Struan F.A. Grant Editor

The Genetics of Obesity



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Editor Struan F.A. Grant Children's Hospital of Philadelphia Research Institute Philadelphia, PA, USA

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Chapter 1 Genetic Variation and Obesity Prior to the Era of Genome-Wide Association Studies

Shana E. McCormack

Abstract The purpose of this chapter is to present a comprehensive review of the evidence for how genetic variation contributes to common obesity (which we have chosen to define as affecting >1 % of obese individuals, likely non-syndromic in etiology, and related to diet and inactivity) in the general population prior to the era of genome wide association studies (GWAS). Twin and adoption studies demonstrate that the tendency to be obese is highly heritable, and also suggest that the cumulative effect of gene-environment interactions on body mass index (BMI) seems to increase with age. Before GWAS, one common approach to dissecting the role of genetic variability in common obesity was to investigate candidate genes. Some of the genes implicated in monogenic or syndromic obesity by traditional linkage analysis, for example, MC4R, have milder variants that appear to be important in common obesity as well. By 2004, over 600 candidate genes or chromosomal regions had been implicated in the pathogenesis of obesity, and 18 of these had multiple lines of supporting evidence. Despite this, much of the heritability in obesity remained to be explained. Next-generation sequencing technology should produce additional insights that extended these seminal investigations but despite this, much of the so-called missing heritability identified prior to the era of GWAS persists.

The global prevalence of obesity has risen rapidly, in particular during the latter part of the twentieth century and the beginning of the twenty-first century. Obesity became a World Health Organization priority after its significant contribution to cardiovascular disease burden as well as all-cause mortality from other conditions, including cancer, came to be appreciated [1]. In the USA, the harbinger of the global

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epidemic, obesity prevalence nearly tripled, from 12 to 33 % in men and 17 to 37 % in women between 1971 and 2006 [2].

Part of the impetus to pursue studies related to the genetics of obesity is an attempt to explain its rising prevalence, and in particular, the rising frequency of extreme and young-onset obesity phenotypes [3]. Nuclear genetic variation alone is unlikely to account for such a recent and rapid change in phenotype; a complex variety of interacting environmental, nutritional, microbial, epigenetic, behavioral, sociocultural, economic, and other factors likely contribute as well [4]. Genetics, however, may help to explain why some individuals are more vulnerable to similar so-called obesogenic environmental influences than others. An understanding of differential genetic susceptibility may inform our search for modifiable or even reversible obesity risk factors, and motivate individualized, targeted prevention efforts.

Prior to the era of genome-wide association studies (GWAS), which will be described in greater detail in subsequent chapters, these efforts took several forms. The purpose of this chapter is to present a comprehensive review of the evidence for how genetic variation contributes to common obesity (which we have chosen to define as affecting >1 % of obese individuals, likely diet-induced, and apparently non-syndromic) in the general population prior to the era of GWAS.

First, twin and adoption studies are two strategies to produce heritability estimates for obesity, and the successes and limitations of this work are considered here. Next, racial and ethnic differences in susceptibility to obesity, along with related traits (body composition and resting energy expenditure) may be related to inherited factors, and some of this is discussed as well. In addition, for individuals who have obesity along with other unique clinical features (e.g., intellectual impairment, dysmorphism, visual or hearing anomalies) and similarly affected family members, an inherited syndrome may be present, and traditional linkage analyses have been used to implicate a particular chromosomal region in these patients. In some cases, the relevant gene(s) and function(s) have been characterized (e.g., *MC4R* and Bardet–Biedl syndrome); the reader is referred to the chapters on monogenic and syndromic obesity for a more detailed discussion of these. For some of these genes where mutations cause severe obesity phenotypes, milder variants seem to play a role in common obesity, and we present some of this evidence here, and in the Table 1.1 below.

Some of the identified genes belong to biological pathways whose integrated function affects the propensity to develop obesity, including, for example, the hypothalamic regulation of appetite and energy balance. As the nature of these pathways has been more fully elucidated in model systems, new candidate genes have been put forth and their contribution to common obesity has been investigated, and some of these findings are reviewed, as well as summarized in the Table 1.1. By 2004, over 600 candidate genes or chromosomal regions had been implicated in the pathogenesis of obesity, and 18 of these had multiple lines of supporting evidence [5]. The advent of next-generation sequencing technology should build on these seminal investigations, and we conclude this chapter by outlining some of the most pressing questions in the field for which, it was hoped, the new techniques would provide much-needed answers.

			Role in common obesity (>1 %) demonstrated prior to era of GWAS (or after, where noted, in candidate
Candidate gene	Mechanism	OMIM #	approach)
Monogenic, obesity-predominant (see t	ext)		
Leptin	Secreted by adipocytes in white	614962	Pre-GWAS studies suggested association, particu-
	adipose tissue [32]; circulating		larly in 5' region, with common obesity [44] that
	levels higher in obesity [39]		may be sex-dependent [45]
Leptin receptor	Hypothalamic receptor for leptin [38]	614963	Mutations may be present in up to 3 % of children with severe, early-onset obesity [42], but consistent results for variation in larger
			populations was lacking
Melanocortin 4 receptor (MC4R)	Hypothalamic G-protein-coupled receptor [49]	155541	Most variation in severe obesity explained by a single locus prior to GWAS [54]
Pro-opiomelanocortin (POMC)	Hypothalamic preprohormone produces α-MSH, a ligand for MC4R [56]	609734	Variants at <i>POMC</i> locus affect BMI in Europeans [59] and Hispanic Americans [60], and influences waist-hip ratio in the normal range [113]
Proprotein convertase subtilisin/kexin	Prohormone convertase, cleaves	600955	Post-GWAS studies demonstrate a role in common
type 1 (<i>PCSK1</i>) Syndromic (selected)	POMC into signaling ligands [56]		obesity [63]
Prader–Willi Syndrome (deletion of paternally imprinted <i>SNRPN</i> ,	Disorder of imprinting/maternal heterodisomy [114]	176270, 182279 (SNRPN), 602117	Possible association with copy number variation at this locus in later candidate study [115];
necedin, possibly others 15q11-q13)		(necedin)	necedin candidate study did not disclose an association [116]
Bardet–Biedl (at least 18 implicated loci)	Disorder of ciliary function [117], hypothalamic appetite dysregula-	209900, many implicated genes	Possible evidence in isolated population [119], relationship of MKKS and common obesity was
	tion [118]	0	investigated, no clear evidence [120], possible
Alstrom syndrome (Alms1) [122, 123]	Ciliary dysgenesis is implicated [124]	203800	Early association studies did not identify a role for variation in common obesity [125]

Table 1.1 Identification of selected candidate genes, and investigation of their contribution to common obesity prior to GWAS

(continued)

Table 1.1 (continued)			
			Role in common obesity (>1 %) demonstrated prior to era of GWAS (or after, where noted, in candidate
Candidate gene	Mechanism	OMIM #	approach)
Associations by other candidate appros <i>Hypothalamic</i>	aches		
AGRP (agouti-related protein)	Cell-signaling protein, antagonist at melanocortin-3 and melanocortin-4 receptors [126]; plasma levels related to obesity [127]	602311	Variation may be related to age-dependent onset of obesity [65]; possible association of a variant with reduced fat mass [66]
Brain-derived natriuretic factor (BDNF)	Neuronal growth factor [128] downstream of MC4R [68]	113505	Association with disordered eating (including bulimia) has been identified [73]
Syndecans 1,3 (Sdc 1,3)	Membrane-bound heparin sulfate proteoglycans, feeding in mice [67, 129]	186355, 186357	Variation associated with obesity in Koreans [130]
Single-minded, drosophila, homolog (<i>Sim1</i>)	Sim1 haploinsufficiency appear to cause hypothalamic obesity in mice [74]	603128	Candidate approach suggests a possible association [76]; association in the Pima Indians where risk allele is major allele [77]
Cocaine- and amphetamine-regulated transcript (CART)	Anorectic peptide activated by leptin [78]; a single family [79] where mutation co-segregated with phenotype	602606	Initial study negative, e.g., [80]; sequencing demonstrated possible association [81]
Mitochondrial			
Uncoupling proteins (nuclear-encoded)	Directing energy to either ATP production or thermogenesis [131]	113730, 602044	REE (racial differences) [22, 23] Lifetime weight gain [132, 133]
Enteroendocrine			
Ghrelin/obestatin (GHRL); growth hormone secretagogue receptor (GHSR)	Growth hormone, secretagogue, role in energy homeostasis [134], may be a role for rare variants [84]; Circulating levels role in long-term weight loss [83]	605353, 601898	Initial work suggesting variation contributing to common obesity not confirmed [85]; later work demonstrates an association of variation in the <i>GHSR</i> and BMI [86]

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Peptide YY (<i>PYY</i>)	Enteroendocrine hormone [88]; rare variant with altered function segregated with obesity in a family [89]	600781	<i>PYY</i> haplotype related to circulating levels and metabolic traits [90]; possible association with obesity in young African-American adults [91]
Fat and glucose utilization			
Peroxisome proliferator activated receptor (PPARs), peroxisome proliferator activated receptor gamma coactivator 1 beta (<i>PPARGCIB</i>)	Lipid sensors, may control adipocyte expansion in relation to BMI [93]	601487, 608886	Association of Pro12Ala variant in <i>PPAR2</i> with obesity in Caucasians [94] and the general population [95], initial observations in <i>PPARGCIB</i> also [96]
Nuclear receptor subfamily 0, group B, member 2 (<i>NR0B2</i>)	Orphan nuclear receptor [97], initial association with HNF4-α caused exploration of DM association	604630	Variation associated with obesity in Japanese [98], but less often in Danish [99]
Ectonucleotide pyrophosphatase/ phosphodiesterase type 1 (ENPP1)	Plasma membrane glycoprotein [135] associated with generalized arterial calcification of infancy, rickets [101, 102]	173335	Ethnic susceptibility to insulin resistance [100], many early studies focus on this aspect of its pathophysiology
Beta-adrenergic receptors (ADRBs)	Beta-adrenergic receptor [103]	109690, 109691	Polymorphisms are frequent, with functional consequences [104], including resistance to catecholamine-induced lipolysis [106]

Evidence for a Genetic Component to Obesity

The obesity epidemic led to a renewed interest in the pathogenesis of obesity. An excess of readily available, highly processed, nutritionally dense foods along with reduced physical activity and increased time spent sitting are all implicated in persistent positive energy balance that leads to overweight [6, 7]. In light of this, the role of genetics remained to be explained in the face of such a rapid change in the prevalence of obesity, especially extreme phenotypes.

One line of thinking invoked the existence of the "thrifty phenotype" originally proposed by Neel in 1962 [8] and cited with increasing frequently as the obesity epidemic progressed e.g., [9]. According to this argument, in prehistoric times there would have been positive selection for traits conferring the ability to store energy efficiently in periods of limited food availability. Alternate views exist, however; although this hypothesis is appealing, it may not explain, for example, observed metabolic responses to physiologic challenges such as famine. In utero epigenetic modifications may produce the observed correlation between intrauterine and postnatal conditions. The "drifty gene" hypothesis has been postulated as well, i.e., that permissive drift is a viable alternative explanation for the existence of genetic variants conferring increased risk for metabolic efficiency and obesity. Clearly, the role of genetics in modifying obesity risk in an era of nutrient excess and deficient activity is complex and far from being completely understood. Estimates of the relative contribution of genetic and individual or shared environmental factors are presented here.

Heritability

The heritability of any condition refers to the proportion of phenotypic variability accounted for by genetics; for a detailed consideration of techniques for heritability estimates, the reader is referred to any number of reviews on the topic e.g., [10]. For a condition that is becoming increasingly prevalent, and apparently occurs more often in relatives, disentangling the contributions of genes and shared adverse environmental influences becomes challenging. Two frequently employed strategies for generating heritability estimates are twin studies and adoption studies.

Twin Studies

The value of twin studies lies in the high degree of genetic similarity between monozygotic (100 %) as compared to nonidentical dizygotic (50 %) twins; both sets of individuals also shared both intrauterine and, to some extent, extrauterine environments. The degree of similarity, or concordance, between monozygotic as compared to dizygotic twins would be expected to be greater in proportion to the

relative importance of their shared genetic information. Indeed, as expected for a heritable trait, estimates of concordance for fat mass between pairs of monozygotic twins are between 70 and 90 %, while for dizygotic twins, they are closer to 35–45 % [11–13]. Fatness is thus considered highly heritable, similar to adult height. The variability in these estimates illustrates a recurrent theme in these studies, that is, the developmental specificity of heritability. In general, estimates of heritability for body mass index (BMI) tend to increase with age, and suggest that interacting gene–environment effects may be cumulative. In another important male twin-pair study, heritability increased from 77 to 84 % over the course of 25 years of longitudinal follow-up [14]. In another study, aggregate data from 23 twin-cohorts demonstrated that heritability for BMI was lowest at birth and increased to over half or more of the variance by as early as 5 months of age [15]. Finally, a study carried out during the obesity epidemic concluded that heritability estimates remained constant, and emphasized the relative importance of individual, non-shared environments [16].

Beyond the biological insights they offer about physiologic regulation of appetite and energy balance, these types of estimates have public health and policy implications, with respect to which individuals to target and by what means to achieve the biggest reduction in overweight and obesity. Adoption studies provide additional information.

Adoption and Family Studies

Adoption studies have provided additional evidence of the genetic contribution to obesity. In one study, adoptees demonstrated more similarity to their biological than their adoptive parents with respect to BMI [17]. In this seminal work, BMI of the biological mother was found to be most closely related to BMI, although a positive association existed for the biological father as well. The authors noted that this association is present across the range of BMI categories (i.e., very lean through obese), pointing to the heritability of low as well as high measures of body fatness. No relationship between adoptees' BMI and the BMI of adoptive parents was identified. In a separate investigation, identical twins raised apart, but not nonidentical twins raised apart, retain some concordance in BMI [18], again indicating the importance of shared genetics in determining common risk for adiposity. Therefore, age-specific methodological effects may be important in such studies.

Racial/Ethnic Differences

Racial and ethnic differences exist in the prevalence of obesity, which also suggests that there may be an effect of shared ancestry, with some populations having an excess risk for obesity, perhaps exacerbated in the context of particular environmental influences. For example, the risk of obesity is 50 % or more in Pima Indians,

conferring an excess risk for diabetes mellitus in this group [19]. By WHO estimates for 2010, the rate of obesity (BMI>30 kg/m²) among men ages 30–100 was substantially less than 1 % in Eritrea but over 80 % in Nauru, a Micronesian island in the South Pacific; similar results were seen in women. (Median percentage of obese individuals over 30 was approximately 12 % for both men and women.)

In the USA, there is variation in the prevalence of obesity by racial group [20], including, for example, ongoing increases in non-Hispanic black and Hispanic women, and a recent Endocrine Society scientific and policy statement emphasized the importance of elucidating the complex interacting social, cultural, biological, and genetic factors that may underlie these differences [21].

Some evidence implicates differences in coupling of oxidative phosphorylation to ATP production (as opposed to thermogenesis, through controlled leak of protons across the mitochondrial matrix) through variation of uncoupling proteins [22, 23]. Although the contribution of mitochondrial variation (either nuclear- or mitochondrialencoded) to obesity is not clear [24], it may be that variation needs to be studied in context of important population and environmental influences including migration patterns, ambient temperature, and altitude [25].

Overall, findings from twin and adoption studies, and differences between racial and ethnic groups do suggest that genetic variation underlies much of the pathogenesis of obesity, but many of the complex mechanism(s) by which this occurs remain elusive.

Previous Genetic Studies in Obesity and the Need for GWAS Approaches

Families with monogenic forms of early onset, apparently isolated childhood obesity have been studied in the context of known candidate genes (see below) and have also yielded loci that remain incompletely characterized. Regarding syndromic obesity (where obesity occurs along with congenital and developmental anomalies), Prader–Willi syndrome [26], Bardet–Biedl syndrome [27–29], and Alström's syndrome [30] are better understood examples, both with respect to underlying molecular mechanisms, and the potential relevance of these for common obesity, as reviewed in the Table 1.1. Overall, candidate gene approaches have yielded some insights prior to the era of GWAS, as summarized by the Table 1.1, but many studies were underpowered to detect more modest effects.

Many candidate genes were chosen for studies on the basis of the known neurophysiology of appetite. The neural circuitry underlying hypothalamic control of appetite and energy balance has been the focus of intensive ongoing research, and seminal work in model systems identified targets for further studies in humans. Indeed, we will describe several important examples of mutations identified in these genes in forms of familial, early-onset morbid obesity where there are either no or only subtle other congenital anomalies or developmental manifestations. These discoveries required comprehensive phenotyping of large numbers of families with apparently isolated severe obesity [31]. We focus in more detail on the genes directly implicated in hypothalamic regulation of appetite, and also include in the Table studies on enteroendocrine regulation of appetite (ghrelin and its receptor, peptide YY) as well as glucose- and lipid-sensing and regulation, including the peroxisome proliferator-activated receptor family (PPARs) and beta-adrenergic receptors.

Hypothalamic Leptin-Melanocortin Pathway

In mice, the *ob* gene is expressed primarily in white fat and encodes a secreted protein called leptin; although this gene was initially cloned and characterized in mice, a human homolog has also identified [32]. Homozygous mutations in the rodent *ob* gene lead to severe obesity, and adult mice outweigh their lean littermates by more than three times [33]. This phenotype is characterized by severe leptin deficiency, and can be rescued by administration of recombinant leptin [34–36]. In contrast, a different strain of mouse with mutations in the primarily hypothalamic leptin receptor (the so-called *db/db* mouse, a model of type 2 diabetes mellitus [37]), exhibit no response to recombinant leptin [38]. Taken together, these results indicate that circulating leptin serves as a homeostatic indicator of the degree of adiposity, and that, physiologically low levels promote food-seeking behavior while high levels inhibit this same behavior. In further support of this proposed mechanism, in humans serum concentrations of leptin do exhibit positive association with obesity [39].

Two children in the same highly consanguineous family with nearly undetectable levels of leptin despite extreme obesity were the first identified human cases of congenital leptin deficiency; they had homozygous frameshift mutations in the leptin gene detected via sequencing of this candidate gene [40]. Later, using a similar strategy, humans with homozygous mutations in the leptin receptor mutation were found [41]. In the latter cases, affected individuals exhibited other endocrinopathies, including hypogonadotropic hypogonadism, and decreased secretion of growth hormone and thyrotropin. When 300 individuals with severe, early-onset obesity all underwent sequencing of the leptin receptor, 3 % had pathogenic mutations, all of which were homozygotes or compound heterozygotes due to a high proportion of individuals from consanguineous families (90 out of 300) [42]. These individuals also displayed hypogonadotropic hypogonadism, as well as defects in immune function.

Although early follow-up studies could not identify evidence for the association between variation within the leptin gene itself and common obesity [43], variation in its 5' region of the gene has been reproducibly associated with propensity for weight loss, as well as common obesity [44]. These pre-GWAS studies suggested some potential sex-dependence of these effects [45]. With respect to the leptin receptor, one small series of Greek individuals identified a modest contribution of variation in the leptin receptor to BMI [46]; like leptin, variation in the leptin receptor may contribute more readily to other physiologic traits like blood pressure [47] or insulin secretion [48]. The difficulty of concluding whether variation in these loci

primarily contribute to common obesity highlights the value of subsequent GWAS studies with much greater power in both discovery and validation cohorts.

As the role of the hypothalamus in regulating appetite and energy balance was investigated further, other components of the so-called leptin-melanocortin pathway were elucidated and characterized, both in model organisms and in humans. The melanocortin family of receptors (MCR) is a G-protein coupled receptor class that, like the leptin receptor, is highly expressed in the hypothalamus and modulates food-seeking behavior. Mice with disruptions in the melanocortin 4 receptor (MC4R) activity are hyperphagic. Unlike the leptin or leptin receptor mutants, obesity develops later in life, and is accompanied by increased, rather than decreased, linear growth; in addition, hyperinsulinemia is also present [49]. Sequencing the gene encoding MC4R in obese humans has led to the discovery of mutations responsible for a co-dominantly inherited form of familial obesity [50-53]. In one seminal series of 500 children with severe, early-onset obesity, 5.8 % were found to have MC4R mutations [54], supporting the conclusion that variation at this locus is the most common genetic cause of obesity identified before the era of GWAS. Similar to the affected rodents, individuals were tall with increased lean mass and hyperinsulinemia. Gene dosage and degree of receptor function modified the phenotype. Later, decreased energy expenditure was also implicated in the etiology of obesity in these individuals [55].

Leptin signaling and MC4R activity are connected via multiple signaling pathways, most notably, pro-opiomelanocortin. Pro-opiomelanocortin is a precursor protein encoded by a gene of the same name. It is cleaved, in part by a prohormone convertase (PC1/3) encoded by the *PCSK1* gene, into melanocortin peptides, including adreoncorticotrophin (ACTH), beta-endorphin, and the melanocyte-stimulating hormones, including anorexigenic α -MSH [56]. This latter is a ligand for MC4R. Multiple ligands for this receptor exist, including agouti-related peptide, an orexigenic peptide.

Expression of POMC is regulated by leptin via activation of its receptor on POMC neurons [57]. Two unrelated individuals of German descent were found to have mutations in *POMC*; their phenotypes included adrenal insufficiency (related to insufficiency in ACTH) and red hair (related to decreased melanin) in addition to early onset obesity [58]. When examined with respect to common obesity, variations in the *POMC* gene have also been related to obesity in individuals of European [59] and Hispanic American [60] descent.

The prohormone convertase 1/3 is encoded by the *PCSK1* gene, and affects processing of other hormones besides POMC, most notably proinsulin, the precursor of insulin C-peptide. A woman who was a compound heterozygote for mutations in PC1 was identified [61] whose phenotype recapitulated that of the so-called *fatlfat* mouse that harbors mutations in the carboxypeptidase E gene, encoding another prohormone convertase. This included early-onset obesity with hyperphagia, pituitary hypofunction (hypogonadotropic hypogonadism and hypocortisolism), as well as disordered glucose homeostasis. Other individuals with mutations in this gene were also described [62]. The consistent role for variation in *PCKS1* in common obesity was not demonstrated until the GWAS era [63].

The initial report of the *MC4R* mutant mouse noted phenotypic similarities to a mouse that overexpresses agouti ("yellow") protein [64], see Table 1.1; disruption of pigment proteins related to POMC leads to this coloring. Variation at the agouti-related protein locus may be related to age-dependent onset of common obesity [65], and there is another possible association of a different variant with reduced fat mass identified in a candidate gene study [66]. The syndecan family of cell surface heparan sulfate proteoglycans can mechanically potentiate the ability of agouti-related protein to inhibit α -MSH; transgenetic alteration of the endogenous hypothalamic syndecans in mice can lead to hyperphagia and maturity-onset obesity suggesting reduced α -MSH signaling [67].

Rodent studies of downstream targets of MC4R have identified brain-derived neurotrophic factor (BDNF), a nerve growth factor expressed in the ventromedial hypothalamus. It has been shown to modulate appetite and energy balance in response to MC4R signaling [68]. Animals missing BDNF, hyperinsulinemic, grow rapidly, and have increased locomotor activity, and the phenotype is rescued with central infusion of BDNF [69]. In humans, loss of only one copy of the *BDNF* gene leads to obesity, hyperphagia, intellectual impairment and hyperactivity [70]. Indeed, it may be haploinsufficiency of *BDNF* that leads to the childhood onset obesity in some individuals with WAGR syndrome (Wilms' tumor, aniridia, genito-urinary abnormalities, and mental retardation) if the extent of the responsible deletion includes that gene [71]. In additional support for the role of this effector, a de novo mutation in a tyrosine kinase receptor downstream of BDNF also produces a similar phenotype [72]. The role for variation in *BDNF* in common obesity per se was not clearly demonstrated, although it was found to be associated with neuropsychiatric conditions also characterized by disordered eating (e.g., [73]).

Sim1 (Single-minded, drosophila, homolog) is another gene whose disruption and/or haploinsufficiency [74] have been reported to cause hypothalamic obesity with hyperphagia. Sim1 (+/–) heterozygous mice demonstrate high levels of leptin, in keeping with their elevated fat mass, as well as hyperinsulinemia and increased linear growth. Human studies demonstrate similar results. A de novo balanced translocation disrupting Sim1 was observed in a girl with severe early-onset obesity and normal resting energy expenditure [75]. Mechanisms invoked have included decreased cellularity of the paraventricular nucleus (PVN) whose development is regulated by Sim1; PVN cells also express MC4R that may be regulated by α -MSH. A candidate gene study suggested a possible association of common variants in Sim1 with BMI and weight gain in individuals of European descent [76]. An association was also identified in the Pima Indians where the risk allele is the major allele [77].

Another focus of investigation has been the *CART* gene (Cocaine- and Amphetamine-Regulated Transcript), which encodes an anorectic peptide activated by leptin [78]. One family has been identified [79] where a mutation in this gene co-segregated with an extreme obesity phenotype. An initial study in Europeans did not find an association with obesity, but did find a relationship with metabolic traits that mediated its effect via fat distribution (waist-to-hip ratio) [80]. Sequencing of the putative promoter region also identified variants that were associated with BMI and, in particular, extreme obesity [81].

Enteroendocrine Hormones

Ghrelin was discovered initially as the endogenous ligand for the growth hormone secretagogue receptor; studies in rodents demonstrated that, like leptin, ghrelin responds to a peripheral signal regarding nutrient availability and appears to convey this information to the hypothalamus. Ghrelin is produced in the stomach, and its level rises with fasting and promotes increased food seeking, as well as decreased fat utilization, and is hypothesized to reflect a signal to increase metabolic efficiency in times of low nutrient availability [82]. Circulating levels of ghrelin have been shown to play a role in long-term weight loss, likely by helping to homeostatically defend a metabolic "set-point" [83]. In one study, 6.3 % of severely obese adult women of European descent were found to be heterozygotes for an amino acid change in the last residue in mature ghrelin; this change was not found in controls, but the functional significance of this change was not confirmed [85]; later work, however, demonstrates an association of variation in the gene encoding ghrelin's receptor (*GHSR*) and BMI [86].

Peptide YY (PYY) is an enteroendocrine hormone present in highest concentrations in the ileum and colon that rises in response to food ingestion, in particular, of fat-containing foods [87]. In addition to inhibiting gastric and pancreatic secretion, it is also expressed in some its target tissues suggesting a paracrine feedback mechanism in addition to its known endocrine function [88]. Rare variants in the *PYY* gene with altered function have been reported to segregate with severe obesity in one family [89]. In addition, a *PYY* haplotype has been related to circulating PYY levels as well as metabolic traits [90]. Later studies have demonstrated a possible association with obesity, particularly in young African-American adults [91].

Fat and Glucose Utilization

The peroxisome proliferator activated receptor family (PPARs) is a class of nuclear hormone-sensing transcription factors that convey signals about nutrient availability and coordinate the appropriate cellular metabolic responses [92]. Of particular relevance for obesity, subsets of PPARs act as lipid sensors and may control adipocyte expansion in relation to BMI [93], and are also the target of insulin-sensitizing thiazolidinediones [92]. In recognition of their importance in lipid homeostasis, variation in the genes encoding PPARs has been examined with respect to adiposity. An early finding of an association of the Pro12Ala variant in *PPAR* γ with obesity in Caucasians [94] was later also demonstrated in the general population [95]. Initial observations about the relevance of variation in one of the PPAR coactivator genes (*PPARGC1B*) have also been published [96].

Another family of nuclear receptors, the nuclear receptor subfamily 0, group B, member 2 (NR0B2), has also been the subject of investigation. The gene encodes an orphan nuclear receptor [97] that is known to interact with HNF4- α , one of the genes mutated in a monogenic form of diabetes. As a result, variation in this gene

was initially studied with respect to risk for diabetes mellitus, but was found instead to confer risk based on its association with obesity. Specifically, in initial studies, variation was associated with obesity in a Japanese population [98], but less often in Danish individuals [99]. Its relationship with insulin resistance [100] was also the motivation to study ectonucleotide pyrophosphatase/phosphodiesterase type 1 (ENPP1), a plasma membrane glycoprotein that may be associated with obesity. Mutations in the gene encoding this glycoprotein have been identified in patients with generalized arterial calcification of infancy and rickets [101, 102]. Its relationship to common obesity remains to be fully characterized.

Another receptor that influences fuel use in response to circulating signals is the beta-adrenergic receptor [103], which coordinates response to catecholamines as part of the physiologic stress response. Polymorphisms in this receptor are frequent, with functional consequences [104]; while other investigations have focused on asthma phenotypes, in particular, response to sympathomimetic bronchodilator therapy, catecholamine-induced lipolysis is relevant in determining obesity risk. In the longitudinal Bogalusa Heart Study, a cohort of individuals of European and African-American descent, males with the Arg16Gly polymorphism demonstrated an increased association of BMI with age over time [105]. This result also demonstrates the developmental specificity of testing for association of genetic variants with BMI. The functional consequence of differential expression, and presumably, receptor action as well, includes resistance to catecholamine-induced lipolysis [106].

Reviews written prior to the era of GWAS describe the success of approaches used to identify monogenic disorders (linkage and candidate gene studies), but acknowledge the limitations of these strategies in addressing the complex pathogenesis of multigenic traits like common obesity, e.g., [7]. Subsequent investigative strategies have varying capacities to identify the "missing heritability," or the genetic variation that remains to be explained after accounting for what is known. With respect to variation in nuclear DNA, the hypothesized risk allele frequency and effect size are important determinants of experimental power of association studies [107]. Other factors, including more complex structural variation like copy-number variants, or epigenetic modification, may require unique approaches.

Interestingly, some of the first GWAS fulfilled the promise of these innovative technologies, but also highlighted new challenges for discovery of novel biologic pathways and explanation of additional variation. One of the first GWAS of BMI identified variation near INSIG2, a gene that encodes a protein that interacts with sterol regulator element binding proteins (SREBPs), transcription factors that control the reverse transport of cholesterol and therefore may have functional significance as well [108]. Inconsistencies and difficulties with replication efforts have been instructive, and remain the subject of ongoing investigation [109]. The first consistently reproducible signal from GWAS for obesity came instead from investigations into type 2 diabetes mellitus; variation near the FTO (fat-mass and obesity associated) locus affected diabetes risk by modulating BMI [110-112]. No matter the strategy to identify additional sources of genetic variation, the functional significance of novel findings, interpreted in the context of what is already understood about the complex regulation of energy homeostasis, will continue to be of primary importance.

References

- 1. James WP (2008) WHO recognition of the global obesity epidemic. Int J Obes (Lond) 32(Suppl 7):S120–S126, Epub 2009/01/16
- Austin GL, Ogden LG, Hill JO (2011) Trends in carbohydrate, fat, and protein intakes and association with energy intake in normal-weight, overweight, and obese individuals: 1971– 2006. Am J Clin Nutr 93(4):836–843, Epub 2011/02/12
- Ogden CL, Carroll MD, Kit BK, Flegal KM (2012) Prevalence of obesity and trends in body mass index among US children and adolescents, 1999–2010. JAMA 307(5):483–490, Epub 2012/01/19
- Wells JC (2012) The evolution of human adiposity and obesity: where did it all go wrong? Dis Model Mech 5(5):595–607, Epub 2012/08/24
- Perusse L, Rankinen T, Zuberi A, Chagnon YC, Weisnagel SJ, Argyropoulos G et al (2005) The human obesity gene map: the 2004 update. Obes Res 13(3):381–490, Epub 2005/04/19
- 6. Friedman JM (2004) Modern science versus the stigma of obesity. Nat Med 10(6):563–569, Epub 2004/06/01
- Lyon HN, Hirschhorn JN (2005) Genetics of common forms of obesity: a brief overview. Am J Clin Nutr 82(1 Suppl):215S–217S, Epub 2005/07/09
- Neel JV (1962) Diabetes mellitus: a "thrifty" genotype rendered detrimental by "progress"? Am J Hum Genet 14:353–362, Epub 1962/12/01
- Baig U, Belsare P, Watve M, Jog M (2011) Can thrifty gene(s) or predictive fetal programming for thriftiness lead to obesity? J Obes 2011:861049, Epub 2011/07/21
- Tenesa A, Haley CS (2013) The heritability of human disease: estimation, uses and abuses. Nat Rev Genet 14(2):139–149, Epub 2013/01/19
- Hebebrand J, Friedel S, Schauble N, Geller F, Hinney A (2003) Perspectives: molecular genetic research in human obesity. Obes Rev 4(3):139–146, Epub 2003/08/15
- Farooqi IS, O'Rahilly S (2005) New advances in the genetics of early onset obesity. Int J Obes (Lond) 29(10):1149–1152, Epub 2005/09/13
- 13. Bell CG, Walley AJ, Froguel P (2005) The genetics of human obesity. Nat Rev Genet 6(3):221–234, Epub 2005/02/11
- Stunkard AJ, Foch TT, Hrubec Z (1986) A twin study of human obesity. JAMA 256(1):51–54, Epub 1986/07/04
- 15. Dubois L, Ohm Kyvik K, Girard M, Tatone-Tokuda F, Perusse D, Hjelmborg J et al (2012) Genetic and environmental contributions to weight, height, and BMI from birth to 19 years of age: an international study of over 12,000 twin pairs. PLoS One 7(2):e30153, Epub 2012/02/22
- Wardle J, Carnell S, Haworth CM, Plomin R (2008) Evidence for a strong genetic influence on childhood adiposity despite the force of the obesogenic environment. Am J Clin Nutr 87(2):398–404, Epub 2008/02/09
- Stunkard AJ, Sorensen TI, Hanis C, Teasdale TW, Chakraborty R, Schull WJ et al (1986) An adoption study of human obesity. N Engl J Med 314(4):193–198, Epub 1986/01/23
- Stunkard AJ, Harris JR, Pedersen NL, McClearn GE (1990) The body-mass index of twins who have been reared apart. N Engl J Med 322(21):1483–1487, Epub 1990/05/24
- 19. Knowler WC, Pettitt DJ, Saad MF, Bennett PH (1990) Diabetes mellitus in the Pima Indians: incidence, risk factors and pathogenesis. Diabetes Metab Rev 6(1):1–27, Epub 1990/02/01
- Flegal KM, Carroll MD, Kit BK, Ogden CL (2012) Prevalence of obesity and trends in the distribution of body mass index among US adults, 1999–2010. JAMA 307(5):491–497, Epub 2012/01/19
- Golden SH, Brown A, Cauley JA, Chin MH, Gary-Webb TL, Kim C et al (2012) Health disparities in endocrine disorders: biological, clinical, and nonclinical factors—an Endocrine Society scientific statement. J Clin Endocrinol Metab 97(9):E1579–E1639, Epub 2012/06/26
- 22. Kimm SY, Glynn NW, Aston CE, Damcott CM, Poehlman ET, Daniels SR et al (2002) Racial differences in the relation between uncoupling protein genes and resting energy expenditure. Am J Clin Nutr 75(4):714–719, Epub 2002/03/28

- Yanovski JA, Diament AL, Sovik KN, Nguyen TT, Li H, Sebring NG et al (2000) Associations between uncoupling protein 2, body composition, and resting energy expenditure in lean and obese African American, white, and Asian children. Am J Clin Nutr 71(6):1405–1420, Epub 2000/06/06
- 24. Saxena R, de Bakker PI, Singer K, Mootha V, Burtt N, Hirschhorn JN et al (2006) Comprehensive association testing of common mitochondrial DNA variation in metabolic disease. Am J Hum Genet 79(1):54–61, Epub 2006/06/15
- Wallace DC (2011) Bioenergetic origins of complexity and disease. Cold Spring Harb Symp Quant Biol 76:1–16, Epub 2011/12/24
- 26. Kondo I, Hamabe J, Yamamoto K, Niikawa N (1990) Exclusion mapping of the Cohen syndrome gene from the Prader-Willi syndrome locus. Clin Genet 38(6):422–426
- 27. Beales PL, Warner AM, Hitman GA, Thakker R, Flinter FA (1997) Bardet-Biedl syndrome: a molecular and phenotypic study of 18 families. J Med Genet 34(2):92–98
- Bruford EA, Riise R, Teague PW, Porter K, Thomson KL, Moore AT et al (1997) Linkage mapping in 29 Bardet-Biedl syndrome families confirms loci in chromosomal regions 11q13, 15q22.3-q23, and 16q21. Genomics 41(1):93–99
- 29. Young TL, Penney L, Woods MO, Parfrey PS, Green JS, Hefferton D et al (1999) A fifth locus for Bardet-Biedl syndrome maps to chromosome 2q31. Am J Hum Genet 64(3):900–904
- 30. Russell-Eggitt IM, Clayton PT, Coffey R, Kriss A, Taylor DS, Taylor JF (1998) Alstrom syndrome. Report of 22 cases and literature review. Ophthalmology 105(7):1274–1280
- Farooqi IS, O'Rahilly S (2000) Recent advances in the genetics of severe childhood obesity. Arch Dis Child 83(1):31–34
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM (1994) Positional cloning of the mouse obese gene and its human homologue. Nature 372(6505):425–432, Epub 1994/12/01
- 33. Ingalls AM, Dickie MM, Snell GD (1950) Obese, a new mutation in the house mouse. J Hered 41(12):317–318
- 34. Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D et al (1995) Weightreducing effects of the plasma protein encoded by the obese gene. Science 269(5223):543–546
- 35. Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P (1995) Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. Science 269(5223):546–549
- 36. Pelleymounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T et al (1995) Effects of the obese gene product on body weight regulation in ob/ob mice. Science 269(5223):540–543
- Lee GH, Proenca R, Montez JM, Carroll KM, Darvishzadeh JG, Lee JI et al (1996) Abnormal splicing of the leptin receptor in diabetic mice. Nature 379(6566):632–635, Epub 1996/02/15
- Chua SC Jr, Chung WK, Wu-Peng XS, Zhang Y, Liu SM, Tartaglia L et al (1996) Phenotypes of mouse diabetes and rat fatty due to mutations in the OB (leptin) receptor. Science 271(5251):994–996
- Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR et al (1996) Serum immunoreactive-leptin concentrations in normal-weight and obese humans. N Engl J Med 334(5):292–295
- 40. Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ et al (1997) Congenital leptin deficiency is associated with severe early-onset obesity in humans. Nature 387(6636):903–908, Epub 1997/06/26
- Clement K, Vaisse C, Lahlou N, Cabrol S, Pelloux V, Cassuto D et al (1998) A mutation in the human leptin receptor gene causes obesity and pituitary dysfunction. Nature 392(6674):398–401, Epub 1998/04/16
- Farooqi IS, Wangensteen T, Collins S, Kimber W, Matarese G, Keogh JM et al (2007) Clinical and molecular genetic spectrum of congenital deficiency of the leptin receptor. N Engl J Med 356(3):237–247, Epub 2007/01/19

- 43. Lucantoni R, Ponti E, Berselli ME, Savia G, Minocci A, Calo G et al (2000) The A19G polymorphism in the 5' untranslated region of the human obese gene does not affect leptin levels in severely obese patients. J Clin Endocrinol Metab 85(10):3589–3591, Epub 2000/11/04
- 44. Mammes O, Betoulle D, Aubert R, Herbeth B, Siest G, Fumeron F (2000) Association of the G-2548A polymorphism in the 5' region of the LEP gene with overweight. Ann Hum Genet 64(Pt 5):391–394, Epub 2001/04/03
- 45. Jiang Y, Wilk JB, Borecki I, Williamson S, DeStefano AL, Xu G et al (2004) Common variants in the 5' region of the leptin gene are associated with body mass index in men from the National Heart, Lung, and Blood Institute Family Heart Study. Am J Hum Genet 75(2):220–230, Epub 2004/06/16
- 46. Yiannakouris N, Yannakoulia M, Melistas L, Chan JL, Klimis-Zacas D, Mantzoros CS (2001) The Q223R polymorphism of the leptin receptor gene is significantly associated with obesity and predicts a small percentage of body weight and body composition variability. J Clin Endocrinol Metab 86(9):4434–4439, Epub 2001/09/11
- 47. Rosmond R, Chagnon YC, Holm G, Chagnon M, Perusse L, Lindell K et al (2000) Hypertension in obesity and the leptin receptor gene locus. J Clin Endocrinol Metab 85(9):3126–3131, Epub 2000/09/22
- 48. Wauters M, Mertens I, Rankinen T, Chagnon M, Bouchard C, Van Gaal L (2001) Leptin receptor gene polymorphisms are associated with insulin in obese women with impaired glucose tolerance. J Clin Endocrinol Metab 86(7):3227–3232, Epub 2001/07/10
- 49. Huszar D, Lynch CA, Fairchild-Huntress V, Dunmore JH, Fang Q, Berkemeier LR et al (1997) Targeted disruption of the melanocortin-4 receptor results in obesity in mice. Cell 88(1):131–141, Epub 1997/01/10
- Yeo GS, Farooqi IS, Aminian S, Halsall DJ, Stanhope RG, O'Rahilly S (1998) A frameshift mutation in MC4R associated with dominantly inherited human obesity. Nat Genet 20(2):111–112, Epub 1998/10/15
- 51. Vaisse C, Clement K, Guy-Grand B, Froguel P (1998) A frameshift mutation in human MC4R is associated with a dominant form of obesity. Nat Genet 20(2):113–114, Epub 1998/10/15
- 52. Gu W, Tu Z, Kleyn PW, Kissebah A, Duprat L, Lee J et al (1999) Identification and functional analysis of novel human melanocortin-4 receptor variants. Diabetes 48(3):635–639, Epub 1999/03/17
- 53. Hinney A, Schmidt A, Nottebom K, Heibult O, Becker I, Ziegler A et al (1999) Several mutations in the melanocortin-4 receptor gene including a nonsense and a frameshift mutation associated with dominantly inherited obesity in humans. J Clin Endocrinol Metab 84(4):1483– 1486, Epub 1999/04/13
- Farooqi IS, Keogh JM, Yeo GS, Lank EJ, Cheetham T, O'Rahilly S (2003) Clinical spectrum of obesity and mutations in the melanocortin 4 receptor gene. N Engl J Med 348(12):1085– 1095, Epub 2003/03/21
- 55. Krakoff J, Ma L, Kobes S, Knowler WC, Hanson RL, Bogardus C et al (2008) Lower metabolic rate in individuals heterozygous for either a frameshift or a functional missense MC4R variant. Diabetes 57(12):3267–3272, Epub 2008/10/07
- 56. Seidah NG, Benjannet S, Hamelin J, Mamarbachi AM, Basak A, Marcinkiewicz J et al (1999) The subtilisin/kexin family of precursor convertases. Emphasis on PC1, PC2/7B2, POMC and the novel enzyme SKI-1. Ann N Y Acad Sci 885:57–74
- Cheung CC, Clifton DK, Steiner RA (1997) Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. Endocrinology 138(10):4489–4492
- Krude H, Biebermann H, Luck W, Horn R, Brabant G, Gruters A (1998) Severe early-onset obesity, adrenal insufficiency and red hair pigmentation caused by POMC mutations in humans. Nat Genet 19(2):155–157, Epub 1998/06/10
- 59. Chen Y, Snieder H, Wang X, Kaviya B, McCaffrey C, Spector TD et al (2005) Proopiomelanocortin gene variants are associated with serum leptin and body fat in a normal female population. Eur J Hum Genet 13(6):772–780, Epub 2005/04/07
- 60. Sutton BS, Langefeld CD, Williams AH, Norris JM, Saad MF, Haffner SM et al (2005) Association of proopiomelanocortin gene polymorphisms with obesity in the IRAS family study. Obes Res 13(9):1491–1498, Epub 2005/10/14

- 61. Jackson RS, Creemers JW, Ohagi S, Raffin-Sanson ML, Sanders L, Montague CT et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 16(3):303–306, Epub 1997/07/01
- 62. Farooqi IS, Volders K, Stanhope R, Heuschkel R, White A, Lank E et al (2007) Hyperphagia and early-onset obesity due to a novel homozygous missense mutation in prohormone convertase 1/3. J Clin Endocrinol Metab 92(9):3369–3373, Epub 2007/06/28
- Benzinou M, Creemers JW, Choquet H, Lobbens S, Dina C, Durand E et al (2008) Common nonsynonymous variants in PCSK1 confer risk of obesity. Nat Genet 40(8):943–945, Epub 2008/07/08
- Fan W, Boston BA, Kesterson RA, Hruby VJ, Cone RD (1997) Role of melanocortinergic neurons in feeding and the agouti obesity syndrome. Nature 385(6612):165–168, Epub 1997/01/09
- 65. Argyropoulos G, Rankinen T, Neufeld DR, Rice T, Province MA, Leon AS et al (2002) A polymorphism in the human agouti-related protein is associated with late-onset obesity. J Clin Endocrinol Metab 87(9):4198–4202, Epub 2002/09/06
- 66. Marks DL, Boucher N, Lanouette CM, Perusse L, Brookhart G, Comuzzie AG et al (2004) Ala67Thr polymorphism in the Agouti-related peptide gene is associated with inherited leanness in humans. Am J Med Genet A 126A(3):267–271, Epub 2004/04/01
- Reizes O, Lincecum J, Wang Z, Goldberger O, Huang L, Kaksonen M et al (2001) Transgenic expression of syndecan-1 uncovers a physiological control of feeding behavior by syndecan-3. Cell 106(1):105–116, Epub 2001/07/20
- Xu B, Goulding EH, Zang K, Cepoi D, Cone RD, Jones KR et al (2003) Brain-derived neurotrophic factor regulates energy balance downstream of melanocortin-4 receptor. Nat Neurosci 6(7):736–742, Epub 2003/06/11
- 69. Rios M, Fan G, Fekete C, Kelly J, Bates B, Kuehn R et al (2001) Conditional deletion of brain-derived neurotrophic factor in the postnatal brain leads to obesity and hyperactivity. Mol Endocrinol 15(10):1748–1757, Epub 2001/10/02
- 70. Gray J, Yeo GS, Cox JJ, Morton J, Adlam AL, Keogh JM et al (2006) Hyperphagia, severe obesity, impaired cognitive function, and hyperactivity associated with functional loss of one copy of the brain-derived neurotrophic factor (BDNF) gene. Diabetes 55(12):3366–3371, Epub 2006/11/30
- Han JC, Liu QR, Jones M, Levinn RL, Menzie CM, Jefferson-George KS et al (2008) Brainderived neurotrophic factor and obesity in the WAGR syndrome. N Engl J Med 359(9):918– 927, Epub 2008/08/30
- 72. Yeo GS, Connie Hung CC, Rochford J, Keogh J, Gray J, Sivaramakrishnan S et al (2004) A de novo mutation affecting human TrkB associated with severe obesity and developmental delay. Nat Neurosci 7(11):1187–1189, Epub 2004/10/21
- 73. Ribases M, Gratacos M, Fernandez-Aranda F, Bellodi L, Boni C, Anderluh M et al (2004) Association of BDNF with anorexia, bulimia and age of onset of weight loss in six European populations. Hum Mol Genet 13(12):1205–1212, Epub 2004/04/30
- Michaud JL, Boucher F, Melnyk A, Gauthier F, Goshu E, Levy E et al (2001) Sim1 haploinsufficiency causes hyperphagia, obesity and reduction of the paraventricular nucleus of the hypothalamus. Hum Mol Genet 10(14):1465–1473, Epub 2001/07/13
- Holder JL Jr, Butte NF, Zinn AR (2000) Profound obesity associated with a balanced translocation that disrupts the SIM1 gene. Hum Mol Genet 9(1):101–108, Epub 1999/12/10
- 76. Hung CC, Luan J, Sims M, Keogh JM, Hall C, Wareham NJ et al (2007) Studies of the SIM1 gene in relation to human obesity and obesity-related traits. Int J Obes (Lond) 31(3):429–434, Epub 2006/08/23
- 77. Traurig M, Mack J, Hanson RL, Ghoussaini M, Meyre D, Knowler WC et al (2009) Common variation in SIM1 is reproducibly associated with BMI in Pima Indians. Diabetes 58(7):1682–1689, Epub 2009/04/30
- Kristensen P, Judge ME, Thim L, Ribel U, Christjansen KN, Wulff BS et al (1998) Hypothalamic CART is a new anorectic peptide regulated by leptin. Nature 393(6680):72–76, Epub 1998/05/20
- 79. del Giudice EM, Santoro N, Cirillo G, D'Urso L, Di Toro R, Perrone L (2001) Mutational screening of the CART gene in obese children: identifying a mutation (Leu34Phe) associated

with reduced resting energy expenditure and cosegregating with obesity phenotype in a large family. Diabetes 50(9):2157–2160, Epub 2001/08/28

- Challis BG, Yeo GS, Farooqi IS, Luan J, Aminian S, Halsall DJ et al (2000) The CART gene and human obesity: mutational analysis and population genetics. Diabetes 49(5):872–875, Epub 2000/07/25
- Yamada K, Yuan X, Otabe S, Koyanagi A, Koyama W, Makita Z (2002) Sequencing of the putative promoter region of the cocaine- and amphetamine-regulated-transcript gene and identification of polymorphic sites associated with obesity. Int J Obes Relat Metab Disord 26(1):132–136, Epub 2002/01/16
- Tschop M, Smiley DL, Heiman ML (2000) Ghrelin induces adiposity in rodents. Nature 407(6806):908–913, Epub 2000/11/01
- Cummings DE, Weigle DS, Frayo RS, Breen PA, Ma MK, Dellinger EP et al (2002) Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. N Engl J Med 346(21):1623–1630, Epub 2002/05/25
- 84. Ukkola O, Ravussin E, Jacobson P, Snyder EE, Chagnon M, Sjostrom L et al (2001) Mutations in the preproghrelin/ghrelin gene associated with obesity in humans. J Clin Endocrinol Metab 86(8):3996–3999, Epub 2001/08/15
- 85. Hinney A, Hoch A, Geller F, Schafer H, Siegfried W, Goldschmidt H et al (2002) Ghrelin gene: identification of missense variants and a frameshift mutation in extremely obese children and adolescents and healthy normal weight students. J Clin Endocrinol Metab 87(6):2716, Epub 2002/06/07
- 86. Baessler A, Hasinoff MJ, Fischer M, Reinhard W, Sonnenberg GE, Olivier M et al (2005) Genetic linkage and association of the growth hormone secretagogue receptor (ghrelin receptor) gene in human obesity. Diabetes 54(1):259–267, Epub 2004/12/24
- Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR (1985) Human distribution and release of a putative new gut hormone, peptide YY. Gastroenterology 89(5):1070–1077, Epub 1985/11/01
- Leiter AB, Toder A, Wolfe HJ, Taylor IL, Cooperman S, Mandel G et al (1987) Peptide YY. Structure of the precursor and expression in exocrine pancreas. J Biol Chem 262(27):12984– 12988, Epub 1987/09/25
- Ahituv N, Kavaslar N, Schackwitz W, Ustaszewska A, Collier JM, Hebert S et al (2006) A PYY Q62P variant linked to human obesity. Hum Mol Genet 15(3):387–391, Epub 2005/12/22
- 90. Shih PA, Wang L, Chiron S, Wen G, Nievergelt C, Mahata M et al (2009) Peptide YY (PYY) gene polymorphisms in the 3'-untranslated and proximal promoter regions regulate cellular gene expression and PYY secretion and metabolic syndrome traits in vivo. J Clin Endocrinol Metab 94(11):4557–4566, Epub 2009/10/13
- Friedlander Y, Li G, Fornage M, Williams OD, Lewis CE, Schreiner P et al (2010) Candidate molecular pathway genes related to appetite regulatory neural network, adipocyte homeostasis and obesity: results from the CARDIA Study. Ann Hum Genet 74(5):387–398, Epub 2010/07/21
- Schupp M, Lazar MA (2010) Endogenous ligands for nuclear receptors: digging deeper. J Biol Chem 285(52):40409–40415, Epub 2010/10/20
- Sewter C, Blows F, Considine R, Vidal-Puig A, O'Rahilly S (2002) Differential effects of adiposity on peroxisomal proliferator-activated receptor gamma1 and gamma2 messenger ribonucleic acid expression in human adipocytes. J Clin Endocrinol Metab 87(9):4203–4207, Epub 2002/09/06
- 94. Beamer BA, Yen CJ, Andersen RE, Muller D, Elahi D, Cheskin LJ et al (1998) Association of the Pro12Ala variant in the peroxisome proliferator-activated receptor-gamma2 gene with obesity in two Caucasian populations. Diabetes 47(11):1806–1808, Epub 1998/10/29
- 95. Deeb SS, Fajas L, Nemoto M, Pihlajamaki J, Mykkanen L, Kuusisto J et al (1998) A Pro12Ala substitution in PPARgamma2 associated with decreased receptor activity, lower body mass index and improved insulin sensitivity. Nat Genet 20(3):284–287, Epub 1998/11/07
- 96. Andersen G, Wegner L, Yangisawa K, Rose CS, Lin J, Glumer C et al (2005) Evidence of an association between genetic variation of the coactivator PGC-1beta and obesity. J Med Genet 42(5):402–7

- Seol W, Choi HS, Moore DD (1996) An orphan nuclear hormone receptor that lacks a DNA binding domain and heterodimerizes with other receptors. Science 272(5266):1336–1339, Epub 1996/05/31
- Nishigori H, Tomura H, Tonooka N, Kanamori M, Yamada S, Sho K et al (2001) Mutations in the small heterodimer partner gene are associated with mild obesity in Japanese subjects. Proc Natl Acad Sci U S A 98(2):575–580, Epub 2001/01/03
- 99. Echwald SM, Andersen KL, Sorensen TI, Larsen LH, Andersen T, Tonooka N et al (2004) Mutation analysis of NR0B2 among 1545 Danish men identifies a novel c.278G>A (p.G93D) variant with reduced functional activity. Hum Mutat 24(5):381–387
- 100. Abate N, Carulli L, Cabo-Chan A Jr, Chandalia M, Snell PG, Grundy SM (2003) Genetic polymorphism PC-1 K121Q and ethnic susceptibility to insulin resistance. J Clin Endocrinol Metab 88(12):5927–5934, Epub 2003/12/13
- 101. Dlamini N, Splitt M, Durkan A, Siddiqui A, Padayachee S, Hobbins S et al (2009) Generalized arterial calcification of infancy: phenotypic spectrum among three siblings including one case without obvious arterial calcifications. Am J Med Genet A 149A(3):456–460, Epub 2009/02/12
- 102. Lorenz-Depiereux B, Schnabel D, Tiosano D, Hausler G, Strom TM (2010) Loss-of-function ENPP1 mutations cause both generalized arterial calcification of infancy and autosomalrecessive hypophosphatemic rickets. Am J Hum Genet 86(2):267–272, Epub 2010/02/09
- 103. Yang-Feng TL, Xue FY, Zhong WW, Cotecchia S, Frielle T, Caron MG et al (1990) Chromosomal organization of adrenergic receptor genes. Proc Natl Acad Sci U S A 87(4):1516–1520, Epub 1990/02/01
- 104. Large V, Hellstrom L, Reynisdottir S, Lonnqvist F, Eriksson P, Lannfelt L et al (1997) Human beta-2 adrenoceptor gene polymorphisms are highly frequent in obesity and associate with altered adipocyte beta-2 adrenoceptor function. J Clin Invest 100(12):3005–3013, Epub 1998/01/31
- 105. Ellsworth DL, Coady SA, Chen W, Srinivasan SR, Elkasabany A, Gustat J et al (2002) Influence of the beta2-adrenergic receptor Arg16Gly polymorphism on longitudinal changes in obesity from childhood through young adulthood in a biracial cohort: the Bogalusa Heart Study. Int J Obes Relat Metab Disord 26(7):928–937, Epub 2002/06/25
- Lonnqvist F, Wahrenberg H, Hellstrom L, Reynisdottir S, Arner P (1992) Lipolytic catecholamine resistance due to decreased beta 2-adrenoceptor expression in fat cells. J Clin Invest 90(6):2175–2186, Epub 1992/12/01
- 107. Blakemore AI, Froguel P (2008) Is obesity our genetic legacy? J Clin Endocrinol Metab 93(11 Suppl 1):S51–S56, Epub 2008/12/04
- 108. Herbert A, Gerry NP, McQueen MB, Heid IM, Pfeufer A, Illig T et al (2006) A common genetic variant is associated with adult and childhood obesity. Science 312(5771):279–283, Epub 2006/04/15
- 109. Heid IM, Huth C, Loos RJ, Kronenberg F, Adamkova V, Anand SS et al (2009) Meta-analysis of the INSIG2 association with obesity including 74,345 individuals: does heterogeneity of estimates relate to study design? PLoS Genet 5(10):e1000694, Epub 2009/10/24
- 110. Dina C, Meyre D, Gallina S, Durand E, Korner A, Jacobson P et al (2007) Variation in FTO contributes to childhood obesity and severe adult obesity. Nat Genet 39(6):724–726, Epub 2007/05/15
- 111. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, Lindgren CM et al (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316(5826):889–894, Epub 2007/04/17
- 112. Gerken T, Girard CA, Tung YC, Webby CJ, Saudek V, Hewitson KS et al (2007) The obesityassociated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. Science 318(5855):1469–1472, Epub 2007/11/10
- 113. Baker M, Gaukrodger N, Mayosi BM, Imrie H, Farrall M, Watkins H et al (2005) Association between common polymorphisms of the proopiomelanocortin gene and body fat distribution: a family study. Diabetes 54(8):2492–2496, Epub 2005/07/28
- 114. Nicholls RD, Knoll JH, Butler MG, Karam S, Lalande M (1989) Genetic imprinting suggested by maternal heterodisomy in nondeletion Prader-Willi syndrome. Nature 342(6247):281–285, Epub 1989/11/16

- 115. Chen Y, Liu YJ, Pei YF, Yang TL, Deng FY, Liu XG et al (2011) Copy number variations at the Prader-Willi syndrome region on chromosome 15 and associations with obesity in whites. Obesity (Silver Spring) 19(6):1229–1234, Epub 2011/01/15
- 116. Oeffner F, Korn T, Roth H, Ziegler A, Hinney A, Goldschmidt H et al (2001) Systematic screening for mutations in the human necdin gene (NDN): identification of two naturally occurring polymorphisms and association analysis in body weight regulation. Int J Obes Relat Metab Disord 25(6):767–769, Epub 2001/07/06
- 117. Jin H, White SR, Shida T, Schulz S, Aguiar M, Gygi SP et al (2010) The conserved Bardet-Biedl syndrome proteins assemble a coat that traffics membrane proteins to cilia. Cell 141(7):1208–1219, Epub 2010/07/07
- 118. Rahmouni K, Fath MA, Seo S, Thedens DR, Berry CJ, Weiss R et al (2008) Leptin resistance contributes to obesity and hypertension in mouse models of Bardet-Biedl syndrome. J Clin Invest 118(4):1458–1467, Epub 2008/03/05
- 119. Benzinou M, Walley A, Lobbens S, Charles MA, Jouret B, Fumeron F et al (2006) Bardet-Biedl syndrome gene variants are associated with both childhood and adult common obesity in French Caucasians. Diabetes 55(10):2876–2882, Epub 2006/09/28
- 120. Andersen KL, Echwald SM, Larsen LH, Hamid YH, Glumer C, Jorgensen T et al (2005) Variation of the McKuisck-Kaufman gene and studies of relationships with common forms of obesity. J Clin Endo Metab 90(1):225–30, Epub 2004/10/13
- 121. Sorensen TI, Boutin P, Taylor MA, Larsen LH, Verdich C, Petersen L et al (2006) Genetic polymorphisms and weight loss in obesity: a randomised trial of hypo-energetic high- versus low-fat diets. PLoS Clin Trials 1(2):e12, Epub 2006/07/28
- 122. Collin GB, Marshall JD, Ikeda A, So WV, Russell-Eggitt I, Maffei P et al (2002) Mutations in ALMS1 cause obesity, type 2 diabetes and neurosensory degeneration in Alstrom syndrome. Nat Genet 31(1):74–78, Epub 2002/04/10
- 123. Hearn T, Renforth GL, Spalluto C, Hanley NA, Piper K, Brickwood S et al (2002) Mutation of ALMS1, a large gene with a tandem repeat encoding 47 amino acids, causes Alstrom syndrome. Nat Genet 31(1):79–83, Epub 2002/04/10
- 124. Li G, Vega R, Nelms K, Gekakis N, Goodnow C, McNamara P et al (2007) A role for Alstrom syndrome protein, alms1, in kidney ciliogenesis and cellular quiescence. PLoS Genet 3(1):e8, Epub 2007/01/09
- 125. Patel S, Minton JA, Weedon MN, Frayling TM, Ricketts C, Hitman GA et al (2006) Common variations in the ALMS1 gene do not contribute to susceptibility to type 2 diabetes in a large white UK population. Diabetologia 49(6):1209–1213, Epub 2006/04/08
- 126. Ollmann MM, Wilson BD, Yang YK, Kerns JA, Chen Y, Gantz I et al (1997) Antagonism of central melanocortin receptors in vitro and in vivo by agouti-related protein. Science 278(5335):135–138, Epub 1997/10/06
- 127. Katsuki A, Sumida Y, Gabazza EC, Murashima S, Tanaka T, Furuta M et al (2001) Plasma levels of agouti-related protein are increased in obese men. J Clin Endocrinol Metab 86(5):1921–1924, Epub 2001/05/10
- 128. Lee R, Kermani P, Teng KK, Hempstead BL (2001) Regulation of cell survival by secreted proneurotrophins. Science 294(5548):1945–1948, Epub 2001/12/01
- Strader AD, Reizes O, Woods SC, Benoit SC, Seeley RJ (2004) Mice lacking the syndecan-3 gene are resistant to diet-induced obesity. J Clin Invest 114(9):1354–1360, Epub 2004/11/03
- 130. Ha E, Kim MJ, Choi BK, Rho JJ, Oh DJ, Rho TH et al (2006) Positive association of obesity with single nucleotide polymorphisms of syndecan 3 in the Korean population. J Clin Endocrinol Metab 91(12):5095–5099, Epub 2006/10/05
- 131. Cassard AM, Bouillaud F, Mattei MG, Hentz E, Raimbault S, Thomas M et al (1990) Human uncoupling protein gene: structure, comparison with rat gene, and assignment to the long arm of chromosome 4. J Cell Biochem 43(3):255–264, Epub 1990/07/01
- 132. Clement K, Ruiz J, Cassard-Doulcier AM, Bouillaud F, Ricquier D, Basdevant A et al (1996) Additive effect of A → G (-3826) variant of the uncoupling protein gene and the Trp64Arg mutation of the beta 3-adrenergic receptor gene on weight gain in morbid obesity. Int J Obes Relat Metab Disord 20(12):1062–1066, Epub 1996/12/01

- 133. Oppert JM, Vohl MC, Chagnon M, Dionne FT, Cassard-Doulcier AM, Ricquier D et al (1994) DNA polymorphism in the uncoupling protein (UCP) gene and human body fat. Int J Obes Relat Metab Disord 18(8):526–531, Epub 1994/08/01
- Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K et al (2001) A role for ghrelin in the central regulation of feeding. Nature 409(6817):194–198, Epub 2001/02/24
- 135. Buckley MF, Loveland KA, McKinstry WJ, Garson OM, Goding JW (1990) Plasma cell membrane glycoprotein PC-1. cDNA cloning of the human molecule, amino acid sequence, and chromosomal location. J Biol Chem 265(29):17506–17511

Chapter 2 Genetic Obesity Syndromes

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Abstract A number of genetic obesity syndromes have been identified by sequencing candidate genes in patients with severe obesity. Many of the initial findings emerged from studying families who displayed a classical Mendelian pattern of inheritance; however, with more comprehensive genome wide approaches, increasingly more complex models of inheritance are likely to emerge. The functional and physiological characterization of the human obesity syndromes has provided information that has diagnostic value (Fig. 2.1), has led to specific treatments in some patients and continues to provide insights into the mechanisms involved in the regulation of body weight in humans.

Introduction

Traditionally, patients affected by genetic forms of obesity were identified as a result of their association with developmental delay, dysmorphic features and/or other developmental abnormalities, i.e. a pattern of clinical features which represented a recognizable syndrome. However, the identification of genetic disorders that disrupt the hypothalamic leptin–melanocortin signalling pathway has led to the recognition that obesity is the predominant presenting feature in a significant subset of individuals. Based on case series of patients with genetic obesity syndromes, childhood onset of obesity is a consistent feature. For the purposes of clinical assessment, it remains useful to categorize the genetic obesity syndromes as those with dysmorphism and/or developmental delay and those without these features; however, in some cases the spectrum of clinical features can be quite variable (Fig. 2.1).

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Fig. 2.1 A diagnostic approach to obesity syndromes

Obesity with Developmental Delay

To date, there are at least 30 disorders where obesity is a consistent clinical feature but often associated with mental retardation, dysmorphic features and organ-specific developmental abnormalities. High-throughput next-generation sequencing technologies, and in particular copy number variant detection, are likely to result in the identification and recognition of multiple new syndromes where obesity and developmental delay are closely associated.

Prader-Willi Syndrome

The Prader–Willi syndrome is the most common obesity syndrome (estimated prevalence of about 1 in 25,000). Key clinical features include hypotonia and failure to thrive in infancy, mental retardation, short stature, hyperphagic obesity and hypogonadotropic hypogonadism [1]. Children with Prader–Willi syndrome (PWS) have reduced lean body mass and increased fat mass, abnormalities which resemble those seen in growth hormone (GH) deficiency; GH treatment decreases body fat and increases linear growth, muscle mass, fat oxidation and energy expenditure [2]. Children and adults with PWS have fasting plasma ghrelin levels that are several-fold higher than equally obese controls and patients with other genetic obesity

syndromes [3]. The significance of this finding and its possible role in the pathogenesis of hyperphagia in these patients is unknown.

PWS is caused by deletion of a critical segment on the paternally inherited copy of chromosome 15q11.2-q12, or loss of the entire paternal chromosome 15 with presence of two maternal copies (uniparental maternal disomy). Most chromosomal abnormalities in PWS occur sporadically. Deletions account for 70–80 % of cases; the majority are interstitial deletions, many of which can be visualized by karyotype analysis. There are distinct differences in DNA methylation of the parental alleles, and DNA methylation can be used as a reliable postnatal diagnostic tool in PWS. Small deletions encompassing only the HBII-85 family of snoRNAs have been reported in association with the cardinal features of PWS including obesity [4, 5], suggesting that these noncoding sequences and the genes they regulate may be important.

Albright Hereditary Osteodystrophy

Mutations in *GNAS1* that decrease expression or function of G alpha s protein result in Albright hereditary osteodystrophy (AHO), which is an autosomal dominant disorder. Maternal transmission of *GNAS1* mutations leads to classical AHO (characterized by short stature, obesity, skeletal defects and impaired olfaction) plus resistance to several hormones (e.g. parathyroid hormone) that activate Gs in their target tissues (pseudohypoparathyroidism type IA), while paternal transmission leads only to AHO (pseudopseudohypoparathyroidism). Studies in both mice and humans demonstrate that GNAS1 is imprinted in a tissue-specific manner, being expressed primarily from the maternal allele in some tissues and biallelically in other tissues; thus multi-hormone resistance occurs only when Gs (alpha) mutations are inherited maternally [6].

Bardet–Biedl Syndrome

Bardet–Biedl syndrome (BBS) is a rare (prevalence <1/100,000), autosomal recessive disease characterized by obesity, mental retardation, dysmorphic extremities (syndactyly, brachydactyly or polydactyly), retinal dystrophy or pigmentary retinopathy, hypogonadism and structural abnormalities of the kidney or functional renal impairment. BBS is a genetically heterogeneous disorder that is now known to map to at least 16 loci, with mutations in more than one locus sometimes required for complete expression of the phenotype. Many BBS genes appear to affect proteins localized to the basal body, a key element of the monocilium thought to be important for intercellular sensing in mammalian cells including neurons [7]. Other disorders of ciliary function (e.g. Alström syndrome and Carpenter syndrome) are also associated with obesity. Recent studies in mice have suggested a connection between ciliary function and leptin signalling [8].

BDNF and TRKB Deficiency

Brain-derived neurotrophic factor (BDNF) is one of several nerve growth factors which activate signalling by the tyrosine kinase receptor tropomycin-related kinase B (TrkB) which may lie distal to melanocortin 4 receptor (MC4R) signalling. We reported a child with severe obesity, impaired short-term memory and developmental delay who had a de novo missense mutation impairing the function of TrkB [9]. We also identified a patient with a de novo chromosomal inversion, which encompasses the *BDNF* locus and disrupts *BDNF* expression [10]. Yanovski and colleagues showed that in patients with WAGR syndrome, a subset of deletions on chromosome 11p.12 which encompass the *BDNF* locus, were associated with early-onset obesity [11].

SIM1 Deficiency

Single-minded 1 (SIM1) is a basic helix-loop-helix transcription factor involved in the development and function of the paraventricular nucleus of the hypothalamus. Obesity has been reported in a patient with a balanced translocation disrupting SIM1 [12] and multiple heterozygous missense mutations have been identified in severely obese patients. SIM1 variants with reduced activity co-segregate with obesity in extended family studies with variable penetrance. The phenotypic similarities between patients with SIM1 deficiency and MC4R deficiency suggests that some of the effects of SIM1 deficiency are mediated by altered melanocortin signalling. In some cases, *SIM1* variant carriers have been reported to exhibit a spectrum of neurobehavioural features including autistic type behaviours. These features are not recognized features of MC4R deficiency but show some overlap with the behavioural phenotypes seen in Prader-Willi Syndrome. As the hyperphagia of sim1 haplo-insufficient mice is partly ameliorated by the central administration of oxytocin [13], a neurotransmitter involved in the modulation of emotion and social interaction, impaired oxytocinergic signalling is one possible mechanism implicated in the obesity and behavioural phenotype seen in SIM1 variant carriers.

Obesity Without Developmental Delay

Severe obesity can result from a multiplicity of defects involving the leptinmelanocortin pathway. Leptin is an adipocyte-derived hormone whose circulating levels correlate closely with fat mass. The physiological effects of leptin are mediated through the long isoform of the leptin receptor which is widely expressed in the hypothalamus and other brain regions involved in energy homeostasis. Leptin stimulates the expression of pro-opiomelanocortin (POMC) in primary neurons located in the arcuate nucleus of the hypothalamus. POMC is extensively post-translationally modified to generate the melanocortin peptides, which activate the melanocortin receptors to modulate diverse functions in the central nervous system, the adrenal gland and the skin. The melanocortins are agonists at melanocortin receptors and suppress food intake. In addition, leptin inhibits orexigenic pathways, mediated by neurons expressing the melanocortin antagonist Agouti-related protein and neuropeptide Y (NPY); NPY can suppress the expression of POMC. These two sets of primary leptin-responsive neurons project to second-order neurons expressing MC4R. Targeted genetic disruption of MC4R in mice leads to increased food intake and increased lean mass and linear growth [14].

Leptin and Leptin Receptor Deficiency

Amongst patients with hyperphagic obesity of early onset from consanguineous families, the prevalence of leptin mutations is approximately 1 % and of leptin receptor mutations, 2-3 %. Leptin receptor mutations have been found in some non-consanguineous families, where both parents are unrelated but happen to carry rare alleles in heterozygous form. Serum leptin is a useful test in patients with severe early onset obesity as an undetectable serum leptin is highly suggestive of a diagnosis of congenital leptin deficiency due to homozygous loss of function mutations in the gene encoding leptin. Serum leptin concentrations are appropriate for the degree of obesity in leptin receptor deficient patients and as such an elevated serum leptin concentration is not necessarily a predictor of leptin receptor deficiency [15].

The clinical phenotypes associated with congenital leptin and leptin receptor deficiencies are similar. Leptin and leptin receptor deficient subjects are born of normal birth weight but exhibit rapid weight gain in the first few months of life resulting in severe obesity [16]. Affected subjects are characterized by intense hyperphagia with food seeking behaviour and aggressive behaviour when food is denied, and energy intake at an ad libitum meal is markedly elevated. While measurable changes in resting metabolic rate or total energy expenditure have not been demonstrated, abnormalities of sympathetic nerve function in leptin deficient adults suggest that autonomic dysfunction may contribute to the obesity phenotype observed. Leptin and leptin receptor deficiency are associated with hypothalamic hypothyroidism; normal pubertal development does not occur in adults with leptin or leptin receptor deficiency, with biochemical evidence of hypogonadotropic hypogonadism. However, there is some evidence for the delayed but spontaneous onset of menses in some leptin and leptin receptor deficient adults. Leptin and leptin receptor deficient children have normal linear growth in childhood and normal IGF1 levels. However, because of the absence of a pubertal growth spurt the final height of adult subjects is reduced. Children with leptin deficiency have impaired T cell number and function, consistent with high rates of childhood infection and a high reported rate of childhood mortality from infection.

Although leptin deficiency appears to be rare, it is entirely treatable with daily subcutaneous injections of recombinant human leptin with beneficial effects on the degree of hyperphagia, reversal of the immune defects and infection risk and permissive effects on the development of puberty [16]. Such treatment is currently available to patients on a named patient basis. The major effect of leptin administration is on food intake, with normalization of hyperphagia and enhanced satiety [16, 17]. Leptin is also involved in mediating food reward [18, 19]. Leptin administration does not result in a change in energy expenditure; however, as weight loss by other means is associated with a decrease in basal metabolic rate, the absence of an effect is notable.

Disorders Affecting Pro-opiomelanocortin (POMC) and POMC Processing

Children who are homozygous or compound heterozygous for mutations in the POMC gene present in neonatal life with adrenal crisis due to ACTH deficiency, as POMC is a precursor of ACTH in the pituitary, and they require long-term corticosteroid replacement [20]. Such children have pale skin and white Caucasians have red hair due to the lack of MSH function at melanocortin 1 receptors in the skin. Although red hair may be an important diagnostic clue in patients of Caucasian origin, its absence in patients originating from other ethnic groups should not result in this diagnostic consideration being excluded as children from different ethnic backgrounds may have a less obvious phenotype such as dark hair with red roots. POMC deficiency results in hyperphagia and early-onset obesity due to loss of melanocortin signalling at the MC4R. The clinical features are comparable to those reported in patients with mutations in the receptor for POMC derived ligands, MC4R (see below).

Heterozygous point mutations in POMC have been described which significantly increase obesity risk but are not invariably associated with obesity. R236G disrupts a di-basic cleavage site between β -MSH and β -endorphin, resulting in a β -MSH/ β -endorphin fusion protein that binds to MC4R but has reduced ability to activate the receptor [21]. A rare missense mutation in the region encoding β -MSH, Tyr221Cys has impaired the ability to bind to and activate signalling from the MC4R, and obese children carrying the Tyr221Cys variant are hyperphagic and showed increased linear growth, features of MC4R deficiency [22]. These observations support a role for β -MSH in the control of human energy homeostasis. Selective MC4R agonists of melanocortin analogues may be feasible therapies for such patients in the future.

PCSK1 Deficiency

Proprotein convertases (PCs) are a family of serine endoproteases that cleave inactive pro-peptides into biologically active peptides [23]. Two family members, Proprotein Convertase Subtilisin/Kexin type 1 and 2 (PCSK1 and PCSK2) are selectively
expressed in neuroendocrine tissues where they cleave prohormones including pro-opiomelanocortin (POMC), prothyrotrophin releasing hormone (TRH), proinsulin, proglucagon and progonadotrophin releasing hormone (GnRH) to release biologically active peptides. Compound heterozygous or homozygous mutations in the *PCSK1* gene, which encodes PC1/3, cause small bowel enteropathy and complex neuroendocrine effects (including diabetes insipidus) due to a failure to process a number of prohormones as well as severe, early onset obesity [24, 25].

MC4R Deficiency

Heterozygous *MC4R* mutations have been reported in obese people from various ethnic groups. The prevalence of pathogenic *MC4R* mutations has varied from 0.5 to 2.5 % of people with a BMI>30 kg/m² in UK and European populations to 5 % in patients with severe childhood obesity [26, 27]. As *MC4R* deficiency is the most common genetic form of obesity, assessment of the sequence of the *MC4R* is increasingly seen as a necessary part of the clinical evaluation of the severely obese child.

Given the large number of potential influences on body weight, it is perhaps not surprising that both genetic and environmental modifiers will have important effects on the severity of obesity associated with MC4R mutations in some pedigrees. Co-dominance, with modulation of expressivity and penetrance of the phenotype, is the most appropriate descriptor for the mode of inheritance.

The clinical features of *MC4R* deficiency include hyperphagia in early childhood. Alongside the increase in fat mass, *MC4R*-deficient subjects also have an increase in lean mass and a marked increase in bone mineral density, thus they often appear "big-boned" [27]. They exhibit accelerated linear growth, which may be a consequence of disproportionate early hyperinsulinemia and effects on pulsatile growth hormone (GH) secretion, which is retained in MC4R-deficient adults in contrast to common forms of obesity [28]. Despite this early hyperinsulinemia, obese adult subjects who are heterozygous for mutations in the *MC4R* gene are not at increased risk of developing glucose intolerance and type 2 diabetes compared to controls of similar age and adiposity [27]. The proportion of visceral to subcutaneous fat is not altered in MC4R deficiency. Reduced sympathetic nervous system activity in MC4R-deficient patients is likely to explain the lower prevalence of hypertension and lower systolic and diastolic blood pressures [29]. Thus, central melanocortin signalling appears to play an important role in the regulation of blood pressure and its coupling to changes in weight.

At present, there is no specific therapy for *MC4R* deficiency, but patients with heterozygous MC4R mutations do respond to Roux-en-Y-bypass surgery [30], which can be considered in adults. As most patients are heterozygotes with one functional allele intact, it is possible that small molecule MC4R agonists or pharma-cological chaperones which improve receptor trafficking to the cell surface might be appropriate treatments for this disorder.

SH2B1 Deficiency

Severe obesity without developmental delay is associated with a significantly increased burden of rare, typically singleton copy number variants (CNVs) [31]. Deletion of a 220-kb segment of 16p11.2 is associated with highly penetrant familial severe early-onset obesity and severe insulin resistance [32]. This deletion includes a small number of genes, one of which is *SH2B1*, known to be involved in leptin and insulin signalling. These patients gain weight in the first years of life, with hyperphagia and fasting plasma insulin levels that are disproportionately elevated compared to age- and obesity-matched controls. Several mutations in the *SH2B1* gene have also been reported in association with early onset obesity, severe insulin resistance and behavioural abnormalities in some patients [33].

Clinical History, Examination and Investigation

The assessment of severely obese children and adults should be directed at screening for potentially treatable endocrine and neurological conditions and identifying genetic conditions so that appropriate genetic counselling and in some cases treatment can be instituted. Much of the information needed can be obtained from a careful medical history and physical examination, which should also address the potential complications of severe obesity such as sleep apnoea [34]. In addition to a general medical history, a specific weight history should be taken carefully establishing the age of onset and the presence of hyperphagia. A careful family history to identify potential consanguineous relationships, the presence of other family members with severe early onset obesity and the ethnic and geographical origin of family members should be taken. The history and examination can then guide the appropriate use of diagnostic tests.

Conclusions

Given the rapid application of next-generation sequencing technologies such as whole exome sequencing, it is very likely that new genes and mechanisms will emerge to explain a variety of previously unrecognized obesity syndromes. As more is learned about these genes and more syndromes are described, it is likely that the need to perform a comprehensive evaluation of severely obese patients will be recognized. Knowledge of the specific molecular mechanisms affected by these genetic disorders may lead to better mechanism-directed, stratified pharmacotherapy in the future.

References

- 1. Goldstone AP (2004) Prader-Willi syndrome: advances in genetics, pathophysiology and treatment. Trends Endocrinol Metab 15:12–20
- 2. Carrel AL, Allen DB (2001) Prader-Willi syndrome: how does growth hormone affect body composition and physical function? J Pediatr Endocrinol Metab 14(Suppl 6):1445–1451
- Haqq AM, Farooqi IS, O'Rahilly S et al (2003) Serum ghrelin levels are inversely correlated with body mass index, age, and insulin concentrations in normal children and are markedly increased in Prader-Willi syndrome. J Clin Endocrinol Metab 88:174–178
- de Smith AJ, Purmann C, Walters RG et al (2009) A deletion of the HBII-85 class of small nucleolar RNAs (snoRNAs) is associated with hyperphagia, obesity and hypogonadism. Hum Mol Genet 18:3257–3265
- 5. Sahoo T, del Gaudio D, German JR et al (2008) Prader-Willi phenotype caused by paternal deficiency for the HBII-85 C/D box small nucleolar RNA cluster. Nat Genet 40:719–721
- Weinstein LS, Chen M, Liu J (2002) Gs(alpha) mutations and imprinting defects in human disease. Ann N Y Acad Sci 968:173–197
- Ansley SJ, Badano JL, Blacque OE et al (2003) Basal body dysfunction is a likely cause of pleiotropic Bardet-Biedl syndrome. Nature 425:628–633
- Seo S, Guo DF, Bugge K, Morgan DA, Rahmouni K, Sheffield VC (2009) Requirement of Bardet-Biedl syndrome proteins for leptin receptor signaling. Hum Mol Genet 18:1323–1331
- 9. Yeo GS, Connie Hung CC, Rochford J et al (2004) A de novo mutation affecting human TrkB associated with severe obesity and developmental delay. Nat Neurosci 7:1187–1189
- Gray J, Yeo GS, Cox JJ et al (2006) Hyperphagia, severe obesity, impaired cognitive function, and hyperactivity associated with functional loss of one copy of the brain-derived neurotrophic factor (BDNF) gene. Diabetes 55:3366–3371
- Han JC, Liu QR, Jones M et al (2008) Brain-derived neurotrophic factor and obesity in the WAGR syndrome. N Engl J Med 359:918–927
- Holder JL Jr, Butte NF, Zinn AR (2000) Profound obesity associated with a balanced translocation that disrupts the SIM1 gene. Hum Mol Genet 9:101–108
- Kublaoui BM, Gemelli T, Tolson KP, Wang Y, Zinn AR (2008) Oxytocin deficiency mediates hyperphagic obesity of Sim1 haploinsufficient mice. Mol Endocrinol 22:1723–1734
- 14. Huszar D, Lynch CA, Fairchild-Huntress V et al (1997) Targeted disruption of the melanocortin-4 receptor results in obesity in mice. Cell 88:131–141
- Farooqi IS, Wangensteen T, Collins S et al (2007) Clinical and molecular genetic spectrum of congenital deficiency of the leptin receptor. N Engl J Med 356:237–247
- Farooqi IS, Matarese G, Lord GM et al (2002) Beneficial effects of leptin on obesity, T cell hyporesponsiveness, and neuroendocrine/metabolic dysfunction of human congenital leptin deficiency. J Clin Invest 110:1093–1103
- Farooqi IS, Jebb SA, Langmack G et al (1999) Effects of recombinant leptin therapy in a child with congenital leptin deficiency. N Engl J Med 341:879–884
- Farooqi IS, Bullmore E, Keogh J, Gillard J, O'Rahilly S, Fletcher PC (2007) Leptin regulates striatal regions and human eating behavior. Science 317:1355
- Rosenbaum M, Sy M, Pavlovich K, Leibel RL, Hirsch J (2008) Leptin reverses weight lossinduced changes in regional neural activity responses to visual food stimuli. J Clin Invest 118:2583–2591
- 20. Krude H, Biebermann H, Schnabel D et al (2003) Obesity due to proopiomelanocortin deficiency: three new cases and treatment trials with thyroid hormone and ACTH4-10. J Clin Endocrinol Metab 88:4633–4640
- 21. Challis BG, Pritchard LE, Creemers JW et al (2002) A missense mutation disrupting a dibasic prohormone processing site in pro-opiomelanocortin (POMC) increases susceptibility to early-onset obesity through a novel molecular mechanism. Hum Mol Genet 11:1997–2004

- 22. Lee YS, Challis BG, Thompson DA et al (2006) A POMC variant implicates beta-melanocytestimulating hormone in the control of human energy balance. Cell Metab 3:135–140
- 23. Seidah NG (2011) The proprotein convertases, 20 years later. Methods Mol Biol 768:23-57
- 24. Jackson RS, Creemers JW, Ohagi S et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene [see comments]. Nat Genet 16:303–306
- 25. Jackson RS, Creemers JW, Farooqi IS et al (2003) Small-intestinal dysfunction accompanies the complex endocrinopathy of human proprotein convertase 1 deficiency. J Clin Invest 112:1550–1560
- 26. Stutzmann F, Tan K, Vatin V et al (2008) Prevalence of melanocortin-4 receptor deficiency in Europeans and their age-dependent penetrance in multigenerational pedigrees. Diabetes 57:2511–2518
- Farooqi IS, Keogh JM, Yeo GS, Lank EJ, Cheetham T, O'Rahilly S (2003) Clinical spectrum of obesity and mutations in the melanocortin 4 receptor gene. N Engl J Med 348:1085–1095
- 28. Martinelli CE, Keogh JM, Greenfield JR et al (2011) Obesity due to melanocortin 4 receptor (MC4R) deficiency is associated with increased linear growth and final height, fasting hyperinsulinemia, and incompletely suppressed growth hormone secretion. J Clin Endocrinol Metab 96:E181–E188
- 29. Greenfield JR, Miller JW, Keogh JM et al (2009) Modulation of blood pressure by central melanocortinergic pathways. N Engl J Med 360:44–52
- Hatoum IJ, Stylopoulos N, Vanhoose AM et al (2012) Melanocortin-4 receptor signaling is required for weight loss after gastric bypass surgery. J Clin Endocrinol Metab 97:E1023–E1031
- Wheeler E, Huang N, Bochukova EG et al (2013) Genome-wide SNP and CNV analysis identifies common and low-frequency variants associated with severe early-onset obesity. Nat Genet 45:513–517
- Bochukova EG, Huang N, Keogh J et al (2010) Large, rare chromosomal deletions associated with severe early-onset obesity. Nature 463:666–670
- Doche ME, Bochukova EG, Su HW et al (2012) Human SH2B1 mutations are associated with maladaptive behaviors and obesity. J Clin Invest 122:4732–4736
- Dietz WH, Robinson TN (2005) Clinical practice. Overweight children and adolescents. N Engl J Med 352:2100–2109

Chapter 3 Genome-Wide Association Studies of Obesity

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Abstract Genome-wide association studies (GWAS) have accelerated the discovery of genetic variants associated with susceptibility to common complex diseases, such as obesity. Following the first robust GWAS of BMI and risk of obesity identified in 2007, GWAS have delivered 70 additional common loci associated with a wide range of obesity-related traits. These loci highlight a variety of molecular and physiological mechanisms involved in shaping these traits. However, even in combination, these loci explain only a small proportion of overall phenotypic heritability indicating that much of the genetic variation in obesity traits remains unexplained. Here, we discuss how the GWAS approach has been applied to the study of anthropometric phenotypes related to overall obesity and fat distribution and describe some of the clues to trait biology that are emerging. We also highlight some of the limitations of this work and future directions for research in this field.

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Abbreviations

BMI	Body mass index			
CNV	Copy number variation			
GIANT	Genetic Investigation of ANthropometric Traits			
GWAS	Genome-wide association studies			
LD	Linkage disequilibrium			
MAF	Minor allele frequency			
SNP	Single nucleotide polymorphism			
T2D	Type 2 Diabetes			
WC	Waist circumference			
WHR	Waist-hip ratio			

The rise in the prevalence of obesity in recent decades has been spectacular: recent estimates indicate that over 500 million adults worldwide are now classed as obese [1]. While the increased prevalence of obesity is almost certainly a reflection of secular changes in environmental and lifestyle factors, including an increased intake of nutrient-dense foods coupled with reduced physical activity [2], the familial aggregation of obesity is consistent with some degree of genetic influence on body mass index (BMI) and individual predisposition to obesity. More conclusive evidence for a genetic component comes from studies that have examined the correlation of BMI between identical twins raised apart and the relationship between the BMI of adoptees and both their biological and adoptive parents [3-5]. These consistently highlight the importance of genetic factors in modulating individual susceptibility to obesity in contemporary environments. Furthermore, in controlled experiments of excessive calorie intake, consequent changes in weight and body composition were highly correlated in monozygotic twins, once again consistent with a powerful role of genetic variation in the regulation of weight [6]. Estimates for the heritability of BMI vary widely between studies, but typical figures range between 0.47 and 0.90 in twin studies and between 0.24 and 0.81 in family-based studies [7].

Other obesity related traits, including measures of fat distribution are also heritable (even after adjusting for BMI). Estimates for the heritability of waist-hip ratio (WHR), a proxy of fat distribution, range between $h^2 \sim 0.31-0.70$; and $\sim 0.22-0.61$ after accounting for BMI [8–11]. The heritability of WHR is higher in women and estimates of genetic correlation of WHR between men and women indicate a sex specific genetic influence on the trait [11].

The Genetics of BMI and Obesity Pre-GWAS

Genetic studies aim to find DNA sequence variants that are causally associated with the trait of interest, in the expectation that such discoveries will help to reveal fundamental mechanisms responsible for human disease. The earliest studies in this field focused on the application of family-based linkage studies to individuals and families with rare monogenic forms of obesity. The rare variants of large effects revealed by these efforts, such as those in *LEP* (encoding the hormone leptin, a crucial component of energy balance mechanisms) [12–14], *LEPR* (encoding the leptin receptor) [15], and *POMC* (encoding the proopiomelanocortin protein which is cleaved to form a number of key neuroendocrine messengers) [16], helped to define components of hypothalamic circuitry involved in body weight regulation in man. However, the application of linkage approaches to population-level variation in BMI and risk of common forms of obesity met with little success in terms of robust, replicated signals even in relatively well-powered meta-analysis [17]. This indicates that the genetic contribution to these traits is not dominated by the kinds of highly penetrant variants which linkage methods are best suited to detect [18].

The shift from linkage to association approaches was initially focused on the analysis of candidate genes [19], a strategy reliant on the quality of the prior biological hypotheses used to select them. One of relatively few successes from this approach was the demonstration that low frequency variants in the gene encoding the melanocortin 4 receptor (MC4R) were associated with severe, early-onset obesity [20]. These variants remain the commonest known genetic cause of morbid obesity contributing to a few percent of these cases [21]. These findings provided confirmation of the role of signalling through the hypothalamic leptin–melanocortin pathway for the maintenance of body mass in man [22]. However, the major impetus to the discovery of BMI- and obesity-associated variants has been provided by the ability to perform genome-wide scans for association.

Genome-Wide Association Studies

Genome-wide association studies (GWAS) (reviewed in [23, 24]) use dense genotyping arrays to determine how variation in genomic sequence (predominantly that due to single nucleotide polymorphisms, SNPs) associates with phenotypic traits of interest. Those traits may be categorical (e.g., obese cases and non-obese controls) or continuous (e.g., BMI or WHR). Array content and the correlation structure of variation across the genome (i.e., linkage disequilibrium) mean that GWAS to date have favored the interrogation of common variants (minor allele frequency [MAF]>5 %). Since GWAS assay such variants across the genome, suitably powered studies enable the discovery of associated loci in an agnostic fashion, without the need for prespecified hypotheses concerning the genomic location of the association and the transcripts through which they may operate.

In the remainder of this chapter, we focus on the loci which have been shown by GWAS to be associated with overall obesity or fat distribution. We distinguish between studies of traits of overall obesity (including BMI, fat percentage, and dichotomized indices of extreme obesity) and those of fat distribution (including WHR, waist circumference (WC), and measures of visceral and subcutaneous fat). In total 70 genome-wide significant loci have been associated with these traits and most of these (50 in number) are common variant loci influencing continuous

obesity-related traits found in European samples. Others derive from equivalent studies in non-European samples (4 loci), and some have emerged exclusively from case–control studies in individuals selected from the extremes of the BMI distribution (9 loci) or by clinical classifications of overweight and obesity (7 loci).

Overall Obesity

Genome-Wide Association Studies of BMI

The first report from a GWA study claiming to have identified variants associated with common forms of overall obesity came in 2006 [25]. The researchers used a two-stage family-based design to identify a signal mapping close to the *INSIG2* encoding insulin induced gene 2 [25]. However, this association has not been proven robust to replication in the much larger samples that have been examined in subsequent studies (see below). In fact, the association p-value observed in this study fell short of the now-widely accepted threshold ($p < 5 \times 10^{-8}$: based on p < 0.05 corrected for a million independent tests [26]), highlighting the value of such stringent criteria as a means of avoiding inflation of the type 1 error, and the attribution of biological significance to loci which, like *INSIG2*, are likely to have been false positives.

The first report of a robust genome-wide significant locus influencing BMI and risk of obesity locus came from Frayling et al. [27] in 2007, and concerned a cluster of common variants close to the *FTO* ("Fat mass and obesity-associated") gene. These variants account for ~0.35 % of the phenotypic variance in BMI in Europeans [28] such that the two groups of homozygotes differ in weight by around 2.5 kg. The BMI association has now been widely replicated [28–33] and it is also clear that the same *FTO* variants are associated with risk of obesity at all grades of severity [27, 34].

Given that the only locus emerging from this first round of GWA studies [25, 27] had a relatively modest effect size, it was clear that larger sample sizes would be needed to extend these discoveries, both to common alleles of lesser effect, and to less frequent risk alleles. This provided the motivation for ever-larger meta-analyses efforts, which have dominated discovery efforts over the past few years. The largest of the studies published to date assembled data from almost 250,000 individuals [28]. The current count of BMI-associated loci detected in Europeans by these studies, most of them conducted under the aegis of the Genetic Investigation of ANthropometric Traits (GIANT) consortium [28–30], is 32 (Fig. 3.1).

The first such meta-analyses uncovered common regulatory variants influencing BMI near *MC4R* (encoding melanocortin receptor 4) [29]: low-frequency coding variants in this same gene had previously been implicated in severe obesity [20]. Subsequently, the parallel publications from GIANT [30] and the deCODE group [31] added nine BMI loci (mapping near *GNPDA2*, *KCTD15*, *MTCH2*, *NEGR1*, *SH2B1*, *TMEM18*, *BDNF*, *ETV5*, and *SEC16B*) to the list. It is of note that *BDNF*, encoding a brain derived neurotrophic factor involved in regulation of development



Fig. 3.1 Overlap of genome-wide significant loci of overall obesity (BMI), fat distribution (BMIadjusted WHR) and BMI extremes (or clinical obesity classes) in European populations. Diagram depicts the overlap of reported GWAS loci ($p < 5 \times 10^{-8}$) of BMI [27–31], BMI-adjusted WHR [98], and BMI extremes or clinical obesity classes II–III [34, 38, 51–53, 55, 120]

of neuronal circuits [35], is also involved in monogenic forms of obesity [36]. The largest meta-analysis of BMI associations added 18 further loci to the tally [28] including regions near known obesity genes such as *POMC* (proopiomelanocortin) [16], known to be involved in neuroendocrine regulation of weight, as well as associations in or near novel genes such as *GPRC5B* (G protein-coupled receptor, family C, group 5, member B), implicated in regulation of adipose inflammatory processes and progression to insulin resistance in obesity in mice [37].

The studies above mostly focused on the analysis of SNPs but there is some evidence that copy number variations (CNVs) may be causal at some loci. For example, in the study by Willer et al. the BMI-associated SNP at the *NEGR1* (neuronal growth regulator 1) locus detected by GWAS was shown to tag a 45 kb deletion that might have stronger functional grounds for being causal [30]. More detailed studies published recently locate the causal allele at this locus to a second 8 kb deletion near *NEGR1* [38]. Rare CNVs have also been implicated in syndromic forms of obesity. For example, a rare deletion in the 16p11.2 region is associated with the combination of severe obesity and mental retardation [39, 40], and duplication of the same region is associated with underweight [41].

To date, most GWAS studies have been performed in populations of European origin but studies in other ethnic groups can help to identify novel loci, to characterize the extent of aetiological overlap, and to fine-map causal variants (such as in the *FTO* locus [42, 43]). Two large GWAS meta-analyses of BMI in East Asian populations were recently published [32, 33]. Between them, seven of the known loci in Europeans could be replicated to genome-wide levels of significance.

Furthermore, evaluating the associations in East Asians of previously reported BMI loci, an additional 11 loci, besides those genome-wide significant, were associated with BMI at lesser levels of significance (Okada et al. [32], p < 0.02; Wen et al. [33], p < 0.05), indicating considerable overlap in signals between East Asian and European populations. As in populations of European origin, the association at *FTO* locus explained the largest proportion of phenotypic variance (~0.2 %).

In addition, these studies identified four novel loci, mapping near *CDKAL1*, *KLF9*, *PCSK1* and *GP2* [32, 33]. Mutations in *PCSK1* cause monogenic obesity [44] and, while a candidate study previously associated nonsynonymous variants in *PCSK1* with common obesity risk in a European population [45], the *PCSK1* signal in East Asians (also nominally associated with BMI in Europeans in GIANT [28]) likely represents an independent signal. Genetic variants in *CDKAL1* (a CDK5 regulatory associated protein 1-like 1 with methythiotransferase function [46]), in strong LD ($r^2 \sim 0.8$) with the BMI GWAS SNPs in East Asians, have previously been associated with increased risk of Type 2 Diabetes (T2D) [47, 48]. The T2D risk allele(s) is associated with decreased glucose-stimulated insulin secretion [47, 49, 50]. Furthermore, the BMI-lowering allele of rs2206734 (also nominally associated with BMI in Europeans) was associated with increased risk of T2D in the same study population [32], indicating that variation near *CDKAL1* may play a complex role with respect to variation in both BMI and T2D-risk.

Case–Control Studies of Dichotomized BMI

In addition to studies of the variance in BMI in population-based studies, a complementary approach treats obesity in terms of a dichotomous "case–control" analysis. A variety of different schemes for this dichotomization are possible (Table 3.1). In the largest study of this type [51], featuring case–control analyses restricted to the "tails" of the BMI distribution using data from studies previously included in GIANT meta-analyses [28], Berndt et al. found considerable overlap in the pattern of association signals seen as compared to those observed in population- or cohortwide analyses. However, where such studies focus on cases of more extreme definitions of obesity and/or leanness (that is, individuals several standard deviations away from the population mean), there may be the opportunity to detect additional, novel, signals that may have limited impact on overall population-level variance and which are therefore difficult or impossible to detect using GWAS approaches. The rare, penetrant variants causal for monogenic and syndromic forms of obesity provide the most obvious example of this phenomenon.

Indeed, whilst several of the loci reaching genome-wide significance in dichotomous analyses focused on extreme obesity in adults overlap with those previously reported (e.g., *BDNF*, *FTO*), there are several signals that appear unique to dichotomous analyses including *KCNMA1*, *NPC1*, *PTER*, and *HS6ST3* (Table 3.1, Fig. 3.1) [51–53]. However, most of these have appeared in a single study and have not, as yet, been replicated, even in other extreme case–control analyses. In equivalent

		Sample size in		
	Selection criteria	stage 1, cases/	Loci not described	
Study type	for cases	controls	in BMI GWAS	Reference
Extreme obesity in children and adults	Early onset obesity (≤ 6 years) and extreme adult obese (BMI ≥ 40)	1,380/1,416	MAF (rs1424233), NPCI ^a (rs1805081), PTER ^a (rs10508503)	[52]
Extreme obesity in children and adolescents	BMI >97 % percentile	1,138/1,120	<i>TNKS/MSRA</i> (rs17150703), <i>SDCCAG8</i> ^a (rs12145833)	[55]
Extreme obesity in adults	$BMI \ge 40$	164/163	KCNMA1 (rs2116830)	[53]
Distributional tails in children	BMI ≥95 % percentiles	5,530/8,313	<i>BC041448</i> (rs4864201), <i>HOXB5</i> (rs9299), <i>OLFM4</i> (rs9568856)	[54]
Extreme obesity in children	BMI standard deviation score (SDS) ≥3, and onset at 10 years	1,509/5,380	LEPR (rs11208659), PACS1 (rs564343), PRKCH (rs1957894) RMST (rs11109072)	[38]
Clinical class: obesity II	BMI≥35	9,889/62,657	HS6ST3 (rs7989336), ZZZ3 (rs17381664)	[51]
Clinical class: obesity I	BMI≥30	32,858/65,839	GNAT2 (rs17024258), HNF4G (rs4735692), MRPS33P4 (rs13041126), ADCY9 (rs2531995)	[51]
Clinical class: overweight	BMI≥25	93,015/65,840	HNF4G (rs4735692), RPTOR (rs7503807)	[51]

Table 3.1 Novel GWAS loci identified in case-control analyses of dichotomized BMI

^aNot genome-wide significant ($p < 5 \times 10^{-8}$)

case–control analyses in children, the more relaxed criteria adopted by Bradfield et al [54] detected many of the known adult BMI association signals but also highlighted novel signals near *OLFM4* and *HOXB5*. In contrast, studies of children selected from the extremes of the distribution have detected signals at (or approaching) genome-wide significance near *LEPR*, *PACS1*, *PRKCH*, *RMST*, *SDCCAG8*, and *TNKS/MSRA* (Table 3.1, Fig. 3.1) [38, 55], the latter locus also detected in some studies of fat distribution [56].

Genome-Wide Association Studies of Fat Percentage

BMI, although a widely used proxy of overall obesity, represents an aggregate measure of the lean and the fat mass of the individual. In an effort to better define the genetic determinants of obesity, Kilpeläinen et al. focused on body fat percentage, as a more direct measure of adiposity, generating a GWAS meta-analysis of 36,626 individuals [57]. As well as detecting *FTO*, these analyses recovered two loci (*IRS1* and *SPRY2*) not previously associated with BMI. The body fat-increasing alleles at the *IRS1* (insulin receptor substrate 1 signalling protein) signal are, intriguingly, associated with a healthy metabolic profile (including reduced risk of T2D [58] and unhealthy lipid profile [59]). The *IRS1* locus is associated with measures of subcutaneous, but not visceral fat, indicating that the effect on fat mass at the *IRS1* locus is through regulation of subcutaneous fat deposition [57, 60]. The *SPRY2* locus has also been implicated in T2D risk [61, 62], though the body fat-associated SNP is not coincident with this previously reported T2D SNP. Contrary to the observations at *IRS1*, the body-fat increasing allele at the *SPRY2* locus is associated with an adverse metabolic profile [57].

Genetic Architecture of Overall Obesity (BMI)

Despite the success in identifying a growing numbers of loci to genome-wide significance, in European populations these signals, in combination, explain no more than 1.5 % of phenotypic variance in BMI. Of the established loci, the *FTO* locus has the largest effect accounting for ~0.35 % of population variance [28]. These numbers fall well short of estimates of the heritability of this trait (see above). The basis for this "missing" genetic variance remains unclear, though there is no lack of possible explanations [63, 64]. At least part of the "missing" genetic variance can be attributed to the effects of additional common variants that lie below the genomewide significance threshold. Using full GWAS data sets (not just the "proven" hits), Yang and colleagues could recover approximately 17 % of the phenotypic variance in BMI that was tagged by common variants [65]. Part of the remaining shortfall likely reflects incomplete linkage disequilibrium between the variants genotyped on GWAS arrays and those which are causally responsible for the BMI associations [66], but other mechanisms are almost certainly involved [63, 67].

There is also the possibility that the estimates of heritability against which these measures of explained variance are evaluated, are themselves inaccurate. For example, intrauterine events that lead to epigenetic modifications with long-term phenotypic impacts can lead to increased sibling resemblance, inflating heritability estimates under some designs. Similarly, estimates derived from the comparison of the phenotypic correlations observed between monozygotic and dizygotic twin pairs are based on the assumption that both types of twin are exposed to a similar degree of shared environment [68], an assumption that may not be appropriate for intrauterine exposures.

Notwithstanding the above, it seems likely that an appreciable component of the genetic variance remains unexplained, and that at least part of this will be attributable to low frequency and rare variants not well captured by GWAS studies to date. The current wave of sequencing studies should shed some light on the extent to which these variants are contributing to inherited risk.

From GWAS Associations to Potential Functional Roles in Overall Obesity

As we have seen, GWAS have powered the identification of many genetic regions associated with BMI and obesity. However, this information is of limited value unless it can be translated into improved understanding of the pathophysiology of disease, and thereby into novel clinical approaches. However, in BMI, as with most other complex traits, the regions revealed by GWAS do not lend themselves to easy biological inference. The effect sizes are modest, and most signals map to non-coding sequence, frustrating efforts to identify the "causal" transcript (that is, the specific gene that is mediating the association signal). At the same time, the extensive local correlations between common variants (that is, linkage disequilibrium) can make fine-mapping of the causal variants challenging.

The *FTO* locus provides an excellent example of the difficulties inherent in moving from an association signal—in this case, a comparatively strong one—to a clear mechanism of action. We have now known for more than 6 years that a cluster of highly correlated common variants in the first intron of the *FTO* gene is associated with BMI and obesity [27]. Epigenetic analyses have suggested that the BMI-associated haplotype may influence local methylation status [69] but fine-mapping efforts have yet to provide compelling localization of the causal variant. When it comes to defining downstream effects, we still have no convincing evidence from man that the *FTO* transcript itself is in any way involved. There is for example, no instance of the co-occurrence of loss of function alleles in *FTO* and severe obesity in humans [70, 71]. On the other hand, the adjacent gene *RPGRIP1L* (or *FTM*), which is known to be coordinately regulated with *FTO* via a common promoter [72], and to display a similar pattern of hypothalamic expression, has an intriguing connection to obesity through its known causal role with respect to monogenic ciliopathies [73] some of which result in marked early obesity.

In fact, the most compelling evidence implicating *FTO* comes from mouse models: transgenic knockdown of the murine homologue *Fto* results in reduced weight, and overexpression to weight gain compared to control mice [74, 75]. One possible explanation consistent with these data is that the common intronic variants within *FTO* identified by GWAS, exert their effects on energy balance in man through coordinate dysregulation of both *FTO* and *RPGRIP1L*.

The identification of the signal at *FTO* naturally prompted interest in the normal function of this transcript. In humans, *FTO* encodes a 2-oxoglutarate-dependent nucleic acid demethylase [76] thought to be involved in nucleic acid repair. In vitro studies have suggested a role for *FTO* demethylation in cellular sensing of amino acids [77], which could be relevant to regulation of appetite control in the hypothalamus. Nonetheless, it is clear that we remain some way from a complete description of how these variants influence BMI and obesity risk.

At certain other BMI GWAS loci, the situation is better understood. At four GWAS loci (near *BDNF*, *PCSK1*, *POMC* and *MC4R*) the common variant associations overlap genes in which coding mutations have been shown to be causal for

monogenic or syndromic forms of obesity [20, 36, 78, 79]. In the case of three of these—*PCSK1* (proprotein convertase 1), *POMC* (proopiomelanocortin), and *MC4R* (melanocortin receptor 4)—there are strong mechanistic ties to the hypothalamic leptin–melanocortin signalling pathways that regulate energy balance [80]. *BDNF* encodes a brain derived neurotrophic factor involved in neurogenesis and thought to be involved in food intake [81]. These GWAS signals therefore demonstrate that the neuroendocrine mechanisms documented in monogenic forms of obesity extend to population level variance in BMI and to more common forms of obesity.

At other BMI-associated GWAS, efforts to define the causal transcript are supported by additional sources of genomic data (regulatory annotations [82] or mRNA expression [83]). For example, it can be very useful through integration with mRNA and/or miRNA transcriptomic data [83–88] to demonstrate that the set of BMI-associated variants at a given locus also drives *cis*-expression of one of the regional transcripts. In the most recent GIANT meta-analysis [28], this approach led to positional candidates being identified at almost half the 32 BMI-associated loci.

These candidacy assignments can often be bolstered by other sources of data. Consider for example the association signal mapping close to the *SH2B1* gene, encoding SH2B adapter protein 1. *Cis*-expression data point to *SH2B1* [28], as does the high expression of this transcript in the hypothalamus [30]. The neuronal isoform of *SH2B1* is involved in regulation of energy balance via effects on leptin and insulin signalling, and systemic deletion of the gene in mice results in severe leptin resistance [89].

For some loci, the data seem to point towards peripheral rather than central mechanisms of action. The BMI association on chromosome 19 lies close to the *GIPR* gene, encoding the gastric inhibitory polypeptide receptor, and the lead SNP is in strong LD with a missense SNP in that transcript (though the functional consequences of that mutation are not yet established). GIPR plays an important role in mediating the incretin response, which augments insulin release in response to the ingestion of food. The same locus has also been shown to associate with glucose response and insulin secretion in response to a glucose challenge [90]. Another example, mentioned earlier, is the mechanistic relationship between insulin signal-ling and obesity implicated by the association between *IRS1* variants and fat percentage [57]. Though both central and peripheral mechanisms may be involved at *IRS1*, the fact that the fat percentage-associated allele is associated with improved insulin sensitivity and a healthy metabolic profile [58, 59] is consistent with enhanced insulin-mediated adipogenesis as the driver of the adiposity.

For several other BMI-associated loci such as *TMEM160-ZC3H4* [28], there are few clues on the biological relevance in obesity, and any one of several transcripts could be responsible. One way of leveraging the combination of genetic and prior biological data to make provisional mechanistic inference in such situations is to perform pathway-based analyses (reviewed in Wang et al. [91]), which test for enrichment of GWAS loci for transcripts that have been mapped to defined biological processes or pathways. Applied to BMI GWAS data, these analyses have tended to support the evidence for broad neuroendocrine involvement, whilst also highlighting processes that are more difficult to assimilate within the current knowledge base (e.g., platelet-derived growth factor signalling) [28].

Fat Distribution

The clinical consequences of adipose tissue excess depend not only on its quantity but also its distribution, with the accumulation of visceral (abdominal) fat leading to particularly adverse metabolic and cardiovascular effects [92, 93]. After accounting for overall obesity (as measured by BMI), fat distribution (commonly measured by WHR) shows substantial residual heritability ($h^2 \sim 0.22-0.61$) consistent with mechanisms of genetic control distinct from those influencing overall energy balance and BMI [9, 10]. The distinct genetic regulation of patterns of fat distribution is also supported by rare monogenic syndromes of selective adipose tissue loss (collectively, the lipodystrophies) [94]. Given the checkered history of efforts to target neuronal pathways related to overall obesity in the search for effective, safe treatments for obesity, there is considerable interest in defining the mechanisms responsible for individual variation in patterns of fat distribution, and in particular, in identifying peripheral (rather than central) targets for therapeutic intervention.

Genome-Wide Association Studies of WHR and WC

Initial efforts to map variants influencing fat distribution focused on the standard clinical traits, WHR and WC. In the first GWAS for WHR, Lindgren et al. discovered an association to a genetic variant on chromosome 1 (close to the *LYPLAL1* gene encoding lysophospholipase-like 1) associated with WHR in women exclusively: this effect was independent of BMI [56]. Studies of WC generated their strongest signals at previously reported BMI loci such as *FTO* and *MC4R*, reflecting the strong correlation between these traits [27, 56, 95, 96]. With the possible exception of the association near *TFAP2B*, at which adjustment with BMI seems to increase the magnitude of the effect on central obesity [97], other WC-associated loci identified by GWAS (*MSRA*, *NRXN3*) are likely to reflect a primary association with BMI [28].

Given these strong trait correlations, more recent fat distribution GWAS efforts have adopted the approach of adjusting WHR (or WC) for BMI before performing the association analyses, thereby seeking to emphasize those signals that influence patterns of relative fat deposition independent of the overall obesity component. In the largest analysis to date, involving data from around 190,000 subjects, Heid et al. [98] used this approach to identify 13 novel loci for BMI-adjusted WHR as well as to replicate the signal near *LYPLAL1*. As might have been expected given the adjustment for BMI, the loci identified by this endeavor were completely distinct from those previously reported to influence overall obesity (Figs. 3.1 and 3.2). In line with the metabolic consequences of visceral fat accumulation, these fat distribution associated variants are also enriched for association with related metabolic traits including fasting insulin, lipids and indices of insulin resistance [98]. The obvious gender dimorphism of WHR prompted efforts to evaluate these signals in terms of their potential for different effects in males and females.



Fig. 3.2 Effect sizes for BMI in GIANT meta-analyses vs. BMI-adjusted WHR in GIANT metaanalyses for genome-wide significant BMI and BMI-adjusted WHR loci. In the *scatterplot*, data for BMI in GIANT meta-analyses [28] are shown on the *X*-axis and data for BMI-adjusted WHR in GIANT meta-analyses [98] on the *Y*-axis. The points are colored according to if they represent loci associated with BMI (*red*) or BMI-adjusted WHR (*blue*)

Half of the 14 loci showed evidence of gender-specific effects: in each case, the effect was stronger in women [98].

In a complementary approach to studies of WHR in population-wide analysis, Berndt et al. restricted analysis to the "tails" of the WHR distribution (upper and lower 5th percentiles) and analyzed WHR in terms of dichotomous "case–control" analyses [51]. This analysis demonstrated a similar pattern of association signals as that of previous population-wide analysis [98], indicating that WHR at the "tails" of the distribution has a similar genetic architecture as that of the full distribution.

Genome-Wide Association Studies of Abdominal Fat Distribution

The use of imprecise, but widely available, clinical measures such as WHR facilitates large meta-analysis, but there is much to be gained by complementary analyses in smaller numbers of more carefully phenotyped subjects. In a recent study, more direct measurements of the extent of abdominal subcutaneous and visceral adiposity were obtained by computed tomography (CT) [60]. This analysis was able to demonstrate that the fat distribution association signal near *LYPLAL1* [51, 56, 98] could also be detected using CT (as the ratio between subcutaneous and visceral fat area). It also highlighted a signal near *THNSL2* that was associated with visceral adiposity in women: this survived adjustment for BMI, and has not previously been associated to obesity traits [60].

Genetic Architecture of Fat Distribution

Combined, the 14 loci for BMI-adjusted WHR uncovered by GWAS account for approximately 1 % of variance in this trait (1.34 % in women; 0.46 % in men) [98]. Using methods analogous to those for BMI described above [65], Vattikuti et al. showed that ~13 % of the overall variance in WHR could be explained by common GWAS SNPs, and thus estimated that 46 % of heritability in WHR may be captured by common variants [99]. A similar range of explanation for the missing genetic variance is possible as for overall obesity [63, 64, 67], and ongoing sequence-based efforts will help to define the extent to which this deficit can be plugged by the contribution of low frequency and rare variants.

From GWAS Associations to Potential Functional Roles in Fat Distribution

As with BMI, progress towards characterization of the mechanisms operating at each of these loci has been patchy. Expression-QTL mapping in adipose tissue, blood, and other tissues has identified promising candidate transcripts at six of the loci (*AA553656*, *GRB14*, *PIGC*, *STAB1*, *TBX15*, and *ZNRF3*) [98].

For several of these transcripts, the genetic data integrates well with the corpus of existing biological data. For example, *GRB14*, encoding a growth factor receptorbinding protein, is known to acts as a negative regulator of insulin receptor signalling [100, 101]. The WHR-associated variant shows directionally consistent associations with triglyceride and insulin levels [98] and other (statistically independent) variants at the same locus influence BMI-adjusted insulin and HDL-cholesterol levels [102, 103]. *TBX15* encodes a mesodermal developmental transcription factor and has been indicated in adipocyte differentiation and triglyceride accumulation [104]. This transcript is also differentially expressed between visceral and subcutaneous adipose tissue, and there is evidence that visceral adipose tissue expression is negatively correlated with BMI [105].

The most consistent signal for fat distribution maps to the *LYPLAL1* locus [51, 56, 60, 98]. As might be expected variants at this locus are associated with a range of related metabolic and anthropometric traits including adiponectin [106], fasting insulin adjusted for BMI [103] and height [107]. So far, there is limited evidence to demonstrate that the signal is mediated through the *LYPLAL1* transcript and the region contains several other potential candidates. However, expression of this gene is induced in subcutaneous fat following obesity [108] and its presumed function as a lysophospholipase is consistent with a causal role.

As with the GWAS loci associated with BMI, pathway enrichment approaches have been applied across the 14 WHR-associated loci. Though the enrichment signals were relatively weak, they highlighted developmental processes and mRNA transcript regulation [98]. The known functions of some of the stronger positional

candidates—such as angiogenesis (*VEGFA*), adipocyte differentiation (*GRB14*) and developmental function (*TBX15*, *HOXC13*)—seem to point towards peripheral mechanisms.

These enrichment patterns, when compared with those seen for BMI, seem consistent with the hypothesis that overall obesity is primarily defined by variation at genes involved in central neuroendocrine regulation, whereas fat distribution is largely influenced by variation at genes, which control peripheral aspects of adipose function and development.

Challenges for the Present and for the Future

Whilst there is no doubt that GWAS studies have accelerated our understanding of the genetics and biology of obesity, there remains much to do. At most of the loci discovered, we have yet to identify the causal variant (or variants) or to define with certainty which regional transcript is responsible for mediating the association effect. The accumulation of transethnic association data [109–111] combined with the growing use of next-generation sequencing to generate reference sets for imputation [112] and to interrogate phenotypically selected individuals (e.g., the morbidly obese) should help to address the former. The latter depends in part on the generation of improved annotations (particularly those from relevant tissues) that connect non-coding variation to transcript regulation, and on the development of appropriate functional assays. As always, the ability to refine the phenotypic consequences of allelic differences at variants of interest in human subjects (for example through imaging of fat tissues) will play a crucial role in defining a mechanistic understanding of these traits.

Missing Genetic Variance

As we have seen the loci identified by GWAS loci explain a surprisingly small proportion of phenotypic variance, far less than appears to be the case for other "similar" quantitative traits, such as height and lipids. Approaches that combine effects across the entire GWAS dataset, rather than considering only those signals reaching genome-wide significance, do a better job of recovering variation (indicating a long "polygenic" tail of common variant susceptibility) but still leave a substantial component of estimated heritability unexplained [65]. To discover further genome-wide significant associations to common genetic variants with increasingly smaller effects would require even larger studies than to date. The latest wave of grand meta-analyses of BMI and BMI-adjusted WHR (involving over 320,000 and 210,000 European samples respectively), currently underway, promise to reveal some of these common variant signals, as do the studies emerging from analyses in a variety of non-European samples.

It has been suggested that rare (MAF <1 %) or low frequency (MAF 1–5 %) variants beyond the range of the historical GWAS approach, may contribute to this missing genetic variance [24, 64, 113, 114]. The rapidly decreasing cost and increasing accuracy of next-generation sequencing are bringing variants in this class under the microscope for the first time [115].

It is clear that individual risk of obesity reflects the integration of genetic and non-genetic factors including variation in food availability and extent of physical exercise [116]. Indeed, these may directly interact such that variant effects are modulated by these lifestyle factors: under some circumstances these interactions may contribute to the missing "genetic" variance [67]. The detection of such interactions at the genome scale requires massive sample sizes, unless the interaction terms are substantial. Nevertheless, there are several examples now emerging of interaction effects at obesity loci: these include an interaction between *FTO* and exercise [117] as well as sex-specific effects reported for WHR [56, 98] and visceral adiposity [60].

Risk Prediction, Intervention and Medication

One might hope that improved knowledge about the genetics of obesity would help to generate predictive models. These might be used to identify individuals at highest future risk of obesity who could be targeted for early intervention, and/or define genetic markers related to treatment outcome that can be used to guide therapeutic choices. However, the common variants so far identified by GWAS have too weak an effect, even in combination, to have value in this respect. Indeed, genetic risk factors are currently outperformed by traditional risk factors [118] including present BMI (a good predictor of future obesity risk [119]).

Instead, the most valuable translational benefits are likely to accrue from the biological knowledge, which grows from the genetics. Currently, there are few effective pharmaceutical treatments for obesity, and the most successful clinical intervention requires radical (bariatric) surgery. The clinical burden of obesity urgently requires the identification of novel validated therapeutic targets based around a better understanding of underlying mechanisms. The wider behavioral effects of drugs acting on central processes such as appetite may continue to prove problematic in this respect and efforts to target peripheral mechanisms of fat distribution, and thereby ameliorate the adverse metabolic consequences of obesity may prove more productive.

References

 Finucane MM, Stevens GA, Cowan MJ, Danaei G, Lin JK, Paciorek CJ et al (2011) National, regional, and global trends in body-mass index since 1980: systematic analysis of health examination surveys and epidemiological studies with 960 country-years and 9.1 million participants. Lancet 377(9765):557–567

- 2. Swinburn BA, Sacks G, Hall KD, McPherson K, Finegood DT, Moodie ML et al (2011) The global obesity pandemic: shaped by global drivers and local environments. Lancet 378(9793):804–814
- 3. Stunkard AJ, Foch TT, Hrubec Z (1986) A twin study of human obesity. JAMA 256(1):51-54
- 4. Stunkard AJ, Harris JR, Pedersen NL, McClearn GE (1990) The body-mass index of twins who have been reared apart. N Engl J Med 322(21):1483–1487
- Stunkard AJ (1991) Genetic contributions to human obesity. Res Publ Assoc Res Nerv Ment Dis 69:205–218
- 6. Bouchard C, Tremblay A, Despres JP, Nadeau A, Lupien PJ, Theriault G et al (1990) The response to long-term overfeeding in identical twins. N Engl J Med 322(21):1477–1482
- 7. Elks CE, den Hoed M, Zhao JH, Sharp SJ, Wareham NJ, Loos RJ et al (2012) Variability in the heritability of body mass index: a systematic review and meta-regression. Front Endocrinol (Lausanne) 3:29
- Rose KM, Newman B, Mayer-Davis EJ, Selby JV (1998) Genetic and behavioral determinants of waist-hip ratio and waist circumference in women twins. Obes Res 6(6):383–392
- 9. Mills GW, Avery PJ, McCarthy MI, Hattersley AT, Levy JC, Hitman GA et al (2004) Heritability estimates for beta cell function and features of the insulin resistance syndrome in UK families with an increased susceptibility to type 2 diabetes. Diabetologia 47(4):732–738
- Souren NY, Paulussen AD, Loos RJ, Gielen M, Beunen G, Fagard R et al (2007) Anthropometry, carbohydrate and lipid metabolism in the East Flanders Prospective Twin Survey: heritabilities. Diabetologia 50(10):2107–2116
- Zillikens MC, Yazdanpanah M, Pardo LM, Rivadeneira F, Aulchenko YS, Oostra BA et al (2008) Sex-specific genetic effects influence variation in body composition. Diabetologia 51(12):2233–2241
- 12. Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ et al (1997) Congenital leptin deficiency is associated with severe early-onset obesity in humans. Nature 387(6636):903–908
- Echwald SM, Rasmussen SB, Sorensen TIA, Andersen T, TybjaergHansen A, Clausen JO et al (1997) Identification of two novel missense mutations in the human OB gene. Int J Obes Relat Metab Disord 21(4):321–326
- 14. Oksanen L, Kainulainen K, Heiman M, Mustajoki P, KauppinenMakelin R, Kontula K (1997) Novel polymorphism of the human ob gene promoter in lean and morbidly obese subjects. Int J Obes Relat Metab Disord 21(6):489–494
- Clement K, Vaisse C, Lahlou N, Cabrol S, Pelloux V, Cassuto D et al (1998) A mutation in the human leptin receptor gene causes obesity and pituitary dysfunction. Nature 392(6674):398–401
- 16. Jackson RS, Creemers JWM, Ohagi S, RaffinSanson ML, Sanders L, Montague CT et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 16(3):303–306
- Saunders CL, Chiodini BD, Sham P, Lewis CM, Abkevich V, Adeyemo AA et al (2007) Meta-analysis of genome-wide linkage studies in BMI and obesity. Obesity (Silver Spring) 15(9):2263–2275
- Botstein D, Risch N (2003) Discovering genotypes underlying human phenotypes: past successes for mendelian disease, future approaches for complex disease. Nat Genet 33(Suppl):228–237
- Tabor HK, Risch NJ, Myers RM (2002) Candidate-gene approaches for studying complex genetic traits: practical considerations. Nat Rev Genet 3(5):391–397
- 20. Farooqi IS, Yeo GS, Keogh JM, Aminian S, Jebb SA, Butler G et al (2000) Dominant and recessive inheritance of morbid obesity associated with melanocortin 4 receptor deficiency. J Clin Invest 106(2):271–279
- Hinney A, Volckmar AL, Knoll N (2013) Melanocortin-4 receptor in energy homeostasis and obesity pathogenesis. Prog Mol Biol Transl Sci 114:147–191
- 22. Farooqi S, O'Rahilly S (2006) Genetics of obesity in humans. Endocr Rev 27(7):710-718
- Hirschhorn JN, Daly MJ (2005) Genome-wide association studies for common diseases and complex traits. Nat Rev Genet 6(2):95–108

- 24. McCarthy MI, Abecasis GR, Cardon LR, Goldstein DB, Little J, Ioannidis JP et al (2008) Genome-wide association studies for complex traits: consensus, uncertainty and challenges. Nat Rev Genet 9(5):356–369
- Herbert A, Gerry NP, McQueen MB (2006) A common genetic variant is associated with adult and childhood obesity. Science 312:279–283
- The International HapMap Consortium (2005) A haplotype map of the human genome. Nature 437(7063):1299–1320
- Frayling TM, Timpson NJ, Weedon MN (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316:889–894
- Speliotes EK, Willer CJ, Berndt SI (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42:937–948
- Loos RJ, Lindgren CM, Li S (2008) Common variants near MC4R are associated with fat mass, weight and risk of obesity. Nat Genet 40:768–775
- Willer CJ, Speliotes EK, Loos RJ (2009) Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet 41:25–34
- Thorleifsson G, Walters GB, Gudbjartsson DF (2009) Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. Nat Genet 41:18–24
- 32. Okada Y, Kubo M, Ohmiya H, Takahashi A, Kumasaka N, Hosono N et al (2012) Common variants at CDKAL1 and KLF9 are associated with body mass index in East Asian populations. Nat Genet 44(3):302–306
- 33. Wen W, Cho YS, Zheng W, Dorajoo R, Kato N, Qi L et al (2012) Meta-analysis identifies common variants associated with body mass index in East Asians. Nat Genet 44(3):307–311
- Cotsapas C, Speliotes EK, Hatoum IJ (2009) Common body mass index-associated variants confer risk of extreme obesity. Hum Mol Genet 18:3502–3507
- Park H, Poo MM (2013) Neurotrophin regulation of neural circuit development and function. Nat Rev Neurosci 14(1):7–23
- Beckers S, Zegers D, Van Gaal LF, Van Hul W (2009) The role of the leptin-melanocortin signalling pathway in the control of food intake. Crit Rev Eukaryot Gene Expr 19(4):267–287
- Kim YJ, Sano T, Nabetani T, Asano Y, Hirabayashi Y (2012) GPRC5B activates obesityassociated inflammatory signaling in adipocytes. Sci Signal 5(251):ra85
- Wheeler E, Huang N, Bochukova EG, Keogh JM, Lindsay S, Garg S et al (2013) Genomewide SNP and CNV analysis identifies common and low-frequency variants associated with severe early-onset obesity. Nat Genet 45:513–517
- Bochukova EG, Huang N, Keogh J, Henning E, Purmann C, Blaszczyk K et al (2010) Large, rare chromosomal deletions associated with severe early-onset obesity. Nature 463(7281): 666–670
- 40. Walters RG, Jacquemont S, Valsesia A, de Smith AJ, Martinet D, Andersson J et al (2010) A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. Nature 463(7281):671–675
- 41. Jacquemont S, Reymond A, Zufferey F, Harewood L, Walters RG, Kutalik Z et al (2011) Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. Nature 478(7367):97–102
- 42. Hassanein MT, Lyon HN, Nguyen TT, Akylbekova EL, Waters K, Lettre G et al (2010) Fine mapping of the association with obesity at the FTO locus in African-derived populations. Hum Mol Genet 19(14):2907–2916
- 43. Peters U, North KE, Sethupathy P, Buyske S, Haessler J, Jiao S et al (2013) A systematic mapping approach of 16q12.2/FTO and BMI in more than 20,000 African Americans narrows in on the underlying functional variation: results from the Population Architecture using Genomics and Epidemiology (PAGE) study. PLoS Genet 9(1):e1003171
- 44. Jackson RS, Creemers JW, Ohagi S, Raffin-Sanson ML, Sanders L, Montague CT et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 16(3):303–306

- 45. Benzinou M, Creemers JW, Choquet H, Lobbens S, Dina C, Durand E et al (2008) Common nonsynonymous variants in PCSK1 confer risk of obesity. Nat Genet 40(8):943–945
- 46. Arragain S, Handelman SK, Forouhar F, Wei FY, Tomizawa K, Hunt JF et al (2010) Identification of eukaryotic and prokaryotic methylthiotransferase for biosynthesis of 2-methylthio-N6-threonylcarbamoyladenosine in tRNA. J Biol Chem 285(37):28425–28433
- 47. Steinthorsdottir V, Thorleifsson G, Reynisdottir I, Benediktsson R, Jonsdottir T, Walters GB et al (2007) A variant in CDKAL1 influences insulin response and risk of type 2 diabetes. Nat Genet 39(6):770–775
- 48. Kamatani Y, Matsuda K, Okada Y, Kubo M, Hosono N, Daigo Y et al (2010) Genome-wide association study of hematological and biochemical traits in a Japanese population. Nat Genet 42(3):210–215
- 49. Groenewoud MJ, Dekker JM, Fritsche A, Reiling E, Nijpels G, Heine RJ et al (2008) Variants of CDKAL1 and IGF2BP2 affect first-phase insulin secretion during hyperglycaemic clamps. Diabetologia 51(9):1659–1663
- 50. Kirchhoff K, Machicao F, Haupt A, Schafer SA, Tschritter O, Staiger H et al (2008) Polymorphisms in the TCF7L2, CDKAL1 and SLC30A8 genes are associated with impaired proinsulin conversion. Diabetologia 51(4):597–601
- 51. Berndt SI, Gustafsson S, Magi R, Ganna A, Wheeler E, Feitosa MF et al (2013) Genomewide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. Nat Genet 45:501–512
- 52. Meyre D, Delplanque J, Chevre JC, Lecoeur C, Lobbens S, Gallina S et al (2009) Genome-wide association study for early-onset and morbid adult obesity identifies three new risk loci in European populations. Nat Genet 41(2):157–159
- 53. Jiao H, Arner P, Hoffstedt J, Brodin D, Dubern B, Czernichow S et al (2011) Genome wide association study identifies KCNMA1 contributing to human obesity. BMC Med Genomics 4:51
- 54. Bradfield JP, Taal HR, Timpson NJ, Scherag A, Lecoeur C, Warrington NM et al (2012) A genome-wide association meta-analysis identifies new childhood obesity loci. Nat Genet 44(5):526
- 55. Scherag A, Dina C, Hinney A, Vatin V, Scherag S, Vogel CIG et al (2010) Two new loci for body-weight regulation identified in a joint analysis of genome-wide association studies for early-onset extreme obesity in French and German study groups. PLoS Genet 6(4):e1000916
- 56. Lindgren CM, Heid IM, Randall JC, Lamina C, Steinthorsdottir V, Qi L et al (2009) Genomewide association scan meta-analysis identifies three Loci influencing adiposity and fat distribution. PLoS Genet 5(6):e1000508
- 57. Kilpelainen TO, Zillikens MC, Stancakova A, Finucane FM, Ried JS, Langenberg C et al (2011) Genetic variation near IRS1 associates with reduced adiposity and an impaired metabolic profile. Nat Genet 43(8):753–760
- 58. Rung J, Cauchi S, Albrechtsen A, Shen L, Rocheleau G, Cavalcanti-Proenca C et al (2009) Genetic variant near IRS1 is associated with type 2 diabetes, insulin resistance and hyperinsulinemia. Nat Genet 41(10):1110–1115
- 59. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M et al (2010) Biological, clinical and population relevance of 95 loci for blood lipids. Nature 466(7307):707–713
- 60. Fox CS, Liu Y, White CC, Feitosa M, Smith AV, Heard-Costa N et al (2012) Genome-wide association for abdominal subcutaneous and visceral adipose reveals a novel locus for visceral fat in women. PLoS Genet 8(5):e1002695
- 61. Shu XO, Long J, Cai Q, Qi L, Xiang YB, Cho YS et al (2010) Identification of new genetic risk variants for type 2 diabetes. PLoS Genet 6(9):e1001127
- 62. Imamura M, Iwata M, Maegawa H, Watada H, Hirose H, Tanaka Y et al (2011) Genetic variants at CDC123/CAMK1D and SPRY2 are associated with susceptibility to type 2 diabetes in the Japanese population. Diabetologia 54(12):3071–3077
- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ et al (2009) Finding the missing heritability of complex diseases. Nature 461(7265):747–753

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- 64. Gibson G (2011) Rare and common variants: twenty arguments. Nat Rev Genet 13(2):135–145
- 65. Yang J, Manolio TA, Pasquale LR, Boerwinkle E, Caporaso N, Cunningham JM et al (2011) Genome partitioning of genetic variation for complex traits using common SNPs. Nat Genet 43(6):519–525
- 66. Yang J, Benyamin B, McEvoy BP, Gordon S, Henders AK, Nyholt DR et al (2010) Common SNPs explain a large proportion of the heritability for human height. Nat Genet 42(7):565–569
- Zuk O, Hechter E, Sunyaev SR, Lander ES (2012) The mystery of missing heritability: genetic interactions create phantom heritability. Proc Natl Acad Sci U S A 109(4):1193–1198
- Visscher PM, Hill WG, Wray NR (2008) Heritability in the genomics era–concepts and misconceptions. Nat Rev Genet 9(4):255–266
- 69. Bell CG, Finer S, Lindgren CM, Wilson GA, Rakyan VK, Teschendorff AE et al (2010) Integrated genetic and epigenetic analysis identifies haplotype-specific methylation in the FTO type 2 diabetes and obesity susceptibility locus. PLoS One 5(11):e14040
- Boissel S, Reish O, Proulx K, Kawagoe-Takaki H, Sedgwick B, Yeo GS et al (2009) Loss-offunction mutation in the dioxygenase-encoding FTO gene causes severe growth retardation and multiple malformations. Am J Hum Genet 85(1):106–111
- Meyre D, Proulx K, Kawagoe-Takaki H, Vatin V, Gutierrez-Aguilar R, Lyon D et al (2010) Prevalence of loss-of-function FTO mutations in lean and obese individuals. Diabetes 59(1):311–318
- Stratigopoulos G, Padilla SL, LeDuc CA, Watson E, Hattersley AT, McCarthy MI et al (2008) Regulation of Fto/Ftm gene expression in mice and humans. Am J Physiol Regul Integr Comp Physiol 294(4):R1185–R1196
- Ansley SJ, Badano JL, Blacque OE, Hill J, Hoskins BE, Leitch CC et al (2003) Basal body dysfunction is a likely cause of pleiotropic Bardet-Biedl syndrome. Nature 425(6958): 628–633
- 74. Church C, Lee S, Bagg EA, McTaggart JS, Deacon R, Gerken T et al (2009) A mouse model for the metabolic effects of the human fat mass and obesity associated FTO gene. PLoS Genet 5(8):e1000599
- Church C, Moir L, McMurray F, Girard C, Banks GT, Teboul L et al (2010) Overexpression of Fto leads to increased food intake and results in obesity. Nat Genet 42(12):1086–1092
- Gerken T, Girard CA, Tung YC, Webby CJ, Saudek V, Hewitson KS et al (2007) The obesityassociated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. Science 318(5855):1469–1472
- 77. Gulati P, Cheung MK, Antrobus R, Church CD, Harding HP, Tung YC et al (2013) Role for the obesity-related FTO gene in the cellular sensing of amino acids. Proc Natl Acad Sci U S A 110:2557–2562
- Hinney A, Schmidt A, Nottebom K, Heibult O, Becker I, Ziegler A et al (1999) Several mutations in the melanocortin-4 receptor gene including a nonsense and a frameshift mutation associated with dominantly inherited obesity in humans. J Clin Endocrinol Metab 84(4):1483–1486
- 79. Krude H, Gruters A (2000) Implications of proopiomelanocortin (POMC) mutations in humans: the POMC deficiency syndrome. Trends Endocrinol Metab 11(1):15–22
- Farooqi IS, O'Rahilly S (2008) Mutations in ligands and receptors of the leptin-melanocortin pathway that lead to obesity. Nat Clin Pract Endocrinol Metab 4(10):569–577
- Vanevski F, Xu B (2013) Molecular and neural bases underlying roles of BDNF in the control of body weight. Front Neurosci 7:37
- Dunham I, Kundaje A, Aldred SF, Collins PJ, Davis CA, Doyle F et al (2012) An integrated encyclopedia of DNA elements in the human genome. Nature 489(7414):57–74
- Stranger BE, Forrest MS, Clark AG, Minichiello MJ, Deutsch S, Lyle R et al (2005) Genomewide associations of gene expression variation in humans. PLoS Genet 1(6):e78
- 84. Dimas AS, Deutsch S, Stranger BE, Montgomery SB, Borel C, Attar-Cohen H et al (2009) Common regulatory variation impacts gene expression in a cell type-dependent manner. Science 325(5945):1246–1250

- 85. Borel C, Deutsch S, Letourneau A, Migliavacca E, Montgomery SB, Dimas AS et al (2011) Identification of cis- and trans-regulatory variation modulating microRNA expression levels in human fibroblasts. Genome Res 21(1):68–73
- 86. Rantalainen M, Herrera BM, Nicholson G, Bowden R, Wills QF, Min JL et al (2011) MicroRNA expression in abdominal and gluteal adipose tissue is associated with mRNA expression levels and partly genetically driven. PLoS One 6(11):e27338
- Grundberg E, Small KS, Hedman AK, Nica AC, Buil A, Keildson S et al (2012) Mapping cis- and trans-regulatory effects across multiple tissues in twins. Nat Genet 44(10): 1084–1089
- Parts L, Hedman AK, Keildson S, Knights AJ, Abreu-Goodger C, van de Bunt M et al (2012) Extent, causes, and consequences of small RNA expression variation in human adipose tissue. PLoS Genet 8(5):e1002704
- Ren D, Zhou Y, Morris D, Li M, Li Z, Rui L (2007) Neuronal SH2B1 is essential for controlling energy and glucose homeostasis. J Clin Invest 117(2):397–406
- 90. Saxena R, Hivert MF, Langenberg C, Tanaka T, Pankow JS, Vollenweider P et al (2010) Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. Nat Genet 42(2):142–148
- Wang K, Li M, Hakonarson H (2010) Analysing biological pathways in genome-wide association studies. Nat Rev Genet 11(12):843–854
- 92. Snijder MB, Dekker JM, Visser M, Bouter LM, Stehouwer CD, Kostense PJ et al (2003) Associations of hip and thigh circumferences independent of waist circumference with the incidence of type 2 diabetes: the Hoorn Study. Am J Clin Nutr 77(5):1192–1197
- 93. Shao J, Yu L, Shen X, Li D, Wang K (2010) Waist-to-height ratio, an optimal predictor for obesity and metabolic syndrome in Chinese adults. J Nutr Health Aging 14(9):782–785
- 94. Garg A (2004) Acquired and inherited lipodystrophies. N Engl J Med 350(12):1220–1234
- 95. Chambers JC, Elliott P, Zabaneh D, Zhang W, Li Y, Froguel P et al (2008) Common genetic variation near MC4R is associated with waist circumference and insulin resistance. Nat Genet 40(6):716–718
- 96. Heard-Costa NL, Zillikens MC, Monda KL, Johansson A, Harris TB, Fu M et al (2009) NRXN3 is a novel locus for waist circumference: a genome-wide association study from the CHARGE Consortium. PLoS Genet 5(6):e1000539
- 97. Bille DS, Banasik K, Justesen JM, Sandholt CH, Sandbaek A, Lauritzen T et al (2011) Implications of central obesity-related variants in LYPLAL1, NRXN3, MSRA, and TFAP2B on quantitative metabolic traits in adult Danes. PLoS One 6(6):e20640
- 98. Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, Steinthorsdottir V et al (2010) Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. Nat Genet 42(11):949–960
- 99. Vattikuti S, Guo J, Chow CC (2012) Heritability and genetic correlations explained by common SNPs for metabolic syndrome traits. PLoS Genet 8(3):e1002637
- 100. Holt LJ, Siddle K (2005) Grb10 and Grb14: enigmatic regulators of insulin action–and more? Biochem J 388(Pt 2):393–406
- 101. Nouaille S, Blanquart C, Zilberfarb V, Boute N, Perdereau D, Roix J et al (2006) Interaction with Grb14 results in site-specific regulation of tyrosine phosphorylation of the insulin receptor. EMBO Rep 7(5):512–518
- 102. Ridker PM, Pare G, Parker AN, Zee RY, Miletich JP, Chasman DI (2009) Polymorphism in the CETP gene region, HDL cholesterol, and risk of future myocardial infarction: genomewide analysis among 18 245 initially healthy women from the Women's Genome Health Study. Circ Cardiovasc Genet 2(1):26–33
- 103. Manning AK, Hivert MF, Scott RA, Grimsby JL, Bouatia-Naji N, Chen H et al (2012) A genome-wide approach accounting for body mass index identifies genetic variants influencing fasting glycemic traits and insulin resistance. Nat Genet 44(6):659–669
- 104. Gesta S, Bezy O, Mori MA, Macotela Y, Lee KY, Kahn CR (2011) Mesodermal developmental gene Tbx15 impairs adipocyte differentiation and mitochondrial respiration. Proc Natl Acad Sci U S A 108(7):2771–2776

- 105. Gesta S, Bluher M, Yamamoto Y, Norris AW, Berndt J, Kralisch S et al (2006) Evidence for a role of developmental genes in the origin of obesity and body fat distribution. Proc Natl Acad Sci U S A 103(17):6676–6681
- 106. Dastani Z, Hivert MF, Timpson N, Perry JR, Yuan X, Scott RA et al (2012) Novel loci for adiponectin levels and their influence on type 2 diabetes and metabolic traits: a multi-ethnic meta-analysis of 45,891 individuals. PLoS Genet 8(3):e1002607
- 107. Lango Allen H, Estrada K, Lettre G, Berndt SI, Weedon MN, Rivadeneira F et al (2010) Hundreds of variants clustered in genomic loci and biological pathways affect human height. Nature 467(7317):832–838
- Steinberg GR, Kemp BE, Watt MJ (2007) Adipocyte triglyceride lipase expression in human obesity. Am J Physiol Endocrinol Metab 293(4):E958–E964
- 109. Zaitlen N, Pasaniuc B, Gur T, Ziv E, Halperin E (2010) Leveraging genetic variability across populations for the identification of causal variants. Am J Hum Genet 86(1):23–33
- Morris AP (2011) Transethnic meta-analysis of genomewide association studies. Genet Epidemiol 35(8):809–822
- 111. Franceschini N, van Rooij FJ, Prins BP, Feitosa MF, Karakas M, Eckfeldt JH et al (2012) Discovery and fine mapping of serum protein loci through transethnic meta-analysis. Am J Hum Genet 91(4):744–753
- 112. Abecasis GR, Auton A, Brooks LD, DePristo MA, Durbin RM, Handsaker RE et al (2012) An integrated map of genetic variation from 1,092 human genomes. Nature 491(7422):56–65
- 113. Pritchard JK (2001) Are rare variants responsible for susceptibility to complex diseases? Am J Hum Genet 69(1):124–137
- 114. Pritchard JK, Cox NJ (2002) The allelic architecture of human disease genes: common disease-common variant...or not? Hum Mol Genet 11(20):2417–2423
- 115. Cirulli ET, Goldstein DB (2010) Uncovering the roles of rare variants in common disease through whole-genome sequencing. Nat Rev Genet 11(6):415–425
- 116. Franco M, Bilal U, Ordunez P, Benet M, Morejon A, Caballero B et al (2013) Population-wide weight loss and regain in relation to diabetes burden and cardiovascular mortality in Cuba 1980-2010: repeated cross sectional surveys and ecological comparison of secular trends. BMJ 346:f1515
- 117. Kilpelainen TO, Qi L, Brage S, Sharp SJ, Sonestedt E, Demerath E et al (2011) Physical activity attenuates the influence of FTO variants on obesity risk: a meta-analysis of 218,166 adults and 19,268 children. PLoS Med 8(11):e1001116
- Loos RJ (2012) Genetic determinants of common obesity and their value in prediction. Best Pract Res Clin Endocrinol Metab 26(2):211–226
- 119. Whitaker RC, Wright JA, Pepe MS, Seidel KD, Dietz WH (1997) Predicting obesity in young adulthood from childhood and parental obesity. N Engl J Med 337(13):869–873
- 120. Paternoster L, Evans DM, Nohr EA, Holst C, Gaborieau V, Brennan P et al (2011) Genomewide population-based association study of extremely overweight young adults-the GOYA study. PLoS One 6(9):e24303

Chapter 4 Copy Number Variants and Their Contribution to the Risk of Obesity

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Abstract Obesity is becoming an increasingly serious health concern, given its associated health risks and the growing number of people affected. Understanding the genetic factors underlying body weight regulation and obesity susceptibility has thus become an issue of paramount importance. Obesity has a high estimated heritability, yet much of this remains unexplained. Copy number variants (CNVs) represent a relatively understudied class of genetic variants which may account for some of this unexplained heritability. This chapter explores how copy number variation contributes to body weight regulation and obesity susceptibility.

Common CNVs associated with body mass index (BMI) and obesity have recently been identified, including variants on chromosomes 1p31.3, 8p21.2, 10q11.22, 11q11, and 16p12.3 at the *NEGR1*, *DOCK5*, *PPYR1*, *OR4P4*, *OR4S2*, and *OR4C6*, and *GPRC5B* loci. A number of rare CNVs have also recently been associated with extreme forms of obesity, including two on chromosome 16p11.2, consisting of a 593 kb deletion whose reciprocal duplication has been associated with increased risk of underweight, as well as a 220 kb deletion encompassing the *SH2B1* gene, which has been associated with overweight and obesity. Several studies have also reported enrichment in the global burden of large, rare CNVs among obese subjects, as well as the presence of several rare CNVs uniquely among obese cases.

In the case of large CNVs encompassing multiple genes, functional studies will be required to establish which gene or genes within each CNV are causative for the

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observed phenotype. Improved methodologies, both laboratory and statistical, are also required to enable systematic investigation of complex structural variants such as VNTRs and multi-allelic loci. While developments in this field are promising, analysis of CNVs, both common and rare, has proven challenging, and reported associations require extensive follow-up in large replication samples in order to confirm their role in obesity susceptibility. It is hoped that such in-depth investigations will provide increased understanding of the role of CNVs in body weight regulation and risk of obesity.

Introduction

The study of the genetic factors underlying obesity susceptibility is a subject which has captured the attention of many within the scientific community, particularly due to the serious health risks faced by affected individuals, and the increased risk of obesity in their relatives. This chapter explores the contribution of copy number variants (CNVs) to body weight regulation and risk of obesity.

The Missing Heritability of Obesity

The current obesogenic environment, characterized by an increased consumption of widely available calorie-dense foods among many other factors, has no doubt driven the recent rise in obesity rates [1]. A question of extreme interest in the study of obesity, however, is why individual risk of obesity differs even between subjects exposed to the same environmental risk factors [1]. The answer to this question lies in the fact that obesity is a complex disease arising from a complex interplay of environmental risk factors, affecting all individuals within any given population, and individual genetic predisposition, which renders certain individuals more susceptible to obesity in the face of these environmental risk factors [1].

Despite this complex interaction, numerous studies have shown obesity to be a highly heritable trait. Several twin, adoption, and family studies examining the heritability of adiposity have reported heritability estimates for obesity ranging from approximately 40–70 %, with increased concordance levels between monozygotic twins, even those reared apart, compared to dizygotic twins [2–7].

Conversely, genetic variants associated with adiposity and obesity identified to date explain only approximately 2–4 % of the heritability of these traits [8, 9], with the vast majority of studies having focussed on the analysis of common single nucleotide polymorphisms (SNPs). This discrepancy between the estimated heritability of corpulence and the proportion of which has been explained to date has raised the important question of whether the heritability of obesity has been overestimated, or whether this "missing heritability" [10] could in fact be accounted for by forms of genetic variation not captured by genome-wide association studies (GWAS) of common SNP variants. One such class of variants that have received increased attention in recent years are CNVs.

Introduction to Copy Number Variation

A CNV is defined as a segment of DNA differing in the number of diploid copies carried by individuals within the population [11–14]. CNVs include simple biallelic deletions and duplications, as well as more complex, multi-allelic variants showing highly polymorphic patterns of copy number distribution at the population level (Fig. 4.1).

CNV discovery studies to date all concur that CNVs are widespread throughout the human genome, and are also observed in phenotypically healthy individuals [11–16]. While precise estimates of CNV frequencies and their average size have differed between studies, in the highest resolution genome-wide CNV discovery study carried out to date [14], a total of 8,599 CNVs above 443 bp, covering approximately 3.7 % of the genome, were independently validated, with a median CNV size of 2.7 kb and a median of 1,117 and 1,488 CNVs in European (CEU) and Yoruban African (YRI) subjects, respectively [14]. Of the approximately 5,000 validated CNVs which were subject to further investigation, 77 % were deletions, 16 % were duplications and 7 % were multi-allelic variants, although it is essential to consider that these frequencies may also be influenced in part by the respective ease of detection of these three forms of structural variation [14].

As shown in Fig. 4.2, CNVs were found to overlap 13.4 % of RefSeq genes, with a smaller proportion of deletions than duplications and multi-allelic variants overlapping genes [14]. CNVs were detected genome-wide, with CNVs shown to result in loss of function mutations at over 260 genes [14]. Any two subjects were found to differ in copy number at an average of approximately 0.78 % of the genome, affecting structure of approximately 2.7 % of gene transcripts [14]. Multiple studies have concurred that common bi-allelic CNVs are well-tagged by surrounding SNPs [13, 14, 17], while significantly less linkage disequilibrium has been detected between duplications and multi-allelic variants and their surrounding SNPs [14, 17]. In addition to tandem duplications, numerous dispersed duplications have also been detected, indicating that this may be an overlooked class of CNV [14] (Fig. 4.3).

Population genetic analyses of genomic structural variation thus suggest that CNVs are widely distributed in the human genome, with the majority of CNVs being of small size, with significant overlap between CNVs detected in different subjects [11–16]. Furthermore, CNV hotspots prone to recurrent recombination



Fig. 4.1 Copy number variant (CNV) classes. CNVs may consist of simple deletions or duplications, or more complex rearrangements such as multi-allelic CNVs, where several allelic configurations exist for the same locus, varying in the number of copies of the duplicated region



Fig. 4.2 Functional impact of CNVs in the genome. (a) Overall functional consequences of CNVs, stratified by level of validation and CNV type. (b) Functional impact of CNVs, stratified by CNV type, frequency, and sample geographic origin. YRI: Yoruba in Ibadan, Nigeria; CEU: CEPH (Utah residents with ancestry from northern and western Europe); ASN: Japanese in Tokyo, Japan+Han Chinese in Beijing, China. Figure reproduced with permission from Conrad et al. (2010) [14]

exist in the genome, particularly in the vicinity of segmental duplications [18] and sequence motifs such as *Alu* repeats [14, 19, 20]. In addition to common CNVs with identical breakpoints shared by multiple individuals, a multitude of rare and recurrent CNVs exist, a higher proportion of which overlap genes than do common structural variants, and might thus contribute significantly to interindividual phenotypic differences [14]. Similarly, a higher degree of overlap exists between genes and



Fig. 4.3 Circular plot of genome-wide CNV distribution reported by Conrad and colleagues [14]. The concentric circles depict, from inside to outside, stacked histograms of the numbers of deletions, duplication, and multi-allelic CNVs in *red, green*, and *blue*, respectively, the number of CNVs by mechanism of formation (NAHR, VNTR, and other shown in *blue, red,* and *grey*, respectively), and the degree of population differentiation between the Yoruban and European study samples of detected CNVs in the outermost circle, with the innermost circle depicting the origin and new location of dispersed duplications in the genome. Figure reproduced with permission from Conrad et al. (2010) [14]

complex structural variants such as multi-allelic CNVs and VNTRs, implicating these complex and understudied variants in phenotypic variability and disease susceptibility [14].

CNVs may influence gene expression levels either directly or indirectly through a number of different mechanisms, including deletion or duplication of entire genes, gene-disrupting CNVs, or through long-range effects mediated through disruption or insertion of regulatory elements such as enhancers or repressors [21, 22]. In the case of multi-allelic CNVs encompassing dosage sensitive genes, expression levels



Fig. 4.4 Dosage-sensitive genes. Dosage-sensitive genes are those at which changes in gene copy number result in changes in the quantity of mRNA produced

may be directly correlated with gene copy number [22] (Fig. 4.4). The phenotypic effects of CNVs and their potential contribution to disease susceptibility have thus become a topic of considerable interest.

Copy Number Variation in Adiposity and Obesity Susceptibility

The Contribution of Common Copy Number Variants to Body Weight Regulation

Given the previously noted potential functional influences of CNVs, a natural progression from CNV discovery studies was the investigation of their potential contribution to human disease susceptibility and the so-called "missing heritability" [10] of common diseases.

A large number of SNP association analyses have been conducted to date in both case–control samples and population cohorts for numerous common diseases [23], and the development of CNV prediction algorithms has enabled CNV prediction using these genome-wide SNP array data [24–27]. This has permitted the reuse of these data for CNV association studies. Similar to SNP GWAS, genome-wide CNV association studies have often focussed on common CNVs, usually defined as those having a population frequency above 5 %, with several associations between common CNVs and complex diseases, including obesity, having been reported in recent years.

Marginal association of a common CNV on chr10q11.22 encompassing the pancreatic polypeptide receptor 1 (*PPYR1*) gene with BMI has been reported in a Chinese population sample, with low copy number associated with increased BMI [28]. *PPYR1* ligands have previously been linked to the regulation of food intake in both human and animal studies [29–31], lending support to a potential role for CNVs encompassing this gene in body weight regulation. Furthermore, a common CNV at 11q11 encompassing the olfactory receptor genes *OR4P4*, *OR4S2*, and *OR4C6* has also been reported to show association with early-onset extreme obesity [32].

As well as CNVs consisting of di-allelic variants, more complex structurally variable regions may also contribute to increased risk of disorders such as obesity [33]. We have recently shown a complex copy number variable region on chromosome

8p21.2 to be significantly associated with susceptibility to severe obesity [33]. The region encompasses two variable number tandem repeats (VNTRs) flanking a 3,975 bp common deletion. Two of these three variants are located within the dedicator of cytokinesis gene (*DOCK5*), and all three structural variants were shown to be significantly associated with *DOCK5* gene expression levels [33]. The *DOCK5* gene is a member of the DOCK family of guanine nucleotide exchange factors (GEFs) [34], which are thought to be involved in a variety of cellular functions such as growth, differentiation, regulation of the actin cytoskeleton, vesicle transport, cell signalling, cell movement, phagocytosis, and apoptosis [35] through their role in the activation of members of the Rho/Rac-family GTPases [34]. Further investigation is required in order to establish the precise mechanism by which CNVs within the *DOCK5* region contribute to obesity susceptibility.

In addition to studies directly measuring copy number, some studies have also identified common CNVs potentially contributing to disease susceptibility through linkage disequilibrium with nearby SNPs. Using this approach, two common CNVs, one upstream of *NEGR1* and another near *GPRC5B*, have been linked to body weight through association of tagging SNPs with BMI in two large meta-analyses [8, 36]. Although the effect sizes observed at each of these loci were small, given the LD between these structural variants and their tagging SNPs, it has been suggested that these variants could potentially be causal variants [8, 36].

In spite of these reports, the role of common CNVs in disease susceptibility remains an issue of contention, with little replication of reported associations. A large study conducted by the Wellcome Trust Case Control Consortium (WTCCC) reported association of common CNVs at IRGM and TSPAN8 with Crohn's disease and type 2 diabetes, respectively, as well as association of copy number at the HLA locus with each of Crohn's disease, rheumatoid arthritis, and type 1 diabetes [37]. However, apart from these reported hits, the authors found little evidence of association between common CNVs included in their analyses and any of the eight complex diseases in their study. The authors did however highlight the complexity of CNV prediction and association studies, reporting the confounding effects of several sources of systematic bias such as DNA source and quality, as well as batch effects, on CNV analyses [37]. Moreover, the authors also acknowledged that due to the extensive challenges in assaying more complex structural variants such as multiallelic CNVs and VNTRs, their study was largely limited to common bi-allelic CNVs [37]. These observations highlight the need for additional investigation of the role of CNVs in complex disease susceptibility, focussing in particular on complex structural variants.

The Role of Rare Genomic Structural Variants in Adiposity and Risk of Obesity

Given the large proportion of the estimated heritability of obesity which remains unexplained, it has been suggested that some of this "missing heritability" [10] may



Fig. 4.5 UCSC genome browser view of the proximal 16p11.2 CNV region. The presence of two segmental duplications with high sequence similarity (depicted in *red*) results in the recurrent occurrence of deletions and duplications of the intervening 593 kb segment of unique DNA sequence in this region. Plot generated using the UCSC genome browser [56]

be accounted for by the collective effect of a large number of individually rare variants, each of large effect size [38]. Consistent with what has been observed in the case of SNPs, an increasing body of evidence is supporting the potential contribution of rare CNVs to susceptibility to complex diseases such as obesity, which will be the focus of this section.

Rare CNVs are generally defined as those with frequencies below 1 % in the general population [39, 40]. The rarity of these CNVs generally means that they are not well-tagged by surrounding common SNPs genotyped on GWAS panels. Given the inherent difficulties in accurately genotyping CNVs, analysis of rare CNVs has also principally focussed on variants of large size, often above 200–500 kb [39, 40]. Several large, rare CNVs have thus been reported to show association with body weight and risk of obesity.

Structural Variants Within the 16p11.2 Region

Several CNVs have been identified within the 16p11.2 region, with CNVs at two loci in this region showing association with either underweight, or increased risk of overweight or obesity [38, 39, 41, 42].

Copy Number Variation at the Proximal 16p11.2 Locus

In 2010, we reported association of a heterozygous deletion on chromosome 16p11.2 (chr16: 29,514,353–30,107,356) with highly increased risk of obesity [38]. The deletion encompasses 593 kb of unique sequence and contains 29 genes (Fig. 4.5), including multiple candidates for the obesity phenotype. The presence of two segmental duplications with high sequence similarity renders this locus prone to de novo structural rearrangements (Fig. 4.5), resulting in the occurrence of both deletions and duplications of the intervening DNA sequence [38].

This deletion was initially identified in our study at a frequency of approximately 2.9 % in a study sample of patients suffering from obesity-plus syndromes, whereby patients presented with obesity coupled with additional clinical features such as developmental delay and/or congenital abnormalities [38]. Further investigation revealed an additional 22 deletion carriers among subjects referred to clinical services for cognitive impairment, and 19 subjects among obesity case–control and population GWAS samples [38]. This variant was also reported concurrently in a study by Bochukova et al. [39].

Deletions at this locus resulted in a 30-fold increase in risk of obesity and 43-fold increased risk of morbid obesity, and were identified in 0.7 % of morbidly obese subjects included in our analysis [38]. The obesity phenotype observed among deletion carriers was frequently coupled with hyperphagia, suggesting it to be of potentially neurological origin. While no gender bias was detected in our analysis, an age-dependent effect for this CNV was observed, where penetrance of the obesity phenotype in deletion carriers was positively correlated with subject age [38]. A 0.4–0.7-fold reduction in gene expression levels was also observed for transcripts of genes located within the deleted segment, suggesting that haploinsufficiency for one or more of these genes may be causative for the obesity phenotype observed in deletion carriers [38]. In addition to increased risk of obesity, the deletion was also associated with increased head circumference [38].

A recent study also confirmed the association of this deletion with macrocephaly, but also reported significantly reduced cognitive functioning and an increased frequency of gross motor delay among deletion carriers [43]. Psychiatric comorbidities were reported in greater than 80 % of deletion carriers, while penetrance of obesity was over 70 % of carriers of the deletion in this study sample [43].

Apart from its association with obesity, copy number variation at this 16p11.2 locus has previously been associated with neurodevelopmental and psychiatric conditions, implicating this locus in a number of phenotypes. Both microdeletions and microduplications of the same locus at 29.5 Mb in the 16p11.2 region were shown to be associated with increased risk of autism spectrum disorders (ASD), accounting for approximately 1 % of ASD cases in one study [44]. On the other hand, duplications, but not deletions, at this locus were also linked to increased susceptibility to schizophrenia [45]. These findings have since been replicated in a number of studies [46–48], confirming the contribution of these loci to increased risk of these disorders, and raising the interesting question of the interrelationship between the obesity and neurodevelopmental and psychiatric phenotypes associated with copy number variation at this locus.

A retrospective analysis of 16p11.2 deletions in a clinical sample of approximately 7,000 subjects—the majority of whom had presented with phenotypes such as developmental delay, autism spectrum disorder (ASD) or dysmorphism—identified 28 deletion carriers among this sample [49]. The age-dependence and juvenile onset of the obesity phenotype was confirmed, with obesity generally developing within the first decade of life. Furthermore, a gender-dependence for the 16p11.2 was reported, with male deletion carriers exhibiting a more severe phenotype than female carriers [49]. The incidence of obesity among deletion carriers diagnosed with ASD was also noted to be higher than among autistic subjects not carrying the deletion, providing further support for the independent association between deletions at this locus and increased risk of obesity [49].

In a second study in 2011, we investigated the impact of the reciprocal 16p11.2 duplication on body mass and head circumference [41]. In a fascinating example of a mirror effect of gene dosage at this locus on phenotype, the reciprocal 16p11.2 duplication was associated with strongly increased risk of being underweight, with carriers of this duplication showed significantly reduced postnatal weight and BMI compared to non-duplication carriers [41]. For the purpose of this study, underweight was defined as a BMI $\leq 18.5 \text{ kg/m}^2$ in adults and BMI z-score ≤ 2 standard deviations from the mean for age and sex in children [41]. Underweight can have serious health repercussions, and is frequently associated with failure to thrive during childhood, eating and feeding disorders, as well as anorexia nervosa. Despite the potentially serious nature of this condition, little is known of the factors underlying its genetic susceptibility [50]. In this analysis, 50 % of the male duplication carriers under the age of 5 were diagnosed with a failure to thrive, while adult carriers of this duplication showed an 8.3-fold increased risk of being clinically underweight [41]. A gender effect was also observed, with males showing a trend towards increased severity. In addition to its observed effect on weight, the duplication was also associated with an increased frequency of restrictive and selective eating behaviors, mirroring the hyperphagic phenotype observed in carriers of the reciprocal deletion [41]. Similarly, duplication carriers were noted to show significant reduction in head circumference, which mirrored the macrocephaly associated with the reciprocal deletion [38].

The 16p11.2 duplication was also observed at a higher frequency among medically ascertained patients, recruited on the basis of developmental and cognitive delay or psychiatric phenotypes, than in non-medically ascertained population cohorts in this study, supporting the previously reported association of this duplication with cognitive, neurodevelopmental, and psychiatric phenotypes [44, 45].

A 220 kb Deletion on Chromosome 16p11.2 Encompassing the SH2B1 Gene

In addition to the previously described proximal 16p11.2 deletion and duplication shown to be associated with body weight regulation, additional CNVs within the 16p11.2 region have also been associated with obesity susceptibility.

A 220 kb deletion in this region encompassing nine genes, including the SH2B adaptor protein 1 (*SH2B1*) gene, has been reported to be associated with severe, hyperphagic, early-onset obesity [39]. Although carriers of this deletion have been reported to exhibit elevated fasting plasma insulin levels, conflicting observations have also been reported [39, 51].

SH2B1 is known to enhance leptin and insulin signalling, and animal studies have shown mice harboring homozygous null mutations in the SH2B1 gene to



Fig. 4.6 The chromosome 16p11.2 region. (a) Association results for SNPs in the 16p11.2 region with BMI in a recent meta-analysis carried out by the GIANT consortium [8] Chromosome 16 genomic coordinates are plotted on the *x* axis, with minus $\log_{10}(P$ -value) plotted on the *y* axis. An association peak can be seen at approximately 28.8 Mb. Plot generated using LocusZoom [57]. (b) The positions of two genomic structural variants associated with adiposity levels are depicted. A 220 kb deletion at chr16: 28.73–28.95 Mb and a 593 kb deletion at chr16: 29.51–30.11 Mb have been associated with obesity [38, 39] while a duplication of the latter 593 kb of unique sequence has also been associated with risk of being underweight [41]. The genes falling within each of the two CNVs are also shown. *CNV* copy number variant, *GIANT* Genetic Investigation of ANthropometric Traits, *SNP* single nucleotide polymorphism. First published in *Nature Reviews Endocrinology*, 2013, doi: 10.1038/nrendo.2013.57 by Nature Publishing Group

exhibit signs of metabolic syndrome, with a phenotype including obesity, hyperphagia and insulin resistance [52]. SNPs within *SH2B1* have also shown association with BMI in several meta-analyses [8, 36, 53], making it a strong candidate for the obesity phenotype observed in carriers of this 220 kb deletion. Figure 4.6 depicts association results for SNPs within the 16p11.2 region from a recent BMI meta-analysis, as well as the positions and gene content of both CNVs within the 16p11.2 region described in this chapter.

In addition to its association with severe obesity, this deletion encompassing *SH2B1* has also been linked to developmental delay. In an analysis of a clinical sample of approximately 23,000 patients referred for array comparative genome hybridization (aCGH) for phenotypic abnormalities including developmental delay and cognitive deficits, this medically ascertained sample was found to be enriched for this deletion, and assessment of additional anthropometric data available for a subset of the deletion carriers supported its association with early-onset obesity [42].

Global Burden of Rare Copy Number Variants in Obesity

In addition to the analysis of individual CNVs and their contribution to obesity susceptibility, another area of particular interest is whether the global burden of large, rare CNVs may be higher among subjects suffering from obesity. This is assessed by comparing the total number of rare CNVs above a defined size threshold observed in obese cases versus normal-weight control subjects.
Large, rare deletions have been reported to be enriched among obese cases compared to normal-weight controls in case–control analyses of global CNV burden [39, 40]. In these analyses, large CNVs were found to be overrepresented among obese cases, with this enrichment driven largely by deletions [39, 40]. Furthermore, a larger effect was observed when the analysis was limited to those CNVs which disrupt genes [40], highlighting the potential significance of genes located within these variants to obesity susceptibility.

Rare CNVs Present Exclusively in Cases

Another method of identifying CNVs which might be relevant to obesity susceptibility is to identify CNVs observed exclusively in obese cases and not in normalweight controls. One study identified 17 CNVs present exclusively in three or more Caucasian obese subjects, eight of which were also observed only among African American obese subjects and no normal-weight controls [54]. Their presence solely in obese cases might suggest a potential role for these variants in obesity susceptibility, and replication of these observations in study samples of different ethnicities provides further support for their relevance to the pathogenesis of obesity [54].

While several studies have provided intriguing evidence for the involvement of rare CNVs on obesity susceptibility, it is essential to note that in the analysis of rare variants, wider replication in larger study samples will be necessary to firmly establish their contribution to disorders such as obesity.

From Genetic Variants to Their Physiological Impact: The Importance of Follow-up Studies in CNV Analyses

The identification of structural variants, both common and rare, associated with obesity susceptibility is providing insight into its pathogenetic origins and helping explain some of the missing heritability of this disorder. However, similar to what is observed in the case of common SNPs, there is often difficulty in translating these genetic findings into clear understanding of the underlying biological pathways and mechanisms responsible for this disease. In the case of CNVs, this problem is compounded by the fact that CNVs are often large and may encompass several genes, making it difficult to decipher which gene or genes are responsible for the phenotypic effects observed. Furthermore, CNVs may also have long-range effects, with variants shown to influence expression levels of genes up to several megabases away.

For this reason, it is important to follow up genetic associations with functional studies which attempt to understand how specific variants affect phenotype. As previously discussed, deletion and duplication of a 593 kb region on chromosome 16p11.2 has been associated with a mirror effect on various phenotypes, one of which is head circumference. Duplication of this region has been associated with microcephaly [41], while the reciprocal deletion has been associated with increased

head circumference. Through systematic over-expression and knockdown of each of the orthologous genes within the CNV region in zebrafish, the gene responsible for the variation in head circumference associated with copy number in this region was shown to be the potassium channel tetramerization domain containing 13 (*KCTD13*) gene [55]. Further studies of this type should be undertaken to identify the causal gene or genes for the obesity phenotype associated with the proximal 16p11.2 CNV. Similarly, functional exploration of other CNVs reported to be associated with obesity susceptibility would help in better delineating their physiological effects and in identifying the causative genes located within them.

Future Directions in the Study of CNVs in Obesity

The contribution of CNVs—both common and rare—to obesity susceptibility is becoming increasingly recognized, with progressively more reports of CNVs associated with adiposity levels. However, in spite of this mounting body of evidence, our understanding of the contribution of structural variants to complex diseases such as obesity remains rudimentary, particularly in the case of rare CNVs.

Extensive replication studies including larger numbers of subjects are now required in order to study reported structural variants more comprehensively, verify their reported associations with obesity susceptibility and provide better estimates of their effect sizes. As previously discussed, functional studies will also be necessary to uncover the underlying mechanisms by which such variants may contribute to body weight regulation.

Furthermore, novel methodologies, both technical and statistical, will be required to enable more systematic investigation of complex CNVs such as multi-allelic CNVs. It is hoped that such further in-depth analyses of structural variation may improve our understanding of the genetics factors underlying susceptibility to complex diseases such as obesity.

References

- 1. Friedman JM (2003) A war on obesity, not the obese. Science 299(5608):856-858
- 2. Stunkard AJ, Foch TT, Hrubec Z (1986) A twin study of human obesity. JAMA 256(1):51-54
- Stunkard AJ, Harris JR, Pedersen NL, McClearn GE (1990) The body-mass index of twins who have been reared apart. N Engl J Med 322(21):1483–1487
- 4. Stunkard AJ, Sorensen TI, Hanis C, Teasdale TW, Chakraborty R, Schull WJ et al (1986) An adoption study of human obesity. N Engl J Med 314(4):193–198
- Turula M, Kaprio J, Rissanen A, Koskenvuo M (1990) Body weight in the Finnish Twin Cohort. Diabetes Res Clin Pract 10(Suppl 1):S33–S36
- 6. Maes HH, Neale MC, Eaves LJ (1997) Genetic and environmental factors in relative body weight and human adiposity. Behav Genet 27(4):325–351
- Wardle J, Carnell S, Haworth CM, Plomin R (2008) Evidence for a strong genetic influence on childhood adiposity despite the force of the obesogenic environment. Am J Clin Nutr 87(2):398–404

- Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU et al (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42(11):937–948
- Day FR, Loos RJ (2011) Developments in obesity genetics in the era of genome-wide association studies. J Nutrigenet Nutrigenomics 4(4):222–238
- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ et al (2009) Finding the missing heritability of complex diseases. Nature 461(7265):747–753
- 11. de Smith AJ, Tsalenko A, Sampas N, Scheffer A, Yamada NA, Tsang P et al (2007) Array CGH analysis of copy number variation identifies 1284 new genes variant in healthy white males: implications for association studies of complex diseases. Hum Mol Genet 16(23): 2783–2794
- Korbel JO, Urban AE, Affourtit JP, Godwin B, Grubert F, Simons JF et al (2007) Paired-end mapping reveals extensive structural variation in the human genome. Science 318(5849): 420–426
- 13. Redon R, Ishikawa S, Fitch KR, Feuk L, Perry GH, Andrews TD et al (2006) Global variation in copy number in the human genome. Nature 444(7118):444–454
- Conrad DF, Pinto D, Redon R, Feuk L, Gokcumen O, Zhang Y et al (2010) Origins and functional impact of copy number variation in the human genome. Nature 464(7289):704–712
- Sebat J, Lakshmi B, Troge J, Alexander J, Young J, Lundin P et al (2004) Large-scale copy number polymorphism in the human genome. Science 305(5683):525–528
- Iafrate AJ, Feuk L, Rivera MN, Listewnik ML, Donahoe PK, Qi Y et al (2004) Detection of large-scale variation in the human genome. Nat Genet 36(9):949–951
- Locke DP, Sharp AJ, McCarroll SA, McGrath SD, Newman TL, Cheng Z et al (2006) Linkage disequilibrium and heritability of copy-number polymorphisms within duplicated regions of the human genome. Am J Hum Genet 79(2):275–290
- Sharp AJ, Locke DP, McGrath SD, Cheng Z, Bailey JA, Vallente RU et al (2005) Segmental duplications and copy-number variation in the human genome. Am J Hum Genet 77(1): 78–88
- de Smith AJ, Walters RG, Coin LJ, Steinfeld I, Yakhini Z, Sladek R et al (2008) Small deletion variants have stable breakpoints commonly associated with alu elements. PLoS One 3(8):e3104
- Conrad DF, Bird C, Blackburne B, Lindsay S, Mamanova L, Lee C et al (2010) Mutation spectrum revealed by breakpoint sequencing of human germline CNVs. Nat Genet 42(5):385–391
- Stranger BE, Forrest MS, Dunning M, Ingle CE, Beazley C, Thorne N et al (2007) Relative impact of nucleotide and copy number variation on gene expression phenotypes. Science 315(5813):848–853
- Pollex RL, Hegele RA (2007) Copy number variation in the human genome and its implications for cardiovascular disease. Circulation 115(24):3130–3138
- 23. Hindorff LA, MacArthur J (European Bioinformatics Institute), Wise A, Junkins HA, Hall PN, Klemm AK, Manolio TA. A Catalog of Published Genome-Wide Association Studies. Available at www.genome.gov/gwastudies. Accessed 11 Jun 2012
- 24. Coin LJ, Asher JE, Walters RG, Moustafa JS, de Smith AJ, Sladek R et al (2010) cnvHap: an integrative population and haplotype-based multiplatform model of SNPs and CNVs. Nat Methods 7(7):541–546
- 25. Wang K, Li M, Hadley D, Liu R, Glessner J, Grant SF et al (2007) PennCNV: an integrated hidden Markov model designed for high-resolution copy number variation detection in whole-genome SNP genotyping data. Genome Res 17(11):1665–1674
- 26. Colella S, Yau C, Taylor JM, Mirza G, Butler H, Clouston P et al (2007) QuantiSNP: an Objective Bayes Hidden-Markov Model to detect and accurately map copy number variation using SNP genotyping data. Nucleic Acids Res 35(6):2013–2025
- Peiffer DA, Le JM, Steemers FJ, Chang W, Jenniges T, Garcia F et al (2006) High-resolution genomic profiling of chromosomal aberrations using Infinium whole-genome genotyping. Genome Res 16(9):1136–1148
- Sha BY, Yang TL, Zhao LJ, Chen XD, Guo Y, Chen Y et al (2009) Genome-wide association study suggested copy number variation may be associated with body mass index in the Chinese population. J Hum Genet 54:199–202

- 29. Batterham RL, Le Roux CW, Cohen MA, Park AJ, Ellis SM, Patterson M et al (2003) Pancreatic polypeptide reduces appetite and food intake in humans. J Clin Endocrinol Metab 88(8):3989–3992
- Schmidt PT, Naslund E, Gryback P, Jacobsson H, Holst JJ, Hilsted L et al (2005) A role for pancreatic polypeptide in the regulation of gastric emptying and short-term metabolic control. J Clin Endocrinol Metab 90(9):5241–5246
- Kamiji MM, Inui A (2007) Neuropeptide y receptor selective ligands in the treatment of obesity. Endocr Rev 28(6):664–684
- 32. Jarick I, Vogel CI, Scherag S, Schafer H, Hebebrand J, Hinney A et al (2011) Novel common copy number variation for early onset extreme obesity on chromosome 11q11 identified by a genome-wide analysis. Hum Mol Genet 20(4):840–852
- 33. El-Sayed Moustafa JS, Eleftherohorinou H, de Smith AJ, Andersson-Assarsson JC, Couto Alves A, Hadjigeorgiou E et al (2012) Novel association approach for variable number tandem repeats (VNTRs) identifies DOCK5 as a susceptibility gene for severe obesity. Hum Mol Genet 21(16):3727–3738
- 34. Cote JF, Vuori K (2002) Identification of an evolutionarily conserved superfamily of DOCK180-related proteins with guanine nucleotide exchange activity. J Cell Sci 115(Pt 24):4901–4913
- 35. Bustelo XR, Sauzeau V, Berenjeno IM (2007) GTP-binding proteins of the Rho/Rac family: regulation, effectors and functions in vivo. Bioessays 29(4):356–370
- 36. Willer CJ, Speliotes EK, Loos RJ, Li S, Lindgren CM, Heid IM et al (2009) Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet 41(1):25–34
- Craddock N, Hurles ME, Cardin N, Pearson RD, Plagnol V, Robson S et al (2010) Genomewide association study of CNVs in 16,000 cases of eight common diseases and 3,000 shared controls. Nature 464(7289):713–720
- 38. Walters RG, Jacquemont S, Valsesia A, de Smith AJ, Martinet D, Andersson J et al (2010) A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. Nature 463(7281):671–675
- Bochukova EG, Huang N, Keogh J, Henning E, Purmann C, Blaszczyk K et al (2010) Large, rare chromosomal deletions associated with severe early-onset obesity. Nature 463(7281): 666–670
- 40. Wang K, Li WD, Glessner JT, Grant SF, Hakonarson H, Price RA (2010) Large copy-number variations are enriched in cases with moderate to extreme obesity. Diabetes 59(10): 2690–2694
- 41. Jacquemont S, Reymond A, Zufferey F, Harewood L, Walters RG, Kutalik Z et al (2011) Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. Nature 478(7367):97–102
- 42. Bachmann-Gagescu R, Mefford HC, Cowan C, Glew GM, Hing AV, Wallace S et al (2010) Recurrent 200-kb deletions of 16p11.2 that include the SH2B1 gene are associated with developmental delay and obesity. Genet Med 12(10):641–647
- 43. Zufferey F, Sherr EH, Beckmann ND, Hanson E, Maillard AM, Hippolyte L et al (2012) A 600 kb deletion syndrome at 16p11.2 leads to energy imbalance and neuropsychiatric disorders. J Med Genet 49(10):660–668
- 44. Weiss LA, Shen Y, Korn JM, Arking DE, Miller DT, Fossdal R et al (2008) Association between microdeletion and microduplication at 16p11.2 and autism. N Engl J Med 358(7):667–675
- 45. McCarthy SE, Makarov V, Kirov G, Addington AM, McClellan J, Yoon S et al (2009) Microduplications of 16p11.2 are associated with schizophrenia. Nat Genet 41(11):1223–1227
- 46. Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J et al (2008) Structural variation of chromosomes in autism spectrum disorder. Am J Hum Genet 82(2):477–488
- 47. Kumar RA, KaraMohamed S, Sudi J, Conrad DF, Brune C, Badner JA et al (2008) Recurrent 16p11.2 microdeletions in autism. Hum Mol Genet 17(4):628–638
- Sebat J, Lakshmi B, Malhotra D, Troge J, Lese-Martin C, Walsh T et al (2007) Strong association of de novo copy number mutations with autism. Science 316(5823):445–449

- 49. Yu Y, Zhu H, Miller DT, Gusella JF, Platt OS, Wu BL et al (2011) Age- and gender-dependent obesity in individuals with 16p11.2 deletion. J Genet Genomics 38(9):403–409
- 50. Bulik CM, Slof-Op't Landt MC, van Furth EF, Sullivan PF (2007) The genetics of anorexia nervosa. Annu Rev Nutr 27:263–275
- 51. Walters R, Coin LJM, Ruokonen A, de Smith AJ, El-Sayed Moustafa JS, Jacquemont S, Elliott P, Esko T, Hartikainen AL, Laitinen J, Männik K, Martinet D, Meyre D, Nauck M, Schurmann C, Sladek R, Thorleifsson G, Thorsteinsdóttir U, Valsesia A, Waeber G, Zufferey F, Balkau B, Pattou F, Metspalu A, Völzke H, Vollenweider P, Stefansson K, Jarvelin MR, Beckmann JS, Froguel P, Blakemore AIF (2013) Rare genomic structural variants in complex disease: lessons from the replication of associations with obesity. PLoS One 8:e58048
- 52. Ren D, Li M, Duan C, Rui L (2005) Identification of SH2-B as a key regulator of leptin sensitivity, energy balance, and body weight in mice. Cell Metab 2(2):95–104
- Doche ME, Bochukova EG, Su HW, Pearce LR, Keogh JM, Henning E et al (2012) Human SH2B1 mutations are associated with maladaptive behaviors and obesity. J Clin Invest 122(12):4732–4736
- 54. Glessner JT, Bradfield JP, Wang K, Takahashi N, Zhang H, Sleiman PM et al (2010) A genomewide study reveals copy number variants exclusive to childhood obesity cases. Am J Hum Genet 87(5):661–666
- 55. Golzio C, Willer J, Talkowski ME, Oh EC, Taniguchi Y, Jacquemont S et al (2012) KCTD13 is a major driver of mirrored neuroanatomical phenotypes of the 16p11.2 copy number variant. Nature 485(7398):363–367
- 56. Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM et al (2002) The human genome browser at UCSC. Genome Res 12(6):996–1006
- Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP et al (2010) LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics 26(18): 2336–2337

Chapter 5 Genetics of Childhood Obesity

Struan F.A. Grant

Abstract Obesity is increasingly becoming a major health issue for both the USA and the rest of the world, and presents health care systems with a huge economic problem. The rate at which children are becoming obese is dramatically increasingly, particularly since the turn of the twenty-first century. Although environmental factors are known to play a key role, childhood obesity is also known to have an underlying genetic component contributing to its complex etiology. Elucidating the genetic architecture of childhood obesity will not only help prevention and treatment of pediatric cases but also will have fundamental implications for diseases that present later on in life. Furthermore, the execution of genome-wide surveys of childhood obesity have uncovered novel loci that turned out not to be within the detection range in an adult setting as a consequence of environmental factor clouding, supporting the notion that the pediatric setting may be optimal for uncovering obesity genes. This new era of genome-wide association studies (GWAS) is delivering compelling signals associated with obesity, particularly with peer research groups sharing a very strong consensus on what the key loci are that contribute to the pathogenesis of this trait. Although we suggest that the pediatric setting can be harnessed for obesity gene discovery, the fact is that most BMI-associated loci identified to date were found in the adult setting, so there is a requirement to elucidate which of these variants contribute early on in life and therefore predisposing an individual to related diseases in later life. In this chapter, we outline what advances have been made in determining which genetic factors are conferring their effects on childhood obesity and which ones go on to have an impact in adulthood.

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Epidemiology of Childhood Obesity

Obesity is considered a major health concern for all industrialized societies, with an ever increasing incidence occurring specifically in children [1]. This disorder, along with the associated insulin resistance [2, 3], is known to be the underpinning of a number of major causes of death in the USA largely due to acting as a key risk factor for type 2 diabetes, cardiovascular disease, and other chronic diseases.

Understanding the early indictors for risk of obesity could play a crucial role in combating this mortality rate. Although an obese adult may not have been necessarily obese as a child, the converse is much more notable, where roughly three quarters of obese adolescents go on to present with obesity in adulthood [4–6]. Apart from the obvious chronic comorbidities and increased overall mortality for obese children in later life [7], where overweight children followed over decades [8, 9] are more likely to have cardiovascular and digestive diseases, they also have to contend with the main direct adverse effects of presenting with this trait, which include orthopedic complications, sleep apnea, and psychosocial disorders [10, 11].

Obesity, as presented as excess in adipose tissue mass, presents when there is imbalance between energy intake and energy expenditure. However, what is now considered a disadvantage and recognized as a disease, could well have been an advantage in previous times, under the "thrifty phenotype" hypothesis, when food availability was much more scarce and physical activity was a more natural part of everyday life [12].

Overweight and obesity is defined by the World Health Organization (WHO) as "abnormal or excessive fat accumulation that may impair health" [13]. In order to ascertain a metric of excess body fat, the most readily available method is to simply leverage an individual's height and weight to determine their body mass index (BMI), defined as kilograms divided by meters squared (kg/m²). Indeed this metric has been shown to correlate well with fat content in most people, where adults with a BMI equal to 25-30 are defined as being overweight while a BMI greater than or equal to 30 is the definition of adult obesity. Although an excepted standard in the adult setting, BMI does not serve the same purpose in children well, due to large fluctuations tracking with pubertal status, age, and gender; instead, a BMI-for-age percentile is used to ascertain a sense of pediatric adiposity, where a BMI from 85th to 95th percentile is considered by the Center for Disease Control and Prevention as overweight, while a BMI equal to or greater than the 95th percentile is considered obese [14, 15]. Once a child reaches late adolescence, these percentiles do start to serve as relatively good proxies for adult definitions, where the 95th BMI-for-age percentile gets closer to representing a BMI of 30 kg/m² [11].

Getting a good handle on the prevalence of childhood obesity has proven challenging due to no internationally accepted definition currently existing. That said, the widely held view is that childhood obesity has reached epidemic levels in the developed world. For instance, in the USA approximately a quarter of all children in the USA are considered overweight and approximately one in ten are obese. More empirically, in the decade separately the two National Health and Nutrition Examination Surveys (NHANES) II (1976–1980) and NHANES III (1988–1991), the prevalence of overweight children in the USA increased by 40% [1]. Indeed, when one looks at many countries, the distribution of BMI is clearly becoming increasingly skewed toward overweight [16], with the lower part of the distribution having changed relatively little, while the upper part has widened substantially. As such, it is increasingly obvious that children are becoming more susceptible to obesity, either as a consequence of genetics or exposures in the environment.

A Genetic Component to Childhood Obesity

Despite societal changes plus strong behavioral and environmental factors, there is very strong evidence that there is a genetic component to obesity pathogenesis [17, 18]. As such, elucidating the genetic architecture of childhood obesity could have fundamental implications for both treatment and prevention of many diseases occurring much later on in life.

Twin studies have revealed much about the genetic component to many complex traits, where monozygotic twins are completely genetically identical while nonidentical dizygotic twins only share 50% of their genetic material. When looking in the context of fat mass, the concordance among monozygotic twins has been shown to be approximately 80% while only approximately 40% in dizygotic twins [19–21].

Adoption and family studies have yielded even further evidence. For instance, adopted children have a strong correlation with the BMI of their biological parents but not their adoptive parents [22]. Furthermore, identical twins are significantly concordant for BMI while their nonidentical counterparts are not [23].

Looking at prevalence difference in racial/ethnic groups presents further clues of a genetic component to obesity, such as 35% or less observed in Caucasian and Asian populations while a prevalence 50% or more is seen in Pima Indians and South Sea Island populations [24].

All this genetic epidemiological evidence points to a substantial inherited component to obesity; however due to the obvious interactions with environmental factors, it has proved challenging to tease apart and characterize the genetic component to this trait.

Pre-GWAS Approaches

Linkage scans in families allow for a non-hypothesis approach to assess regions of the genome shared within and across families presenting with a given trait. In the case of the common form of childhood obesity, a number of loci have been reported, but the underlying causative event has still to be elucidated. On the other hand, syndromic forms of childhood obesity have been readily solved using this approach, with chromosomal loci for Prader–Willi syndrome [25], Alström's syndrome [26], and Bardet–Biedl syndrome [27–29] having been mapped.

Single gene disorders that present with obesity features have given us the first insight in to the genetic etiology of this trait, with early studies in rodents shedding much needed light on the issue.

The *ob/ob* mutant mouse [30, 31], exhibiting excess adipose tissue, revealed a mutation in the leptin gene [32, 33], with another strain of severely obese mice, *db/db*, revealing a mutation in its receptor [34]. Variants within these genes have subsequently been reported for human obesity-related traits [35–39], in particular a skipped exon 16 in the human leptin receptor gene leading to impaired growth hormone secretion, early-onset morbid obesity, and failure of pubertal development [40].

A notable developmental trajectory for children with disturbances of the hypothalamic leptin–melanocortin pathway as a whole has been reported. Mutations in the pro-opiomelanocortin (*POMC*) gene have been shown to impact metrics of early onset obesity in children [41–44]. In addition, individuals with genetic mutations in the *PCSK1* gene, which encodes neuroendocrine-specific prohormone convertase 1/3 (PC1/3), present with childhood obesity, hyperphagia, diarrhea, pituitary hypofunction, and disordered glucose homeostasis [45–47].

MC4R is widely considered to be the first established gene to confer morbid human obesity when mutated. Its encoded protein also plays a vital role in the hypothalamic leptin–melanocortin pathway. Multiple nonsense and missense mutations have now been reported in MC4R, many of which are strongly correlated with obesity related traits [48–50].

Brain-derived neurotrophic factor (BDNF) is a downstream target of MC4R activity and has also been implicated in the pathogenesis of childhood obesity, most notably a chromosomal inversion leading to the loss of one functional copy of *BDNF* in an 8-year-old girl, resulting in increased food intake, severe early-onset obesity, hyperactivity, and cognitive impairment [51].

Other loci implicated include a Y722C missense variant in *NTRK2* causing severe obesity and impaired memory in an 8-year-old boy [52] and haploinsufficiency of *SIM1* leading to severe early-onset human obesity due to a balanced translocation between chromosomes 1p22.1 and 6q16.2 [53].

More pronounced syndromes of obesity were our only means to isolate genetic factors before genome-wide association studies (GWAS) emerged after such technology became available around 2005. These classical approaches provided key insights in to the underlying mechanisms involved in energy homoeostasis and are now being complemented by the findings arising from GWAS.

Genome-Wide Association Studies

As outlined above, it has become relatively clear that family-based linkage analyses have had limited success in isolating genes contributing to obesity, particularly the common form of the disease, largely due to the fact that this approach is not well suited to detect common variants in the population conferring relatively modest risk [54, 55]. Candidate gene association studies have also struggled to bear fruit as such

approaches are limited to known biology of the given trait mechanisms; indeed from GWAS approaches described below, many of the key loci identified were never on anybody's candidate gene list.

Conversely, the GWAS approach has empowered investigators to execute a more comprehensive and unbiased strategy to identify causal genes related to complex traits, including obesity, through non-hypothesis based methodologies.

GWAS was made possible by the International HapMap project, which arose out of the human genome sequencing project. This large-scale effort went about systematically characterizing human sequence variation, a vital precursor to comprehensively investigate the genetic basis of complex disease [54–56]. Genome-wide genotyping of in excess of 500,000 single nucleotide polymorphisms (SNPs) can now be readily achieved in an efficient, cost effective and highly accurate manner [57–60]. These SNPs represented on the arrays coming out of these efforts are not selected based being putatively causal, rather they are statistically selected to simply act as "tag-SNPs" for capture of common haplotypic variation information stored in a given region of the human genome. This approach has much higher resolution than the previous linkage approaches for complex traits, where a given signal signifies that the underlying causative variant is typically within just a few hundred kilobases of the tag-SNP.

Unlike the linkage and candidate gene eras that tackled complex traits, GWAS has proven to be a very successful approach yielding robust associations that fellow researchers can replicate and agree on (see the continually updated NIH Catalog of Published Genome-Wide Association Studies at http://www.genome.gov/gwastudies).

Findings from First GWAS Analyses of Obesity

In the past 6 years, tens of genetic loci have been implicated and established for BMI from the outcomes of GWAS, but primarily in adults. These findings will be briefly outlined below in order to give context to the pediatric findings made subsequently.

The first GWAS-implicated locus for obesity was close to the insulin-induced gene 2 (*INSIG2*) gene employing only employing 100,000 SNPs [61]. The tag-SNP, rs7566605, captured the association, which represented a common genetic event with modest relative risk (relative risk=~1.2). The locus was reported to be associated with both adult and childhood obesity and in individuals of both European and African American and ancestry. However, this study has been largely not replicated by other investigative groups [62–66] and disagreement on this observation remains in the obesity research community.

On the other hand, the second obesity locus to be reported, within the fat massand obesity-associated gene (*FTO*) gene [67], has been extensively replicated [68–71], including children [72]. *FTO* is now widely regarded as the most strongly associated obesity locus reported to date [69]. Of note, this locus was actually implicated in type 2 diabetes initially, from one of the first GWAS of that disease [73, 74] but it became very obvious early on in the analyses that the primary trait was obesity susceptibility which was in turn impairing glycemic control [67]. It was subsequently shown that the minor allele of the *FTO* tag-SNP, rs9939609, is correlated almost exclusively to greater fat mass and that it influences fat distribution [75].

Like almost every other GWAS-implicated locus, the causative variant at the *FTO* locus has still to be determined. In addition, the mechanism by which *FTO* confers its effect on the pathogenesis of obesity is still far from clear. It is known that the gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase [76], is expressed in areas of the brain that influence appetite [77] and as such may explain its association with increased energy intake [78]. Compelling studies of both *FTO* knockout and *FTO* over-expressing mice strongly support the role of this gene in the regulation of energy intake and metabolism, showing that a lack of *FTO* expression leads to leanness and the converse promotes obesity [79, 80].

A sequencing effort in Caucasians, consisting of primarily adults reported a set of exonic mutations in FTO; however, these variants largely did not confer risk for obesity [81]; in addition, a comparable sequencing endeavor in African American children drew the same conclusion [82]. These data show that exonic events are unlikely to be the process by which this gene confers its effect, rather it is more likely to harbor a causative event somewhere in the regulatory machinery of FTO.

Meta-analyses

With the sample sizes required and the cost to run the arrays, GWAS represents a sizeable investment. In order to get the maximum from their datasets, investigators subsequently combine their datasets with other groups in order to discover additional loci as a consequence of the extra statistical power gained. In addition, imputation can enable an increase in the number of SNPs available for analysis in these existing datasets [83] through computationally inferring them based on neighboring variant frequencies.

Although these "meta-analyses" represent a substantial statistical power gain, the additional loci detected do have substantially smaller effects than *FTO*, but do provide additional insights in to the biology of the BMI/obesity phenotype.

The first GWAS meta-analysis of BMI, again primarily in adults, revealed a signal that coincided with the well-known *MC4R* gene [84]. The GIANT consortium then revealed six more genes [transmembrane protein 18 (*TMEM18*), potassium channel tetramerization domain containing 15 (*KCTD15*), glucosamine-6-phosphate deaminase 2 (*GNPDA2*), SH2B adaptor protein 1 (*SH2B1*), mitochondrial carrier 2 (*MTCH2*), and neuronal growth regulator 1 (*NEGR1*)] [85], five of which were confirmed in an Icelandic GWAS (but not *GNPDA2* due to an unavailable proxy SNP), who also uncovered and reported loci on 1q25, 3q27, and 12q13 [86] and verified association with the brain-derived neurotrophic factor (*BDNF*) gene [87].

The largest meta-analysis reported to data, by the GIANT consortium, revealed multiple additional BMI loci through the leveraging of data available on 249,796

individuals [88]. Thirty-two loci reached genome-wide significance, of which ten were known from the BMI studies described above, four were known loci from previous studies of weight and/or waist-hip ratio, namely, *SEC16B*, *TFAP2B*, *FAIM2*, *NRXN3*, and eighteen were entirely novel BMI loci, namely, *RBJ-ADCY3-POMC*, *GPRC5B-IQCK*, *MAP2K5-LBXCOR1*, *QPCTL-GIPR*, *TNNI3K*, *SLC39A8*, *FLJ35779-HMGCR*, *LRRN6C*, *TMEM160-ZC3H4*, *FANCL*, *CADM2*, *PRKD1*, *LRP1B*, *PTBP2*, *MTIF3-GTF3A*, *ZNF608*, *RPL27A-TUB*, and *NUDT3-HMGA1*. Interestingly, apart from the *GPRC5B* association to SNPs, a 21 kb associated deletion was identified 50 kb upstream of this gene. This study also made use of a pediatric cohort to provide further support for their findings.

The same study group subsequently went on to look at extremes of the distribution in 263,407 individuals of European ancestry and identified 7 additional loci (*HNF4G*, *RPTOR*, *GNAT2*, *MRPS33P4*, *ADCY9*, *HS6ST3* and *ZZZ3*) that contributed to clinical classes of obesity [89].

Testing Adult-Discovered Loci in Children

There is increasing evidence that that many of the common complex diseases observed in adults have their developmental origins in childhood, in particular obesity, and the path to these disorders are laid out at a young age, or even *in utero* [90, 91]. As described above, a number of genetic loci have now been established to be robustly associated with adult BMI so it would be interesting to know how these loci operate in childhood to see if they confer risk for the pediatric form of obesity.

Leveraging an existing GWAS dataset of pediatric BMI variation from 6,000 children, investigators were able to ask if these SNPs influenced this trait [92]. Nine of the loci in fact did reveal evidence of association with pediatric BMI, of which the *FTO* locus was the strongest. *TMEM18* followed by *GNPDA2* were the next most strongly associated adult-implicated loci, showing a similar magnitude to that of *FTO* in this pediatric setting. The remaining weaker loci were *INSIG2*, *MC4R*, *NEGR1*, 1q25, *BDNF* and *KCTD15* (Table 5.1). These findings were much in line with the findings made in the initial adult report, where they checked in a smaller pediatric setting [85].

Going on to check the full 32 loci reported in the more recent GIANT metaanalysis through the leveraging of 1,097 childhood obesity cases (BMI \geq 95th percentile CDC definition), together with 2,760 lean controls (defined as BMI <50th percentile), aged between 2 and 18 years old [93], the same investigative group reported evidence of association for nine of the loci, namely, at *FTO*, *TMEM18*, *NRXN3*, *MC4R*, *SEC16B*, *GNPDA2*, *TNN13K*, *QPCTL*, and *BDNF*. Overall, 28 of the 32 loci revealed directionally consistent effects to that of the adult BMI metaanalysis. As such, it is abundantly clear that the majority of obesity-conferring variants initially uncovered in adults are indeed operating early on in life; however these adult-discovered loci only explain less than 2% of the total variation for BMI in children, which is less striking than reported by the GIANT consortium in their adult cohorts [85] so many more genes need to be characterized.

Category	Loci	Citations
Adult BMI GWAS loci also associated with childhood BMI/obesity in independent studies	FTO, TMEM18, GNPDA2, INSIG2, MC4R, NEGR1, 1q25, BDNF, KCTD15, POMC, FAIM2, TNNI3K, SEC16B, GNPDA2, BDNF, NRXN3, QPCTL	[92, 93, 109]
Adult 2 type diabetes GWAS loci also associated with childhood BMI/obesity	HHEX-IDE	[94]
GWAS of extreme childhood obesity—novel loci	SDCCAG8, TNKS-MSRA	[108]
GWAS of common childhood obesity (BMI≥95th percentile)—novel loci	OLFM4, HOXB5	[109]
CNV analyses of childhood obesity—novel loci	SH2B1, EDIL3, S1PR5, FOXP2, TBCA, ABCB5, ZPLD1, KIF2B, ARL15, EPHA6-UNQ6114, OR4P4-OR4S2-OR4C6	[126, 127, 129 131]

 Table 5.1
 Childhood obesity loci that have been identified to date and the context by which they were implicated

Similarly, a number of loci have been implicated by GWAS for type 2 diabetes and their mechanism of action in this regard is equally far from clear. Using a similar tactic, the same investigative group analyzed the role of these loci with respect to childhood BMI [94]. Interestingly, only a single locus showed association, where the same variant in *IDE-HHEX* that increases type 2 diabetes risk also showed evidence of association with increased pediatric BMI. This finding provides some guidance on how this particular locus is conferring its diabetic effect.

Subsequently, a comparable study was carried out with loci implicated by GWAS for influencing adult bone mineral density and/or osteoporosis risk [95–99] and found that the same variation near *Osterix* that increases BMD is also associated with higher BMI in girls [100], further implicating body size and skeletal loading in its potential mechanism of action.

FTO in Childhood

As *FTO* represents the first robustly established locus associated with obesity, it has received much more attention to date than other similarly found loci due to the fact that it has been known longest, including in the pediatric setting.

It has become clear from cross-sectional studies that there is an age-dependent effect of *FTO* genetic variation on BMI. Although there is consistent evidence that *FTO* has no obvious impact on fetal growth or birth weight, there is marked effect by the age 7 years old [67]; however, one study has suggested that there is in fact a negative association between *FTO* and BMI before the age of 2 years old, but a

positive correlation after that age, peaking at approximately 20 years old followed by a decline [101].

Longitudinal cohort studies have added context to some of these initial reports. Specific statistical modeling has already been developed that can characterize important milestones in pediatric BMI trajectories. For children with normal growth patterns, BMI rises steeply after birth until reaching "infancy peak" [102], which has been suggested to be at a median of 7 months old [103]. Following this maximum, BMI declines gradually, and then reaches a low point called the "adiposity rebound" [104], which occurs just before the onset of puberty, and then continues to rise until adulthood. Adiposity rebound can occur anywhere in the range of 2-7 years old, but typically in the 4–5 year old age band [103]. Children carrying the obesity risk variants harbored at the FTO locus do not just present with a higher BMI throughout their lives, but also present with a specific developmental trajectory that is known to lead to higher incidences of future obesity [105] and comorbidities [106]. In a meta-analysis of eight Caucasian pediatric cohorts, adult obesity associated FTO alleles were associated with lower BMI at adiposity peak, higher BMI at adiposity rebound, and most interestingly, with an earlier age of adiposity rebound [107], thus supporting the initial study suggesting a negative association in the less than 2 years old age band. This in turn reflects a possible steeper downward inflection of BMI trajectory after infancy peak leading towards an earlier adiposity rebound in those genetically predisposed individuals [107].

Loci Specifically Identified in Childhood Obesity GWAS Analyses

A widely recognized notion is that the distillation of the genetic component for a number of complex traits, including obesity, should be easier in children, where the period of environmental exposure is substantially. As such, if a child is presenting with a given trait that is also seen in adults, it is more likely that has presented due to a genetic predisposition rather than a prolonged environmental stressor.

Although multiple studies have revealed many loci in the context of relatively simple syndromic forms of the disease, there has been relatively little progress in identifying genes that directly impact the less extreme, common form of childhood obesity; after all this is the primary trait that is on the massive increase observed over recent years.

The first attempt at a full GWAS of childhood obesity was carried out in 2010, where French and German study groups carried out a joint analysis of genome-wide genotyped data generated on their early-onset obesity cohort [108], defined as BMI in the range of the 97th to the 99th percentile; as a consequence, two novel loci were identified, namely, *SDCCAG8* and *TNKS/MSRA* (Table 5.1). In the initial step of the analyses, association was tested for both genotyped and imputed SNPs in a combined French and German sample consisting of 1,138 extremely obese children plus 1,120 normal or underweight controls. For the follow-up replication

attempts, all SNPs that yielded a degree of strong evidence for association were genotyped and tested in an independent cohort of 1,181 obese children and 1,960 normal- or underweight controls plus 715 nuclear families with at least one extremely obese offspring. Although this observation for these two loci was relatively compelling, their effect within the most recent adult GIANT meta-analysis [88] was marginally associated at best, suggesting that the effect was limited to a relatively extreme pediatric setting.

Subsequently, a large-scale meta-analysis was carried out with a definition of the disease with a less extreme cutoff, i.e., BMI >95th percentile, in order to address the more common form of childhood obesity [109]. This study consisted of 14 existing GWAS datasets, made up of 5,530 cases plus 8,318 controls defined at the <50th percentile of BMI. Apart from robustly detecting seven known loci, namely, FTO, TMEM18, POMC, MC4R, FAIM2, TNNI3K, and SEC16B, this study also detected two novel loci when taking any signal that reached a $P < 5 \times 10^{-5}$ in the discovery stage in to the replication stage, namely, rs9568856 near OLFM4 and rs9299 within HOXB5. It became clear that these loci were also associated with adult BMI when querying the large GIANT meta-analysis dataset, but were not detected by those investigators in that setting as the signals were below detection at the genome-wide level. As such, this pediatric model did detect additional variants that were below the bandwidth in the environmentally more complex setting of adulthood; in addition, the study "rediscovered" loci like TNNI3K at the fraction of the cohort size that was required in the adult setting. These two loci are also functionally interesting where studies in mice have shown that OLFM4 plays a role in the host gastric mucosa immune response against H. pylori infection [110], while HOXB5 is a member of the homeobox transcription factor family and has been implicated in gut development and fat loss [111, 112]. As such, these findings point to a possible role of the gut in determining BMI in early childhood and beyond, and presents investigators with possibly novel therapeutic entry points in to both preventing and treating the disease.

A subsequent study performed SNP association analyses in 1,509 children at the very extreme end of the obesity tail, i.e., greater than 3 standard deviations from the mean of the BMI distribution plus 5,380 controls [113]. Following up 29 SNPs that reached $P < 1 \times 10^{-5}$ in the discovery stage in an additional 971 severely obese children and 1,990 controls revealed four new loci, namely, *LEPR*, *PRKCH*, *PACS1*, and *RMST*. This is clearly different from what was observed in the more common form of the disease and could be possibly used in the future to partition the disease in to different forms of obesity, i.e., sub-categorization.

Other Ethnicities

Genomes from people of different ancestries can provide geneticists with a lot of information. In particular if a locus is found to confer risk of a disease in multiple populations, then that genomic region will be considered to have more global relevance to the disease and thus may be more attractive targets and pathways from a potential diagnostic and therapeutic point-of-view. In addition, cohorts of African ancestry have been leveraged on multiple occasions to fine map a locus down to the closest point to the underlying causative variant by taking advantage of the fact that these populations are more ancient, have had more time for genetic recombination and therefore have, on average, shorter stretches of linkage disequilibrium, e.g., the association of T2D with *TCF7L2* [114] has been refined utilizing a West African patient cohort [115].

With respect to outcomes form GWAS analyses of obesity, the bulk of those studies have been carried out in Caucasians to date. This is partly due to the relatively low haplotypic complexity of genomes from European ancestral populations and thus less SNPs are needed on the array to capture the bulk of the common variation, but is also partly to best deal with issues related to admixture. When considering the well-established association between *FTO* and obesity, and the fact that it shows the strongest association with BMI among children European ancestry [92], the picture has been less clear investigating the correlation in populations of African ancestry [70, 116]. However, there is growing consensus from recent large cohort studies in both adults [117] and children [72], that SNP rs3751812 captures the *FTO* association with the trait in both ethnicities.

GWAS meta-analyses for BMI and obesity are now emerging for other ethnicities, and with respect to the first African American effort [118], where a pediatric cohort of the same ethnicity was used to support the findings. In addition to supporting 32 of the 36 loci previously reported in Caucasians, the large group of investigators found robust evidence for association at loci harboring *GALNT10* and *MIR148A-NFE2L3* plus suggestive support for association at *KLHL32*.

Missing Heritability

A major restriction built in to any GWAS approach in that it is based on the concept of the common disease, common variant hypothesis, where it is presumed that the genetic component to the complex trait under study falls in to that category. It has turned out to be partly correct, where a proportion of such a genetic component is due to a moderate number of common variants, but which individually only explain a small proportion of predicted genetic susceptibility to the trait in a population. Like most complex and common diseases, tens of loci have now been identified for obesity, but only a small proportion of the calculated heritability has been explained, representing approximately 10% of the estimated heritability in most disease settings. In fact, the statistically robust obesity associations with *FTO* and *MC4R* only account for less than 2% of the variance in adult BMI, with the combined results of all obesity GWAS loci found to date still only accounting for a very small proportion of the heritability of BMI [85, 119]. This has led to a great deal of debate on what the missing heritability could look like [120], with the main consensus settling around the notion that it will be made up of much rarer variants, copy number variants and epigenetic changes which are not within the detection range of GWAS [121], rather they will need to be characterized through the use of whole genome sequencing related approaches.

Interestingly, there is a counter argument to this prevailing view, where it could be said that the current estimates of missing heritability are incorrect. In particular, Zuk et al. [122] suggested that the estimated missing heritability not detected by GWAS may in fact be hugely overinflated, due to the fact that the community has assumed incorrectly that genetic variants contributing to a given trait do not interact with each other. Even if this interaction occurred at a relatively modest level, then missing heritability estimates would need to be substantially reassessed.

Irrespective of some of these estimate, it is clear that additional variants need to be found that cannot be readily picked up by GWAS and some alternative approaches are outlined below.

Rare SNPs

Commercially available genotyping arrays have typically restricted content to simply feature products comprehensive coverage of common variants with a minor allele frequency greater than 5% based on information gained from the HapMap. However, this changing to a degree as more information comes out from the successor to the HapMap, the 1000 Genomes project, where newer arrays are ensuring comprehensive coverage down to a 1% frequency plus exome coverage. However this information is still far from exhaustive as not every variant known in the genome can as yet be represented on this platform and so signals may be missed. For instance, a rare SNP within intron 4 of *NAMPT* has been implicated in patients with severe obesity [123] but would not be detected with standard high throughput genotyping methods. As such, genotyping array products will need to be much more comprehensive in genomic coverage going forward or extremely large sample sizes will be required to overcome multiple testing statistical power issues in whole genome sequencing setting in order to elucidate the remaining genetic component of obesity, the latter of which remains cost prohibitive.

Copy Number Variation

Genomic rearrangements resulting in deletions, duplications, inversions, and translocations are collectively referred to as copy number variants (CNVs). As described above, much of the genetic diversity in the human genome is due to single base pair variations, but there is also variation in copy number throughout the genome too [124]. With the array technology available for GWAS-related approaches, the underlying single-base extension (SBE) biochemistry and hybridization/detection to synthetic oligonucleotides allows for accurate genotyping and quantitation of allelic copy number [58, 125]. As such SNP arrays allow for characterization of CNVs genome-wide for a given individual [57, 58, 125].

CNVs have been particularly implicated in subjects with the combination of extreme obesity and coexisting developmental delay (Table 5.1). Analyses in this regard has revealed what is now considered the most established CNV in the obesity field to date, namely, a large, rare chromosomal deletion of at least 593 kb at 16p11.2. Two groups in the UK plus their collaborators independently detected rare deletions at this chromosomal location, observing that they were present in excess among extreme obese cases when compared with normal and regular obese subjects [126, 127]. Heterozygotes for this deletion at this location are significantly enriched in Caucasian patients with severe early-onset obesity and developmental delay [127, 128]. These deletions, albeit very rare, have been shown not to be present in healthy non-obese controls but in 0.7% of morbidly obese cases (BMI >40 kg/m²), resulting in a striking odds ratio of 43.0. This deleted region, estimated to be in the size range of 220 kb to 1.7 Mb, and thus coincides with a number of genes making it more challenging to determine which is the causative gene. However, Bochukova et al. [126] observed that SH2B1 was the only consistently impacted genes when deleted regions were overlaid among the five cases studied. In addition, it is well known that SH2B1 plays a role in leptin and insulin signaling and energy homeostasis [126], plus common SNPs near SH2B1 locus have been strongly associated with BMI in GWAS analyses [85, 126].

Examining CNVs in common childhood obesity is a good complement to what was described above in the more extreme setting. Investigators examined children in the upper 5th percentile of BMI but excluded any subject greater than 3 standard deviations in order to rigorously adhere to the pursuit of genetic factors underpinning the common form the disease rather than rare syndromes [129] (Table 5.1). Through the use of a European American pediatric cohort at the discovery stage to detect any associated events, a very conservative bar was set for the replication stage, where the observed CNVs also had to be exclusive to cases in a second ethnicity, i.e., African Americans. 34 putative CNVR loci (15 deletions and 19 duplications) that were exclusive to EA cases were detected in a cohort of European American (EA) childhood obesity cases ($n = \sim 1,000$) and lean controls (n = 2,500). However, following a physical validation step to ensure the events were real, three of the putative deletions turned out to be false positives during the quantitative PCR (qPCR) checks. Ultimately 17 of these CNV impacted loci were unique to the cohort, not appearing in any public databases, so those findings were further evaluated in the independent African American (AA) cohort (n = -1,500) of childhood obesity cases and lean controls ($n = \sim 1,500$). Interestingly, almost half of these variants were also exclusive to AA cases (6 deletions and 2 duplications). The established loci from this study that were impacted by a deletion were EDIL3, S1PR5, FOXP2, TBCA, ABCB5, and ZPLD1, while KIF2B and ARL15 were impacted by a duplication event. When the AA cohort was used as the discovery set, evidence was also gained for a deletion at the EPHA6-UNQ6114 locus.

As with findings from GWAS, most of the loci represented novel biology for obesity, with no reports of these genes playing a role in the disease previously.

Although it was comforting to see, from a positive control point of view, that one locus came up that had been implicated in a related trait previously, i.e., *ARL15* in a GWAS of adiponectin levels, a known risk factor for CVD and T2D [130].

In the same study, the presence of large rare deletions in childhood obesity was assessed, where they had to present in <1% of individuals and >500 kb in size. The result was that no excess of large rare deletions was observed genome-wide. This is consistent with the 16p11.2 story, where significance in this regard was only achieved once developmental delay subjects were included but not severe early-onset obesity on its own [126, 127].

A novel common CNV for early onset extreme obesity was subsequently reported on chromosome 11q11, harboring a cluster of related genes OR4P4, OR4S2, and OR4C6 genes [131] (Table 5.1). Furthermore, a very recent study performed CNV association analyses in 1,509 children at the extreme tail obesity (>3 standard deviations from the mean) plus 5,380 controls [113]. A previously reported 43 kb deletion at the *NEGR1* locus was significantly associated with the trait but it appeared that this signal was entirely driven by a flanking 8 kb deletion; absence of this deletion increased the risk for obesity dramatically. They also reported significant burden of rare, single CNVs in severely obese cases (P < 0.0001), while integrative gene network pathway analysis of rare deletions indicated enrichment of genes affecting G protein-coupled receptors (GPCRs) involved in the neuronal regulation of energy homeostasis. This pathway is of course great relevance to the pathogenesis of obesity.

As greater and greater resolution genome-wide scans are executed, one should envisage further reports of such findings. The hunt for CNVs in the context of childhood obesity has proven fruitful up to now and it has become quite clear they contribute to the missing heritability for the trait.

Epigenetics

Looking beyond the underlying DNA sequence to explain the reminder of the missing heritability of childhood obesity and other complex traits, one should also consider heritable changes in gene expression or cellular phenotype, in particular when they are caused by DNA methylation and histone modification.

Prader–Willi is a syndrome that presents with developmental defects, cognitive disabilities, excessive eating and pronounced life-threatening obesity. This imprinted disorder is due to genetic and epigenetic errors in the region of chromosome 15q11-q13. The phenotype occurs when the paternal copy is deleted, while the maternal copy is inactivated by methylation [132, 133].

Although the underlying genes have yet to be identified, linkage scans of European and African ancestral populations have revealed that several obesity-related genetic loci have different parental effects or maternal effects [134]. In addition, variation in DNA methylation patterns on the human leptin promoter has been

observed between alleles and cells, suggesting that imprinting occurs at this locus and that its actions are cell type specific.

Observations like these strongly point to epigenetic mechanisms contributing to obesity risk, but of course this still needs to be fully elucidated. New techniques, such as ChIP-Seq [135] and whole genome sequencing, will provide us with much needed insight in to global methylation patterns and histone modifications in the context of childhood obesity.

Conclusions

Only until recently, genetic studies of childhood obesity were largely reliant on family studies, animal models and candidate gene efforts, with monogenic obesity providing crucial clues. However, GWAS has now taken on much of the heavy lifting with respect to gene discovery in the context of the more polygenic form of the disease, with its unbiased, whole genome scan approach.

Although such studies have revealed several new biomolecular pathways not previously associated with obesity, it is still a concern to many complex trait geneticists that the well-established and robust associations only explain very little of the genetic risk for a given complex, in particular childhood obesity, suggesting that more loci are out there to find but may be conferring their effects more subtly or through rare variants.

The current situation has given the genetics community pause for thought on what the best strategy is to adopt in order to fill in the gaps and elucidate the full repertoire of the genetic component of a given trait so that the current "missing heri-tability" [120] can ultimately be all accounted for. It is becoming increasingly clear that larger and larger cohorts combined in to mega meta-analyses will be required, but new discoveries can only be made when leveraging the ultimate resolution of whole genome sequencing technologies rather than the variant snapshots we get with GWAS. Although not a reality yet, these approaches will become feasible as the price of sequencing continues to drop.

In addition, as GWAS is designed with representative tag-SNPs present on the array, the signals that we are seeing and robustly associated with given traits are in no way to be considered the underlying causal susceptibility variant at a given loci. The move from association to causality remains a big challenge for common complex diseases like obesity so new approaches to characterize the true causative genes will need to be developed and subsequent functional studies will be required to fully understand how they feed in to the risk profile for childhood obesity. This in turn will help us produce more efficacious therapies and will guide us on the path to personalized medicine.

Clearly using the pediatric model can aid in elucidating the role of many variants first shown to be associated with an adult trait. As shown above, many of the loci initially found for BMI determination in adults turn out to operate early on in life and influence childhood obesity risk. And as we suggest, leveraging the pediatric model may be a more efficient way to distill out genetic variants underpinning many complex traits, being more cost effective in sample collection and genotyping, due to the fact that the environmental confounder is substantially less in the childhood setting. As such, recognizing this angle can increase the efficiency of efforts to detect, treat, and ultimately, prevent obesity and its comorbidities.

References

- Troiano RP, Flegal KM (1998) Overweight children and adolescents: description, epidemiology, and demographics. Pediatrics 101:497–504
- Reaven GM (1988) Banting lecture 1988. Role of insulin resistance in human disease. Diabetes 37:1595–1607
- DeFronzo RA, Ferrannini E (1991) Insulin resistance. A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. Diabetes Care 14:173–194
- 4. Nicklas TA, Baranowski T, Cullen KW, Berenson G (2001) Eating patterns, dietary quality and obesity. J Am Coll Nutr 20:599–608
- Whitaker RC, Wright JA, Pepe MS, Seidel KD, Dietz WH (1997) Predicting obesity in young adulthood from childhood and parental obesity. N Engl J Med 337:869–873
- 6. Parsons TJ, Power C, Logan S, Summerbell CD (1999) Childhood predictors of adult obesity: a systematic review. Int J Obes Relat Metab Disord 23(Suppl 8):S1–S107
- 7. Must A (2003) Does overweight in childhood have an impact on adult health? Nutr Rev 61:139–142
- 8. Mossberg HO (1989) 40-year follow-up of overweight children. Lancet 2:491-493
- Must A, Jacques PF, Dallal GE, Bajema CJ, Dietz WH (1992) Long-term morbidity and mortality of overweight adolescents. A follow-up of the Harvard Growth Study of 1922 to 1935. N Engl J Med 327:1350–1355
- Dietz WH (1998) Health consequences of obesity in youth: childhood predictors of adult disease. Pediatrics 101:518–525
- 11. Daniels SR et al (2005) Overweight in children and adolescents: pathophysiology, consequences, prevention, and treatment. Circulation 111:1999–2012
- Eckel RH (2003) Obesity: a disease or a physiologic adaptation for survival? In: Eckel RH (ed) Obesity mechanisms and clinical management. Lippincott, Williams & Wilkins, Philadelphia, PA, pp 3–30
- 13. World Health Organization (2012) Obesity and overweight. Fact sheet no 311. WHO, Geneva
- Flegal KM, Wei R, Ogden C (2002) Weight-for-stature compared with body mass index-forage growth charts for the United States from the Centers for Disease Control and Prevention. Am J Clin Nutr 75:761–766
- 15. Himes JH, Dietz WH (1994) Guidelines for overweight in adolescent preventive services: recommendations from an expert committee. The Expert Committee on Clinical Guidelines for Overweight in Adolescent Preventive Services. Am J Clin Nutr 59:307–316
- Flegal KM, Troiano RP (2000) Changes in the distribution of body mass index of adults and children in the US population. Int J Obes Relat Metab Disord 24:807–818
- 17. Friedman JM (2004) Modern science versus the stigma of obesity. Nat Med 10:563-569
- Lyon HN, Hirschhorn JN (2005) Genetics of common forms of obesity: a brief overview. Am J Clin Nutr 82:215S–217S
- Hebebrand J, Friedel S, Schauble N, Geller F, Hinney A (2003) Perspectives: molecular genetic research in human obesity. Obes Rev 4:139–146
- Farooqi IS, O'Rahilly S (2005) New advances in the genetics of early onset obesity. Int J Obes (Lond) 29(1149–52)

5 Genetics of Childhood Obesity

- 21. Bell CG, Walley AJ, Froguel P (2005) The genetics of human obesity. Nat Rev Genet 6:221–234
- 22. Stunkard AJ et al (1986) An adoption study of human obesity. N Engl J Med 314:193-198
- Stunkard AJ, Harris JR, Pedersen NL, McClearn GE (1990) The body-mass index of twins who have been reared apart. N Engl J Med 322:1483–1487
- 24. Knowler WC, Pettitt DJ, Saad MF, Bennett PH (1990) Diabetes mellitus in the Pima Indians: incidence, risk factors and pathogenesis. Diabetes Metab Rev 6:1–27
- Kondo I, Hamabe J, Yamamoto K, Niikawa N (1990) Exclusion mapping of the Cohen syndrome gene from the Prader-Willi syndrome locus. Clin Genet 38:422–426
- 26. Russell-Eggitt IM et al (1998) Alstrom syndrome. Report of 22 cases and literature review. Ophthalmology 105:1274–1280
- 27. Beales PL, Warner AM, Hitman GA, Thakker R, Flinter FA (1997) Bardet-Biedl syndrome: a molecular and phenotypic study of 18 families. J Med Genet 34:92–98
- 28. Bruford EA et al (1997) Linkage mapping in 29 Bardet-Biedl syndrome families confirms loci in chromosomal regions 11q13, 15q22.3-q23, and 16q21. Genomics 41:93–99
- Young TL et al (1999) A fifth locus for Bardet-Biedl syndrome maps to chromosome 2q31. Am J Hum Genet 64:900–904
- Ingalls AM, Dickie MM, Snell GD (1996) Obese, a new mutation in the house mouse. Obes Res 4:101
- Ingalls AM, Dickie MM, Snell GD (1950) Obese, a new mutation in the house mouse. J Hered 41:317–318
- Halaas JL et al (1995) Weight-reducing effects of the plasma protein encoded by the obese gene. Science 269:543–546
- Zhang Y et al (1994) Positional cloning of the mouse obese gene and its human homologue. Nature 372:425–432
- 34. Chua SC Jr et al (1996) Phenotypes of mouse diabetes and rat fatty due to mutations in the OB (leptin) receptor. Science 271:994–996
- 35. Considine RV et al (1996) Serum immunoreactive-leptin concentrations in normal-weight and obese humans. N Engl J Med 334:292–295
- Montague CT et al (1997) Congenital leptin deficiency is associated with severe early-onset obesity in humans. Nature 387:903–908
- Echwald SM et al (1997) Identification of two novel missense mutations in the human OB gene. Int J Obes Relat Metab Disord 21:321–326
- Oksanen L et al (1997) Novel polymorphism of the human ob gene promoter in lean and morbidly obese subjects. Int J Obes Relat Metab Disord 21:489–494
- Masuo K et al (2008) Leptin-receptor polymorphisms relate to obesity through blunted leptin-mediated sympathetic nerve activation in a Caucasian male population. Hypertens Res 31:1093–1100
- 40. Clement K et al (1998) A mutation in the human leptin receptor gene causes obesity and pituitary dysfunction. Nature 392:398–401
- 41. Krude H et al (2003) Obesity due to proopiomelanocortin deficiency: three new cases and treatment trials with thyroid hormone and ACTH4-10. J Clin Endocrinol Metab 88:4633–4640
- 42. Flickinger TW, Salz HK (1994) The Drosophila sex determination gene snf encodes a nuclear protein with sequence and functional similarity to the mammalian U1A snRNP protein. Genes Dev 8:914–925
- 43. Krude H et al (1998) Severe early-onset obesity, adrenal insufficiency and red hair pigmentation caused by POMC mutations in humans. Nat Genet 19:155–157
- 44. Challis BG et al (2002) A missense mutation disrupting a dibasic prohormone processing site in pro-opiomelanocortin (POMC) increases susceptibility to early-onset obesity through a novel molecular mechanism. Hum Mol Genet 11:1997–2004
- 45. Jackson RS et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 16:303–306
- 46. Farooqi IS et al (2007) Hyperphagia and early-onset obesity due to a novel homozygous missense mutation in prohormone convertase 1/3. J Clin Endocrinol Metab 92:3369–3373

- Benzinou M et al (2008) Common nonsynonymous variants in PCSK1 confer risk of obesity. Nat Genet 40:943–945
- Farooqi IS et al (2000) Dominant and recessive inheritance of morbid obesity associated with melanocortin 4 receptor deficiency. J Clin Invest 106:271–279
- 49. Farooqi IS et al (2003) Clinical spectrum of obesity and mutations in the melanocortin 4 receptor gene. N Engl J Med 348:1085–1095
- 50. Krakoff J et al (2008) Lower metabolic rate in individuals heterozygous for either a frameshift or a functional missense MC4R variant. Diabetes 57:3267–3272
- 51. Malakooti J et al (1999) Molecular cloning, tissue distribution, and functional expression of the human Na(+)/H(+) exchanger NHE2. Am J Physiol 277:G383–G390
- 52. Yeo GS et al (2004) A de novo mutation affecting human TrkB associated with severe obesity and developmental delay. Nat Neurosci 7:1187–1189
- Holder JL Jr, Butte NF, Zinn AR (2000) Profound obesity associated with a balanced translocation that disrupts the SIM1 gene. Hum Mol Genet 9:101–108
- Hirschhorn JN, Daly MJ (2005) Genome-wide association studies for common diseases and complex traits. Nat Rev Genet 6:95–108
- Carlson CS, Eberle MA, Kruglyak L, Nickerson DA (2004) Mapping complex disease loci in whole-genome association studies. Nature 429:446–452
- 56. The International HapMap Consortium (2005) A haplotype map of the human genome. Nature 437:1299–1320
- 57. Reich D et al (2005) A whole-genome admixture scan finds a candidate locus for multiple sclerosis susceptibility. Nat Genet 37:1113–1118
- Steemers FJ et al (2006) Whole-genome genotyping with the single-base extension assay. Nat Methods 3:31–33
- 59. The International HapMap Consortium (2003) The International HapMap Project. Nature 426:789–796
- Manolio TA, Collins FS (2009) The HapMap and genome-wide association studies in diagnosis and therapy. Annu Rev Med 60:443–456
- Herbert A et al (2006) A common genetic variant is associated with adult and childhood obesity. Science 312:279–283
- 62. Loos RJ, Barroso I, O'Rahilly S, Wareham NJ (2007) Comment on "A common genetic variant is associated with adult and childhood obesity". Science 315:187, author reply 187
- 63. Dina C et al (2007) Comment on "A common genetic variant is associated with adult and childhood obesity". Science 315:187, author reply 187
- 64. Rosskopf D et al (2007) Comment on "A common genetic variant is associated with adult and childhood obesity". Science 315:187, author reply 187
- 65. Lyon HN et al (2007) The association of a SNP upstream of INSIG2 with body mass index is reproduced in several but not all cohorts. PLoS Genet 3:e61
- Hotta K et al (2008) INSIG2 gene rs7566605 polymorphism is associated with severe obesity in Japanese. J Hum Genet 53:857–862
- 67. Frayling TM et al (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316:889–894
- 68. Hinney A et al (2007) Genome Wide Association (GWA) study for early onset extreme obesity supports the role of fat mass and obesity associated gene (FTO) variants. PLoS One 2:e1361
- 69. Dina C et al (2007) Variation in FTO contributes to childhood obesity and severe adult obesity. Nat Genet 39:724–726
- 70. Scuteri A et al (2007) Genome-Wide Association scan shows genetic variants in the FTO gene are associated with obesity-related traits. PLoS Genet 3:e115
- 71. Fawcett KA, Barroso I (2010) The genetics of obesity: FTO leads the way. Trends Genet 26:266–274
- 72. Grant SF et al (2008) Association analysis of the FTO gene with obesity in children of Caucasian and African ancestry reveals a common tagging SNP. PLoS One 3:e1746

5 Genetics of Childhood Obesity

- 73. Wellcome Trust Case Control Consortium (2007) Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature 447:661–678
- 74. Zeggini E et al (2007) Replication of genome-wide association signals in UK samples reveals risk loci for type 2 diabetes. Science 316:1336–1341
- 75. Fox CS et al (2012) Genome-wide association for abdominal subcutaneous and visceral adipose reveals a novel locus for visceral fat in women. PLoS Genet 8:e1002695
- 76. Gerken T et al (2007) The obesity-associated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. Science 318:1469–1472
- 77. Lein ES et al (2007) Genome-wide atlas of gene expression in the adult mouse brain. Nature 445:168–176
- Cecil JE, Tavendale R, Watt P, Hetherington MM, Palmer CN (2008) An obesity-associated FTO gene variant and increased energy intake in children. N Engl J Med 359:2558–2566
- Church C et al (2010) Overexpression of Fto leads to increased food intake and results in obesity. Nat Genet 42:1086–1092
- 80. Fischer J et al (2009) Inactivation of the Fto gene protects from obesity. Nature 458: 894–898
- Meyre D et al (2010) Prevalence of loss-of-function FTO mutations in lean and obese individuals. Diabetes 59:311–318
- Deliard S et al (2013) The missense variation landscape of FTO, MC4R, and TMEM18 in obese children of African Ancestry. Obesity (Silver Spring) 21(159–63)
- Stephens M, Smith NJ, Donnelly P (2001) A new statistical method for haplotype reconstruction from population data. Am J Hum Genet 68:978–989
- Loos RJ et al (2008) Common variants near MC4R are associated with fat mass, weight and risk of obesity. Nat Genet 40:768–775
- 85. Willer CJ et al (2009) Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet 41:25–34
- 86. Thorleifsson G et al (2009) Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. Nat Genet 41:18–24
- Gunstad J et al (2006) BDNF Val66Met polymorphism is associated with body mass index in healthy adults. Neuropsychobiology 53:153–156
- Speliotes EK et al (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42:937–948
- Berndt SI et al (2013) Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. Nat Genet 45:501–512
- Barker DJ (2012) Sir Richard Doll lecture. Developmental origins of chronic disease. Public Health 126:185–189
- Gluckman PD, Hanson MA, Cooper C, Thornburg KL (2008) Effect of in utero and early-life conditions on adult health and disease. N Engl J Med 359:61–73
- 92. Zhao J et al (2009) The role of obesity-associated loci identified in genome-wide association studies in the determination of pediatric BMI. Obesity (Silver Spring) 17(2254–2257)
- Zhao J et al (2011) Role of BMI-associated loci identified in GWAS meta-analyses in the context of common childhood obesity in European Americans. Obesity (Silver Spring) 19(2436–9)
- 94. Zhao J et al (2010) Examination of all type 2 diabetes GWAS loci reveals HHEX-IDE as a locus influencing pediatric BMI. Diabetes 59:751–755
- 95. Styrkarsdottir U et al (2009) New sequence variants associated with bone mineral density. Nat Genet 41:15–17
- 96. Styrkarsdottir U et al (2008) Multiple genetic loci for bone mineral density and fractures. N Engl J Med 358:2355–2365
- 97. Richards JB et al (2008) Bone mineral density, osteoporosis, and osteoporotic fractures: a genome-wide association study. Lancet 371:1505–1512
- Estrada K et al (2012) Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture. Nat Genet 44:491–501

- Richards JB, Zheng HF, Spector TD (2012) Genetics of osteoporosis from genome-wide association studies: advances and challenges. Nat Rev Genet 13:576–588
- 100. Zhao J et al (2011) BMD-associated variation at the Osterix locus is correlated with childhood obesity in females. Obesity (Silver Spring) 19(1311–4)
- 101. Hardy R et al (2010) Life course variations in the associations between FTO and MC4R gene variants and body size. Hum Mol Genet 19:545–552
- 102. Silverwood RJ, De Stavola BL, Cole TJ, Leon DA (2009) BMI peak in infancy as a predictor for later BMI in the Uppsala Family Study. Int J Obes (Lond) 33(929–37)
- 103. Wen X, Kleinman K, Gillman MW, Rifas-Shiman SL, Taveras EM (2012) Childhood body mass index trajectories: modeling, characterizing, pairwise correlations and sociodemographic predictors of trajectory characteristics. BMC Med Res Methodol 12:38
- 104. Rolland-Cachera MF et al (1984) Adiposity rebound in children: a simple indicator for predicting obesity. Am J Clin Nutr 39:129–135
- 105. Rolland-Cachera MF, Deheeger M, Maillot M, Bellisle F (2006) Early adiposity rebound: causes and consequences for obesity in children and adults. Int J Obes (Lond) 30(Suppl 4): S11–S17
- 106. Barker DJ, Osmond C, Forsen TJ, Kajantie E, Eriksson JG (2005) Trajectories of growth among children who have coronary events as adults. N Engl J Med 353:1802–1809
- 107. Sovio U et al (2011) Association between common variation at the FTO locus and changes in body mass index from infancy to late childhood: the complex nature of genetic association through growth and development. PLoS Genet 7:e1001307
- 108. Scherag A et al (2010) Two new Loci for body-weight regulation identified in a joint analysis of genome-wide association studies for early-onset extreme obesity in French and german study groups. PLoS Genet 6:e1000916
- Bradfield JP et al (2012) A genome-wide association meta-analysis identifies new childhood obesity loci. Nat Genet 44:526–531
- 110. Liu W et al (2010) Olfactomedin 4 down-regulates innate immunity against Helicobacter pylori infection. Proc Natl Acad Sci U S A 107:11056–11061
- 111. Fu M, Lui VC, Sham MH, Cheung AN, Tam PK (2003) HOXB5 expression is spatially and temporarily regulated in human embryonic gut during neural crest cell colonization and differentiation of enteric neuroblasts. Dev Dyn 228:1–10
- 112. Dankel SN et al (2010) Switch from stress response to homeobox transcription factors in adipose tissue after profound fat loss. PLoS One 5:e11033
- 113. Wheeler E et al (2013) Genome-wide SNP and CNV analysis identifies common and low-frequency variants associated with severe early-onset obesity. Nat Genet 45:513–517
- 114. Grant SF et al (2006) Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of type 2 diabetes. Nat Genet 38:320–323
- 115. Helgason A et al (2007) Refining the impact of TCF7L2 gene variants on type 2 diabetes and adaptive evolution. Nat Genet 39:218–225
- 116. Adeyemo A et al (2010) FTO genetic variation and association with obesity in West Africans and African Americans. Diabetes 59:1549–1554
- 117. Hassanein MT et al (2010) Fine mapping of the association with obesity at the FTO locus in African-derived populations. Hum Mol Genet 19:2907–2916
- 118. Monda KL et al (2013) A meta-analysis identifies new loci associated with body mass index in individuals of African ancestry. Nat Genet 45:690–696
- 119. Bogardus C (2009) Missing heritability and GWAS utility. Obesity (Silver Spring) 17(209–10)
- 120. Manolio TA et al (2009) Finding the missing heritability of complex diseases. Nature 461:747–753
- 121. Walley AJ, Asher JE, Froguel P (2009) The genetic contribution to non-syndromic human obesity. Nat Rev Genet 10:431–442
- 122. Zuk O, Hechter E, Sunyaev SR, Lander ES (2012) The mystery of missing heritability: genetic interactions create phantom heritability. Proc Natl Acad Sci U S A 109:1193–1198
- 123. Blakemore AI et al (2009) A rare variant in the visfatin gene (NAMPT/PBEF1) is associated with protection from obesity. Obesity (Silver Spring) 17(1549–53)

- 124. Redon R et al (2006) Global variation in copy number in the human genome. Nature 444: 444–454
- 125. Gunderson KL, Steemers FJ, Lee G, Mendoza LG, Chee MS (2005) A genome-wide scalable SNP genotyping assay using microarray technology. Nat Genet 37:549–554
- Bochukova EG et al (2010) Large, rare chromosomal deletions associated with severe earlyonset obesity. Nature 463:666–670
- 127. Walters RG et al (2010) A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. Nature 463:671–675
- 128. Jacquemont S et al (2011) Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. Nature 478:97–102
- 129. Glessner JT et al (2010) A genome-wide study reveals copy number variants exclusive to childhood obesity cases. Am J Hum Genet 87:661–666
- 130. Richards JB et al (2009) A genome-wide association study reveals variants in ARL15 that influence adiponectin levels. PLoS Genet 5:e1000768
- 131. Jarick I et al (2011) Novel common copy number variation for early onset extreme obesity on chromosome 11q11 identified by a genome-wide analysis. Hum Mol Genet 20:840–852
- 132. Leff SE et al (1992) Maternal imprinting of the mouse Snrpn gene and conserved linkage homology with the human Prader-Willi syndrome region. Nat Genet 2:259–264
- 133. Nicholls RD, Saitoh S, Horsthemke B (1998) Imprinting in Prader-Willi and Angelman syndromes. Trends Genet 14:194–200
- 134. Dong C et al (2005) Possible genomic imprinting of three human obesity-related genetic loci. Am J Hum Genet 76:427–437
- 135. Deliard S, Zhao J, Xia Q, Grant SF (2013) Generation of high quality chromatin immunoprecipitation DNA template for high-throughput sequencing (ChIP-seq). J Vis Exp 74

Chapter 6 Genetic Pleiotropies of Obesity

Bratati Kahali and Elizabeth K. Speliotes

Abstract Advances in the knowledge of variation in the human genome and reproducible results from the Genome-wide Association Studies (GWAS) have led to the identification of susceptible loci contributing to obesity and related disorders. Obesity is associated with and may contribute to the development of many metabolic diseases including, but not limited to, diabetes, hypertension, dyslipidemia, liver disease, and cardiovascular disease, often leading to morbidity and mortality; however the mechanisms linking the genetic polymorphisms associated with obesity to these metabolic complications are extremely complex and remains to be fully elucidated. In this chapter, we review a number of genetic perturbations that predispose to obesity as well as obesity-associated metabolic complications. Further understanding on how these variants act may help toward personalized treatment for obesity-related comorbidities based on individual needs.

Obesity Is a Worldwide Epidemic with Much Comorbidity

Obesity has become an epidemic that threatens the health of billions of people worldwide. Obesity is associated with and may contribute to the development of many metabolic diseases including, but not limited to, diabetes, hypertension, dyslipidemia, liver disease, cardiovascular disease, and cancer (Fig. 6.1). Worldwide, more than 700 million adults are obese and global projections estimate there will be 2.16 billion overweight and 1.12 billion obese individuals by 2030 [47]. According to

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Fig. 6.1 Obesity-related metabolic diseases

data released by the National Bureau of Economic Research in 2010, the annual health costs related to obesity in the USA reach \$168 billion, and nearly 17 % of the medical costs in the USA result from obesity. An obese person generates health costs of \$3,271 annually compared with \$512 for a nonobese person [11]. A better understanding of the causes of obesity and its related comorbidities is essential to mitigate the immense medical and economic impact of obesity worldwide.

Interestingly, obesity and related metabolic diseases are in part genetically influenced. Recent work into the genetic basis of these conditions is beginning to reveal a possible common genetic basis to obesity and related traits, sometimes in directions not predicted by epidemiological relationships. Here we discuss the genetic basis of some obesity related traits as they relate to and possibly contribute to associated metabolic diseases.

Types of Adiposity and Their Measures

Adiposity can increase throughout the whole body or in particular depots. Specialized methods to measure body-fat percentage, like dual energy X-ray absorptiometry (DXA—formerly DEXA), and air displacement plethysmography (ADP) are specific but expensive and cumbersome to implement for large groups. Body mass index (BMI) is commonly used as an inexpensive, noninvasive measurement of adiposity, but it is less specific since it also includes lean body mass. Internationally, BMI is commonly used to classify adult obesity. The World Health Organization, considers individuals of European ancestry with a BMI \geq 30 kg/m² to be obese and those with a BMI \geq 40 kg/m² to be extremely obese. Interestingly, comorbidities can develop at lower BMIs in some people of non-European ancestry; individuals of Indian, Chinese, Korean, and Japanese ancestry have a higher prevalence than those of Caucasian ancestry, of diabetes and other metabolic conditions with similar BMI and waist circumference measurements [41]. One possible explanation for this disparity is fat distribution. Visceral and hepatic fat deposition is more strongly

associated with the presence of metabolic disease than generalized adiposity [41]. Individuals of Asian ancestry then may have a greater proportion of visceral fat deposition than individuals of European ancestry [41]. One pathophysiological explanation for how increased visceral fat accumulation may lead to an increased risk for type 2 diabetes is the production by visceral fat of cytokines that promote insulin resistance [8]. Noninvasive, inexpensive measures of visceral adiposity include waist circumference (WC) and waist-to-hip ratio (WHR) but these are not as specific for measuring this and other specific depots as using imaging modalities such as magnetic resonance imaging, computed tomography, or ultrasound.

Obesity Is Associated with and May Contribute to the Development of Other Diseases

Obesity is excess body fat accumulation caused by a chronic positive energy balance (energy intake exceeding energy expenditure) due to deregulation in the complex process of energy homeostasis [73]. Unfortunately, the increased prevalence of obesity worldwide has led to an increase in the prevalence of diabetes, hypertension, dyslipidemia, coronary artery disease, nonalcoholic fatty liver disease, and some forms of cancer [53]. In individuals of Caucasian ancestry with a BMI above 29 versus below this cutoff, there is a greater risk of developing these diseases, independent of gender [63, 112, 113]. Increased waist circumference is a measure of abdominal obesity. Men and women that have a waist circumference greater than or equal to 102 cm and 88 cm respectively are at substantially increased risk for metabolic complications like coronary heart disease, hypertension, and dyslipidemia [32] compared to those with a waist circumference less than these values. Increased adiposity is associated with elevated fasting plasma insulin and an exaggerated insulin response to an oral glucose load, and may contribute to the development of type 2 diabetes [52]. In the Nurses Cohort Study of 116,000 women, the risk of diabetes increased fivefold for women with a BMI of 25, 28-fold for women with a BMI of 30, and 93-fold for women with a BMI of 35 or higher, compared with women with a BMI of less than 21, even after adjustment for age [16]. In the Health Professionals Study of 51,529 men, the risk of diabetes, adjusted for age, is increased 2.2-fold for men with a BMI between 25 and 26.9, 6.7-fold for men with a BMI between 29 and 30, and 42-fold for men with a BMI of 35 or higher, compared with men with a BMI of less than 21 [13]. Obesity is also a primary risk factor for nonalcoholic fatty liver disease (NAFLD), but not all obese individuals are affected [21]. NAFLD includes a spectrum of disease in individuals without a history of excessive alcohol ingestion: from fatty infiltration of the liver (steatosis), to histological evidence of inflammation (nonalcoholic steatohepatitis, NASH), to fibrosis or cirrhosis [15]. NAFLD can lead to liver failure and is accompanied by substantial morbidity and mortality, with few known effective treatments [33]. Dyslipoproteinemia (abnormal blood lipid concentrations) with decreased HDL cholesterol, increased LDL triglyceride, and increased LDL concentrations is associated with increased adiposity (reviewed by Kopelman [53]).

Genetic Basis of Obesity

Although decreased physical activity and excess calorie intake may partially explain the increased prevalence of obesity in modern times, some individuals are more susceptible to these lifestyle changes than are others, suggesting that they may have a genetic predisposition to obesity [75]. Indeed, results from twin studies suggest that genetic factors explain 50-90 % of the variance in BMI [62]. Individual variation in waist-to-hip ratio is heritable, even after accounting for BMI, with heritability estimates ranging from 22–61 % [36]. Hepatic adiposity is also 26–27 % heritable [98]. In the last decade much has been learned about the genetic basis of adiposity. Specific genetic variants have been associated with measures of human obesity. These include rare chromosomal aberrations and gene defects that have a large effect on afflicted individuals, as well as common variants that have smaller effects but influence the overall distribution of obesity in the population [96]. Some obesity conditions are caused by large chromosomal aberrations; whereas in many obesity conditions the causal variant has been narrowed down to particular intervals in the genome. This work suggests that there is an allelic spectrum of genetic variants that contribute to human obesity.

Obesity Syndromes with Developmental Delay or Mental Retardation

Over 30 pleiotropic syndromes have been reported that include obesity and developmental delay or mental retardation as components (Refer to other chapter in book). The genetic basis for some of these has been identified. A large chromosomal aberration involving multiple genes is sometimes the cause. One example is Prader Willi syndrome which causes obesity together with hyperphagia, hypotonia, mental retardation, learning disabilities, short stature, and hypogonadotropic hypogonadism. Complications co-occurring with the syndrome include abnormal glucose tolerance, type 2 diabetes, right-sided heart failure, and bone problems. Prader–Willi syndrome is a contiguous gene syndrome resulting from deletion of paternal copies of the imprinted *SNRPN* gene, the *NDN* (necdin) gene, and possibly other missing genes on part of chromosome 15 (15q11-13) (OMIM); which phenotypes are due to disturbance of which genes remains to be determined.

For other syndromes, such as Bardet–Biedl syndrome (where there are 15 BBS genes and 3 modifier genes identified), we have a better understanding of how disruption in the functioning of one of these genes in the set results in pleiotropic phenotypes. BBS is characterized by retinal degeneration, obesity, polydactyly, and hypogonadism, as well as developmental delay, speech disorder, anosmia, ataxia/imbalance, behavioral problems, polycystic kidneys, hearing loss, congenital heart defects, situs inversus, and Hirschsprung disease ([105]; a clinical synopsis available in OMIM id 209900). Afflicted individuals are at increased risk for diabetes mellitus,

hypertension, and congenital heart disease [70, 71]. BBS genes are involved in basal body and centrosome function, relating to ciliary development and function in many tissues. Defects in ciliary processes in many tissues are thought to underlie the pleiotropic phenotypes observed in this syndrome [69, 72]. For example, dysfunction of BBS genes in preadipocytes and the hypothalamus may contribute to the obesity seen in BBS [64]. Human preadipocytes form a transient primary cilium during differentiation. BBS10 and BBS12 localize to the basal body of this primary cilium and knockdown of BBS10 and BBS12 expression reduces the number of ciliated cells and promotes adipogenesis. Furthermore, differentiation of BBS10 and BBS12 patients' fibroblasts into fat-accumulating cells contain increased triglycerides compared with control cells [64], supporting the assertion that primary dysfunction of adipogenesis may contribute to the obesity seen in BBS. A second mechanism by which BBS genes contribute to obesity may involve altered leptin receptor (LEPR) signaling in the hypothalamus leading to deregulation of energy balance. Bbs2 -/-, Bbs4 -/-, and *Bbs6* –/– mice are resistant to the action of leptin to reduce body weight and food intake regardless of serum leptin levels and obesity [90]. Activation of hypothalamic Stat3 by leptin, but not downstream MC4R signaling, was significantly decreased in Bbs2 -/-, Bbs4 -/-, and Bbs6 -/- mice indicating that LEPR signaling was specifically impaired in these animals. The human BBS1 protein physically interacts with LEPR, and loss of BBS proteins perturbs LEPR trafficking and decreases proopiomelanocortin (POMC) expression in human cells, suggesting a mechanism by which some BBS proteins may mediate their effects. Some BBS proteins are also required for proper trafficking of G protein-coupled receptors (GPCRs) in and out of neuronal cilia, suggesting disrupted ciliary GPCR trafficking is the basis for the neurological defects in BBS [7]. Better understanding of ciliopathies like Bardet-Biedl syndrome allow the identification of signaling pathways potentially involved in common diseases that share phenotypic features like obesity.

Obesity Syndromes Without Developmental Delay or Mental Retardation

Obesity that segregates within families but that is not associated with severe developmental delay or mental retardation has been reported and characterized. Genes that are part of a hypothalamic axis for appetite regulation include leptin, leptin receptor, *POMC*, and *MC4R*. Defects in leptin in humans lead to severe early-onset obesity and intense hyperphagia, as well as hypogonadotropic hypogonadism [77]. *POMC* mutations are characterized by hyperphagia, and impaired melanocortin signaling in the hypothalamus with early-onset obesity [54]. Heterozygous mutations in *MC4R* have been reported to cause a dominantly inherited obesity in various ethnic groups. Mutations in *MC4R* are by far the most common forms of monogenic obesity in humans. Defects in *MC4R* have been reported in multiple families, and associated phenotypes include obesity, an increase in lean body mass and bone mineral density, an increase in linear growth throughout childhood, hyperphagia, and severe hyperinsulinemia; homozygotes are more severely affected than heterozygotes [22]. MC4R is expressed in human and rodent brain and targeted disruption of Mc4r in rodents leads to increased food intake, obesity, severe early hyperinsulinemia, and increased linear growth; heterozygotes have an intermediate phenotype between homozygous mutant and wild-type mice [43]. The results of MC4R disruption are similar in humans, suggesting that aspects of the MC4R pathway may be conserved between rodents and humans. These genes may have a common effect on appetite because they act on the central leptin-melanocortin axis in the hypothalamus, thus playing a critical role in the regulation of feeding behavior. However, individuals with defects in leptin, leptin receptor, POMC, and MC4R do not display the same set of related pleiotropisms. For example, individuals with defects in MC4R show increased lean body mass, hyperinsulinemia, and increased linear growth, which are not seen in individuals with the defects in the other genes [74]. Individuals with defects in leptin/leptin receptor have hypogonadotropic hypogonadism and central hypothyroidism, which are not seen with defects in the other genes [74]. Adrenal crisis and pale skin are seen in individuals with POMC defects [54] but not with defects in the other genes [74]. The basis for gene unique phenotypes may be due to particular genes having effects in other neurons/cells outside of the feeding pathways or to them having effects via downstream mediators that are not part of the core feeding circuit. For example, MC4R is expressed in many neurons in the hypothalamus in addition to those that control feeding [92]. It is therefore probable that its effects on non-obesity phenotypes are mediated through neurons in non-feeding pathways. Alternatively, POMC undergoes extensive and tissue-specific posttranslational processing by prohormone convertases (PCs) to yield a range of biologically active peptides [17]. Specifically, POMC is cleaved by PC1 and PC2 to make α -, β -, and γ -MSH (the melanocortins) and ACTH respectively. The expression of PC2 within the hypothalamus leads to the production of α -, β -, and γ -MSH (the melanocortins), which mediate the effect of POMC on feeding. In contrast, adrenocorticotropic hormone (ACTH) is the predominant peptide produced from POMC in pituitary corticotrophs that express PC1 and mediate its effect on the adrenal axis. Therefore, expression of the genes themselves or of the downstream mediators of their effects in particular cell types may help to explain the diversity of pleiotropisms observed when these genes are disrupted.

Common Variants Affect Adiposity and Related Metabolic Traits in Human Populations

BMI, WC, WHR, and Obesity-Associated Variants

Genome-wide association studies (GWAS) have identified genetic variants that associate with measures of overall obesity (e.g., BMI) that also have effects on related metabolic traits. Interestingly, some of these common variants fall in or near genes

GWAS trait	Variant	Nearby Gene	HDL-	LDL-	Triglyce	Fasting	2-hour	Fasting	HOMA-IR	Type 2	NASH	Neuropsychiatric
	identified		cholesterol	cholesterol	rides	Glucose	Glucose	Insulin		Diabetes		traits***
BMI	rs2229616,	MC4R							•			
	rs129/0134		•						T.			
DMI	rs1//82313**	ETO										
DIVII	181556902	FIO						1	1	Τ		
BMI	rs7359397	SH2B1				1		1				[@] 1
BMI	rs10767664*	BDNF										4
WC	rs10146997	NRXN3										1
BMI	rs2287019	GIPR				1						
BMI	rs6232	PCSK1				1	•		4			
WHR	rs10195252	GRB14		1	1			1	1			
WHR	rs2605100	LYPLAL1			1				#			
Body Fat Percentage	rs2943650	IRSI	1		•					. ↑		
VAT (Women only)	rs1659258	THNSL2	¥			1						
Liver steatosis	rs780094	GCKR		1	1				4		1	
Liver steatosis	rs738409	PNPLA3									1	
Liver steatosis	rs12137855	LYPLAL1									1 A	
Liver steatosis	rs4240624^	PPP1R3B	•	•		1			1			
Liver steatosis	rs2228603	NCAN		•	•						1	

Fig. 6.2 The associations with other metabolic traits for the trait-increasing allele for various measures of obesity

[^]Variant associates with CT hepatic steatosis but not histologic fibrosis (Speliotes et al., PLoS Genetics, 2011)

*Variant is in r2 = 0.76 with rs6265 which encodes a Val66Met missense change in BDNF. A strong association of the BDNF 196G/A (Val66Met) polymorphism has been demonstrated with the restricting type of anorexia nervosa and binge-eating/purging type of bulimia nervosa (Ribases et al., Mol Psychiatry, 2003; Koizumi et al., Am J Med Genet, 2004).

** rs17782313 identified as in a large-scale GWAS as susceptible to fat mass, overweight and risk of obesity in European population, these authors suggest that rs17782313 influence BMI independently from rs2229616 V103I polymorphism (Loos et al., Nature Genetics, 2008); rs2229616 encodes the V103I missense change in MC4R (Heid et al., Obesity, 2008). The association of rs17782313 with insulin resistance was assessed in a recent study population based study (Povel et al., Cardiovasc Diabetol, 2012). rs12970134 (r2 = 0.81 with rs17782313) associated significantly with waist circumference and insulin resistance in Indian Asians and Europeans (Chambers et al., Nature Genetics, 2008).

*** Implies Anorexia nervosa and Bulimia nervosa for the variant near BDNF and Impulsivity for the variant near NRXN3.

[@] Aggressive behavior for SH2B1 loss-of-function mutations.

[#]rs4846567 G allele carriers showed a 5.2% lower HOMA-IR in women, this SNP is in moderate LD (r2 = 0.64) with the WHR SNP.

that have been previously identified to play a role in obesity because when they are deregulated in humans or model organisms, they result in obesity. In particular, common variants have been identified in or near *MC4R* and *POMC* that affect population-based BMI. As noted above, severe perturbation of these genes results in monogenic obesity syndromes. Indeed, for lipid and height traits when monogenic disease genes that affect these traits are present near a GWAS variant that affect these same traits, the monogenic disease gene is the nearest gene two-thirds of the time [55, 102]. This suggests that examining genes near GWAS signals may give us insights into genes that may affect the biology of the trait of interest. Interestingly, common variants identified by large-scale GWAS near *MC4R* [60] and *POMC* [96,

97] that associate with higher BMI also associate with higher and lower height, respectively, consistent with observations in individuals with monogenic mutations affecting these genes. Thus, GWAS-associated variants can provide direct insights into the genetic basis of obesity pleiotropisms. For example, variants near *MC4R* have been found to associate with increased risk of metabolic syndrome and increased insulin resistance in human populations of diverse ancestry [12, 37, 78] (Fig. 6.2).

The first common variants identified by GWAS to associate with overall fat measures were located in the first intron of *FTO* (fat mass and obesity-associated gene). Variants in high linkage disequilibrium with each other were associated with BMI, severe childhood and adult obesity, as well as fat mass: rs9939609 [111]; rs9930506 [89]; rs1421085 [66]; rs8050136 [49]. The BMI-increasing allele (rs9939609) at this locus is strongly associated with increased risk for type 2 diabetes [26]. Further, the BMI-increasing alleles at the FTO locus are also significantly associated with increased fasting insulin, increased homeostatic measure for insulin resistance (HOMA-IR) [96, 97], elevated diastolic blood pressure [104], and increased risk for metabolic syndrome [93] (Fig. 6.2). *FTO* affects adiposity; ubiquitous overexpression of *FTO* leads to a dose-dependent increase in body and fat mass in mice fed either a standard or high-fat diet [14]. FTO has a potential role in nucleic acid demethylation and is highly expressed in parts of the brain that govern energy balance and feeding behavior [27, 28], which make it a likely candidate through which these variants exert their effects on BMI.

The common variant rs7359397 near SH2B1 (Src homology 2 [SH2] domaincontaining putative adaptor protein B1) not only associates with population-measured BMI but also associates with increased fasting levels of insulin and plasma glucose [96, 97] (Fig. 6.2). This variant is in perfect linkage disequilibrium with missense variant rs7498665 in SH2B1, [111] which affects amino acid polarity (Thr484Ala) and falls into a highly conserved protein segment of SH2B1 containing a class II SH3 domain-binding site. Carriers of a large, rare chromosomal deletion of SH2B1 exhibit hyperphagia and severe insulin resistance disproportionate to the degree of obesity [9, 107]. Doche et al. [19] also identified SH2B1 loss-of-function mutations in a large cohort of 300 patients with severe early-onset obesity. Mutation carriers exhibited hyperphagia, childhood-onset obesity, disproportionate insulin resistance, and reduced final height as adults. Behavioral abnormalities, including social isolation and aggression, were also reported. SH2B1 modulates signaling by a variety of ligands that bind to receptor tyrosine kinases or JAK-associated cytokine receptors, including leptin, insulin, growth hormone, and nerve growth factor [85]. In mice, targeted deletion of SH2B1 results in increased food intake, obesity, and insulin resistance; heterozygous null mice fed a high-fat diet have an intermediate phenotype. SH2B1 can act as an insulin sensitizer because it directly binds to insulin receptor substrates IRS1 and IRS2, and enhances insulin sensitivity by promoting insulin receptor catalytic activity and inhibiting tyrosine dephosphorylation of IRS protein [56, 68]. Interestingly, the expression of only neuronal SH2B1 in these knockout mice rescues some of the phenotypes seen in knockout mice and this suggest that SH2B1 regulates energy balance, body weight, peripheral insulin sensitivity, and glucose homeostasis at least in part by enhancing hypothalamic leptin sensitivity [81]. These data suggest that *SH2B1* plays a critical role in the control of body weight, food intake, leptin-insulin signaling, and a putative role in maladaptive human behavior.

The lead SNP (rs10767664) identified near BDNF in a GWAS for BMI-susceptible loci in 249,796 individuals of European ancestry [96, 97] is in high linkage disequilibrium $(r^2=0.76)$ with rs6265, which results in a Val66Met missense change in BDNF. The BDNF 196G/A (Val66Met) polymorphism is strongly associated with the restricting type of anorexia nervosa and binge-eating/purging type of bulimia nervosa [51, 81] (Fig. 6.2), suggesting that the Met allele may be a susceptibility factor for eating disorders. Variants in or near BDNF, including rs6265 (Val66Met), that are associated with an increased BMI have also been associated with substancerelated disorders, alcohol dependence, and mood disorders [31]. These effects may be due to BDNF disruption causing interference with dopamine neurotransmission in pathways involved in reward effects, motivation, and decision making [31, 34, 96, 97, 103]. Further, heterozygous BDNF knockout mice exhibited aggressiveness and hyperphagia accompanied by significant weight gain in early adulthood [61], and abnormal locomotor activity and infusion with BDNF in these mice can transiently reverse the eating behavior and obesity [48]; supporting that altering BDNF function can mediate both obesity and psychiatric behavioral phenotypes.

The variant rs10146997 in neurexin-3-alpha (NRXN3) was significantly associated with waist circumference in a large-scale GWAS of a European population [35]. Although it is unlikely that a different gene accounts for this finding as there are no other genes within a distance of more than several hundred kilobases of this SNP, additional research is required to prove that rs10146997 acts through NRXN3 [35]. The rs760288-rs8019381-rs2293847 haplotype near NRXN3 associates with genderspecific alcohol dependence, cocaine addiction, and illegal substance abuse in 332 alcohol-dependent human participants [39]; however, these variants are in very low linkage disequilibrium with rs10146997. Weak association between rs10146697 and impulsivity in women has been reported [101] (Fig. 6.2). Therefore, although a direct link between variants that associate with obesity phenotypes and those that associate with substance abuse has not been shown or confirmed, these phenotypes may all result from perturbations of NRXN3. NRXN3, which belongs to a family of proteins that function as receptors and cell adhesion molecules in the nervous system, may play a role in the development and function of synapses [58]. How polymorphisms near NRXN3 confer vulnerability to addictions and obesity remains to be determined.

The BMI-increasing genome-wide significantly associated variant near the gastric inhibitory polypeptide receptor (*GIPR*, also known as the glucose-dependent insulinotropic polypeptide receptor) [96, 97] is in strong linkage disequilibrium (r^2 =0.83) with a missense SNP in *GIPR* (rs1800437, p.Glu354Gln) that has been shown to influence glucose and insulin responses to an oral glucose challenge [88]. The BMI-increasing allele is associated with increased fasting glucose levels and decreased 2-h glucose levels [96, 97] (Fig. 6.2). *GIPR* is widely distributed in peripheral organs, including the pancreas, gut, and adipose tissue, suggesting a role for peripheral biology in obesity. GIP, which is expressed in the K cells of the duodenum and intestine, is an incretin hormone that mediates insulin secretion in response to oral intake of glucose. Mice with disruption of *GIPR* are resistant to diet-induced obesity [67], further supporting a role of this pathway in affecting BMI. Interestingly, GIP and its receptor may constitute a link between the consumption of energy-rich high-fat diets and the development of obesity (reviewed by Irwin and Flatt [45] and McIntosh et al. [65]).

A GWAS of early-onset and morbid adult obesity in Europeans identified a coding missense variant in the Niemann–Pick C1 gene *NPC1* (rs1805081 [H215R]) that was genome-widely significantly associated with this trait. This variant had a population attributable risk for obesity of 9.6 % in children and 13.6 % in adults [66]. NPC1 is a protein involved in endosomal cholesterol trafficking in the central nervous system, liver, and macrophages [3, 44, 106]. *NPC1* mRNA was significantly increased in obese individuals in subcutaneous white adipose tissue (scWAT) and omental WAT (omWAT) and downregulated by weight loss, implicating *NPC1* in adipocyte biology [4]. Although rs1805081 has not been associated with the Niemann Pick C phenotype, *Npc1*-null mice exhibit late-onset weight loss and poor food intake, as well as neurological deficits and a cellular defect in cholesterol transport [20, 66, 108], suggesting that alteration of this gene results in pleiotropic effects.

The nonsynonymous variants rs6232[G] for PCSK1 (prohormone convertase (PC) 1/3), encoding N221D, and rs6234-rs6235[C], encoding the Q665E-S690T changes, have been consistently associated with obesity in adults and children $(P=7.27\times10^{-8} \text{ and } P=2.31\times10^{-12}, \text{ respectively})$ [6]. Rare mutations as well as common SNPs in PCSK1 cause childhood obesity and abnormal glucose homeostasis with elevated proinsulin concentrations in Europeans [6, 23, 46, 84]. A largescale GWAS in East Asians identified a common variant rs261967 near PCSK1 that associates with BMI [109]. Precursor polypeptides, such as *POMC*, proglucagon, and proinsulin, which are involved in the regulation of energy metabolism, serve as substrates for PCSK1 in various metabolic processes. The missense variant rs6235[G] coding for S690T in PCSK1 is significantly associated with fasting proinsulin levels on a genome-wide scale [101], consistent with the function of PCSK1 being the first enzymatic step in the insulin processing pathway. In addition, the obesity risk allele of SNP rs6232 (encoding missense variant N221D) was associated with reduced HOMA-IR, increased fasting glucose, and reduced 120-min glucose levels (Fig. 6.2), independently of BMI and proinsulin conversion in Europeans [38]. Functional analysis showed that the N221D-mutant PC1/3 protein has impaired catalytic activity [6]. Gjesing et al. [29] proposed that a direct or indirect consequence of having the C-allele of rs6235 for PCSK1 is an increased GIP level that might lead to increased levels of circulating insulin, reduce glucose levels, and possibly protect against diabetes. Thus, PCSK1 arbitrates its effect on obesity and related metabolic disorders through cleaving many precursor hormones in multiple tissues thus causing these pleiotropic effects in a parallel fashion.

The waist-to-hip ratio (WHR)-increasing allele at *GRB14* (rs10195252) shows a strong association with increased triglycerides ($P=7.4 \times 10^{-9}$), fasting insulin levels ($P=5.0 \times 10^{-6}$), and insulin resistance ($P=1.9 \times 10^{-6}$) [36] (Fig. 6.2). Growth factor receptor-bound protein 14 (GRB14) is a member of a family of SH2-containing adaptor proteins and binds directly to the insulin receptor [18, 40]. *GRB14* expression is increased in the adipose tissue of insulin-resistant animal models and type 2 diabetic human patients, suggesting the effect of *GRB14* is by modulating insulin sensitivity [10] to exert its effects.
A meta-analysis of genome-wide association data for central adiposity identified rs2605100 (near LYPLAL1—lysophospholipase-like protein 1) as an association in females only for WHR [57]. These authors also found an association between the WHR-increasing G-allele of rs2605100 and increased fasting triglycerides. Although the subcutaneous and visceral adipose tissue of 16 obese subjects had elevated mRNA expression of LYPLAL1 [99], the functional and molecular details of how rs2605100 actually exerts its effects to result in increased WHR effect remain to be determined.

Body Fat Percentage

Variant rs2943650 was identified to be significantly associated at genome-wide levels with body fat percentage in a recent meta-analysis study. rs2943650 is near IRS1 and shows an association with fat mass, greater in men than women [49]. The insulin receptor substrate protein IRS1 is a key target of the insulin receptor tyrosine kinase and is required for hormonal control of metabolism. Variants in high linkage disequilibrium with the fat mass-associated variant at *IRS1* also associate with type 2 diabetes [30]; cardiovascular disease [87]; and plasma lipid levels [102]; and is in perfect linkage disequilibrium with a variant (rs2943641) identified to be associated with insulin resistance and hyperinsulinemia in European population-based cohorts [86]. The C-allele for rs2943641 was also associated with reduced basal levels of IRS1 protein and decreased insulin induction of IRS1-associated phosphatidylinositol-3-OH kinase activity in human skeletal muscle biopsies. Interestingly, the fat percentage decreasing allele was associated with increased risk for type 2 diabetes and cardiovascular disease, and with an impaired lipid profile [1, 91, 102] (Fig. 6.2). Further analyses showed that the body fat percentage decreasing allele lowered the subcutaneous fat-but not the more harmful visceral fat-and also lowered adiponectin levels. This suggests that this variant may act by affecting fat distribution and in particular it may promote metabolic disease development by increasing visceral versus subcutaneous fat [49, 59].

Visceral Adipose Tissue (VAT)

A higher ratio of visceral to subcutaneous adipose tissue is known to be associated with a higher risk for cardiovascular and metabolic diseases [25, 50]. A genomewide association study of abdominal, subcutaneous, and visceral adipose tissue in individuals of European ancestry revealed a novel genome-wide significant variant rs1659258 near *THNSL2* (threonine synthase-like 2) that was significantly associated with visceral adipose tissue (VAT) only in women [24]. The VAT-increasing allele at *THNSL2* was associated with lower HDL and increased fasting glucose consistent (Fig. 6.2), and increased visceral adiposity promotes metabolic syndrome traits [24].

Liver Steatosis

A genome-wide association scan revealed a missense variant rs738409 (I148M) in PNPLA3 (Patatin-like phospholipase domain-containing protein 3) to be significantly associated with hepatic fat accumulation and hepatic inflammation in Hispanics, European Americans, and African Americans [82]. PNPLA3 plays a role in the hydrolysis of glycerolipids and the I148M substitution causes a loss of function [42]. The rs738409 (I148M) in PNPLA3 is also associated with severe histologic complications of NAFLD, such as portal inflammation, lobular inflammation, Mallory-Denk bodies, and fibrosis [83, 96, 97]. Genome-wide association scans for hepatic steatosis measured using computed tomography scanning or magnetic resonance spectroscopy has revealed that variants in or near PNPLA3, NCAN, GCKR, LYPLAL1, and PPP1R3B associate with this trait [98]. Variants in or near PNPLA3, NCAN, GCKR, and LYPLAL1 but not near PPP1R3B associate with NASH/fibrosis on a genome-wide significant level [98]. The hepatic steatosis-increasing allele at NCAN was associated with lower triglycerides and plasma LDL cholesterol levels [98] (Fig. 6.2) but how this variant exerts these effects is unclear. The nearest gene NCAN is a brain chondroitin sulfate proteoglycan that acts as an adhesion molecule [79] but it is not clear that the variant rs2228603 acts through this gene. The hepatic steatosis-increasing allele at GCKR (rs780094) was associated with higher levels of plasma LDL cholesterol and triglycerides, lower fasting glucose, lower HOMA-IR [98] (Fig. 6.2). GCKR inhibits glucokinase in the liver, pancreatic islet cells, and possibly other tissues and prevents the phosphorylation of glucose, which is required for its use as a substrate for de novo lipogenesis. Carriers of the common L446 allele for GCKR have higher triglyceride levels and lower fasting plasma glucose levels [76, 94]. The P446L coding change in the protein product of GCKR leads to a reduction in GCK-inhibition by the variant regulatory protein. This is predicted to increase glycolytic flux and hence glucose uptake by the liver. This enhanced rate of glycolysis may increase other liver metabolites such as malonyl-CoA, increasing triglyceride levels by two mechanisms-acting as a substrate for de novo lipogenesis, and also by blocking fatty acid oxidation [5]. This perturbation of hepatic metabolism may account for lowered glucose and raised triglycerides levels seen in L446carriers of GCKR. In this way it may possibly promote hepatic steatosis. Similarly, since PPP1R3B encodes a protein that promotes the breakdown of liver glycogen to phosphorylated glucose [2], variants near this gene may increase its activity and predispose carriers to increased hepatic steatosis by increasing substrates for de novo lipogenesis. Indeed, the hepatic steatosis associated variants (rs4240624) at this locus increase PPP1R3B expression in the liver which is consistent with this model [98]. How these hepatic steatosis variants at PPP1R3B also cause increased HDL and LDL cholesterol and decreased fasting glucose levels (Fig. 6.2) remains to be determined. Finally, variants associated with hepatic steatosis in or near PNPLA3 (rs738409) (encoded protein is a triacylglycerol lipase that mediates triacylglycerol hydrolysis in adipocytes) and LYPLAL1 (rs12137855) (lysophospholipase-like 1 protein) show association with NASH/fibrosis but do not show significant association for several lipid and glycemic traits tested. Thus, *PNPLA3* and *LYPLAL1* may be involved in lipid metabolism and exert their effects within the liver in ways that are not well reflected in serum glucose or lipid measurements [99]. These results were some of the first to show that genetics can dissociate epidemiologically correlated traits. Targeting the genes through which associated variants act may yield diverse outcomes, leading to personalized disease treatments.

Future Elucidation of More Obesity-Susceptible Loci Having Pleiotropic Association with Other Complex Diseases

Genome-wide association studies of many human phenotypes are beginning to uncover the unique and shared genetic basis of human traits. This information can complement studies of obesity syndromes to uncover the mechanisms by which genetic defects promote adiposity. Already we have learned that some obesity pleiotropisms may be caused by genetic defects that are affecting more than one gene, as occurs in Prader–Willi syndrome. Other pleiotropisms may arise from affected genes being expressed and acting in multiple tissues, as occurs with the BBS genes. A gene product may also have different effects in different tissues based on the expression of downstream mediators, as is seen for *POMC*; the expression of PC1 or PC2 determines how it is cleaved into different peptides with disparate effects, only some of which relate to the development of obesity. The mechanism by which many of the new GWAS-associated variants exert their pleiotropic effects remains to be determined.

Conclusion

With the increased prevalence of obesity worldwide, there has been a corresponding increase in many obesity-related diseases, with resultant morbidity and mortality, and for which there are few treatments and no cures. Genetic studies suggest that the development of obesity comorbidities may be influenced through different metabolic pathways that may represent distinct therapeutic targets. A better understanding of the unique and common genetic bases of these diseases may help us to improve personalized treatment based on individual needs. The integration of genetic data across phenotypes with data that include but are not limited to metabolomics, gene expression, or protein–protein interaction information can help us to better understand the mechanisms by which these genetic variants exert their effects and in this way help us to develop much needed new therapeutics for these conditions.

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References

- 1. Adeyemo A, Bentley AR, Meilleur KG, Doumatey AP, Chen G, Zhou J et al (2012) Transferability and fine mapping of genome-wide associated loci for lipids in African Americans. BMC Med Genet 13:88
- Agius L (2008) Glucokinase and molecular aspects of liver glycogen metabolism. Biochem J 414(1):1–18
- Amigo L, Mendoza H, Castro J, Quinones V, Miquel JF, Zanlungo S (2002) Relevance of Niemann-Pick type C1 protein expression in controlling plasma cholesterol and biliary lipid secretion in mice. Hepatology 36(4 Pt 1):819–828
- 4. Bambace C, Dahlman I, Arner P, Kulyte A (2013) NPC1 in human white adipose tissue and obesity. BMC Endocr Disord 13:5
- Beer NL, Tribble ND, McCulloch LJ, Roos C, Johnson PR, Orho-Melander M et al (2009) The P446L variant in GCKR associated with fasting plasma glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. Hum Mol Genet 18(21): 4081–4088
- Benzinou M, Creemers JW, Choquet H, Lobbens S, Dina C, Durand E et al (2008) Common nonsynonymous variants in PCSK1 confer risk of obesity. Nat Genet 40(8):943–945
- Berbari NF, Lewis JS, Bishop GA, Askwith CC, Mykytyn K (2008) Bardet-Biedl syndrome proteins are required for the localization of G protein-coupled receptors to primary cilia. Proc Natl Acad Sci U S A 105(11):4242–4246
- Bergman RN, Kim SP, Catalano KJ, Hsu IR, Chiu JD, Kabir M et al (2006) Why visceral fat is bad: mechanisms of the metabolic syndrome. Obesity (Silver Spring) 14(Suppl 1):16S–19S
- Bochukova EG, Huang N, Keogh J, Henning E, Purmann C, Blaszczyk K et al (2010) Large, rare chromosomal deletions associated with severe early-onset obesity. Nature 463(7281):666–670
- Cariou B, Capitaine N, Le Marcis V, Vega N, Bereziat V, Kergoat M et al (2004) Increased adipose tissue expression of Grb14 in several models of insulin resistance. FASEB J 18(9):965–967
- Cawley J, Meyerhoefer C (2012) The medical care costs of obesity: an instrumental variables approach. J Health Econ 31(1):219–230
- Chambers JC, Elliott P, Zabaneh D, Zhang W, Li Y, Froguel P et al (2008) Common genetic variation near MC4R is associated with waist circumference and insulin resistance. Nat Genet 40(6):716–718
- Chan JM, Rimm EB, Colditz GA, Stampfer MJ, Willett WC (1994) Obesity, fat distribution, and weight gain as risk factors for clinical diabetes in men. Diabetes Care 17(9):961–969
- Church C, Moir L, McMurray F, Girard C, Banks GT, Teboul L et al (2010) Overexpression of Fto leads to increased food intake and results in obesity. Nat Genet 42(12):1086–1092
- Clark JM (2006) The epidemiology of nonalcoholic fatty liver disease in adults. J Clin Gastroenterol 40(Suppl 1):S5–S10
- Colditz GA, Willett WC, Rotnitzky A, Manson JE (1995) Weight gain as a risk factor for clinical diabetes mellitus in women. Ann Intern Med 122(7):481–486
- Coll AP, Farooqi IS, Challis BG, Yeo GS, O'Rahilly S (2004) Proopiomelanocortin and energy balance: insights from human and murine genetics. J Clin Endocrinol Metab 89(6):2557–2562
- Depetris RS, Hu J, Gimpelevich I, Holt LJ, Daly RJ, Hubbard SR (2005) Structural basis for inhibition of the insulin receptor by the adaptor protein Grb14. Mol Cell 20(2):325–333
- Doche ME, Bochukova EG, Su HW, Pearce LR, Keogh JM, Henning E et al (2012) Human SH2B1 mutations are associated with maladaptive behaviors and obesity. J Clin Invest 122(12):4732–4736
- Elrick MJ, Pacheco CD, Yu T, Dadgar N, Shakkottai VG, Ware C et al (2010) Conditional Niemann-Pick C mice demonstrate cell autonomous Purkinje cell neurodegeneration. Hum Mol Genet 19(5):837–847

- Fan JG, Zhu J, Li XJ, Chen L, Lu YS, Li L et al (2005) Fatty liver and the metabolic syndrome among Shanghai adults. J Gastroenterol Hepatol 20(12):1825–1832
- Farooqi IS, Keogh JM, Yeo GS, Lank EJ, Cheetham T, O'Rahilly S (2003) Clinical spectrum of obesity and mutations in the melanocortin 4 receptor gene. N Eng J Med 348(12): 1085–1095
- 23. Farooqi IS, Volders K, Stanhope R, Heuschkel R, White A, Lank E et al (2007) Hyperphagia and early-onset obesity due to a novel homozygous missense mutation in prohormone convertase 1/3. J Clin Endocrinol Metab 92(9):3369–3373
- 24. Fox CS, Liu Y, White CC, Feitosa M, Smith AV, Heard-Costa N et al (2012) Genome-wide association for abdominal subcutaneous and visceral adipose reveals a novel locus for visceral fat in women. PLoS Genet 8(5):e1002695
- 25. Fox CS, Massaro JM, Hoffmann U, Pou KM, Maurovich-Horvat P, Liu CY et al (2007) Abdominal visceral and subcutaneous adipose tissue compartments: association with metabolic risk factors in the Framingham Heart Study. Circulation 116(1):39–48
- 26. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, Lindgren CM et al (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316(5826):889–894
- Fredriksson R, Hagglund M, Olszewski PK, Stephansson O, Jacobsson JA, Olszewska AM et al (2008) The obesity gene, FTO, is of ancient origin, up-regulated during food deprivation and expressed in neurons of feeding-related nuclei of the brain. Endocrinology 149(5):2062–2071
- Gerken T, Girard CA, Tung YC, Webby CJ, Saudek V, Hewitson KS et al (2007) The obesityassociated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. Science 318(5855):1469–1472
- 29. Gjesing AP, Vestmar MA, Jorgensen T, Heni M, Holst JJ, Witte DR et al (2011) The effect of PCSK1 variants on waist, waist-hip ratio and glucose metabolism is modified by sex and glucose tolerance status. PLoS One 6(9):e23907
- Grarup N, Sparso T, Hansen T (2010) Physiologic characterization of type 2 diabetes-related loci. Curr Diab Rep 10(6):485–497
- Gratacos M, Gonzalez JR, Mercader JM, de Cid R, Urretavizcaya M, Estivill X (2007) Brainderived neurotrophic factor Val66Met and psychiatric disorders: meta-analysis of casecontrol studies confirm association to substance-related disorders, eating disorders, and schizophrenia. Biol Psychiatry 61(7):911–922
- 32. Han TS, van Leer EM, Seidell JC, Lean ME (1995) Waist circumference action levels in the identification of cardiovascular risk factors: prevalence study in a random sample. BMJ 311(7017):1401–1405
- Harrison SA, Neuschwander-Tetri BA (2004) Nonalcoholic fatty liver disease and nonalcoholic steatohepatitis. Clin Liver Dis 8(4):861–879
- 34. Hashimoto K, Koizumi H, Nakazato M, Shimizu E, Iyo M (2005) Role of brain-derived neurotrophic factor in eating disorders: recent findings and its pathophysiological implications. Prog Neuropsychopharmacol Biol Psychiatry 29(4):499–504
- 35. Heard-Costa NL, Zillikens MC, Monda KL, Johansson A, Harris TB, Fu M et al (2009) NRXN3 is a novel locus for waist circumference: a genome-wide association study from the CHARGE Consortium. PLoS Genet 5(6):e1000539
- 36. Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, Steinthorsdottir V et al (2010) Metaanalysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. Nat Genet 42(11):949–960
- Heid IM, Vollmert C, Kronenberg F, Huth C, Ankerst DP, Luchner A et al (2008) Association of the MC4R V103I polymorphism with the metabolic syndrome: the KORA Study. Obesity (Silver Spring) 16(2):369–376
- Heni M, Haupt A, Schafer SA, Ketterer C, Thamer C, Machicao F et al (2010) Association of obesity risk SNPs in PCSK1 with insulin sensitivity and proinsulin conversion. BMC Med Genet 11:86
- Hishimoto A, Liu QR, Drgon T, Pletnikova O, Walther D, Zhu XG et al (2007) Neurexin 3 polymorphisms are associated with alcohol dependence and altered expression of specific isoforms. Hum Mol Genet 16(23):2880–2891

- Holt LJ, Siddle K (2005) Grb10 and Grb14: enigmatic regulators of insulin action—and more? Biochem J 388(Pt 2):393–406
- 41. Hsu WC, Boyko EJ, Fujimoto WY, Kanaya A, Karmally W, Karter A et al (2012) Pathophysiologic differences among Asians, native Hawaiians, and other Pacific Islanders and treatment implications. Diabetes Care 35(5):1189–1198
- Huang Y, Cohen JC, Hobbs HH (2011) Expression and characterization of a PNPLA3 protein isoform (I148M) associated with nonalcoholic fatty liver disease. J Biol Chem 286(43): 37085–37093
- 43. Huszar D, Lynch CA, Fairchild-Huntress V, Dunmore JH, Fang Q, Berkemeier LR et al (1997) Targeted disruption of the melanocortin-4 receptor results in obesity in mice. Cell 88(1):131–141
- 44. Ikonen E (2008) Cellular cholesterol trafficking and compartmentalization. Nat Rev Mol Cell Biol 9(2):125–138
- 45. Irwin N, Flatt PR (2009) Evidence for beneficial effects of compromised gastric inhibitory polypeptide action in obesity-related diabetes and possible therapeutic implications. Diabetologia 52(9):1724–1731
- 46. Jackson RS, Creemers JW, Ohagi S, Raffin-Sanson ML, Sanders L, Montague CT et al (1997) Obesity and impaired prohormone processing associated with mutations in the human prohormone convertase 1 gene. Nat Genet 16(3):303–306
- Kelly T, Yang W, Chen CS, Reynolds K, He J (2008) Global burden of obesity in 2005 and projections to 2030. Int J Obes 32(9):1431–1437
- Kernie SG, Liebl DJ, Parada LF (2000) BDNF regulates eating behavior and locomotor activity in mice. EMBO J 19(6):1290–1300
- Kilpelainen TO, Zillikens MC, Stancakova A, Finucane FM, Ried JS, Langenberg C et al (2011) Genetic variation near IRS1 associates with reduced adiposity and an impaired metabolic profile. Nat Genet 43(8):753–760
- 50. Kim S, Cho B, Lee H, Choi K, Hwang SS, Kim D et al (2011) Distribution of abdominal visceral and subcutaneous adipose tissue and metabolic syndrome in a Korean population. Diabetes Care 34(2):504–506
- 51. Koizumi H, Hashimoto K, Itoh K, Nakazato M, Shimizu E, Ohgake S et al (2004) Association between the brain-derived neurotrophic factor 196G/A polymorphism and eating disorders. Am J Med Genet B Neuropsychiatr Genet 127B(1):125–127
- Kolterman OG, Insel J, Saekow M, Olefsky JM (1980) Mechanisms of insulin resistance in human obesity: evidence for receptor and postreceptor defects. J Clin Invest 65(6):1272–1284
- 53. Kopelman PG (2000) Obesity as a medical problem. Nature 404(6778):635-643
- 54. Krude H, Biebermann H, Luck W, Horn R, Brabant G, Gruters A (1998) Severe early-onset obesity, adrenal insufficiency and red hair pigmentation caused by POMC mutations in humans. Nat Genet 19(2):155–157
- 55. Lango Allen H, Estrada K, Lettre G, Berndt SI, Weedon MN, Rivadeneira F et al (2010) Hundreds of variants clustered in genomic loci and biological pathways affect human height. Nature 467(7317):832–838
- 56. Li Z, Zhou Y, Carter-Su C, Myers MG Jr, Rui L (2007) SH2B1 enhances leptin signaling by both Janus kinase 2 Tyr813 phosphorylation-dependent and -independent mechanisms. Mol Endocrinol 21(9):2270–2281
- Lindgren CM, Heid IM, Randall JC, Lamina C, Steinthorsdottir V, Qi L et al (2009) Genomewide association scan meta-analysis identifies three Loci influencing adiposity and fat distribution. PLoS Genet 5(6):e1000508
- Liu QR, Drgon T, Walther D, Johnson C, Poleskaya O, Hess J et al (2005) Pooled association genome scanning: validation and use to identify addiction vulnerability loci in two samples. Proc Natl Acad Sci U S A 102(33):11864–11869
- Loos RJ (2012) Genetic determinants of common obesity and their value in prediction. Best Pract Res Clin Endocrinol Metab 26(2):211–226
- Loos RJ, Lindgren CM, Li S, Wheeler E, Zhao JH, Prokopenko I et al (2008) Common variants near MC4R are associated with fat mass, weight and risk of obesity. Nat Genet 40(6):768–775

- 6 Genetic Pleiotropies of Obesity
 - 61. Lyons WE, Mamounas LA, Ricaurte GA, Coppola V, Reid SW, Bora SH et al (1999) Brain-derived neurotrophic factor-deficient mice develop aggressiveness and hyperphagia in conjunction with brain serotonergic abnormalities. Proc Natl Acad Sci U S A 96(26): 15239–15244
 - Maes HH, Neale MC, Eaves LJ (1997) Genetic and environmental factors in relative body weight and human adiposity. Behav Genet 27(4):325–351
 - Manson JE, Willett WC, Stampfer MJ, Colditz GA, Hunter DJ, Hankinson SE et al (1995) Body weight and mortality among women. N Eng J Med 333(11):677–685
 - 64. Marion V, Stoetzel C, Schlicht D, Messaddeq N, Koch M, Flori E et al (2009) Transient ciliogenesis involving Bardet-Biedl syndrome proteins is a fundamental characteristic of adipogenic differentiation. Proc Natl Acad Sci U S A 106(6):1820–1825
 - McIntosh CH, Widenmaier S, Kim SJ (2009) Glucose-dependent insulinotropic polypeptide (Gastric Inhibitory Polypeptide; GIP). Vitam Horm 80:409–471
 - 66. Meyre D, Delplanque J, Chevre JC, Lecoeur C, Lobbens S, Gallina S et al (2009) Genomewide association study for early-onset and morbid adult obesity identifies three new risk loci in European populations. Nat Genet 41(2):157–159
 - 67. Miyawaki K, Yamada Y, Ban N, Ihara Y, Tsukiyama K, Zhou H et al (2002) Inhibition of gastric inhibitory polypeptide signaling prevents obesity. Nat Med 8(7):738–742
 - Morris DL, Cho KW, Zhou Y, Rui L (2009) SH2B1 enhances insulin sensitivity by both stimulating the insulin receptor and inhibiting tyrosine dephosphorylation of insulin receptor substrate proteins. Diabetes 58(9):2039–2047
 - 69. Muller J, Stoetzel C, Vincent MC, Leitch CC, Laurier V, Danse JM et al (2010) Identification of 28 novel mutations in the Bardet-Biedl syndrome genes: the burden of private mutations in an extensively heterogeneous disease. Hum Genet 127(5):583–593
 - Mykytyn K, Braun T, Carmi R, Haider NB, Searby CC, Shastri M et al (2001) Identification of the gene that, when mutated, causes the human obesity syndrome BBS4. Nat Genet 28(2): 188–191
 - Mykytyn K, Nishimura DY, Searby CC, Shastri M, Yen HJ, Beck JS et al (2002) Identification of the gene (BBS1) most commonly involved in Bardet-Biedl syndrome, a complex human obesity syndrome. Nat Genet 31(4):435–438
 - Mykytyn K, Sheffield VC (2004) Establishing a connection between cilia and Bardet-Biedl Syndrome. Trends Mol Med 10(3):106–109
 - 73. O'Rahilly S (2009) Human genetics illuminates the paths to metabolic disease. Nature 462(7271):307–314
 - 74. O'Rahilly S, Farooqi IS (2006) Genetics of obesity. Philos Trans R Soc Lond B Biol Sci 361(1471):1095–1105
 - 75. O'Rahilly S, Farooqi IS (2008) Human obesity: a heritable neurobehavioral disorder that is highly sensitive to environmental conditions. Diabetes 57(11):2905–2910
 - 76. Orho-Melander M, Melander O, Guiducci C, Perez-Martinez P, Corella D, Roos C et al (2008) Common missense variant in the glucokinase regulatory protein gene is associated with increased plasma triglyceride and C-reactive protein but lower fasting glucose concentrations. Diabetes 57(11):3112–3121
 - 77. Ozata M, Ozdemir IC, Licinio J (1999) Human leptin deficiency caused by a missense mutation: multiple endocrine defects, decreased sympathetic tone, and immune system dysfunction indicate new targets for leptin action, greater central than peripheral resistance to the effects of leptin, and spontaneous correction of leptin-mediated defects. J Clin Endocrinol Metab 84(10):3686–3695
 - 78. Povel CM, Boer JM, Onland-Moret NC, Dolle ME, Feskens EJ, van der Schouw YT (2012) Single nucleotide polymorphisms (SNPs) involved in insulin resistance, weight regulation, lipid metabolism and inflammation in relation to metabolic syndrome: an epidemiological study. Cardiovasc Diabetol 11:133
 - Rauch U, Feng K, Zhou XH (2001) Neurocan: a brain chondroitin sulfate proteoglycan. Cell Mol Life Sci 58(12–13):1842–1856
 - Ren D, Zhou Y, Morris D, Li M, Li Z, Rui L (2007) Neuronal SH2B1 is essential for controlling energy and glucose homeostasis. J Clin Invest 117(2):397–406

- Ribases M, Gratacos M, Armengol L, de Cid R, Badia A, Jimenez L et al (2003) Met66 in the brain-derived neurotrophic factor (BDNF) precursor is associated with anorexia nervosa restrictive type. Mol Psychiatry 8(8):745–751
- Romeo S, Kozlitina J, Xing C, Pertsemlidis A, Cox D, Pennacchio LA et al (2008) Genetic variation in PNPLA3 confers susceptibility to nonalcoholic fatty liver disease. Nat Genet 40(12):1461–1465
- Rotman Y, Koh C, Zmuda JM, Kleiner DE, Liang TJ (2010) The association of genetic variability in patatin-like phospholipase domain-containing protein 3 (PNPLA3) with histological severity of nonalcoholic fatty liver disease. Hepatology 52(3):894–903
- 84. Rouille Y, Duguay SJ, Lund K, Furuta M, Gong Q, Lipkind G et al (1995) Proteolytic processing mechanisms in the biosynthesis of neuroendocrine peptides: the subtilisin-like proprotein convertases. Front Neuroendocrinol 16(4):322–361
- Rui L, Carter-Su C (1999) Identification of SH2-bbeta as a potent cytoplasmic activator of the tyrosine kinase Janus kinase 2. Proc Natl Acad Sci U S A 96(13):7172–7177
- 86. Rung J, Cauchi S, Albrechtsen A, Shen L, Rocheleau G, Cavalcanti-Proenca C et al (2009) Genetic variant near IRS1 is associated with type 2 diabetes, insulin resistance and hyperinsulinemia. Nat Genet 41(10):1110–1115
- Samani NJ, Erdmann J, Hall AS, Hengstenberg C, Mangino M, Mayer B et al (2007) Genomewide association analysis of coronary artery disease. N Eng J Med 357(5):443–453
- 88. Saxena R, Hivert MF, Langenberg C, Tanaka T, Pankow JS, Vollenweider P et al (2010) Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. Nat Genet 42(2):142–148
- 89. Scuteri A, Sanna S, Chen WM, Uda M, Albai G, Strait J et al (2007) Genome-wide association scan shows genetic variants in the FTO gene are associated with obesity-related traits. PLoS Genet 3(7):e115
- Seo S, Guo DF, Bugge K, Morgan DA, Rahmouni K, Sheffield VC (2009) Requirement of Bardet-Biedl syndrome proteins for leptin receptor signaling. Hum Mol Genet 18(7): 1323–1331
- 91. Sharma R, Prudente S, Andreozzi F, Powers C, Mannino G, Bacci S et al (2011) The type 2 diabetes and insulin-resistance locus near IRS1 is a determinant of HDL cholesterol and triglycerides levels among diabetic subjects. Atherosclerosis 216(1):157–160
- 92. Siljee JE, Unmehopa UA, Kalsbeek A, Swaab DF, Fliers E, Alkemade A (2013) Melanocortin 4 receptor distribution in the human hypothalamus. Eur J Endocrinol 168(3):361–369
- 93. Sjogren M, Lyssenko V, Jonsson A, Berglund G, Nilsson P, Groop L et al (2008) The search for putative unifying genetic factors for components of the metabolic syndrome. Diabetologia 51(12):2242–2251
- 94. Sparso T, Andersen G, Nielsen T, Burgdorf KS, Gjesing AP, Nielsen AL et al (2008) The GCKR rs780094 polymorphism is associated with elevated fasting serum triacylglycerol, reduced fasting and OGTT-related insulinaemia, and reduced risk of type 2 diabetes. Diabetologia 51(1):70–75
- 95. Speliotes EK (2009) The genetic determinants of common human obesity. Curr Cardiovasc Risk Rep 3:411–417
- Speliotes EK, Butler JL, Palmer CD, Voight BF, Hirschhorn JN (2010) PNPLA3 variants specifically confer increased risk for histologic nonalcoholic fatty liver disease but not metabolic disease. Hepatology 52(3):904–912
- 97. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU et al (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42(11):937–948
- 98. Speliotes EK, Yerges-Armstrong LM, Wu J, Hernaez R, Kim LJ, Palmer CD et al (2011) Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. PLoS Genet 7(3):e1001324
- Steinberg GR, Kemp BE, Watt MJ (2007) Adipocyte triglyceride lipase expression in human obesity. Am J Physiol Endocrinol Metab 293(4):E958–E964

- Stoltenberg SF, Lehmann MK, Christ CC, Hersrud SL, Davies GE (2011) Associations among types of impulsivity, substance use problems and neurexin-3 polymorphisms. Drug Alcohol Depend 119(3):e31–e38
- 101. Strawbridge RJ, Dupuis J, Prokopenko I, Barker A, Ahlqvist E, Rybin D et al (2011) Genomewide association identifies nine common variants associated with fasting proinsulin levels and provides new insights into the pathophysiology of type 2 diabetes. Diabetes 60(10): 2624–2634
- 102. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M et al (2010) Biological, clinical and population relevance of 95 loci for blood lipids. Nature 466(7307): 707–713
- 103. Thorleifsson G, Walters GB, Gudbjartsson DF, Steinthorsdottir V, Sulem P, Helgadottir A et al (2009) Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. Nat Genet 41(1):18–24
- 104. Timpson NJ, Harbord R, Davey Smith G, Zacho J, Tybjaerg-Hansen A, Nordestgaard BG (2009) Does greater adiposity increase blood pressure and hypertension risk?: Mendelian randomization using the FTO/MC4R genotype. Hypertension 54(1):84–90
- 105. Tobin JL, Beales PL (2007) Bardet-Biedl syndrome: beyond the cilium. Pediatr Nephrol 22(7):926–936
- 106. Vance JE (2006) Lipid imbalance in the neurological disorder. Niemann-Pick C disease. FEBS Lett 580(23):5518–5524
- 107. Walters RG, Jacquemont S, Valsesia A, de Smith AJ, Martinet D, Andersson J et al (2010) A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. Nature 463(7281):671–675
- 108. Watari H, Blanchette-Mackie EJ, Dwyer NK, Watari M, Neufeld EB, Patel S et al (1999) Mutations in the leucine zipper motif and sterol-sensing domain inactivate the Niemann-Pick C1 glycoprotein. J Biol Chem 274(31):21861–21866
- 109. Wen W, Cho YS, Zheng W, Dorajoo R, Kato N, Qi L et al (2012) Meta-analysis identifies common variants associated with body mass index in east Asians. Nat Genet 44(3):307–311
- 110. Willer CJ, Speliotes EK, Loos RJ, Li S, Lindgren CM, Heid IM et al (2009) Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet 41(1):25–34
- 111. Willett WC, Dietz WH, Colditz GA (1999) Guidelines for healthy weight. N Eng J Med 341(6):427–434
- 112. Willett WC, Manson JE, Stampfer MJ, Colditz GA, Rosner B, Speizer FE et al (1995) Weight, weight change, and coronary heart disease in women. Risk within the 'normal' weight range. JAMA 273(6):461–465

Chapter 7 Functional Follow-up of Genetic Variants Using *FTO* as the Prime Example

Stefanie Seehaus and Ulrich Rüther

Abstract Obesity is the result of lifestyle and genetic predisposition. Tens of genes have been found to be associated with obesity. Among those, *FTO* showed the highest effect on body weight, but the function of the gene product was unknown at the time of discovering this association. As a consequence, several hundreds of studies have now been performed in just the last 6 years to unravel the biological role of *FTO* as an obesity gene. Although FTO is very likely to act as a RNA demethylase, the process by which FTO influences body weight is still unknown. In this review we have collected and evaluated most of the recent results which contribute to the relatively small but nevertheless important progress in FTO research with respect to obesity.

Abbreviations

BMI	Body mass index
CUX1	Cut-like homeobox 1
FTO	Fat mass and obesity associated
GWAS	Genome-wide association studies
m ⁶ A	N6-methyladenosine
SNP	Single nucleotide polymorphism
alkB	Alpha-ketoglutarate-dependent dioxygenase
ALKBH	Mammalian alkB homolog

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Introduction

As soon as genome-wide association studies (GWAS) were efficiently possible, several groups screened for predispositions of increased body mass index (BMI). Up to date, tens of loci have been identified [1]. The locus with the best association was found to be the "*fat mass and obesity associated*" (*FTO*) gene [1]. Subsequently, this association could be reproduced worldwide for populations of different ethnicities [2–5]. Since the function of FTO was unknown, this review highlights the *FTO* gene as an example to illustrate the complex investigations necessary after the initial identification using GWAS. In principle, several fundamental questions have to be addressed which are equally valid for any other GWAS detected genes of unknown function:

- Is *FTO* really the obesity gene?
- What is the function of the gene product?
- What is causing the difference between the risk and the non-risk allele?
- Is FTO at all a potential drug target for common obesity?

On the History of *Fto*

In mice, *Fto* was identified via positional cloning as far back as 1999 [6]. At that time there was little clue that *FTO* would eventually become of immense interest to the obesity field. It is one of the six genes affected by the mouse mutation, Fused toes (*Ft*) [7]. Mice heterozygous for this 1.6 megabase deletion showed partial syndactyly of their forelimbs with fused toes and tremendous thymic hyperplasia, both caused most likely through impaired apoptosis. Homozygous *Ft*/*Ft* embryos died between embryonic day 10.5 and 12.5 due to severe developmental malformations.

Fto encodes a protein of 502 amino acid residues with a bipartite nuclear localization signal at the N terminus, suggesting a role in the nucleus. However, the 58-kDa protein showed no similarities to known proteins [6]. It is a large gene of nine exons present in all vertebrates and, surprisingly, also in marine algae [8, 9].

When the era of GWAS began, *FTO* was found to be associated with increased body mass index (BMI). Carriers homozygous for the risk allele weighted 3 kg more on average and had a 1.67-fold higher risk to become obese in comparison to non-risk allele carriers [10]. Of the several single nucleotide polymorphisms (SNPs) in the first intron identified so far, most are correlated with increased body weight [10–13]. Thus, all these data suggest *FTO* is indeed an important gene for obesity.

Originally *Fto* was given the name "Fatso", being the biggest gene affected by the *Ft* mutation [6]. However, after the discovery of *FTO* being associated with obesity [10, 13], *FTO* was renamed to the more appropriate designation "*fat mass and obesity associated*".

Is FTO Really the Obesity Gene?

Since most GWAS only indicate a loci and not a specific gene, the discovery raised the question: Is *FTO* really the obesity gene at this location or is it a neighbour of *FTO*? A possible candidate would be *RPGRIP1L* located in very close proximity 5' of *FTO*, and transcribed in the opposite direction. RPGRIP1L has been shown to be essential for embryonic development through its involvement in cilia function [14]. Just a few publications attempted to address this question, which are summarized here.

Ex vivo and in vitro analyses studied the expression of *FTO* and *RPGRIP1L* in subcutaneous fat of lean and obese females, as well as preadipocyte differentiation. *FTO* was found to correlate positively with BMI but the same was not the case with *RPGRIP1L*. However, both *FTO* and *RPGRIP1L* were down-regulated with progression in adipocyte differentiation, suggesting that both might be involved in adipocyte differentiation [15].

Stratigopoulos and colleagues identified the transcription factor cut-like homeobox 1 (CUX1) as a potential regulator of *FTO* and *RPGRIP1L* [16, 17]. In vitro, CUX1 was found to bind to SNP rs8050136 of *FTO* and regulate the expression of *FTO* and *RPGRIP1L* using isolated human fibroblasts [17]. This SNP is part of the linkage disequilibrium region that harbours the first intronic SNP set found to be associated with obesity. This regulation of *FTO* and *RPGRIP1L* expression is suggested to be controlled via two isoforms of CUX1: P110 and P200. The authors suggested a regulatory role for CUX1 in modifying *FTO* and *RPGRIP1L* expression. P110 might operate as an activator of *FTO* and *RPGRIP1L* transcription, whereas P200 might act as a repressor of *FTO* transcription [16]. However, analysis of human *FTO* and *RPGRIP1L* mRNA expression levels from subcutaneous and visceral fat could not support the findings of Stratigopoulos and colleagues. There, expression of both *FTO* and *RPGRIP1L* in adipose tissue was not associated with rs8050136 genotypic status [18].

Fischer and colleagues addressed changes in regulation of *Rpgrip1l* due to structural alterations of the *Fto* gene in the course of its inactivation [19]. Within *Fto*deficient mice, *Rpgrip1l* mRNA levels were unaltered in the hypothalamus, liver and white and brown adipose tissue. This was accomplished by data of Berulava and Horsthemke, showing that intronic SNPs in *FTO* did not affect *RPGRIP1L* expression [20], which is in line with data from a large association study showing no link between *RPGRIP1L* variation and BMI [21]. Hence, *RPGRIP1L* being the gene responsible for obesity in carriers of *FTO* susceptibility-conferring variants is unlikely.

In principle, even the gene at the 3' site of *FTO*, the transcription factor *Iroqouis* 3 (*IRX3*) has to be considered as a potential candidate. This, however, has not been investigated by any researcher to date.

FTO Expression Profile in Relation to Nutritional States

As the function of FTO was initially unknown, expression characteristics of *Fto/FTO* had to be established first, in particular in the context of energy metabolism. Several studies showed ubiquitous expression of murine and human Fto/FTO

within nuclei of various cell types and tissues, revealing a generalized function for FTO. The highest expression of *Fto/FTO* was found in the brain, especially in the hypothalamus and cerebellum [6, 10, 22]. These results, together with the association of *FTO* with obesity, led researchers to focus on a possible role of FTO in regulating energy metabolism [23].

Fto expression in the hypothalamus has been shown to be concentrated in the dorsomedial (DMN), ventromedial (VMN), paraventricular (PVN) and arcuate (ARC) nuclei [22]. Furthermore, *Fto* mRNA has been reported to be localized in neuropeptide-regulating neurons guiding to a control of food intake [24]. Unfortunately, experiments altering nutritional states delivered controversial results in mammals regarding *Fto* expression within the hypothalamus. *Fto* mRNA expression of fasted rodents was found to be reduced [17, 22, 25], unaltered [26, 27] and even increased [24, 28]. In rats, fed on a high fat diet for several weeks, *Fto* mRNA appeared to be differentially expressed as well. In one study, *Fto* expression was up-regulated [29], whereas it was down-regulated in another [30]. In addition, mice receiving a solvent sucrose or Intralipid (fat emulsion) diet to their standard chow for 48 h exhibited unaltered *Fto* expression within the hypothalamus [28]. Olszewski and colleagues distinguished between mice partaking less calories (eating less) and mice taking in more calories (eating more). "Small eaters" were associated with up-regulated *Fto* mRNA expression in comparison to "big eaters" [28].

The discrepancy between Fto mRNA expression levels within the hypothalamus of different mammals made it rather complicated to understand the presumptive role of FTO in regulating appetite and food intake. Several aspects have to be taken in account. First of all, comparing results between different mammals is relatively challenging. Mice are much more sensitive to long-term fasting of 48 h than rats, showing enormous loss of fat stores. Secondly, results are dependent on experimental procedure. Sacrificing fasted rodents at different time points of the circadian system can influence study results. Importantly, comparing *Fto* mRNA expression levels in the entire hypothalamus between fasted and standard chow fed mammals has proven relatively meaningless. Different orexic and anorexic neuropeptides within different hypothalamic nuclei are responsible for controlling food intake [31]; therefore, it is of importance to detect alterations in FTO protein expression within appetite stimulating or inhibiting neuropeptide-harbouring cells due to changes in nutritional state. To our knowledge only McTaggart and colleagues addressed this in part, where they compared the percentage of FTO-positive anorexic proopiomelanocortin (Pomc) cells between fasted and standard chow fed mice without finding a significant difference [26]. Further analyses regarding the number of FTO-positive orexic neuropeptide-harbouring cells remained elusive. Finally, the main question is: "Are FTO expression levels altered between orexic and anorexic neuropeptide-containing cells during changing the nutritional state?" However, due to limitations in the experimental procedure this still remains to be answered.

Nevertheless, other publications tried to investigate factors involved in *FTO* regulation. Guo and colleagues designed 5' deletion constructs of the *FTO* promoter to identify transcription factors relevant to expression of *FTO* [32]. Forkhead box A2 (FOXA2) appeared to bind to the promoter of *FTO* and thereby inhibiting the

expression of *FTO*. Interestingly, FOXA2 can bind also to *melanin-concentrating hormone* (*MCH*) and *Orexin* promoters and therefore regulating the expression of neuropeptides which are responsible for controlling food intake [33].

Functional Studies In Vitro

The Demethylase FTO

The first evidence of FTO function was achieved by in silico analysis, which suggested FTO to be a member of the superfamily of non-heme dioxygenases, depending on both Fe(II) as a co-factor and 2-oxogluterate as a co-substrate [9, 22]. Members of this superfamily are involved in posttranslational modifications, DNA repair and histone modifications and act as cellular sensors for metabolism and oxygen [34, 35]. Analyzing the highly conserved N-terminal region and secondary structures, FTO shows high sequence similarity to alpha-ketoglutarate-dependent dioxygenase alkB of E. coli and its mammalian homologues ALKBH2 and ALKBH3 [9, 22]. Thus, FTO appears to be a member of mammalian alpha-ketoglutaratedependent dioxygenase alkB homologues (ALKBH1-8). AlkB and ALKBH3 prefer single stranded DNA and RNA (ssDNA, ssRNA) as their substrates whereas ALKBH2 prefers double-stranded DNA (dsDNA) [36]. Analyzing substrate specificity, FTO seemed to demethylate 3-methylthymine (m³T) in the ssDNA setting as being the most likely scenario [22]. At this point it was already postulated that FTO may acts as a RNA demethylase, like the mammalian homologue ALKBH3 [22, 37]. Shortly after, in vitro analysis showed demethylation of m³T in ssDNA and 3-methyluracil (m³U) in ssRNA using recombinant human and mouse FTO [37]. Therefore, FTO favoured demethylation of m³U in ssRNA over m³T in ssDNA. Of note, FTO catalyzed demethylation of m³U and m³T with low efficiency; furthermore, m³T methylation of DNA in mammalian had not been described up to that point. Thus, FTO functioning as a DNA demethylase is very unlikely.

Nevertheless, while the N terminus of FTO (32-326 aa residues) exhibits high sequence similarity to alkB and mammalian homologues [9], the C-terminal domain (327-498 aa residues) shows no sequence similarity to any gene. Han et al. shed some light on the function of FTO revealing its crystal structure [38]. As a consequence, the group confirmed the β sheet containing N terminus as the catalytic domain, by showing a core of six anti-parallel β sheet structures forming the so-called jelly-roll motif with an extra loop around the motif. This extra loop, which does not exist in any other alkB member, was suggested to be responsible for FTO selection for ssDNA or ssRNA. Binding of FTO to double-stranded nucleic acids would not be impossible, as the extra loop of FTO would compete with the unmethylated strand of the DNA duplex. Furthermore, it was shown that the alpha sheet containing C terminus formed bulky hydrophobic contacts with the N terminus, suggesting a role for N terminus-stabilization and therefore involving a catalytic role as well.

Recent findings identified N6-methyladenosine (m⁶A) as another substrate for FTO in ssRNA, with a 50-fold higher substrate specificity over m³U under physiological conditions in vitro [39]. Furthermore, FTO was detected in nuclear speckles [40] and was shown to co-localize with nuclear speckle factors, thus providing a hint for FTO in pre-mRNA processing [39]. Overexpression of FTO led to decreased quantity of m⁶A in mRNA, whereas 90 % knockdown of FTO led to an increase in m⁶A quantity in mRNA. This discovery raised questions regarding the role of m⁶A for biological processes. Mapping m⁶A enabled researchers to uncover the human and mouse m⁶A RNA methylomes. Furthermore, m⁶A modifications appeared to be dynamically and reversibly regulated [41, 42]. Collectively, more than 12,000 highly conserved m⁶A sites were identified in the transcripts of more than 7,000 genes, preferentially within 3'UTRs and near stop codons within mRNA. The corresponding genes were found to be involved in transcriptional regulation, RNA metabolism and intracellular signalling cascades. In addition, 67 % of m⁶A-containing 3'UTRs also contained microRNA-binding sites, suggesting a role of m⁶A in microRNA pathways [42]. In addition, m⁶A around stop codons could affect translational efficiency [41]. However, the two RNA methylome publications came to different conclusions. Meyer et al. suggested a localization of m^6A only in mature mRNA [42]. They could not show an enrichment of m⁶A at splice junctions so they asked if previous studies, which used non-specific methylation inhibitors, pointed to the wrong direction proposing a role of m⁶A in mRNA processing. Meanwhile, Dominissini's group [41] based their analysis on a possible role of m6A on RNA splicing. A knockdown of the only known m⁶A methylase, methyltransferase like3 (METTL3), affected alternative splicing of pre-mRNA, resulting in altered signalling pathways and apoptosis via alteration of P53 in a human hepatocellular carcinoma cell line. Therefore, the authors proposed a role of m⁶A modulating pre-mRNA splicing which was supported by the notion that FTO and METTL3 co-localize with splicing proteins in the nucleus, in particular nuclear speckles [39, 43, 44]. Nevertheless, a role for m⁶A in modulating pre-mRNA splicing remained unclear. Recently, a second m⁶A demethylase, the mammalian alkB homolog 5 (ALKBH5), was identified with localization to splicing proteins [44]. Certainly, further studies regarding FTO and ALKBH5 will bring clarity to this issue. Finally, it cannot be excluded that demethylation of m⁶A via FTO could also affect rRNA and tRNA [45, 46].

Role of FTO in Amino Acid Sensing?

Other in vitro studies using human and mouse cell lines suggest a role for FTO in amino acid sensing [47, 48]. Essential amino acid-deprived human and mouse cell lines exhibited down-regulation of *FTO* mRNA and protein levels [47]. In addition, analyses of *Fto*-deficient mouse embryonic fibroblasts (MEFs) and human HEK cells showed decreased cell growth, impaired mRNA translation, more autophagic flux and reduced protein levels of tRNA synthetases, especially leucyl tRNA synthetase [48], thus defining a role for FTO in mammalian target of rapamycin complex 1

(mTORC1) signalling, among others, as an important protein complex for controlling food intake [49]. Future experiments are needed to further investigate how FTO is affected by amino acid sensing and thereby controlling protein biosynthesis.

Loss- and Gain-of-function-analyses of Fto in Mice and Rats

In recent years, *Fto* mouse mutants with loss or gain of function were designed to enable a better understanding of the role of FTO *in vivo*. For such an approach we assigned mouse line numbers for the different *Fto*-deficient mouse mutants based on appearance (mouse lines 1–7). Concordance and differences between mouse lines are illustrated in Table 7.1. Interestingly, constitutive *Fto*-deficient mice (mouse line 1) appeared to be protected from obesity, with decreased body weight and an increase in energy expenditure, although exhibiting relative hyperphagia and somewhat decreased spontaneous locomotor activity [19].

Analyzing body composition using magnetic resonance imaging (MRI) revealed a reduction in fat as well as lean mass. In addition, male mice were more affected then female mice. Furthermore, increased serum adiponectin and decreased serum leptin levels, markers for body weight regulation, underlined the lean phenotype due to loss of FTO within mouse line 1. Even on a high fat diet, constitutive mice of mouse line 1 did not gain weight compared to their wild type littermates. Finally, mice were growth retarded and showed a higher frequency of postnatal death. These results led to the suggestion that FTO is involved in body weight regulation and energy homoeostasis.

Almost as expected, a mouse line overexpressing FTO revealed gain in body weight due to increased fat mass but not lean mass in comparison to their wild type counterparts [50]. A dose-dependent effect with the highest increase in body mass was verified in male and female mice with four *Fto*-copies (two additional copies), respectively weighting an additional 10 or 22 %. Furthermore, mice exhibited an increase in food intake and energy expenditure but no alterations in physical activity or circadian rhythm.

A second mouse line with partial loss of FTO function due to a dominant missense mutation (mouse line 2; Table 7.1) exhibited the lean phenotype as seen in mouse line 1 [19] but to a milder extend [51]. However, several differences persisted between these two *Fto* mouse mutants. For instance, mouse line 2 did not show postnatal death, growth retardation or alterations in food intake and physical activity.

Since then, several other *Fto* mouse mutants were generated. Another constitutive (mouse line 3) and a nervous system-specific *Fto*-deficient mouse mutant (mouse line 4) displayed very similar phenotypes as mouse lines 1 and 2 [19, 51] but with emphasis on postnatal growth retardation [52]. Surprisingly, the lower body weight in mouse line 3 and mouse line 4 (Table 7.1) was found to be due to decreased lean mass. Fat mass was even increased in females of mouse line 3 and both male and female mice of mouse line 4. The authors suggested that FTO exerts its function in the brain and therefore could regulate growth and presumably food intake over

	Mouse line						
	1	2	6	4	5	6	7
	Fischer et al. 2009 [19]	Church et al. 2009 [51]	Gao et al. 2010 [52]	Gao et al. 2010 [52]	McMurray et al. 2013 [53]	McMurray et al. 2013 [53]	McMurray et al. 2013 [53]
<i>Fto</i> -deficiency:	Constitutive	Partial	Constitutive ^a	Neural-specific ^a	Constitutive	Adult onset constitutive	Adult onset hypothalamus-specific ^b
Body weight	\rightarrow	→	\rightarrow	\rightarrow	\rightarrow	\rightarrow	¢
Lean mass	\rightarrow	\$	\rightarrow	\rightarrow	\rightarrow	\rightarrow	¢
Fat mass	\rightarrow	\rightarrow	0;;↓0	~	\rightarrow	\leftrightarrow young; \uparrow old	¢
Postnatal death	←	\$	←	n. a.	←	\$	n. a.
Growth retarded	Yes	No	Yes	Yes	Yes	No	n. a.
Food intake	←	\$	←	←	¢	\$	→
Energy expenditure	←	¢	←	←	¢	¢	¢
Compared to wild tyl	be situation: ↓=dec	crease, ↑=increase	or appearance me	ore often, ↔ = no diff	erence; n.a. not addr	essed	
^a Efficiency of <i>Fto</i> -ini	ctivation could no	t be shown					
^b Only partial inactiva	tion of Fto due to	delivery of Cre by	adenoviral infecti	on			

 Table 7.1
 Overview of published *Fto*-deficient mouse lines

energy expenditure [52]. However, taking a closer look at the study of Gao and colleagues, in both mouse lines (3 and 4) FTO was still found to be expressed within the hypothalamus, especially in neural-specific *Fto*-deficient mouse mutants [52].

Another publication presented three more *Fto* mouse lines (Table 7.1), namely, constitutive (mouse line 5), adult onset (mouse line 6) and adult onset hypothalamic (mouse line 7) Fto-deficient mouse mutants [53]. Comparing the new constitutive mouse line 5 with previously published constitutive Fto-deficient mice (mouse line 1 and 3), a very similar phenotype was found, pointing to a role of FTO in body weight and growth regulation. Adult onset Fto-deficient mice (mouse line 6) were protected from postnatal death, which was not the case for constitutive Fto-deficient mice (mouse line 1, 3 and 5) [19, 52, 53], suggesting FTO as being essential soon after birth. Surprisingly, fat mass of older adult onset Fto-deficient mice (mouse line 6) was found to be increased. It would be of interest to see if fat mass of young adult onset mice (mouse line 6) was increased as well by taking the lower body weight into account (percent fat). Another interesting observation was the decrease in food intake of adult onset hypothalamic Fto-deficient mice (mouse line 7). In this case the authors injected adeno-associated viral vectors encoding Cre recombinase into the hypothalamus of *Fto* floxed mice to regionally delete FTO expression and found decreased food intake. Using the same virus system to knock down FTO expression via short hairpin RNA in the hypothalamus, the same authors reported an increase in food intake in rats. Furthermore, in both rodent lines protein levels of FTO within the hypothalamus or ARC and PVN were found to be reduced by less than 50 % in comparison to sham operated rodents [29, 53]. Thus, the interpretation of these results is extremely complicated and challenging.

Finally, 4 years and 7 *Fto*-deficient mouse mutants later, the role of FTO in obesity is still unclear, especially after the report that an adult onset loss of FTO led to a gain in fat mass. Whether loss or gain of FTO function is involved in obesity still needs to be clarified.

Analysis of Individuals Carrying Genetic Variants of FTO

Several studies have tried to unravel the differences of the risk and non-risk allele of *FTO* for developing obesity. Whereas association between *FTO* variants and increased BMI and body weight were clearly identified [10, 11, 13], the question whether increased, decreased or deregulated FTO expression is responsible for the effect is still open and cannot yet be answered by the use of *Fto* mouse mutants. In addition, variants of the protein could attribute to the gain of body weight. While trying to address this question, most investigators have concentrated on the link between *FTO* risk allele carriers and *FTO* expression levels in adipose tissue. As a consequence, no association between *FTO* genotype and *FTO* expression levels in adipose tissue was found [18, 54, 55]. Furthermore, *FTO* expression levels were found to be either increased or decreased in adipose tissue of obese patients [15, 55–57]. Moreover, inconsistency was found in regard to *FTO* expression in subcutaneous and visceral adipose tissue of obese individuals [18, 55, 56].

Berulava and Horsthemke were so far the only investigators trying to address whether an allele specific difference of *FTO* expression can be found. Instead of analyzing *FTO* mRNA levels they analyzed levels in unspliced nuclear RNA of heterozygous risk allele carriers. Risk allelic expression of *FTO* was found to be more abundant than non-risk allele expression [20]. However, this result was only shown for fibroblasts; nevertheless, this study suggests that *FTO* overexpression might lead to an increase in body weight.

One study addressed variants that may impact the FTO protein by screening the nine exons of *FTO* in 2,866 individuals and identifying heterozygous non-synonymous variants [58]. However, these were equally frequent between lean and obese individuals. Furthermore, loss-of-function mutations of *FTO* were found in lean and obese people. Thus, showing clearly that non-synonymous variants in the coding region of *FTO*, and therefore of the protein, are not linked to obesity.

FTO as a Therapeutic Target for Obesity?

Even with the fragmentary knowledge of FTO function as a demethylase, the first mechanistic study was published presenting an effective cell-active inhibitor for human FTO, called rhein [59]. Rhein binds FTO reversibly, thus blocking the recognition site of FTO for m⁶A substrates. However, considering the little knowledge we still have of the biochemical role of FTO, its ubiquitous expression in every tissue and the severe phenotype documented in humans being deficient for FTO demethylase activity [60], application of any drug-based anti-obesity therapy is currently still extremely risky. In other words, elucidation of the role of FTO is indispensable to design a tissue or signalling pathway-specific drug for FTO targeting.

How to Continue Functional Studies on FTO

So overall, what is known about FTO today? FTO is a member of the alkB demethylase family. Loss of FTO function leads to reduced body weight and growth retardation. However, whether the demethylase activity of FTO is responsible for the effects seen in mutant mice still remains unclear. If the demethylase activity of FTO is responsible, what are the targets of FTO and are these targets organ specific? Further, if FTO is involved in epigenetic regulation of the mammalian transcriptome and therefore might demethylate a high number of targets, which of those targets are relevant for obesity? Answering these questions might help to develop a therapy against obesity which in turn reduces the side effects to a minimum.

Much effort was put into uncovering the function of FTO and how it may be involved in becoming obese. However, still little is known. But at the end of the day, independent of our genetic status we should not forget: Do not eat more than your body burns, otherwise you will gain weight. Acknowledgements We would like to thank Cindy Thron and Renate Dildrop for their help in the context of manuscript preparation.

References

- Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU et al (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42(11):937–948
- Cornes BK, Lind PA, Medland SE, Montgomery GW, Nyholt DR, Martin NG (2009) Replication of the association of common rs9939609 variant of FTO with increased BMI in an Australian adult twin population but no evidence for gene by environment (G x E) interaction. Int J Obes (Lond) 33(1):75–79
- Hotta K, Nakata Y, Matsuo T, Kamohara S, Kotani K, Komatsu R et al (2008) Variations in the FTO gene are associated with severe obesity in the Japanese. J Hum Genet 53(6):546–553
- Tan JT, Dorajoo R, Seielstad M, Sim XL, Ong RT, Chia KS et al (2008) FTO variants are associated with obesity in the Chinese and Malay populations in Singapore. Diabetes 57(10):2851–2857
- Villalobos-Comparan M, Teresa Flores-Dorantes M, Teresa Villarreal-Molina M, Rodriguez-Cruz M, Garcia-Ulloa AC, Robles L et al (2008) The FTO gene is associated with adulthood obesity in the Mexican population. Obesity (Silver Spring) 16(10):2296–2301
- 6. Peters T, Ausmeier K, Rüther U (1999) Cloning of Fatso (Fto), a novel gene deleted by the Fused toes (Ft) mouse mutation. Mamm Genome 10(10):983–986
- 7. van der Hoeven F, Schimmang T, Volkmann A, Mattei MG, Kyewski B, Rüther U (1994) Programmed cell death is affected in the novel mouse mutant Fused toes (Ft). Development 120(9):2601–2607
- Robbens S, Rouze P, Cock JM, Spring J, Worden AZ, Van de Peer Y (2008) The FTO gene, implicated in human obesity, is found only in vertebrates and marine algae. J Mol Evol 66(1):80–84
- Sanchez-Pulido L, Andrade-Navarro MA (2007) The FTO (fat mass and obesity associated) gene codes for a novel member of the non-heme dioxygenase superfamily. BMC Biochem 8:23
- Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, Lindgren CM et al (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316(5826):889–894
- Dina C, Meyre D, Gallina S, Durand E, Korner A, Jacobson P et al (2007) Variation in FTO contributes to childhood obesity and severe adult obesity. Nat Genet 39(6):724–726
- 12. Hinney A, Nguyen TT, Scherag A, Friedel S, Bronner G, Muller TD et al (2007) Genome wide association (GWA) study for early onset extreme obesity supports the role of fat mass and obesity associated gene (FTO) variants. PLoS One 2(12):e1361
- Scuteri A, Sanna S, Chen WM, Uda M, Albai G, Strait J et al (2007) Genome-wide association scan shows genetic variants in the FTO gene are associated with obesity-related traits. PLoS Genet 3(7):e115
- Vierkotten J, Dildrop R, Peters T, Wang B, Rüther U (2007) Ftm is a novel basal body protein of cilia involved in Shh signalling. Development 134(14):2569–2577
- 15. Tews D, Fischer-Posovszky P, Wabitsch M (2011) Regulation of FTO and FTM expression during human preadipocyte differentiation. Horm Metab Res 43(1):17–21
- Stratigopoulos G, LeDuc CA, Cremona ML, Chung WK, Leibel RL (2011) Cut-like homeobox 1 (CUX1) regulates expression of the fat mass and obesity-associated and retinitis pigmentosa GTPase regulator-interacting protein-1-like (RPGRIP1L) genes and coordinates leptin receptor signaling. J Biol Chem 286(3):2155–2170
- Stratigopoulos G, Padilla SL, LeDuc CA, Watson E, Hattersley AT, McCarthy MI et al (2008) Regulation of Fto/Ftm gene expression in mice and humans. Am J Physiol Regul Integr Comp Physiol 294(4):R1185–R1196

- Klöting N, Schleinitz D, Ruschke K, Berndt J, Fasshauer M, Tonjes A et al (2008) Inverse relationship between obesity and FTO gene expression in visceral adipose tissue in humans. Diabetologia 51(4):641–647
- Fischer J, Koch L, Emmerling C, Vierkotten J, Peters T, Brüning JC et al (2009) Inactivation of the Fto gene protects from obesity. Nature 458(7240):894–898
- Berulava T, Horsthemke B (2010) The obesity-associated SNPs in intron 1 of the FTO gene affect primary transcript levels. Eur J Hum Genet 18(9):1054–1056
- 21. Jacobsson JA, Danielsson P, Svensson V, Klovins J, Gyllensten U, Marcus C et al (2008) Major gender difference in association of FTO gene variant among severely obese children with obesity and obesity related phenotypes. Biochem Biophys Res Commun 368(3):476–482
- Gerken T, Girard CA, Tung YC, Webby CJ, Saudek V, Hewitson KS et al (2007) The obesityassociated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. Science 318(5855):1469–1472
- 23. Suzuki K, Jayasena CN, Bloom SR (2012) Obesity and appetite control. Exp Diabetes Res 2012:824305
- 24. Fredriksson R, Hagglund M, Olszewski PK, Stephansson O, Jacobsson JA, Olszewska AM et al (2008) The obesity gene, FTO, is of ancient origin, up-regulated during food deprivation and expressed in neurons of feeding-related nuclei of the brain. Endocrinology 149(5):2062–2071
- 25. Poritsanos NJ, Lew PS, Fischer J, Mobbs CV, Nagy JI, Wong D et al (2011) Impaired hypothalamic Fto expression in response to fasting and glucose in obese mice. Nutr Diabetes 1:e19
- McTaggart JS, Lee S, Iberl M, Church C, Cox RD, Ashcroft FM (2011) FTO is expressed in neurones throughout the brain and its expression is unaltered by fasting. PLoS One 6(11):e27968
- 27. Olszewski PK, Radomska KJ, Ghimire K, Klockars A, Ingman C, Olszewska AM et al (2011) Fto immunoreactivity is widespread in the rodent brain and abundant in feeding-related sites, but the number of Fto-positive cells is not affected by changes in energy balance. Physiol Behav 103(2):248–253
- Olszewski PK, Fredriksson R, Olszewska AM, Stephansson O, Alsio J, Radomska KJ et al (2009) Hypothalamic FTO is associated with the regulation of energy intake not feeding reward. BMC Neurosci 10:129
- 29. Tung YC, Ayuso E, Shan X, Bosch F, O'Rahilly S, Coll AP et al (2010) Hypothalamic-specific manipulation of Fto, the ortholog of the human obesity gene FTO, affects food intake in rats. PLoS One 5(1):e8771
- Gutierrez-Aguilar R, Kim DH, Woods SC, Seeley RJ (2012) Expression of new loci associated with obesity in diet-induced obese rats: from genetics to physiology. Obesity (Silver Spring) 20(2):306–312
- Parker JA, Bloom SR (2012) Hypothalamic neuropeptides and the regulation of appetite. Neuropharmacology 63(1):18–30
- 32. Guo J, Ren W, Ding Y, Li A, Jia L, Su D et al (2012) Fat mass and obesity associated gene (FTO) expression is regulated negatively by the transcription factor Foxa2. PLoS One 7(12):e51082
- Silva JP, von Meyenn F, Howell J, Thorens B, Wolfrum C, Stoffel M (2009) Regulation of adaptive behaviour during fasting by hypothalamic Foxa2. Nature 462(7273):646–650
- 34. Clifton IJ, McDonough MA, Ehrismann D, Kershaw NJ, Granatino N, Schofield CJ (2006) Structural studies on 2-oxoglutarate oxygenases and related double-stranded beta-helix fold proteins. J Inorg Biochem 100(4):644–669
- 35. Ozer A, Bruick RK (2007) Non-heme dioxygenases: cellular sensors and regulators jelly rolled into one? Nat Chem Biol 3(3):144–153
- 36. Aas PA, Otterlei M, Falnes PO, Vagbo CB, Skorpen F, Akbari M et al (2003) Human and bacterial oxidative demethylases repair alkylation damage in both RNA and DNA. Nature 421(6925):859–863
- 37. Jia G, Yang CG, Yang S, Jian X, Yi C, Zhou Z et al (2008) Oxidative demethylation of 3-methylthymine and 3-methyluracil in single-stranded DNA and RNA by mouse and human FTO. FEBS Lett 582(23–24):3313–3319
- Han Z, Niu T, Chang J, Lei X, Zhao M, Wang Q et al (2010) Crystal structure of the FTO protein reveals basis for its substrate specificity. Nature 464(7292):1205–1209
- 39. Jia G, Fu Y, Zhao X, Dai Q, Zheng G, Yang Y et al (2011) N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. Nat Chem Biol 7(12):885–887

- 40. Berulava T, Ziehe M, Klein-Hitpass L, Mladenov E, Thomale J, Rüther U et al (2012) FTO levels affect RNA modification and the transcriptome. Eur J Hum Genet 21(3):317–323
- 41. Dominissini D, Moshitch-Moshkovitz S, Schwartz S, Salmon-Divon M, Ungar L, Osenberg S et al (2012) Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq. Nature 485(7397):201–206
- 42. Meyer KD, Saletore Y, Zumbo P, Elemento O, Mason CE, Jaffrey SR (2012) Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. Cell 149(7):1635–1646
- 43. Bokar JA, Shambaugh ME, Polayes D, Matera AG, Rottman FM (1997) Purification and cDNA cloning of the AdoMet-binding subunit of the human mRNA (N6-adenosine)methyltransferase. RNA 3(11):1233–1247
- 44. Zheng G, Dahl JA, Niu Y, Fedorcsak P, Huang CM, Li CJ et al (2013) ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. Mol Cell 49(1):18–29
- 45. Ougland R, Zhang CM, Liiv A, Johansen RF, Seeberg E, Hou YM et al (2004) AlkB restores the biological function of mRNA and tRNA inactivated by chemical methylation. Mol Cell 16(1):107–116
- 46. Jia G, Fu Y, He C (2013) Reversible RNA adenosine methylation in biological regulation. Trends Genet 29(2):108–115
- Cheung MK, Gulati P, O'Rahilly S, Yeo GS (2013) FTO expression is regulated by availability of essential amino acids. Int J Obes (Lond) 37(5):744–747
- 48. Gulati P, Cheung MK, Antrobus R, Church CD, Harding HP, Tung YC et al (2013) Role for the obesity-related FTO gene in the cellular sensing of amino acids. Proc Natl Acad Sci U S A 110(7):2557–2562
- Cota D, Proulx K, Smith KA, Kozma SC, Thomas G, Woods SC et al (2006) Hypothalamic mTOR signaling regulates food intake. Science 312(5775):927–930
- Church C, Moir L, McMurray F, Girard C, Banks GT, Teboul L et al (2010) Overexpression of Fto leads to increased food intake and results in obesity. Nat Genet 42(12):1086–1092
- 51. Church C, Lee S, Bagg EA, McTaggart JS, Deacon R, Gerken T et al (2009) A mouse model for the metabolic effects of the human fat mass and obesity associated FTO gene. PLoS Genet 5(8):e1000599
- 52. Gao X, Shin YH, Li M, Wang F, Tong Q, Zhang P (2010) The fat mass and obesity associated gene FTO functions in the brain to regulate postnatal growth in mice. PLoS One 5(11):e14005
- 53. McMurray F, Church CD, Larder R, Nicholson G, Wells S, Teboul L et al (2013) Adult onset global loss of the fto gene alters body composition and metabolism in the mouse. PLoS Genet 9(1):e1003166
- 54. Grunnet LG, Nilsson E, Ling C, Hansen T, Pedersen O, Groop L et al (2009) Regulation and function of FTO mRNA expression in human skeletal muscle and subcutaneous adipose tissue. Diabetes 58(10):2402–2408
- 55. Zabena C, Gonzalez-Sanchez JL, Martinez-Larrad MT, Torres-Garcia A, Alvarez-Fernandez-Represa J, Corbaton-Anchuelo A et al (2009) The FTO obesity gene. Genotyping and gene expression analysis in morbidly obese patients. Obes Surg 19(1):87–95
- 56. Terra X, Auguet T, Porras JA, Quintero Y, Aguilar C, Luna AM et al (2010) Anti-inflammatory profile of FTO gene expression in adipose tissues from morbidly obese women. Cell Physiol Biochem 26(6):1041–1050
- Wahlen K, Sjolin E, Hoffstedt J (2008) The common rs9939609 gene variant of the fat massand obesity-associated gene FTO is related to fat cell lipolysis. J Lipid Res 49(3):607–611
- 58. Meyre D, Proulx K, Kawagoe-Takaki H, Vatin V, Gutierrez-Aguilar R, Lyon D et al (2010) Prevalence of loss-of-function FTO mutations in lean and obese individuals. Diabetes 59(1):311–318
- Chen B, Ye F, Yu L, Jia G, Huang X, Zhang X et al (2012) Development of cell-active N6-methyladenosine RNA demethylase FTO inhibitor. J Am Chem Soc 134(43):17963–17971
- 60. Boissel S, Reish O, Proulx K, Kawagoe-Takaki H, Sedgwick B, Yeo GS et al (2009) Loss-offunction mutation in the dioxygenase-encoding FTO gene causes severe growth retardation and multiple malformations. Am J Hum Genet 85(1):106–111

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