Chapter 24 The Effect of Basic Assumptions on the Tissue Oxygen Saturation Value of Near Infrared Spectroscopy

Andreas Jaakko Metz*, Martin Biallas, Carmen Jenny, **Thomas Muehlemann, and Martin Wolf**

Abstract Tissue oxygen saturation $(StO₂)$, a potentially important parameter in clinical practice, can be measured by near infrared spectroscopy (NIRS). Various devices use the multi-distance approach based on the diffusion approximation of the radiative transport equation [1, 2]. When determining the absorption coefficient (μ_{a}) by the slope over multiple distances a common assumption is to neglect μ_a in the diffusion constant, or to assume the scattering coefficient (μ_s) to be constant over the wavelength. Also the water influence can be modeled by simply subtracting a water term from the absorption. This gives five approaches $A1-A5$. The aim was to test how these different methods influence the $StO₂$ values. One data set of 30 newborn infants measured on the head and another of eight adults measured on the nondominant forearm were analyzed. The calculated average StO_2 values measured on the head were (mean ± SD): A1: 79.99 ± 4.47%, A2: 81.44 ± 4.08%, A3: 84.77 ± 4.87%, A4: 85.69 ± 4.38%, and A5: 72.85 ± 4.81%. The StO₂ values for the adult forearms are: A1: $58.14 \pm 5.69\%$, A2: $73.85 \pm 4.77\%$, A3: $58.99 \pm 5.67\%$, A4: 74.21 \pm 4.76%, and A5: 63.49 \pm 5.11%. Our results indicate that StO₂ depends strongly on the assumptions. Since StO_2 is an absolute value, comparability between different studies is reduced if the assumptions of the algorithms are not published.

Keywords Absorption coefficient • Methodology • Near-infrared spectroscopy • Tissue oxygen saturation

^{*} Andreas Jaakko Metz is the member of the Ph.D. Program imMed

A.J. Metz $(\boxtimes) \cdot M$. Biallas $\cdot C$. Jenny $\cdot T$. Muehlemann $\cdot M$. Wolf Biomedical Optics Research Laboratory, Division of Neonatology, University Hospital Zurich, Zurich, Switzerland

Zurich Center for Integrative Physiology, University Zurich, Frauenklinikstrasse 10 , 8091 Zürich , Switzerland e-mail: andreas.metz@usz.ch

W.J. Welch et al. (eds.), *Oxygen Transport to Tissue XXXIV*, Advances in Experimental 169 Medicine and Biology 765, DOI 10.1007/978-1-4614-4989-8_24, © Springer Science+Business Media New York 2013

1 Introduction

Tissue oxygen saturation (StO_2) has a great potential to become an important clinical parameter, especially in neonatology $[3, 4]$. It is related to the oxygen metabolism in the tissue on an absolute scale. Slightly different approaches are used to calculate StO_2 , depending on the manufacturer. This is reflected in different naming, e.g., tissue oxygenation index for the NIRO (Hamamatsu Photonics, Japan) [5] or regional oxygen saturation for the INVOS (Somanetics Corp., USA) or Critikon (Johnson & Johnson, UK). Studies have been published, which compared the values obtained from the three different devices and found differences between INVOS and Critikon [6] and agreement between the NIRO and INVOS [7, 8]. However, both found *unacceptable* baseline differences. Several reasons were given as explanation: Differences in the technical setup, the effect of extracranial blood flow and differences in the algorithm.

However, the influence of the algorithm itself has to our knowledge not been evaluated. Our aim was to test the influence of basic assumptions of the multidistance approach $[1]$, which is similar to spatially resolved spectroscopy $[2]$. Using the different approaches on the same data sets excludes the instrumentation or extracranial blood flow as a source of differences.

2 Methods

2.1 Subjects

 Data sets from two different studies have been evaluated. First, 30 newborn infants have been studied previously in our group with the aim to identify precision of NIRS [9]. Second, eight adult subjects (all male, age range 26–45, median 29.5) were investigated within a still ongoing study. Both studies were approved by the ethical committee of the Kanton of Zurich and informed consent was obtained prior to the study.

2.2 Protocol

Neonatal group . The frontal and temporal cerebral region was measured four times for approximately 1 min. The sensor was repositioned between the measurements [9].

Adult group. Five repeated measurements per subject were taken from the nondominant forearm, near to musculus brachioradialis. The sensor was fixated with an elastic bandage around the forearm. Each measurement took 1 min, in between measurements the bandage was completely removed and the sensor was repositioned to approximately the same place as before.

2.3 NIRS Measurement

 The neonatal group was measured with the MCPII, which is described in detail elsewhere $[10]$. It uses three wavelengths $(750, 800,$ and $875 \text{ nm})$ at distances of 1.25 and 2.5 cm.

 The adult group was assessed by a novel continuous wave NIRS device, the $OxyPrem$, which is similar to previous wireless sensors [11]. It measures light attenuation at 760 and 870 nm, at distances of 1.5 and 2.5 cm.

2.4 Theory

 Tissue oxygen saturation was calculated by a self-calibrating multi-distance approach [1] based on the diffusion approximation of the radiative transport equation and using two sources and two detectors. The light intensity decreases with the distance. This relation is linear (semi-infinite boundary condition).

$$
\ln(dc(r)r^2) = rS1_{dc}(\mu_a, \mu_s) + \ln_{dc}(D, K_{dc}),
$$
\n(24.1)

 $dc(r)$ is the average light intensity as a function of distance r, SL_c the slope of the intensity loss and In'_{dc} the intercept. μ_{a} and μ_{s}' are the absorption and the reduced scattering coefficient, respectively. K_d is a constant. The diffusion constant *D* equals

$$
D = \frac{1}{3\mu_{a} + 3\mu_{s}'} \approx \frac{1}{3\mu_{s}'}.
$$
 (24.2)

 μ _a is often neglected because tissue scattering is much larger than absorption $(\mu_{\rm s})$. However, here we distinguish between simplified and exact diffusion constant (as seen below).

When evaluating (24.1) at two distances r_L and r_S and subtracting them, the slope can be calculated from the ratio of the measured intensities.

$$
SI_{dc} = \frac{\frac{1}{2} \ln \left(\frac{dc_1(r_L)dc_2(r_L)}{dc_1(r_s)dc_2(r_s)} \right) + 2 \ln \left(\frac{r_L}{r_s} \right)}{r_L - r_s},
$$
(24.3)

where r_L is the longer source–detector distance and r_S the shorter one, respectively. Equation (24.3) is a special self-calibrating form, whereby the use of two source– detector pairs [giving the four intensity values $dc_{1,2}(r_L, r_S)$] the coupling factors between the tissue and source/detector cancel out [1]. Then μ _a can be calculated as

$$
\mu_a = \mathrm{SI}_{dc}^2 D. \tag{24.4}
$$

 When the absorption is determined at least at two wavelengths, concentrations of oxygenated $([O_2Hb])$ and deoxygenated hemoglobin $([HHb])$ and the tissue oxygen

saturation can be calculated. We used the absorption coefficients from Matcher et al. [12], averaged over the measured intensity spectrum of each light source. Coefficients for scattering were taken from Matcher et al. [[13](#page-6-0)] for the adult arm and from ISS OxyPlex measurements on 36 term infants $[14]$ for the neonates, extrapolated to 750, 800, and 875 nm (3.81, 3.49, and 3.01[cm⁻¹]).

$$
\begin{bmatrix}\n[HHb] \\
[O_2Hb]\n\end{bmatrix} = \mathbf{A}^{-1} \begin{bmatrix}\n\mu_a(\lambda_1) \\
\mu_a(\lambda_2)\n\end{bmatrix}, \quad \mathbf{A} = \begin{bmatrix}\na_{HHb,\lambda_1} & a_{O_2Hb,\lambda_1} \\
a_{HHb,\lambda_2} & a_{O_2Hb,\lambda_2}\n\end{bmatrix}
$$
\n(24.5)

where a_{ij} is the absorption coefficient for $i = ($ [HHb], [O₂Hb]) at the wavelength *j*. StO₂ is calculated as $[O_2Hb]/([O_2Hb] + [HHb])$. We examine five different assumptions A1–A5 for the determination of the absorption:

A1.
$$
\mu_{a} = -\frac{\mu'_{s}}{2} + \sqrt{\frac{1}{4}\mu_{s}^{2} + \frac{1}{3}SI_{dc}^{2}} - a_{H_{2}0,\lambda}55.5 M \frac{p_{H_{2}0}}{100\%},
$$
 (24.6)

A2.
$$
\mu_{a} = -\frac{\mu'_{s}}{2} + \sqrt{\frac{1}{4}\mu''_{s}^{2} + \frac{1}{3}SI_{dc}^{2}},
$$
 (24.7)

A3.
$$
\mu_{a} = \frac{SI_{dc}^{2}}{3\mu_{s}} - a_{H_{2}O,\lambda} 55.5 M \frac{p_{H_{2}O}}{100\%},
$$
 (24.8)

A4.
$$
\mu_{a} = \frac{SI_{dc}^{2}}{3\mu_{s}^{'}},
$$
 (24.9)

A5.
$$
StO_{2} = \frac{a_{HHb,\lambda_{1}} - a_{HHb,\lambda_{2}} \left(\frac{Sl_{dc}(\lambda_{1})}{Sl_{dc}(\lambda_{2})} \right)^{2}}{(a_{HHb,\lambda_{1}} - a_{O_{2}Hb,\lambda_{1}}) - (a_{HHb,\lambda_{2}} - a_{O_{2}Hb,\lambda_{2}}) \left(\frac{Sl_{dc}(\lambda_{1})}{Sl_{dc}(\lambda_{2})} \right)^{2}},
$$
(24.10)

Equations (24.6) and (24.7) use the exact diffusion constant, while (24.8) – (24.10) use the simplified one. In (24.10), μ_{s} is assumed to be constant over the wavelength. Hence, it cancels out in StO_2 calculation as shown. Equations (24.6) and (24.8) are accounting for water in tissue. Here $a_{H2O,\lambda}$ is the absorption of water at the wavelength λ in 1/(M^{*}cm) and p_{H2O} is the amount of water in the tissue. We used 70% for the adult forearm and 90% for the neonatal head. Water contains approximately 55.5 mol atoms/l.

2.5 Statistics

 Between-subject variability and within-subject variability were determined using *R* (version 2.6.1, *R* Development Core Team, Austria) with its linear mixed effects function LME. StO_2 was the random variable and subject the factor.

| \ldots . The contract of \ldots and \ldots and \ldots are \ldots and \ldots are \ldots and \ldots are \ldots | | | | | | | | |
|--|------------|------------------|------------------|------------------|------------------|--|--|--|
| | ΑI | А2 | А3 | A4 | А5 | | | |
| $StO, \pm SD [\%]$ | 79.99±4.47 | 81.44 ± 4.08 | 84.77 ± 4.87 | 85.69 ± 4.38 | 72.85 ± 4.81 | | | |
| $\text{Var}_{\text{het}} [\%]$ | 4.2 | 3.84 | 4.64 | 4.16 | 4.56 | | | |
| $Var_{within} [\%]$ | 2.76 | 2.55 | 2.73 | 2.51 | 2.83 | | | |

Table 24.1 StO₂, within-subject variability and between-subject variability for 30 newborn infants measured on the head for the five different assumptions $A1-A5$

Table 24.2 StO₂, within-subject variability and between-subject variability for eight adults measured on the forearm for the five different assumptions $A1-A5$

| | ΑI | А2 | A3 | A4 | A5 |
|--------------------------------|------------------|------------------|------------------|------------------|------------------|
| $StO, \pm SD [\%]$ | 58.14 ± 5.69 | 73.85 ± 4.77 | 58.99 ± 5.67 | 74.21 ± 4.76 | 63.49 ± 5.11 |
| $\text{Var}_{\text{het}} [\%]$ | 5.54 | 4.65 | 5.52 | 4.64 | 4.98 |
| $Var_{within} [\%]$ | 2.96 | 2.43 | 2.95 | 2.42 | 2.60 |

3 Results

For the neonatal head measurements the mean $StO_2 \pm$ standard deviation (SD), the within-subject variability (Var $_{\text{within}}$) and the between-subject variability (Var $_{\text{ber}}$) are given in Table 24.1 . In Table 24.2 the values for the adult group are shown.

In both adults and the neonates assumption A5 deviates in value \sim 10%. In neonates including a water term (A1 vs. A2, A3 vs. A4) has a minor effect on StO_2 , but the use of the exact or simplified diffusion constant $(A1 vs. A3, A2 vs. A4)$ induces a change in StO_2 by 5%. In contrast, on the adult arm, the water term makes a large difference of \sim 15%, while the diffusion constant assumption does induce smaller changes. For both groups, between-subject variability and within-subject variability are smaller when not including the water term (A2 and A4). Both variables are ~0.3% larger when additionally assuming μ_s to be constant (A5 against A2, A4).

4 Discussion and Conclusion

Our results show, that slight differences in the assumptions have a relevant influence on the final StO_2 value. This difference is also dependent on the measured tissue. The water term seems to have a smaller influence in neonates than the tissue homogeneity $(\mu_a \ll \mu_s)$. This may reflect the influence of the cerebral spinal fluid in the brain [15]. Since the water term only induces a small correction of StO_2 we believe that the water correction is more or less correct. However, the variability within and in between subjects is smaller when not including the water term, although only by $\sim 0.2\%$.

 Regarding the arm tissue of the adults, the concentration of lipid is higher and the water concentration is lower than for the neonatal head. While the diffusion constant assumption does not affect the $StO₂$ value, the water term makes a difference of \sim 15%. Since no real reference value exists for StO₂, it is not possible to state if one assumption is more valid than another. From a mathematical point of view, the water term has no relevant influence if the slope (24.3) is much larger than the water term. Hence, the ratio between the long and short distances is much smaller than 1. If the ratio is close to 1, the slope will be small and the water term (usually in the order of 10^{-2}) dominates. This means the ratio is closer to 1 when measuring the adult arm. This may be due to the lipid concentration in the arm, which has not been taken into account, or due to the 70% water assumption, which may be too high, or both. We calculated the body mass index (BMI) for the subjects, which correlated with the change in StO_2 (data not shown), i.e., the higher the BMI and hence the lipid concentration, the higher the change of $StO₂$ when taking water into account. The latter is supported by the fact that not subtracting the water lowers the variability. The additional assumption A5 lowers the $StO₂$ values, compared to A2 and A4. This suggests that this assumption is not valid, neither in the neonatal head nor in the adult arm.

In conclusion, we investigated the effect of the assumptions μ_s , $\mu_a \ll \mu_s$ = constant over the wavelengths and the water contribution and their combinations when using the multi-distance approach of StO_2 calculation. We found significant differences in StO_2 and its variability, depending on the assumptions made and the tissue investigated.

Acknowledgments This work was financially supported by the Zurich Center of Integrative Human Physiology (ZIHP), University of Zurich, Switzerland. The authors would like to thank Raphael Zimmermann for very helpful discussions.

References

- 1. Hueber DM, Fantini S, Cerussi AE et al (1999) New optical probe designs for absolute (selfcalibrating) NIR tissue hemoglobin measurements. Proc SPIE 3597:618–631
- 2. Matcher SJ, Kirkpatrick P, Nahid K et al (1995) Absolute quantification methods in tissue near infrared spectroscopy. Proc SPIE 2359:486–495
- 3. van Bel F, Lemmers P, Naulaers G (2008) Monitoring neonatal regional cerebral oxygen saturation in clinical practice: value and pitfalls. Neonatology 94(4):237–244
- 4. Wolf M, Greisen G (2009) Advances in near-infrared spectroscopy to study the brain of the preterm and term neonate. Clin Perinatol 36(4):807–834
- 5. Suzuki S, Takasaki S, Ozaki T et al (1999) A tissue oxygenation monitor using NIR spatially resolved spectroscopy. Proc SPIE 3597:582–592
- 6. McKeating EG, Monjardino JR, Signorini DF et al (1997) A comparison of the Invos 3100 and the Critikon 2020 near-infrared spectrophotometers as monitors of cerebral oxygenation. Anaesthesia 52(2):136–140
- 7. Thavasothy M, Broadhead M, Elwell C et al (2002) A comparison of cerebral oxygenation as measured by the NIRO 300 and the INVOS 5100 Near-Infrared Spectrophotometers. Anaesthesia 57(10):999–1006
- 8. Yoshitani K, Kawaguchi M, Tatsumi K et al (2002) A comparison of the INVOS 4100 and the NIRO 300 near-infrared spectrophotometers. Anesth Analg 94(3):586–590
- 9. Jenny C, Biallas M, Trajkovic I et al (2011) Reproducibility of cerebral tissue oxygenation saturation measurements by near infrared spectroscopy in newborn infants. J Biomed Opt 16(9): 097004
- 24 Basic assumptions on the tissue oxygen saturation 175
- 10. Haensse D, Szabo P, Brown D et al (2005) New multichannel near infrared spectrophotometry system for functional studies of the brain in adults and neonates. Opt Express 13(12): 4525–4538
- 11. Muehlemann T, Haensse D, Wolf M (2008) Wireless miniaturized in-vivo near infrared imaging. Opt Express 16(14):10323–10330
- 12. Matcher SJ, Elwell CE, Cooper CE et al (1995) Performance comparison of several published tissue near-infrared spectroscopy algorithms. Anal Biochem 227(1):54–68
- 13. Matcher SJ, Cope M, Delpy DT (1997) In vivo measurements of the wavelength dependence of tissue-scattering coefficients between 760 and 900 nm measured with time-resolved spectroscopy. Appl Opt 36(1):386–396
- 14. Arri SJ, Muehlemann T, Biallas M et al (2011) Precision of cerebral oxygenation and hemoglobin concentration measurements in neonates measured by near-infrared spectroscopy. J Biomed Opt 16(4):047005
- 15. Wolf M, Keel M, Dietz V et al (1999) The influence of a clear layer on near-infrared spectrophotometry measurements using a liquid neonatal head phantom. Phys Med Biol 44(7): 1743–1753