# **Molecular Pathology of Breast Cancer**

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#### **Introduction**

• Breast cancer is the most common cancer affecting women, with an estimated 250,000 new cases in 2011 in the US alone and 1.5 million worldwide. It is one of the first major diseases where basic laboratory research has had a large impact on the routine clinical management of patients, ranging from detection, to diagnosis, to therapy. Molecular approaches to pathology, in particular, have had an enormous influence, especially in the areas of diagnosis and therapeutic decision-making. The topic of molecular pathology in breast cancer is very large and evolving far too rapidly to cover completely in a chapter of this nature. This chapter will primarily focus on reviewing aspects that are already in routine clinical use, some of the more promising applications on the near horizon, and scientific questions that are currently at the forefront of translational research. From an etiological point of view, the molecular pathology of breast cancer is the result of molecular abnormalities occurring in important normal processes, including the gross, microscopic, and molecular anatomy of the breast, breast development, and adult physiology—which is where we begin

#### **Normal Characteristics of the Female Human Breast**

#### **Gross, Microscopic, and Molecular Anatomy**

- Grossly, the size of the adult female breast varies enormously. On average, it is about 10–12 cm in diameter, 5–8 cm in thickness, and weighs about 700 g. Weight may almost double during pregnancy and lactation. Pathologists typically divide the breast into four quadrants (Q): upper outer (UOQ), upper inner (UIQ), lower outer (LOQ), and lower inner (LIQ). Other important regions are the areola/nipple complex and the lymph nodes in axillary tail extending from the UOQ. Lymphatic (and vascular) drainage is important as the main pathway for breast cancer cells to metastasize. Most regions of the breast, especially the UOQ and LOQ, drain to the axillary nodes, although the LIQ and UIQ also drain to a chain of internal mammary nodes beneath the sternum and extending upwards
- Internally, the breast is composed of 15–20 segments or lobes, somewhat analogous to segments of an orange, but less well defined. Each lobe contains thousands of lobules, which are small grape-like clusters of glands lined by epithelial cells specialized to produce milk. Small ducts that join to form larger ducts that eventually exit through the nipple, transmitting milk to nourish our young, interconnect the lobules. All known precursors of breast cancer, also referred to as premalignant lesions, develop and progress from abnormal cells within the ductal system, primarily in the lobules and smallest ducts connected to them, referred to as the terminal duct lobular unit (TDLU)
- The entire normal ductal and lobular system is delineated from the mesenchymal stroma ("connective tissue") by a continuous basement membrane (BM) which is an important barrier which must be breached for cancer cells to invade and metastasize
- The lumens of the ducts and lobules are generally lined by two distinct layers of cells; an

outer layer directly on top of the BM referred to a myoepithelial cells (MECs), and an inner layer directly on top of the MECs referred to a luminal epithelial cells (LECs)—although LECs also have many subtle points of attachment with the BM interspersed with the MECs

- Nearly, all LECs typically express large amounts of keratin proteins, particularly CK8, CK18, and CK19. MECs express abundant CK5 and CK6, but are generally negative for keratins found in LECs, and they do not express ER or PR. MECs also typically express several other molecules distinct from LECs, including smooth muscle actin (SMA), calponin, S100, p63, CD10, and stratifin (SFN), which appear to be important in certain specialized normal functions such as contraction of duct lumens to expel milk, and to maintain normal cell polarity within ducts, which can actively suppress the invasion of cancer cells
- These keratins play an important role in a new molecular classification of breast cancersthe so-called intrinsic molecular subtypes, which is discussed in more detail later. Briefly, the most common subtype is referred to as "luminal" breast cancers, primarily because they have many similarities at the gene expression level with normal LECs, including these keratins. Another important subtype, referred to as "basal" breast cancers, expresses keratins normally associated with MECs, which historically have been referred to as "basal" cells because of their location in ducts and lobules. There is a common misconception that luminal and basal breast cancers evolve from genetically altered LECs and MECs, respectively, partly because of molecular similarities including keratins—which is probably not true, although the "stem" cell origin of all breast cancers is far from clear and a topic of much debate and research
- A proportion of LECs (20–30%) also express nuclear estrogen receptors (ER) and progesterone receptors (PR). ER and PR are important mediators of growth and differentiation stimulated by the hormones estrogen and progesterone. The majority of cancer cells also

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 **Fig. 6.1** Anatomy of the adult mature human breast. Correlation between compartments and different distinct pathologic processes arising in the breast

express these receptors, which may promote tumor growth

Recent studies have shown that histologically normal appearing breast epithelial cells are not always normal at the molecular level, and some of these morphologically silent biological abnormalities may predispose the cells to premalignant or malignant transformation. For example, chromosomal gains and losses have been observed in normal breast epithelium. Although the overall frequency of imbalances is quite low, it is significantly higher in normal cells adjacent to cancer cells than normal cells at a distance. Some of these genetic defects may be shared with the adjacent cancer, although the majority are not and appear to be random. Other studies have shown that breast tissue, especially in women at high risk for breast cancer, may contain patches of histologically normal appearing cells in which activity of the p16 tumor suppressor gene is suppressed. Compared to adjacent cells with normal p16 function, these cells show increased proliferation and elevated expression of cyclooxygenase 2 (COX2), and the latter appears to be associated with the development of many types of cancers. There are likely to be many other acquired and inherited molecular abnormalities in otherwise nor-mal appearing cells (Figs. 6.1 and [6.2](#page-3-0))

#### **Breast Development**

The molecular mechanisms responsible for human breast development are poorly understood because it is extremely difficult to study directly. Most of what we know is inferred

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 **Fig. 6.2** Breast histology. Differences between luminal epithelial cells (LEC) and myoepithelial cell layer (MEC) compartments. GATA 3 is a representative marker of LEC in both TDLU and TD. CK14 is a distinctive marker of MEC. Lower: Histological changes associated with lacta-

tion and menopause. During lactation, the acini are closely packed, with reduced amount of stroma; secretory material in the lumens is seen. With menopause, there is a marked reduction of acini and ducts, with replacement by fat

from animal studies, particularly involving genetically engineered mice, where the effect of altering specific genes on breast development can be directly observed. However, there are probably many important parallels in breast development among all mammals, and studies using mice and other models almost certainly reveal molecular mechanisms shared with humans. Many normal developmental mechanisms play a central role in the development and progression of breast cancers. For example, cells in the earliest potential precursors of breast cancer, referred to as hyperplasias, demonstrate suppression of molecular pathways involved in adult differentiation, and reactivation of embryonic pathways, which is also true of later stages such as the progression of ductal carcinoma in situ (DCIS) to invasive breast cancer (IBC)

• Mammary glands are derived from ectodermal buds or ingrowths along mammary lines in the embryo. Between 14 and 18 weeks of gestation, distinct mesenchymal and ductal compartments start to develop. By 28 weeks, there are two clearly defined cell compartments (LECs and MECs). The ductal and lobular system continues to develop and mature throughout the second half of gestation, as well as the areola and nipple. Many genes are known to play critical roles in regulating development. For example, BCL2, which suppresses apoptosis, increases dramatically beginning at about 18 weeks, and plays a important role in duct formation by inducing

Gene	<b>Disease</b>	Pathway	Clinical features
TBX3	Ulnar mammary syndrome	Linked to FGF pathway	Abnormalities in limbs and apocrine glands TBX3 overexpression linked to breast carcinomas
PTHR1	<b>Blomstrand</b> chondrodysplasia	Mutation in receptor or parathormone. (PTH) mediates cross-talk between epithelium and mesenchyme in early mammary bud	Dwarfism PTHrP is commonly secreted in breast cancers
Ectodysplasin	Hypohidrotic ectodermal dysplasias	Development of ectodermal appendages	X-linked ectodermal dysplasia receptor (which binds ectodyspla- sin) promoter methylation is linked to breast cancer
Ska (neuroregulin 3)	None known	Affects patterning of mammary glands, along the body axes. NRG-3 is a ligand to EGFR family	Upregulated in breast cancer, particularly those with HER2 overexpression
WNT		LEF1, the transcriptional mediator of WNT signaling at placode stage	LEF1-/- embryos placodes 2 and 3 do not form; the other placodes develop into small buds and degenerate. Corresponding ducts and nipples are missing in newborn

 **Table 6.1** Genes involved in breast development

Clinical consequences of mutations involving those genes and associated clinical syndromes

cells in the center of solid cords of primitive epithelial cells to die, forming patent lumens. Ductal budding and branching depends on prolactin which sensitizes cells to the growth stimulating effects of insulin. Aldosterone promotes differentiation of buds into ducts and lobules, forming primitive TDLUs. ER is expressed in LECs by third trimester and PR, 2–3 months after birth. Genetic alterations of these regulatory molecules can play important roles in the development and progression of breast cancer, in general, by promoting "embryonic" growth in an inappropriate setting. Other important genes are discussed later in the context of what happens to breast development when they are altered in transgenic and knockout mice (Tables 6.1 and 6.2)

• There are no structural or known molecular differences between male and female breasts during the postnatal period. At birth, nipple ducts finally open onto the surface. Closely after birth, prolactin, estrogen, and progesterone decrease dramatically, resulting in involution of newborn breast tissue. During this time, apocrine and cystic changes become prominent, which are also common in postmenopausal breasts. Between 2 years of age and puberty, the breasts are very small, and the main constituents are scattered small ducts embedded within a dense collagenous stroma. Pubertal changes are characterized by greatly increased growth of stroma, MECs, and LECs, which are prominently caused by increased levels of estrogen, although full differentiation requires other hormones and growth factors as well, including insulin, cortisol, thyroxin, prolactin, and growth hormone. ER is necessary for duct elongation, and ER knockout mice only develop rudimentary ducts without terminal end buds or alveolar buds. Interestingly, these glands are highly resistant to cancer development. PR is necessary for duct elongation and alveolar development, which are lacking in PR knockout mice. After menarche, prominent cyclical developmental changes occur with the menstrual cycle. Early on,

Gene (KO or			
overexpression)	Pathway	Clinical features	
<b>BRCA1 KO</b>	BRCA1 and p53	Increased mammary tumor development in BRCA1 KO that was p53 heterozygous (p53+/-) suggested that BRCA1 loss may induce tumor development due to genetic instability causing LOH LOH in p53 was seen in majority of BRCA1 KO mice	
$Era$ OE	ER	Mammary carcinomas with similarities to human breast cancer and $ER+$ phenotype	
Aromatase OE	Aromatase	Male mice developed gynecomastia, and homozygous mice were infertile and developed Leydig testicular cancers Females developed ductal hyperplasias and dysplasia. However, no mammary tumors were seen Mice exhibited increased $ER\alpha$ and $ER\beta$ levels, as well as PR, cyclin D1, and cyclin E levels (cyclin D1 overexpression correlates with ER+ phenotype in human cancers) DMBA treated mice with AO developed mammary tumors, whereas WT only showed hyperplasia. Letrozole effectively inhibited dysplastic growth in MMTV-aromatase mice	
$TGF\alpha/HER2$	$TGF\alpha$ and HER2	Double transgenic mice developed significantly less breast tumors than parental lines. Double transgenic mice with HER2 aromatase overex- pression show less hyperplasias	
$ER\alpha$ KO	ER	Mammary glands resembled prepubertal wild-type mice. $ER\alpha$ KO mammary epithelium underwent ductal morphogenesis when trans- planted to wild-type mice. Transgenic mice with MMTV-aromatase/ $ER\alpha$ KO did not develop hyperplastic growth and exhibited morphol- ogy similar to ER $\alpha$ KO mice. ER $\alpha$ mediated growth of the mammary duct network is a prerequisite for aromatase induced changes within the transgenic mammary gland	
WNT	WNT	Mice developed ductal hyperplasias early in life and mammary adenocarcinomas in most animals by 1 year of age. MMTV-wnt/ $ER\alpha$ $KO$ –/– exhibited stunted growth similar to parental $ER\alpha$ KO mice	
PELP-1	Coactivator of ER, PR, AR. Mediates G1-S transition. Aromatase pathway	MMTV-PELP1 developed mammary tumors in over 40% of cases. Tumors show ER and aromatase expression. Human breast cancers commonly show PELP1 overexpression and are associated with poor response to tamoxifen	
AIB1 KO/OE	Binds to steroid receptors and transcription factors	AIB-1 levels have been correlated with poor prognosis in breast cancer. Coinduction of AIB1 and HER2 was associated with decreased DFS and tamoxifen resistance. AIB1 KO resulted in decreased oncogenesis with decreased HRAS, HER2, and IGF1 expression. MMTV-AIB1 resulted in tumor development in 48% of mice. The carcinogenic potential was abrogated in double transgenic mice with MMTV-AIB1 $ER\alpha$ KO (ER is important in the AIB1 pathway). Induction of IGF1 signaling in the mammary gland is typical of the AIB1 transgenic model. Treatment with the mTOR inhibitor RAD001 resulted in block of hyperplasia and atypia in the AIB1 transgenic model	
CSF1	CSF1	The macrophage colony-stimulating factor, CSF1, is commonly elevated in breast cancer. CSF1 op/op is deficient in lactation and develops osteopetrosis. Cross-linked species of MMTV-PYMT CSF op/ op showed less progression of disease and lung metastases compared to the parental strain of MMTV-PYMT	

<span id="page-5-0"></span> **Table 6.2** Animal transgenic models in breast carcinogenesis

TDLUs develop more alveoli with each successive cycle. From menarche on, the mammary gland is fully anatomically and functionally developed to support pregnancy and lactation

- During pregnancy, proliferation of essentially all types of cells, especially LECs, dramatically increases, mediated by increasing levels of estrogen, progesterone, ER, and PR. After delivery circulating ER and PR decrease to low levels, in preparation for lactation. Once lactation begins, cell proliferation ceases as the cells terminally differentiate to produce milk. When lactation ceases, secretory LECs undergo apoptosis, alveoli collapse, and the mammary gland involutes back to the nonpregnant condition, although the ductal system postpregnancy retains a somewhat more complex ductal framework than prior to pregnancy. In the adult female breast, there is a relatively large reserve of normal stem cells which support the dynamic changes in growth and differentiation associated with menstrual cycling, pregnancy, and lactation. Presumably, various genetic alterations of normal stem cells may give rise to precancerous or cancer-stem cells, which eventually grow uncontrollably. However, there are probably other sources of cancer-stem cells, including dedifferentiation of mature LECs due to specific mutations
- After menopause, both lobules and ducts are decreased in number. Intralobular stroma is replaced by collagen and the breast stroma undergoes replacement by fat (Fig. [6.3](#page-7-0))

## **Molecular Biomarkers in Routine Clinical Practice**

#### **Estrogen Receptor and Progesterone Receptor: Molecular and Clinical Aspects**

The measurement of ER and PR has become a standard of practice in the evaluation of patients with primary breast cancer. The measurements can be performed accurately on formalin-fixed paraffin-embedded (FFPE) tissue by using immunohistochemistry (IHC) and the results have good correlation with those of biochemical testing

- The ER is the paradigm tumor marker for management of patients with cancer. It dates back to at least 1896 when G Batson reported regression of advanced breast cancer in women who underwent oophorectomy
- ER controls essential developmental and physiological processes. It interacts with the receptor as estradiol, regulates growth and differentiation, and helps maintain homeostasis. Studies have shown that dysregulation of ER and PR during development are important in carcinogenesis
- The effects and actions of estradiol are mediated through interaction with two nuclear receptor proteins,  $ER\alpha$  and  $ER\beta$ , located in chromosomes 6q and 14q, respectively, which are encoded by two separate genes *ESR1* and *ESR2*, respectively. Both,  $ER\alpha$  and  $ER\beta$  show substantial homology in the DNA binding domain. Role of  $ER\beta$  in breast cancer has not yet been determined. Hereafter,  $ER\alpha$  will simply be referred to as ER
- The "classical" function of ER involves binding of 17 B estradiol to ER located in the cell nucleus. This induces receptor dimerization, which binds to estrogen response elements (EREs) on many other genes, which are then indirectly regulated by estrogen and  $ER\alpha$
- ERE activated genes perform many important functions, including inhibition of apoptosis and stimulation of the cell cycle. There is cross talk with other mitogenic pathways (ras, raf, cyclin D1)
- Activation of estrogen target genes is accomplished through direct hormonal binding with the ER. This recruits protein regulators known as coactivators and repressors. Coregulators are responsible for chromatin remodeling to facilitate binding of RNA-polymerase. Histone acetylation, through acetyl transferases, correlates with a more actively transcribed state of chromatin regulation, whereas methylation favors more tightly coiled chromatin, which is less accessible to transcription and less gene expression

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**Fig. 6.3** Morphological stages in the embryonic development of the mammary gland in the mice: Around embryonic day 10 (E10) of mouse development, the milk line (*orange*) is defined by a slight thickening and stratification of the ectoderm (*gray*) as depicted here in this series of cross sections through the trunk. On E11.5, the milk line breaks up into individual placodes ( *orange* ) and the underlying mammary mesenchyme (*blue*) starts to condense. Over the following days, the placodes sink deeper into the dermis and the mammary mesenchyme becomes organized in concentric layers around the mammary bud (*orange*). Starting on E15.5, the mammary epithelium (*orange*) starts to proliferate at the tip and the primary sprouts pushes through the mammary mesenchyme towards the fat pad (*green*). On E18.5, the elongating duct has grown into the fat pad and has branched into a small ductal system. The cells of the mammary mesenchyme have formed the nipple, which is made of specialized epidermal cells (*purple*). *Lower*: The schematic diagram shows the position of the milk line, placodes, and mammary buds along the lateral body wall of early mouse embryos. Secreted molecules, receptors, and transcription factors that are important at the different stages are listed in the table below. At the mammary bud stage, proteins that are expressed in the epithelium and in the mesenchyme are listed separately. *BMP* bone morphogenic protein; *ERBB* erythroblastic leukemia viral oncogene homologue; *FGF* fibroblast growth factor; *FGFR1* fibroblast growth factor receptor; *GLI* Gli-Kruppel family member; *IGF* insulin growth factor; *IGFR* insulin growth factor receptor; *LEF* lymphoid enhancer-binding protein; *MSX* muscle segmentation homeobox; *NRG* neuroregulin; *PTHLH* parathyroid hormone-like hormone; *PTHR1* parathyroid hormone receptor; *TBX* T-box; *WNT* wingless-related MMTV integration site. (Reprinted with permission of Nature publishing group)

- ER status is highly predictive of clinical benefit from endocrine therapy in both adjuvant and metastatic disease settings. ER-positive tumors are more likely to respond to hormonal therapy, and have a better prognosis, when compared to ER− tumors
	- Harvey et al. showed in a cohort of 1,982 patients, using ligand binding assays (LBA) >3 fmol/mg and, retrospectively IHC (Allred Score >2 or 1–10% weakly positive cells), showed IHC to be a stronger predictor of disease-free survival (DFS) in patients receiving endocrine therapy when compared to LBA
	- Elledge et al., in a cohort of 205 patients, showed significant correlation of IHC ER and clinical response in patients with advanced metastatic disease (ER negative 25%, intermediate 46%, and high 66%)
- Accurate measurements of ER are of considerable importance, because it represents one of the strongest predictive factors of responsiveness to endocrine management. In some cases, endocrine therapy alone is an option, without additional cytotoxic therapy. About 70–80% of breast cancers are ER-positive and 20–30% are ER-negative. Only 70% of ER-positive tumors show clinical response to estrogen manipulation, but measuring ER expression alone is insufficient to distinguish responders from nonresponders. A significant fraction of patients with ER-positive disease eventually develop resistance to endocrine therapy
- Clinical progression of the ER-positive breast cancer typically correlates with hormone resistance. Loss of response and decreased ER expression are associated with a more aggressive clinical course. Epigenetic alterations of the ER promoter, including methylation of *ESR1* gene, are thought to be important events in the development of ER-negative breast cancers
- In the last decade, prospective randomized clinical trials have shown the superiority of aromatase inhibitors over tamoxifen in postmenopausal receptor-positive women
- Tamoxifen is a partial agonist (both antagonistic and agonistic effects) of the ER receptor,

and induces dimerization and nuclear translocation and is designated as a selective ER modulator (SERM)

- Fulvestrant directly binds to ER monomers, inhibits dimerization, and suppresses activation, thereby functioning as a pure antiestrogen. Its benefits have been demonstrated in the metastatic setting, and ongoing trials are underway in the adjuvant setting
- Anastrozole, letrozole, and exemestane are aromatase inhibitors (AI) which block the conversion of adrenally produced precursor compounds to estrogenic molecules. Recent trials also showed the benefits of estrogen deprivation persist for many years even after completion of the initial hormonal therapy in reducing both unilateral and contralateral breast cancers
- Recently, the Women's health Initiative Estrogen-Alone trial, analyzed, prospectively, the use of equine-conjugated estrogen (CEE) among patients with prior hysterectomy. The trial was stopped earlier and showed a decreased risk of breast cancer in the treatment group
- Progesterone has an essential role in regulating breast maturation. A clear role in carcinogenesis has been shown in animal models, particularly in respect to induction, maintenance, and progression of the neoplastic phenotype. An increased risk of breast cancer is documented in long-term users of progestinonly containing hormone-replacement therapy (HRT) regimens
- ER is important for regulating PR expression. Colocalization studies show that PR expressing cells also express ER. In fact, PR expression is regarded as a marker of an intact ER axis. However, discrepancies exist: the relative risk of disease recurrence is higher in patients with ER+/PR– cancers, compared to ER+/PR+ tumors
- About 60% of breast cancers express PR. This expression is regarded as a marker of intact ER function. PR receptor is also nuclear. Progesterone effects are mediated through the intracellular proteins PRA and PRB. Both are coded from the same gene using two distinct translation initiation sites
- Expression of PR in breast cancer is also associated with higher responsiveness to endocrine therapy. The majority of HER2-positive cancers are PR-negative, suggesting that nuclear  $ER\alpha$  may be nonfunctional in these cases. However, membrane ER appears to remain functional and promotes tumor cell proliferation in cooperation with overexpressed *HER2* . In this setting, tamoxifen (as a partial agonist) may theoretically help induce cell proliferation. In this setting, AI will remain beneficial. A role for highly quantitative assessment of PR might be helpful in more precisely predicting response in patients with ER-positive/HER2-positive tumors
- Most testing for ER and PR today is done using IHC. However, errors have been problematical when using IHC. For example, the United Kingdom National External Quality Assessment Service (UK NEQAS) evaluated the frequency of hormone-receptor-positive cancers in more than 7,000 patients, highlighting significant variation in ER and PR positivity rates. Similar results were obtained by the Royal College of Pathologists of Australasia  $(n=8,000 \text{ patients})$ . Approximately one-third of 1,023 ER tests performed on patients, in Canada, between 1997 and 2005 were scored falsely negative, which was revealed by retesting in an expert central laboratory in Ontario. More than 100 of these patients have since died and a class action lawsuit ensued claiming negligence in ER testing and failure to provide Tamoxifen to these patients. Investigation into the matter identified many causes of false negative IHC results, including: poor sample fixation, improper staining procedures, and improper interpretation:
	- The International Breast Cancer Study Group (IBCSG) conducted a series of studies comparing chemo and endocrine treatment to endocrine treatment alone in years before the establishment of IHC testing: Most studies of ER testing used LBA or ELISA. They compared with results obtained after the primary tumor blocks were collected and reanalyzed in a single central lab using IHC. Discordant ER

results between institutional and central results were 16% (ER+) and 24% (ER−) for specimens from premenopausal women, and 9% (ER+) and 24% (ER−) from postmenopausal women. Overall concordance rate was 82 and 88% for pre- and postmenopausal women, respectively

- In the ECOG 2197 trial, 11% of local ER− tests were scored positive on central testing, with an overall concordance rate of 90%
- In the ALTTO trial (5,000 patients from countries worldwide), so far, 4.3% of tumors that tested ER+ in local laboratories were found to be negative (false-positive) on central review. More than 20% of tumors exhibited at least some expression of ER (false-negative) on central review

### **Guidelines for Estrogen Receptor and Progesterone Receptor Testing by Immunohistochemistry**

- In an effort to improve the quality of testing for ER and PR by IHC, the American Society of Oncologists (ASCO) and College of American Pathologists (CAP) jointly developed and recently published guidelines for pathologists to follow (Fig.  $6.4$  and  $6.5$ ). Compliance with the guidelines is now mandatory for laboratories in the US to receive CAP accreditation
- Immunohistochemistry on FFPE tissue replaced LBAs for testing in the late 1980s. Harvey et al. compared the predictive abilities of LBA and IHC using the 6F11 antibody in a large cohort of patients with newly diagnosed breast cancer. This cohort received a variety of types of adjuvant therapy that ranged from none to endocrine alone, chemotherapy alone, or a combination of the above. Receptor status was scored as the sum of the proportion and average intensity scores of positive staining tumor cells (Allred Score on a scale ranging from 0 to 8). On the basis of clinical outcome in patients with adjuvant endocrine therapy, patients with Allred Score >3 (corresponding

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Total Score  $(TS) = PS + IS$  (range 0-8)

 **Fig. 6.4** Algorithm for scoring biomarkers (ER, PR) according to recent ASCO guidelines. Allred Score. A combination of number of cells (Proportion Score) and

intensity of staining (Intensity Score) is used. (Adapted from Allred et al.)



 **Fig. 6.5** Schematic example of ER interpretation

to as few as 1–10% positive cells) had a substantially and statistically significant better prognosis than patients with scores less than 3 (<1% positive cells). The predictive ability of IHC was superior to LBA previously performed in the same tumors

- There is no gold standard available for IHC assays of ER and PR. A relevant standard would be any assay whose specific preanalytic and analytic components conformed to assays whose results have been validated against clinical benefit from endocrine therapy (clinical validation). Several assays meet this criteria such as the methods described in the publication by Harvey et al. and Mohsin et al., the FDA 510(k)-cleared ER/PR pharmDx assay kit (Dako, Glostrup, Denmark)
- ER status can also be determined at the RNA level. The Oncotype  $DX^{\circledast}$  Assay measures RNA expression of 21 genes to determine a recurrence score (RS). ER and PR are among the most prevalent genes in the signature. Comparison between measures of the ER/PR protein by IHC and of mRNA by RT-PCR showed a discordance rate of 9% and 12%, respectively. There are no published correlations of the individual measures of ER and PR mRNA from the 21-gene signature with the clinical outcome
- A laboratory that performs ER testing should validate its proposed or existing assay against one of the clinically validated assays and demonstrate acceptable concordance. To be considered acceptable, the results of the assay must be initially 90% concordant with those of the clinically validated assay for the ER-positive and PR-positive categories, and 95% concordant for the ER-negative or PR-negative categories
- The cutoff from distinguishing a "positive" from "negative" cases should be  $\geq 1\%$  ER+ positive tumor cells. Patients whose breast tumors show at least 1% ER+ cells are candidates for endocrine therapy and those with less are not. Percentage of stained tumor cells provides valuable predictive and prognostic information to inform treatment strategies
- Eight studies described the relationship between hormone-receptor levels and patient outcomes. Overall survival, DFS, recurrence/ relapse-free survival, 5-year survival, time to treatment failure, response to endocrine therapy, and time to recurrence were positively related to ER levels
- PR status provides additional predictive value independent of ER values, especially among premenopausal women. Its predictive value has been demonstrated in retrospective studies using 1% as cutoff point. Among patients who received adjuvant endocrine therapy, the best cutoff for both DFS  $(P=0.0021)$  and OS  $(P=0.0014)$  was a total PR Allred Score >2, which corresponds to greater than 1% of carcinoma cells exhibiting weakly positive staining. In patients with metastatic breast cancer who received first-line endocrine therapy on relapse, a correlation with PR status and response to endocrine therapy was found at a 1% staining threshold  $(P=0.044)$  or response to tamoxifen at  $10\%$  ( $P = 0.021$ ). Patients with carcinomas >1% PR staining had a better survival after relapse  $(P=0.0008)$
- Reporting results for ER, PR, and HER2: The percentage and proportion of tumor cells staining positively should be recorded and reported. All tumor areas of the tissue section on the slide should be evaluated. This can be achieved manually by counting cells or through image analysis
- The intensity of the staining should be recorded and reported as weak, moderate, or strong. This measurement should represent an estimate of the average staining of the intensity of the positively stained tumor cells on the entire section relative to the intensity of the positive controls run on the same batch. A cutoff of a minimum of 1% of the tumor cells positive for ER/PR for a specimen is considered to be positive. The term equivocal must not be used
- Less than 1% of the tumor cells positive for ER/PR for a specimen is considered to be negative. Such patients do not receive meaningful benefit from endocrine therapy
- Any specimen lacking intrinsic elements (normal breast epithelium) that is negative on ER and/or PR assay should be repeated using another tumor block or another specimen, and reported as not interpretable rather than as negative
- "Not interpretable" receptor results refer to samples that did not conform to preanalytic specifications of the guidelines, were processed using procedures that did not conform to guideline specifications of the lab operating procedures, or the assay used to analyze the specimen was not validated and controlled as specific in the guideline. Examples of circumstances leading to not interpretable results include testing of needle biopsies or cytology samples fixed in alcohol, use of fixatives other than 10% NBF, biopsies fixed for intervals shorter than 6 h or longer than 72 h, samples where fixation was delayed more than 1 h, samples with prior decalcification, and samples without internal or external controls
- Negative ER and PR interpretations in tumors that characteristically have an ER+ phenotype (e.g., lobular, tubular, and mucinous carcinomas) should be confirmed by retesting
- ER and PR should be documented in all newly diagnosed breast cancers. Recurrences should also always be tested to exclude prior false negatives, and to document changes in biologic behavior. In the routine practice, DCIS is also commonly tested for ER and PR based on the NSABP-24 clinical trials. The trial compared placebo versus tamoxifen after lumpectomy and radiation. There was a significant reduction  $(40-50\%)$  in subsequent breast cancer (ipsilateral and contralateral) restricted to patients with DCIS ER+ at 10 years followup

### **Human Epidermal Growth Factor Receptor 2 Gene: Molecular and Clinical Aspects**

• The human epidermal growth factor receptor 2 gene, more commonly referred to as *HER2* , is amplified in 15–25% of human breast cancers. *HER2* amplification and overexpression are highly correlated, which are significantly associated with aggressive disease (i.e., poor prognostic factors), and are the molecular targets for specific therapies, such as trastuzumab

- *HER2* is a protooncogene located on chromosome 17. It encodes a tyrosine–kinase receptor residing in the surface membrane of breast epithelial cells. It forms complexes with similar proteins (erbB1, erbB3, and erbB4) and acts as receptors for several ligands, such as EGF, heregulin, and amphiregulin. It regulates many normal cell functions, including proliferation, survival, and apoptosis
- The overall relationship between HER2 and clinical outcome is complex and varies with the clinical setting. A weak but significant association between poor outcome and a positive HER2 (overexpression or amplification) in patients receiving no additional therapy after initial surgery is seen. But this only represents a small fraction of patients today. The majority of patients typically receive some form of adjuvant treatment. Some studies have shown that HER2+ breast cancers are resistant to certain types of cytotoxic chemotherapy (e.g., the combination of cyclophosphamide, methotrexate, and 5-fluorouracil) but sensitive to others (e.g., anthracyclines and taxanes). In general, it is accepted that HER2+ cancers appear to be associated with relative, but no absolute, resistance to endocrine therapies in general. However, this issue remains very controversial. The most promising and useful findings are based on recent studies showing that HER2+ cancers respond favorably to new antibody-based therapies, targeting specifically the HER2 protein, such as trastuzumab. Although this therapy was originally demonstrated effective in patients with metastatic disease, more recent clinical trials have shown significant benefits in the adjuvant setting for patients with less advanced disease. The NSABP-B31 clinical trial, which randomized patients with HER2+ cancer to adjuvant chemotherapy +/− trastuzumab, showed a 52% improvement in disease-free survival with the monoclonal antibody
- A long and persistent controversy in the evaluation of the HER2 status by protein expression through IHC, or gene amplification by FISH exists. However, many studies have shown that, when properly performed, a very strong correlation between the two methods exists, and they are equivalent and complimentary in the clinical practice
	- Owens et al. observed a similar frequency of HER2 amplified cases by IHC  $(20\%)$ among 116,736 specimens and FISH (22%) among 6,556 specimens
	- Most clinical trials using trastuzumab enroll patients with IHC positive, or reflex FISH positive, or ISH alone
- In general, approximately 70% of breast cancers show little or no protein overexpression, a normal gene copy number, and do not respond to trastuzumab. Roughly 15% show low to intermediate levels of protein expression, and the gene is amplified in nearly a third of those cases. There is still uncertainty of how well these patients respond to the drug. The remaining 15% of cases show very strong membrane staining, indicating high levels of protein expression and the gene is nearly always amplified. This is the population who shows best response to trastuzumab

### **Guidelines for HER2 Testing in Breast Cancer**

- ASCO and the CAP jointly developed and published guidelines to improve the quality of HER2 testing (Fig. 6.6)
- A positive HER2 test is defined as a result of 3+ surface protein expression (formed as uniform intense membrane staining of >30% of invasive tumor cells) or FISH result of amplified *HER2* gene copy number (average of >6 copies/nucleus for test systems without internal control probe) or *HER2/CEP17* ratio of more than 2.2, where CEP17 is a centromeric probe for chromosome 17 on which the HER2 gene resides
- Originally, FISH testing results were reported as either positive or negative, but an interme-

diate range (referred as equivocal range) has since been described and its clinical significance remains unclear. Much of the confusion using this term comes from the need to define the need for trastuzumab treatment. There is also significant variation in the intermediate (equivocal) ranges for both the IHC and FISH assays. The equivocal range for IHC consists of samples scored 2+, which includes up to 15% of samples. An equivocal result (2+) is complete membrane staining that is either nonuniform or weak in intensity but with obvious circumferential distribution in at least 10% of cells. Some, but not all of these samples may have *HER2* gene amplification and require additional testing to define the true HER2 status. The equivocal range for FISH assays is defined as  $HER2/$ *CEP17* ratios from 1.8 to 2.2 or average gene copy numbers between 4.0 and 6.0 for systems without an internal control probe. About 3% of patients have ratios of 2.0–2.2 and were previously included in treatment arms with trastuzumab. Polysomy 17 is a vague term, seen in up to 8% of tumors. If polysomy 17 is defined as three or more copies of CEP17, most are not associated with protein or mRNA overexpression

- Discordant results (IHC3+/FISH– or IHC<3+/ FISH+) have been documented in approximately  $4\%$  of cases. The significance of this is unclear. Equivocal results of a single test require additional action, which should be specified in the report. Equivocal results by IHC should follow confirmatory FISH analysis. Counting additional cells or repeating the test confirms equivocal FISH results. If the results remain equivocal, confirmatory IHC is recommended
- A negative HER2 test is defined as either an IHC result of 0 or 1+ for cellular membrane protein expression (no staining or weak, incomplete membrane staining in any proportion of tumor cells), or a FISH result showing *HER2/CEP17* ratio of less than 1.8 or an average of fewer than four copies of *HER2* gene per nucleus for systems without an internal control probe

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**Fig. 6.6** Algorithm for scoring HER2 according to recent ASCO guidelines

• The ASCO/CAP guidelines establish that, in order to classify a test as positive or negative, the laboratory must have performed concordance testing with a validated FISH assay and confirmed that only  $5\%$  or less of samples classified as either + or  $-$  disagree with the validated assay on an ongoing basis. Equivocal cases are not expected to be 95% concordant, but rather subjected to a confirmatory test

## **Recent Advances in the Molecular Pathology of Breast Cancer of Clinical Signi fi cance**

#### **Multigene Prognostic Indices**

• Oncotype  $DX^{\circ}$  is a prognostic test measuring the RNA expression of 21 genes, which provides a recurrence score (RS; range 0–100) using FFPE tumor samples. The genes include proliferation markers ( *Ki67, survivin, cyclin* 

*D1* ), invasion-related ( *MMP11, cathepsin* ), *HER2*, ER, PR, and others (*GSTM1*, *CD68*, *BCL2*), as well as five housekeeping genes used to normalize expression overall. The RS quantifies the likelihood of disease recurrence based on studies in women with early stage hormone estrogen receptor (ER) positive only breast cancer, and assesses the likely benefit from certain types of chemotherapy. Scores are reported as: low (<18), intermediate (18– 31), or high (>31) relative to risk of recurrence. Typically, patients in the high risk receive chemotherapy and those in the low risk do not. Studies have demonstrated that treatment is modified in  $31\%$  of patients who are tested by Oncotype  $DX^{\circledast}$ , including omission of presumed unnecessary chemotherapy in  $22\%$ . Based on these findings, it is estimated that the cost of gene expression against the relative costs of ER, PR, and HER2 are likely to result in an overall cost saving, as well as reduced toxicity and quality of life

 improvements for patients. Recently, the test has also shown similar prognostic and predictive significance in women with receptor-positive node-positive received adjuvant treatment with the aromatase inhibitor anastrozole, and in cancer patients receiving neoadjuvant hormonal therapy and chemotherapy. There is an important ongoing phase III clinical trial, referred to as the TAILORx study, designed to help optimize the use of adjuvant endocrine and chemotherapy in patients with receptorpositive breast cancer. Based on their recurrence score, women will be assigned to three different treatment groups: women with a recurrence score higher than 25 will receive chemotherapy plus hormonal therapy (the standard of care); women with a recurrence score lower than 11 will receive hormonal therapy alone; and women with a recurrence score of 11–25 will be randomly assigned to receive adjuvant hormonal therapy, with or without chemotherapy. The study is primarily designed to evaluate the effect of chemotherapy on those with a recurrence score of 11–25. Because the degree of benefit of chemotherapy for women with recurrence scores between 11 and 25 is uncertain, strong preliminary evidence suggests that may only require endocrine therapy, which would be an important **benefit** 

The Mammaprint<sup>®</sup>: 70-gene prognostic index was validated as clinically useful in studies of younger women with node-negative breast cancer by classifying them into low risk and high risk for disease recurrence. It requires frozen tumor samples. Genes involved in the regulation of cell cycle, invasion, and angiogenesis heavily weight it. Genes of interest do not include known prognostic markers such as ER, PR, and HER2. High risk patients are most likely to benefit from cytotoxic chemotherapy. In contrast, the low risk group typically responds very well to endocrine therapy without chemotherapy. The prospective validation of the MammaPrint<sup>®</sup> signature's prognostic value is currently ongoing through the Microarray in Node-Negative Disease May Avoid Chemotherapy

(MINDACT) trial. This trial opened in February 2007 as has enrolled over 6,000 patients from five European countries. It assesses all patients by the standard clinicopathologic prognostic factors included in adjuvant setting and by the 70-gene signature assay. If both traditional and molecular assays predict a high risk status, the patient receives adjuvant cytotoxic chemotherapy and also hormonal therapy if ER positive. If both assays indicate a low risk, no chemotherapy is given and ER-positive patients are given adjuvant hormonal therapy only. When there is discordance between the traditional clinicopathologic prognostic factor prediction of risk and the 70-gene signature prediction of risk, the patients are randomized to receive treatment based on either the genomic or the clinical prediction results. The primary goal of the study is to confirm that breast cancer patients with a "low risk" molecular prognosis by MammaPrint<sup>®</sup> and "high risk" clinical prognosis can be safely spared chemotherapy without affecting distant metastases-free survival (DMFS)

PAM50 assay: was developed to efficiently determine intrinsic molecular subtypes based on evaluating 50 carefully selected genes using next generation sequencing and FFPE tissue samples. It is currently performed in a commercial reference laboratory, but an instrument dedicated to perform this will be available to pathology laboratories in the future. The PAM50 test provides a risk of relapse score (ROR) initially based on studies of patients with node-negative breast cancer who did not receive adjuvant systemic therapy. The ability of ROR to predict prognosis has recently been confirmed as useful in an independent set of 786 patients with ER+ treated only with tamoxifen. In these studies, ROR was a better predictor than standard clinicopathologic variables, including Ki67, PR, and histological grade. Most recently, PAM50 outperformed Oncotype $DX^{\circ}$  for predicting response to endocrine therapy in a large prospective clinical trial of receptor-positive node-negative patients



Fig. 6.7 Taxonomy of breast cancer. WHO classification of common histologic subtypes

- MapQuant  $Dx^{\circledast}$  genomic grade: is a predictor test derived by identifying 97 differentially expressed genes from grade 1 and 3 breast cancers using a training set of 64 ER+ tumors. Most genes are cell cycle regulators and proliferation. Genomic grade index (GGI) was strongly associated with risk of recurrence among patients with grade 2 tumors. It requires fresh tissue, similar to Mammaprint
- Breast cancer index (BCI): provides assessment of likelihood of distant recurrence in patients with ER+, node-negative breast cancer treated with endocrine therapy (primarily tamoxifen). BCI was developed from a combination of two indices: HOXB13:IL17BR and a proliferation related five-gene molecular grade index. Technically, it involves using a qRT-PCR assay with FFPE tissue samples
- The clinical use of Mammaprint<sup>®</sup>, Oncotype  $DX^{\circ}$ , BCI, PAM50 assays have all been proven most useful in studies of patients with receptor-positive node-negative breast cancer, which are highly enriched with luminal A molecular subtypes, which may explain why the prognostic ability of these different gene expression-based assays is similar, as most of them are differentiating luminal A from all other subtypes (Figs. 6.7 and [6.8 \)](#page-17-0)

#### **Intrinsic Molecular Subtypes of Breast Cancer**

• Understanding the more recent advances in the molecular biology of breast carcinogenesis, imply acknowledging the major contribution of

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 **Fig. 6.8** Breast cancer pathology, common histologic subtypes. Grading invasive carcinomas depend on the degree of tubular formation, nuclear features, and mitotic index. Invasive ductal carcinoma of no special type. 8a-Grade 1; 8b-Grade 2; 8c-Grade 3. 8d-8i, common

histologic types. 8d and 8e: invasive lobular carcinoma; 8f: invasive tubular carcinoma; 8g: invasive mucinous carcinoma; 8h: invasive medullary carcinoma; 8i: invasive ductal carcinoma with mucinous features

Perou et al. in the description of the molecular intrinsic subtypes of breast carcinomas. This work represents the first molecular classification of tumors, not considering the histology but a description of gene expression profiles of different breast tumors

- Four molecular subtypes were originally described: luminal, normal breast-like, HER2, and basal like. Subsequently luminals were further subdivided into Luminal A and Luminal B
- • Luminal tumors are reminiscent of "normal luminal epithelial cells," including CK8/18+. Lum A are ER+ and enriched with genes associated with active ER pathway, low levels of proliferation related genes, low histological grade, and generally good prognosis. The Lum

B tumors are typically higher grade, with high proliferation indexes, and worse outcome, and a significant proportion are HER2+. Recent data show no good separation between Lum A and Lum B based on proliferation

- The normal breast-like subtype has gene expression profiles similar to fibroadenomas and normal breast enriched in adipose tissue genes. They are relatively poorly characterized and their prognostic significance is unclear. Recent studies suggest that the normal breastlike group may be an artifact caused by contamination of samples with normal tissue
- The HER2+ subtype shows amplification or 3+ reactivity by IHC, and expresses many other genes associated with the HER2 pathway.

However, a good number of *HER2* amplified, ER+ cancers fall into Lum B category

- The basal subtype expresses genes found in normal basal/MECs of breast, such as *CK5, CK14, p-cadherin, caveolins 1–2, CD44* , and *EGFR*. A minority has *EGFR* amplification. However, unlike MECs, they also express certain proteins characteristic of LECs, such as CK8, CK18, and KIT. Basal-like carcinomas are usually high histological grade tumors with high proliferation, necrosis, pushing borders, and lymphocytic infiltrate. Histological subtypes commonly seen in this category include medullary or metaplastic carcinomas. The basal-like subtype more commonly happen in younger individuals, often of African– American or Hispanic decent. The tumors usually show high initial response to cytotoxic chemotherapy, although the majority relapses and overall prognosis is very poor. These features are similar to those seen in tumors of patients with *BRCA1* mutation and the BRCA1 pathway is dysfunctional in basal-like cancers
- Three new ER-negative molecular subtypes have recently been described: One, referred to as "Molecular apocrine," is similar to HER2 subtype but shows activation of androgen receptor signaling; another, referred to "Interferon subtype," are characterized STAT1; and the third are referred to as the "claudinlow" group, which typically demonstrate a cancer-stem cell like phenotype
- Recently, several studies have questioned whether intrinsic subtyping is reproducible or stable, and whether it has any useful clinical significance
- The relationship of intrinsic molecular subtypes to special histological subtypes of breast cancer: Some studies, mainly using microarray-based technology, have shown that at the transcriptional level, tubular, mucinous, and lobular subtypes are more homogeneous than invasive ductal carcinomas of no special type (IDC/NST). Tubular, mucinous, and neuroendocrine carcinomas are typically included in the luminal phenotype. Adenocystic, medullary, and metaplastic are basal-like in agreement with previous studies
- The use of IHC has recently been advocated as a surrogate to microarray analysis to define the intrinsic molecular subtypes  $(Fig. 6.11)$ : Expression by IHC of ER, PR, and luminal CKs (CK8 and CK18), lack of HER2 overexpression, and low Ki67 are typical of Lum A. Expression of ER, PR, and luminal CKs, and HER2 overexpression are seen in Lum B. Absence of ER and PR, and HER2, and expression of basal CKs  $(CK5/6)$  define basal-like tumors
- In the neoadjuvant settings, pathologic complete response (pCR) has been used to determine response to chemotherapy. pCR is only seen in 20–30% of patients (with use of standard anthracycline and taxane-based chemotherapy): Different rates have been shown across the different molecular subtypes: rates are 7% for Lum A, 17% for Lum B, 36% for HER2, and 43% for basal-like. This is one of the few scenarios where the use of molecular subtypes is advocated to translate into clinical practice. It is important to understand that molecular subtypes do not add much additional information of prognostic significance compared to the current standards of histologic subtypes and pathologic grading
- Even though the molecular classification has been one of the greatest advances in breast cancer in the last two decades, differences in molecular aspects of common histologic subtypes have been also recognized. Here are some examples: Medullary carcinomas show a prominent T helper cell immune response. Adenoid cystic carcinomas of the breast show a characteristic translocation  $t(6,9)$ , which creates a *MYB–NFIB* fusion transcript. Secretory carcinomas also have an associated translocation,  $t(12;15)$  with the conformation of a *ETV6–NTRK3* fusion transcript. Micropapillary carcinomas have a high rate of lymph node metastasis and are typically included in the luminal B subtype, but a distinct set of gene clusters on their own, including high rate *FGFR1* amplification. Metaplastic breast cancers are a mixture of adenocarcinoma with metaplastic elements, homologous (squamous and spindle metaplasia) or heterologous (chondroid, osteoid, skeletal muscle). They

are typically associated with *PI3K/AKT* mutations—over 90% are HER2 and ER negative, and typically show a basal-like immunophenotype. A dysfunctional BRCA1 pathway is seen with over 60% of metaplastic carcinomas, which is caused by methylation silencing of the *BRCA1* gene promoter. In addition, a mouse model with *BRCA1* inactivation and wild-type allele of *TP53* show classical morphologic features of metaplastic carcinomas, including HER2 and basal markers (CK14 and EGFR), as well as activation of WNT pathway (Figs.  $6.9, 6.10,$  $6.9, 6.10,$  and  $6.11$ )

#### **Important Somatic Mutations in Breast Cancer**

• *TP53* is mutated in up to 30% of sporadic breast cancers, as well as many other types of cancers. The gene is located on chr 17 and encodes a nuclear transcription factor normally involved in cellular pathways activated in response to stress by inhibiting the proliferation, and inducing apoptosis, of cell damaged in a variety of ways. P53 acts as a transcriptional activator of genes involved in inhibition of the cell cycle, blood vessel formation, stimulation of apoptosis, and promotion of DNA repair. Currently, 2,500 different inactivating *TP53* mutations have been described in breast cancer. About 75% are single nucleotide substitutions leading to substitution of a single amino acid, and the remaining 25% are insertions, deletions, and nonsense mutations. Mutations in one allele are associated with inactivation of the other one by loss of heterozygosity (LOH) in most affected breast cancers. Mutation of the gene often correlates with increased nuclear p53 expression by IHC, which can be used as an easy surrogate assay in certain situations. Somatic mutations of *TP53* occur in IBCs and DCIS. In both settings, they are associated with increased tumor size and grade, as well as axillary metastasis and the rate of *TP53* mutations is very high in *BRCA1/BRAC2* carriers. The presence of *TP53* mutations is associated with poor prognosis: shorter DFS and OS in both node-negative and node-positive cancers. However, one study has shown an advantage in survival in node-negative breast cancer with mutated *TP53* treated with XRT compared to node-negative with WT *TP53*

- *ESR1* mutations:  $ER\alpha$  has been reported as mutated and amplified in low percentage of breast cancers. Those with an ER A86V mutation are associated with lower activity of the receptor. The ER K303R mutation makes the receptor hypersensitive to activation by estrogen, which may promote tumor progression. An ER 437 stop codon mutation has been identified in metastatic breast cancers, and may be important in promoting metastatic spread, although the mutation is very rare
- Gene copy number alterations (referred to as allelic imbalance): AI is very common in breast cancers, occurring in as many as 50%. Gene amplification is a pathologic change commonly associated with increased mRNA transcription and protein expression of affected genes. Gene deletions are associated with loss of expression and function. Amplification of several regions in the breast cancer genome contains genes coding for oncogenes. For example, the chromosome 17q12 amplicon contains the *HER2* gene, the 8p24 amplicon the *MYC* gene, the 11q13 amplicon the *CCND1* gene, and the 6p11 the *ESR1* gene
	- Amplification of *HER2* is common in breast cancer and was discussed in detail above
	- Amplification of *ESR1* on chromosome 6p occurs in 5–20% of breast cancer, it is associated with increased ER expression, and it appears to increase responsiveness to tamoxifen therapy—so determining this feature may help optimize the use of endocrine therapy
	- 8q24 *MYC* on chromosome 8q24 is frequently amplified. MYC regulates cell growth and proliferation, and amplification is associated with higher histological grade, high proliferation rate, early recurrence, and death. Coamplification of MYC and *HER2* is very common, and trastuzumab is

<span id="page-20-0"></span>Gene expression patterns of 85 experimental samples representing 78 carcinomas, three benign tumors, and four normal tissues, analyzed by hierarchical clustering using the 476 cDNA intrinsic clone set.



 **Fig. 6.9** Breast cancer molecular intrinsic subtypes. Gene expression patterns of 85 experimental samples representing 78 carcinomas, 3 benign tumors, and 4 normal tissues, analyzed by hierarchical clustering using the 476 cDNA intrinsic clone set. (a) The tumor specimens were divided into five (or six) subtypes based on differences in gene expression. The cluster dendrogram showing the five (or six) subtypes of tumors are colored as: luminal subtype A, *dark blue*; luminal subtype B, *yellow*;

luminal subtype C, *light blue*; normal breast-like, *green*; basal-like, *red*; and ERBB2+, *pink*. (b) The full cluster diagram scaled down. The *colored bars* on the right represent the *inserts* presented in **c–g**. (**c**) ERBB2 amplicon cluster. (d) Novel unknown cluster. (e) Basal epithelial cell-enriched cluster. (f) Normal breast-like cluster. ( **g** ) Luminal epithelial gene cluster containing ER. (Copyright 2001 National Academy of Sciences, USA, with permission)

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 **Fig. 6.10** Overall and relapse-free survival analysis of 49 breast cancer patients, uniformly treated in a prospective study, based on different gene expression classification. Overall and relapse-free survival analysis of the 49 breast cancer patients, uniformly treated in a prospective study, based on different gene expression classification. (a) Overall survival and (**b**) relapse-free survival for the five

expression-based tumor subtypes based on the classification presented in Fig.  $6.9$  (luminals B and C were considered one group). (c) Overall survival estimated for the six-subtype classification with the three different lumi-nal subtypes presented in Fig. [6.1](#page-2-0). (d) Overall survival based on the five-subtype classification. (Copyright 2001 National Academy of Sciences, USA, with permission)

associated with improved outcome when coamplification exists compared to tumors with amplified *HER2* alone

– *CCND1* on chromosome 11q13 encodes a cell cycle regulatory protein that plays an important role in normal mammary gland development. The amplification is seen in up to 20% of breast cancers, which is significantly higher in lobular and with ER+/PR+ tumors. Coamplification of *MYC* and *CCND*1 occurs and is associated with aggressive phenotype. Coamplification with *FGFR1* has also been reported and is associated with worse outcome

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Fig. 6.11 Use of IHC in determination of molecular intrinsic subtypes. Representative cases for each molecular subtype. Hematoxylin and eosin and immunohistochemical stains of estrogen receptor, HER2, CK5/6,

and epidermal growth factor receptor for luminal A ( **a–e** ), luminal B ( **f–j** ), HER2 ( **k–o** ), basal ( **p–t** ), and unclassified (**u–y**). (Adapted from Tang et al., 2009)

- 8p11.3 *FGFR1* on chromosome 8p11.3 is amplified in about 10% of breast cancers, and is associated with poor clinical outcome. Typically, it is associated with an ER+, PR+, and HER2− phenotype. In addition, *FGFR1* amplification is associated with resistance to endocrine therapy. *FGFR1* inhibitors have shown clinical response in patients with metastatic breast cancer, as an adjuvant to chemotherapy
- *MDM2* amplification has been reported in breast cancer and is associated with worse outcome in patients with node-negative disease
- Complex amplicons, as commonly observed with *HER2* on 17q22 (HER2) and *FGFR1* on8p11.3, typically involve a large

number of adjacent genes that might also be important in the pathogenesis of breast cancer. For example, *TOP2A, RARA,* and *PPARB*. Coamplification with *TOP2A* is associated with responsiveness to anthracycline chemotherapy

#### **Hereditary Breast Cancer BRCA1 and BRCA2**

• Hereditary breast cancer (HBC) means that an alteration in a single major gene strongly contributes to the development of cancer or cancer-related conditions within the family. HBC was brought first to the medical literature by the surgeon Paul Broca, who accounted for his wife pedigree in 1865 showing four generations of breast cancer and occurrences of cancer of the GI tract. In 1990, Hall et al. described a linkage specific site of breast cancer on chromosome 17q. *BRCA1* gene was later cloned. Subsequently, a second gene located in chromosome 13q was cloned, *BRCA2. BRCA1* and *BRCA2* are the major well characterized genes contributing to HBC, but others are known (but very rare), but it is likely that there are more yet to be discovered. In general, HBC is characterized by a significant earlier onset of breast cancer (average, 45, beginning at the age of 20), an excess of bilateralism, a greater frequency of multiple primary cancers (such as breast and ovary), and an autosomal dominant pattern of inheritance. In females, about 45% of HBC and 80% of hereditary breast and ovarian cancers are associated with *BRCA1* mutations. Most of the remaining HBCs are attributable to *BRCA2* mutations. The lifetime risk of breast cancer in *BRCA1* and *BRCA2* mutation carriers is about 85%. The risk of ovarian cancer is 40–60% for *BRCA1* and 15% for *BRCA2* . It is estimated that about two-thirds of male breast cancer are linked to *BRCA2* , and one-third to *BRCA1* mutations. Overall, the prognosis of *BRCA1/2* mutated population appears to be similar to non-BRCA patients, although there is still controversy on this issue. For example, Ashkenazi Jews with *BRCA1/2* mutations appear to have relatively poor outcomes. Some new studies suggest that *BRCA1* patients may even have better survival than matched non-BRCA patients, and that *BRCA2* prognosis is worse

• *BRCA1* : 1,643 mutations have been described, of which 890 have been reported only once. For *BRCA2* approximately 1,856 mutations have been identified. BRCA shows two variants of penetrance, high (84% by 70 years of age) and low (32% by 70 years). Phenotypically, most *BRCA1* mutated tumors are basal-like breast cancers: highly proliferative, poorly differentiated, and genomically unstable. Most studies find *BRCA1* HBC to have a triple negative phenotype (ER−/PR–/HER2−). They are also associated with higher histological grade. A much higher prevalence of typical and atypical medullary carcinomas is also observed compared to sporadic breast cancers (35.3 vs. 3.4% for age matched controls and *BRCA1* mutated cancers). A lower prevalence of low grade tumors is seen in *BRCA1* mutated cancers compared to sporadic cancers, including ILC, tubulolobular, tubular, and invasive cribriform types. Indeed ILC commonly lack alterations at the *BRCA1* site. Aneuploidy is common among *BRCA1* mutated tumors. The frequency *of TP53* mutations is increased in *BRCA1* tumors compared to non-HBC and *BRCA2* tumors. Tamoxifen has been shown to be beneficial in reducing the risk of contralateral breast cancer in *BRCA1* patients, suggesting that they evolve from ER-positive precursors

• *BRCA2* mutated cancers have a more variable phenotypes than *BRCA1*, including a much higher proportion of luminal subtypes, and a much proportion of basal subtypes. Most studies show that the age of onset is older than in *BRCA1* . Some studies have shown higher prevalence of ILC associated with *BRCA2* than *BRCA1* . *BRCA2* also tend to show lesser aneuploidy and S phase. In *BRCA2*, ER/PR expression appears to be similar to non-BRCA cancer—a single study has even shown higher levels. Mutations of the *BRCA2* gene are also linked to other types of cancer, including pancreatic, prostate, and melanoma

#### **Hereditary Breast Cancer Non-BRCA**

- Non-BRCA HBC represents approximately 50% of cases in the general population. Overall, their clinical pathological features are statistically similar to sporadic breast cancer patients overall, including histological subtypes and grade, proliferation, p53 status, and intrinsic subtypes
- Germline mutations of *CDH1* (E-cadherin), which are very rare, confer a 40–70% lifetime risk of hereditary diffuse gastric carcinoma, and a 39–52% of ILC. E-cadherin is an adhe-

sion protein, which is lost in sporadic ILC through somatic mutations

- Li–Fraumeni syndrome: Lynch et al. described an extended kindred with a broad spectrum of cancers: sarcoma, breast cancer and brain tumors, lung and laryngeal cancers, leukemia, lymphoma, and adrenocortical carcinomas (SBLA syndrome). It is caused by a *TP53* germline mutation. The penetrance is variable with two age specific models: one in childhood and the second in adult life
- Cowden syndrome is a cancer associated genodermatosis, also referred as multiple hamartoma syndrome. It has an autosomal dominant pattern of inheritance, and is associated with distinctive mucocutaneous lesions and cancer of the breast, thyroid, and female genitourinary tract
- Germline mutations of the *PTEN* gene (also seen in Bannayan–Riley–Ruvalcaba syndrome). Cutaneous manifestations include trichilemmomas, which are pathognomonic. Also, multiple facial papules, acral and palmoplantar keratosis, skin tags and lipomas. Merkel cell carcinoma can occur. Thirty percent of women show breast carcinomas, and one-third shows bilateral disease. Patients with the mutation are candidates for prophylactic bilateral mastectomy

#### **Familial Breast Cancer**

• Familial breast cancer (FBC) is described as breast cancer within a family history of one or more first or second degree relatives affected. A patient with one or more first degree relatives with breast cancer in this category has a substantial excess lifetime risk of breast cancer when compared to patients in the general population. The relative risk increases from 1.80, 2.93, and 3.90 with one, two, and three first degree relatives compared to women without affected pedigree. FBC suggests a clustering of cancers that probably occurred by chance. In other words, there may be a combination of genetic and nongenetic (i.e., environmental) factors that contributed to the development of cancers within a family. In such instances, where an alteration in a single major gene is not likely or is not identified, individuals may still face elevated risks of cancer

#### **Genome Sequencing of Breast Cancers**

• Whole genome sequencing (WGS): The use of rapidly evolving techniques that combines whole genome, deep generation sequencing, and next generation sequencing have provided novel insights into the understanding of mutational analysis in breast cancer. Although these studies are in their infancy, it is already clear that essentially all breast cancers have an enormous number of mutations, far more than originally imagined—suggesting that developing widely successful targeted therapies will be extremely difficult. The seminal study by Sjoblom, based on outdated sequencing technology, found more than 100 distinct mutations in just 11 breast cancers. A more recent study Ding et al., using newer higher resolution technology, found an average of 50 somatic point mutations (including *JAK2, PTCH2, CSMD1, NRK, TP53* , *MAP3K8* ), 28 large deletions, 6 inversions, and 7 translocations in a single case of basal-like breast cancer. One of the next major challenges in breast cancer research will be to determine which of the mutations are the "drivers" for developing breast cancer

## **Suggested Reading**

- Abd El-Rehim DM, Pinder SE, Paish CE, et al. Expression of luminal and basal cytokeratins in human breast carcinoma. J Pathol. 2004;203:661–71.
- Al-Kuraya K, Schraml P, Torhorst J, et al. Prognostic relevance of gene amplifications and coamplifications in breast cancer. Cancer Res. 2004;64:8534–40.
- Allred DC. Issues and updates: evaluating estrogen receptor-alpha, progesterone receptor, and HER2 in breast cancer. Mod Pathol. 2010;23 Suppl 2:S52–9.
- Allred DC, Swanson PE. Testing for erbB-2 by immunohistochemistry in breast cancer. Am J Clin Pathol. 2000;113:171–5.
- Allred DC, Mohsin SK, Fuqua SA. Histological and biological evolution of human premalignant breast disease. Endocr Relat Cancer. 2001;8:47–61.
- Anderson E. The role of oestrogen and progesterone receptors in human mammary development and tumorigenesis. Breast Cancer Res. 2002;4:197–201.
- Anderson E, Clarke RB. Steroid receptors and cell cycle in normal mammary epithelium. J Mammary Gland Biol Neoplasia. 2004;9:3–13.
- Armes JE, Trute L, White D, et al. Distinct molecular pathogeneses of early-onset breast cancers in BRCA1 and BRCA2 mutation carriers: a population-based study. Cancer Res. 1999;59:2011–7.
- Arriola E, Rodriguez-Pinilla SM, Lambros MB, et al. Topoisomerase II alpha amplification may predict benefit from adjuvant anthracyclines in HER2 positive early breast cancer. Breast Cancer Res Treat. 2007;106:181–9.
- Badve SS, Baehner FL, Gray RP, et al. Estrogen- and progesterone-receptor status in ECOG 2197: comparison of immunohistochemistry by local and central laboratories and quantitative reverse transcription polymerase chain reaction by central laboratory. J Clin Oncol. 2008;26:2473–81.
- Bane AL, Beck JC, Bleiweiss I, et al. BRCA2 mutationassociated breast cancers exhibit a distinguishing phenotype based on morphology and molecular profiles from tissue microarrays. Am J Surg Pathol. 2007;31:121–8.
- Barnes DM, Harris WH, Smith P, Millis RR, Rubens RD. Immunohistochemical determination of oestrogen receptor: comparison of different methods of assessment of staining and correlation with clinical outcome of breast cancer patients. Br J Cancer. 1996;74: 1445–51.
- Baselga J, Campone M, Piccart M, et al. Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer. N Engl J Med. 2012;366:520–9.
- Bean GR, Bryson AD, Pilie PG, et al. Morphologically normal-appearing mammary epithelial cells obtained from high-risk women exhibit methylation silencing of INK4a/ARF. Clin Cancer Res. 2007;13:6834–41.
- Bedard PL, Cardoso F, Piccart-Gebhart MJ. Stemming resistance to HER-2 targeted therapy. J Mammary Gland Biol Neoplasia. 2009;14:55–66.
- Bertucci F, Finetti P, Cervera N, et al. Gene expression profiling shows medullary breast cancer is a subgroup of basal breast cancers. Cancer Res. 2006;66:4636–44.
- Bjornstrom L, Sjoberg M. Mechanisms of estrogen receptor signaling: convergence of genomic and nongenomic actions on target genes. Mol Endocrinol. 2005;19:833–42.
- Borresen-Dale AL. TP53 and breast cancer. Hum Mutat. 2003;21:292–300.
- Brekelmans CT, Tilanus-Linthorst MM, Seynaeve C, et al. Tumour characteristics, survival and prognostic factors of hereditary breast cancer from BRCA2-, BRCA1 and non-BRCA1/2 families as compared to sporadic breast cancer cases. Eur J Cancer. 2007;43:867–76.
- Brownstein MH, Wolf M, Bikowski JB. Cowden's disease: a cutaneous marker of breast cancer. Cancer. 1978;41:2393–8.
- Buyse M, Loi S, van't Veer LJ, et al. Validation and clinical utility of a 70-gene prognostic signature for women with node-negative breast cancer. J Natl Cancer Inst. 2006;98:1183–92.
- Caldas C, Aparicio SA. The molecular outlook. Nature. 2002;415:484–5.
- Cardoso F, Van't Veer L, Rutgers E, Loi S, Mook S, Piccart-Gebhart MJ. Clinical application of the 70-gene profile: the MINDACT trial. J Clin Oncol. 2008;26:729–35.
- Ceriani RL, Contesso GP, Nataf BM. Hormone requirement for growth and differentiation of the human mammary gland in organ culture. Cancer Res. 1972; 32:2190–6.
- Clarke CL, Sutherland RL. Progestin regulation of cellular proliferation. Endocr Rev. 1990;11:266–301.
- Clarke RB, Howell A, Potten CS, Anderson E. Dissociation between steroid receptor expression and cell proliferation in the human breast. Cancer Res. 1997;57:4987–91.
- Clarke RB, Anderson E, Howell A, Potten CS. Regulation of human breast epithelial stem cells. Cell Prolif. 2003;36:45–58.
- Clarke C, Sandle J, Lakhani SR. Myoepithelial cells: pathology, cell separation and markers of myoepithelial differentiation. J Mammary Gland Biol Neoplasia. 2005;10:273–80.
- Coletta RD, Christensen K, Reichenberger KJ, et al. The Six1 homeoprotein stimulates tumorigenesis by reactivation of cyclin A1. Proc Natl Acad Sci USA. 2004;101:6478–83.
- Conneely OM, Lydon JP. Progesterone receptors in reproduction: functional impact of the A and B isoforms. Steroids. 2000;65:571–7.
- Correa Geyer F, Reis-Filho JS. Microarray-based gene expression profiling as a clinical tool for breast cancer management: are we there yet? Int J Surg Pathol. 2009;17:285–302.
- Courjal F, Cuny M, Simony-Lafontaine J, et al. Mapping of DNA amplifications at 15 chromosomal localizations in 1875 breast tumors: definition of phenotypic groups. Cancer Res. 1997;57:4360–7.
- Coussens L, Yang-Feng TL, Liao YC, et al. Tyrosine kinase receptor with extensive homology to EGF receptor shares chromosomal location with neu oncogene. Science. 1985;230:1132–9.
- Cowen PN, Teasdale J, Jackson P, Reid BJ. Oestrogen receptor in breast cancer: prognostic studies using a new immunohistochemical assay. Histopathology. 1990;17:319–25.
- Crawford YG, Gauthier ML, Joubel A, et al. Histologically normal human mammary epithelia with silenced p16(INK4a) overexpress COX-2, promoting a premalignant program. Cancer Cell. 2004;5:263–73.
- Cui X, Schiff R, Arpino G, Osborne CK, Lee AV. Biology of progesterone receptor loss in breast cancer and its implications for endocrine therapy. J Clin Oncol. 2005;23:7721–35.
- Cuny M, Kramar A, Courjal F, et al. Relating genotype and phenotype in breast cancer: an analysis of the prognostic significance of amplification at eight different genes or loci and of p53 mutations. Cancer Res. 2000;60:1077–83.
- Dahabreh IJ, Linardou H, Siannis F, Fountzilas G, Murray S. Trastuzumab in the adjuvant treatment of earlystage breast cancer: a systematic review and metaanalysis of randomized controlled trials. Oncologist. 2008;13:620–30.
- Dal Lago L, Durbecq V, Desmedt C, et al. Correction for chromosome-17 is critical for the determination of true Her-2/neu gene amplification status in breast cancer. Mol Cancer Ther. 2006;5:2572–9.
- Dean-Colomb W, Esteva FJ. Her2-positive breast cancer: herceptin and beyond. Eur J Cancer. 2008;44:2806-12.
- Deng G, Lu Y, Zlotnikov G, Thor AD, Smith HS. Loss of heterozygosity in normal tissue adjacent to breast carcinomas. Science. 1996;274:2057–9.
- Deroo BJ, Hewitt SC, Collins JB, Grissom SF, Hamilton KJ, Korach KS. Profile of estrogen-responsive genes in an estrogen-specific mammary gland outgrowth model. Mol Reprod Dev. 2009;76:733–50.
- Dewar R, Fadare O, Gilmore H, Gown AM. Best practices in diagnostic immunohistochemistry: myoepithelial markers in breast pathology. Arch Pathol Lab Med. 2011;135:422–9.
- Dhesy-Thind B, Pritchard KI, Messersmith H, O'Malley F, Elavathil L, Trudeau M. HER2/neu in systemic therapy for women with breast cancer: a systematic review. Breast Cancer Res Treat. 2008;109:209–29.
- Ding L, Ellis MJ, Li S, et al. Genome remodelling in a basal-like breast cancer metastasis and xenograft. Nature. 2010;464:999–1005.
- Doane AS, Danso M, Lal P, et al. An estrogen receptornegative breast cancer subset characterized by a hormonally regulated transcriptional program and response to androgen. Oncogene. 2006;25:3994–4008.
- Downs-Kelly E, Yoder BJ, Stoler M, et al. The influence of polysomy 17 on HER2 gene and protein expression in adenocarcinoma of the breast: a fluorescent in situ hybridization, immunohistochemical, and isotopic mRNA in situ hybridization study. Am J Surg Pathol. 2005;29:1221–7.
- Dowsett M, Allred C, Knox J, et al. Relationship between quantitative estrogen and progesterone receptor expression and human epidermal growth factor receptor 2 (HER-2) status with recurrence in the Arimidex, Tamoxifen, Alone or in Combination trial. J Clin Oncol. 2008;26:1059–65.
- Dybdal N, Leiberman G, Anderson S, et al. Determination of HER2 gene amplification by fluorescence in situ hybridization and concordance with the clinical trials immunohistochemical assay in women with metastatic breast cancer evaluated for treatment with trastuzumab. Breast Cancer Res Treat. 2005;93:3–11.
- Easton DF, Ford D, Bishop DT. Breast and ovarian cancer incidence in BRCA1-mutation carriers. Breast Cancer Linkage Consortium. Am J Hum Genet. 1995;56: 265–71.
- Eisinger F, Stoppa-Lyonnet D, Longy M, et al. Germ line mutation at BRCA1 affects the histoprognostic grade in hereditary breast cancer. Cancer Res. 1996;56: 471–4.
- Elbauomy Elsheikh S, Green AR, Lambros MB, et al. FGFR1 amplification in breast carcinomas: a chromogenic in situ hybridisation analysis. Breast Cancer Res. 2007;9:R23.
- Elledge RM, Green S, Pugh R, et al. Estrogen receptor (ER) and progesterone receptor (PR), by ligand-binding assay compared with ER, PR and pS2, by immunohistochemistry in predicting response to tamoxifen in metastatic breast cancer: a Southwest Oncology Group Study. Int J Cancer. 2000;89:111–7.
- Engel RH, Kaklamani VG. HER2-positive breast cancer: current and future treatment strategies. Drugs. 2007; 67:1329–41.
- Esteban JM, Ahn C, Battifora H, Felder B. Quantitative immunohistochemical assay for hormonal receptors: technical aspects and biological significance. J Cell Biochem Suppl. 1994;19:138–45.
- Fan C, Oh DS, Wessels L, et al. Concordance among gene-expression-based predictors for breast cancer. N Engl J Med. 2006;355:560–9.
- Fan C, Prat A, Parker JS, et al. Building prognostic models for breast cancer patients using clinical variables and hundreds of gene expression signatures. BMC Med Genomics. 2011;4:3.
- Fantl V, Stamp G, Andrews A, Rosewell I, Dickson C. Mice lacking cyclin D1 are small and show defects in eye and mammary gland development. Genes Dev. 1995;9:2364–72.
- Farmer P, Bonnefoi H, Becette V, et al. Identification of molecular apocrine breast tumours by microarray analysis. Oncogene. 2005;24:4660–71.
- Faverly D, Holland R, Burgers L. An original stereomicroscopic analysis of the mammary glandular tree. Virchows Arch A Pathol Anat Histopathol. 1992;421: 115–9.
- Fendrick JL, Raafat AM, Haslam SZ. Mammary gland growth and development from the postnatal period to postmenopause: ovarian steroid receptor ontogeny and regulation in the mouse. J Mammary Gland Biol Neoplasia. 1998;3:7–22.
- Fisher B, Redmond C, Brown A, et al. Treatment of primary breast cancer with chemotherapy and tamoxifen. N Engl J Med. 1981;305:1–6.
- Fisher B, Redmond C, Brown A, et al. Influence of tumor estrogen and progesterone receptor levels on the response to tamoxifen and chemotherapy in primary breast cancer. J Clin Oncol. 1983;1:227–41.
- Fitzgibbons PL, Page DL, Weaver D, et al. Prognostic factors in breast cancer. College of American Pathologists Consensus Statement 1999. Arch Pathol Lab Med. 2000;124:966–78.
- Ford D, Easton DF. The genetics of breast and ovarian cancer. Br J Cancer. 1995;72:805–12.
- Foulkes WD, Stefansson IM, Chappuis PO, et al. Germline BRCA1 mutations and a basal epithelial phenotype in breast cancer. J Natl Cancer Inst. 2003;95:1482–5.
- Francis GD, Dimech M, Giles L, Hopkins A. Frequency and reliability of oestrogen receptor, progesterone receptor and HER2 in breast carcinoma determined by immunohistochemistry in Australasia: results of the RCPA Quality Assurance Program. J Clin Pathol. 2007;60:1277–83.
- Frank TS, Deffenbaugh AM, Reid JE, et al. Clinical characteristics of individuals with germline mutations in BRCA1 and BRCA2: analysis of 10,000 individuals. J Clin Oncol. 2002;20:1480–90.
- Fulford LG, Easton DF, Reis-Filho JS, et al. Specific morphological features predictive for the basal phenotype in grade 3 invasive ductal carcinoma of breast. Histopathology. 2006;49:22–34.
- Gaffney DK, Brohet RM, Lewis CM, et al. Response to radiation therapy and prognosis in breast cancer patients with BRCA1 and BRCA2 mutations. Radiother Oncol. 1998;47:129–36.
- Geisler S, Borresen-Dale AL, Johnsen H, et al. TP53 gene mutations predict the response to neoadjuvant treatment with 5-fluorouracil and mitomycin in locally advanced breast cancer. Clin Cancer Res. 2003; 9:5582–8.
- Gelber RD, Gelber S. Facilitating consensus by examining patterns of treatment effects. Breast. 2009;18 Suppl 3:S2–8.
- Gianni L, Zambetti M, Clark K, et al. Gene expression profiles in paraffin-embedded core biopsy tissue predict response to chemotherapy in women with locally advanced breast cancer. J Clin Oncol. 2005;23: 7265–77.
- Goepfert TM, McCarthy M, Kittrell FS, et al. Progesterone facilitates chromosome instability (aneuploidy) in p53 null normal mammary epithelial cells. FASEB J. 2000;14:2221–9.
- Goodwin PJ, Phillips KA, West DW, et al. Breast cancer prognosis in BRCA1 and BRCA2 mutation carriers: an International Prospective Breast Cancer Family Registry population-based cohort study. J Clin Oncol. 2012;30:19–26.
- Greene GL, Press MF. Structure and dynamics of the estrogen receptor. J Steroid Biochem. 1986;24:1–7.
- Gudjonsson T, Adriance MC, Sternlicht MD, Petersen OW, Bissell MJ. Myoepithelial cells: their origin and function in breast morphogenesis and neoplasia. J Mammary Gland Biol Neoplasia. 2005;10:261–72.
- Gusterson B. Do 'basal-like' breast cancers really exist? Nat Rev Cancer. 2009;9:128–34.
- Gusterson BA, Ross DT, Heath VJ, Stein T. Basal cytokeratins and their relationship to the cellular origin and functional classification of breast cancer. Breast Cancer Res. 2005;7:143–8.
- Hall JM, Lee MK, Newman B, et al. Linkage of earlyonset familial breast cancer to chromosome 17q21. Science. 1990;250:1684–9.
- Hamann U, Sinn HP. Survival and tumor characteristics of German hereditary breast cancer patients. Breast Cancer Res Treat. 2000;59:185–92.
- Hammond ME, Fitzgibbons PL, Compton CC, et al. College of American Pathologists Conference XXXV:

solid tumor prognostic factors-which, how and so what? Summary document and recommendations for implementation. Cancer Committee and Conference Participants. Arch Pathol Lab Med. 2000;124:958–65.

- Hammond ME, Hayes DF, Dowsett M, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. Arch Pathol Lab Med. 2010a;134:907–22.
- Hammond ME, Hayes DF, Dowsett M, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer (unabridged version). Arch Pathol Lab Med. 2010b;134:e48–72.
- Harari D, Yarden Y. Molecular mechanisms underlying ErbB2/HER2 action in breast cancer. Oncogene. 2000;19:6102–14.
- Hart LL, Davie JR. The estrogen receptor: more than the average transcription factor. Biochem Cell Biol. 2002;80:335–41.
- Harvey JM, Clark GM, Osborne CK, Allred DC. Estrogen receptor status by immunohistochemistry is superior to the ligand-binding assay for predicting response to adjuvant endocrine therapy in breast cancer. J Clin Oncol. 1999;17:1474–81.
- Hayes MJ, Thomas D, Emmons A, Giordano TJ, Kleer CG. Genetic changes of Wnt pathway genes are common events in metaplastic carcinomas of the breast. Clin Cancer Res. 2008;14:4038–44.
- Hennessy BT, Gonzalez-Angulo AM, Stemke-Hale K, et al. Characterization of a naturally occurring breast cancer subset enriched in epithelial-to-mesenchymal transition and stem cell characteristics. Cancer Res. 2009;69:4116–24.
- Herschkowitz JI, Simin K, Weigman VJ, et al. Identification of conserved gene expression features between murine mammary carcinoma models and human breast tumors. Genome Biol. 2007;8:R76.
- Herynk MH, Fuqua SA. Estrogen receptor mutations in human disease. Endocr Rev. 2004;25:869–98.
- Hewitt SC, Harrell JC, Korach KS. Lessons in estrogen biology from knockout and transgenic animals. Annu Rev Physiol. 2005;67:285–308.
- Hill CB, Yeh IT. Myoepithelial cell staining patterns of papillary breast lesions: from intraductal papillomas to invasive papillary carcinomas. Am J Clin Pathol. 2005;123:36–44.
- Holst CR, Nuovo GJ, Esteller M, et al. Methylation of p16(INK4a) promoters occurs in vivo in histologically normal human mammary epithelia. Cancer Res. 2003;63:1596–601.
- Holst F, Stahl PR, Ruiz C, et al. Estrogen receptor alpha (ESR1) gene amplification is frequent in breast cancer. Nat Genet. 2007;39:655–60.
- Honrado E, Osorio A, Palacios J, Benitez J. Pathology and gene expression of hereditary breast tumors associated with BRCA1, BRCA2 and CHEK2 gene mutations. Oncogene. 2006;25:5837–45.
- Honrado E, Osorio A, Milne RL, et al. Immunohistochemical classification of non-BRCA1/2 tumors identifies different groups that demonstrate the heterogeneity of BRCAX families. Mod Pathol. 2007;20:1298–306.
- Hovey RC, Trott JF, Vonderhaar BK. Establishing a framework for the functional mammary gland: from endocrinology to morphology. J Mammary Gland Biol Neoplasia. 2002;7:17–38.
- Howell A, Cuzick J, Baum M, et al. Results of the ATAC (Arimidex, Tamoxifen, Alone or in Combination) trial after completion of 5 years' adjuvant treatment for breast cancer. Lancet. 2005;365:60–2.
- Hu Z, Fan C, Oh DS, et al. The molecular portraits of breast tumors are conserved across microarray platforms. BMC Genomics. 2006;7:96.
- Hu M, Yao J, Carroll DK, et al. Regulation of in situ to invasive breast carcinoma transition. Cancer Cell. 2008;13:394–406.
- Hussain SP, Harris CC. Molecular epidemiology of human cancer: contribution of mutation spectra studies of tumor suppressor genes. Cancer Res. 1998;58: 4023–37.
- Hutson SW, Cowen PN, Bird CC. Morphometric studies of age related changes in normal human breast and their significance for evolution of mammary cancer. J Clin Pathol. 1985;38:281–7.
- Huvos AG, Lucas Jr JC, Foote Jr FW. Metaplastic breast carcinoma. Rare form of mammary cancer. N Y State J Med. 1973;73:1078–82.
- Jansson T, Inganas M, Sjogren S, et al. p53 Status predicts survival in breast cancer patients treated with or without postoperative radiotherapy: a novel hypothesis based on clinical findings. J Clin Oncol. 1995;13: 2745–51.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA Cancer J Clin. 2011;61:69–90.
- Jerevall PL, Ma XJ, Li H, et al. Prognostic utility of HOXB13:IL17BR and molecular grade index in earlystage breast cancer patients from the Stockholm trial. Br J Cancer. 2011;104:1762–9.
- Johannsson OT, Idvall I, Anderson C, et al. Tumour biological features of BRCA1-induced breast and ovarian cancer. Eur J Cancer. 1997;33:362–71.
- Jordan C. Historical perspective on hormonal therapy of advanced breast cancer. Clin Ther. 2002;24(Suppl A): A3–16.
- Jordan VC, Wolf MF, Mirecki DM, Whitford DA, Welshons WV. Hormone receptor assays: clinical usefulness in the management of carcinoma of the breast. Crit Rev Clin Lab Sci. 1988;26:97–152.
- Kallioniemi A, Kallioniemi OP, Piper J, et al. Detection and mapping of amplified DNA sequences in breast cancer by comparative genomic hybridization. Proc Natl Acad Sci USA. 1994;91:2156–60.
- Kampa M, Pelekanou V, Castanas E. Membrane-initiated steroid action in breast and prostate cancer. Steroids. 2008;73:953–60.
- Karni R, Jove R, Levitzki A. Inhibition of pp 60c-Src reduces Bcl-XL expression and reverses the trans-

formed phenotype of cells overexpressing EGF and HER-2 receptors. Oncogene. 1999;18:4654–62.

- Kaurah P, MacMillan A, Boyd N, et al. Founder and recurrent CDH1 mutations in families with hereditary diffuse gastric cancer. JAMA. 2007;297:2360–72.
- Kauraniemi P, Kallioniemi A. Activation of multiple cancer-associated genes at the ERBB2 amplicon in breast cancer. Endocr Relat Cancer. 2006;13:39–49.
- Keaveney M, Klug J, Dawson MT, et al. Evidence for a previously unidentified upstream exon in the human oestrogen receptor gene. J Mol Endocrinol. 1991;6:111–5.
- Keith WN, Douglas F, Wishart GC, et al. Co-amplification of erbB2, topoisomerase II alpha and retinoic acid receptor alpha genes in breast cancer and allelic loss at topoisomerase I on chromosome 20. Eur J Cancer. 1993;29A:1469–75.
- Keller G, Vogelsang H, Becker I, et al. Diffuse type gastric and lobular breast carcinoma in a familial gastric cancer patient with an E-cadherin germline mutation. Am J Pathol. 1999;155:337–42.
- Klingbeil P, Natrajan R, Everitt G, et al. CD44 is overexpressed in basal-like breast cancers but is not a driver of 11p13 amplification. Breast Cancer Res Treat. 2010;120:95–109.
- Konecny G, Pauletti G, Pegram M, et al. Quantitative association between HER-2/neu and steroid hormone receptors in hormone receptor-positive primary breast cancer. J Natl Cancer Inst. 2003;95:142–53.
- Kuter I, Gee JM, Hegg R, et al. Dose-dependent change in biomarkers during neoadjuvant endocrine therapy with fulvestrant: results from NEWEST, a randomized Phase II study. Breast Cancer Res Treat. 2012;133(1): 237–46.
- LaCroix AZ, Chlebowski RT, Manson JE, et al. Health outcomes after stopping conjugated equine estrogens among postmenopausal women with prior hysterectomy: a randomized controlled trial. JAMA. 2011;305: 1305–14.
- Lacroix-Triki M, Geyer FC, Lambros MB, et al. Betacatenin/Wnt signalling pathway in fibromatosis, metaplastic carcinomas and phyllodes tumours of the breast. Mod Pathol. 2010;23:1438–48.
- Lakhani SR, Jacquemier J, Sloane JP, et al. Multifactorial analysis of differences between sporadic breast cancers and cancers involving BRCA1 and BRCA2 mutations. J Natl Cancer Inst. 1998;90:1138–45.
- Lakhani SR, Van De Vijver MJ, Jacquemier J, et al. The pathology of familial breast cancer: predictive value of immunohistochemical markers estrogen receptor, progesterone receptor, HER-2, and p53 in patients with mutations in BRCA1 and BRCA2. J Clin Oncol. 2002;20:2310–8.
- Larson PS, de las Morenas A, Bennett SR, Cupples LA, Rosenberg CL. Loss of heterozygosity or allele imbalance in histologically normal breast epithelium is distinct from loss of heterozygosity or allele imbalance in co-existing carcinomas. Am J Pathol. 2002;161: 283–90.
- Larson PS, Schlechter BL, de las Morenas A, Garber JE, Cupples LA, Rosenberg CL. Allele imbalance, or loss

of heterozygosity, in normal breast epithelium of sporadic breast cancer cases and BRCA1 gene mutation carriers is increased compared with reduction mammoplasty tissues. J Clin Oncol. 2005;23:8613–9.

- Lee S, Medina D, Tsimelzon A, et al. Alterations of gene expression in the development of early hyperplastic precursors of breast cancer. Am J Pathol. 2007;171: 252–62.
- Lee S, Stewart S, Nagtegaal I, et al. Differentially expressed genes regulating the progression of ductal carcinoma in situ to invasive breast cancer. Cancer Res 2012;72(17):4574–86.
- Levin ER, Pietras RJ. Estrogen receptors outside the nucleus in breast cancer. Breast Cancer Res. Treat. 2008;108:351–61.
- Liaw D, Marsh DJ, Li J, et al. Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. Nat Genet. 1997;16:64–7.
- Liedtke C, Hatzis C, Symmans WF, et al. Genomic grade index is associated with response to chemotherapy in patients with breast cancer. J Clin Pathol. 2009;27: 3185–91.
- Liu S, Ginestier C, Charafe-Jauffret E, et al. BRCA1 regulates human mammary stem/progenitor cell fate. Proc Natl Acad Sci USA. 2008;105:1680–5.
- Livasy CA, Karaca G, Nanda R, et al. Phenotypic evaluation of the basal-like subtype of invasive breast carcinoma. Mod Pathol. 2006;19:264–71.
- Lockwood CA, Ricciardelli C, Raymond WA, Seshadri R, McCaul K, Horsfall DJ. A simple index using video image analysis to predict disease outcome in primary breast cancer. Int J Cancer. 1999;84:203–8.
- Loi S, Haibe-Kains B, Desmedt C, et al. Definition of clinically distinct molecular subtypes in estrogen receptor-positive breast carcinomas through genomic grade. J Clin Oncol. 2007;25:1239–46.
- Lubinski J, Gorski B, Huzarski T, et al. BRCA1-positive breast cancers in young women from Poland. Breast Cancer Res Treat. 2006;99:71–6.
- Lukas J, Niu N, Press MF. p53 Mutations and expression in breast carcinoma in situ. Am J Pathol. 2000;156:183–91.
- Lydon JP, Ge G, Kittrell FS, Medina D, O'Malley BW. Murine mammary gland carcinogenesis is critically dependent on progesterone receptor function. Cancer Res. 1999;59:4276–84.
- Lynch HT, Mulcahy GM, Harris RE, Guirgis HA, Lynch JF. Genetic and pathologic findings in a kindred with hereditary sarcoma, breast cancer, brain tumors, leukemia, lung, laryngeal, and adrenal cortical carcinoma. Cancer. 1978;41:2055–64.
- Lynch HT, Watson P, Conway TA, et al. DNA screening for breast/ovarian cancer susceptibility based on linked markers. A family study. Arch Intern Med. 1993;153: 1979–87.
- Lynch HT, Lynch J, Conway T, et al. Hereditary breast cancer and family cancer syndromes. World J Surg. 1994;18:21–31.
- Lynch BJ, Holden JA, Buys SS, Neuhausen SL, Gaffney DK. Pathobiologic characteristics of hereditary breast cancer. Hum Pathol. 1998;29:1140–4.
- Lynch HT, McComb RD, Osborn NK, et al. Predominance of brain tumors in an extended Li-Fraumeni (SBLA) kindred, including a case of Sturge-Weber syndrome. Cancer. 2000;88:433–9.
- Lynch HT, Kaurah P, Wirtzfeld D, et al. Hereditary diffuse gastric cancer: diagnosis, genetic counseling, and prophylactic total gastrectomy. Cancer. 2008;112: 2655–63.
- Ma XJ, Wang Z, Ryan PD, et al. A two-gene expression ratio predicts clinical outcome in breast cancer patients treated with tamoxifen. Cancer Cell. 2004;5:607–16.
- Ma XJ, Salunga R, Dahiya S, et al. A five-gene molecular grade index and HOXB13:IL17BR are complementary prognostic factors in early stage breast cancer. Clin Cancer Res. 2008;14:2601–8.
- Ma CX, Sanchez CG, Ellis MJ. Predicting endocrine therapy responsiveness in breast cancer. Oncology (Williston Park). 2009;23:133–42.
- Malkin D, Li FP, Strong LC, et al. Germ line p53 mutations in a familial syndrome of breast cancer, sarcomas, and other neoplasms. Science. 1990;250: 1233–8.
- Malone KE, Daling JR, Doody DR, et al. Prevalence and predictors of BRCA1 and BRCA2 mutations in a population-based study of breast cancer in white and black American women ages 35 to 64 years. Cancer Res. 2006;66:8297–308.
- Marchio C, Iravani M, Natrajan R, et al. Mixed micropapillary-ductal carcinomas of the breast: a genomic and immunohistochemical analysis of morphologically distinct components. J Pathol. 2009;218:301–15.
- Marcus JN, Watson P, Page DL, et al. Hereditary breast cancer: pathobiology, prognosis, and BRCA1 and BRCA2 gene linkage. Cancer. 1996;77:697–709.
- Marcus JN, Watson P, Page DL, et al. BRCA2 hereditary breast cancer pathophenotype. Breast Cancer Res Treat. 1997;44:275–7.
- Mariani G, Fasolo A, De Benedictis E, Gianni L. Trastuzumab as adjuvant systemic therapy for HER2 positive breast cancer. Nat Clin Pract Oncol. 2009;6: 93–104.
- Marotti JD, Collins LC, Hu R, Tamimi RM. Estrogen receptor-beta expression in invasive breast cancer in relation to molecular phenotype: results from the Nurses' Health Study. Mod Pathol. 2010;23:197–204.
- Marsh DJ, Dahia PL, Caron S, et al. Germline PTEN mutations in Cowden syndrome-like families. J Med Genet. 1998;35:881–5.
- Mauri D, Pavlidis N, Polyzos NP, Ioannidis JP. Survival with aromatase inhibitors and inactivators versus standard hormonal therapy in advanced breast cancer: meta-analysis. J Natl Cancer Inst. 2006;98:1285–91.
- McBryan J, Howlin J, Kenny PA, Shioda T, Martin F. ERalpha-CITED1 co-regulated genes expressed during pubertal mammary gland development: implications for breast cancer prognosis. Oncogene. 2007; 26:6406–19.
- McCarthy A, Savage K, Gabriel A, Naceur C, Reis-Filho JS, Ashworth A. A mouse model of basal-like breast carcinoma with metaplastic elements. J Pathol. 2007;211:389–98.
- Menard S, Tagliabue E, Campiglio M, Pupa SM. Role of HER2 gene overexpression in breast carcinoma. J Cell Physiol. 2000;182:150–62.
- Menard S, Valagussa P, Pilotti S, et al. Response to cyclophosphamide, methotrexate, and fluorouracil in lymph node-positive breast cancer according to HER2 overexpression and other tumor biologic variables. J Clin Oncol. 2001;19:329–35.
- Miki Y, Swensen J, Shattuck-Eidens D, et al. A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. Science. 1994;266:66–71.
- Mitani Y, Li J, Weber RS, et al. Expression and regulation of the DeltaN and TAp63 isoforms in salivary gland tumorigenesis clinical and experimental findings. Am J Pathol. 2011;179:391–9.
- Moelans CB, de Weger RA, Monsuur HN, Vijzelaar R, van Diest PJ. Molecular profiling of invasive breast cancer by multiplex ligation-dependent probe amplificationbased copy number analysis of tumor suppressor and oncogenes. Mod Pathol. 2010;23:1029–39.
- Mohsin SK, Weiss H, Havighurst T, et al. Progesterone receptor by immunohistochemistry and clinical outcome in breast cancer: a validation study. Mod Pathol. 2004;17:1545–54.
- Moll UM, Riou G, Levine AJ. Two distinct mechanisms alter p53 in breast cancer: mutation and nuclear exclusion. Proc Natl Acad Sci USA. 1992;89:7262–6.
- Mueller SO, Clark JA, Myers PH, Korach KS. Mammary gland development in adult mice requires epithelial and stromal estrogen receptor alpha. Endocrinology. 2002;143:2357–65.
- Muggerud AA, Ronneberg JA, Warnberg F, et al. Frequent aberrant DNA methylation of ABCB1, FOXC1, PPP2R2B and PTEN in ductal carcinoma in situ and early invasive breast cancer. Breast Cancer Res. 2010;12:R3.
- Naccarato AG, Viacava P, Vignati S, et al. Biomorphological events in the development of the human female mammary gland from fetal age to puberty. Virchows Arch. 2000;436:431–8.
- Narod SA, Feunteun J, Lynch HT, et al. Familial breastovarian cancer locus on chromosome 17q12-q23. Lancet. 1991;338:82–3.
- Narod SA, Brunet JS, Ghadirian P, et al. Tamoxifen and risk of contralateral breast cancer in BRCA1 and BRCA2 mutation carriers: a case–control study. Hereditary Breast Cancer Clinical Study Group. Lancet. 2000;356:1876–81.
- Nass SJ, Dickson RB. Defining a role for c-Myc in breast tumorigenesis. Breast Cancer Res Treat. 1997;44:1–22.
- Nassar H. Carcinomas with micropapillary morphology: clinical significance and current concepts. Adv Anat Pathol. 2004;11:297–303.
- Nathan B, Anbazhagan R, Clarkson P, Bartkova J, Gusterson B. Expression of BCL-2 in the developing human fetal and infant breast. Histopathology. 1994;24:73–6.
- Nelen MR, van Staveren WC, Peeters EA, et al. Germline mutations in the PTEN/MMAC1 gene in patients with Cowden disease. Hum Mol Genet. 1997;6:1383–7.
- Nielsen TO, Hsu FD, Jensen K, et al. Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma. Clin Cancer Res. 2004;10:5367–74.
- Nielsen TO, Parker JS, Leung S, et al. A comparison of PAM50 intrinsic subtyping with immunohistochemistry and clinical prognostic factors in tamoxifen-treated estrogen receptor-positive breast cancer. Clin Cancer Res. 2010;16:5222–32.
- Ogawa Y, Moriya T, Kato Y, et al. Immunohistochemical assessment for estrogen receptor and progesterone receptor status in breast cancer: analysis for a cut-off point as the predictor for endocrine therapy. Breast Cancer. 2004;11:267–75.
- Olivier M, Hainaut P. TP53 mutation patterns in breast cancers: searching for clues of environmental carcinogenesis. Semin Cancer Biol. 2001;11:353–60.
- Osborne RJ, Merlo GR, Mitsudomi T, et al. Mutations in the p53 gene in primary human breast cancers. Cancer Res. 1991;51:6194–8.
- Owens MA, Horten BC, Da Silva MM. HER2 amplification ratios by fluorescence in situ hybridization and correlation with immunohistochemistry in a cohort of 6556 breast cancer tissues. Clin Breast Cancer. 2004;5:63–9.
- Paik S, Shak S, Tang G, et al. A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. N Engl J Med. 2004;351:2817–26.
- Paik S, Tang G, Shak S, et al. Gene expression and benefit of chemotherapy in women with node-negative, estrogen receptor-positive breast cancer. J Clin Oncol. 2006;24:3726–34.
- Palacios J, Honrado E, Osorio A, et al. Immunohistochemical characteristics defined by tissue microarray of hereditary breast cancer not attributable to BRCA1 or BRCA2 mutations: differences from breast carcinomas arising in BRCA1 and BRCA2 mutation carriers. Clin Cancer Res. 2003;9:3606–14.
- Panoff JE, Hurley J, Takita C, et al. Risk of locoregional recurrence by receptor status in breast cancer patients receiving modern systemic therapy and post-mastectomy radiation. Breast Cancer Res Treat. 2011;128:899–906.
- Park K, Kwak K, Kim J, Lim S, Han S. c-myc amplification is associated with HER2 amplification and closely linked with cell proliferation in tissue microarray of nonselected breast cancers. Hum Pathol. 2005;36: 634–9.
- Parker JS, Mullins M, Cheang MC, et al. Supervised risk predictor of breast cancer based on intrinsic subtypes. J Clin Oncol. 2009;27:1160–7.
- Peppercorn J, Perou CM, Carey LA. Molecular subtypes in breast cancer evaluation and management: divide and conquer. Cancer Invest. 2008;26:1–10.
- Perez EA, Roche PC, Jenkins RB, et al. HER2 testing in patients with breast cancer: poor correlation between weak positivity by immunohistochemistry and gene amplification by fluorescence in situ hybridization. Mayo Clin Proc. 2002;77:148–54.
- Perez EA, Suman VJ, Davidson NE, et al. HER2 testing by local, central, and reference laboratories in

 specimens from the North Central Cancer Treatment Group N9831 intergroup adjuvant trial. J Clin Oncol. 2006;24:3032–8.

- Perez EA, Jenkins RB, Dueck AC, et al. C-MYC alterations and association with patient outcome in earlystage HER2-positive breast cancer from the north central cancer treatment group N9831 adjuvant trastuzumab trial. J Clin Oncol. 2011;29:651–9.
- Perou CM, Sorlie T, Eisen MB, et al. Molecular portraits of human breast tumours. Nature. 2000;406:747–52.
- Persons DL, Tubbs RR, Cooley LD, et al. HER-2 fluorescence in situ hybridization: results from the survey program of the College of American Pathologists. Arch Pathol Lab Med. 2006;130:325–31.
- Persson M, Andren Y, Mark J, Horlings HM, Persson F, Stenman G. Recurrent fusion of MYB and NFIB transcription factor genes in carcinomas of the breast and head and neck. Proc Natl Acad Sci USA. 2009;106:18740–4.
- Petersen OW, Polyak K. Stem cells in the human breast. Cold Spring Harb Perspect Biol. 2010;2:a003160.
- Petersen OW, Hoyer PE, van Deurs B. Frequency and distribution of estrogen receptor-positive cells in normal, nonlactating human breast tissue. Cancer Res. 1987; 47:5748–51.
- Pharoah PD, Guilford P, Caldas C. Incidence of gastric cancer and breast cancer in CDH1 (E-cadherin) mutation carriers from hereditary diffuse gastric cancer families. Gastroenterology. 2001;121:1348–53.
- Piva R, Bianchi N, Aguiari GL, Gambari R, del Senno L. Sequencing of an RNA transcript of the human estrogen receptor gene: evidence for a new transcriptional event. J Steroid Biochem Mol Biol. 1993;46:531–8.
- Polyak K, Hu M. Do myoepithelial cells hold the key for breast tumor progression? J Mammary Gland Biol Neoplasia. 2005;10:231–47.
- Porter DE, Cohen BB, Wallace MR, et al. Breast cancer incidence, penetrance and survival in probable carriers of BRCA1 gene mutation in families linked to BRCA1 on chromosome 17q12-21. Br J Surg. 1994;81:1512–5.
- Prat A, Perou CM. Deconstructing the molecular portraits of breast cancer. Mol Oncol. 2011;5:5–23.
- Press MF, Sauter G, Bernstein L, et al. Diagnostic evaluation of HER-2 as a molecular target: an assessment of accuracy and reproducibility of laboratory testing in large, prospective, randomized clinical trials. Clin Cancer Res. 2005;11:6598–607.
- Pritchard KI, Shepherd LE, O'Malley FP, et al. HER2 and responsiveness of breast cancer to adjuvant chemotherapy. N Engl J Med. 2006;354:2103–11.
- Pusztai L, Mazouni C, Anderson K, Wu Y, Symmans WF. Molecular classification of breast cancer: limitations and potential. Oncologist. 2006;11:868–77.
- Rakha E, Ellis I, Reis-Filho J. Are triple-negative and basal-like breast cancer synonymous? Clin Cancer Res. 2008;14:618; author reply 618–9.
- Reddy JC, Reimann JD, Anderson SM, Klein PM. Concordance between central and local laboratory HER2 testing from a community-based clinical study. Clin Breast Cancer. 2006;7:153–7.
- Regan MM, Viale G, Mastropasqua MG, et al. Re-evaluating adjuvant breast cancer trials: assessing hormone receptor status by immunohistochemical versus extraction assays. J Natl Cancer Inst. 2006;98: 1571–81.
- Reis-Filho JS, Tutt AN. Triple negative tumours: a critical review. Histopathology. 2008;52:108–18.
- Reis-Filho JS, Milanezi F, Steele D, et al. Metaplastic breast carcinomas are basal-like tumours. Histopathology. 2006;49:10–21.
- Reis-Filho JS, Drury S, Lambros MB, et al. ESR1 gene amplification in breast cancer: a common phenomenon? Nat Genet. 2008;40:809–10; author reply 810–2.
- Rhodes A, Jasani B, Balaton AJ, Barnes DM, Miller KD. Frequency of oestrogen and progesterone receptor positivity by immunohistochemical analysis in 7016 breast carcinomas: correlation with patient age, assay sensitivity, threshold value, and mammographic screening. J Clin Pathol. 2000a;53:688–96.
- Rhodes A, Jasani B, Barnes DM, Bobrow LG, Miller KD. Reliability of immunohistochemical demonstration of oestrogen receptors in routine practice: interlaboratory variance in the sensitivity of detection and evaluation of scoring systems. J Clin Pathol. 2000b;53: 125–30.
- Richer JK, Jacobsen BM, Manning NG, Abel MG, Wolf DM, Horwitz KB. Differential gene regulation by the two progesterone receptor isoforms in human breast cancer cells. J Biol Chem. 2002;277:5209–18.
- Robinson GW. Identification of signaling pathways in early mammary gland development by mouse genetics. Breast Cancer Res. 2004;6:105–8.
- Robinson GW. Cooperation of signalling pathways in embryonic mammary gland development. Nat Rev Genet. 2007;8:963–72.
- Rossouw JE, Anderson GL, Prentice RL, et al. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. JAMA. 2002;288:321–33.
- Rouzier R, Perou CM, Symmans WF, et al. Breast cancer molecular subtypes respond differently to preoperative chemotherapy. Clin Cancer Res. 2005;11:5678–85.
- Schechter AL, Stern DF, Vaidyanathan L, et al. The neu oncogene: an erb-B-related gene encoding a 185,000- Mr tumour antigen. Nature. 1984;312:513–6.
- Schiff R, Massarweh SA, Shou J, Bharwani L, Mohsin SK, Osborne CK. Cross-talk between estrogen receptor and growth factor pathways as a molecular target for overcoming endocrine resistance. Clin Cancer Res. 2004;10:331S–6.
- Schnitt SJ. Classification and prognosis of invasive breast cancer: from morphology to molecular taxonomy. Mod Pathol. 2010;23 Suppl 2:S60–4.
- Serova O, Montagna M, Torchard D, et al. A high incidence of BRCA1 mutations in 20 breast-ovarian cancer families. Am J Hum Genet. 1996;58:42–51.
- Shattuck-Eidens D, McClure M, Simard J, et al. A collaborative survey of 80 mutations in the BRCA1 breast and ovarian cancer susceptibility gene. Implications

for presymptomatic testing and screening. JAMA. 1995;273:535–41.

- Shiang CY, Qi Y, Wang B, et al. Amplification of fibroblast growth factor receptor-1 in breast cancer and the effects of brivanib alaninate. Breast Cancer Res Treat. 2010;123:747–55.
- Silva CM, Shupnik MA. Integration of steroid and growth factor pathways in breast cancer: focus on signal transducers and activators of transcription and their potential role in resistance. Mol Endocrinol. 2007;21:1499–512.
- Silver DP, Richardson AL, Eklund AC, et al. Efficacy of neoadjuvant Cisplatin in triple-negative breast cancer. J Clin Oncol. 2010;28:1145–53.
- Sjoblom T, Jones S, Wood LD, et al. The consensus coding sequences of human breast and colorectal cancers. Science. 2006;314:268–74.
- Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/ neu oncogene. Science. 1987;235:177–82.
- Smith CL, O'Malley BW. Coregulator function: a key to understanding tissue specificity of selective receptor modulators. Endocr Rev. 2004;25:45–71.
- Sorlie T, Perou CM, Tibshirani R, et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA. 2001;98:10869–74.
- Sorlie T, Tibshirani R, Parker J, et al. Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA. 2003;100:8418–23.
- Stendahl M, Ryden L, Nordenskjold B, Jonsson PE, Landberg G, Jirstrom K. High progesterone receptor expression correlates to the effect of adjuvant tamoxifen in premenopausal breast cancer patients. Clin Cancer Res. 2006;12:4614–8.
- Straver ME, Glas AM, Hannemann J, et al. The 70-gene signature as a response predictor for neoadjuvant chemotherapy in breast cancer. Breast Cancer Res Treat. 2010;119:551–8.
- Surmacz E, Bartucci M. Role of estrogen receptor alpha in modulating IGF-I receptor signaling and function in breast cancer. J Exp Clin Cancer Res. 2004;23: 385–94.
- Tang P, Skinner KA, Hicks DG. Molecular classification of breast carcinomas by immunohistochemical analysis: are we ready? Diagn Mol Pathol. 2009;18: 125–32.
- Tanner MM, Tirkkonen M, Kallioniemi A, et al. Independent amplification and frequent co-amplification of three nonsyntenic regions on the long arm of chromosome 20 in human breast cancer. Cancer Res. 1996;56:3441–5.
- Tanner M, Isola J, Wiklund T, et al. Topoisomerase II alpha gene amplification predicts favorable treatment response to tailored and dose-escalated anthracyclinebased adjuvant chemotherapy in HER-2/neu-amplified breast cancer: Scandinavian Breast Group Trial 9401. J Clin Oncol. 2006;24:2428–36.
- Thurlimann B, Keshaviah A, Coates AS, et al. A comparison of letrozole and tamoxifen in postmenopausal women with early breast cancer. N Engl J Med. 2005;353:2747–57.
- Tognon C, Knezevich SR, Huntsman D, et al. Expression of the ETV6-NTRK3 gene fusion as a primary event in human secretory breast carcinoma. Cancer Cell. 2002; 2:367–76.
- Tubbs RR, Pettay JD, Roche PC, Stoler MH, Jenkins RB, Grogan TM. Discrepancies in clinical laboratory testing of eligibility for trastuzumab therapy: apparent immunohistochemical false-positives do not get the message. J Clin Oncol. 2001;19:2714–21.
- Turner NC, Reis-Filho JS. Basal-like breast cancer and the BRCA1 phenotype. Oncogene. 2006;25:5846–53.
- Turner NC, Reis-Filho JS, Russell AM, et al. BRCA1 dysfunction in sporadic basal-like breast cancer. Oncogene. 2007;26:2126–32.
- Turner N, Pearson A, Sharpe R, et al. FGFR1 amplification drives endocrine therapy resistance and is a therapeutic target in breast cancer. Cancer Res. 2010;70:2085–94.
- van de Vijver MJ, He YD, van't Veer LJ, et al. A geneexpression signature as a predictor of survival in breast cancer. N Engl J Med. 2002;347:1999–2009.
- Van Tine BA, Crowder RJ, Ellis MJ. ER and PI3K independently modulate endocrine resistance in ER positive breast cancer. Cancer Discov. 2011;1:287–8.
- Vici P, Viola G, Botti C, et al. Docetaxel in the adjuvant therapy of HER-2 positive breast cancer patients. Clin Ter. 2008;159:449–52.
- Walton BJ, Morain WD, Baughman RD, Jordan A, Crichlow RW. Cowden's disease: a further indication for prophylactic mastectomy. Surgery. 1986;99:82–6.
- Wang Y, Thakur A, Sun Y, et al. Synergistic effect of cyclin D1 and c-Myc leads to more aggressive and invasive mammary tumors in severe combined immunodeficient mice. Cancer Res. 2007;67:3698–707.
- Wapnir IL, Dignam JJ, Fisher B, et al. Long-term outcomes of invasive ipsilateral breast tumor recurrences after lumpectomy in NSABP B-17 and B-24 randomized clinical trials for DCIS. J Natl Cancer Inst. 2011;103:478–88.
- Watson P, Marcus JN, Lynch HT. Prognosis of BRCA1 hereditary breast cancer. Lancet. 1998;351:304-5.
- Weigelt B, Hu Z, He X, et al. Molecular portraits and 70-gene prognosis signature are preserved throughout the metastatic process of breast cancer. Cancer Res. 2005;65:9155–8.
- Weigelt B, Horlings HM, Kreike B, et al. Refinement of breast cancer classification by molecular characterization of histological special types. J Pathol. 2008;216: 141–50.
- Weigelt B, Kreike B, Reis-Filho JS. Metaplastic breast carcinomas are basal-like breast cancers: a genomic profiling analysis. Breast Cancer Res Treat. 2009;117:273–80.
- Weigelt B, Baehner FL, Reis-Filho JS. The contribution of gene expression profiling to breast cancer classification, prognostication and prediction: a retrospective of the last decade. J Pathol. 2010a;220:263–80.
- Weigelt B, Geyer FC, Reis-Filho JS. Histological types of breast cancer: how special are they? Mol Oncol. 2010b;4:192–208.
- Westbury CB, Reis-Filho JS, Dexter T, et al. Genomewide transcriptomic profiling of microdissected human breast tissue reveals differential expression of KIT (c-Kit, CD117) and oestrogen receptor-alpha (ERalpha) in response to therapeutic radiation. J Pathol. 2009;219:131–40.
- Williard W, Borgen P, Bol R, Tiwari R, Osborne M. Cowden's disease. A case report with analyses at the molecular level. Cancer. 1992;69:2969–74.
- Winer EP, Hudis C, Burstein HJ, et al. American Society of Clinical Oncology technology assessment on the use of aromatase inhibitors as adjuvant therapy for postmenopausal women with hormone receptor-positive breast cancer: status report 2004. J Clin Oncol. 2005;23:619–29.
- Wirapati P, Sotiriou C, Kunkel S, et al. Meta-analysis of gene expression profiles in breast cancer: toward a unified understanding of breast cancer subtyping and prognosis signatures. Breast Cancer Res. 2008;10:R65.
- Wolaniuk A. Hormonal regulation of the development and function of the breasts and mammae. Postepy Hig Med Dosw. 1982;36:375–84.
- Wolff AC, Hammond ME, Schwartz JN, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for human epidermal growth factor receptor 2 testing in breast cancer. J Clin Oncol. 2007;25:118–45.
- Wooster R, Neuhausen SL, Mangion J, et al. Localization of a breast cancer susceptibility gene, BRCA2, to chromosome 13q12-13. Science. 1994;265:2088–90.
- Wooster R, Bignell G, Lancaster J, et al. Identification of the breast cancer susceptibility gene BRCA2. Nature. 1995;378:789–92.