# Chapter 3 Mitochondrion: A Missing Link in Asthma Pathogenesis

Ulaganathan Mabalirajan, Anurag Agrawal, and Balaram Ghosh

**Abstract** Asthma is a multifactorial airway disease with airway hyperresponsiveness, airway inflammation, goblet cell metaplasia, and structural changes including airway smooth muscle proliferation and subepithelial fibrosis. Airway epithelial injury and apoptosis is an important triggering and amplification point in asthma pathogenesis, and mitochondrial dysfunction in epithelial cells appears to play an important role. On the other hand, mitochondrial biogenesis is an important aspect of smooth muscle hypertrophy and fibroblast proliferation, which leads to airway remodeling and hyperresponsiveness. In mice, preexisting mitochondrial dysfunction has been shown to potentiate allergic experimental asthma. In this review, we summarize the current understanding on the involvement of mitochondria in asthma pathogenesis, discuss the probable points of intersection between lung pathobiology and mitochondrial biology, and speculate regarding the road ahead. Mitochondrial influence on cellular oxidative and nitrative stress, apoptosis, and calcium homeostasis is covered in detail, as well as the role of molecules like nitric oxide synthase, asymmetric dimethyl arginine (ADMA), and peroxynitrite on mitochondrial function, epithelial injury, and asthma. Potential therapeutic strategies involving coenzyme Q, vitamin E, and esculetin that influence mitochondrial function and alleviate features of asthma are also discussed.

Keywords Asthma • Mitochondria • Mitochondrial dysfunction • Apoptosis

B. Ghosh, PhD, FNA, FNASc, FASc (🖂)

U. Mabalirajan, MBBS, PhD • A. Agrawal, MD, PhD

Molecular Immunogenetics Laboratory, Centre of Excellence for Translational Research in Asthma and Lung Disease, CSIR- Institute of Genomics and Integrative Biology, Delhi, India

Molecular Immunogenetics Laboratory, CSIR- Institute of Genomics and Integrative Biology, Delhi University Campus (North), Mall Road, Delhi 110007, India e-mail: bghosh@igib.res.in

V. Natarajan, N.L. Parinandi (eds.), *Mitochondrial Function in Lung Health and Disease*, Respiratory Medicine 15, DOI 10.1007/978-1-4939-0829-5\_3,
© Springer Science+Business Media New York 2014

# Introduction

Albert von Kölliker described some intracellular parts (now mitochondria) present in muscle cell as "granules" in 1857, then Richard Altman stained and named them as "bioblasts" in 1886, and finally Carl Benda coined them as "*mitochondria*" (mitos: thread, chondros: granule) [1]. Today we know that the humble mitochondrion, a primitive structure believed to have evolved from  $\alpha$ -proteobacteria, is not merely a powerhouse of the cell but its function could regulate the function of whole organism [2]. This has led to its emergence as a key organelle in several chronic and metabolic diseases. Both basic and applied researchers are excited in exploring mitochondrial biology, thanks to the recognition of novel functions and features of mitochondria in addition to energy synthesis such as apoptosis, oxidative stress, calcium regulation, and mitochondrial DNA mutation-mediated diseases.

The mitochondrion has solute permeable outer membrane, intermembranous space, solute impermeable inner membrane, and matrix. The inner mitochondrial membrane is embedded with respiratory chain enzymes that utilize electrons from NADH or FADH2 as energy to create a proton gradient in the intermembranous space, which further drives ATP production by ATP synthase [2]. While the mitochondria have a separate genome that replicates independently, mitochondrial DNA can synthesize only 13 proteins for electron transport chain, whereas proper mitochondrial structure and function need more than a thousand proteins; so mitochondria have sophisticated protein import machineries on both membranes to import nucleus-encoded mitochondrial precursor proteins with mitochondrial targeting signals from cytosol [3]. In addition to the nucleus, the mitochondrion has a tight functional connection with most of the compartments of the cell and physical contact with few crucial organelles such as the endoplasmic reticulum, indicating that mitochondrion could be the point of convergence for cellular stress signaling [4–6].

Asthma (is a Greek word meaning "panting") affects a significant portion of the population worldwide and represents a significant health and economic burden [7, 8]. Asthma is a syndrome rather than a single disease as it has various phenotypes; however, most of these phenotypic features converge in episodic airway obstruction and hyperresponsiveness [9]. Asthma is symptomatically characterized by difficulty in breathing, wheezing, coughing, increased sputum production, and chest pain. It is pathophysiologically characterized by airway hyperresponsiveness, airway inflammation including airway eosinophilia, increased IgE, mucus hypersecretion, and structural changes of the airway called airway remodeling [10]. Current definition of asthma is based on three components: chronic airway inflammation, reversible airflow obstruction, and enhanced bronchial reactivity [10]. Asthma symptoms are commonly triggered by airway irritants such as air pollution and smoke, allergens, respiratory infections, certain drugs, exercise, etc. Although the majority of asthmatics can be managed with available anti-asthma medications, complex phenotypes such as obese asthma, neutrophilic asthma, and steroid-resistant asthma are poorly understood and challenging to manage effectively [11–14]. Thus, there is a need to identify alternate pathogenetic pathways to unravel the complexity and devise new therapies.

Asthma is not considered to be a mitochondrial disease or syndrome. Most mitochondrial syndromes are characterized by neurological and/or muscular abnormalities, since the high metabolism of neurons and muscle cells requires a large functional mitochondrial mass, without which they would lose function. While neurons and smooth muscle cells are implicated in asthma pathogenesis (neurohumoral control, bronchial smooth muscle, and cellular dysfunction with increased release of mediators), the observed defects are of excessive function, e.g., excessive vagal tone or smooth muscle contraction [15, 16]. While other commonalities between asthma pathogenesis and mitochondrial biology exist, namely, (a) maternal transmission in some studies, (b) oxidative/nitrative stress and antioxidant defense, (c) apoptosis/necrosis, and (d) calcium ion homeostasis, these are largely nonspecific. Reliable associations between mitochondrial dysfunction and asthma came from reports of abnormal mitochondria in the bronchial epithelia of asthmatic children and mice [17, 18]. The bronchial epithelium is a primary site of injury in asthma, and epithelial cell apoptosis is a critical event in asthma pathogenesis. Since apoptosis signaling is classically composed of two pathways, a direct pathway from death receptor ligation and an alternative pathway mediated by mitochondria during cell stress, mitochondrial abnormality could precipitate into the development of asthma. This was experimentally shown in mice, when mice with preexisting mitochondrial dysfunction were found to be more susceptible to allergic airway inflammation and developed more severe disease [19, 20].

The findings discussed above provided an association between mitochondrial dysfunction and asthma as well as a possible mechanism, opening the possibility that restoration of mitochondrial health may be beneficial in asthma. It had previously been shown that a mitochondria-targeted antioxidant, coenzyme Q10, reduced corticosteroid requirement in asthmatic patients [21]. The benefits of targeting mitochondria with antioxidants such as vitamin E and N-acetyl-L-cysteine (NAC) in allergic diseases [22, 23] supported the possible involvement of mitochondria in allergic diathesis. In this chapter, we discuss comprehensively the role of mitochondria in asthma pathogenesis based on the findings from our lab and others.

#### **Mitochondrial Genetics and Asthma**

Every mitochondrion contains 2–10 mtDNA copies, and in humans each copy has approximately 16,569 nucleotides, which can generate only 37 genes in its own: 22 tRNAs, 13 polypeptides which are important in electron transport chain, and two rRNAs (12S and 16S rRNA) (Fig. 3.1). Mitochondrial DNA has a higher tendency for mutations as the mitochondrion does not have efficient DNA repair mechanisms and the mitochondrion itself is the major source of oxidative free radicals [24]. As mitochondria from paternal sperm are not contributing in the generation of fertilized ovum, at least in mammals, maternal inheritance is the common feature in many mitochondrial DNA mutation-mediated syndromes [25]. From a genetic standpoint, asthma is a complex disorder with more than a hundred genes



**Fig. 3.1** Schematic diagram of human mitochondrial genome to show the mtDNA-derived 13 polypeptides, 22 tRNAs (each was named based on respective amino acid), and two rRNAs (12S and 16S). It also shows the reported mutations/polymorphisms in asthmatics along with respective associated asthma phenotypes

implicated so far. While asthma runs in families, it does not always follow Mendelian patterns, and different genes may be relevant in different individuals [26-28]. Though asthma is not considered as a mitochondrial disease, maternal history of asthma is one of the substantial risk factors for the development of asthma [29, 30] compared to paternal. Similar maternal influence has been observed in other atopic diseases such as atopic dermatitis, allergic rhinitis, etc. [31, 32]. Since maternal transmission is the hallmark of mitochondrial diseases, mitochondria could be important in the inheritance of asthma or atopy risk. Interestingly, maternal transmission of asthma features, such as airway hyperresponsiveness and increased susceptibility to allergic airway inflammation, was noted in mice. Interleukin-4 (IL-4), one of the Th2 cytokines crucial for most of the features of asthma, was shown to be involved in this transmission [33]. We have shown that administration of IL-4 monoclonal antibody reduces mitochondrial dysfunction and ultrastructural changes of bronchial epithelial mitochondria, suggesting that IL-4 may influence mitochondrial function [19]. IL-4 is also involved in IgE class switching, and importantly maternal IgE levels are well correlated with child cord blood IgE levels [34]. However, while this supports a possible role for mitochondria, various environmental factors such as transplacental passage of molecules and epigenetic factors are also likely to be involved in maternal mode of inheritance [35]. Interestingly, recent microarray studies performed in the placenta have revealed that various crucial mitochondrial genes are altered in the existence of maternal asthma during pregnancy, and glucocorticoid usage for asthma treatment during pregnancy also altered mitochondrial genes [36].

In support of a direct role for mitochondria in asthma, common mitochondrial haplogroups, especially haplogroup U from European population, have shown association with increased serum IgE levels [35]. Another study found five mutations in mitochondrial gene-encoding tRNAs (A12308G in tRNA leucine (CUN), G15928A in tRNA threonine, 595insC in tRNA phenylalanine, T10448C in tRNA arginine, and A8343G in tRNA lysine) and five more mutations in tRNA flanking regions [37]. Importantly, A12308G mutation is part of the mitochondrial haplogroup U, which was found to be significantly associated with total serum IgE levels. Another case-control study in Caucasian European children has found an association of the genomic region containing the ATPAF1 (ATP synthase mitochondrial F1 complex assembly factor 1) gene in Caucasian European children [38]. Further, they found that ATPAF1 gene expression was increased in asthmatic bronchial biopsies [38]. In other studies, peripheral blood mononuclear cells (PBMCs) of allergic patients were found to differentially express nine mitochondrial genes, including II and III subunits of cytochrome c oxidase [39]; cytochrome b gene polymorphism was linked with nonallergic bronchial asthma [40]; and substitution polymorphism at 195 site and insertion polymorphism at 309 site of hypervariable regions of D loop of mitochondria were associated with bronchitis in children [41]. Interestingly, A3243G tRNALeu(UUR) mutation which is associated with MIDD (maternally inherited diabetes and deafness syndrome) and MERRF (myoclonic epilepsy and ragged-red fibers) syndromes has been found in rare forms of asthma along with other comorbidity conditions such as maculopathy or depression [42, 43]. It is to be noted that A3243G tRNALeu(UUR) leads to severe combined respiratory chain assembly defect with abnormal incorporation of amino acids, alteration in stabilization, methylation, aminoacylation, and codon recognition of tRNA [44]. These indicate the necessity of initiating a study of mitochondrial various in various populations to understand the exact role of mitochondria in the pathogenesis of asthma and other diseases.

#### **Mitochondrial Function and Asthma**

It is also necessary to note that mitochondrial research has seen periodic surges related to discoveries of new functions of mitochondria, starting from initial descriptions as energy sources to critical orchestrators of cell death [45]. Meanwhile, the understanding and management of asthma have also seen various evolutions (from the concept of smooth muscle defect to airway remodeling), thanks to the rapid



**Fig. 3.2** Schematic diagram shows the respiratory organelle of the cell on one side and respiratory organ of body on the other side to indicate the possible common aspects between mitochondrial biology and asthma pathogenesis. These are ATP generation, 15-lipoxygensase (*15-LOX*), asymmetric dimethyl L-arginine (*ADMA*), apoptosis, reactive oxygen free radicals (*ROS*), reactive nitrogen free radicals (*RNS*), and calcium homeostasis. *OMM* outer mitochondrial membrane, *IMS* intermembranous space, *IMM* inner mitochondrial membrane, *g.c.* goblet cell, *c.e.* ciliated epithelia, *s.e.f.* subepithelial fibrosis, *a.s.m.* airway smooth muscle

advances in the field of immunology and molecular biology [46, 47]. While a link between the two was not realized until recently, many common features can be seen between mitochondrial pathobiology and asthma pathophysiology (Fig. 3.2).

# **Mitochondrial Dysfunction**

Before our lab demonstrated the potential role of mitochondrial dysfunction in asthma, there were scattered reports indicating increased number of mitochondria in the bronchial epithelia of asthmatic children and experimental allergic mice [17, 18]. The functional status of such mitochondria was not known. We found that allergic murine lungs show features of mitochondrial dysfunction like decreased cytochrome c oxidase activity in lung mitochondria, decrease in the expressions of the third subunit of cytochrome c oxidase and 17 kDa subunit of complex I in bronchial epithelium, decreased lung ATP levels, and increased cytochrome c in lung cytosol [19].



Healthy Mouse

Asthmatic Mouse

Fig. 3.3 Transmission electron microscopy of bronchial epithelia of asthmatic mice

Importantly, this was associated with mitochondrial ultrastructural changes such as mitochondrial swelling and loss of cristae. Further, IL-4 monoclonal antibody administration reduced these mitochondrial dysfunction and ultrastructural changes, indicating that mitochondrial dysfunction is associated with asthma pathogenesis (Fig. 3.3) [19]. In another study, mice exposed to environmental irritants such as tobacco smoke and printer emissions exhibited damage to lung mitochondria along with increased generation of reactive oxygen species (ROS) [48]. However, whether the observed mitochondrial dysfunction was the cause or effect in asthma pathogenesis remained uncertain. Findings from Boldogh lab shed some light in this context, where they demonstrated that increase in ROS generation from mitochondrial respiratory complex III augments Th2 responses independent of the adaptive immune response [49]. Further, they demonstrated that preexisting mitochondrial dysfunction in airway epithelia which was induced by deficiency of UQCRC2, ubiquinolcytochrome c reductase core II protein, worsened asthma features in ragweed pollen-induced allergic mice [20]. These data indicate that mitochondrial dysfunction may be an important component of asthma pathogenesis and that restoration of normal mitochondrial function may be an important target for improving asthma. In support of these findings from mouse studies, dysfunctional mitochondria have also been observed in human asthmatic bronchial epithelia [50].

# **Reactive Oxygen Species**

It is a well-known fact that mitochondria-mediated aerobic metabolism is, bioenergetically, at least 19 times more efficient than anaerobic glycolysis [51]. However, this energetically favorable system also introduces oxidative free radicals as unwanted by-products. Mitochondria are the major endogenous source of oxidative free radicals, and oxidative stress is tightly linked with airway inflammation. Additionally, asthmatic airway is bombarded with various proinflammatory mediators from various recruited inflammatory cells such as eosinophil, lymphocytes, macrophages, neutrophil, mast cells, etc. This leads to increased oxidative stress in the lung. For example, eosinophil secretes various toxic proteins such as eosinophil peroxidase, major basic protein, eosinophil cationic protein, and eosinophil-derived neurotoxin [52]. These various proinflammatory mediators create local oxidative milieu in the airway [53]. The fate of generated superoxide ( $O_2^-$ ) or free radicals is largely dependent on the local microenvironment. If there are sufficient dismutase enzymes inside the mitochondria, superoxide ( $O_2^-$ ) is converted to  $H_2O_2$ , which is relatively stable and can be further broken down by catalase. However, superoxide dismutase has been shown to be reduced in asthmatic airway [54, 55]. Under these conditions, unstable superoxide may directly damage the mitochondria. This is consistent with the reports of mitochondrial dysfunction in asthma [19, 20].

The molecular targets of mitochondria-generated superoxide ( $O_2^-$ ) are similar to any other parts of the cell such as nucleic acids, proteins, and lipids, similar to the NADPH oxidase-derived ROS. However, since mitochondria lack sophisticated DNA repair mechanisms, mitochondrial DNA mutations can accumulate [56]. Mitochondrial damage due to accumulated mitochondrial DNA mutations is an attractive theory of aging [57]. However, whether the reduction in the mitochondrial function due to accumulated mutations leads to aggravation of existing asthma features or induction of new asthma is yet to be investigated. To support this, various studies have found association of mitochondrial gene variations with asthma [35–42].

Oxidative damage to mitochondrial lipids such as cardiolipin, which is vulnerable due to its unsaturated state, leads to inhibition of the respiratory complexes [58, 59]. Oxidative stress also induces prooxidant enzymes such as 15-lipoxygenase (15-LOX), a lipid-peroxidizing enzyme, which is present in cytosol [60]. 15-LOX is unique since it directly hydrolyzes phospholipid esters present in biomembranes such as mitochondrial membrane even without the prior action of phospholipase A<sub>2</sub> (PLA<sub>2</sub>) [60]. A number of evidences show that 15-LOX may be the common denominator between asthma pathogenesis and mitochondrial dysfunction. 15-LOX is the main enzyme in degrading mitochondria in reticulocyte during RBC maturation, attacks mitochondria under oxidative stress conditions, and can lead to mitochondria-mediated apoptosis [59]. In the lungs, 15-LOX is inducible by important Th2 cytokines such as IL-4 and IL-13, which are sufficient to induce an asthmalike state. Further, 15-LOX is predominantly expressed in the bronchial epithelium, where mitochondrial dysfunction and apoptosis have been noted [60]. In mice, deficiency of 12/15-LOX has been shown to alleviate the features of asthma [61, 62]. Together, this suggests that IL-4- or IL-13-mediated increase in 15-LOX may cause epithelial mitochondrial dysfunction and injury, which in turn lead to asthma. However, neither the exact role of 15-LOX in mitochondrial degeneration nor the consequences of epithelial mitochondrial dysfunction that leads towards asthma pathogenesis are well understood.

#### **Reactive Nitrogen Species**

Nitric oxide synthase (NOS) generates NO from substrate L-arginine, a semiessential amino acid, and there are different isoforms of NOS: constitutive and inducible [63]. The role of NO in asthma is controversial and depends on its cellular source, isoform, and microenvironment [63, 64]. In normal airways, NO from constitutively expressed NOS such as eNOS (endothelial NOS) reduces airway tone by bronchodilation via cGMP pathway and increases mitochondrial biogenesis [63, 64]. In asthmatic airways, high levels of arginase consume most of the endogenous L-arginine and thus lead to reduced substrate availability to eNOS, inhibiting its bronchodilator action. Increased expression of iNOS during inflammation along with high levels of ROS leads to formation of reactive nitrogen species, peroxynitrite, which can cause numerous cytotoxic activities including mitochondrial dysfunction and is also a potent bronchoconstrictor [63]. NO and peroxynitrite inhibit cytochrome c oxidase (COX), a key oxidative enzyme of mitochondrial electron transport chain [65, 66]. Peroxynitrite not only affects cytochrome c oxidase but also affects many other mitochondrial enzymes such as NADH/coenzyme-Q reductase, aconitase, cytochrome c reductase, ATP synthase, and succinate dehydrogenase [67]. Thus, peroxynitrite is more effective to cause mitochondrial damage compared to NO. Thus, NO has a dual role in asthma pathophysiology as well as mitochondrial biology. In this context, therapeutic strategies for increasing eNOS and/or decreasing iNOS and peroxynitrite would be beneficial for both asthma and mitochondria. In this context, a high dose of L-arginine (250 mg/kg) reduces the formation of peroxynitrite by regulating NO and asymmetric dimethylarginine (ADMA) metabolism and alleviates the asthma features along with the reduction in mitochondrial dysfunction and epithelial damage [64, 68-70]. Since the existence of mitochondrial NOS is still under debate, we are not discussing that here [71].

Increased peroxynitrite formation in asthma can also be explained by the high levels of asymmetric dimethyl arginine (ADMA). ADMA binds to NOS, displacing L-arginine, and cannot be oxidized to NO, instead leading to formation of superoxide by uncoupling electron flow and NO production [69]. Although it has been suggested that ADMA can inhibit all forms of NOS, it predominantly inhibits eNOS in airway epithelia compared to inducible NOS [63, 68, 69]. The uncoupling of NOS leads to production of superoxide and peroxynitrite, which leads to mitochondrial dysfunction in pulmonary arterial endothelial cells [72]; however, the exact role of ADMA in asthma is yet to be explored. We have shown increased ADMA levels, along with increased expression of ADMA-synthesizing enzyme and decrease in the expression of ADMA-degrading enzyme, in bronchial epithelia of asthmatic lungs [69]. Importantly, ADMA levels were found to be increased in lung mitochondria in asthmatic mice [69]. Also, ADMA infusion leads to increased collagen deposition and deterioration of lung function in mice [73]. Further, ADMA increases the levels of both superoxide free radicals and peroxynitrite in bronchial epithelia (Fig. 3.4) [74].



**Fig. 3.4** Schematic diagram to show the dual role of nitric oxide (*NO*) in bronchial asthma and mitochondrial biology. Endogenous L-arginine is the common substrate for both eNOS (*endothelial NOS*) and iNOS (*inducible NOS*). NO derived from eNOS activates cGMP which dilate the airway; on the other hand cGMP also leads to mitochondrial biogenesis, whereas NO derived from iNOS reacts with superoxide free radicals to form peroxynitrite (ONOO<sup>-</sup>) which is the powerful bronchoconstrictor and also causes mitochondrial dysfunction. Increased ADMA leads to increase in the formation of superoxide free radicals and peroxynitrite

#### Apoptosis

Epithelial injury and shedding are increased in asthma. This may relate to increased mitochondria-mediated apoptosis in bronchial epithelia as oxidative stress in the lung could lead to cardiolipin peroxidation that dissociate cytochrome c from cardiolipin and release into cytosol where cytochrome c complexes with Apaf-1 and deoxy-ATP to form apoptosome which further activates caspase 9 and caspase 3 to execute apoptosis [75, 76]. In this context, increase of both caspase 9 and DNA fragmentation in asthmatic patients and OVA-induced mice were observed [77]. Further, the disturbance in the balance between apoptotic and antiapoptotic members of Bcl2 has been demonstrated in asthma [78]. It has been shown that exposure of bronchial epithelial cells to TNF-alpha and IFN-gamma leads to the damage of mitochondria and consequently epithelial injury [79]. Supportively, mucous cell apoptosis has been induced by IFN-gamma by BAX upregulation and its translocation into mitochondria [80], indicating that mitochondria-mediated apoptosis could be independent of Th1/Th2 types of inflammation. However, mitochondrialindependent pathways are also likely to be important in epithelial apoptosis. The mitochondrial-apoptotic pathway is also crucial for programmed death of



**Fig. 3.5** Schematic diagram to show the involvement of mitochondria at various stages of asthma pathophysiology. Mitochondrial dysfunction occurs in mast cells leading to generate ROS which degranulates mast cell to release IL-4 that converts naïve T cells to Th2 cells. These Th2 cells release various cytokines and recruit various inflammatory cells such as eosinophils, neutrophils, and lymphocytes. Recruited inflammatory cells release several proinflammatory mediators which injure airway epithelia. Epithelial injury activates epithelial mesenchymal trophic unit (EMTU) to initiate airway remodeling which consists of hyperplasia and hypertrophy of smooth muscle, goblet cell metaplasia, and subepithelial fibrosis. Asthmatic bronchial epithelia had shown the features of mitochondrial biogenesis

immune cells during resolution of asthma [81, 82]. Interestingly, glucocorticoids have been shown to have proapoptotic effect on human eosinophils via the mitochondrial pathway [83]. Translocation of Bax to the mitochondria, release of cytochrome c, caspase-independent collapse of the mitochondrial membrane, and subsequent activation of caspases are involved in spontaneous eosinophil apoptosis [84]. Failure of inducing eosinophil apoptosis by steroid could be one of the mechanisms of steroid resistance in severe asthmatic patients [85, 86]. This subgroup of asthmatic patients is difficult to treat and consumes a disproportionately large share of health resources [87]. Thus, investigating the role of mitochondria in steroid resistance is an important area of research (Fig. 3.5).

#### **Airway Remodeling**

The important sequelae of bronchial epithelial injury is airway remodeling [88–90]. Repeated allergen exposures lead to structural alterations in the airway, called airway remodeling. Airway epithelia have been postulated to contribute to a large extent to the remodeling process [91]. The airway epithelium in asthmatics is injured by inhaled pollutants, allergens, viruses, and also by endogenous factors such as proteolytic enzymes [89]. Injured epithelium activates epithelial mesenchymal trophic unit (EMTU) [90]. EMTU is active in the lung development at the embryonic

stage and it maintains the relationship between epithelia and mesenchymal cells with various growth factors such as TGF-beta, FGF, VEGF, etc. EMTU remains inactive in the adult lung and reactivated by epithelial injury, and its activation in asthmatic airway releases various growth factors which are responsible for various components of airway remodeling. Airway remodeling, a complex phenomenon, consists of alteration in the number and/or type of resident cells and the proportion/ quantitation of various components of extracellular matrix [90]. Airway remodeling includes epithelial damage, epithelial hypertrophy and hyperplasia, goblet cell metaplasia, airway myofibroblast transformation, increased airway collagen deposition, hyperplasia, and hypertrophy of airway smooth muscle and bronchial blood vessels [91, 92]. Among these features, hyperplasia and hypertrophy of airway smooth muscle and subepithelial fibrosis are crucial components. On the one hand, there is more of proline causing subepithelial fibrosis [93], and on the other hand, proline leads to inhibit cytochrome c oxidase [94]. Bronchial smooth muscle of asthmatics showed more number of mitochondria with increase in the factors required for mitochondrial biogenesis such as peroxisome proliferator-activated receptor-y coactivator, nuclear respiratory factor 1, and mitochondrial transcription factor [95].

It was further demonstrated that mitochondria are responsible for smooth muscle proliferation which emphasizes the importance of mitochondria in airway remodeling [95]. Also, mitochondrial biogenesis and increased mitochondrial activity have been reported in asthmatic airway smooth muscle along with altered calcium homeostasis, and it has been suggested that these are crucial in smooth muscle hypertrophy of airway remodeling [95]. However, mitochondrial dysfunction was not observed in those mitochondria from bronchial smooth muscle [95]. In contrast, our lab and others found mitochondrial dysfunction in bronchial epithelia [19, 20]. These indicate the possibility of differential roles of mitochondria in the lung. Thus, it seems there is a differential status of mitochondria in different cell types of asthmatic lungs, and thus careful interpretations are required to understand the precise role of mitochondria in asthma pathogenesis. This is very crucial in converting this knowledge into therapeutics. For example, theophylline causes cAMP-dependent tyrosine phosphorylation of subunit I of cytochrome c oxidase (COX) leading to its inhibition in smooth muscle cells [96]. If this is not cell-specific, it also could inhibit COX in bronchial epithelia and might create havoc in epithelial homeostasis. Thus, meticulous investigations are needed to take mitochondrial research in asthma from bench to the bedside in the form of mitochondrial medicine. It also has been suggested that both mitochondrial dysfunction and mitochondrial biogenesis are interrelated with each other as the latter feature is the possible compensatory mechanism of the former. In addition, it has been suggested that the dense localization of benzodiazepine receptor (BR) on the outer mitochondrial membrane of airway smooth muscle might have a role in asthmatic AHR [97]. Interestingly, airway smooth muscle is known to have high density of mitochondrial benzodiazepine receptor (MBR), a peripheral benzodiazepine receptor (PBR). There are two major types of BR, CBR (central type) and PBR. CBR is present exclusively in neurons, located at the plasma membrane, and regulates chloride channel intrinsic to the GABAA receptor, which is responsible for the antianxiety, anticonvulsant, and muscle relaxant effects of benzodiazepines [97].

PBR is distributed ubiquitously and predominantly located at the outer mitochondrial membrane; due to this reason PBR is generally called as MBR. The exact physiological role of PBR is not clear but has been postulated to relate to mitochondrial heme synthesis and steroid synthesis. Indeed, specific MBR ligands have been shown to affect the airway smooth muscle contraction in basal as well as pharmacologically stimulated conditions [97]. Also, these MBRs are densely localized in various structures in the lung other than airway smooth muscle, such as airway epithelium and submucosal glands. Further studies are needed to better define the potential role of MBR in the modulation of bronchial caliber and in the pathogenesis of asthma.

## **Calcium Homeostasis and Asthma**

It is well known that calcium plays an integrated role in asthma pathogenesis as most of the pathophysiological features such as airway smooth muscle contraction, mast cell degranulation, mucus secretion, recruitment of inflammatory cells, release of proinflammatory mediators from inflammatory cells, and vagal neurotransmission are dependent on calcium signaling. Indeed, calcium channel blockers have been used in asthmatic patients where beta adrenergic receptor agonists were contraindicated or ineffective [98]. Calcium signals also influence most of the functions of mitochondria such as oxidative phosphorylation, apoptosis, etc. [99]. Other than signaling, the mitochondrion acts as an intelligent calcium buffering system since they can sense and fine-tune intracellular calcium levels. Increased intracellular calcium leads to enhanced mitochondrial uptake of calcium via uniporters present in the inner mitochondrial membrane, and this uptake further regulates the calcium release from the plasma membrane and endoplasmic reticulum. Thus, mitochondria can act as buffering organelles in a spatiotemporal manner to maintain the calcium homeostasis inside the cell [100]. This mitochondrial calcium uptake leads to reduction of intracellular calcium, preventing excitotoxicity in cells. However, mitochondrial calcium overload itself may lead to mitochondrial damage and possibly cell death if there is significant damage caused to many mitochondria in a cell [101, 102]. Though calcium signaling is important for both mitochondrial physiology and asthma pathogenesis, detailed studies of this aspect are still lacking. It is to be noted that ORMDL3, one of the very few genes identified through genome-wide association study in asthma, binds and inhibits sarco-endoplasmic reticulum Ca2+ (SERCA) pump and alters endoplasmic reticulum-mediated calcium homeostasis [103].

#### Viral Infections and Asthma

Respiratory viral infection is one of the risk factors for asthma development in children [104]. Concurrent viral infections in asthma aggravate asthma features, cause acute exacerbations in chronic asthmatic conditions, and also lead to irreversible stage of airway remodeling [105]. Though it is very clear that viral infections caused by respiratory syncytial viruses, and few other viruses of the paramyxoviridae family, worsen airway inflammation and airway hyperresponsiveness, the mechanisms underlying this remain poorly understood. The bronchial epithelium is the major site of viral infections in the airway and participates in innate immune responses to viral infections [106]. Interestingly, mitochondria are suggested as surveillance sites for viruses due to the presence of antiviral proteins in mitochondria [107-109] such as MAVS (named for its *m*itochondrial *a*ntiviral signaling). Nucleic acids of various paramyxoviruses are recognized intracellularly by cytoplasmic receptors such as by RIG-I (retinoic acid-inducible gene I)/melanoma differentiation-associated protein 5 (MDA5) which further binds with MAVS present in outer mitochondrial membrane through CARD-CARD (caspase activation and recruitment *domains*) interactions [110, 111]. MAVS further recruits other molecules such as Tom70, and TNFreceptor-associated factors to assemble signalosome complex and activate IKK and TBK-1 complexes to signal both NF-κB and IRF3 signaling pathways to further initiate type I interferon responses to evade viral pathogens [110, 111]. In this signaling, mitochondrial localization of MAVS is very crucial as removal of MAVS from mitochondria evades the host immune system. Whether viral infection-induced mitochondrial stress triggers asthma related pathways, or mitochondrial dysfunction in asthmatic airway epithelium accentuates viral infection, remains to be studied.

## Therapeutics

The studies discussed above indicate strongly that mitochondrial dysfunction is an important aspect of asthma pathophysiology. It is therefore important to evolve strategies for mitochondrial protection as part of efforts to promote lung health and alleviate asthma. At this time, there are many gaps in the available knowledge, and further investigations are required to develop a sufficient understanding of the interface between mitochondrial biology and lung disease. Numerous efforts are being taken to bring mitochondrial bench research to bedside to control/cure various diseases [112–114]. As a result "mitochondrial medicine" was introduced with the recognition of new target sites in mitochondria. There are two major types of drug targets in mitochondria: direct or primary which acts specifically on mitochondrial enzymes and metabolic pathways and indirect or secondary which acts on mitochondria in nonspecific or accidental manner [115]. Possible reasons for secondary mechanisms are structural homology between mitochondria and other cellular parts, bacterial origin of mitochondria, functional or structural cross talk between mitochondria with other cellular compartments, etc. For example, the secondary effects of various antibiotics like chloramphenicol which affect protein synthesis of both bacteria and mitochondria may be explained by the bacterial origin of mitochondria [116]. The capacity of stem cells in donating mitochondria in rescuing injured airway epithelia in acute lung injury condition indicates another emerging target through the healthy mitochondrial transfer to damaged cells [117]. Though various reports demonstrated the therapeutic capacity of stem cells in asthma [118, 119], whether this is directly through the donation of mitochondria is yet to be demonstrated.

Our lab showed that mitochondrial dysfunction and mitochondrial structural changes, in asthma, were reversible with treatment of allergic mice with plantderived coumarins (esculetin), vitamin E, or statins [120–122]. Esculetin treatment to experimentally allergic mice reduced asthma features such as airway hyperresponsiveness, airway inflammation, Th2 response, eotaxin, and airway eosinophilia along with restoration of mitochondrial function and structure in bronchial epithelia. It is important to note that both esculetin and vitamin E are not only antioxidants but they reduced the expression and activity of reticulocyte lipoxygenase in the lung. So we believe that esculetin might restore mitochondrial function through its primary cytosolic target, reticulocyte lipoxygenase. Coenzyme Q and vitamin E are mitochondria-targeted antioxidants. Notably, coenzyme Q10 is known to reduce the requirement of corticosteroids in asthmatic patients [21]. Further, using experimental mouse models, we have shown that L-arginine and statins alleviate asthma features and improve mitochondrial function in lungs, possibly due to restoration of homeostatic nitric oxide metabolism in bronchial epithelia [69, 122]. However, each of these observations is limited by inability to distinguish between primary and secondary effects. Thus, further efforts have to be taken to develop "mitochondrial medicine" for effective management of chronic respiratory diseases.

# References

- 1. Ernster L, Schatz G. Mitochondria: a historical review. J Cell Biol. 1981;91:227-55.
- 2. Nunnari J, Suomalainen A. Mitochondria: in sickness and in health. Cell. 2012;148: 1145–59.
- Chacinska A, Koehler CM, Milenkovic D, Lithgow T, Pfanner N. Importing mitochondrial proteins: machineries and mechanisms. Cell. 2009;138:628–44.
- 4. Kroemer G, Galluzzi L, Brenner C. Mitochondrial membrane permeabilization in cell death. Physiol Rev. 2007;87:99–163.
- Crow MT, Mani K, Nam YJ, Kitsis RN. The mitochondrial death pathway and cardiac myocyte apoptosis. Circ Res. 2004;95:957–70.
- Tal MC, Iwasaki A. Mitoxosome: a mitochondrial platform for cross-talk between cellular stress and antiviral signaling. Immunol Rev. 2011;243:215–34.
- Masoli M, Fabian D, Holt S, Beasley R. Global Initiative for Asthma (GINA) program: the global burden of asthma: executive summary of the GINA Dissemination Committee report. Allergy. 2004;59:469–78.
- 8. Braman SS. The global burden of asthma. Chest. 2006;130:4S-12.
- King CS, Moores LK. Clinical asthma syndromes and important asthma mimics. Respir Care. 2008;53:568–80.
- 10. The Global Asthma Report 2011. Paris: The International Union Against Tuberculosis and Lung Disease; 2011.
- 11. Agrawal A, Mabalirajan U, Ahmad T, Ghosh B. Emerging interface between metabolic syndrome and asthma. Am J Respir Cell Mol Biol. 2011;44:270–5. Review.
- Fahy JV. Eosinophilic and neutrophilic inflammation in asthma: insights from clinical studies. Proc Am Thorac Soc. 2009;6:256–9.
- 13. Monteseirín J. Neutrophils and asthma. J Investig Allergol Clin Immunol. 2009;19:340-54.

- Wang W, Li JJ, Foster PS, Hansbro PM, Yang M. Potential therapeutic targets for steroidresistant asthma. Curr Drug Targets. 2010;11:957–70.
- Tattersfield AE. The site of the defect in asthma. Neurohumoral, mediator or smooth muscle? Chest. 1987;91:184S–9.
- 16. Hirst SJ, Lee TH. Airway smooth muscle as a target of glucocorticoid action in the treatment of asthma. Am J Respir Crit Care Med. 1998;158:S201–6.
- 17. Konradova V, Copova C, Sukova B, Houstek J. Ultrastructure of the bronchial epithelium in three children with asthma. Pediatr Pulmonol. 1985;1:182–7.
- Hayashi T, Ishii A, Nakai S, Hasegawa K. Ultrastructure of goblet-cell metaplasia from Clara cell in the allergic asthmatic airway inflammation in a mouse model of asthma in vivo. Virchows Arch. 2004;444:66–73.
- Mabalirajan U, Dinda AK, Kumar S, Roshan R, Gupta P, Sharma SK, Ghosh B. Mitochondrial structural changes and dysfunction are associated with experimental allergic asthma. J Immunol. 2008;181:3540–8.
- Aguilera-Aguirre L, Bacsi A, Saavedra-Molina A, Kurosky A, Sur S, Boldogh I. Mitochondrial dysfunction increases allergic airway inflammation. J Immunol. 2009;183:5379–87.
- Gvozdjáková A, Kucharská J, Bartkovjaková M, Gazdíková K, Gazdík FE. Coenzyme Q10 supplementation reduces corticosteroids dosage in patients with bronchial asthma. Biofactors. 2005;25:235–40.
- Fogarty A, Lewis S, Weiss S, Britton J. Dietary vitamin E, IgE concentration, and atopy. Lancet. 2000;356:1573–4.
- Blesa S, Cortijo J, Mata M, Serrano A, Closa D, Santangelo F, et al. Oral N-acetylcysteine attenuates the rat pulmonary inflammatory response to antigen. Eur Respir J. 2003;21: 394–400.
- Gredilla R. DNA damage and base excision repair in mitochondria and their role in aging. J Aging Res. 2010;2011:257093.
- Taylor RW, Turnbull DM. Mitochondrial DNA mutations in human disease. Nat Rev Genet. 2005;6:389–402.
- Ghosh B, Batra J, Sharma S, Kumar A, Sharma M, Chatterjee R, Mabalirajan U. Genetic components of asthma: current status and future goals. Int Rev Asthma (Jpn). 2006;8:67–88.
- 27. Kumar A, Ghosh B. Genetics of asthma: a molecular biologist perspective. Clin Mol Allergy. 2009;7:7.
- Ober C, Hoffjan S. Asthma genetics 2006: the long and winding road to gene discovery. Genes Immun. 2006;7:95–100.
- 29. Oliveti JF, Kercsmar CM, Redline S. Pre- and perinatal risk factors for asthma in inner city African-American children. Am J Epidemiol. 1996;143:570–7.
- Litonjua AA, Carey VJ, Burge HA, Weiss ST, Gold DR. Parental history and the risk for childhood asthma. Does mother confer more risk than father? Am J Respir Crit Care Med. 1998;158:176–81.
- Soto-Quiros ME, Silverman EK, Hanson LA, Weiss ST, Celedon JC. Maternal history, sensitization to allergens, and current wheezing, rhinitis, and eczema among children in Costa Rica. Pediatr Pulmonol. 2002;33:237–43.
- 32. Kurukulaaratchy RJ, Matthews S, Arshad SH. Relationship between childhood atopy and wheeze: what mediates wheezing in atopic phenotypes? Ann Allergy Asthma Immunol. 2006;97:84–91.
- Hamada K, Suzaki Y, Goldman A, Ning YY, Goldsmith C, Palecanda A, et al. Allergenindependent maternal transmission of asthma susceptibility. J Immunol. 2003;170:1683–9.
- 34. Liu CA, Wang CL, Chuang H, Ou CY, Hsu TY, Yang KD. Prediction of elevated cord blood IgE levels by maternal IgE levels, and the neonate's gender and gestational age. Chang Gung Med J. 2003;26:561–9.
- Raby BA, Klanderman B, Murphy A, Mazza S, Camargo Jr CA, Silverman EK, et al. A common mitochondrial haplogroup is associated with elevated total serum IgE levels. J Allergy Clin Immunol. 2007;120:351–8.

- Clifton VL, Davies M, Moore V, Wright IM, Ali Z, Hodyl NA. Developmental perturbation induced by maternal asthma during pregnancy: the short- and long-term impacts on offspring. J Pregnancy. 2012;2012:741613. Epub 2012 Jul 8.
- Zifa E, Daniil Z, Skoumi E, Stavrou M, Papadimitriou K, Terzenidou M, et al. Mitochondrial genetic background plays a role in increasing risk to asthma. Mol Biol Rep. 2012;39: 4697–708.
- Schauberger EM, Ewart SL, Arshad SH, Huebner M, Karmaus W, Holloway JW, et al. Identification of ATPAF1 as a novel candidate gene for asthma in children. J Allergy Clin Immunol. 2011;128:753–60.
- Fukuda T, Mochida S, Fukushima Y, Makino S. Detection of allergen-induced genes in peripheral blood mononuclear cells of patients with allergic asthma using subtractive hybridization. J Allergy Clin Immunol. 1995;96:1076–82.
- Polonikov AV, Ivanov VP, Solodilova MA, Kozhukhov MA, Panfilov VI, Bulgakova VI. Polymorphism -930A > G of the cytochrome b gene is a novel genetic marker of predisposition to bronchial asthma. Ter Arkh. 2009;81:31–5.
- Schmuczerova J, Brdicka R, Dostal M, Sram RJ, Topinka J. Genetic variability of HVRII mtDNA in cord blood and respiratory morbidity in children. Mutat Res. 2009;666:1–7.
- 42. Jones M, Mitchell P, Wang JJ, Sue C. MELAS A3243G mitochondrial DNA mutation and age related maculopathy. Am J Ophthalmol. 2004;138:1051–3.
- Shanske AL, Shanske S, Silvestri G, Tanji K, Wertheim D, Lipper S. MELAS point mutation with unusual clinical presentation. Neuromuscul Disord. 1993;3:191–3.
- 44. Finsterer J. Genetic, pathogenetic, and phenotypic implications of the mitochondrial A3243G tRNALeu(UUR)mutation. Acta Neurol Scand. 2007;116:1–14.
- 45. Marchetti P, Castedo M, Susin SA, Zamzami N, Hirsch T, Macho A, et al. Mitochondrial permeability transition is a central coordinating event of apoptosis. J Exp Med. 1996;184: 1155–60.
- 46. Elias JA, Zhu Z, Chupp G, Homer RJ. Airway remodeling in asthma. J Clin Invest. 1999;104:1001-6.
- Elias JA, Lee CG, Zheng T, Ma B, Homer RJ, Zhu Z. New insights into the pathogenesis of asthma. J Clin Invest. 2003;111:291–7.
- Konga DB, Kim Y, Hong SC, Roh YM, Lee CM, Kim KY, et al. Oxidative stress and antioxidant defenses in asthmatic murine model exposed to printer emissions and environmental tobacco smoke. J Environ Pathol Toxicol Oncol. 2009;28:325–40.
- 49. Chodaczek G, Bacsi A, Dharajiya N, Sur S, Hazra TK, Boldogh I. Ragweed pollen-mediated IgE-independent release of biogenic amines from mast cells via induction of mitochondrial dysfunction. Mol Immunol. 2009;46:2505–14.
- Xu W, Comhair SAA, Janocha AJ, Mavrakis LA, Erzurum SC. Alteration of nitric oxide synthesis related to abnormal cellular bioenergetics in asthmatic airway epithelium. Am J Respir Crit Care Med. 2010;181:A1436.
- Ramakrishna R, Edwards JS, McCulloch A, Palsson BO. Flux balance analysis of mitochondrial energy metabolism: consequences of systemic stoichiometric constraints. Am J Physiol Regul Integr Comp Physiol. 2001;280:R695–704.
- 52. Weller PF, Ackerman SJ, Smith JA. Eosinophil granule cationic proteins: major basic protein is distinct from the smaller subunit of eosinophil peroxidase. Leukoc Biol. 1988;43:1–4.
- Wood LG, Gibson PG, Garg ML. Biomarkers of lipid peroxidation, airway inflammation and asthma. Eur Respir J. 2003;21:177–86.
- Macmillan-Crow LA, Jahangir DL. Invited review: manganese superoxide dismutase in disease. Free Radic Res. 2001;34:325–36.
- 55. Comhair SA, Ricci KS, Arroliga M, Lara AR, Dweik RA, Song W, et al. Correlation of systemic superoxide dismutase deficiency to airflow obstruction in asthma. Am J Respir Crit Care Med. 2005;172:306–13.
- Larsen NB, Rasmussen M, Rasmussen LJ. Nuclear and mitochondrial DNA repair: similar pathways? Mitochondrion. 2005;5:89–108.

- 57. Ames BN, Shigenaga MK, Hagen TM. Mitochondrial decay in aging. Biochim Biophys Acta. 1995;1271:165–70.
- 58. Schewe T. 15-lipoxygenase-1: a prooxidant enzyme. Biol Chem. 2002;383:365-74.
- Géminard C, de Gassart A, Vidal M. Reticulocyte maturation: mitoptosis and exosome release. Biocell. 2002;26:205–15.
- Chanez P, Bonnans C, Chavis C, Vachier I. 15-lipoxygenase: a Janus enzyme? Am J Respir Cell Mol Biol. 2002;27:655–8.
- Andersson CK, Claesson HE, Rydell-Törmänen K, Swedmark S, Hällgren A, Erjefält JS. Mice lacking 12/15-lipoxygenase have attenuated airway allergic inflammation and remodeling. Am J Respir Cell Mol Biol. 2008;39:648–56.
- 62. Hajek AR, Lindley AR, Favoreto Jr S, Carter R, Schleimer RP, Kuperman DA. 12/15-Lipoxygenase deficiency protects mice from allergic airways inflammation and increases secretory IgA levels. J Allergy Clin Immunol. 2008;122:633–9.
- 63. Ricciardolo FL. cNOS-iNOS paradigm and arginase in asthma. Trends Pharmacol Sci. 2003;24:560–1.
- 64. Mabalirajan U, Ahmad T, Leishangthem GD, Joseph DA, Dinda AK, Agrawal A, et al. Beneficial effects of high dose of L-Arginine on airway hyperresponsiveness and airway inflammation in a murine model of asthma. J Allergy Clin Immunol. 2010;125:626–35.
- 65. Shiva S, Crawford JH, Ramachandran A, Ceaser EK, Hillson T, Brookes PS, et al. Mechanisms of the interaction of nitroxyl with mitochondria. Biochem J. 2004;379:359–66.
- 66. Capaldi RA. Structure and function of cytochrome oxidase. Annu Rev Biochem. 1992;59: 569–96.
- 67. Szabó C, Ischiropoulos H, Radi R. Peroxynitrite: biochemistry, pathophysiology and development of therapeutics. Nat Rev Drug Discov. 2007;6:662–80.
- Ahmad T, Mabalirajan U, Ghosh B, Agrawal A. Altered assymetric dimethyl arginine metabolism in bronchial epithelium of allergically inflamed lungs. Am J Resp cell Mol Biol. 2010;42:3–8.
- 69. Mabalirajan U, Ahmad T, Leishangthem G, Dinda AK, Agrawal A, Ghosh B. L-arginine reduces mitochondrial dysfunction and epithelial injury in murine allergic airway inflammation. Int Immunopharmacol. 2010;10:1514–9.
- Scott JA, North ML, Rafii M, Huang H, Pencharz P, Subbarao P, Belik J, Grasemann H. Asymmetric dimethylarginine is increased in asthma. Am J Respir Crit Care Med. 2011;184:779–85.
- 71. Lacza Z, Pankotai E, Csordás A, Gero D, Kiss L, Horváth EM, et al. Mitochondrial NO and reactive nitrogen species production: does mtNOS exist? Nitric Oxide. 2006;14:162–8.
- Sud N, Wells SM, Sharma S, Wiseman DA, Wilham J, Black SM. Asymmetric dimethylarginine inhibits HSP90 activity in pulmonary arterial endothelial cells: role of mitochondrial dysfunction. Am J Physiol Cell Physiol. 2008;294:1407–18.
- Wells SM, Buford MC, Migliaccio CT, Holian A. Elevated asymmetric dimethylarginine alters lung function and induces collagen deposition in mice. Am J Respir Cell Mol Biol. 2009;40:179–88.
- 74. Wells SM, Holian A. Asymmetric dimethylarginine induces oxidative and nitrosative stress in murine lung epithelial cells. Am J Respir Cell Mol Biol. 2007;36:520–8.
- Robinson NC. Functional binding of cardiolipin to cytochrome c oxidase. J Bioenerg Biomembr. 1993;25:153–63.
- Ott M, Zhivotovsky B, Orrenius S. Role of cardiolipin in cytochrome c release from mitochondria. Cell Death Differ. 2007;14:1243–7.
- 77. Comhair SA, Xu W, Ghosh S, Thunnissen FB, Almasan A, Calhoun WJ, et al. Superoxide dismutase inactivation in pathophysiology of asthmatic airway remodeling and reactivity. Am J Pathol. 2005;166:663–74.
- Grigoraş CC, Grigoraş A, Mihăescu T, Floarea-Stra A, Cozma L, Vereş L. Expression of the Bax proapoptotic factor in asthmatic patients. Pneumologia. 2009;58:8–12.
- 79. Kampf C, Relova AJ, Sandler S, Roomans GM. Effects of TNF-alpha, IFN-gamma and IL-beta on normal human bronchial epithelial cells. Eur Respir J. 1999;14:84–91.

- Shi ZQ, Feng Y, Hou YK, Liu T, Xiu QY. A study of interferon-gamma induced airway mucous cell apoptosis and its mechanisms. Zhonghua Jie He Hu Xi Za Zhi. 2005;28:160–3.
- Chang KC, Lo CW, Fan TC, Chang MD, Shu CW, Chang CH, et al. TNF-alpha mediates eosinophil cationic protein-induced apoptosis in BEAS-2B cells. BMC Cell Biol. 2010;11:6.
- Serradell MC, Guasconi L, Masih DT. Involvement of a mitochondrial pathway and key role of hydrogen peroxide during eosinophil apoptosis induced by excretory-secretory products from Fasciola hepatica. Mol Biochem Parasitol. 2009;163:95–106.
- Gardai SJ, Hoontrakoon R, Goddard CD, Day BJ, Chang LY, Henson PM, et al. Oxidantmediated mitochondrial injury in eosinophil apoptosis: enhancement by glucocorticoids and inhibition by granulocyte-macrophage colony-stimulating factor. J Immunol. 2003;170: 556–66.
- Dewson G, Cohen GM, Wardlaw AJ. Interleukin-5 inhibits translocation of Bax to the mitochondria, cytochrome c release, and activation of caspases in human eosinophils. Blood. 2001;98:2239–47.
- Druilhe A, Létuvé S, Pretolani M. Glucocorticoid-induced apoptosis in human eosinophils: mechanisms of action. Apoptosis. 2003;8:481–95.
- Walsh GM, Sexton DW, Blaylock MG. Corticosteroids, eosinophils and bronchial epithelial cells: new insights into the resolution of inflammation in asthma. J Endocrinol. 2003;178(1): 37–43.
- Cruz AA, Bousquet PJ. The unbearable cost of severe asthma in underprivileged populations. Allergy. 2009;64(3):319–21.
- Holgate ST, Lackie P, Wilson S, Roche W, Davies D. Bronchial epithelium as a key regulator of airway allergen sensitization and remodeling in asthma. Am J Respir Crit Care Med. 2000;162:S113–7.
- O'Sullivan MP, Tyner JW, Holtzman MJ. Apoptosis in the airways: another balancing act in the epithelial program. Am J Respir Cell Mol Biol. 2003;29:3–7.
- Holgate ST. The inflammation-repair cycle in asthma: the pivotal role of the airway epithelium. Clin Exp Allergy. 1998;28 Suppl 5:97–103.
- Holgate ST, Holloway J, Wilson S, Bucchieri F, Puddicombe S, Davies DE. Epithelialmesenchymal communication in the pathogenesis of chronic asthma. Proc Am Thorac Soc. 2004;1:93–8.
- Davies DE, Holgate ST. Asthma: the importance of epithelial mesenchymal communication in pathogenesis. Inflammation and the airway epithelium in asthma. Int J Biochem Cell Biol. 2002;34:1520–6.
- Zimmermann N, King NE, Laporte J, Yang M, Mishra A, Pope SM, et al. Dissection of experimental asthma with DNA microarray analysis identifies arginase in asthma pathogenesis. J Clin Invest. 2003;111:1863–74.
- Delwing D, Delwing D, Chiarani F, Kurek AG, Wyse AT. Proline reduces brain cytochrome c oxidase: prevention by antioxidants. Int J Dev Neurosci. 2007;25:17–22.
- Trian T, Benard G, Begueret H, Rossignol R, Girodet PO, Ghosh D, et al. Bronchial smooth muscle remodeling involves calcium-dependent enhanced mitochondrial biogenesis in asthma. J Exp Med. 2007;204:3173–81.
- Lee I, Salomon AR, Ficarro S, Mathes I, Lottspeich F, Grossman LI, et al. cAMP-dependent tyrosine phosphorylation of subunit I inhibits cytochrome c oxidase activity. J Biol Chem. 2005;280:6094–100.
- Pelaia G, Di Paola ED, De Sarro G, Marsico SA. Is the mitochondrial benzodiazepine receptor involved in the control of airway smooth muscle tone? Gen Pharmacol. 1997;28:495–8.
- Ten Broeke R, Blalock JE, Nijkamp FP, Folkerts G. Calcium sensors as new therapeutic targets for asthma and chronic obstructive pulmonary disease. Clin Exp Allergy. 2004;34: 170–6.
- Duchen MR. Contributions of mitochondria to animal physiology: from homeostatic sensor to calcium signalling and cell death. J Physiol. 1999;516:1–17.

- 100. Herrington J, Park YB, Babcock DF, Hille B. Dominant role of mitochondria in clearance of large Ca<sup>2+</sup> loads from rat adrenal chromaffin cells. Neuron. 1996;16:219–28.
- Peng TI, Jou MJ. Oxidative stress caused by mitochondrial calcium overload. Ann N Y Acad Sci. 2010;1201:183–8.
- 102. Feissner RF, Skalska J, Gaum WE, Sheu SS. Crosstalk signaling between mitochondrial Ca2+ and ROS. Front Biosci. 2009;14:1197–218.
- 103. Cantero-Recasens G, Fandos C, Rubio-Moscardo F, Valverde MA, Vicente R. The asthmaassociated ORMDL3 gene product regulates endoplasmic reticulum-mediated calcium signaling and cellular stress. Hum Mol Genet. 2010;19:111–21.
- Folkerts G, Busse WW, Nijkamp FP, Sorkness R, Gern JE. Virus induced airway hyperresponsiveness and asthma. Am J Respir Crit Care Med. 1998;157:1708–20.
- 105. You D, Becnel D, Wang K, Ripple M, Daly M, Cormier SA. Exposure of neonates to respiratory syncytial virus is critical in determining subsequent airway response in adults. Respir Res. 2006;7:107.
- 106. Punyadarsaniya D, Liang CH, Winter C, Petersen H, Rautenschlein S, Hennig-Pauka I, et al. Infection of differentiated porcine airway epithelial cells by influenza virus: differential susceptibility to infection by porcine and avian viruses. PLoS One. 2011;6:e28429.
- 107. Seth RB, Sun L, Ea CK, Chen ZJ. Identification and characterization of MAVS, a mitochondrial antiviral signaling protein that activates NF-kappaB and IRF 3. Cell. 2005;122:669–82.
- Qi B, Huang Y, Rowe D, Halliday G. VISA–a pass to innate immunity. Int J Biochem Cell Biol. 2007;39:287–91.
- Tang ED, Wang CY. MAVS self-association mediates antiviral innate immune signaling. J Virol. 2009;83:3420–8.
- 110. Seth RB, Sun L, Chen ZJ. Antiviral innate immunity pathways. Cell Res. 2006;16:141-7.
- 111. Rehwinkel J, Reis e Sousa C. RIGorous detection: exposing virus through RNA sensing. Science. 2010;327:284–6.
- 112. Land JM, Morgan-Hughes JA, Hargreaves I, Heales SJ. Mitochondrial disease: a historical, biochemical, and London perspective. Neurochem Res. 2004;29:483–91.
- Szeto HH. Mitochondria-targeted peptide antioxidants: novel neuroprotective agents. AAPS J. 2006;8:521–31.
- 114. Armstrong JS. Mitochondrial medicine: pharmacological targeting of mitochondria in disease. Br J Pharmacol. 2007;151:1154–65.
- 115. Szewczyk A, Wojtczak L. Mitochondria as a pharmacological target. Pharmacol Rev. 2002;54:101–27.
- Barnhill AE, Brewer MT, Carlson SA. Adverse effects of antimicrobials via predictable or idiosyncratic inhibition of host mitochondrial components. Antimicrob Agents Chemother. 2012;56:4046–51.
- 117. Islam MN, Das SR, Emin MT, Wei M, Sun L, Westphalen K, et al. Mitochondrial transfer from bone-marrow derived stromal cells to pulmonary alveoli protects against acute lung injury. Nat Med. 2012;18:759–65.
- 118. Jones CP, Rankin SM. Bone marrow-derived stem cells and respiratory disease. Chest. 2011;140:205–11.
- 119. Kapoor S, Patel SA, Kartan S, Axelrod D, Capitle E, Rameshwar P. Tolerance-like mediated suppression by mesenchymal stem cells in patients with dust mite allergy-induced asthma. J Allergy Clin Immunol. 2012;129:1094–101.
- Mabalirajan U, Dinda AK, Sharma SK, Ghosh B. Esculetin restores mitochondrial dysfunction and reduces allergic asthma features in experimental murine model. J Immunol. 2009;183:2059–67.
- 121. Mabalirajan U, Aich J, Sharma SK, Ghosh B. Effects of vitamin E on mitochondrial dysfunction and asthma features in an experimental allergic murine model. J Appl Physiol. 2009;107:1285–92.
- 122. Ahmad T, Mabalirajan U, Sharma A, Ghosh B, Agrawal A. Simvastatin improves epithelial dysfunction and airway hyperresponsiveness: from ADMA to asthma. Am J Resp Cell Mol Biol. 2011;44:531–9.