the CNS [\[87](#page-16-0)] and are taken up by glia and, to a lesser extent, by neurons, where they are thought to comprise a major energy source in cellular metabolism, particularly during early brain

development [[88–90\]](#page-16-0). They also play a role in cell signalling [[91\]](#page-16-0) and neurotransmitter synthesis and release [[92\]](#page-16-0). Moreover, SCFA have been shown to increase the synthesis of dopamine and its related catecholamines through induction of tyrosine hydroxylase, a key enzyme in the synthesis of catecholamines [[93](#page-16-0)]. Propionate, in particular has been shown to alter dopamine, serotonin, and glutamate systems in a manner similar to that observed in autism spectrum disorders (ASD) [[94,](#page-16-0) [95\]](#page-16-0). Furthermore, intraventricular infusion of propionate in rats was shown to impair social behaviour and cause brain abnormalities, similar to those detected in human autism [\[96–98](#page-16-0)] and furthermore to alter brain phospholipid composition [[99\]](#page-16-0). Some clinical studies have also found that a subset of ASD patients have high levels of Clostridial and Bacteriodetes species in the gut [[100,](#page-17-0) [101](#page-17-0)], species which are efficient propionate-producers [[19\]](#page-12-0). This highlights that although propionate is beneficial at appropriate levels, such as lowering lipogenesis, serum cholesterol levels and improving insulin sensitivity [[102\]](#page-17-0), excessive propionate may have negative effects on health and behaviour.

Butyrate is known to exhibit many important physiological functions in eukaryotic cells [\[19](#page-12-0)]. One of the most recognised cellular mechanisms for the action of butyrate is its effects on histone acetylation $[103]$ $[103]$, where the inhibition of histone deacetylase facilitates hyperacetylation of histone proteins to occur, thus facilitating the access of DNA repair enzymes. Interestingly, sodium butyrate has been demonstrated to elicit an antidepressant effect in the murine brain [[104\]](#page-17-0). When injected systemically, sodium butyrate induced a short-lasting, transient acetylation of histones in frontal cortex and hippocampus, in conjunction with dynamic changes in expression of brain-derived neurotrophic factor (BDNF), thereby resulting in an antidepressant-like behavioral response [\[104](#page-17-0)].

Long-Chain Fatty Acids

Long-chain fatty acids (LC-PUFAs) play numerous roles in the brain, including structural (forming the physico-chemical properties in the lipid bilayer of cellular membranes) and signalling functions. Moreover, they influence neurogenesis and neurotransmission within the nervous tissue. Arachidonic acid (AA, C20:4n-6) and docosahexaenoic acid (DHA, C22:6n-3) are highly concentrated in the brain and are vital fatty acids for neurological development [[105,](#page-17-0) [106\]](#page-17-0). Furthermore, there is a growing body of evidence for their role in mental health across the lifespan [\[107](#page-17-0)]. Being the major structural components of brain cells [\[108](#page-17-0)], AA and DHA influence cell membrane physical properties, enzyme activity, regulation of ion channels and neuroreceptors and their signalling (neurotransmission) [[109,](#page-17-0) [110](#page-17-0)].

Recently, we have reported that administration of a *Bifidobacterium breve* strain, B. breve NCIMB702258 to mice had a significant impact on the fatty acid composition of brain $[111]$ $[111]$. Mice that received this strain for 8 weeks exhibited significantly higher concentrations of AA and DHA in the brain when compared to unsupplemented mice. Interestingly, this effect was bacterial strain-dependent, as it was not induced by the B. breve strain DPC6330. The mechanism by which the B. breve strain alters the fatty acid composition is currently unknown. Possible explanations include modulations of fat-absorption processes in the small intestine and/or desaturase activities involved in the metabolism of fatty acids to the longerchain unsaturated derivatives caused either directly by the strain administered or by alterations in the gut microbiota. Interestingly, it was previously postulated that different members of the gut microbiota promote fatty acid absorption via distinct mechanisms $[112]$ $[112]$. Semova et al. $[112]$ $[112]$ used the zebrafish model to investigate how microbiota and diet interact to regulate lipid absorption in the gut epithelium. By comparing GF zebrafish with conventional zebrafish, the authors demonstrated that the microbiota stimulate fatty acid uptake and lipid droplet formation in both the intestinal epithelium and liver [\[112](#page-17-0)]. Previous studies have also demonstrated that manipulation of the gut microbiota by probiotics resulted in altered fat composition in the host $[113-115]$. Although the adult microbiome is not known to be particularly enriched in genes involved in fatty acid metabolism [[116\]](#page-17-0), there are indications that interactions between fatty acids and components of the gut microbiota occur which could affect the biological roles of both. However, a deeper knowledge of such interactions and what consequences they have for the host are warranted.

Conjugated Linoleic Acid

Conjugated linoleic acid (CLA) comprises a mixture of positional and geometric isomers of linoleic acid (cis-9, cis-12 C18:2n-6) characterised by the presence of conjugated double bonds with cis or trans configurations. CLA is a natural component of ruminant milk and tissue fat as a result of the action of the ruminal microbiota on dietary linoleic acid [\[117](#page-17-0)]. A range of health-promoting activities have been attributed to the consumption of CLA, most notably anticarcinogenic, immune-modulatory, anti-obesity and anti-atherosclerotic activities [[118–](#page-17-0)[121\]](#page-18-0). In contrast, in certain conditions, some CLA isomers can also exert potentially negative effects such as liver steatosis and insulin resistance [[122](#page-18-0), [123\]](#page-18-0). Regarding the action of CLA on the CNS, it is known that CLA crosses the blood-brain barrier and is incorporated and metabolized in the brain [\[124\]](#page-18-0). Dietary CLA has been shown to reduce cerebral prostaglandin E2 in the peripheral and CNS [\[125](#page-18-0)], and to exert antiangiogenic actions in the brain [[126\]](#page-18-0). Furthermore, Hunt et al. [\[127](#page-18-0)] showed that CLA protects cortical neurons from excitotoxicity at concentrations likely achieved by consumption of CLA as a dietary supplement.

Evidence of the role of the gut microbiota in the endogenous production of CLA was first reported by Chin et al. [[128\]](#page-18-0) who observed that increasing the amount of linoleic acid in the diet increased the tissue content of CLA in conventional rats but not in GF rats. Since then, commensal lactobacilli and bifidobacteria from the human GIT, most notably *B. breve* strains and *L. plantarum* strains, have been shown to produce CLA, predominantly the *cis-9*, *trans-11* (c9, t11) isomer from free linoleic acid [\[21–23,](#page-13-0) [129\]](#page-18-0). These bacteria convert linoleic acid to CLA in a similar manner to ruminant bacteria via the action of the enzyme linoleic acid isomerase [[130\]](#page-18-0), which also has been sequenced in some *Lactobacillus* and Bifidobacterium strains [\[131](#page-18-0), [132](#page-18-0)]. We have reported that the CLA-producing bacterium, B. breve NCIMB702258 converts linoleic acid to CLA in the murine gut, resulting in significantly elevated c9, t11 CLA in the liver [\[133](#page-18-0)]. Moreover, Bassaganya-Riera et al. [[134\]](#page-18-0) demonstrated that administration of the probiotic mixture VSL#3 to mice with colitis resulted in high colonic concentrations of c9, t11 CLA and that this locally produced CLA improved colitis by activating peroxisome proliferator-activated receptor gamma (PPAR γ) in macrophages. Two studies have also reported the in vivo production of the CLA isomer, trans-10, cis-12 (t10, c12) CLA, by using two strains of human origin, L. rhamnosus PL60 and L. plantarum PL62. Administration of these two strains to mice resulted in increased t10, c12 CLA concentrations in sera with subsequent reductions in adipose tissue and body weight [\[135](#page-18-0), [136](#page-18-0)]. Druart et al. [\[137](#page-18-0)] further demonstrated that prebiotic supplementation increased CLA content in caecal tissue, by increasing substrate availability and by modulating gut microbiota composition.

Although dietary CLA has been reported to affect the CNS, the consequences of bacterially-produced CLA on nervous system function are yet to be discovered.

Conclusion

Although we are still at the very early stages of understanding the complex communication systems between gut bacteria and the brain, we know that certain bacteria within the human gut have the ability to produce molecules with neuroactive functions which could affect the brain in a direct manner. However, only cultivable bacteria have been tested for their capacity to produce neuroactive compounds in vitro and only a limited number of bacterial strains have been tested up to now. Moreover, in complex microbial ecosystems such as in the human gut, interactions and competition exist between bacteria, which are not studied upon simple culture conditions in vitro. This highlights the need for in vivo studies to elucidate the role of metabolite-producing bacteria and what effect such bacteria, and their components, have on nervous system function and behaviour. Such future studies may also facilitate our understanding of the consequences of neuroactive compound production by the microbiota and how probiotic bacteria can influence the CNS, and could furthermore identify the potential for neurochemical containing/producing probiotic bacteria as a therapeutic strategy in the treatment of certain neurological and neurophysiological conditions. Given that molecular tools have now been developed for many intestinal organisms, the possibility exists now to overproduce neuroactive compounds and/or to regulate their production in response to gut metabolites such as bile.

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References

- 1. Dinan TG, Stanton C, Cryan JF (2013) Psychobiotics: a novel class of psychotropic. Biol Psychiatry 74(10):720–726
- 2. Collins SM, Surette M, Bercik P (2012) The interplay between the intestinal microbiota and the brain. Nat Rev Microbiol 10(11):735–742
- 3. Heijtz RD, Wang S, Anuar F, Qian Y, Bjorkholm B, Samuelsson A, Hibberd ML, Forssberg H, Pettersson S (2011) Normal gut microbiota modulates brain development and behavior. Proc Natl Acad Sci U S A 108:3047–3052
- 4. Neufeld K, Kang N, Bienenstock J, Foster J (2011) Reduced anxiety-like behavior and central neurochemical change in germ-free mice. Neurogastroenterol Motil 23:255–264
- 5. Rhee SH, Pothoulakis C, Mayer EA (2009) Principles and clinical implications of the braingut-enteric microbiota axis. Nat Rev Gastroenterol Hepatol 6:306–314
- 6. O'Mahony SM, Hyland NP, Dinan TG, Cryan JF (2011) Maternal separation as a model of brain-gut axis dysfunction. Psychopharmacology (Berl) 214:71–88
- 7. Cryan JF, O'Mahony SM (2011) The microbiome-gut-brain axis: from bowel to behaviour. Neurogastroenterol Motil 23:187–192
- 8. FAO/WHO (2001) Report on Joint FAO/WHO Expert Consultation on Evaluation of Health and Nutritional Properties of Probiotics in Food including Powder Milk with Live Lactic Acid Bacteria. ftp://ftp.fao.org/es/esn/food/probio_report_en.pdf
- 9. Lyte M (2011) Probiotics function mechanistically as delivery vehicles of neuroactive compounds: microbial endocrinology in the design and use of probiotics. Bioessays 33(8): 574–581
- 10. Barrett E, Ross RP, O'Toole PW, Fitzgerald GF, Stanton C (2012) γ-Amino butyric acid production by culturable bacteria from the human intestine. J Appl Microbiol 113:411–417
- 11. Thomas CA, Hong T, van Pijkeren JP, Hemarajata P, Trinh DV, Hu W, Britton RA, Kalkum M, Versalovic J (2012) Histamine derived from probiotic Lactobacillus reuteri suppress TNF via modulation of PKA and ERK signalling. PLoS One 7(2):e31951
- 12. Kawashima K, Misawa H, Moriwaki Y, Fujii YX, Fujii T, Horiuchi Y, Yamada T, Imanaka T, Kamekura M (2007) Ubiquitous expression of acetylcholine and its biological functions in life forms without nervous systems. Life Sci 80:2206–2209
- 13. Marquardt P, Spitznagel G (1959) Bakterielle Acetylcholine Bildung in Kunstlichen Nahrboden. Arzneimittelforschung 9:456–465
- 14. Tsavkelova EA, Botvinko IV, Kudrin VS, Oleskin AV (2000) Detection of neurotransmitter amines in microorganisms using of high performance liquid chromatography. Dokl Biochem 372:115–117 (in Russian issue 840–842)
- 15. Forsythe P, Kunze WA (2013) Voices from within: gut microbes and the CNS. Cell Mol Life Sci 70:55–69
- 16. Nishino R, Mikami K, Takahashi H, Tomonaga S, Furuse M, Hiramoto T, Aiba Y, Koga Y, Sudo N (2013) Commensal microbiota modulate murine behaviors in a strictly contamination-free environment confirmed by culture-based methods. Neurogastroenterol Motil 25:521–e371
- 17. Özogul F (2011) Effects of specific lactic acid bacteria species on biogenic amine production by foodborne pathogens. Int J Food Sci Technol 46(3):478–484
- 18. Roshchina VV (2010) Evolutionary considerations of neurotransmitters in microbial, plant, and animal cells. In: Lyte M, Freestone PPE (eds) Microbial endocrinology: interkingdom signaling in infectious disease and health. Springer, New York, pp 17–52
- 19. Macfarlane GT, Macfarlane S (2012) Bacteria, colonic fermentation, and gastrointestinal health. J AOAC Int 95(1):50–60
- 20. Coakley M, Ross RP, Nordgren M, Fitzgerald G, Devery R, Stanton C (2003) Conjugated linoleic acid biosynthesis by human-derived Bifidobacterium species. J Appl Microbiol 94: 138–145
- 21. Barrett E, Ross RP, Fitzgerald GF, Stanton C (2007) Rapid screening method for analyzing the conjugated linoleic acid production capabilities of bacterial cultures. Appl Environ Microbiol 73:2333–2337
- 22. Rosberg-Cody E, Ross RP, Hussey S, Ryan CA, Murphy BP, Fitzgerald GF, Devery R, Stanton C (2004) Mining the microbiota of the neonatal gastrointestinal tract for CLA-producing bifidobacteria. Appl Environ Microbiol 70:4635–4641
- 23. Hughes DT, Sperandio V (2008) Inter-kingdom signalling: communication between bacteria and their hosts. Nat Rev Microbiol 6:111–120
- 24. Boontham P, Robins A, Chandran P, Pritchard D, Camara M, Williams P, Chuthapisith S, McKechnie A, Rowlands BJ, Eremin O (2008) Significant immunomodulatory effects of Pseudomonas aeruginosa quorum-sensing signal molecules: possible link in human sepsis. Clin Sci (Lond) 115:343–351
- 25. Telford G, Wheeler D, Williams P, Tomkins PT, Appleby P, Sewell H, Stewart GS, Bycroft BW, Pritchard DI (1998) The Pseudomonas aeruginosa quorum-sensing signal molecule N- (3-oxododecanoyl)-L-homoserine lactone has immunomodulatory activity. Infect Immun 66: 36–42
- 26. Clarke MB, Hughes DT, Zhu C, Boedeker EC, Sperandio V (2006) The QseC sensor kinase: a bacterial adrenergic receptor. Proc Natl Acad Sci U S A 103:10420–10425
- 27. Schousboe A, Waagepetersen HS (2007) GABA: homeostatic and pharmacological aspects. In: Tepper JM, Abercrombie ED, Bolam JP (eds) Gaba and the basal ganglia: from molecules to systems. Elsevier Science, Amsterdam, pp 9–19
- 28. Bienenstock J, Forsythe P, Karimi K, Kunze W (2010) Neuroimmune aspects of food intake. Int Dairy J 20:253–258
- 29. Komatsuzaki N, Nakamura T, Kimura T, Shima J (2008) Characterization of glutamate decarboxylase from a high gamma-aminobutyric acid (GABA)-producer, Lactobacillus paracasei. Biosci Biotechnol Biochem 72:278–285
- 30. Higuchi T, Hayashi H, Abe K (1997) Exchange of glutamate and gammaaminobutyrate in a Lactobacillus strain. J Bacteriol 179:3362–3364
- 31. Siragusa S, De Angelis M, Di Cagno R, Rizzello CG, Coda R, Gobbetti M (2007) Synthesis of gammaaminobutyric acid by lactic acid bacteria isolated from a variety of Italian cheeses. Appl Environ Microbiol 73:7283–7290
- 32. Hiraga K, Ueno YH, Oda KH (2008) Glutamate decarboxylase from Lactobacillus brevis: activation by ammonium sulfate. Biosci Biotechnol Biochem 72:1299–1306
- 33. Park KB, Oh SH (2006) Isolation and characterization of Lactobacillus buchneri strains with high gamma-aminobutyric acid producing capacity from naturally aged cheese. Food Sci Biotechnol 15:86–90
- 34. Rizzello CG, Cassone A, Di Cagno R, Gobbetti M (2008) Synthesis of angiotensin I-converting enzyme (ACE)-inhibitory peptides and gamma-aminobutyric acid (GABA) during sourdough fermentation by selected lactic acid bacteria. J Agric Food Chem 56: 6936–6943
- 35. Komatsuzaki N, Shima J, Kawamoto S, Momose H (2005) Production of gammaaminobutyric acid (GABA) by Lactobacillus paracasei isolated from traditional fermented foods. Food Microbiol 22:497–504
- 36. Li HX, Gao DD, Cao YS, Xu HY (2008) A high gamma-aminobutyric acid-producing Lactobacillus brevis isolated from Chinese traditional paocai. Ann Microbiol 58:649–653
- 37. Ko CY, Victor Lin HT, Tsai GJ (2013) Gamma-aminobutyric acid production in black soybean milk by *Lactobacillus brevis* FPA 3709 and the antidepressant effect of the fermented product on a forced swimming rat model. Process Biochem 48(4):559–568
- 38. Krantis A (2000) GABA in the mammalian enteric nervous system. News Physiol Sci 15: 284–290
- 39. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, Bienestock J, Cryan JF (2011) Ingestion of Lactobacillus strain regulates emotional behavior and central

GABA receptor expression in a mouse via the vagus nerve. Proc Natl Acad Sci U S A 108: 16050–16055

- 40. Jakobs C, Jaeken J, Gibson KM (1993) Inherited disorders of GABA metabolism. J Inherit Metab Dis 16:704–715
- 41. Wong CG, Bottiglieri T, Snead OC (2003) GABA, gammahydroxybutyric acid, and neurological disease. Ann Neurol 54:S3–S12
- 42. Bjurstom H, Wang J, Ericsson I, Bengtsson M, Liu Y, Kumar-Mendu S, Issazadeh-Navikas S, Birnir B (2008) GABA, a natural immunomodulator of T lymphocytes. J Neuroimmunol 205: 44–50
- 43. Page AJ, O'Donnell TA, Blackshaw LA (2006) Inhibition of mechanosensitivity in visceral primary afferents by GABAB receptors involves calcium and potassium channels. Neuroscience 137:627–636
- 44. Kema IP, de Vries EG, Muskiet FA (2000) Clinical chemistry of serotonin and metabolites. J Chromatogr B Biomed Sci Appl 747(1–2):33–48
- 45. Wikoff WR, Anfora AT, Liu J, Schultz PG, Lesley SA, Peters EC, Siuzdak G (2009) Metabolomics analysis reveals large effects of gut microflora on mammalian blood metabolites. Proc Natl Acad Sci U S A 106:3698–3703
- 46. Clarke G, Grenham S, Scully P, Fitzgerald P, Moloney RD, Shanahan F, Dinan TG, Cryan JF (2013) The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. Mol Psychiatry 18:666–673
- 47. Desbonnet L, Garrett L, Clarke G, Bienenstock J, Dinan TG (2008) The probiotic Bifidobacteria infantis: an assessment of potential antidepressant properties in the rat. J Psychiatr Res 43:164–174
- 48. Forsythe P, Sudo N, Dinan T, Taylor VH, Bienenstock J (2010) Mood and gut feelings. Brain Behav Immun 24:9–16
- 49. Shishov VA, Kirovskaia TA, Kudrin VS, Oleskin AV (2009) Amine neuromediators, their precursors, and oxidation products in the culture of Escherichia coli K-12. Prikl Biokhim Mikrobiol 45(5):550–554
- 50. Kobayashi K (2001) Role of catecholamine signalling in brain and nervous system functions: new insights from mouse molecular genetic study. J Investig Dermatol Symp Proc 6:115–121
- 51. Calabresi P, Castrioto A, Di Filippo M, Picconi B (2013) New experimental and clinical links between the hippocampus and the dopaminergic system in Parkinson's disease. Lancet Neurol 12:811–821
- 52. Robertson IH (2013) A noradrenergic theory of cognitive reserve: implications for Alzheimers disease. Neurobiol Aging 34(1):298–308
- 53. Hamon M, Blier P (2013) Monoamine neurocircuitry in depression and strategies for new treatments. Prog Neuropsychopharmacol Biol Psychiatry 45:54–63
- 54. Asano Y, Hiramoto T, Nishino R, Aiba Y, Kimura T, Yoshihara K (2012) Critical role of gut microbiota in the production of biologically active, free catecholamines in the gut lumen of mice. Am J Physiol Gastrointest Liver Physiol 303(11):G1288–G1295
- 55. Kruk ZL, Pycock CJ (1990) Neurotransmitters and drugs. Chapman and Hall, New York
- 56. Tucˇek S (1988) Choline acetyltransferase and the synthesis of acetylcholine. In: Whittaker VP (ed) Handbook of experimental pharmacology. The cholinergic synapse, vol 86. Springer, Berlin, pp 125–165
- 57. Tucˇek S (1882) The synthesis of acetylcholine in skeletal muscles of the rat. J Physiol 322:53–69
- 58. Wessler I, Kirkpatrick CJ, Racke K (1999) Cholinergic "pitfall": acetylcholine, a universal cell molecule widely distributed in biological systems: expression and function in humans. Clin Exp Pharmacol Physiol 26:198–205
- 59. Girvin GT, Stevenson JW (1954) Cell free "choline acetylase" from Lactobacillus plantarum. Can J Biochem Physiol 32:131–146
- 60. Rowatt E (1948) The relation of pantothenic acid to acetylcholine formation by a strain of Lactobacillus plantarum. J Gen Microbiol 2:25–30
- 61. Horiuchi Y, Kimura R, Kato N, Fujii T, Seki M, Endo T, Kato T, Kawashima K (2003) Evolutional study on acetylcholine expression. Life Sci 72:1745–1756
- 62. Panula P, Nuutinen S (2013) The histaminergic network in the brain: basic organization and role in disease. Nat Rev Neurosci 14:472–487
- 63. Alvarez EO (2009) The role of histamine on cognition. Behav Brain Res 199:183–189
- 64. Airaksinen MS, Paetau A, Paljarvi L, Reinikainen K, Riekkinen P, Suomalainen R, Panula P (1991) Histamine neurons in human hypothalamus: anatomy in normal and Alzheimer diseased brains. Neuroscience 44:465–481
- 65. Wu G (2009) Amino acids: metabolism, functions, and nutrition. Amino Acids 37:1–17
- 66. Landete JM, de las Rivas B, Marcobal A, Munoz R (2008) Updated molecular knowledge about histamine biosynthesis by bacteria. Crit Rev Food Sci Nutr 48:697–714
- 67. Coton E, Rollan G, Bertrand A, Lonvaud-Funel A (1998) Histamine producing lactic acid bacteria: early detection, frequency and distribution. Am J Enol Vitic 49:199–204
- 68. Shalaby AR (1996) Significance of biogenic amines in food safety and human health. Food Res Int 29:675–690
- 69. Morita I, Kawamoto M, Yoshida H (1992) Difference in the concentration of tryptophan metabolites between maternal and umbilical foetal blood. J Chromatogr 576:334–339
- 70. Young S, Anderson GM, Gauthier S, Purdy WC (1980) The origin of indoleacetic and indolepropionic acid in rat and human cerebrospinal fluid. J Neurochem 34:1087–1092
- 71. Karbownik M, Reiter RJ, Garcia JJ, Cabrera J, Burkhardt S, Osuna C, Lewinski A (2001) Indole-3-propionic acid, a melatonin-related molecule, protects hepatic microsomal membranes from iron-induced oxidative damage: relevance to cancer reduction. J Cell Biochem 81:507–513
- 72. Bendheim PE, Poeggeler B, Neria E, Ziv V, Pappolla MA, Chain DG (2002) Development of indole-3-propionic acid (OXIGON) for Alzheimer's disease. J Mol Neurosci 19:213–217
- 73. Hwang IK, Yoo KY, Li H, Park OK, Lee CH, Choi JH, Jeong YG, Lee YL, Kim YM, Kwon YG, Won MH (2009) Indole-3-propionic acid attenuates neuronal damage and oxidative stress in the ischemic hippocampus. J Neurosci Res 87:2126–2137
- 74. Chyan YJ, Poeggeler B, Omar RA, Chain DG, Frangione B, Ghiso J, Pappolla MA (1999) Potent neuroprotective properties against the Alzheimer β -amyloid by an exogenous melatonin-related indole structure, indole-3-propionic acid. J Biol Chem 274(31): 21937–21942
- 75. Jellet JJ, Forrest TP, Macdonald IA, Marrie TJ, Holdeman LV (1980) Production of indole-3 propionic acid and 3-(p-hydroxyphenyl) propionic acid by Clostridium sporogenes: a convenient thin-layer chromatography detection system. Can J Microbiol 26(4):448–453
- 76. Smith EA, Macfarlane GT (1997) Formation of phenolic and indolic compounds by anaerobic bacteria in the human large intestine. Microb Ecol 33:180–188
- 77. Kovatcheva-Datchary P, Zoetendal EG, Venema K, de Vos WM, Smidt H (2009) Review: tools for the tract: understanding the functionality of the gastrointestinal tract. Therap Adv Gastroenterol 2:s9–s22
- 78. Cummings JH (1981) Short chain fatty-acids in the human-colon. Gut 22:763–779
- 79. Cummings JH (1995) In: Gibson GR, Macfarlane GT (eds) Human colonic bacteria: role in nutrition, physiology and health. CRC, Boca Raton, pp 101–130
- 80. Topping DL, Clifton PM (2001) Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. Physiol Rev 81(3):1031–1064
- 81. Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, Muir AI, Wigglesworth MJ, Kinghorn I, Fraser NJ, Pike NB, Strum JC, Steplewski KM, Murdock PR, Holder JC, Marshall FH, Szekeres PG, Wilson S, Ignar DM, Foord SM, Wise A, Dowell SJ (2003) The orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. J Biol Chem 278:11312–11319
- 82. Xiong Y, Miyamoto N, Shibata K, Valasek MA, Motoike T, Kedzierski RM, Yanagisawa M (2004) Short-chain fatty acids stimulate leptin production in adipocytes through the G protein-coupled receptor GPR41. Proc Natl Acad Sci U S A 101:1045–1050
- 83. Maslowski KM, Vieira AT, Ng A, Kranich J, Sierro F, Yu D, Schilter HC, Rolph MS, Mackay F, Artis D, Xavier RJ, Teixeira MM, Mackay CR (2009) Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. Nature 461:1282–1286
- 84. Ge H, Li X, Weiszmann J, Wang P, Baribault H, Chen JL, Tian H, Li Y (2008) Activation of G protein-coupled receptor 43 in adipocytes leads to inhibition of lipolysis and suppression of plasma free fatty acids. Endocrinology 149(9):4519–4526
- 85. Hong Y, Nishimura HY, Hishikawa D, Tsuzuki H, Miyahara H, Gotoh C, Choi KC, Feng DD, Chen C, Lee HG, Katoh K, Roh SG, Sasaki S (2005) Acetate and propionate short chain fatty acids stimulate adipogenesis via GPCR43. Endocrinology 146(12):5092–5099
- 86. Samuel BS, Shaito A, Motoike T, Rey FE, Backhed F, Manchester JK, Hammer RE, Williams SC, Crowley J, Yanagizawa M, Gordon JI (2008) Effects of the gut microbiota on host adiposity are modulated by the short-chain fatty acid binding G protein-coupled receptor, Gpr41. Proc Natl Acad Sci U S A 105(43):16767–16772
- 87. Karuri AR, Dobrowsky E, Tannock IF (1993) Selective cellular acidification and toxicity of weak organic acids in an acidic microenvironment. Br J Cancer 68:1080–1087
- 88. Maurer MH, Canis M, Kuschinsky W, Duelli R (2004) Correlation between local monocarboxylate transporter 1 (MCT1) and glucose transporter 1 (GLUT1) densities in the adult rat brain. Neurosci Lett 355:105–108
- 89. Rafiki A, Boulland JL, Halestrap AP, Ottersen OP, Bergersen L (2003) Highly differential expression of the monocarboxylate transporters MCT2 and MCT4 in the developing rat brain. Neuroscience 122:677–688
- 90. Peinado A, Yuste R, Katz LC (1993) Extensive dye coupling between rat neocortical neurons during the period of circuit formation. Neuron 10:103–114
- 91. Nakao S, Moriya Y, Furuyama S, Niederman R, Sugiya H (1998) Propionic acid stimulates superoxide generation in human neutrophils. Cell Biol Int 22:331–337
- 92. DeCastro M, Nankova BB, Shah P, Patel P, Mally PV, Mishra R, La Gamma EF (2005) Short chain fatty acids regulate tyrosine hydroxylase gene expression through a cAMP-dependent signaling pathway. Brain Res Mol Brain Res 142:28–38
- 93. Shah P, Nankova BB, Parab S, La Gamma EF (2006) Short chain fatty acids induce TH gene expression via ERK-dependent phosphorylation of CREB protein. Brain Res 1107:13–23
- 94. El-Ansary AK, Ben BA, Kotb M (2012) Etiology of autistic features: the persisting neurotoxic effects of propionic acid. J Neuroinflammation 9:74
- 95. Mitsui R, Ono S, Karaki S, Kuwahara A (2005) Neural and nonneural mediation of propionate-induced contractile responses in the rat distal colon. Neurogastroenterol Motil 17:585–594
- 96. Schultz SR, MacFabe DF, Martin S, Jackson J, Taylor R, Boon F, Ossenkopp KP, Caiin DP (2009) Intracerebroventricular injections of the enteric bacterial metabolic product propionic acid impair cognition and sensorimotor ability in the Long-Evans rat: further development of a rodent model of autism. Behav Brain Res 200:33–41
- 97. Schultz SR, MacFabe DF, Ossenkopp KP, Scratch S, Whelan J, Taylor R, Cain DP (2008) Intracerebroventricular injection of propionic acid, an enteric bacterial metabolic end-product, impairs social behavior in the rat: implications for an animal model of autism. Neuropharmacology 54:901–911
- 98. MacFabe DF, Cain DP, Rodriguez-Capote K, Franklin AE, Hoffmann JE, Boon F, Taylor AR, Kavaliers M, Ossenkopp KP (2007) Neurobiological effects of intraventricular propionic acid in rats: possible role of short chain fatty acids on the pathogenesis and characteristics of autism spectrum disorders. Behav Brain Res 176:149–169
- 99. Thomas RH, Meeking MM, Mepham JR, Tichenoff L, Possmayer F, Liu S, MacFabe DF (2012) The enteric bacterial metabolite propionic acid alters brain and plasma phospholipid molecular species: further development of a rodent model of autism spectrum disorder. J Neuroinflammation 9:153
- 100. Finegold SM, Dowd SE, Gontcharova V, Liu CX, Henley KE, Wolcott RD (2010) Pyrosequencing study of fecal microflora of autistic and control children. Anaerobe 16: 444–453
- 101. Song Y, Liu C, Finegold SM (2004) Real-time PCR quantification of clostridia in feces of autistic children. Appl Environ Microbiol 70:6459–6465
- 102. Hosseini E, Grootaert C, Verstraete W, Van de Wiele T (2011) Propionate as a healthpromoting microbial metabolite in the human gut. Nutr Rev 69(5):245–258
- 103. Kruh J, Defer N, Tichonicky L (1995) In: Cummings JH, Rombeau JL, Sakata T (eds) Physiological and clinical aspects of short chain fatty acids. Cambridge University Press, Cambridge, pp 275–288
- 104. Schroeder FA, Lin CL, Crusio WE, Akbarian S (2007) Antidepressant- like effects of the histone deacetylase inhibitor, sodium butyrate, in the mouse. Biol Psychiatry 62:55–64
- 105. Haag M (2003) Essential fatty acids and the brain. Can J Psychiatry 48(3):195–203
- 106. Lauritzen L, Hansen HS, Jørgensen MH, Michaelsen KF (2001) The essentiality of long chain n-3 fatty acids in relation to development and function of the brain and retina. Prog Lipid Res 40:1–94
- 107. Sinn N, Milte C, Howe PRC (2010) Oiling the brain: a review of randomised controlled trials of omega-3 fatty acids in psychopathology across the lifespan. Nutrients 2(2):128–170
- 108. O'Brien JS, Sampson EL (1965) Lipid composition of the normal human brain: gray matter, white matter, and myelin. J Lipid Res 6:537–544
- 109. Heinrichs SC (2010) Dietary ω-3 fatty acid supplementation for optimizing neuronal structure and function. Mol Nutr Food Res 54:447–456
- 110. Chalon S (2006) Omega-3 fatty acids and monoamine neurotransmission. Prostaglandins Leukot Essent Fatty Acids 75:259–269
- 111. Wall R, Marques TM, O'Sullivan O, Ross RP, Shanahan F, Quigley EM, Dinan TG, Kiely B, Fitzgerald GF, Cotter PD, Fouhy F, Stanton C (2012) Contrasting effects of Bifidobacterium breve NCIMB 702258 and Bifidobacterium breve DPC 6330 on the composition of murine brain fatty acids and gut microbiota. Am J Clin Nutr 95(5):1278–1287
- 112. Semova I, Carten JD, Stombaugh J, Mackey LC, Knight R, Farber SA, Rawls JF (2012) Microbiota regulate intestinal absorption and metabolism of fatty acids in the zebrafish. Cell Host Microbe 12:277–288
- 113. Hoppu U, Isolauri E, Laakso P, Matomäki J, Laitinen K (2012) Probiotics and dietary counselling targeting maternal dietary fat intake modifies breast milk fatty acids and cytokines. Eur J Nutr 51(2):211–219
- 114. Kaplas N, Isolauri E, Lampi AM, Ojala T, Laitinen K (2007) Dietary counselling and probiotic supplementation during pregnancy modify placental phospholipid fatty acids. Lipids 45:865–870
- 115. Kankaanpaa PE, Yang B, Kallio HP, Isolauri E, Salminen SJ (2002) Influence of probiotic supplemented infant formula on composition of plasma lipids in atopic infants. J Nutr Biochem 13:364–369
- 116. Kurokawa K, Itoh T, Kuwahara T, Oshima K, Toh H, Toyoda A, Takami H, Morita H, Sharma VK, Srivastava TP, Taylor TD, Noguchi H, Mori H, Ogura Y, Ehrlich DS, Itoh K, Takagi T, Sakaki Y, Hayashi T, Hattori M (2007) Comparative metagenomics revealed commonly enriched gene sets in human gut microbiomes. DNA Res 14:169–181
- 117. Kepler CR, Hirons KP, McNeill JJ, Tove SB (1966) Intermediates and products of the biohydrogenation of linoleic acid by Butyrinvibrio fibrisolvens. J Biol Chem 241:1350–1354
- 118. Gaullier JM, Halse J, Hoye K, Kristiansen K, Fagertun H, Vik H, Gudmundsen O (2004) Conjugated linoleic acid supplementation for 1 y reduces body fat mass in healthy overweight humans. Am J Clin Nutr 79:1118–1125
- 119. Ip MM, Masso-Welch PA, Ip C (2003) Prevention of mammary cancer with conjugated linoleic acid: role of the stroma and the epithelium. J Mammary Gland Biol Neoplasia 8: 103–118
- 120. Bassaganya-Riera J, Hontecillas R, Beitz DC (2002) Colonic anti-inflammatory mechanisms of conjugated linoleic acid. Clin Nutr 21:451–459
- 121. Kritchevsky D, Tepper SA, Wright S, Tso P, Czarnecki SK (2000) Influence of conjugated linoleic acid (CLA) on establishment and progression of atherosclerosis in rabbits. J Am Coll Nutr 19:472S–477S
- 122. Taylor CG, Zahradka P (2004) Dietary conjugated linoleic acid and insulin sensitivity and resistance in rodent models. Am J Clin Nutr 79:1164S–1168S
- 123. Clement L, Poirier H, Niot I, Bocher V, Guerro-Millo M, Krief S, Staels B, Besnard P (2002) Dietary trans-10, cis-12 conjugated linoleic acid induces hyperinsulinemia and fatty liver in the mouse. J Lipid Res 43:1400–1409
- 124. Fa M, Diana A, Carta G, Cordeddu L, Melis MP, Murru E, Sogos V, Banni S (2005) Incorporation and metabolism of c9, t11 and t10, c12 conjugated linoleic acid (CLA) isomers in rat brain. Biochim Biophys Acta 1736:61–66
- 125. Nakanishi T, Koutoku T, Kawahara S, Murai A, Furuse M (2003) Dietary conjugated linoleic acid reduces cerebral prostaglandin E2 in mice. Neurosci Lett 341:135–138
- 126. Sikorski AM, Hebert N, Swain RA (2008) Conjugated linoleic acid (CLA) inhibits new vessel growth in the mammalian brain. Brain Res 1213:35–40
- 127. Hunt WT, Kamboj A, Anderson HD, Anderson CM (2010) Protection of cortical neurons from excitotoxicity by conjugated linoleic acid. J Neurochem 115:123–130
- 128. Chin SF, Storkson JM, Liu W, Albright KJ, Pariza MW (1994) Conjugated linoleic acid (9,11- and 10,12-octadecadienoic acid) is produced in conventional but not germ free rats fed linoleic acid. J Nutr 124:694–701
- 129. Kishino S, Ogawa J, Omura Y, Matsumura K, Shimizu S (2002) Conjugated linoleic acid production from linoleic acid by lactic acid bacteria. JAOCS 79:159–163
- 130. Lin TY, Lin CW, Wang YJ (2002) Linoleic acid isomerase activity in enzyme extracts from Lactobacillus acidophilus and Propionibacterium freudenreichii ssp. shermanii. J Food Sci 67:1502–1505
- 131. Macouzet M, Robert N, Lee BH (2010) Genetic and functional aspects of linoleate isomerase in Lactobacillus acidophilus. Appl Microbiol Biotechnol 87:1737–1742
- 132. Macouzet M, Lee BH, Robert N (2010) Genetic and structural comparison of linoleate isomerases from selected food-grade bacteria. J Appl Microbiol 109:2128–2134
- 133. Wall R, Ross RP, Shanahan F, O'Mahony L, O'Mahony C, Coakley M, Hart O, Lawlor P, Quigley EM, Kiely B, Fitzgerald GF, Stanton C (2009) Metabolic activity of the enteric microbiota influences the fatty acid composition of murine and porcine liver and adipose tissues. Am J Clin Nutr 89(5):1393–1401
- 134. Bassaganya-Riera J, Viladomiu M, Pedragosa M, De Simone C, Carbo A, Shaykhutdinov R, Jobin C, Arthur JC, Corl BA, Vogel H, Storr M, Hontecillas R (2012) Probiotic bacteria produce conjugated linoleic acid locally in the gut that targets macrophage PPAR $γ$ to suppress colitis. PLoS One 7(2):e31238
- 135. Lee K, Paek K, Lee HY, Park JH, Lee Y (2007) Antiobestity effect of trans-10, cis-12 conjugated linoleic acid-producing Lactobacillus plantarum PL62 on diet-induced obese mice. J Appl Microbiol 103:1140–1146
- 136. Lee HY, Park JH, Seok SH, Baek MW, Kim DJ, Lee KE, Paek KS, Lee Y, Park JH (2006) Human originated bacteria, Lactobacillus rhamnosus PL60, produce conjugated linoleic acid and show anti-obesity effects in diet-induced obese mice. Biochim Biophys Acta 1761: 736–744
- 137. Druart C, Neyrinck AM, Dewulf EM, De Backer FC, Possemiers S, Van De Wiele T, Moens F, De Vuyst L, Cani PD, Larondelle Y, Delzenne NM (2013) Implication of fermentable carbohydrates targeting the gut microbiota on conjugated linoleic acid production in high-fat-fed mice. Br J Nutr 110:998–1011