

## Chapter 9

# The Immune System of Breast Milk: Antimicrobial and Anti-inflammatory Properties

Philippe Lepage and Philippe Van de Perre

### Introduction

Human breast milk is recognized as the optimal infant feeding. Human milk contains the nutrients necessary to support the infant's development [1–3]. Breast milk also contains components that protect young children against various infectious diseases and, as more recently described, constituents necessary to support the development of the infant's immune system (ontogeny) [1–3]. This includes various antimicrobial substances, constituents that promote tolerance and priming of the infant immune system, as well as anti-inflammatory components. It has recently become clearer that protection provided through breast milk against some infections extends well beyond weaning [4].

The immune system of the neonate differs considerably from that of an adult [5]. At birth, cells of the innate immune system (dendritic cells, macrophages, and neutrophils) and IgM- and IgG-producing cells are present in the infant's intestinal mucosae, but IgA secreting cells are extremely rare [6]. Because at least 90% of microorganisms infecting human beings use the mucosae as portal of entry [7, 8], young infants are exposed to a large number of microorganisms and are at increased risk for infection [9]. However, the intestinal immune system develops rapidly in the early postnatal period. During this critical period of immunological vulnerability, resistance to infection relies both on the protective factors in milk and on the infant developing his own innate and adaptive immunity. Breast milk provides the missing components while also actively stimulating maturation of the infant's own intestinal defense [3].

In addition to eliminating infectious agents and minimizing the damage they cause, the neonatal immune system must develop the ability to acquire tolerance. This means to discriminate between antigens that are harmless and those that are potentially dangerous. Induction of tolerance is believed to occur primarily in the gut and is facilitated by the specialized B and T cells, the production of secretory IgA (SIgA), and the skewed Th2 response [10, 11]. Failure to regulate tolerance and

---

P. Lepage, M.D., Ph.D. (✉)

Department of Pediatrics, Hôpital Universitaire des Enfants Reine Fabiola,  
Université Libre de Bruxelles, 15, av JJ Crocq, 1020 Brussels, Belgium  
e-mail: philippe.lepage@huderf.be

P. Van de Perre, M.D., Ph.D.

Department of Bacteriology-Virology, INSERM U 1058 "Infection by HIV and by Agents  
with Mucocutaneous Tropism: From Pathogenesis to Prevention", University Montpellier 1,  
CHRU Montpellier, 371 Avenue du Doyen Gaston Giraud, 34295 Montpellier Cedex 5, France  
e-mail: p-van\_de\_perre@chu-montpellier.fr

active immune responses is hypothesized to contribute to food-related allergy, autoimmunity, and inflammatory bowel disorders. Human milk promotes oral tolerance in the infant [12] and exclusive breast-feeding may even prevent or delay the onset of atopic illness such as allergies.

## Immunological Components of Human Milk

### *Breast Milk Cells*

The cellular content of breast milk has certainly not yet revealed all its surprises. Recent studies demonstrated that, particularly in mature milk, epithelial cell adhesion molecule (EPCAM+) cells from the mammary gland are the most represented cell type in breast milk [13]. The exact role of these cells ingested by neonates and breast-feeding infants remains unknown. Stem/progenitor cells have also been identified by their specific surface markers in breast milk [14]. Their physiological role, if any, is equally mysterious.

Excluding epithelial cells, neutrophils comprise roughly 80% of breast milk cells, macrophages 15%, and lymphocytes 4% [15].

### **Lymphocytes**

Various lymphocyte types are coexisting in breast milk: CD3+ T cells (representing roughly 83% of lymphocytes, almost equally distributed in CD4+ and CD8+ lymphocytes), gdT cells (11%), CD16+ NK cells (3–4%), and B cells (2%). Breast milk T and B lymphocytes share three major characteristics that differentiate them from circulating lymphocytes. First, memory T and B lymphocytes are overrepresented in breast milk, the majority of cells failing to express CD45RA receptor characterizing naïve cells [16, 17]. More than 70% of breast milk B cells are IgD-CD27+ memory B cells [18]. Second, many T and B lymphocytes from breast milk are activated, as they frequently express markers such as HLA-DR and CD38 [16–20]. This is paradoxical since human milk per se is remarkably not prone to immune activation. As compared with other media including plasma, blood lymphocytes incubated in the presence of human milk display considerably less frequently activation markers at their surface (E. Tuailon, personal communication). Most likely, breast milk lymphocytes become activated through the extravasation process and/or their transepithelial migration [18–21]. Breast milk B cells are frequently switched memory cells primed to secrete antibodies with overrepresentation of large-sized B cells, plasmablasts, and plasma cells [18] and are not expressing complement receptor [22]. Third, breast milk T [16] and B cells [18] predominantly harbor mucosal homing markers (CD49f, b7 integrin, CD103, CD44) confirming that most of these cells migrated from the highly compartmentalized mucosal-associated lymphoid tissue (MALT) to the mammary gland as an effector site. At least in milk-derived B cells, migration seems to have occurred preferentially from gut-associated lymphoid tissue (GALT) confirming the predominant entero-mammary axis of the mucosal immune system. Migrating B cells, mainly those primed to secrete IgA antibodies, are attracted to and anchored within the mammary acini by the mucosae-associated epithelial chemokine CCL28 [23], other cells being released in breast milk.

CD8+ cytotoxic T lymphocytes can be found in human milk. These cells result from antigenic exposure of maternal mucosal surfaces and are thought to be functional [24–27].

These characteristics of breast milk lymphocytes reinforce the idea that human milk provides neonates and infants with a supplemental, highly immunoreactive system primed to recognize the maternal environment and to protect from potential pathogens the mother–infant dyad may encounter.

**Table 9.1** Distribution of immunoglobulins and other soluble substances in the colostrum and milk delivered to the breast-fed infant during a 24-h period

Soluble product	Concentration in mg/day at postpartum			
	<1 week	1–2 weeks	3–4 weeks	>4 weeks
IgG	50	25	25	10
IgA	5,000	1,000	1,000	1,000
IgM	70	30	15	10
Lysozyme	50	60	60	100
Lactoferrin	1,500	2,000	2,000	1,200

Adapted from ref. 34

## Macrophages

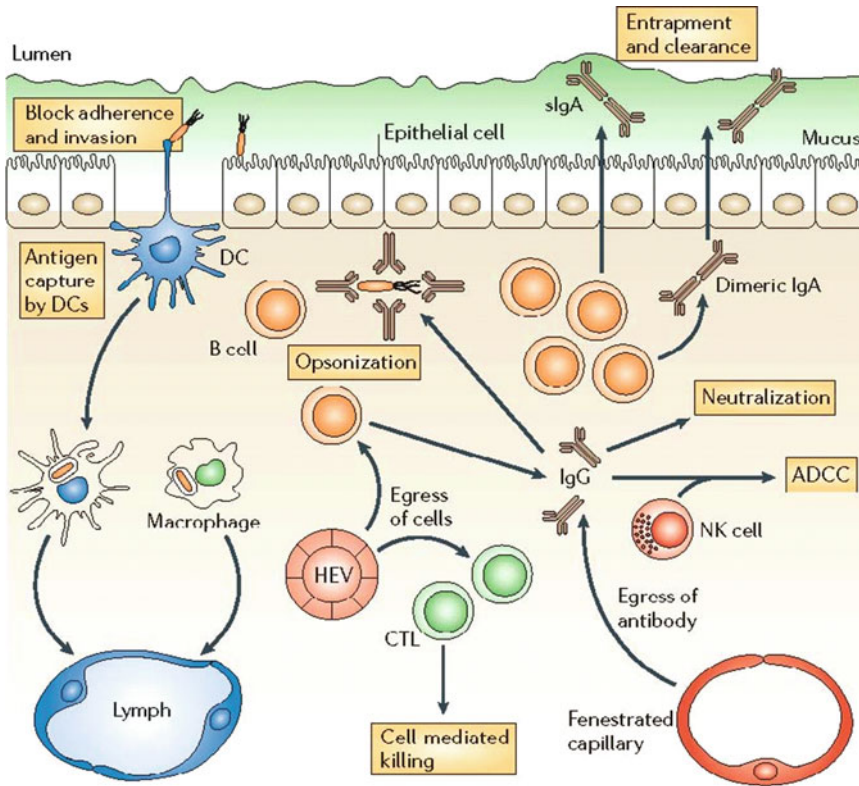
Macrophages have long been considered to represent the major cell type in colostrum and breast milk consisting of 40% of breast milk leukocytes in early lactation and up to 85% in mature milk [19, 28, 29]. However, in these studies, macrophages were defined by morphologic characteristics and by using forward and scatter flow cytometry analyses. More recent studies strongly suggest that the number of breast milk macrophages has been overestimated since only a small proportion of breast milk cells harbor surface markers of macrophages such as CD14 [13–15]. Indeed, macrophages may represent only 15% of breast milk leukocytes [15].

Also, breast milk macrophages are distinct in terms of phenotype and functions from blood monocytes/macrophages conferring them a higher phagocytic capacity and a more efficacious defense against pathogens [30]. Breast milk macrophages are frequently activated [31] and their motility is much enhanced as compared with blood monocytes/macrophages [32]. Breast milk macrophages are morphologically distinct from blood macrophages and spontaneously secrete granulocyte–macrophage colony-stimulating factor (GM-CSF), an important cell growth factor that enhances effector functions and cell signaling pathways [30]. They express DC-SIGN gene and protein (a DC-specific lectin that mediates interaction with HIV-1), and differentiate into CD1+ dendritic cells after incubation with IL-4 [30]. In addition, breast milk macrophages contain secretory IgA that can be released during the phagocytosis process [33] and are profuse secretors of soluble factors such as lactoferrin and complement factors C3 and C4 [29].

## Immunoglobulins

The mammary gland produces SIgA in high quantity with lesser amounts of SIgM and SIgG (Table 9.1). They are one of the predominant proteins in breast milk and found at highest level in colostrum (5 mg/mL in colostrum, about 1 mg/mL in mature milk, [35]). IgA is synthesized in human milk by resident B-cells anchored in the mammary gland tissue through CCL28. These cells have migrated from the mother's intestine via the enteromammary axis [36]. IgA are synthesized as a dimer and linked to a secretory component (SC). SIgA is relatively resistant to proteolytic enzymes from the infant's gastrointestinal tract, allowing to provide a supply of IgA antibodies to the infant's gut [2]. Human milk is not only a rich source of SIgA that could be involved in situ in immune exclusion, but the specificity of the SIgA is directed against microbes common to both the mother and her infant [1].

SIgA provides antimicrobial defense in three ways (Fig. 9.1): preventing bacteria and viruses from attaching to mucosal surfaces by immune exclusion and mucosal painting, as well as neutralizing microbial toxins. Through these mechanisms, SIgA would prevent the establishment of bacterial colonies in the intestine and/or translocation across the mucosal barrier, thereby preventing an inflammatory response that would be damaging to the infant. This provides a potential mechanistic



**Fig. 9.1** Mechanisms of immune protection of human mucosae. From ref. 37, with permission. Abbreviations: ADCC antibody-dependent cellular cytotoxicity, NK cell natural killer cell, CTL cytotoxic lymphocyte responses, DC dendritic cells, sIgA secretory immunoglobulin A, HEV high endothelial venules

explanation for the reduced neonatal septicemia associated with breast-feeding [1]. SIgA have been identified in breast milk against many bacterial pathogens including *Escherichia coli*, *Vibrio cholerae*, *Campylobacter*, *Shigella*, *Giardia lamblia*, *Haemophilus influenzae*, *Clostridium difficile*, and *Streptococcus pneumoniae* [4], viruses such as Rotavirus, CMV, HIV, Influenza virus, respiratory syncytial virus, and yeast [36]. SIgA against maternal microflora and dietary proteins have also been identified in human milk [38]. Vaccination of mothers in late gestation will produce specific SIgA in her breast milk that may protect her infant against exposure.

Animal experiments strongly suggest that SIgA is the crucial protective component of breast milk [39]. Knock-out mice lacking SIgA and SIgM show reduced protection against certain epithelial infections [6]. However, in striking contrast to humans, immunoglobulins from breast milk in many animal species (rodents, bovines, cats, ferrets, etc.) are transported across the intestinal epithelium into the neonatal circulation [40]. This is an evident limitation in translating observations made from animal models into humans.

### Secretory Component

The secretory component (SC) is a glycoprotein present in many external secretions that is usually coupled with polymeric immunoglobulins. SC is also abundant in human milk [41]. It may interfere with the mucosal adhesion of enterotoxigenic *E. coli*, salmonellae, and *C. difficile* toxin A, as well as interact with a pneumococcal surface antigen [6]. SC may inhibit certain toxins, such as *C. difficile* toxin A [6].

## ***Lactoferrin***

Lactoferrin is a predominant whey protein in mature milk ( $\approx 1$  g/L), its maximal concentration is found in colostrum ( $\approx 7$  g/L). The concentration of lactoferrin in breast milk is controlled by the reproductive hormones prolactin and estrogen [1, 3].

Lactoferrin is an iron-binding protein. It is believed that its ability to withhold iron from pathogens explains a great part of its antimicrobial potency [42]. However, this protein has antibacterial activities that are independent of iron chelation. It may also act against some pro-inflammatory cytokines such as IL-1 $\beta$ , TNF- $\alpha$ , and IL-6 [1].

Lactoferrin may inhibit the attachment of bacteria to intestinal cells and has a direct activity against a wide range of Gram-positive and Gram-negative bacteria, fungi, viruses, including HIV-1 [43], and tumor cells [44, 45].

## ***Lysozyme***

Lysozyme is also a major whey protein of human milk. Together with SIgA, lactoferrin, and other antimicrobial compounds, this protein plays a major role in the immune defense of the infant. Lactoferrin removes lipopolysaccharide from the outer cell membrane so that lysozyme can enter and degrade the inner proteoglycan matrix of the membrane of invading cells [46].

## ***Oligosaccharides***

Nondigestible oligosaccharides are an abundant constituent of human milk. They are found in high concentration in colostrum ( $\approx 22$  g/L) and are less abundant in mature milk ( $\approx 12$  g/L). The types of oligosaccharides also change during lactation. Oligosaccharides are produced by an antigen-independent mechanism in epithelial cells in the mammary gland. Oligosaccharides resist hydrolysis by gastro-intestinal enzymes, indicating that they would remain intact in the small intestine.

These compounds function like bacterial receptor analogues by competing with pathogens for binding sites on the epithelial surfaces of the intestine. They can bind pathogens present on the mucosal epithelia, such as *E. coli*, *Campylobacter jejuni*, and *S. pneumoniae* [44].

## ***Other Nonantibody Protein Defense Agents and Other Innate Immune Components***

Many microorganisms express surface molecules that are believed to act like oligosaccharides [47]. These include lactadherin, mucins, and antisecretory lectins.

Lactadherin is a mucin-related glycoprotein produced by mammary epithelial cells during lactation. Its concentration in milk peaks immediately postpartum and declines thereafter. It has been reported to bind human rotavirus by preventing viral attachment to host cell receptors [48, 49]. Its concentration has been inversely associated with rotavirus symptoms in infected breast-fed infants [50].

In addition, several hydrolysis products of casein and of  $\alpha$ -lactalbumin have antimicrobial activity against *E. coli*, *Klebsiella pneumoniae*, staphylococci, streptococci, and *C. albicans* [51].

The protein haptocorrin has been shown in vitro to resist digestion and to inhibit the growth of enterotoxigenic *E. coli* [51].

The secretory leukocyte protease inhibitor (SLPI), a serine protease inhibitor, is present at potentially active concentrations (1,100 ng/mL) in colostrum and transition milk, and it can inhibit HIV-1 entry into host cells in vitro [52].

Human milk contains active lactoperoxidase. The antibacterial properties of lactoperoxidase are closely connected to the oxidation of thiocyanate, both in vitro and in milk.

Mature milk is reported to contain a soluble form of CD14 (sCD14), a cell receptor expressed on macrophages. sCD14 is a glycoprotein that binds bacterial wall components; it is produced by mammary epithelial cells in concentration 20-fold higher than in serum [1].

Several components (C3 and C4), receptors (CF2, CD21), and activation fragments of complement are found in human milk. They might participate in immune bacteriolysis, neutralization of viruses, immune adherence, cytolysis, and enhanced phagocytosis in the infant's intestine.

Human  $\beta$ -defensin-1 has been demonstrated to have antimicrobial activity against *E. coli* and might also be involved in upregulating the adaptative immune system in the gut [53].

Toll-like receptors, lacking in the neonate's gut, have recently been reported in breast milk. They play a crucial "sensing" role in the early innate immune response to invading pathogens [1].

### ***Fatty Acids***

The fat content of breast milk is 3–4 g/L in colostrum, transitional and mature milk, with 93–97% of the lipids in the form of triglycerides. In addition to their nutritive and developmental benefits, milk fats have been demonstrated to provide antimicrobial activity in infant gut [54]. The mechanism for antimicrobial effects of fatty acids and monoglycerides has not been established, but it has been suggested that free fatty acids damage bacteria by disrupting their cell membranes or by changing intracellular pH.

### ***Compounds that Promote "Healthy" Intestinal Microflora***

It is well documented that the colon of breast-fed infants contains fewer potentially pathogenic bacteria, such as *E. coli*, *Bacteroides*, *Campylobacter*, and *Streptococcus*, and more beneficial bacteria, such as *Lactobacillus* and *Bifidobacterium*, as compared to formula-fed infants [55]. Activation of SIgA-producing plasma cells in the young infant's gut is dependent of the colonization of the gut by *Bifidobacteria* and *Lactobacilli* stimulated by fermentation of nondigestible oligosaccharides which are found in large amounts in human milk [10]. These are bifidus factors (compounds that enhance the growth of bifidobacteria in the intestine), prebiotics, and compounds with prebiotic activity that stimulate the growth of beneficial bacteria. Bifidogenic peptides from lactoferrin have been isolated from human milk. Lactoferricin, a cationic peptide derived from digested lactoferrin has been found to be structurally related to certain bifidogenic peptides and is hypothesized to participate in the colonization of newborn infants' flora [56]. The SC of IgA may also have some prebiotic properties [56]. It has also been hypothesized that the long-chain polyunsaturated fatty acids (LCPUFAs) in human milk facilitate the adhesion of probiotics to mucosal surface [57].

### **Effect of Human Milk on the Infant's Immune Development (Table 9.2)**

The infant's intestinal immune system develops rapidly in the early postnatal period as it comes into contact with breast milk components as well as dietary and microbial antigens [6]. During the first 4–6 weeks of life, subepithelial plasma cells are unable to secrete a sufficient amount of SIgA to protect the infant mucosae [6]. This has to be compensated by breast milk intake.

**Table 9.2** Compounds in human milk with possible influence on the infant’s immune development

---

Maternal mammary epithelial cells
Maternal immune cells
Macrophages and dendritic cells
Neutrophils
Natural Killer cells
T-cells
B-cells and their immunoglobulins
Stem/progenitor cells
Cytokines
Nucleotides
Other immune components
Chemokines
Other soluble factors
Long-chain polyunsaturated fatty acids
Compounds that promote microbiological colonization of the infant’s colon
Hormones and bioactive peptides

---

Adapted from ref. 1

**Table 9.3** Cytokines found in human milk

---

TNF- $\alpha$	IL-7
TGF- $\beta$	IL-8
IFN- $\gamma$	IL-10
IL-1 $\beta$	IL-12
IL-2	IL-13
IL-4	IL-16
IL-6	IL-18

---

Adapted from ref. 1

### Cells

The immune cells present in mother’s milk play a pivotal role in bridging the gap between birth and the child’s development of a fully functional immune system.

These cells may secrete immunoregulatory factors such as cytokines. The cytokines may stimulate IgA production by peripheral blood lymphocytes [2]. The same cytokines are found in breast milk, and the presence of transforming growth factor- $\beta$  (TGF- $\beta$ ), IL-6, IL-7, and IL-10 is important for immunologic development and differentiation of IgA-producing cells in the infant [6, 58, 59].

Breast milk neutrophils are also present in activated form, but seem to have limited functional capacity once secreted into milk.

### Cytokines

Human milk contains an array of cytokines, some in concentrations that could potentially influence immune function (Table 9.3). The exact source of the cytokines present in the aqueous fraction of breast milk remains to be determined, though it is suspected that the primary source is from cells present in the mammary gland [60]. The extent to which cytokines survive passage through the infant stomach is largely unknown, but recent work suggests that some cytokines may be sequestered and

protected until they reach the intestine [61]. For example, IL-6 may be unable to survive the passage through the gastro-intestinal tract, unless it is released from milk macrophages in the neonatal gut and thereby enhance the development of the mucosal immunity.

In general, the concentration of cytokines varies widely in human milk, changes through lactation, and is influenced by the mother's health, making it difficult to assess their roles in the development of the infant's immune system.

The intake of cytokines through human milk has the potential to influence the maturation and development of immune cells in the infant. Neonates have a limited ability to produce many cytokines found in breast milk [1]. For example, TGF- $\beta$  is present in breast milk and has been proposed to stimulate appropriate maturation of naïve infant intestinal immune system [4, 62, 63]. IL-6 is hypothesized to enhance the development of the infant's mucosal immunity [6]. Additionally, the cytokines present in milk assist both the transport of maternal leukocytes into milk and across the infant's gut epithelium [64]. There are many factors in breast milk that could either facilitate or inhibit cytokine activities that are not accounted for in studies conducted in vitro.

### ***Hormones***

Little is known about the activity of these compounds on the naïve immune system of an infant when delivered orally. However, it can be suspected that they impact immune development in some synergistic manner [1].

### ***Nucleotides***

Nucleotides are present in small amounts in human milk. It is likely that they play an important role for the rapid early expansion of the lymphoid system as well as the early rapid growth of the gut of the young infant. Dietary nucleotides are reported to benefit the immune system by promoting lymphocyte proliferation, NK activity, macrophage activation, and by producing a variety of other immunomodulatory factors [65]. It is suggested that nucleotides promote a Th1 response and modulate the differentiation of T and B cells [65].

### ***Long-Chain Polyunsaturated Fatty Acids***

Docosahexaenoic acid (DHA) and arachidonic acid (AA) constitute a small fraction of LCPUFAs in human breast milk, but have recently been suggested to participate in immune development [66]. Adding the LCPUFAs DHA and AA to preterm formula resulted in lymphocyte populations and cytokines more similar to human milk-fed infants than to infants who received unsupplemented formula [66].

### **Immune Tolerance**

In addition to eliminating infectious agents and minimizing the damage they cause, the infant's immune system also has to process harmless antigens and has evolved intricate mechanisms to discriminate between antigens with the potential to cause damage and those without (tolerance). Induction of tolerance is believed to occur primarily in the gut by unique features of GALT, including specialized B- and



T-cells, the production of IgA, and the Th2 response [11, 67]. Failure to regulate tolerance and active immune responses can lead to diseases such as food-related allergy, autoimmunity, and inflammatory bowel disorders.

Although it is well established that dietary antigens/potential allergens are present in human milk (reviewed in ref. 68), the consequences on the infant's immune system remain unclear. It is hypothesized that breast milk promotes tolerance to dietary antigens and microflora via immunosuppressive cytokines (IL-10 and TGF- $\beta$ ) and antigens present in breast milk [6].

### ***Compounds in Human Milk that May Be Involved in the Induction of Tolerance***

#### 1. TGF- $\beta$ and IL-10

Acquisition of tolerance involves the downregulation of the immune system through the secretion of cytokines such as TGF- $\beta$  and IL-10. Low levels of TGF- $\beta$  in breast milk have been related to an increased risk of atopic illness in infants, which supports the role of this cytokine in the development of tolerance [67]. Other dietary nutrients may influence the concentration of TGF- $\beta$  in human milk, as levels of TGF- $\beta$  were reported to be positively correlated to polyunsaturated fatty acids (PUFA) content [69]. IL-10 present in breast milk is bioactive. Human milk inhibits the proliferation of human blood lymphocytes in vitro and this inhibition is lessened by adding antihuman IL-10 antibodies in the milk [70]. IL-10 plays also a role in the synthesis of IgA [70].

#### 2. *n*-6 and *n*-3 fatty acids

In human milk, there are roughly ten different long chain (LC)-PUFAs consistently detected, representing both the *n*-3 and the *n*-6 series and including AA and DHA [71]. Maternal intake of PUFA prior to and during lactation is reflected in breast milk PUFA content [72]. Membrane phospholipids fatty acid composition in infants is strongly influenced by maternal diet and can alter the function of the immune cells [73]. Low *n*-6/*n*-3 ratios (due to higher content of *n*-3) in breast milk have been related to lower risk of atopic illness in the breast-fed population [4]. Animal studies suggest that the essential fatty acid content of the maternal diet significantly affects the development of immunological tolerance to food antigens in the suckling offspring [74]. Immunoregulatory benefits have been attributed to *n*-3 LCPUFA [68]. Dietary intervention during critical early stages of immune development before the establishment of allergic responses is thought to be most preferable [68]. Cohort studies have shown lower levels of *n*-3 PUFA in the milk of mothers whose infants showed symptoms of atopic illness before the age of 18 months [68].

### ***Priming of the Immune System***

A balance between tolerance and sensitization (priming) is necessary for the gut immune system to discriminate between harmless antigens and those associated with pathogenic and nonpathogenic microbes [67]. Antigen exposure via mother's milk has been shown to prime the immune response of the suckling pup against that antigen in rats [11]. Clinical trials have shown that breast-fed babies have enhanced specific antibody titer to some, but not all vaccines [4, 5]. The ability to transfer vaccinations from mother to infant via her milk is of great interest because it could eliminate potential problems associated with directly vaccinating the infant [76]. One possible explanation for the ability to immunize infants with their mother's milk has been attributed to the presence of anti-idiotypic antibodies in their breast milk [4]. Anti-idiotypic antibodies are antibodies with specificity against other autologous antibodies [10]. Therefore, anti-idiotypic antibodies, if present in breast milk, could have the capability of priming the infant's antibody response against the antigen the idio type is directed to [40].

## Anti-inflammatory Properties of Breast Milk

Inflammation is a necessary part of the immune response that helps protect the infant from infection. The inflammatory response traps pathogens and signals the arrival of immune cells to destroy the antigen. However, this process results in a great deal of adverse events on healthy tissue if it is not controlled [1].

The immature human intestine overreacts to both indigenous (IL-1 $\beta$ ) and exogenous (Lipopolysaccharide-LPS) inflammatory stimuli to produce an excessive inflammatory (i.e., IL-8) response [3]. In preterm infants, this excessive inflammatory response may contribute to the high incidence of necrotizing enterocolitis. Furthermore, the immature human intestinal epithelium cannot distinguish between pathogens and commensal colonizing bacteria, and reacts to both with an inflammatory response [77]. This immature inflammatory response has to be controlled in order to prevent premature newborns from being in a chronic state of inflammation. Breast milk contains many protective anti-inflammatory components that prevent excessive inflammation until the infant can develop its own mature anti-inflammatory mechanisms.

In a human intestinal model, colostrum whey incubated with a human small intestinal xenograft not only reduces the epithelial secretion of IL-8 (inflammatory response), but also downregulates the luminal expression of TLR-4 expression, thereby reducing innate inflammation.

These studies indicate that breast milk protection from infection could be due in part to anti-inflammatory components in the human milk that prevent inappropriate immune response of the infant against nondeleterious antigens.

There have been very few experimental studies on the anti-inflammatory properties of human milk. Neutrophils are the main immune cells involved in the inflammatory process and *in vitro* studies have shown that human milk can limit the oxidative injury produced by them (reduced cytochrome *c* and consumed H<sub>2</sub>O<sub>2</sub>) [78].

The explanation for the prophylactic nature of human milk is not currently known. However, some components of human milk have potential anti-inflammatory effects; these include cytokines (as well as their receptors and antagonists), antioxidants, antiproteases, and fatty acids [79].

## Cytokines

### IL-10

IL-10 inhibits the production of pro-inflammatory cytokines, providing the necessary balance to ensure that the inflammatory response is limited to destroying the pathogen and not healthy tissue. *In vivo* evidence of the necessity of IL-10 as an anti-inflammatory cytokine is provided by genetically altered mice that are not able to produce IL-10. These mice mount an immune response to the normal microflora in their gut, but without the IL-10 to suppress the inflammation, they develop enterocolitis (similar to ulcerative colitis and celiac disease in humans) [80]. This suggests that IL-10 in human milk might help regulate aberrant immune responses in the infant.

### TGF- $\beta$

TGF- $\beta$  inhibits the production of inflammatory cytokines and promotes the healing of intestinal cells damaged by injury or infection. A feeding trial examining the effectiveness of a diet with proteins and supplemented with TGF- $\beta$  in the management of pediatric Crohn's disease provides the most

convincing *in vivo* evidence of the anti-inflammatory properties of TGF- $\beta$  [81]. The enteral diet containing high levels of TGF- $\beta$  resulted in decreased mucosal IL-1 mRNA (pro-inflammatory cytokine) and clinical remission in 79% of the children [81].

### **IL-1 Receptor Antagonist**

The IL-1 receptor antagonist (IL-1ra) is present in human milk. IL-1ra limits inflammation by competing with IL-1 (pro-inflammatory cytokine) for receptor binding. The reduced inflammatory response in rats with colitis fed human milk compared to formula was similar to the inflammatory response in rats fed infant formula supplemented with IL-1ra [82]. These results suggest that the IL-1ra content of human milk contributes to its anti-inflammatory properties.

### ***Antioxidants***

Free radicals, or reactive oxygen species, are produced during the normal metabolic activity of cells. These free radicals can damage cells by lipid peroxidation and alteration of protein and/or nucleic acid structures leading to oxidative stress [83]. Antioxidants in both milk and formula prevent significant lipid oxidation, breast milk suppresses oxidative DNA damage better than formula does. We have yet to identify all of the compounds in human milk that have antioxidant properties; however, there are several antioxidants in human milk that can scavenge free radicals and thereby limit the damage caused by oxidative stress. These compounds include  $\alpha$ -tocopherol,  $\beta$ -carotene, cysteine, ascorbic acid, catalase, and glutathione peroxidase. The ability of human milk to resist oxidative stress is greater than formula [84]. *In vitro* studies have shown that human milk degrades the naturally occurring hydrogen peroxide as well as that produced by neutrophils. This is possibly due to the catalase content of human milk [79].

Lactoferrin has been shown to inhibit the production of proinflammatory cytokines (IL-6 and TNF- $\alpha$ ) as well as inflammatory mediators (nitric oxide, GM-CSF [4]). The anti-inflammatory activity of lactoferrin is generally attributed to its ability to search out free iron, which is a potent oxidizer.

### ***Antiproteases***

Inflammatory cells produce proteases, which allow the cells to enter the affected area. Some pathogens also produce proteases in order to enter the body. Human milk contains active protease inhibitors (e.g.,  $\alpha$ -1-antitrypsin,  $\alpha$ -1-antichymotrypsin, and elastase inhibitor) that can limit the ability of pathogens to gain entry into the body and limit the inflammation caused by the inflammatory response.

### ***Long-Chain Polyunsaturated Fatty Acids***

It is hypothesized that the effects of LCPUFA *n*-3 fatty acids on immune function are mediated by their ability to compete with the metabolism of the *n*-6 fatty AA [85]. AA can be metabolized into the pro-inflammatory prostaglandin-E2 (PGE2) or leukotriene-B4 (LTB4) [86].

Prostaglandin E2 is one of the most important prostaglandins formed as it initiates the typical symptoms associated with inflammation: pain, fever, and swelling [87]. The metabolism of AA to

yield PGE2 and LTB4 can be inhibited by DHA, thereby decreasing the capacity of immune cells to synthesize eicosanoids from AA. DHA will then give rise to PGE3 and LTB5, which are considered less biologically potent than the eicosanoids derived from AA. However, their activities have not yet been fully investigated.

## ***Others***

Other substances in breast milk can also produce anti-inflammatory effects in the newborn intestine. For example, lactoferrin can reduce the production of inflammatory cytokines in monocytes by inhibiting nuclear factor kappa light-chain enhancer of activated B cell (NFκB) activation.

## **Protection Against Infection**

The adoption of exclusive breast-feeding for the first 6 months of life has been estimated to be the single most effective preventive strategy for saving lives of young children in low-income settings, with a potential estimated reduction of 13% in infant mortality rate [82]. A large part of this protective effect can contribute to a dramatic reduction in infectious diseases incidence, as it has been observed for decades [88–92].

Protection by breast-feeding against death, hospitalization, diarrheal diseases, and acute lower respiratory tract infections (ALRI) is undisputed in children living in conditions of poor hygiene, where mucosal pathogens are major killers in young children.

In developing nations, it has been demonstrated for many years that the risk of dying from diarrhea is dramatically reduced in breast-fed children [7, 93–95]. For example, Scrimshaw and Taylor [96], in the years 1955–1959, demonstrated that the infant mortality rate in villages from Punjab, India, was 950 per 1,000 live births in formula-fed babies compared with 120 in the breast-fed ones. In Rwanda, case-fatality rates for diarrhea, pneumonia, and measles were significantly lower in breastfed, than in completely weaned, hospitalized children, for all three diseases [92].

Additional data from developing countries have confirmed and extended these early observations [97–106]. In communities with a high prevalence of malnutrition, such as rural Bangladesh, breast-feeding may substantially enhance child survival up to 3 years of age [98]. Lack of breast-feeding is an important risk factor for pneumonia and ALRI mortality in developing countries [101, 103–105]. In a population-based study of infant mortality in two urban areas of southern Brazil, the type of milk in an infant's diet was found to be an important risk factor for deaths from diarrheal and respiratory infections [102]. Compared with infants who were breast-fed with no milk supplements, and after adjusting for confounding variables, those completely weaned had 3.6 times the risk of death from respiratory infections [102]. In a prospective observational study conducted in slum areas of Dhaka, Bangladesh [103], partial or no breast-feeding was associated with a 2.23-fold higher risk of infant deaths resulting from all causes and 2.40- and 3.94-fold higher risk of deaths attributable to ALRI and diarrhea, respectively, when compared with exclusive breast-feeding. Betrán et al. [104] have strongly suggested that 5% of infant deaths from diarrheal disease and acute respiratory infections in Latin America are preventable by exclusive breast-feeding among infants aged 0–3 months and partial breast-feeding throughout the remainder of infancy. A strong association between delayed initiation of breast-feeding and increased neonatal mortality has been shown in a large observational study in rural Ghana [99, 100].

A beneficial effect of breast-feeding has also been demonstrated in the industrialized world. Numerous careful studies in children from wealthy nations have now confirmed that breast-feeding

protects against several common diseases such as diarrhea, acute otitis media (AOM), respiratory tract infections, invasive *Haemophilus influenzae* type B infections, neonatal septicaemia, and necrotising enterocolitis [6, 8, 107–109]. Chen and Rogan [110] have assessed the effect of breast-feeding on postneonatal mortality in the USA using recent (1988) National Maternal and Infant Health Survey data. Breast-feeding was associated with a statistically significant reduction in the risk for postneonatal death [104]. In a study of the epidemiology of middle ear disease in Massachusetts children, absence of breast-feeding was a risk factor for AOM and/or recurrent AOM (together with male gender, a sibling history of recurrent AOM, and early onset of otitis) [111].

In industrialized countries, breast-feeding has been shown to reduce the severity of lower respiratory infection in the first 6 months of life and an inverse relation between the duration of ALRI symptoms and the length of exclusive breast-feeding has been demonstrated [112]. In these countries, an effect is seen principally among infants living in crowded conditions in lower socioeconomic strata [112].

In the Republic of Belarus, an experimental intervention (modeled on the Baby-Friendly Hospital Initiative of WHO and UNICEF and emphasizing health care worker assistance with initiating and maintaining breast-feeding and lactation and postnatal breast-feeding support) was shown to decrease the risk of gastrointestinal tract infection in the first year of life [113]. In the UK, breastfeeding, particularly when exclusive and prolonged, protected against severe morbidity from diarrhea and ALRI in a large cohort of healthy, singleton, term infants born from 2000 to 2002 [114]. In Alicante, Spain, full breast-feeding was shown to lower the risk for hospital admission as a result of infections among infants younger than 1 year [115].

## Conclusion

Human milk is extraordinary complex in composition. Notably, its content evolves over time. In early lactation phases, SIgA, SIgM, antiinflammatory factors and, most probably immunologically active cells provided by breast milk are substituting for the relatively immature neonatal immune system. Thereafter, breast milk continues to adapt remarkably to the infant ontogeny, to its immune protection needs and to its nutritional requirements. In particular, breast-milk antibodies are targeted against potentially deleterious infectious agents and antigens that are present in the maternal environment, which are those likely to be encountered by the infant. In that sense, it is legitimate to consider the maternal and the infant's immune systems as a continuum, with the placenta and the mammary glands/breast milk as interfaces, now known as the mother–offspring immune dyad [116].

## References

1. Blewett HJH, Cicalo MC, Holland CD, Field CJ (2008) The immunological components of human milk. *Adv Food Nutr* 54:45–80
2. Brandtzaeg P (2010) The mucosal immune system and its integration in the mammary glands. *J Pediatr* 156:S8–15
3. Walker W (2010) Breast milk as the gold standard for protective nutrients. *J Pediatr* 156:S3–S7
4. Hanson LA, Korotkova M, Lundin S et al (2003) The transfer of immunity from mother to child. *Ann NY Acad Sci* 987:199–206
5. Kelly D, Coutts AG (2000) Early nutrition and the development of immune function in the neonate. *Proc Nutr Soc* 59:177–185
6. Brandtzaeg P (2003) Mucosal immunity: integration between mother and the breast-fed infant. *Vaccine* 21: 3382–3388
7. Anonymous (1994) A warm chain for breastfeeding. *Lancet* 344:1239–1241
8. Evidence Report/Technology Assessment Report No. 153, U.S. Department of Health: breastfeeding and maternal and infant health outcomes in developed countries. Agency for Healthcare Research and Quality (AHRQ), Publication No. 07-E007, April 2007

9. Brandtzaeg P, Nilssen DE, Rognum TO, Thrane PS (1991) Ontogeny of the mucosal immune system and IgA deficiency. *Gastroenterol Clin North Am* 20:397–439
10. Field CJ (2005) The immunological components of human milk and their effect on the immune development in infants. *J Nutr* 135:1–4
11. Strobel S (2001) Immunity induced after a feed of antigen during early life: oral tolerance v. sensitisation. *Proc Nutr Soc* 60:437–442
12. van Odijk J, Kull I, Borres MP et al (2003) Breastfeeding and allergic disease: a multidisciplinary review of the literature (1966–2001) on the mode of early feeding in infancy and its impact on later atopic manifestations. *Allergy* 58:833–843
13. Petitjean G, Becquart P, Tuaille E et al (2007) Isolation and characterization of resting CD4+ T lymphocytes in breast milk. *J Clin Virol* 39:1–8
14. Fan Y, Chong YS, Choolani MA, Cregan MD, Chan JKY (2010) Unravelling the mystery of stem/progenitor cells in human breast milk. *PLoS One* 5:e14421
15. Goldman AS, Chheda S, Garofalo R (1998) Evolution of immunologic functions of the mammary gland and the postnatal development of immunity. *Pediatr Res* 43:155–162
16. Bertotto A, Gerli R, Fabietti G et al (1990) Human breast milk T cells display the phenotype and functional characteristics of memory T cells. *Eur J Immunol* 20:1877–1880
17. Valéa D, Tuaille E, Al Tabaa Y et al (2011) CD4+ T cells spontaneously producing human immunodeficiency virus type I in breast milk from women with or without antiretroviral drugs. *Retrovirology* 8:34
18. Tuaille E, Valea D, Becquart P et al (2009) Human milk-derived B cells: a highly activated switched memory cell population primed to secrete antibodies. *J Immunol* 182:7155–7162
19. Wirt DP, Adkins LT, Palkowetz KH et al (1992) Activated and memory T lymphocytes in human milk. *Cytometry* 13:282–290
20. Richie ER, Steinmetz KD, Meistrich ML, Ramirez I, Hilliard JK (1980) T lymphocytes in colostrum and peripheral blood differ in their capacity to form thermostable E-rosettes. *J Immunol* 125:2344–2346
21. Richie ER, Bass R, Meistrich ML, Dennison DK (1982) Distribution of T lymphocyte subsets in human colostrum. *J Immunol* 129:1116–1119
22. Bush JF, Beer AE (1979) Analysis of complement receptors on B-lymphocytes in human milk. *Am J Obstet Gynecol* 133:708–712
23. Wilson E, Butcher EC (2004) CCL28 controls immunoglobulin (Ig)A plasma cell accumulation in the lactating mammary gland and IgA antibody transfer to the neonate. *J Exp Med* 200:805–809
24. Losonsky GA, Fishaut JM, Strussenberg J, Ogra PL (1982) Effect of immunization against rubella on lactation products. I. Development and characterization of specific immunologic reactivity in breast milk. *J Infect Dis* 145:654–660
25. Ruben FL, Holzman IR, Fireman P (1982) Responses of lymphocytes from human colostrum or milk to influenza antigens. *Am J Obstet Gynecol* 143:518–522
26. Sabbaj S, Edwards BH, Ghosh MK et al (2002) Human immunodeficiency virus-specific CD8(+) T cells in human breast milk. *J Virol* 76:7365–7373
27. Lohman BL, Slyker J, Mbori-Ngacha D et al (2003) Prevalence and magnitude of human immunodeficiency virus (HIV) type 1-specific lymphocyte responses in breast milk from HIV-1-seropositive women. *J Infect Dis* 188:1666–1674
28. Crago SS, Prince SJ, Pretlow TG et al (1979) Human colostrum cells. I. Separation and characterization. *Clin Exp Immunol* 38:585–597
29. Pitt J (1979) The milk mononuclear phagocyte. *Pediatrics* 64:745–749
30. Ichikawa M, Sugita M, Takahashi M, Satomi M, Takeshita T, Araki T, Takahashi H (2003) Breast milk macrophages spontaneously produce granulocyte-macrophage colony-stimulating factor and differentiate into dendritic cells in the presence of exogenous interleukin-4 alone. *Immunology* 108:189–195
31. Rivas RA, El Mohandes AA, Katona IM et al (1994) Mononuclear phagocytic cells in human milk: HLA-DR and Fc $\gamma$ R ligand expression. *Biol Neonate* 66:195–204
32. Ozkaragöz F, Rudloff HB, Rajaraman S, Mushtaha AA, Schmalstieg FC, Goldman AS (1988) The motility of human milk macrophages in collagen gels. *Pediatr Res* 23:449–452
33. Weaver EA, Goldblum RM, Davis CP, Goldman AS (1981) Enhanced immunoglobulin A release from human colostrum cells during phagocytosis. *Infect Immun* 34:498–502
34. Ogra SS, Ogra PL (1978) Immunologic aspects of human colostrum and milk. I. Distribution characteristics and concentrations of immunoglobulins at different times after the onset of lactation. *J Pediatr* 92:546–549
35. Goldman AS, Garza C, Nichols BL, Goldblum RM (1982) Immunologic factors in human milk during the first year of lactation. *J Pediatr* 100:563–567
36. Hanson LA, Korotkova M (2002) The role of breast-feeding in prevention of neonatal infection. *Semin Neonatol* 7:275–281
37. Neutra MR, Kozlowski PA (2006) Mucosal vaccines: the promise and the challenge. *Nat Rev Immunol* 6:148–158

38. Hanson LA, Korotkova M, Telemo E (2003) Breast-feeding, infant formulas and the immune system. *Ann Allergy Asthma Immunol* 90:59–63
39. Dickinson EC, Gorga JC, Garrett M et al (1998) Immunoglobulin A supplementation abrogates bacterial translocation and preserves the architecture of the intestinal epithelium. *Surgery* 124:284–290
40. Van de Perre P (2003) Transfer of antibody via mother's milk. *Vaccine* 21:3374–3376
41. Brandtzaeg P (1983) The secretory immune system of lactating human mammary glands compared with other exocrine organs. *Ann NY Acad Sci* 409:353–381
42. Lonnerdal B (2003) Nutritional and physiologic significance of human milk proteins. *Am J Clin Nutr* 77:1537S–1543S
43. Harmsen MC, Swart PJ, de Bethune MP et al (1995) Antiviral effect of plasma and milk proteins: lactoferrin shows potent activity against both human immunodeficiency virus and cytomegalovirus replication in vitro. *J Infect Dis* 172:380–388
44. Shah NP (2000) Effects of milk-derived bioactives: an overview. *Br J Nutr* 84(suppl 1):S3–S10
45. Bernt KM, Walker WA (1999) Human milk as carrier of biochemical message. *Acta Paediatr Suppl* 88:27–41
46. Ellison RT III, Giehl TJ (1991) Killing of gram-negative bacteria by lactoferrin and lysozyme. *J Clin Invest* 88:1080–1091
47. Gopal PK, Gill HS (2000) Oligosaccharides and glycoconjugates in bovine milk and colostrum. *Br J Nutr* 84(suppl 1):S69–S74
48. Kvistgaard AS, Pallesen LT, Arias CF, Lopez S, Petersen TE, Heegaard CW, Rasmussen JT (2004) Inhibitory effects of human and bovine milk constituents on rotavirus infections. *J Dairy Sci* 87:4088–4096
49. Yolken RH, Peterson JA, Vonderfecht SL, Fouts ET, Midthun K, Newburg DS (1992) Human milk mucin inhibits rotavirus replication and prevents experimental gastroenteritis. *J Clin Invest* 90:1984–1991
50. Newburg DS, Peterson JA, Ruiz-Palacios GM et al (1998) Role of human-milk lactadherin in protection against symptomatic rotavirus infection. *Lancet* 351:1160–1164
51. Pellegrini A, Thomas U, Bramaz N, Hunziker P, von Fellenberg R (1999) Isolation and identification of three bactericidal domains in the bovine alpha-lactalbumin molecule. *Biochim Biophys Acta* 1426:439–448
52. Wahl SM, McNeely TB, Janoff EN et al (1997) Secretory leukocyte protease inhibitor (SLPI) in mucosal fluids inhibits HIV-1. *Oral Dis* 3(suppl 1):S64–69
53. Jia HP, Starner T, Ackermann M, Kirby P, Tack BF, McCray PB Jr (2001) Abundant human beta-defensin-1 expression in milk and mammary gland epithelium. *J Pediatr* 138:109–112
54. German JB, Dillard CJ (2006) Composition, structure and absorption of milk lipids: a source of energy, fat-soluble nutrients and bioactive molecules. *Crit Rev Food Sci Nutr* 46:57–92
55. Kleessen B, Bunke H, Tovar K, Noack J, Sawatzki G (1995) Influence of two infant formulas and human milk on the development of the faecal flora in newborn infants. *Acta Paediatr* 84:1347–1356
56. Liepke C, Adermann K, Raida M, Magert HJ, Forssmann WG, Zucht HD (2002) Human milk provides peptides highly stimulating the growth of bifidobacteria. *Eur J Biochem* 269:712–718
57. Das UN (2002) Essential fatty acids as possible enhancers of the beneficial actions of probiotics. *Nutrition* 18:786
58. Brandtzaeg P, Johansen F-E (2005) Mucosal B cells: phenotypic characteristics, transcriptional regulation, and homing properties. *Immunol Rev* 206:32–63
59. Hanson LA (2007) Breast-feeding and immune function. *Proc Nutr Soc* 66:384–396
60. Bryan DL, Forsyth KD, Gibson RA, Hawkes JS (2006) Interleukin-2 in human milk: a potential modulator of lymphocyte development in breastfed infants. *Cytokine* 33:289–293
61. Calhoun DA, Lunoe M, Du Y, Staba SL, Christensen RD (1999) Concentrations of granulocyte colony-stimulating factor in human milk after in vitro stimulations of digestion. *Pediatr Res* 46:767–771
62. Bottcher MF, Jenmalm MC, Garofalo RP, Bjoksten B (2000) Cytokines in breast milk from allergic and non allergic mothers. *Pediatr Res* 47:157–162
63. Donnet-Hughes A, Duc N, Serrant P, Vidal K, Schiffrin EJ (2000) Bioactive molecules in milk and their role in health and disease: the role of transforming growth factor-beta. *Immunol Cell Biol* 78:74–79
64. Ustundag B, Yilmaz E, Dogan Y et al (2005) Levels of cytokines (IL-1beta, IL-2, IL-6, IL-8, TNF-alpha) and trace elements (Zn, Cu) in breast milk from mothers of preterm and term infants. *Mediators Inflamm* 2005:331–336
65. Aggrett P, Leach JL, Rueda R, MacLean J (2003) Innovation in infant formula development: a reassessment of ribonucleotides in 2002. *Nutrition* 19:375–384
66. Field CJ, Clandinin MT, Van Aerde JE (2001) Polyunsaturated fatty acids and T-cell function: implications for the neonate. *Lipids* 36:1025–1032
67. Mowat AM (2003) Anatomical basis of tolerance and immunity to intestinal antigens. *Nat Rev Immunol* 3:331–341
68. Palmer DJ, Makrides M (2006) Diet of lactating women and allergic reactions in their infants. *Curr Opin Clin Nutr Metab Care* 9:284–288
69. Laiho K, Lampi AM, Hamalainen M, Moilanen E, Piironen V, Arvola T, Syrjanen S, Isolauri E (2003) Breast milk fatty acids, eicosanoids, and cytokines in mothers with and without allergic disease. *Pediatr Res* 53:642–647

70. Garofalo R, Chheda S, Mei F et al (1995) Interleukin-10 in human milk. *Pediatr Res* 37:444–449
71. Koletzko B, Rodriguez-Palmero M, Demmelmair H, Fidler N, Jensen R, Sauerwald T (2001) Physiological aspects of human milk lipids. *Earl Hum Dev* 65 Suppl S3-S18
72. Hawkes JS, Bryan DL, Gibson RA (2002) Cytokine production by hm cells and peripheral blood mononuclear cells from the same mothers. *J Clin Immunol* 22:338–344
73. Field CJ, Thomson CA, Van Aerde JE, Parrott A, Euler A, Lien E, Clandinin MT (2000) Lower proportion of CD45RO+ cells and deficient interleukin-10 production by formula-fed infants, compared with human-fed, is corrected with supplementation of long-chain polyunsaturated fatty acids. *J Pediatr Gastroenterol Nutr* 31:291–299
74. Korotkova M, Telemo E, Hanson LA, Strandvik B (2004) Modulation of neonatal immunological tolerance to ovalbumin by maternal essential fatty acid intake. *Pediatr Allergy Immunol* 15:112–122
75. Harbig LS, Fisher BA (2001) Dietary fatty acid modulation of mucosally-induced tolerogenic immune responses. *Proc Nutr Soc* 60:449–456
76. Gust DA, Strine TW, Maurice E et al (2004) Underimmunization among children: effects of vaccine safety concerns on immunization status. *Pediatrics* 114:e16–e22
77. Goldman AS (1993) The immune system of human milk: antimicrobial, anti-inflammatory and immunomodulating properties. *Pediatr Infect Dis J* 12:664–671
78. Grazioso CF, Buescher ES (1996) Inhibition of neutrophil function by human milk. *Cell Immunol* 168:125–132
79. Garofalo RP, Goldman AS (1999) Expression of functional immunomodulatory and anti-inflammatory factors in human milk. *Clin Perinatol* 26:361–377
80. Sydora BC, Tavernini MM, Wessler A, Jewell LD, Fedorak RN (2003) Lack of interleukin-10 leads to intestinal inflammation, independent of the time at which luminal microbial colonization occurs. *Inflamm Bowel Dis* 9:87–97
81. Fell JM, Paintin M, Arnaud-Battandier F et al (2000) Mucosal healing and a fall in mucosal pro-inflammatory cytokine mRNA induced by a specific oral polymeric diet in paediatric Crohn's disease. *Aliment Pharmacol Ther* 14:281–289
82. Grazioso CF, Werner AL, Alling DW, Bishop PR, Buescher ES (1997) Antiinflammatory effects of human milk on chemically induced colitis in rats. *Pediatr Res* 42:639–643
83. Sharda B (2006) Free radicals: emerging challenge in environmental health research in childhood and neonatal disorders. *Int J Environ Res Public Health* 3:286–291
84. Friel JK, Martin SM, Langdon M, Herzberg GR, Buettner GR (2002) Milk from mothers of both premature and full-term infants provides better antioxidant protection than does infant formula. *Pediatr Res* 51:612–618
85. Gottrand F (2008) Long-chain polyunsaturated fatty acids influence the immune system of infants. *J Nutr* 38:1807S–1812S
86. Calder PC, Grimble RF (2002) Polyunsaturated fatty acids, inflammation and immunity. *Eur J Clin Nutr* 56(suppl 3):S14–S19
87. Wahle KW, Heys SD, Rotondo D (2004) Conjugated linoleic acids: are they beneficial or detrimental to health? *Prog Lipid Res* 43:553–587
88. Jones G, Steketee RW, Black RE, Bhutta ZA, Morris SS (2003) How many child deaths can we prevent this year? *Lancet* 362:65–71
89. Chien PF, Howie PW (2001) Breast milk and the risk of opportunistic infection in infancy in industrialized and non-industrialized settings. *Adv Nutr Res* 10:69–104
90. Cunningham AS, Jelliffe DB, Jelliffe EF (1991) Breastfeeding and health in the 1980s: a global epidemiologic review. *J Pediatr* 118:659–666
91. Grulee C, Sandord H, Schwartz H (1935) Breast and artificially fed infants; study of the age incidence in the morbidity and mortality in 20,000 cases. *JAMA* 104:1986–1988
92. Lepage P, Munyakazi C, Hennart P (1981) Breastfeeding and hospital mortality in children in Rwanda. *Lancet* 2:409–11
93. Feachem R, Koblinski MA (1984) Interventions for the control of diarrhoeal diseases among young children: promotion of breast-feeding. *Bull WHO* 62:271–291
94. Plank SJ, Milanese MI (1973) Infant feeding and infant mortality in rural Chile. *Bull WHO* 48:203–210
95. Puffer RR, Serrano CV (1973) Patterns of mortality in childhood. Scientific publication no. 262. Pan American Health Organization 1973, Washington DC
96. Scrimshaw NS, Taylor CE (1968) Interaction of nutrition and infection. WHO monograph 1968; No. 29, Geneva
97. Bennish ML, Harris JR, Wojtyniak BJ, Struelens M (1990) Death in shigellosis: incidence and risk factors in hospitalized patients. *J Infect Dis* 16:500–506
98. Briend A, Wojtyniak B, Rowland MGM (1988) Breast feeding, nutritional state, and child survival in rural Bangladesh. *BMJ* 296:879–882
99. Edmond KM, Zandoh C, Quigley MA, Amenga-Etego S, Owusu-Agyei S, Kirkwood BR (2006) Delayed breast-feeding initiation increases risk of neonatal mortality. *Pediatrics* 117:380–386



100. Edmond KE, Kirkwood BR, Amenga-Etego S, Owusu-Agyei S, Hurt LS (2007) Effect of early infant feeding practices on infection-specific neonatal mortality: an investigation of the causal links with observational data from rural Ghana. *Am J Clin Nutr* 86:1126–1131
101. Victora CG, Kirkwood BR, Ashworth A et al (1999) Potential interventions for the prevention of childhood pneumonia in developing countries: improving nutrition. *Am J Clin Nutr* 70:309–320
102. Victora CG, Smith PG, Vaughan JP et al (1987) Evidence for protection by breast-feeding against infant deaths from infectious diseases in Brazil. *Lancet* 2(8554):319–322
103. Arifeen S, Black RE, Antelman G, Baqui A, Caulfield L, Becker S (2001) Exclusive breastfeeding reduces acute respiratory infection and diarrhea deaths among infants in Dhaka slums. *Pediatrics* 108:e67
104. Betrán AP, de Onís M, Lauer JA, Villar J (2001) Ecological study of effect of breast feeding on infant mortality in Latin America. *BMJ* 323:1–5
105. Yoon PW, Black RE, Moulton LH, Becker S (1996) Effect of not breastfeeding on the risk of diarrheal and respiratory mortality in children under 2 years of age in Metro Cebu, The Philippines. *Am J Epidemiol* 143:1142–1148
106. Ashraf RN, Jalil F, Zaman S et al (1991) Breastfeeding and protection against neonatal sepsis in a high risk population. *Arch Dis Child* 66:488–490
107. Hanson LA, Silfverdal S-A, Stromback L, Erling V, Zaman S, Olcén P, Telemo E (2001) The immunological role of breast feeding. *Pediatr Allergy Immunol* 12(suppl 14):15–19
108. Hanson L, Telemo E (2008) Immunobiology and epidemiology of breastfeeding in relation to prevention of infections from a global perspective. In: Ogra P, Harris WS (eds) *n-3 fatty acids in health: DaVinci's code*. *Am J Clin Nutr* vol 88, pp. 595–596
109. Howie PW, Forsyth JS, Ogston SA, Clark A, Florey CD (1990) Protective effect of breast feeding against infection. *BMJ* 300:11–16
110. Chen A, Rogan WJ (2004) Breastfeeding and the risk of postneonatal death in the United States. *Pediatrics* 113:e435–e439
111. Pelton SI, Leibovitz E (2009) Recent advances in otitis media. *Pediatr Infect Dis J* 28:S133–S137
112. Simoes EAF (2003) Environmental and demographic risk factors for respiratory syncytial virus lower respiratory tract disease. *J Pediatr* 143:S118–S126
113. Kramer MS, Chalmers B, Hodnett ED et al (2001) Promotion of breastfeeding intervention trial (PROBIT). A randomized trial in the Republic of Belarus. *JAMA* 285:413–420
114. Quigley MA, Kelly YQ, Sacker A (2007) Breastfeeding and hospitalization for diarrheal and respiratory infection in the United Kingdom Millennium Cohort Study. *Pediatrics* 119:e837–e842
115. Talayero JMP, Lizán-García M, Puime AO et al (2006) Full breastfeeding and hospitalization as a result of infections in the first year of life. *Pediatrics* 118:e92–e99
116. Hanson LA (2000) The mother-offspring dyad and the immune system. *Acta Paediatr* 89:252–258