Platelets and Coronary Artery Disease

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Introduction

Platelets play a critical role in thrombosis and hemostasis [1, 2]. Beyond that, platelets have been recognized to act as an interface between thrombosis and inflammation [3]. Previous research has shown that inflammation plays a key role in the development of coronary artery disease (CAD) and other manifestations of atherosclerosis [4]. Recently, platelets have been recognized to trigger atherosclerotic lesion formation, to favor plaque instability, and to form thrombosis at areas of vulnerable plaques, resulting in myocardial infarction and tissue ischemia. During atherogenesis platelets interact with a variety of vascular and blood-borne cells (e.g., endothelial cells and leukocytes) and regulate chemotaxis, migration, and cytokine/chemokine release, which further propagate inflammation within vascular lesions [3, 5]. Understanding the role of platelets in atherogenesis will allow developing new strategies in the treatment of coronary artery disease.

Platelets Interact with Endothelium and Propagate Inflammation

In the past, numerous studies have shown that platelets can adhere to the intact endothelial monolayer and substantially modulate endothelial cell function [6–8]. Thus, under certain pathophysiological circumstances, endothelial denudation and exposure of the subendothelial matrix are not required for platelet adhesion to the vascular wall. Adherent platelets release a variety of proinflammatory mediators and growth hormones and have the potential to modify signaling cascades in vascular cells, inducing the expression of endothelial adhesion receptors and the release of endothelial chemoattractants [3]. In this

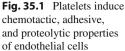
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manner, they might regulate the adhesion and infiltration of leukocytes, in particular of monocytes, into the vascular wall, a process, that is thought to play a key role in acute and chronic inflammation. Normal "resting" endothelium represents a nonadhesive and non-thrombogenic surface that prevents extravasation of circulating blood cells. In contrast, activated endothelial cells are pro-adhesive and promote the adhesion of circulating blood platelets. Adhesion of platelets to the intact but activated endothelium in the absence of previous endothelial denudation involves a surface receptor-dependent process that allows "capturing" of circulating platelets toward the vessel wall even under high shear stress. Similar to the recruitment of leukocytes [5], the adhesion of platelets to the vascular endothelial surface is a multistep process in which platelets are tethered to the vascular wall followed by platelet rolling and subsequent firm adhesion. Whereas the adhesion receptors involved in platelet attachment to the subendothelial matrix, e.g., following rupture of an atherosclerotic plaque, have been well defined during the past decade, few studies have focused on the molecular determinants that promote the interaction between platelets and the intact vascular endothelium.

During the adhesion process, platelets are activated and release an arsenal of potent proinflammatory and promitogenic substances into the local microenvironment, thereby altering chemotactic, adhesive, and proteolytic properties of endothelial cells (Fig. 35.1). These platelet-induced alterations of the endothelial phenotype support chemotaxis, adhesion, and transmigration of monocytes to the site of inflammation. Among the various platelet-derived proinflammatory proteins, IL-1 β has been identified as a major mediator of platelet-induced activation of endothelial cells. The IL-1 β activity expressed by platelets appears to be associated with the platelet surface [9], and co-incubation of endothelial cells with thrombin-activated platelets induces IL-1β-dependent secretion of IL-6 and IL-8 from endothelial cells. Furthermore, incubation of cultured endothelial cells with thrombin-stimulated platelets significantly enhances the secretion of endothelial monocyte chemoattractant protein-1 (MCP-1) in an IL-1 β -dependent manner [10, 11].

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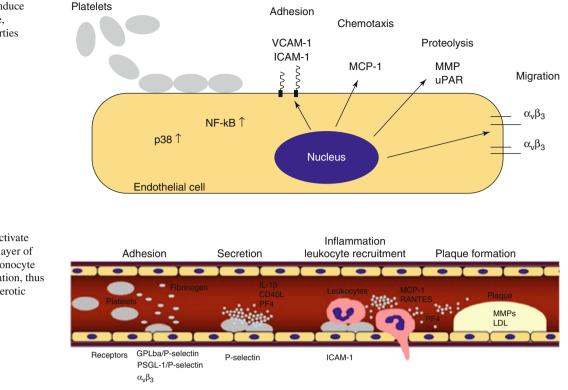


Fig. 35.2 Platelets activate the endothelial monolayer of arteries and induce monocyte chemotaxis and migration, thus propagating atherosclerotic lesion formation

However, platelet IL-1 β not only modifies endothelial release of chemotactic proteins, but it also has the potential to increase endothelial expression of adhesion molecules. Surface expression of ICAM-1 and $\alpha_{1}\beta_{2}$ on endothelial cells is significantly enhanced by activated platelets via IL-1ß [10, 12]. Both enhanced chemokine release and upregulation of endothelial adhesion molecules through platelet-derived IL-1β act in concert and promote neutrophil and monocyte adhesion to the endothelium. IL-1B-dependent expression of early inflammatory genes, such as MCP-1 or ICAM-1, involves the activation of the transcription factor nuclear factor kappa B (NF- κ B). Transient adhesion of platelets to the endothelium initiates degradation of IkB and supports activation of NF-kB in endothelial cells, thereby inducing NF-κB-dependent chemokine gene transcription [11]. Parallel to this finding, transfection of "decoy" kB oligonucleotides or a dominant negative IKK mutant attenuates platelet-induced nuclear translocation of NF-KB and MCP-1 secretion in endothelial cells [10]. Likewise, platelet-induced NF- κ B-activation was largely reduced by IL-1 β antagonists, supporting the notion that platelet IL-1 β is the molecular determinant of platelet-dependent activation of the transcription factor. Taken together, platelet-derived IL-1 β initiates NF-kB-dependent expression of chemotactic and adhesive proteins in endothelial cells. In this manner, platelets promote

the recruitment of both neutrophils and monocytes to the endothelial cell surface, thus inducing inflammation.

Another platelet-derived chemokine is RANTES, which has been identified to trigger monocyte arrest on inflamed and atherosclerotic endothelium [13]. Deposition of platelet RANTES induces monocyte recruitment mediated by P-selectin. Furthermore, release of platelet-derived CD40 ligand induces inflammatory responses in endothelium. CD154 (CD40L), a 30-33 kDa protein, belongs to the TNF family of cytokines, which includes TNF- α and Fas ligand. CD40L was originally thought to be restricted to CD4+ T-lymphocytes, mast cells, and basophils. Henn et al. [14] showed that platelets store CD40L in high amounts and release CD40L within seconds following activation in vitro and in vivo. Ligation of CD40 on endothelial cells by CD40L expressed on the surface of activated platelets increased the release of IL-8 and MCP-1, the principal chemoattractants for neutrophils and monocytes. In addition, platelet CD40L enhanced the expression of endothelial adhesion receptors including E-selectin, VCAM-1, and ICAM-1, all molecules that mediate the attachment of neutrophils, monocytes, and lymphocytes to the inflamed vessel wall (Fig. 35.2). Hence, like IL-1β, CD40L expressed on platelets induces endothelial cells to release chemokines and to express adhesion molecules, thereby generating signals for the recruitment of leukocytes

in the process of inflammation. CD40 ligation on endothelial cells, smooth muscle cells, and macrophages initiates the expression and release of matrix degrading enzymes, the matrix metalloproteinases (MMPs). These enzymes, which degrade extracellular matrix proteins, significantly contribute to destruction and remodeling of inflamed tissue. Adhesion of activated platelets to endothelial cells results in generation and secretion of MMP-9 and of the protease receptor uPAR on cultured endothelium [15]. The endothelial release of MMP-9 was dependent on both the fibrinogen receptor GPIIb-IIIa and CD40L because inhibition of either mechanism resulted in reduction of platelet-induced matrix degradation activity of endothelial cells. Moreover, GPIIb-IIIa ligation resulted in substantial release of CD40L in the absence of any further platelet agonist. These results propose that the release of platelet-derived proinflammatory mediators such as CD40L is dependent on GPIIb-IIIa-mediated adhesion. This mechanism may be pathophysiologically important to localize platelet-induced inflammation of the endothelium at a site of platelet-endothelium adhesion.

Platelets Interact with Leukocytes

Platelet adhesion to the endothelium or the subendothelial matrix induces platelet activation and the release of substances that are able to cause chemotaxis and migration of circulating leukocytes toward the site of platelet accumulation. Similar to platelet adhesion to the vessel wall, leukocyte recruitment to vascular endothelium requires multistep adhesive and signaling events, including selectin-mediated rolling, leukocyte activation, and integrin-mediated firm adhesion and diapedesis [5]. On leukocytes, members of the β_{2} -integrin family, LFA-1, MAC-1, and p150.95, as well as β ,-integrins interact with endothelial counterligands such as ICAM-1, surface-associated fibrinogen [16], or vascular cell adhesion molecule-1 (VCAM-1) to mediate the described heterotypic cell interaction. At sites of platelet adhesion to the endothelium or subendothelium, leukocyte infiltration can occur through interactions with platelets and fibrin [5]. Similar to the leukocyte-endothelium adhesion, a sequential adhesion process of leukocytes to adherent platelets has been proposed. Leukocyte adhesion to platelets involves surface expression of P-selectin on activated platelets and binding to PSGL-1, the counterreceptor present on neutrophils and monocytes [17]. Diacovo et al. [18, 19] have previously demonstrated that leukocytes tether, roll, and subsequently rest on activated platelet monolayers via sequential action of platelet P-selectin and ICAM-2 binding to their leukocyte counterreceptors PSGL-1 and CD11b/CD18, respectively.

This suggests that platelets attached to the vessel wall may recruit leukocytes. In addition, P-selectin/PSGL-1-dependent platelet-leukocyte interaction brings platelets into close vicinity with neutrophils and may facilitate leukocyte activation by platelet proinflammatory mediators. Earlier studies indicated that GPIba and JAM-3 on platelets are potential counterreceptors for MAC-1 [20] and that they mediate mechanism of platelet-leukocyte adhesion. Furthermore, ICAM-2 and $\alpha_{IIB}\beta_3$ -associated fibrinogen have also been proposed to mediate MAC-1-dependent platelet-leukocyte adhesion. Thus, platelets either immobilized on a surface or activated in suspension express a complete machinery to recruit leukocytes: (1) platelet P-selectin is a mediator of the first contact (tethering), (2) interaction of platelet P-selectin with its counterreceptor PSGL-1 on leukocytes induces signaling events relevant for MAC-1 activation, and (3) the activated β_{a} -integrin (MAC-1) on leukocytes allows and reinforces firm platelet-leukocyte adhesion through binding to counterreceptors (ICAM-2, fibrinogen bound to GPIIb-IIIa, GPIbα, JAM-3) present on the platelet surface.

Platelets Interact with Circulating Progenitor Cells

There is increasing evidence showing that circulating progenitor cells contribute to vascular repair mechanisms and limit atheroprogression. Impairment of progenitor-dependent vascular repair due to either low numbers of circulating progenitor cells or dysfunctional progenitor cells leads to inadequate vascular healing and atheroprogression. Recently, the role of platelets for recruitment and subsequent differentiation of progenitor cells has been recognized [21-23]. Adherent platelets recruit circulating progenitor cells and induce differentiation of the latter into endothelial cells or macrophages and foam cells [21-23]. Further, the combination of platelets and fibrin promoted CD34⁺ cell migration even to a greater extent than vascular endothelial growth factor in vitro [24]. Moreover, the chemokine stromal cell-derived factor-1 (SDF-1) was found to be secreted by activated platelets, which supports chemotaxis and primary recruitment of progenitor cells on the surface of arterial thrombi in vivo [22, 23]. Moreover, cytokine-mediated deployment of SDF-1 coming from activated platelets induces revascularization through mobilization of CXCR4 hemangiocytes [25].

Adhesion of human CD34⁺ cells to immobilized platelets is significantly attenuated in the presence of blocking mAbs anti-CD162 or anti-CD62P, indicating that the platelet P-selectin interacts with the endothelial progenitor cells (EPCs) through interaction with P-selectin glycoprotein ligand-1 [24, 26, 27]. Thus, platelets act as an intermediate mediator to tether progenitor cells, indicating that platelets are a prerequisite for the initial step of the homing process of CD34⁺ cells to vascular injury.

Platelets play a critical part not only in the capture, but also in the subsequent differentiation of murine EPCs, inducing the differentiation of the latter into spindle-shaped cells that are positive for vWF [21]. Furthermore, human CD34 progenitor cells can form colonies on immobilized platelets similar to immobilized fibronectin and further differentiate into mature endothelial cells [23]. Under distinct circumstances, however, in vitro co-culture experiments between platelets and human CD34⁺ cells induce distinct morphological changes of the latter and differentiation into macrophages at an early phase and later on into foam cells [26].

Inhibition of Platelet Adhesion Attenuates Atheroprogression

Enhanced chronic interaction of platelets with the arterial wall results in endothelial inflammation and atheroprogression [3]. The platelet von Willebrand receptor GPIb α and the collagen receptor GPVI have been demonstrated to largely contribute to endothelial platelet adhesion in vivo in *ApoE^{-/-}* mice [28], making them good candidates for inhibition. Inhibition of GP1b prevented adhesion of circulating platelets to endothelial cells at the carotid artery of *ApoE^{-/-}* mice.

Inhibition of platelet adhesion by blocking monoclonal anti-GPIb antibodies substantially attenuated atheroprogression in $ApoE^{-/-}$ mice [28]. Next, we studied the role of GPVImediated platelet adhesion for atheroprogression. Prolonged administration of the soluble form of GPVI [28] substantially reduced atheroprogression in ApoE-deficient mice. Further, gene transfer of GPVI-Fc to the carotid vascular wall significantly attenuated atheroprogression and endothelial dysfunction in atherosclerotic rabbits in vivo [29]. In addition, administration of soluble GPVI-Fc preferentially bound to sites of vascular injury and was able to inhibit neointimal formation after wire-induced vascular injury in $ApoE^{-/-}$ mice [30] (Fig. 35.3). Thus, inhibition of platelet adhesion via GPIb- or GPVI-blockers might be a promising strategy to attenuate lesion progression [31].

Conclusion

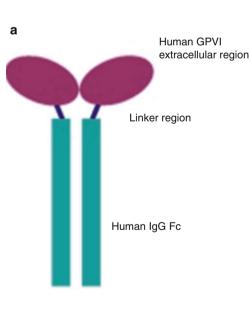
Platelets are central players in thrombosis and inflammation and contribute significantly to development of coronary artery disease. Understanding the molecular mechanisms of platelet-mediated inflammatory reactions within the vessel wall discloses important aspects of the pathophysiology of coronary artery disease. Platelet-mediated vascular inflammation is an attractive therapeutic strategy to limit atheroprogression.

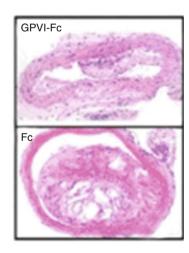
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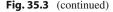
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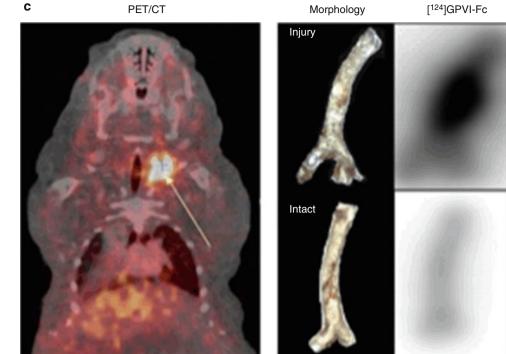
Fig. 35.3 Glycoprotein VI binds to vascular lesions and reduces atherosclerotic lesion formation. Glycoprotein VI binds to vascular lesions and reduces atherosclerotic lesion formation. (**a**) Schematic of the recombinant GPVI-Fc. (**b**) Reduced neointimal formation in GPVI-Fc-treated *ApoE^{-/-}* mice after wire-induced injury of the carotid artery (H&E staining, *bar* 100 μm) [30]. (**c**) Binding

of radiolabeled GPVI-Fc to injured carotid artery shown by PET/CT (*left*) and autoradiography (*right*) [30]









References

- 1. Davi G, Patrono C. Platelet activation and atherothrombosis. N Engl J Med. 2007;357:2482-94.
- 2. Ruggeri ZM. Platelets in atherothrombosis. Nat Med. 2002;8: 1227 - 34
- 3. Gawaz M, Langer H, May AE. Platelets in inflammation and atherogenesis. J Clin Invest. 2005;115:3378-84.
- 4. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med. 2005;352:1685-95.
- 5. May AE, Seizer P, Gawaz M. Platelets: inflammatory firebugs of vascular walls. Arterioscler Thromb Vasc Biol. 2008;28:S5-10.
- 6. Bombeli T, Schwartz BR, Harlan JM. Adhesion of activated platelets to endothelial cells: evidence for a GPIIbIIIa-dependent bridging mechanism and novel roles for endothelial intercellular adhesion molecule 1 (ICAM-1), alphavbeta3 integrin, and GPIbalpha. J Exp Med. 1998;187:329-39.
- 7. Gawaz M, Neumann FJ, Dickfeld T, Reininger A, Adelsberger H, Gebhardt A, et al. Vitronectin receptor (alpha(v)beta3) mediates platelet adhesion to the luminal aspect of endothelial cells: implications for reperfusion in acute myocardial infarction. Circulation. 1997;96:1809-18.
- 8. Gawaz M. Role of platelets in coronary thrombosis and reperfusion of ischemic myocardium. Cardiovasc Res. 2004;61:498-511.
- 9. Hawrylowicz CM, Howells GL, Feldmann M. Platelet-derived interleukin 1 induces human endothelial adhesion molecule expression and cytokine production. J Exp Med. 1991;174:785-90.
- 10. Gawaz M, Neumann FJ, Dickfeld T, Koch W, Laugwitz KL, Adelsberger H, et al. Activated platelets induce monocyte chemotactic protein-1 secretion and surface expression of intercellular adhesion molecule-1 on endothelial cells. Circulation. 1998;98:1164-71.
- 11. Gawaz M, Page S, Massberg S, Nothdurfter C, Weber M, Fisher C, et al. Transient platelet interaction induces MCP-1 production by endothelial cells via I kappa B kinase complex activation. Thromb Haemost. 2002;88:307-14.

- 12. Gawaz M, Brand K, Dickfeld T, Pogatsa-Murray G, Page S, Bogner C, et al. Platelets induce alterations of chemotactic and adhesive properties of endothelial cells mediated through an interleukin-1-dependent mechanism. Implications for atherogenesis. Atherosclerosis. 2000;148:75-85.
- 13. von Hundelshausen P, Weber KS, Huo Y, Proudfoot AE, Nelson PJ, Ley K, et al. RANTES deposition by platelets triggers monocyte arrest on inflamed and atherosclerotic endothelium. Circulation. 2001;103:1772-7.
- 14. Henn V, Slupsky JR, Grafe M, Anagnostopoulos I, Forster R, Muller-Berghaus G, et al. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. Nature. 1998;391: 591-4.
- 15. May AE, Kalsch T, Massberg S, Herouy Y, Schmidt R, Gawaz M. Engagement of glycoprotein IIb/IIIa (alpha(IIb)beta3) on platelets upregulates CD40L and triggers CD40L-dependent matrix degradation by endothelial cells. Circulation. 2002;106:2111-7.
- 16. Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. Cell. 1994;76: 301 - 14.
- 17. Gardiner EE, De Luca M, McNally T, Michelson AD, Andrews RK, Berndt MC. Regulation of P-selectin binding to the neutrophil P-selectin counter-receptor P-selectin glycoprotein ligand-1 by neutrophil elastase and cathepsin G. Blood. 2001;98:1440-7.
- 18. Diacovo TG, DeFougerolles AR, Bainton DF, Springer TA. A functional integrin ligand on the surface of platelets: intercellular adhesion molecule-2. J Clin Invest. 1994;94:1243-51.
- 19. Diacovo TG, Roth SJ, Buccola JM, Bainton DF, Springer TA. Neutrophil rolling, arrest, and transmigration across activated, surface-adherent platelets via sequential action of P-selectin and the beta 2-integrin CD11b/CD18. Blood. 1996;88:146-57.
- 20. Santoso S, Sachs UJ, Kroll H, Linder M, Ruf A, Preissner KT, et al. The junctional adhesion molecule 3 (JAM-3) on human platelets is a counterreceptor for the leukocyte integrin Mac-1. J Exp Med. 2002;196:679-91.

- 21. Langer H, May AE, Daub K, Heinzmann U, Lang P, Schumm M, et al. Adherent platelets recruit and induce differentiation of murine embryonic endothelial progenitor cells to mature endothelial cells in vitro. Circ Res. 2006;98:E2–10.
- 22. Massberg S, Konrad I, Schurzinger K, Lorenz M, Schneider S, Zohlnhoefer D, et al. Platelets secrete stromal cell-derived factor lalpha and recruit bone marrow-derived progenitor cells to arterial thrombi in vivo. J Exp Med. 2006;203:1221–33.
- 23. Stellos K, Langer H, Daub K, Schoenberger T, Gauss A, Geisler T, et al. Platelet-derived stromal cell-derived factor-1 regulates adhesion and promotes differentiation of human CD34+ cells to endothelial progenitor cells. Circulation. 2008;117:206–15.
- 24. de Boer HC, Verseyden C, Ulfman LH, Zwaginga JJ, Bot I, Biessen EA, et al. Fibrin and activated platelets cooperatively guide stem cells to a vascular injury and promote differentiation towards an endothelial cell phenotype. Arterioscler Thromb Vasc Biol. 2006;26:1653–9.
- Jin DK, Shido K, Kopp HG, Petit I, Shmelkov SV, Young LM, et al. Cytokine-mediated deployment of SDF-1 induces revascularization through recruitment of CXCR4(+) hemangiocytes. Nat Med. 2006;12:557–67.
- 26. Daub K, Langer H, Seizer P, Stellos K, May AE, Goyal P, et al. Platelets induce differentiation of human CD34(+) progenitor

cells into foam cells and endothelial cells. FASEB J. 2006; 20:2559.

- 27. Lev EI, Patel R, Karim A, Kleiman A, Badimon JJ, Kleiman NS. Anti-thrombotic effect of bivalirudin compared with eptifibatide and unfractionated heparin in diabetic patients: an ex vivo human study. Thromb Haemost. 2006;95:441–6.
- Massberg S, Brand K, Gruner S, Page S, Muller E, Muller I, et al. A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. J Exp Med. 2002;196:887–96.
- Bultmann A, Li ZM, Wagner S, Peluso M, Schonberger T, Weis C, et al. Impact of glycoprotein VI and platelet adhesion on atherosclerosis-A possible role of fibronectin. J Mol Cell Cardiol. 2010;49: 532–42.
- Schonberger T, Siegel-Axel D, Bussl R, Richter S, Judenhofer MS, Haubner R, et al. The immunoadhesin glycoprotein VI-Fc regulates arterial remodelling after mechanical injury in ApoE-/- mice. Cardiovasc Res. 2008;80:131–7.
- 31. Ungerer M, Rosport K, Bultmann A, Piechatzek R, Uhland K, Schlieper P, et al. Novel antiplatelet drug revacept (Dimeric Glycoprotein VI-Fc) specifically and efficiently inhibited collageninduced platelet aggregation without affecting general hemostasis in humans. Circulation. 2011;123:1891–9.