

Caroline Jadlowiec and Alan Dardik

The Molecular-Cellular Basis for Critical Limb Ischemia

Endothelial Cell Physiology

Progressive atherosclerotic stenosis of vessels commonly leads to the development of critical limb and myocardial ischemia. When possible and appropriate, surgical revascularization is attempted, and it is here that we clinically observe the pathological processes of ischemia and reperfusion and their complex effects [1]. Understanding of the role and function of the vascular endothelium has undergone significant changes over the past several decades. In the 1960s Willms-Kretschmer and colleagues [2] and Pober [3] referred to altered endothelial cells as being activated and, in doing so, implied a functional consequence to the altered cell morphology. This dynamic view of the endothelium, however, did not ensue into the following decade when, again, it was believed that endothelial cells were nothing more than a passive barrier. It would not be until the 1980s that Bevilacqua et al [4] would reexamine the scientific principle and ultimately prove that the vascular endothelium is both dynamic and integral to vascular and systemic equilibrium. The scientific process to better understand the endothelium dates back to the 1800s when von Recklinghausen recognized that vessels were not merely inert tunnels passing through tissue, but living entities lined by cells [5]. The endothelial monolayer comprises the entirety of the vascular system, and it is now recognized that the diversity of these cells is not merely limited by cell type alone, but rather is a function of anatomic hemodynamic variation. The unique interface formed

by the endothelium between blood and the surrounding vessel wall allows it to function as a primary mediator in response to shear stress alterations.

Studies have found that steady laminar shear stress is protective and, under such conditions, the endothelium is found to occupy a quiescent state where it exhibits both low proliferation and apoptosis [6]. Perhaps the most extensively investigated subject to date has been the influence of fluid shear stress applied to confluent monolayers of cultured endothelial cells [7]. Cellular morphology, under higher shear stress conditions, finds endothelial cells to have an elongated appearance and increased alignment, whereas low-flow, turbulent conditions produce rounded, nonaxially oriented endothelial cells that possess a higher cell turnover. The first in vivo documentation of flow-altered endothelial cellular morphology was conducted in the early 1970s, and the findings from this study showed that, in uniform laminar flow vessel segments, endothelial nuclei were oriented parallel to the axis of the blood vessel [8]. In contrast nonaxial, less-ordered nuclear orientation was found at vessel branch points and bifurcations where flow is recognized to be turbulent [9]. Further studies have since confirmed the morphologic observation that unidirectional laminar shear stress applied to cultured endothelial monolayers induces time- and force-dependent cytoskeletal reorganization. This restructuring produces changes in cell shape and alignment and is reversible with flow interruption [10].

Current studies identify hemodynamic shear stress as an important, if not primary, determinant of endothelial function and phenotype (Fig. 15.1). High shear stress, typically regarded as being greater than 15 dyn/cm², such as is found in arterial circulation, promotes endothelial cell quiescence and expression of atheroprotective genes. In contrast, low shear stress, defined as less than 4 dyn/cm², stimulates an activated atherogenic endothelial phenotype [11]. Non-laminar flow correlates with endothelial cell activation and the development of atherosclerosis and neointimal hyperplasia, and, in vitro, replication of disturbed flow has shown increased endothelial cell proliferation and apoptosis. With

C. Jadlowiec, MD • A. Dardik, MD, PhD (✉)
Department of Surgery and the Interdepartmental Program in Vascular Biology and Therapeutics, Yale University School of Medicine,
10 Amistad Street, Room 437, P.O. Box 208089,
New Haven, CT 06520-8089, USA

VA Connecticut Healthcare System,
West Haven, CT, USA
e-mail: alan.dardik@yale.edu

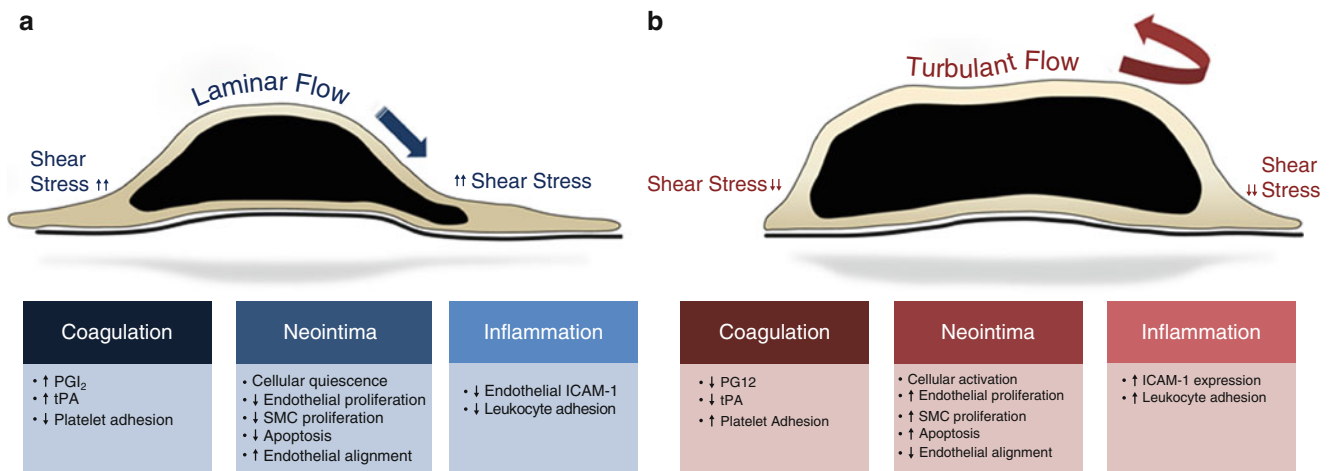


Fig. 15.1 Alterations in endothelial cell function and phenotype as determined by environmental hemodynamics. **(a)** Endothelial cell responses to high shear stress. **(b)** Endothelial cell responses to low shear stress

regard to anticoagulant properties, activated endothelium displays increased intercellular adhesion molecule expression and platelet aggregation as well as reduced production of antithrombotic substances, such as prostacyclin (PGI₂) and tissue plasminogen activator (tPA) [7]. These qualities are in stark contrast to quiescent endothelium where leukocyte adhesion is diminished, PGI₂ and tPA production is increased, and platelet aggregation is reduced [7, 12].

Normal laminar shear stress is critical in maintaining normal physiologic vascular function including thromboresistance, barrier function, and vascular tone. In contrast, low or oscillatory shear stress results in disturbed flow conditions and plays an important role in atherogenesis and bypass failure [13] (Fig. 15.1). Work done by Gimbrone and colleagues has demonstrated the relationship between decreased shear stress and atherogenesis [14]. Decreases in shear stress are accompanied by disturbances in normal endothelial cell function, and repair mechanisms within the endothelium are reduced along with the production of endothelial nitric oxide synthase (eNOS). Moreover, systemic risk factors such as hypertension and hyperlipidemia exacerbate endothelial intrinsic dysfunction. On a cellular level, this results in increased production of reactive oxygen species (ROS), altered lipoprotein permeability, leukocyte adhesion, and cellular proliferation [15].

Hypoxic Physiology

Hypoxia, depletion of the circulating oxygen content, is recognized as the driving force behind ischemic injury and, on a cellular level, endothelial cells respond to this insult by undergoing the phenotypic change now universally recognized as endothelial cell activation. This results in a series of alterations including the release of stored inflammatory mediators, changes in endothelial cell surface protein

expression, and the conversion from aerobic metabolism to anaerobic alternative pathways [16]. The vascular endothelium is responsible for regulating membrane permeability, vascular tone, coagulation, and inflammation, and it does so by dynamically responding to any systemic alterations, such as hypoxia, within the vascular environment [17]. Yet, despite its wide range of adaptability, the vascular endothelium likewise remains very sensitive to hypoxic insults, inflammatory stimuli, and physical injury from surgical manipulation or hemodynamic stress, and it is this interplay between adaptation and injury that makes cardiovascular surgery a challenging paradox [16] (Fig. 15.2).

In contrast to other cell types, the vascular endothelium is comparatively resistant to decreases in oxygen content [17]. Under ischemic conditions, aerobic cellular energy depletion leads to an atypical accumulation of cytoplasmic metabolites and failure of oxygen-dependent membrane transport systems. Of most significance is the documented intracellular increase in calcium ion concentration and the upregulation of xanthine oxidase (XO) transcription and synthesis. Although restoration of blood flow after prolonged ischemia is essential for possible physiologic salvage, reperfusion itself exacerbates endothelial cell injury. XO, an endothelial cell-associated enzyme, increases under hypoxic conditions and, with the restoration of blood flow and oxygen, leads to the intracellular buildup of reactive oxygen species (ROS) [18]. Once produced, ROS cause localized and systemic cellular damage through both direct injury to membranes and proteins and indirectly through activation of proapoptotic pathways [19]. This reperfusion damage is further augmented as damaged cellular membranes are exposed to a replenished intravascular supply of calcium. Acting as a second messenger, calcium triggers activation of various enzymes crucial to the production of proinflammatory mediators [20].

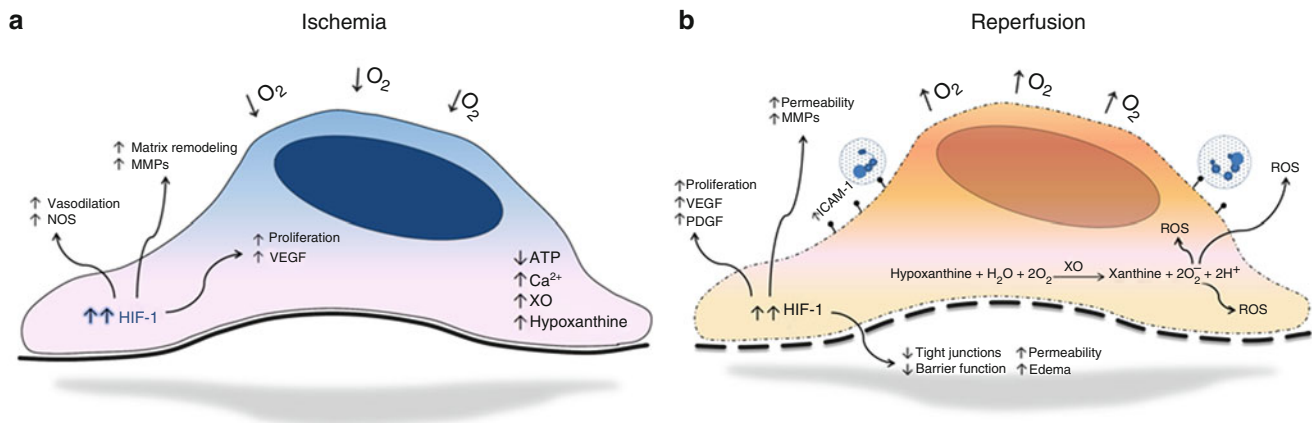


Fig. 15.2 Endothelial cell responses to ischemia. (a) Endothelial cell hypoxic physiology. (b) Endothelial cell responses to reperfusion

Under normal conditions endothelial cells form an overlapping monolayer that permits only controlled passage of molecules. This monolayer is further enhanced through the presence of tight junctions that reinforce endothelial cell barrier function. Hypoxic conditions lead to alterations in endothelial cell barrier function and vessel permeability. In response to decreased oxygen content, there is loss of tight junctions between adjacent endothelial cells leading to gap formation and increased permeability between cells. These changes are likewise accompanied by decreases in intracellular cyclic AMP, which is essential for maintenance of the actin-based cytoskeletal architecture and normal barrier function [4, 21]. This increased permeability results in what is clinically observed as ischemia reperfusion edema.

Hypoxic adaptation in endothelium leads to the transcriptional induction of a series of genes that participate in angiogenesis, metabolism, and cell proliferation, and the primary mediator of this response is hypoxia-inducible factor-1 (HIF-1), an oxygen-sensitive transcriptional activator [22]. The HIF-1 heterodimeric protein consists of two subunits, a beta subunit that is constitutively expressed (HIF-1 β) and an oxygen-regulated alpha unit (HIF-1 α). The stability and activity of the alpha subunit are regulated by its post-translational modifications, and, under normoxic conditions, this subunit is degraded. Conversely, in hypoxia, the beta subunit becomes stable resulting in the regulation of target gene expression [22]. Previous studies have demonstrated that HIF-1 plays a critical role in endothelial angiogenesis through both paracrine and autocrine mechanisms [23, 24]. Recent discoveries have shown that hypoxia-activated HIFs induce endothelial expression of several critical angiogenic factors, including vascular endothelial growth factor (VEGF), nitric oxide synthase (NOS), platelet-derived growth factor (PDGF), Ang2, and others [25]. HIFs have also been found to be mediators of endothelial survival pathways where hypoxic upregulation results in cellular proliferation. Clinically, this can result in a wide spectrum of remodeling, including angiogenesis and neointimal hyperplasia (Fig. 15.2).

Shear Stress and Arteriogenesis

Study of arteriogenesis, the enlargement of preexisting arterial connections into true collateral arteries, began in the 1700s with the work of Albrecht von Haller, a Swiss anatomist, who dissected human hearts and found that coronary arteries provide a system of interarterially connected vessels on the side of high arterial pressure. His findings suggested that these conduits were functional arteries, larger in size than capillaries [26]. Today, this process is recognized as arteriogenesis, and, unlike angiogenesis, which is primarily mediated by hypoxia, initiation of collateral artery growth and remodeling is dependent on alterations in hemodynamic forces. Circumferential wall stress and fluid shear stress are recognized and the two primary forces responsible for this complex process; however, uncertainties continue with regard to the specific contributions and overall importance of either toward arteriogenesis. Some uncertainty comes from the observation that fluid shear stress, with a typical range of 20–30 dyn/cm², is a weak force when compared with circumferential wall stress, which is 106 times greater [27].

In the early 1920s, Murray proposed that the vascular system's branching configuration exists so as to minimize the amount of mechanical and metabolic work to provide adequate blood flow. Using this model, it can be surmised that fluid shear stress remains constant throughout the vasculature and that blood flow is proportional to the cube of each individual vessel's diameter. Additionally, shear stress remains proportional to blood flow and inversely related to the cube of the radius [27, 28]. Accordingly, it is hypothesized that shear stress regulates the acute early phase of arteriogenesis where small increases in collateral artery diameter result in significant decreases in shear stress [29]. The formation of collateral circulation after an arterial occlusion correlates well with the observed increase in shear stress where increased collateral flow is caused by the pressure redistribution from pre-existent, now occluded, vessels [30]. It is also recognized that increases in collateral diameter end once shear stress normalizes.

Interestingly, despite early and abrupt normalization, cellular remodeling of collaterals persists beyond this acute period, and a shift from quiescence toward proliferation is observed in both endothelial and vascular smooth muscle cells [31]. As a result, it has been proposed that circumferential wall stress, which remains elevated, is the more dominant force in this later stage of arteriogenesis. Regardless of the dominant mediators, collateral remodeling frequently reaches a plateau, at which point hemodynamic alterations are unable to compensate for progressive atherogenesis. Indeed, when these compensatory systems fail or are overwhelmed, what follows next is the clinical entity of critical limb ischemia.

Surgical Solutions for CLI

Autologous Vein Grafts

Today, autologous vein grafting is the recognized standard for infrainguinal arterial revascularization, yet prior to the pioneering work done in the early 1900s, the predominating belief was that repair of major vasculature was beyond the capacity of surgical technique. Unconvinced that the 1894 penetrating portal venous injury of French president Carnot was not treatable, Carrel would undertake the task of refining vessel anastomosis. He would go on to establish the modern fundamentals of bypass surgery with his combined techniques of delicate vessel handling, fine suture material, and triangulation [21]. The future of bypass surgery would then be further progressed by Kunlin, who in 1948 would be the first to successfully use a reversed saphenous vein graft in the treatment of lower extremity ischemia [32]. In 1962, Sabiston would then go on to use a modification of this technique in what we recognize today as the modern coronary bypass. Interestingly, although much has changed from the pioneering days of surgical vascular research, what remains is that the conduit of choice for lower extremity bypass continues to be the greater saphenous vein.

Physiology of Vein Graft Adaptation

Venous anatomy echoes arterial, and the intima, media, and adventitia are easily identified as the major anatomic vessel wall components. It is well known that, unlike arteries, veins have a reduced medial component, which results in a thinner vessel wall and typically allows for increased compliance. From prior work, it is known that vein compliance is not a static property, but rather dependent on its environment. Although it is commonly assumed that native veins are more compliant than arteries, in actuality, this increased compliance only remains present up to pressures of 35–50 mmHg [33]. Under typical arterial pressures, vein grafts become stiff and lose compliance, while matched arteries remain moderately distensible. Accordingly, a more accurate description is that veins are more

compliant than arteries at low pressures, but less compliant than arteries when subjected to arterial pressures.

Low pressure and low flow are the dominant hemodynamic entities in the normal venous environment, and, unlike the physiology observed in arterial circulation, veins are not subjected to pulsatility, high flow, or high pressures. All vessels, venous and arterial, are subject to mechanical forces in the form of shear and circumferential wall stress. Effects of circumferential strain directly correlate with blood pressure, and in the venous system, these maximal pressures are understandably reduced. The circumferential venous forces that remain go on to be counteracted by the entirety of the vessel wall, and the cellular components in the intima, media, and adventitia are uniformly affected. Conversely, shear stress, the frictional force of blood along the intima is distinctive in that it exerts its force primarily on endothelial cells. Fluid shear stress likewise displays significant variation between venous and arterial systems, and prior studies have found 1–6 dyn/cm² to be typical of venous shear stress. This is in large contrast to arterial vasculature where anywhere from 10 to 70 dyn/cm² is the prevailing norm [11]. This abrupt hemodynamic alteration has been implicated as the mechanism responsible for initial endothelial desquamation, which is observed to occur in the first week of graft placement. In such situations, graft endothelium regeneration appears to occur quickly with complete healing observed in some veins at 48 h [34]. Nonetheless, concerns regarding the long-term effects of acute arterial disruption of venous endothelium remain with respect to graft patency.

Indeed, long-term patency for autologous bypass is largely dependent on how successfully venous conduits adapt to arterialization. As first suggested by Owens et al., the process of successful vein graft adaptation appears to involve at least two distinct temporal phases: early outward remodeling of the lumen and delayed acquisition of wall stiffness [35]. In 2006, in a prospective human study, the authors showed that 72 % of venous grafts dilate during the first post-surgical month and that no appreciable changes occur beyond this time. During the first 6 months, a nearly 40 % increase in conduit stiffness was found with the greatest relative increase occurring during the initial first 3 postoperative months. Clinically, grafts that demonstrated early positive remodeling in the form of lumen dilatation appeared to have increased primary patency, and a trend toward greater wall stiffness at 1 month was noted in grafts that failed [35]. Further work done by the same authors in 2008 underscored the critical role of systemic inflammation in vein graft remodeling. In this study, there was positive correlation noted between vein graft diameter and initial shear stress. This shear-dependent response, however, was disrupted in patients with an elevated baseline high-sensitivity C-reactive protein (hsCRP). Moreover, despite similar vein diameter and shear stress at implantation, grafts in the elevated hsCRP group demonstrated less positive remodeling within the first month and likewise had a propensity to be stiffer [36]. Accordingly,

although intricacies and specific mediators remain to be identified, early positive adaptation of vein grafts lends itself to successful long-term patency, and the importance of controlling systemic inflammation deserves emphasis.

Responses of Venous Endothelial Cells to Shear Stress

Hemodynamic shear stress, possibly the most influential component of vascular remodeling and pathology, is the frictional force created by the flow of blood in relation to the luminal vessel wall and endothelial surface. It is in this regard that the creation of a surgical bypass, using a venous conduit, becomes one of the most dynamic processes responsible for endothelial remodeling. Although the technicalities of performing a successful bypass are well established, much of the final outcome hinges on how successful the venous conduit will be in adapting to the high-pressure pulsatile flow arterial circulation where it is acutely placed [37]. Much remains to be learned about the underlying mechanisms responsible for both successful as well as pathologic endothelial adaptation. At present, the majority of our insight into endothelial response to hemodynamics comes from animal experiments in which shear stress is acutely or chronically altered [11].

Classically, increases in shear stress, both acute and chronic, are observed to result in upregulation of endothelial nitric oxide synthase (eNOS) mRNA and protein production [38]. Such increases in eNOS activity result in vessel lumen dilation. Studies investigating this role for nitric oxide (NO) have shown that through the use of L-NAME, a nitric oxide synthase inhibitor, flow-induced vessel dilation is inhibited [39]. In 1997, Clowes and colleagues demonstrated an important relationship between shear stress and neointimal hyperplasia. Using a primate model, the authors inserted PTFE grafts into aortoiliac circulation bilaterally. A neointima was allowed to develop for 2 months, and, at that time, half of the animals were killed. For the remaining group, graft flow was increased through the creation of a femoral arteriovenous fistula. These animals were then killed 2 months post-fistula creation. The findings from this study showed regression of the neointima in those grafts exposed to the additional 2 months of high shear stress. From prior work, using this same model, the authors were also able to show that high shear stress inhibits neointimal growth [39].

Work examining the effects of high and low flow on graft adaptation have shown increased smooth muscle cell proliferation in grafts exposed to low shear stress, and similar findings have also been observed in endothelial cells [40]. In a rat model, where unilateral ligation of the internal and external carotid arteries results in the creation of high- and low-flow arteries, increased endothelial cell proliferation is seen at 24, 48, and 72 h post-ligation in the reduced flow carotids. With this model, the achieved alteration in flow reflected true arterial

and venous circulation, with a comparative shear stress of 33.4 ± 1.1 dyn/cm² in the high-flow carotid versus 1.4 ± 0.2 dyn/cm² in the ligated, low-flow complement [41]. These correlations between shear stress and neointimal remodeling provide insight into the mechanisms involved in venous bypass and likewise demonstrate the paradoxical effects that high flow and shear stress can exert on endothelial homeostasis.

Graft Characteristics and Long-Term Patency

Using the best available autologous conduit is universally accepted as the fitting approach for peripheral bypass surgery, and the conduit of choice is the ipsilateral, single-segment greater saphenous vein secondary to its documented patency in lower extremity bypass [42]. Other characteristics that define the best autologous conduit however continue to be debated, and much of the data available comes from single-institution retrospective studies that attempted to define meaningful graft characteristics in regard to long-term patency. Traditionally, three surgical configurations, reversed, nonreversed, and in situ, have been accepted for use in infrainguinal vascular bypass work, and each is believed to confer different effects on endothelial cells.

Reversed Vein Grafts

Vein excision with reversal of anatomical orientation and maintenance of intact valves is the essence of a reversed vein grafting technique. Considered to be somewhat basic, usage of reversed vein grafts is typically well adapted for most surgical bypass settings. The reversed technique eliminates the need for valve lysis and thus limits intraluminal manipulation. Concerns with regard to the altered flow dynamic in reversed vein grafts have come from studies showing intraluminal turbulence secondary to retained valves. The surgical creation of a reversed vein graft introduces valves into the arterial system and, by doing so, adds a potential source of turbulence. Such deviations from laminar flow hold the potential to initiate pathological endothelial cell activation. Despite having a reversed anatomy, venous valves do not lie flush with the graft wall, and once subjected to pulsatile arterial flow, these valves are observed to close during diastole [43]. Coupled with the cardiac cycle, valves have the potential to modify flow dynamics by becoming active and preventing the natural arterial backflow, and it has been speculated that midgraft stenotic lesions occur secondary to retained valve leaflets [44]. Other surgical concerns arise from the stasis observed within the valve cusps and the potential for them to function as a thrombogenic nidus [43].

Nonreversed Vein Grafts

In nonreversed technique, the vein is excised, its valves are subjected to mechanical lysis, and it is then oriented in its original nonreversed configuration. Observed advantages

come from the preservation of a nonreversed orientation, thus improving the circumferential accord of the arterial graft anastomosis. Intraluminal valve lysis, however, is a requisite and results in endothelial cell trauma and denudation. Venous valve lysis results in significant endothelial trauma, and, from prior work, it is known that the preservation of endothelial cell integrity in venous grafts prevents subsequent morphologic changes, namely thickening of the venous intima and media in response to arterial dynamics [45]. Remarkably, despite the inherent need for more manipulation and handling with nonreversed grafts, improved hemodynamic flow is the resulting outcome following valve lysis. Studies have shown that lysis of valves increases graft flow rates anywhere from 30 to 60 % as compared to grafts with intact valves [46, 47]. Clinically, the improved bypass hemodynamics, shown through higher shear stress and flow velocity, are atheroprotective and correlate with increased long-term graft patency.

In Situ Vein Grafts

The initial concept of utilizing an autologous vein graft while minimizing surgical manipulation is attributed to the work of both Rob and Hall in the early 1960s. At that time, advanced valvulotomes had yet to be developed, and valve lysis was achieved through serial transverse venotomies and individual valve excision [48]. The modern in situ vein graft is created through limited mobilization of both the proximal and distal vein segments, thus allowing them to be used in arterial bypass but simultaneously reducing total vein manipulation and damage to the outer vessel layers, including the vasa vasorum, during harvesting. Some studies have suggested improved patency outcomes in vein bypasses harvested en-bloc with surrounding tissue where venous spasm is believed to be reduced [49]. Prior studies have also demonstrated that the disruption of the vasa vasorum results in an acute decrease in vessel distensibility as well as long-term structural changes and delayed deterioration of vessel elastic properties [50]. Based on such evidence, it has been proposed that by diminishing spasm, the need for high pressure distention, a known mediator of vein wall and endothelial damage, is reduced [49]. There is also evidence to suggest an inverse relationship between vasa vasorum preservation and neointimal hyperplasia. Findings by Gossl et al. showed that vessel areas with diminished vasa vasorum density were more susceptible to hypoxia, oxidative stress, and microinflammation, all factors known to potentiate early atherogenesis [51]. Accordingly, there is modest evidence present that the use of atraumatic techniques, which minimize damage to the adventitia and the vasa vasorum, improve bypass patency.

Although there have been many studies, mainly single center and retrospective, that have attempted to address aspects of surgical technique in relation to lower extremity bypass outcomes, the exact contribution of specific technical factors remains to be fully defined. The most comprehensive

data available come from the 2006 Project of Ex-Vivo Vein Graft Engineering via Transfection III (PREVENT III) Study, which was a randomized, double-blinded, placebo-controlled trial of edifoligide for prevention of vein graft failure in patients undergoing lower extremity revascularization for critical limb ischemia. Although the primary and secondary endpoints for PREVENT III did little to modify current medical strategies for limb revascularization, the study was able to provide perhaps the most comprehensive multicenter data available to date with regard to long-debated conduit and technique characteristics, such as vessel diameter, graft type, and conduit orientation [52, 53].

Results for early 30-day graft failure identified several significant technical predictors of early loss of primary patency. Small-conduit diameter was identified as one such technical factor, and primary patency in grafts greater than 3.5 mm was observed to be 93.2 % vs. 85.7 % in grafts with diameters less than 3.0 mm. Not surprisingly, composite grafts showed worse outcomes when compared to single-segment grafts with a primary patency of 84.1 % vs. 92.2 % in non-spliced greater saphenous vein. Bypasses originating from a more distal location showed improved short-term outcomes. Grafts that originated from the popliteal artery had primary patency rates of 93.9 % as compared to those with more proximal anastomoses, such as the common or superficial femoral arteries, where rates were 91.7 % and 87.7 %, respectively. Graft length and the site of distal anastomosis were not found to be predictors of primary patency loss. Although a slight difference was observed between the early failure rates of reversed and nonreversed grafts, 91.6 % patency as compared to 93.3 %, this finding was not statistically significant [53]. At 1 year, graft patency was again found to be detrimentally affected by small conduit diameter with primary patency observed to be 68.4 % in grafts with diameters greater than 3.5 mm vs. 42.4 % for those less than 3.0 mm. Poorer outcomes were observed in bypasses composed of composite spliced vein, those greater than 60 cm in length, and more proximal bypass origination. Moreover, at 1 year, the primary patency rates for reversed (65.0 %) and nonreversed (63.3 %) orientation grafts were equivalent and not statistically significant. From these data, significant identifiable technical predictors of early and late graft failure include use of a small conduit diameter or a composite vein; however, surgical variation with the reversed and nonreversed technique does not appear to be a primary factor in bypass success [53].

Prosthetic

While the saphenous vein continues to be the superior unrivaled arterial substitute for lower extremity bypass, a significant portion of patients with critical limb ischemia does not possess an autologous vein that is usable. It is in

such difficult situations that an alternative arterial conduit is frequently employed. The history regarding synthetic conduits, like much of surgical history, is long and much indebted to research pioneers. In 1952, Voorhees and Blakemore published a preliminary report describing the successful use of Vinyon "N" cloth prostheses in the infrarenal aorta of mongrel dogs [54]. Despite these limited successes, long-term durability would remain limited until the successful use of the modern polymers polytetrafluoroethylene (PTFE) and polyethylene terephthalate (Dacron).

Dacron

Within the same time period of the early 1950s, DeBakey went on to create the first Dacron tube graft for aortic reconstruction [55]. Unlike the preceding experiments using nylon, Dacron, a synthetic multifilament yarn, proved to be both a durable and suitable material for vascular reconstruction. Traditionally, Dacron grafts have either been woven or knitted, and modern grafts of both varieties continue to be manufactured and used. Woven grafts do not rely on the looping of yarn around a needle, and because of this the resulting material has a decreased porosity [56]. The more compact nature of woven grafts results in stronger fabric, but comes at the cost of decreased compliance. The stiffness of the woven graft is believed to make it more difficult to handle and suture, and the fiber orientation enhances the potential of fraying when cut in the operating room [56]. Conversely, knitted Dacron results in a softer, more compliant material that likewise is more porous. The increased porosity of these grafts makes them more susceptible to bleeding, and traditionally knitted tube grafts were subjected to preclotting where the patient's blood was passed through the graft prior to surgical interposition so as to coat the pores with fibrin and minimize operative bleeding [57]. Modern knitted grafts are now manufactured with precoated materials, such as gelatin or collagen.

PTFE

Polytetrafluoroethylene (PTFE) first came into production in the 1930s, and by the early 1970s, an expanded form, ePTFE, began to be used in animal models as an alternative to Dacron in arterial reconstruction [58]. The first successful reported clinical use followed soon after in 1976 when Campbell et al. published their experience in using ePTFE in infrainguinal bypass [59]. Today, PTFE is predominantly processed into expanded PTFE (ePTFE), and the extrusion process results in a porous morphology that microscopically is characterized by interconnected nodes and fibrils. Unlike the woven or knitted characteristics of Dacron, ePTFE is a single seamless structure. Despite being microporous, ePTFE has a low friction coefficient making its surface smooth and hydrophobic. Although ePTFE's hydrophobic property prevents hematogenous graft permeation, plasma and platelet surface

adherence is not inhibited, and the overall host response has been found to be similar to that of Dacron [60]. More recently, heparin-bonded ePTFE grafts have become available for clinical use. Early data have suggested that these grafts decrease platelet adherence and acute thrombus formation surface [61]. Animal models have also shown a reduction in anastomotic intimal hyperplasia [62]. In 2009, The Propaten European Product Evaluation (PEPE II) study showed that heparin-bonded ePTFE grafts yield patency rates that are comparable to those obtained with other graft material in infrainguinal bypass surgery [63]. At present, however, improvements to graft patency remain to be demonstrated, and further clinical data will be required to adequately determine this. Despite such fundamental differences in material composition, prospective randomized trials have found PTFE and Dacron to be equally suitable for infrainguinal bypass. Results from a trial published in 2001 showed no differences between Dacron and PTFE with regard to primary and secondary patency or limb salvage [64]. Consequently, the surgeon's choice of PTFE versus Dacron for infrainguinal bypass in the operating room is truly based on preference and not on clinical differences in graft performance and patency.

Endothelial Cell Seeding

Early work with Dacron from the 1960s raised excitement over the finding of a complete endothelial lining present in prosthetic grafts implanted into various animal species [65]. In stark contrast to this mature endothelium found within the arterial prostheses of baboons and pigs, however, was the realization that humans grafts fail to achieve this re-endothelialization [66]. Knowledge that endothelial cells possess anti-thrombotic properties and function to limit intimal hyperplasia, both primary obstacles to graft patency, led to research investigating the potential role for endothelial cell transplantation onto synthetic vascular grafts. In 1978, Herring et al. published their work using a canine model on arterial prostheses seeded with autogenous vascular endothelium. The authors achieved this result by harvesting endothelial cells from saphenous vein using steel wool pledgets and using the admixture to preclot Dacron grafts [67]. These results were promising, and over the ensuing decade much effort would be placed into endothelial cell harvest optimization and transplant, a process that came to be referred to as cell seeding.

Further enthusiasm toward cell seeding would come from later animal studies demonstrating that ex-vivo endothelial cell incorporation into synthetic grafts enhanced patency [68]. These results only further reinforced the belief that development of mature endothelial cell surface was the missing link to maintaining long-term patency in synthetic arterial

substitutes [68]. Concerns that remained to be reconciled, however, included whether any of the benefits ascribed to cell seeding in animal models actually conferred analogous human benefit [69]. In 1985, Rosenman et al. published important work relating to the kinetics of endothelial cell adhesion to synthetic graft materials. Using carotid interposition ePTFE seeded grafts with 111 indium oxide-labeled endothelial cells, the authors were able to examine cell retention following arterial implantation. Immediately following implantation, only 19.8 % of the originally seeded cells were found to be present, and, at 30 min, 70.2 % of these remaining cells desquamated. Cell loss continued over the next 24 h, at which time only 4.4 % of originally seeded cells remained present [70]. These results were obviously unfavorable, and for much of the next decade, experimental work would focus on methods to improve endothelial cell adhesion.

The findings of poor endothelial cell retention coupled with, at best, modest improvements in graft patency greatly diminished the surgical community's enthusiasm for cell seeding. As noted by Zilla, those who continued work with endothelial cell transplant were prepared to accept both a long and arduous route with the knowledge that their contribution would be a quiet one [71]. Not surprisingly, research using collagen or fibronectin pretreatment to synthetic grafts found that endothelial cells adhered poorly to and did not grow on untreated Dacron and PTFE, while protein-treated materials facilitated cellular retention [72]. Differences were also noted between pre-treated Dacron and PTFE. Cells seeded onto protein-treated PTFE were noted to form a confluent monolayer within 9 days, while the more irregular surface of Dacron showed inferior results [73]. The following decade would bring with it the next generation of advancement in the quest to create a biologically functional vascular prosthesis. It was observed that chronic in vitro culture of aortic endothelial cells with flow was associated with cytoskeletal reorganization and increased endocytic activity when compared to conventional static culture conditions. These data implied that, through the control of flow, differentiation of endothelial cells could be regulated [74]. It was therefore hypothesized that endothelial cell adhesion could be stimulated and enhanced by shear stress. The results of this hypothesis became apparent in 1999, when Dardik and colleagues published their work investigating the role of shear stress in endothelial cell retention. PTFE endothelial cell seeded grafts were treated with 0, 1, or 25 dyn/cm² shear stress in vitro and then implanted. The authors found that pretreatment with 25 dyn/cm² resulted in fully confluent endothelial cell retention at 24 h post in vivo implantation. Furthermore, this confluency and retention continued to be observed up to a 3-month time point. In comparison, this same observation did not hold true for low flow conditions [75].

Despite great progress in understanding the biomechanics of endothelial cell adaptation, clinical studies to date have

been less successful in providing similar insight and progress. The first report of human cell seeding came from Herring et al. in 1984. Using seeded and unseeded ePTFE, the authors documented a 100 % patency for seeded femoral-popliteal bypasses at 18 months in comparison to 60 % in unseeded grafts [76]. Overall, however, results from further human clinical trials have been mixed, with many investigators finding no benefit [77]. The most promising long-term clinical cell seeding data come from work done by Zilla and colleagues. In a study published in 1999, the authors showed a notable primary 9-year patency rate of 65 % in endothelialized PTFE grafts versus 16 % in control non-seeded PTFE grafts [78]. More recently in 2009, the same authors published their long-term results using autologous in vitro endothelialization of infrainguinal ePTFE grafts. Over a span of 15 years, and inclusive of 341 infrainguinal endothelialized ePTFE grafts, the authors showed an overall femoropopliteal primary patency rate of 69 % at 5 years and 61 % at 10 years [79]. Unexpectedly, examination of retrieved ePTFE samples from this cohort revealed the presence of an endothelium on all samples after 38.9 ± 17.8 months, thus challenging prior cell seeding outcomes where long-term cell retention appeared difficult, if not impossible, to achieve [79]. The results of this group go on to suggest that in vitro long-term endothelialization of synthetic grafts is not only clinically feasible, but that it also provides a patency benefit in challenging patients without suitable autogenous vein.

References

1. Verrier ED, Boyle Jr EM. Endothelial cell injury in cardiovascular surgery. *Ann Thorac Surg.* 1996;62(3):915–22.
2. Willms-Kretchmer K, Flax MH, Cotran RS. The fine structure of the vascular response in haptan-specific delayed hypersensitivity and contact dermatitis. *Lab Invest.* 1967;17(3):334–49.
3. Pober JS, Warner-Lambert/Parke-Davis award lecture. Cytokine-mediated activation of vascular endothelium. *Physiology and pathology.* *Am J Pathol.* 1988;133(3):426–33.
4. Bevilacqua MP, Pober JS, Wheeler ME. Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes, and related leukocyte cell lines. *J Clin Invest.* 1985;76:2003–11.
5. Cines DB, Pollak ES, Buck CA, et al. Endothelial cells in physiology and in the pathophysiology of vascular disorders. *Blood.* 1998;91(10):3527–61.
6. Paszkowiak JJ, Dardik A. Arterial wall shear stress: observations from the bench to the bedside. *Vasc Endovascular Surg.* 2003;37(1):47–57.
7. Dardik A, Chen L, Frattini J, et al. Differential effects of orbital and laminar shear stress on endothelial cells. *J Vasc Surg.* 2005;41(5):869–80.
8. Davies PF. Flow-mediated endothelial mechanotransduction. *Physiol Rev.* 1995;75(3):519–60.
9. Flaherty JT, Pierce JE, Ferrans VJ, et al. Endothelial nuclear patterns in the canine arterial tree with particular reference to hemodynamic events. *Circ Res.* 1972;30(1):23–33.

10. Resnick N, Gimbrone MA. Hemodynamic forces are complex regulators of endothelial gene expression. *FASEB J*. 1995;9(10):874–82.
11. Malek AM, Alper AL, Izumo S. Hemodynamic shear stress and its role in atherosclerosis. *JAMA*. 1999;282(21):2035–42.
12. Caplan BA, Schwartz CJ. Increased endothelial cell turnover in areas of in vivo Evans blue uptake in the pig aorta. *Atherosclerosis*. 1973;17:401–17.
13. Cunningham KS, Gotlieb AI. The role of shear stress in the pathogenesis of atherosclerosis. *Lab Invest*. 2005;85:9–23.
14. Gimbrone Jr MA, Nagel T, Topper JN. Biomechanical activation: an emerging paradigm in endothelial adhesion biology. *J Clin Invest*. 1997;99(8):1809–13.
15. Gimbrone MA, Topper JN, Nagel T, et al. Endothelial dysfunction, hemodynamic forces, and atherogenesis. *Atherosclerosis V: the Fifth Saratoga Conference*. *Ann N Y Acad Sci*. 2000;902:230–40.
16. Verrier ED, Morgan EN. Endothelial response to cardiopulmonary bypass surgery. *Ann Thorac Surg*. 1998;66(S1):S17–9.
17. Ten VS, Pinsky DJ. Endothelial response to hypoxia: physiologic adaptation and pathologic dysfunction. *Curr Opin Crit Care*. 2002;8(3):242–50.
18. Ratych RE, Chuknyiska RS, Bulkley GB. The primary localization of free radical generation after anoxia/reoxygenation in isolated endothelial cells. *Surgery*. 1987;102(2):122–31.
19. Braunersreuther V, Jaquet V. Reactive oxygen species in myocardial reperfusion injury: from physiopathology to therapeutic approaches. *Curr Pharm Biotechnol*. 2012;13(1):97–114.
20. Kerrigan CL, Stotland MA. Ischemia reperfusion injury: a review. *Microsurgery*. 1993;14(3):165–75.
21. Cooley BC. History of vein grafting. *Microsurgery*. 1998;18(4):234–6.
22. Ke Q, Costa M. Hypoxia-inducible factor-1 (HIF-1). *Mol Pharmacol*. 2006;70(5):1469–80.
23. Pugh CW, Ratcliffe PJ. Regulation of angiogenesis by hypoxia: role of the HIF system. *Nat Med*. 2003;9:677–84.
24. Manalo JD, Rowan A, Lavoie T, et al. Transcriptional regulation of vascular endothelial cell responses to hypoxia by HIF-1. *Blood*. 2005;105(2):659–69.
25. Carmeliet P, Jain RK. Angiogenesis in cancer and other disease. *Nature*. 2000;407:249–57.
26. Buschmann I, Schaper W. The pathophysiology of the collateral circulation (arteriogenesis). *J Pathol*. 2000;190(3):338–42.
27. Schaper W, Scholz D. Factors regulation arteriogenesis. *Arterioscler Thromb Vasc Biol*. 2003;23(7):1143–51.
28. Murray CD. The physiological principle of minimum work applied to the angle of branching of arteries. *J Gen Physiol*. 1926;9(6):835–41.
29. Grundmann S, Piek JJ, Pasterkamp G, et al. Arteriogenesis: basic mechanisms and therapeutic stimulation. *Eur J Clin Invest*. 2007;37(10):755–66.
30. Pipp F, Boehm S, Cai WJ, et al. Elevated fluid shear stress enhances postocclusive collateral artery growth and gene expression in the pig hind limb. *Arterioscler Thromb Vasc Biol*. 2004;24(9):1664–8.
31. Heil M, Eitenmüller I, Schmitz-Rixen T. Arteriogenesis versus angiogenesis: similarities and differences. *J Cell Mol Med*. 2006;10(1):45–55.
32. Kunlin J. Le traitement de l'arterite oblitérante par la greffe veineuse. *Arch Mal Coeur*. 1949;42:371–2.
33. Dobrin PB, Littooy FN, Golan J, et al. Mechanical and histologic changes in canine vein grafts. *J Surg Res*. 1988;44(3):259–65.
34. Krupski W, Thal ER, Gewertz BL, et al. Endothelial response to venous injury. *Am Med Assoc*. 1979;114(11):1240–8.
35. Owens CD, Wake N, Jacot JG, et al. Early biomechanical changes in lower extremity vein grafts—distinct temporal phases of remodeling and wall stiffness. *J Vasc Surg*. 2006;44(4):740–6.
36. Owens CD, Rybicki FJ, Wake N. Early remodeling of lower extremity vein grafts: inflammation influences biomechanical adaptation. *J Vasc Surg*. 2008;47(6):1235–42.
37. Golledge J, Turner RJ, Harley SL, et al. Circumferential deformation and shear stress induce differential responses in saphenous vein endothelium exposed to arterial flow. *J Clin Invest*. 1997;99(11):2719–26.
38. Nadaud S, Philippe M, Arnal JF, et al. Sustained increase in aortic endothelial nitric oxide synthase expression in vivo in a model of chronic high blood flow. *Circ Res*. 1996;79:857–63.
39. Guzman RJ, Abe K, Zarins CK. Flow-induced arterial enlargement is inhibited by suppression of nitric oxide synthase activity in vivo. *Surgery*. 1997;122:273–80.
40. Kraiss LW, Geary RL, Mattsson EJ, et al. Acute reductions in blood flow and shear stress induce platelet-derived growth factor-A expression in baboon prosthetic grafts. *Circ Res*. 1996;79:45–53.
41. Mondy JS, Lindner V, Miyashiro JK, et al. Platelet-derived growth factor ligand and receptor expression in response to altered blood flow in vivo. *Circ Res*. 1997;81:320–7.
42. Masser PA, Taylor LM, Moneta GL, et al. Technique of reversed vein bypass for lower extremity ischemia. *Ann Vasc Surg*. 1996;10(2):190–200.
43. McCaughan Jr JJ, Walsh DB, Edgcomb LP, et al. In vitro observations of greater saphenous vein valves during pulsatile and nonpulsatile flow and following lysis. *J Vasc Surg*. 1984;1(2):356–61.
44. Vesti BR, Primožich J, Bergelin RO, et al. Follow-up of valves in saphenous vein bypass grafts with duplex ultrasonography. *J Vasc Surg*. 2001;33(2):369–74.
45. Quist WC, LoGerfo FW. Prevention of smooth muscle cell phenotypic modulation in vein grafts: a histomorphometric study. *J Vasc Surg*. 1992;16(2):225–31.
46. Ku DN, Klafta JM, Gewertz BL, et al. The contribution of valves to saphenous vein graft resistance. *J Vasc Surg*. 1987;6:274–9.
47. Chin AK, Mayer DN, Goldman RK, et al. The effect of valvulotomy on flow rate through the saphenous vein graft: clinical implications. *J Vasc Surg*. 1988;8(3):316–20.
48. May AG, DeWeese JA, Rob CG. Arterialization in situ saphenous vein. *Arch Surg*. 1965;91(5):743–50.
49. Souza SRD, Bomfim V, Skoglund H, et al. Early patency of saphenous vein graft for coronary artery bypass harvested with surrounding tissue. *Ann Thorac Surg*. 2001;71:797–800.
50. Stefanadis C, Vlachopoulos C, Karayannacos P, et al. Effect of vasa vasorum flow on structure and function of the aorta in experimental animals. *Circulation*. 1995;91:2669–78.
51. Gossli M, Versari D, Lerman LO, et al. Low vasa vasorum densities correlate with inflammation and subintimal thickening: potential role in location—determination of atherogenesis. *Atherosclerosis*. 2009;206:362–8.
52. Conte MS, Bandyk DK, Clowed AW, et al. Results of PREVENT III: a multicenter, randomized trial of edifoligide for the prevention of vein graft failure in lower extremity bypass surgery. *J Vasc Surg*. 2006;43(4):742–51.
53. Schanzer A, Hevelone N, Owens CD, et al. Technical factors affecting autogenous vein graft failure: observations from a large multicenter trial. *J Vasc Surg*. 2007;46(6):1180–90.
54. Voorhees AB, Jaretzki A, Blakemore AH. The use of tubes constructed from vinyon “N” cloth in bridging arterial defects: a preliminary. *Ann Surg*. 1952;135(3):332–3.
55. Kapadia MR, Popowich DA, Kibbe MR. Modified prosthetic vascular conduits. *Circulation*. 2008;117(14):1873–82.
56. Quarumby JW, Burnand KG, Lockhart SJM, et al. Prospective randomized trial of woven versus collagen-impregnated knitted prosthetic Dacron grafts in aortoiliac surgery. *Br J Surg*. 1998;85(6):775–7.

57. Guidoin R, Gosselin C, Domurado D. Dacron as arterial prosthetic material: nature, properties, brands, fate and perspectives. *Biomater Med Devices Artif Organs*. 1977;5(2):177–203.
58. Soyer T, Lempinen M, Cooper P, et al. A new venous prosthesis. *Surgery*. 1972;72(6):864–72.
59. Campbell CD, Brooks DH, Webster MW. The use of expanded microporous polytetrafluoroethylene for limb salvage: a preliminary report. *Surgery*. 1976;79(5):485–91.
60. Blumenberg RM, Anderson JM, Gelfand ML, et al. Histologic evaluation of Dacron® and PTFE graft material explanted from humans after 4 to 20 years in vivo. *Vasc Endovascular Surg*. 2000;34(6):505–12.
61. Heyligers JMM, Lisman T, Weeterings C, et al. Heparin immobilization reduces thrombogenicity on small-caliber ePTFE grafts. *J Vasc Surg*. 2006;43(3):587–91.
62. Lin PH, Chen C, Bush RL, et al. Small-caliber heparincoated ePTFE grafts reduce platelet deposition and neointimal hyperplasia in a baboon model. *J Vasc Surg*. 2004;39(6):1322–8.
63. Hugl B, Nevelsteen A, Daenens K, et al. PEPE II-a multicenter study with an end-point heparin-bonded expanded polytetrafluoroethylene vascular graft for above and below knee bypass surgery: determinants of patency. *J Cardiovasc Surg*. 2009;50(2):195–203.
64. Post S, Kraus T, Muller-Reinartz U, et al. Dacron vs polytetrafluoroethylene grafts for femoropopliteal bypass: a prospective randomised multicentre trial. *Eur J Vasc Endovasc Surg*. 2001;22(3):226–31.
65. Stump MM, Jordan GL, DeBakey ME, et al. Endothelium grown from circulating blood on isolated intravascular Dacron hub. *Am J Pathol*. 1963;43(3):361–7.
66. Berger K, Sauvage LR, Rao AM, et al. Healing of arterial prostheses in man: its incompleteness. *Ann Surg*. 1972;175(1):118–27.
67. Herring MB, Dilley R, Jersild RA. Seeding arterial prostheses with vascular endothelium. The nature of the lining. *Ann Surg*. 1979;190(1):84–90.
68. Stanley JC, Burkel WE, Ford JW, et al. Enhanced patency of small-diameter, externally supported Dacron iliofemoral grafts seeded with endothelial cells. *Surgery*. 1982;92(6):994–1005.
69. Smyth JV, Welch M, Carr HMH, et al. Fibrinolysis profiles and platelet activation after endothelial cell seeding of prosthetic vascular grafts. *Ann Vasc Surg*. 1995;9(6):542–6.
70. Rosenman JE, Kempczinski RF, Pearce WH, et al. Kinetics of endothelial cell seeding. *J Vasc Surg*. 1985;2(6):778–84.
71. Zilla P. In vitro endothelialization: its contribution towards an ideal vascular replacement. *Madame Curie Bioscience Database* [Internet]. Austin: Landes Bioscience; 2000.
72. Radomski JS, Jarrell BE, Williams SK, et al. Initial adherence of human capillary endothelial cells to Dacron. *J Surg Res*. 1987;42(2):133–40.
73. Foxall TL, Auger KR, Callow AD, et al. Adult human endothelial cell coverage of small-caliber Dacron and polytetrafluoroethylene vascular prostheses in vitro. *J Surg Res*. 1986;41(2):158–72.
74. Ott MJ, Ballermann BJ. Shear stress-conditioned, endothelial cell-seeded vascular grafts: improved cell adherence in response to in vitro shear stress. *Surgery*. 1995;117(3):334–9.
75. Dardik A, Liu A, Ballermann BJ. Chronic in vitro shear stress stimulates endothelial cell retention on prosthetic vascular grafts and reduces subsequent in vivo neointimal thickness. *J Vasc Surg*. 1999;29(1):157–67.
76. Herring M, Gardner A, Glover J. Seeding human arterial prostheses with mechanically derived endothelium: the detrimental effect of smoking. *J Vasc Surg*. 1984;1(2):279–89.
77. Herring MH, Comiton RS, Legrand DR, et al. Endothelial seeding of polytetrafluoroethylene popliteal bypasses: a preliminary report. *J Vasc Surg*. 1987;6(2):114–8.
78. Deutsch M, Meinhart J, Fischlein T, et al. Clinical autologous in vitro endothelialization of infrainguinal ePTFE grafts in 100 patients: a 9-year experience. *Surgery*. 1999;126(5):847–55.
79. Deutsch M, Meinhart J, Zilla P, et al. Long-term experience in autologous in vitro endothelialization of infrainguinal ePTFE grafts. *J Vasc Surg*. 2009;49(2):352–62.