

Chapter 12

Implications of Oxygen Homeostasis for Tumor Biology and Treatment

Boyan K. Garvalov and Till Acker

Abstract Tumors serve as a prototype system to study the role of the hypoxic microenvironment and gain insight in the regulation of oxygen homeostasis. A series of biochemical and cell biological studies have significantly extended our knowledge of how tumor cells activate key regulatory mechanisms of oxygen homeostasis not only to adapt to the hostile tumor microenvironment but also to acquire a more aggressive tumor phenotype. Reduced oxygen levels and tumor-specific genetic alterations synergistically drive tumor progression by activating a key transcriptional system, the hypoxia-inducible factors (HIFs). HIFs trigger a set of adaptive responses commonly associated with tumor malignancy including tumor angiogenesis, a shift in metabolism, proliferation, invasion, and metastasis. We and others could demonstrate that cancer stem cells are controlled by HIFs within a hypoxic niche, establishing an intriguing link between the well-known function of hypoxia in tumor growth and stem cell biology. Additionally, HIF activation potentially conveys resistance to current tumor therapies including the evasive resistance phenotype observed after anti-angiogenic treatment. Together, these findings provide strong evidence that activation of the HIF system is a decisive step in cancer progression that critically shapes therapy response and clinical outcome. Recent insight into the precise mechanisms of oxygen sensing and signalling has offered new promising and potentially selective strategies to counteract this crucial pathway.

Keywords Anti-angiogenesis • Cancer stem cells (CSCs) • Epithelial mesenchymal transition (EMT) • Glioblastoma • Hypoxia • Hypoxia-inducible factor (HIF) • Prolyl hydroxylase (PHD) • Tumor therapy

B.K. Garvalov • T. Acker (✉)
Institute of Neuropathology, Justus Liebig University, Giessen 35392, Germany
e-mail: till.acker@patho.med.uni-giessen.de

12.1 Introduction

Oxygen is an indispensable substrate for aerobic metabolism and is therefore essential for the normal development and functioning of higher organisms. The efficient distribution of oxygen to tissues is a main function of the respiratory and cardiovascular systems. However, oxygen availability can vary greatly among individual cells and organs or during specific stages of development. While the arterial oxygen partial pressure is 90 mmHg (corresponding to ~12.5 %) under physiological conditions, differences in vascularization, tissue diffusion properties, and cell-specific oxygen consumption create a heterogeneous O₂ distribution, so that most tissues are exposed to lower oxygen concentrations. In particular, the brain experiences low O₂ levels down to 1 mmHg [24]. Reduced levels of oxygen (hypoxia) activate a set of adaptive responses which either enhance oxygen delivery or decrease oxygen consumption to promote survival under low oxygen conditions. Importantly, hypoxia arises not only in physiological situations, but is also a characteristic feature of various pathological conditions [1]. Particularly prominent among those is the process of neoplastic transformation and progression. Tumor growth and progression occurs as a result of the cumulative acquisition of genetic and epigenetic alterations in individual cells, followed by the selection of tumor cell clones with enhanced proliferation and survival potential. Once a tumor is formed, it creates a specialized microenvironment, which critically controls tumor progression. Highly proliferating tumors frequently outstrip their vascular supply leading to a tumor microenvironment characterized by low oxygen tension, low glucose levels, and an acidic pH. Tumor hypoxia is associated with an increased frequency of tumor invasion and metastasis and a poor therapy outcome. Notably, tumor cells not only adapt to survive under low oxygen, but also exploit hypoxia-induced mechanisms in order to promote their own growth and dissemination. Indeed, tumor hypoxia has become one of the main settings to study the mechanisms and functions of hypoxic signaling. Here, we will briefly summarize the mechanisms of cellular oxygen homeostasis and will focus on the function of hypoxic signaling in various aspects of cancer progression and resistance, as well as on possible strategies to target the hypoxic response as an anti-tumor therapy.

12.2 The Hypoxic Response and HIF

Since their discovery in 1995 [105], the hypoxia inducible transcription factors (HIFs) have emerged as the key transcriptional system initiating adaptive responses to hypoxia. HIFs act as heterodimers composed of a shared, stable HIF β subunit and specific oxygen-regulated HIF α subunits. The stability of the α subunits is mainly controlled by prolyl hydroxylase domain proteins (PHDs), which use O₂ as a substrate to hydroxylate HIF α and target it for proteasomal degradation [15, 23, 62], following ubiquitination by the E3 ubiquitin ligase pVHL [48, 70]. An additional level of oxygen-dependent control is conferred by another hydroxylase, factor

inhibiting HIF-1 α (FIH1), which modifies asparaginyl residues and inhibits the interaction between HIF α and its transcriptional coactivators p300/CBP [51]. More recently a number of additional mechanisms that modulate the HIF pathway have been identified, including the heat shock proteins HSP90 and HSP70, the histone deacetylases Sirt1 and Sirt6, TCA cycle-related metabolites, nitric oxide, the PHD E3 ubiquitin ligases Siah1/2, the microRNAs miR-17-92, and miR-107, as well as a number of oncogenes or tumor suppressor genes, e.g. PI3K/Akt, mTOR, Ras, p53 (reviewed in [69]). This high level of complexity of HIF regulation, which often involves an elaborate set of negative and positive feedback mechanisms [41], allows for precise, fine-tuned control of hypoxia-mediated responses and highlights the central importance of HIF signaling in cellular homeostasis [3]. In addition, hypoxia initiates adaptive responses independent of HIF through other redox sensitive systems, including activation of the NF- κ B pathway, or global protein synthesis inhibition through the AMPK/mTOR or PERK/eIF2 α pathways [11, 100], although the mechanisms mediating the O₂ dependence of these processes are less well understood.

There are two principal HIF α subunits, HIF-1 α and HIF-2 α , which mediate the hypoxic response through transcriptional regulation of an ever-growing number of genes. Although the functions of HIF-1 α and HIF-2 α partially overlap, it is now clear that the two isoforms are differently regulated by oxygen, are expressed in distinct normal and neoplastic cell types, possess different target specificities and generally appear to have complementary rather than redundant functions, as described in more detail in the following sections.

12.3 The Hypoxic Response and the Hallmarks of Cancer

Hypoxic signaling activates a large number of downstream biological responses, which together promote most of the defining properties of tumors (Fig. 12.1.) [4, 11]. One of the primary effects of hypoxia is the induction of a shift in cellular metabolism from oxidative phosphorylation to anaerobic glycolysis via HIF-mediated upregulation of glucose transporters and glycolytic enzymes [60]. This is accompanied by the upregulation of carbonic anhydrase IX (CA IX), transporters for lactate, H⁺, HCO₃⁻ and other ions, as an adaptive mechanism of pH homeostasis, leading to a net acidification of the extracellular tumor environment. A decreased pH is a characteristic feature of many tumor types and has been shown to promote tumor growth and metastatic spread, at least in part through activation of extracellular matrix-degrading enzymes [18]. Another key response to tumor hypoxia is the stimulation of angiogenesis in order to improve the blood and oxygen supply of tumor cells. It is well established that tumors induce the formation of new vasculature as a key event in multistage carcinogenesis, a phenomenon termed “angiogenic switch” [8]. This is mainly accomplished through the upregulation of multiple angiogenic genes as direct HIF targets, including components of the VEGF, angiopoietin, and SDF-1 pathways [85].

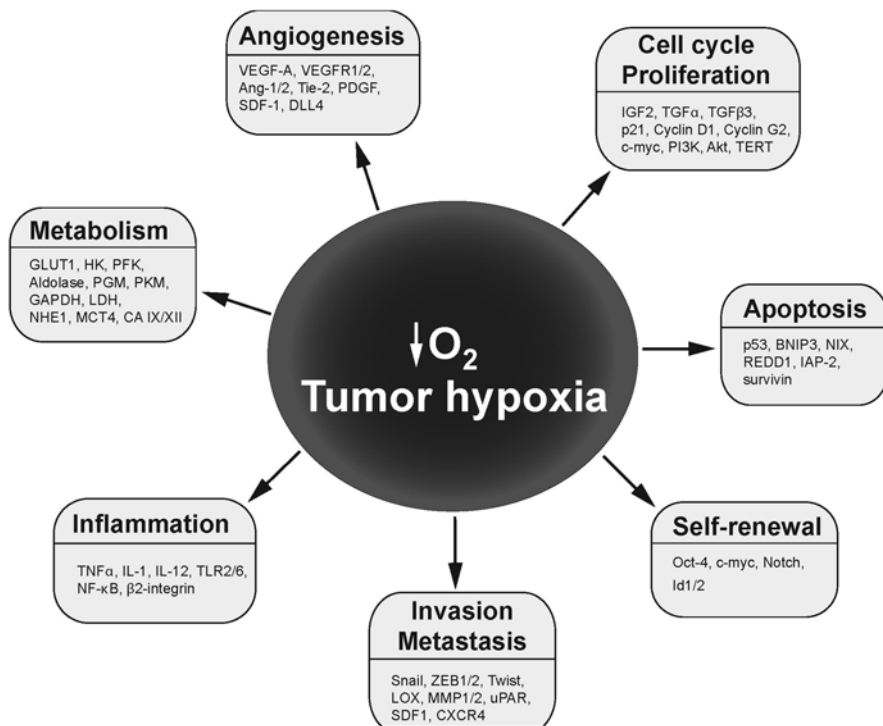


Fig. 12.1 Major aspects of tumor biology regulated by hypoxia. HIFs regulate multiple cellular processes relevant to tumor progression in response to hypoxia through transcriptional upregulation of HIF-1 α and/or HIF-2 α target genes. In some cases, however, alternative mechanisms are involved, such as HIF-mediated protein stabilization (p53) or modulation of transcriptional activity (c-myc, Notch). *Ang-1/2* angiopoietin 1/2, *BNIP3* BCL2/adenovirus E1B 19 kDa interacting protein 3, *CA IX/XII* carbonic anhydrase IX/XII, *CXCR4* C-X-C chemokine receptor type 4, *DLL4* delta-like protein 4, *GAPDH* glyceraldehyde-3-phosphate-dehydrogenase, *GLUT1* glucose transporter 1, *HK* hexokinase, *IAP-2* inhibitor of apoptosis protein 2, *ID-1/2* inhibitor of DNA binding 1/2, *IGF2* insulin-like growth factor 2, *IL-1/12* interleukin 1/12, *LDH* lactate dehydrogenase, *LOX* lysyl oxidase, *MCT4* monocarboxylate transporter 4, *MMP1/2* matrix metalloproteinase 1/2, *NHE1* Na⁺/H⁺ exchanger 1, *NIX* NIP3-like protein X, *NF- κ B* nuclear factor kappa-light-chain-enhancer of activated B cells, *PDGF* platelet-derived growth factor, *PFK* phosphofructokinase, *PGM* phosphoglycerate mutase, *PKM* pyruvate kinase M, *PI3K* phosphatidylinositol 3-kinase, *REDD1* protein regulated in development and DNA damage response 1, *TERT* telomerase reverse transcriptase, *TGF* transforming growth factor, *Tie-2* tunica interna endothelial cell kinase 2, *TLR2/6* Toll-like receptor 2/6, *TNF* tumor necrosis factor, *SDF-1* stromal cell-derived factor 1, *uPAR* urokinase plasminogen activator receptor, *VEGF-A* vascular endothelial growth factor A, *VEGFR1/2* vascular endothelial growth factor receptor 1/2, *ZEB1/2* zinc finger E-box-binding homeobox 1/2

Hypoxia also plays an important role in the regulation of tumor cell proliferation and cell cycle progression, via the control of growth factors (e.g. TGFs, IGFs), oncogenes (c-myc), the PI3K/Akt pathway, p21, cyclins, and telomerase [88, 92]. The regulation of tumor cell death is another major aspect of cancer cell biology modulated by the hypoxic response. HIFs can activate proapoptotic genes such as

BNIP3, NIX, and REDD1 [92] and can stabilize p53 [6]. However, tumor cells develop various mechanisms to evade hypoxia-induced cell death, e.g. by HIF-dependent upregulation of anti-apoptotic molecules such as IAP-2 and survivin or by exerting a selective pressure to acquire p53 mutations under hypoxia [34, 36, 78]. Moreover, we have shown that HIF-1/2 α induce the expression of PHD2 and PHD3, which act in a negative feedback loop even under low O₂ concentrations to protect tumor cells against hypoxia-induced cell death by dampening the HIF response [42].

Solid tumors trigger an intrinsic smoldering inflammatory response modulated so as to create a protumorigenic environment, which plays a critical role at different stages of tumor progression [37]. Hypoxia/HIFs are central mediators of this process by modulating key aspects of immune cell function and inflammation to promote an immunosuppressive environment. This is mediated for example through the control of immune cell adhesion (via induction of β 2 integrin), expression of toll-like receptors, production of NO, proinflammatory cytokines (e.g. TNF, IL-1, IL-12), and activation of NF- κ B signaling [47, 76]. An additional central aspect of cancer progression that is under the control of hypoxia is tumor cell invasion and metastasis. Hypoxia can induce the process of epithelial-mesenchymal transition (EMT), which is thought to be an essential early determinant of metastatic dissemination [109]. Hypoxic signaling suppresses the expression of the epithelial cell adhesion molecule E-cadherin, an essential step of EMT [26], through HIF-1 α or HIF-2 α -dependent upregulation of EMT transcriptional repressors of the SNAI, ZEB, or TWIST families [27, 46, 59, 61, 110]. Furthermore, hypoxia can promote metastasis via upregulation of pro-invasive and metastatic HIF target genes including extracellular matrix-remodelling proteins such as lysyl oxidase (LOX) and urokinase plasminogen activator receptor (uPAR) [25, 58], matrix metalloproteinases [94] and the prometastatic chemokine receptor CXCR4 [79, 98, 111].

Additionally, hypoxia plays an essential role in the regulation of the self-renewal and differentiation of physiological stem cells in a variety of tissues, at least in part through the transcriptional activation of central stem cell regulators such as Oct4, Notch, and c-myc [53]. Importantly, work by us and others has demonstrated that hypoxia also has a critical function in the maintenance of cancer stem cells (CSCs), a population of tumor cells with properties of stem cells, that drives tumor initiation and progression [30]. Hypoxia promotes the self-renewal of CSCs, particularly in glioblastoma [40, 72, 91], while HIF knockdown blocks this effect and reduces CSC-mediated tumor growth [65, 73, 91, 95]. Current evidence indicates that HIF-2 α is the main isoform to promote CSC maintenance as HIF-2 α , but not HIF-1 α , is highly expressed and strongly upregulated by hypoxia in glioma CSCs [65], enhances the CSC phenotype [91] and promotes tumor growth [40]. Moreover, HIF-2 α regulates crucial signaling pathways that are involved in stem cell maintenance. HIF-2 α interacts with and stabilizes the Notch ICD (intracellular domain), enhancing Notch signalling to control cellular differentiation [38]. Additionally, HIF-2 α transcriptionally regulates Oct4 [21], a transcription factor that is important for the maintenance of the self renewal of embryonic stem cells and one of four factors necessary to induce pluripotency [99].

12.4 The Hypoxic Response and Tumor Progression

The brief overview provided above highlights the central role of hypoxic signaling in promoting the “hallmarks of cancer” [39, 88]. It is therefore not surprising that hypoxia and HIFs have been associated with tumor initiation and progression and a worse clinical outcome. It has been shown for a number of different cancer types that hypoxia correlates with a more aggressive tumor phenotype, including enhanced angiogenesis, metastasis, recurrence, therapy resistance, and decreased patient survival [49]. In comparison to adjacent tissue widespread HIF activation can be seen in various tumors types, correlating with tumor growth and progression. HIF overexpression in tumors is a result of hypoxia-dependent and hypoxia-independent mechanism such as oncogenic mutations or enhanced growth factor signalling. Various oncogenic mutations can directly lead to HIF α stabilization. For example, genetic alterations of pVHL are a characteristic feature of clear cell renal cell carcinomas and are linked to accumulation predominantly of HIF-2 α [50]. In addition, activating mutations of PI3K, Akt, and Ras, as well as inactivating mutations of PTEN and TSC2 result in enhanced HIF-1 α transcription, translation, or stabilization [14, 31]. An elevated expression of HIFs has been associated with multiple cancers, based on immunohistochemical analysis. HIF-1 α has been found to be upregulated compared to the nonmalignant tissue in a broad variety of tumor types, including oligodendroglioma, breast, cervical, colon, ovarian, endometrial, lung, prostate, bladder, pancreatic, and oropharyngeal cancer [49, 92]. In most of these cancers higher HIF-1 α levels have been associated with poor patient survival [11]. Elevated HIF-2 α , on the other hand has been linked to worse prognosis in a distinct set of tumor entities, including clear cell renal carcinoma, non-small-cell lung carcinoma, head and neck squamous cell carcinoma, neuroblastoma, and glioma [65, 82].

In support of a distinct, nonredundant function of the two main HIF α isoforms, HIF-1 α overexpression correlated with decreased patient mortality in head and neck cancer and non-small-cell lung cancer, in both of which HIF-2 α showed the opposite association [92]. Furthermore, silencing of HIF-2 α , but not HIF-1 α , in a number of cancer cell lines reduced cell proliferation and tumor growth [29]. Such functional differences between HIF-1 α and HIF-2 α could be due to differential expression in the different tumor entities, as well as to the distinct sets of target genes controlled by the two isoforms [41]. In addition, the two HIF α subunits can modulate the activity of key oncogenes or tumor suppressors in opposing fashion. For example, HIF-1 α antagonizes myc transcriptional activity, while HIF-2 α promotes it [32]. Renal cell carcinomas that express HIF-2 α but not HIF-1 α upregulate myc target genes, have increased proliferation and enhanced resistance to replication stress [33]. HIF-1 α and HIF-2 α also have contrasting effects of the function of p53: while HIF-1 α binds to p53 and stabilizes it [6, 75], HIF-2 α indirectly suppresses p53, promoting radio- and chemoresistance [10, 86]. Compared to HIF-1 α , HIF-2 α accumulates at higher oxygen concentrations [43, 108], which more closely resemble the *in vivo* conditions under which tumors arise and grow. At the same time,

while HIF-1 α gets only transiently upregulated under chronic hypoxia, HIF-2 α levels remain elevated in these conditions [44]. In addition, HIF-2 α appears to be the primary isoform regulating the self-renewal capacity of the CSC pool [65, 91], which may have different contributions to the progression of distinct tumor entities [54, 83].

Although HIFs are typically perceived as protumorigenic molecules, in some settings they can also act as tumor suppressors. For example, HIF-1 α -deficient teratomas grow faster due to the refractoriness of the mutant tumor cells to stress-induced apoptosis [16]. Similarly, HIF-1 α and HIF-2 α were stabilized in Vhl-/- ES cells, but the resulting teratomas were smaller than in controls [68]. Furthermore, HIF-2 α overexpression in glioma cells enhanced apoptosis and decreased tumor growth, whereas HIF-2 α inhibition or genetic deletion had the reverse effect [2]. The complexity of HIF function in tumorigenesis was further highlighted by studies using a Kras mutant lung tumor model. Expression of nondegradable HIF-2 α in this system increased tumor burden, angiogenesis, EMT, and decreased survival [55]. Paradoxically, deletion of HIF-2 α in the same model also promoted tumorigenesis, whereas HIF-1 α deletion had no apparent effect [71]. Such contrasting results indicate that the effect of HIFs on tumor progression is likely to depend on the cellular context as well as the precise extent of functional inhibition or activation of specific isoforms and the balance of competing signaling pathways that can be activated by their stimulation or suppression [69].

In glioma, hypoxia plays a prominent role in several aspects. First, the characteristic necrotic regions which represent one of the key criteria for the histological diagnosis of glioblastoma (GBM) are associated with hypoxia [49]. Hypoxia and the activation of HIF also contribute to the second characteristic feature of GBM, the high degree of vascularization [2]; the GBM microcirculation, however, is leaky and functionally inefficient, failing to restore normal oxygenation [103]. Glioma cells overexpress HIF-1 α and HIF-2 α both in culture and in situ, especially in the perinecrotic pseudopalisading areas [49, 91]. Interestingly, the cells found in those regions have been implicated in hypoxia-induced migration away from the necrotic areas [12, 84]. Furthermore, the expression of classical HIF target genes like CA IX and glucose transporter 1 (Glut-1) correlates with higher brain tumor grade and poor response to treatment [28, 45, 52, 67, 89, 96, 102, 112]. Finally, as discussed above, hypoxia and HIF-2 α play a particularly prominent role in the maintenance of glioma CSCs within a hypoxic niche, which is thought to be responsible for determining key aspects of GBM malignancy [30].

12.5 Hypoxia and Therapy Resistance

The capacity of hypoxia to protect tumor cells from radiation damage was first noted in the 1950s and has been extensively corroborated since then (reviewed in [11]). In GBM, for example, elevated hypoxia before radiotherapy is strongly

associated with decreased time to progression and patient survival [97]. In addition, hypoxic cells have an increased resistance to a variety of standard chemotherapeutic agents [101]. Hypoxic signaling converges on multiple pathways that contribute to therapy resistance. For example, hypoxia selects for cancer cell clones with mutant p53, a key mediator of therapy-induced apoptosis [34]. Additionally, in GBM cells hypoxia induces the activation of the antiapoptotic protein Bad and subsequent inhibition of programmed cell death [74]. Moreover, the multidrug resistance gene MDR1 is a direct HIF-1 α target, which can mediate the efflux of chemotherapeutic drugs [20, 106]. The ability of hypoxia to increase the CSC pool, as discussed above, may provide an additional explanation for the decreased sensitivity of hypoxic tumors to treatment. Indeed, a series of studies have demonstrated that CSC have enhanced resistance to chemo- and radiotherapy [30]. This is due to a combination of properties characteristic of CSCs, including the high expression of ABC drug pumps, relative quiescence, resistance to oxidative DNA damage and enhanced DNA repair capacity [7, 90]. The increased resistance of CSCs, combined with the ability of only a very small number of CSCs to reinitiate tumor growth is thought to be a major reason for cancer persistence and relapse after treatment.

Anti-angiogenic therapies have become an established tool in the treatment of several cancers, including colorectal, lung, breast cancer, and GBM [66]. However, following the initial wave of enthusiasm, it has become clear that the inhibition of angiogenesis has complex consequences and smaller than expected benefits for cancer patients. Typically tumor shrinkage is initially observed, but this is followed by adaptation and renewed growth of the tumor, often resulting only in extension of progression-free survival, but not overall survival. The major problem underlying the relative inefficacy of angiogenic therapies is that tumors quickly adapt and manage to circumvent them. The mechanisms of this “evasive resistance” are poorly understood, but preclinical studies have started to suggest several possible explanations. By definition, anti-angiogenic agents are designed to curtail the blood supply of the tumor, thus inducing tumor hypoxia. This may provide one key explanation of the limited therapeutic efficiency of current anti-angiogenic drugs, since hypoxia activates a number of mechanisms that contribute to the evasive resistance following anti-angiogenic therapy. As discussed above, several alternative proangiogenic signals are HIF target genes. In addition, hypoxia enhances the recruitment of bone marrow derived cells to the tumor, which can promote the formation of blood vessels either through secretion of cytokines and growth factors or through direct differentiation into blood vessel cells, such as endothelial cells or pericytes [8]. Interestingly, inhibition of angiogenesis also elicits enhanced local invasion and distant metastasis in different tumor types, including breast tumors, pancreatic neuroendocrine tumors, melanoma, and glioma [22, 77]. The elevated hypoxia observed under these conditions [77] suggests several possible mechanisms for anti-angiogenesis driven metastasis, as outlined in the previous sections, however, the precise pathways involved in the evasive resistance phenotype remain to be elucidated.

12.6 Strategies for Therapeutic Targeting of Tumor Hypoxia

Based on the realization that hypoxia plays a key role at various steps of tumor progression and resistance, substantial effort has been invested in targeting or exploiting tumor hypoxia as an anti-cancer therapeutic strategy (Fig. 12.2). Early attempts were aimed at preventing hypoxia by increasing tumor oxygenation during irradiation, but the clinical efficacy of such interventions was unsatisfactory [11]. Given the relevance of the hypoxic tumor fraction in shaping the tumor phenotype, a different strategy is to take advantage of the hypoxic state of tumor cells in order to selectively eliminate them. Several chemical classes of agents have been proposed which can be specifically converted from a nontoxic to a toxic form by reduction under low oxygen conditions [13]. An example of such a hypoxia activated prodrug is tirapazamine. In hypoxic cells, it gives rise to free radical species which block the function of topoisomerase II and lead to double-stranded DNA breaks. Phase III clinical trials of tirapazamine in combination with chemotherapy have demonstrated benefits in lung cancer patients [104]. Another bioreductive prodrug, the potent DNA intercalator and topoisomerase poison AQ4N, has shown selective activation in hypoxic regions in phase I trials [5]. Other classes of drugs (e.g. CB 1954 or SN 23862) are designed to release more stable cytotoxins upon reduction, which can diffuse away from hypoxic cells and kill additional cells in the tumor, in a “bystander effect” [13].

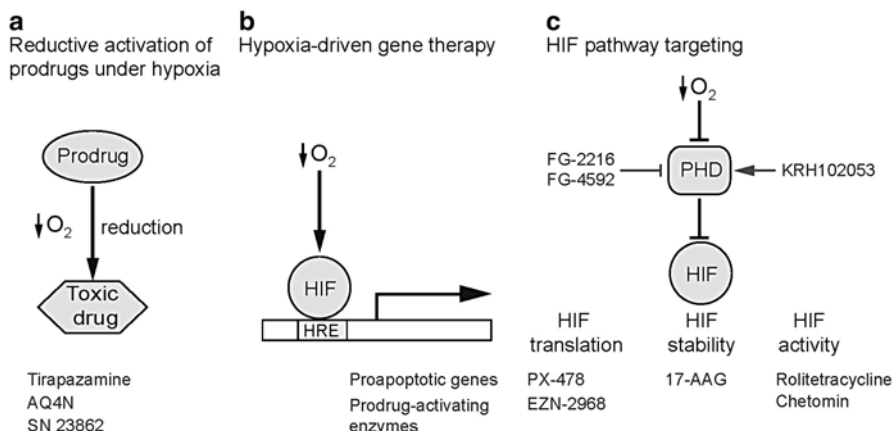


Fig. 12.2 Strategies for therapeutic targeting of tumor hypoxia (a) The hypoxic state of tumor cells can be exploited for the chemical conversion of prodrugs into a toxic form under the reducing conditions created by the shortage of O₂. (b) The stabilization of HIFs under hypoxia can also be used for driving the expression of cytotoxic gene products in hypoxic cells. (c) A third strategy is to interfere with the tumor promoting functions of hypoxic signalling, e.g. by specific targeting of components of the HIF pathway. Shown below the schemes are specific compounds or types of genes that are being explored in preclinical studies or clinical trials. FG-2216 and FG-4592 are PHD inhibitors currently in clinical trials for renal anemia [9, 17, 69]

An alternative set of approaches in preclinical development aims to exploit tumor hypoxia for the selective activation of gene expression. Such hypoxia-targeted gene therapy approaches involve constructs containing HIF-binding sequences (hypoxia response elements (HREs)), which are virally transduced into tumor cells and drive transcription of therapeutic genes in cells that experience hypoxia. Examples of such strategies include the hypoxia-dependent expression of proapoptotic genes [87] or of prodrug-activating enzymes [93]. Others have designed conditionally replicative oncolytic viruses that are specifically activated in hypoxic cells, causing their lysis [81]. A somewhat different approach consists in the coupling of diphtheria toxin to the oxygen-dependent degradation domain (ODD) of HIF-1 α [57]. Under normoxic conditions such a fusion protein would be targeted for proteasomal degradation following ubiquitination of the ODD, however, under hypoxia it would be stabilized allowing the toxin to kill the hypoxic cell.

The largest group of hypoxia-targeting agents in current development are centered around HIF and the molecules that regulate its stability and function. Being a transcription factor, HIF represents a challenging target, but several different approaches have provided interesting hits. Interestingly, a surprisingly broad array of established drugs has been shown to suppress HIF α stability or activity. Examples include the HSP90 antagonists geldanamycin and 17-AAG, the histone deacetylase inhibitors trichostatin A and FK228, the DNA intercalating agents doxorubicin, daunorubicin, and acriflavine, the topoisomerase inhibitor topotecan, cardiac glycosides, as well as inhibitors of central signal transduction pathways like Ras/MAPK, PI3K/Akt, and mTOR [63, 64, 80]. While there is evidence that some of the anticancer effects of these drugs may be mediated by HIF inhibition [63, 64], the diversity of “nonselective” compounds that block HIF may rather be seen as an indication of the central role of this protein in the control of cellular homeostasis, than as an optimal strategy for the design of HIF-targeted therapies. More specific strategies to suppress HIF activity include the blockade of HIF-1 α /HIF-1 β dimerization or HIF binding to p300/CBP, which has been achieved with small molecule inhibitors such as rolitetracycline, chetomin, or YC-1 [80]. A specific inhibitor of HIF-1 α translation, PX-478, exhibited antitumor activity against human xenografts and is currently in phase I clinical trials [56, 107]. The RNA antagonist of HIF-1 α , EZN-2968, inhibits tumor cell growth and is also being tested in phase I clinical trials [35, 69]. A further possibility for suppressing HIF function is to promote activation of PHDs. For instance, a potent small molecule activator of PHD2, KRH102053, has been shown to decrease HIF-1 α levels in tumor cells [19]. In principle, a variety of RNAi or gene therapy approaches, e.g. aimed at the silencing of HIFs, expression of dominant negative HIF mutants or overexpression of PHDs are potentially powerful alternative treatment strategies, provided that safe and efficient methods for clinical delivery become available.

It has to be noted that nearly all therapeutic agents so far have been targeted against the more ubiquitous family member HIF-1 α . However, as discussed in the previous sections, HIF-2 α plays a dominant role in some tumor types or subpopulations of tumor cells, such as CSCs. Small molecule inhibitors have been identified, which suppress HIF-2 α translation in renal cell carcinoma cells through a mecha-

nism dependent on an iron response element in the 5' UTR of the HIF-2 α mRNA; however, the same compounds also decreased the levels of HIF-1 α , albeit via unrelated mechanisms [113]. Therefore concentrating greater efforts on specifically targeting the HIF-2 α isoform remains an important objective for future drug discovery screens. In addition, given that under some circumstances HIFs can also elicit tumor suppressive functions (see above), exploring HIF activating strategies, e.g. by using PHD inhibitors, may in some cases also prove valuable. Our own work, for example, has shown that PHD inhibition in GBM cells facilitates cell death induction by staurosporine or TRAIL [42].

Our growing understanding of the mechanisms mediating the hypoxic response and the signaling pathways involved in its regulation have allowed us to more fully comprehend central aspects of tumor cell biology and malignant progression. Our deepened knowledge of O₂ homeostasis in tumors has formed the basis for the design of novel therapeutic strategies targeted at hypoxic signaling, which carry the potential to become a powerful weapon in our battle against cancer.

Acknowledgements This work was supported by grants from the DFG (KFO210, AC 110/4-1; EXC 147/1), LOEWE (OSF), the Deutsche Krebshilfe (111719), the von Behring-Röntgen Foundation (58-0069; 59-0037), the RKA-Förderpool and the UKGM Kooperationsvertrag.

References

1. Acker T, Acker H. Cellular oxygen sensing need in CNS function: physiological and pathological implications. *J Exp Biol.* 2004;207:3171–88.
2. Acker T, Diez-Juan A, Aragones J, Tjwa M, Brusselmans K, Moons L, Fukumura D, Moreno-Murciano MP, Herbert JM, Burger A, Riedel J, Elvert G, Flamme I, Maxwell PH, Collen D, Dewerchin M, Jain RK, Plate KH, Carmeliet P. Genetic evidence for a tumor suppressor role of HIF-2 α . *Cancer Cell.* 2005;8:131–41.
3. Acker T, Fandrey J, Acker H. The good, the bad and the ugly in oxygen-sensing: ROS, cytochromes and prolyl-hydroxylases. *Cardiovasc Res.* 2006;71:195–207.
4. Acker T, Plate KH. A role for hypoxia and hypoxia-inducible transcription factors in tumor physiology. *J Mol Med.* 2002;80:562–75.
5. Albertella MR, Loadman PM, Jones PH, Phillips RM, Rampling R, Burnet N, Alcock C, Anthony A, Vjaters E, Dunk CR, Harris PA, Wong A, Lalani AS, Twelves CJ. Hypoxia-selective targeting by the bioreductive prodrug AQ4N in patients with solid tumors: results of a phase I study. *Clin Cancer Res.* 2008;14:1096–104.
6. An WG, Kanekal M, Simon MC, Maltepe E, Blagosklonny MV, Neckers LM. Stabilization of wild-type p53 by hypoxia-inducible factor 1 α . *Nature.* 1998;392:405–8.
7. Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, Dewhirst MW, Bigner DD, Rich JN. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature.* 2006;444:756–60.
8. Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer.* 2008;8:592–603.
9. Bernhardt WM, Wiesener MS, Scigalla P, Chou J, Schmieder RE, Gunzler V, Eckardt KU. Inhibition of prolyl hydroxylases increases erythropoietin production in ESRD. *J Am Soc Nephrol.* 2010;21:2151–6.

10. Bertout JA, Majmundar AJ, Gordan JD, Lam JC, Ditsworth D, Keith B, Brown EJ, Nathanson KL, Simon MC. HIF2 α inhibition promotes p53 pathway activity, tumor cell death, and radiation responses. *Proc Natl Acad Sci U S A*. 2009;106:14391–6.
11. Bertout JA, Patel SA, Simon MC. The impact of O₂ availability on human cancer. *Nat Rev Cancer*. 2008;8:967–75.
12. Brat DJ, Castellano-Sanchez AA, Hunter SB, Pecot M, Cohen C, Hammond EH, Devi SN, Kaur B, Van Meir EG. Pseudopalisades in glioblastoma are hypoxic, express extracellular matrix proteases, and are formed by an actively migrating cell population. *Cancer Res*. 2004;64:920–7.
13. Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. *Nat Rev Cancer*. 2004;4:437–47.
14. Brugarolas J, Kaelin Jr WG. Dysregulation of HIF and VEGF is a unifying feature of the familial hamartoma syndromes. *Cancer Cell*. 2004;6:7–10.
15. Bruick RK, McKnight SL. A conserved family of prolyl-4-hydroxylases that modify HIF. *Science*. 2001;294:1337–40.
16. Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M, Neeman M, Bono F, Abramovitch R, Maxwell P, Koch CJ, Ratcliffe P, Moons L, Jain RK, Collen D, Keshert E. Role of HIF-1 α in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature*. 1998;394:485–90.
17. Cases A. The latest advances in kidney diseases and related disorders. *Drug News Perspect*. 2007;20:647–54.
18. Chiche J, Brahimi-Horn MC, Pouyssegur J. Tumour hypoxia induces a metabolic shift causing acidosis: a common feature in cancer. *J Cell Mol Med*. 2010;14:771–94.
19. Choi HJ, Song BJ, Gong YD, Gwak WJ, Soh Y. Rapid degradation of hypoxia-inducible factor-1 α by KRH102053, a new activator of prolyl hydroxylase 2. *Br J Pharmacol*. 2008;154:114–25.
20. Comerford KM, Wallace TJ, Karhausen J, Louis NA, Montalto MC, Colgan SP. Hypoxia-inducible factor-1-dependent regulation of the multidrug resistance (MDR1) gene. *Cancer Res*. 2002;62:3387–94.
21. Covelto KL, Kehler J, Yu H, Gordan JD, Arsham AM, Hu CJ, Labosky PA, Simon MC, Keith B. HIF-2 α regulates Oct-4: effects of hypoxia on stem cell function, embryonic development, and tumor growth. *Genes Dev*. 2006;20:557–70.
22. Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG, Kerbel RS. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. *Cancer Cell*. 2009;15:232–9.
23. Epstein AC, Gleadle JM, McNeill LA, Hewitson KS, O'Rourke J, Mole DR, Mukherji M, Metzen E, Wilson MI, Dhanda A, Tian YM, Masson N, Hamilton DL, Jaakkola P, Barstead R, Hodgkin J, Maxwell PH, Pugh CW, Schofield CJ, Ratcliffe PJ. *C. elegans* EGL-9 and mammalian homologs define a family of dioxygenases that regulate HIF by prolyl hydroxylation. *Cell*. 2001;107:43–54.
24. Erecinska M, Silver IA. Tissue oxygen tension and brain sensitivity to hypoxia. *Respir Physiol*. 2001;128:263–76.
25. Erler JT, Bennewith KL, Nicolau M, Dornhofer N, Kong C, Le QT, Chi JT, Jeffrey SS, Giaccia AJ. Lysyl oxidase is essential for hypoxia-induced metastasis. *Nature*. 2006;440:1222–6.
26. Esteban MA, Tran MG, Harten SK, Hill P, Castellanos MC, Chandra A, Raval R, O'Brien TS, Maxwell PH. Regulation of E-cadherin expression by VHL and hypoxia-inducible factor. *Cancer Res*. 2006;66:3567–75.
27. Evans AJ, Russell RC, Roche O, Burry TN, Fish JE, Chow VW, Kim WY, Saravanan A, Maynard MA, Gervais ML, Sufan RI, Roberts AM, Wilson LA, Betten M, Vandewalle C, Bex G, Marsden PA, Irwin MS, Teh BT, Jewett MA, Ohh M. VHL promotes E2 box-dependent E-cadherin transcription by HIF-mediated regulation of SIP1 and snail. *Mol Cell Biol*. 2007;27:157–69.
28. Flynn JR, Wang L, Gillespie DL, Stoddard GJ, Reid JK, Owens J, Ellsworth GB, Salzman KL, Kinney AY, Jensen RL. Hypoxia-regulated protein expression, patient characteristics,

- and preoperative imaging as predictors of survival in adults with glioblastoma multiforme. *Cancer*. 2008;113:1032–42.
29. Franovic A, Holterman CE, Payette J, Lee S. Human cancers converge at the HIF-2 α oncogenic axis. *Proc Natl Acad Sci U S A*. 2009;106:21306–11.
 30. Garvalov BK, Acker T. Cancer stem cells: a new framework for the design of tumor therapies. *J Mol Med*. 2011;89:95–107.
 31. Gerald D, Berra E, Frapart YM, Chan DA, Giaccia AJ, Mansuy D, Pouyssegur J, Yaniv M, Mechta-Grigoriou F. JunD reduces tumor angiogenesis by protecting cells from oxidative stress. *Cell*. 2004;118:781–94.
 32. Gordan JD, Bertout JA, Hu CJ, Diehl JA, Simon MC. HIF-2 α promotes hypoxic cell proliferation by enhancing c-myc transcriptional activity. *Cancer Cell*. 2007;11:335–47.
 33. Gordan JD, Lal P, Dondeti VR, Letrero R, Parekh KN, Oquendo CE, Greenberg RA, Flaherty KT, Rathmell WK, Keith B, Simon MC, Nathanson KL. HIF- α effects on c-Myc distinguish two subtypes of sporadic VHL-deficient clear cell renal carcinoma. *Cancer Cell*. 2008;14:435–46.
 34. Graeber TG, Osmanian C, Jacks T, Housman DE, Koch CJ, Lowe SW, Giaccia AJ. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature*. 1996;379:88–91.
 35. Greenberger LM, Horak ID, Filpula D, Sapra P, Westergaard M, Frydenlund HF, Albaek C, Schroder H, Orum H. A RNA antagonist of hypoxia-inducible factor-1 α , EZN-2968, inhibits tumor cell growth. *Mol Cancer Ther*. 2008;7:3598–608.
 36. Greijer AE, van der Wall E. The role of hypoxia inducible factor 1 (HIF-1) in hypoxia induced apoptosis. *J Clin Pathol*. 2004;57:1009–14.
 37. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell*. 2010;140:883–99.
 38. Gustafsson MV, Zheng X, Pereira T, Gradin K, Jin S, Lundkvist J, Ruas JL, Poellinger L, Lendahl U, Bondesson M. Hypoxia requires notch signaling to maintain the undifferentiated cell state. *Dev Cell*. 2005;9:617–28.
 39. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144:646–74.
 40. Heddleston JM, Li Z, McLendon RE, Hjelmeland AB, Rich JN. The hypoxic microenvironment maintains glioblastoma stem cells and promotes reprogramming towards a cancer stem cell phenotype. *Cell Cycle*. 2009;8:3274–84.
 41. Henze AT, Acker T. Feedback regulators of hypoxia-inducible factors and their role in cancer biology. *Cell Cycle*. 2010;9:2749–63.
 42. Henze AT, Riedel J, Diem T, Wenner J, Flamme I, Pouysegur J, Plate KH, Acker T. Prolyl hydroxylases 2 and 3 act in gliomas as protective negative feedback regulators of hypoxia-inducible factors. *Cancer Res*. 2010;70:357–66.
 43. Holmquist L, Jogi A, Pahlman S. Phenotypic persistence after reoxygenation of hypoxic neuroblastoma cells. *Int J Cancer*. 2005;116:218–25.
 44. Holmquist-Mengelbier L, Fredlund E, Lofstedt T, Noguera R, Navarro S, Nilsson H, Pietras A, Vallon-Christersson J, Borg A, Gradin K, Poellinger L, Pahlman S. Recruitment of HIF-1 α and HIF-2 α to common target genes is differentially regulated in neuroblastoma: HIF-2 α promotes an aggressive phenotype. *Cancer Cell*. 2006;10:413–23.
 45. Ihnatko R, Kubes M, Takacova M, Sedlakova O, Sedlak J, Pastorek J, Kopacek J, Pastorekova S. Extracellular acidosis elevates carbonic anhydrase IX in human glioblastoma cells via transcriptional modulation that does not depend on hypoxia. *Int J Oncol*. 2006;29:1025–33.
 46. Imai T, Horiuchi A, Wang C, Oka K, Ohira S, Nikaido T, Konishi I. Hypoxia attenuates the expression of E-cadherin via up-regulation of SNAIL in ovarian carcinoma cells. *Am J Pathol*. 2003;163:1437–47.
 47. Imtiyaz HZ, Simon MC. Hypoxia-inducible factors as essential regulators of inflammation. *Curr Top Microbiol Immunol*. 2010;345:105–20.
 48. Ivan M, Kondo K, Yang H, Kim W, Valiando J, Ohh M, Salic A, Asara JM, Lane WS, Kaelin Jr WG. HIF α targeted for VHL-mediated destruction by proline hydroxylation: implications for O₂ sensing. *Science*. 2001;292:464–8.

49. Jensen RL. Brain tumor hypoxia: tumorigenesis, angiogenesis, imaging, pseudoprogression, and as a therapeutic target. *J Neurooncol.* 2009;92:317–35.
50. Kaelin WG. Von Hippel-Lindau disease. *Annu Rev Pathol.* 2007;2:145–73.
51. Kaelin Jr WG, Ratcliffe PJ. Oxygen sensing by metazoans: the central role of the HIF hydroxylase pathway. *Mol Cell.* 2008;30:393–402.
52. Kaynar MY, Sanus GZ, Hnimoglu H, Kacira T, Kemerdere R, Atukeren P, Gumustas K, Canbaz B, Tanriverdi T. Expression of hypoxia inducible factor-1 α in tumors of patients with glioblastoma multiforme and transitional meningioma. *J Clin Neurosci.* 2008;15:1036–42.
53. Keith B, Simon MC. Hypoxia-inducible factors, stem cells, and cancer. *Cell.* 2007;129:465–72.
54. Kelly PN, Dakic A, Adams JM, Nutt SL, Strasser A. Tumor growth need not be driven by rare cancer stem cells. *Science.* 2007;317:337.
55. Kim WY, Perera S, Zhou B, Carretero J, Yeh JJ, Heathcote SA, Jackson AL, Nikolinakos P, Ospina B, Naumov G, Brandstetter KA, Weigman VJ, Zaghul S, Hayes DN, Padera RF, Heymach JV, Kung AL, Sharpless NE, Kaelin Jr WG, Wong KK. HIF2 α cooperates with RAS to promote lung tumorigenesis in mice. *J Clin Invest.* 2009;119:2160–70.
56. Koh MY, Spivak-Kroizman T, Venturini S, Welsh S, Williams RR, Kirkpatrick DL, Powis G. Molecular mechanisms for the activity of PX-478, an antitumor inhibitor of the hypoxia-inducible factor-1 α . *Mol Cancer Ther.* 2008;7:90–100.
57. Koshikawa N, Takenaga K. Hypoxia-regulated expression of attenuated diphtheria toxin A fused with hypoxia-inducible factor-1 α oxygen-dependent degradation domain preferentially induces apoptosis of hypoxic cells in solid tumor. *Cancer Res.* 2005;65:11622–30.
58. Krishnamachary B, Berg-Dixon S, Kelly B, Agani F, Feldser D, Ferreira G, Iyer N, LaRusch J, Pak B, Taghavi P, Semenza GL. Regulation of colon carcinoma cell invasion by hypoxia-inducible factor 1. *Cancer Res.* 2003;63:1138–43.
59. Krishnamachary B, Zagzag D, Nagasawa H, Rainey K, Okuyama H, Baek JH, Semenza GL. Hypoxia-inducible factor-1-dependent repression of E-cadherin in von Hippel-Lindau tumor suppressor-null renal cell carcinoma mediated by TCF3, ZFHX1A, and ZFHX1B. *Cancer Res.* 2006;66:2725–31.
60. Kroemer G, Pouyssegur J. Tumor cell metabolism: cancer's Achilles' heel. *Cancer Cell.* 2008;13:472–82.
61. Kurrey NK, KA, Bapat SA. Snail and Slug are major determinants of ovarian cancer invasiveness at the transcription level. *Gynecol Oncol.* 2005;97:155–65.
62. Lando D, Peet DJ, Gorman JJ, Whelan DA, Whitelaw ML, Bruick RK. FIH-1 is an asparaginyl hydroxylase enzyme that regulates the transcriptional activity of hypoxia-inducible factor. *Genes Dev.* 2002;16:1466–71.
63. Lee K, Qian DZ, Rey S, Wei H, Liu JO, Semenza GL. Anthracycline chemotherapy inhibits HIF-1 transcriptional activity and tumor-induced mobilization of circulating angiogenic cells. *Proc Natl Acad Sci U S A.* 2009;106:2353–8.
64. Lee K, Zhang H, Qian DZ, Rey S, Liu JO, Semenza GL. Acriflavine inhibits HIF-1 dimerization, tumor growth, and vascularization. *Proc Natl Acad Sci U S A.* 2009;106:17910–5.
65. Li Z, Bao S, Wu Q, Wang H, Eyler C, Sathornsumetee S, Shi Q, Cao Y, Lathia J, McLendon RE, Hjelmeland AB, Rich JN. Hypoxia-inducible factors regulate tumorigenic capacity of glioma stem cells. *Cancer Cell.* 2009;15:501–13.
66. Loges S, Schmidt T, Carmeliet P. Mechanisms of resistance to anti-angiogenic therapy and development of third-generation anti-angiogenic drug candidates. *Genes Cancer.* 2010;1:12–25.
67. Lund EL, Hog A, Olsen MW, Hansen LT, Engelholm SA, Kristjansen PE. Differential regulation of VEGF, HIF1 α and angiopoietin-1, -2 and -4 by hypoxia and ionizing radiation in human glioblastoma. *Int J Cancer.* 2004;108:833–8.
68. Mack FA, Rathmell WK, Arsham AM, Gnarr J, Keith B, Simon MC. Loss of pVHL is sufficient to cause HIF dysregulation in primary cells but does not promote tumor growth. *Cancer Cell.* 2003;3:75–88.

69. Majmundar AJ, Wong WJ, Simon MC. Hypoxia-inducible factors and the response to hypoxic stress. *Mol Cell*. 2010;40:294–309.
70. Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, Wykoff CC, Pugh CW, Maher ER, Ratcliffe PJ. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature*. 1999;399:271–5.
71. Mazumdar J, Hickey MM, Pant DK, Durham AC, Sweet-Cordero A, Vachani A, Jacks T, Chodosh LA, Kissil JL, Simon MC, Keith B. HIF-2 α deletion promotes Kras-driven lung tumor development. *Proc Natl Acad Sci U S A*. 2010;107:14182–7.
72. McCord AM, Jamal M, Shankavaram UT, Lang FF, Camphausen K, Tofilon PJ. Physiologic oxygen concentration enhances the stem-like properties of CD133+ human glioblastoma cells in vitro. *Mol Cancer Res*. 2009;7:489–97.
73. Mendez O, Zavadil J, Esencay M, Lukyanov Y, Santovasi D, Wang SC, Newcomb EW, Zagzag D. Knock down of HIF-1 α in glioma cells reduces migration in vitro and invasion in vivo and impairs their ability to form tumor spheres. *Mol Cancer*. 2010;9:133.
74. Merighi S, Benini A, Mirandola P, Gessi S, Varani K, Leung E, MacLennan S, Baraldi PG, Borea PA. Hypoxia inhibits paclitaxel-induced apoptosis through adenosine-mediated phosphorylation of bad in glioblastoma cells. *Mol Pharmacol*. 2007;72:162–72.
75. Moeller BJ, Dreher MR, Rabbani ZN, Schroeder T, Cao Y, Li CY, Dewhirst MW. Pleiotropic effects of HIF-1 blockade on tumor radiosensitivity. *Cancer Cell*. 2005;8:99–110.
76. Nizet V, Johnson RS. Interdependence of hypoxic and innate immune responses. *Nat Rev Immunol*. 2009;9:609–17.
77. Páez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Viñals F, Inoue M, Bergers G, Hanahan D, Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell*. 2009;15:220–31.
78. Peng XH, Karna P, Cao Z, Jiang BH, Zhou M, Yang L. Cross-talk between epidermal growth factor receptor and hypoxia-inducible factor-1 α signal pathways increases resistance to apoptosis by up-regulating survivin gene expression. *J Biol Chem*. 2006;281:25903–14.
79. Phillips RJ, Mestas J, Gharaee-Kermani M, Burdick MD, Sica A, Belperio JA, Keane MP, Strieter RM. Epidermal growth factor and hypoxia-induced expression of CXC chemokine receptor 4 on non-small cell lung cancer cells is regulated by the phosphatidylinositol 3-kinase/PTEN/AKT/mammalian target of rapamycin signaling pathway and activation of hypoxia inducible factor-1 α . *J Biol Chem*. 2005;280:22473–81.
80. Poon E, Harris AL, Ashcroft M. Targeting the hypoxia-inducible factor (HIF) pathway in cancer. *Expert Rev Mol Med*. 2009;11, e26.
81. Post DE, Van Meir EG. A novel hypoxia-inducible factor (HIF) activated oncolytic adenovirus for cancer therapy. *Oncogene*. 2003;22:2065–72.
82. Qing G, Simon MC. Hypoxia inducible factor-2 α : a critical mediator of aggressive tumor phenotypes. *Curr Opin Genet Dev*. 2009;19:60–6.
83. Quintana E, Shackleton M, Sabel MS, Fullen DR, Johnson TM, Morrison SJ. Efficient tumour formation by single human melanoma cells. *Nature*. 2008;456:593–8.
84. Raza SM, Fuller GN, Rhee CH, Huang S, Hess K, Zhang W, Sawaya R. Identification of necrosis-associated genes in glioblastoma by cDNA microarray analysis. *Clin Cancer Res*. 2004;10:212–21.
85. Rey S, Semenza GL. Hypoxia-inducible factor-1-dependent mechanisms of vascularization and vascular remodelling. *Cardiovasc Res*. 2010;86:236–42.
86. Roberts AM, Watson IR, Evans AJ, Foster DA, Irwin MS, Ohh M. Suppression of hypoxia-inducible factor 2 α restores p53 activity via Hdm2 and reverses chemoresistance of renal carcinoma cells. *Cancer Res*. 2009;69:9056–64.
87. Ruan H, Wang J, Hu L, Lin CS, Lamborn KR, Deen DF. Killing of brain tumor cells by hypoxia-responsive element mediated expression of BAX. *Neoplasia*. 1999;1:431–7.
88. Ruan K, Song G, Ouyang G. Role of hypoxia in the hallmarks of human cancer. *J Cell Biochem*. 2009;107:1053–62.
89. Sathornsumetee S, Cao Y, Marcello JE, Herndon 2nd JE, McLendon RE, Desjardins A, Friedman HS, Dewhirst MW, Vredenburgh JJ, Rich JN. Tumor angiogenic and hypoxic pro-

- files predict radiographic response and survival in malignant astrocytoma patients treated with bevacizumab and irinotecan. *J Clin Oncol.* 2008;26:271–8.
90. Schatton T, Murphy GF, Frank NY, Yamaura K, Waaga-Gasser AM, Gasser M, Zhan Q, Jordan S, Duncan LM, Weishaupt C, Fuhlbrigge RC, Kupper TS, Sayegh MH, Frank MH. Identification of cells initiating human melanoma. *Nature.* 2008;451:345–9.
 91. Seidel S, Garvalov BK, Wirta V, von Stechow L, Schänzer A, Meletis K, Wolter M, Sommerlad D, Henze AT, Nistér M, Reifenberger G, Lundeberg J, Frisén J, Acker T. A hypoxic niche regulates glioblastoma stem cells through hypoxia inducible factor 2 α . *Brain.* 2010;133:983–95.
 92. Semenza GL. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer.* 2003;3:721–32.
 93. Shibata T, Giaccia AJ, Brown JM. Hypoxia-inducible regulation of a prodrug-activating enzyme for tumor-specific gene therapy. *Neoplasia.* 2002;4:40–8.
 94. Shyu KG, Hsu FL, Wang MJ, Wang BW, Lin S. Hypoxia-inducible factor 1 α regulates lung adenocarcinoma cell invasion. *Exp Cell Res.* 2007;313:1181–91.
 95. Soeda A, Park M, Lee D, Mintz A, Androutsellis-Theotokis A, McKay RD, Engh J, Iwama T, Kunisada T, Kassam AB, Pollack IF, Park DM. Hypoxia promotes expansion of the CD133-positive glioma stem cells through activation of HIF-1 α . *Oncogene.* 2009;28:3949–59.
 96. Sondergaard KL, Hilton DA, Penney M, Ollerenshaw M, Demaine AG. Expression of hypoxia-inducible factor 1 α in tumours of patients with glioblastoma. *Neuropathol Appl Neurobiol.* 2002;28:210–7.
 97. Spence AM, Muzi M, Swanson KR, O’Sullivan F, Rockhill JK, Rajendran JG, Adamsen TC, Link JM, Swanson PE, Yagle KJ, Rostomily RC, Silbergeld DL, Krohn KA. Regional hypoxia in glioblastoma multiforme quantified with [18 F]fluoromisonidazole positron emission tomography before radiotherapy: correlation with time to progression and survival. *Clin Cancer Res.* 2008;14:2623–30.
 98. Staller P, Sulitkova J, Lisztwan J, Moch H, Oakeley EJ, Krek W. Chemokine receptor CXCR4 downregulated by von Hippel-Lindau tumour suppressor pVHL. *Nature.* 2003;425:307–11.
 99. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell.* 2006;126:663–76.
 100. Taylor CT, Cummins EP. The role of NF-kappaB in hypoxia-induced gene expression. *Ann N Y Acad Sci.* 2009;1177:178–84.
 101. Teicher BA. Hypoxia and drug resistance. *Cancer Metastasis Rev.* 1994;13:139–68.
 102. Tsukamoto H, Boado RJ, Pardridge WM. Differential expression in glioblastoma multiforme and cerebral hemangioblastoma of cytoplasmic proteins that bind two different domains within the 3’-untranslated region of the human glucose transporter 1 (GLUT1) messenger RNA. *J Clin Invest.* 1996;97:2823–32.
 103. Vajkoczy P, Menger MD. Vascular microenvironment in gliomas. *Cancer Treat Res.* 2004;117:249–62.
 104. von Pawel J, von Roemeling R, Gatzemeier U, Boyer M, Elisson LO, Clark P, Talbot D, Rey A, Butler TW, Hirsh V, Olver I, Bergman B, Ayoub J, Richardson G, Dunlop D, Arcenas A, Vescio R, Viallet J, Treat J. Tirapazamine plus cisplatin versus cisplatin in advanced non-small-cell lung cancer: a report of the international CATAPULT I study group. Cisplatin and tirapazamine in subjects with advanced previously untreated non-small-cell lung tumors. *J Clin Oncol.* 2000;18:1351–9.
 105. Wang GL, Jiang BH, Rue EA, Semenza GL. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O₂ tension. *Proc Natl Acad Sci U S A.* 1995;92:5510–4.
 106. Wartenberg M, Ling FC, Muschen M, Klein F, Acker H, Gassmann M, Petrat K, Putz V, Hescheler J, Sauer H. Regulation of the multidrug resistance transporter P-glycoprotein in multicellular tumor spheroids by hypoxia-inducible factor (HIF-1) and reactive oxygen species. *FASEB J.* 2003;17:503–5.

107. Welsh S, Williams R, Kirkpatrick L, Paine-Murrieta G, Powis G. Antitumor activity and pharmacodynamic properties of PX-478, an inhibitor of hypoxia-inducible factor-1 α . *Mol Cancer Ther.* 2004;3:233–44.
108. Wiesener MS, Turley H, Allen WE, Willam C, Eckardt KU, Talks KL, Wood SM, Gatter KC, Harris AL, Pugh CW, Ratcliffe PJ, Maxwell PH. Induction of endothelial PAS domain protein-1 by hypoxia: characterization and comparison with hypoxia-inducible factor-1 α . *Blood.* 1998;92:2260–8.
109. Yang J, Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell.* 2008;14:818–29.
110. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. Direct regulation of TWIST by HIF-1 α promotes metastasis. *Nat Cell Biol.* 2008;10:295–305.
111. Zagzag D, Lukyanov Y, Lan L, Ali MA, Esencay M, Mendez O, Yee H, Voura EB, Newcomb EW. Hypoxia-inducible factor 1 and VEGF upregulate CXCR4 in glioblastoma: implications for angiogenesis and glioma cell invasion. *Lab Invest.* 2006;86:1221–32.
112. Zagzag D, Zhong H, Scalzitti JM, Laughner E, Simons JW, Semenza GL. Expression of hypoxia-inducible factor 1 α in brain tumors: association with angiogenesis, invasion, and progression. *Cancer.* 2000;88:2606–18.
113. Zimmer M, Ebert BL, Neil C, Brenner K, Papaioannou I, Melas A, Tolliday N, Lamb J, Pantopoulos K, Golub T, Iliopoulos O. Small-molecule inhibitors of HIF-2 α translation link its 5'UTR iron-responsive element to oxygen sensing. *Mol Cell.* 2008;32:838–48.