# Chapter 30 Physiology of Stem Cells

Jos Domen and Kimberly Gandy

## **Stem Cells: History**

Mammalian life, at conception, starts with a single zygote that has the ability to form all of the cells of the body (as well as extra-embryonic tissues). This cell by definition is a pluripotent stem cell, that is, it is capable of assuming all possible cell fates. It has long been recognized that certain tissues, such as blood, skin and gut epithelium, have a high turnover throughout life, and need to be replenished continuously from progenitor or stem cells. The need for stem cells in adults is less well understood, and only recently, has stem cell potential been identified in mature tissues that do not have rapid turnover.

A leading impetus in the search for stem cells was the aftermath of the atomic bomb explosions in World War II. It became clear that exposure to radiation could kill the ability of the body to generate new blood cells, and at somewhat higher doses, could destroy the ability of the intestinal tract to regenerate. Both conditions result in death. Mouse experiments confirmed the lethal consequences of irradiation and demonstrated that the ability to generate blood cells could be preserved if just one limb was shielded from radiation [1]. This was followed by the equally critical observation that transfer of non-irradiated bone marrow cells could restore the ability to generate blood cells [2]. It was then determined that irradiation followed by bone marrow transfer resulted in the appearance of colonies of myeloid and erythroid cells in the spleen and that all of the cells in a colony were derived from one

J. Domen, PhD (🖂)

K. Gandy, MD, PhD Department of Biomedical and Health Informatics, University of Missouri, 4451 Francis Street, Kansas City, KS 66103, USA e-mail: kgandy@playithealth.com

Section of Cardiac Surgery, Children's Mercy Hospital and Clinics and University of Missouri, Room 3730-08, 2401 Gilham Road, Kansas City, MO 64108, USA e-mail: adomen@cmh.edu

cell [3]. Hematopoietic Stem Cells are the small subset of spleen colony-forming cells that can give rise to both secondary colonies, as well as lymphoid offspring [4, 5]. Specific labeling methods and reconstitution assays developed since [6, 7] have greatly improved our ability to study these cells.

Not only has a great deal been learned about these hematopoietic stem cells in the last 50 years, but they have also found widespread application in the clinic. Mimicking the original observation that bone marrow could rescue the hematopoietic system in an otherwise lethally irradiated recipient, bone marrow transplantation has become standard of care to treat cancer patients that undergo high dose chemo and/or radiation therapy, or that suffer from hematopoietic malignancies. Worldwide, more than 50,000 people receive bone marrow transplantations each year [8]. Approximately 21,000 are allogeneic, the rest are autologous transplants. Increasingly, mobilized peripheral blood or umbilical cord blood is used as the source.

Stem cells in most other systems are less well characterized, and in many cases, have only been recognized fairly recently as being present in adults. Clinical use is limited, at best, to trials.

#### **Stem Cells: Definition**

Stem cells are defined as cells that have, at the individual cell level, the ability to both self-renew (make more copies of themselves) and differentiate into one, many or all cell fates of the body (Fig. 30.1). Stem cells are typically rare and are often quiescent.

Self-renewal in its strictest sense implies unlimited proliferative capacity, since the daughters are identical to the parent cell. This is impossible to verify experimentally, at least without unlimited funding and time. A more practical definition would be the ability to function and produce cells beyond the normal lifespan of the organ-



**Fig. 30.1** Stem and progenitor cells defined by proliferative and differentiation capacity. This figure depicts the reduction in proliferative potential and differentiation potential during cell maturation. It should be kept in mind that these differences are not absolute. Some mature cells, e.g. lymphocytes, can proliferate extensively, and some mature cells retain the potential to differentiate further. B cell class switching would be an example

ism. E.g. murine hematopoietic stem cells can be transplanted serially, and repopulate new hosts, at least five times [9-11]. While this is not unlimited, it clearly suffices for the animal, even if the need for cells is higher than normal, such as following (repeated) serious injury and blood loss. In addition, it is important to note that the proliferative limitations encountered may well be a result of the experimental conditions (repeated transplantation into an irradiated environment) and do not necessarily represent the true limitations inherent to the stem cells themselves.

Cells that come much closer to demonstrating unlimited proliferative capacity are cells that can be maintained in culture, such as embryonic stem cells. Many of the murine ES cell lines have been in culture now for over 20 years, and have been used and expanded extensively by many different laboratories, without losing the ability to act as pluripotent stem cells with the capacity to regenerate a mouse.

The differentiation capacity is the main distinguishing feature between different types of stem cells. Pluripotent stem cells such as embryonic stem cells and iPS cells have the ability to generate all the different cells in the body (and can generate the organism as a whole). These cells represent germ line stem cells, the cells that allow life to pass from one generation to the next. Other types of stem cells in the adult body (sometimes referred to as "adult stem cells") are more restricted; they can differentiate into many (multipotent) or a few (oligopotent) cell lineages within the germ layer from which they are derived. Mesenchymal stem cells and hematopoietic stem cells, for instance, can make subsets of mature mesoderm cells. The ability of stem cells to produce cells derived from other germ layers was hotly debated a decade ago, a debate that was referred to as the stem cell plasticity debate. It was claimed, for instance, that blood cells could differentiate into brain cells, or liver cells [12, 13]. More extensive analysis revealed, however, that much of this apparent plasticity was due to experimental artifacts, such as cell fusion. Plasticity or transdifferentiation--differentiation into a lineage that is not part of the normal repertoire of the stem cell, seems to happen rarely, if at all, under normal conditions [14–16]. Interestingly, what has surfaced since this debate is the ability to reprogram cells, even regular somatic cells without stem cell ability, into pluripotent stem cells (the so-called iPS cells) with the ability to produce cells from endodermal, mesodermal and ectodermal origin [17–21].

Once stem cells commit to differentiation, and leave the stem cell pool, they typically go through an intermediate stage often referred to as progenitors (Fig. 30.2). In some systems other names are used for these cells: In skin, for instance, they are called transit-amplifying cells. Practically, the difference between stem and progenitor cells can be hard to make. Progenitor cells may still have the potential to generate several different types of mature cells. Progenitor cells often have extensive proliferative capacity, and much of the expansion needed to amplify the offspring of the rare stem cells into the many mature cells needed in tissues such as skin, blood and gut epithelium, occurs at the progenitor level. However, progenitor cells do not self-renew. With every cell division they move closer to the mature state. New cells from any given progenitor cell are only generated for a relatively brief period of time, whereas stem cells may produce new cells throughout life. However, the distinction between newly generated and older pre-existing cells can be hard to make when the mature cells are long-lived.



**Fig. 30.2** Classical stem cell differentiation pattern. This would apply to hematopoietic cell differentiation, but also to other types of cells, e.g. skin [22]. Maintenance of stem cells is assured by stem cell self-renewal. Much of the proliferation needed to obtain the required number of mature cells occurs at the progenitor level. Progenitors can initially be oligopotent (more than one cell fate possible) or can be committed to a single outcome. Full commitment of progenitors to a new more restricted stage can take several days. Mature cells, depending on type, may retain the capacity for extensive proliferation (e.g. lymphocytes) or may not be able to divide at all (red blood cells, muscle cells)

Diagrams, such as the tree shown in Fig. 30.2 often depict progenitors as very distinct entities, with differentiation steps taking them from more primitive to less primitive, more restricted, progenitors, eventually resulting in mature cells. These entities are based on purification methods such as Fluorescence Activated Cell Sorting (FACS) or assays such as colony assays. The ability to purify, study and use progenitors with specific potential is very useful and informative. The cells themselves, however, can be thought of as being on a continuum, gradually shifting from one phenotype to the next, gradually restricting their differentiation potential.

#### **Stem Cells: Niche and Regulation**

Obviously, in view of their proliferative potential, stem cells need extensive regulation. While some stem cells, such as neuronal stem cells, are mostly quiescent and generate few offspring under normal conditions, other stem cells have to continuously generate large numbers of mature cells. For example, in the hematopoietic system more than 10<sup>11</sup> mature cells need to be produced every day [23–25]. To control this expansion, and prevent it from escaping control, stem cells typically need specific signals to maintain their stem cell potential. In the absence of these signals the cells either differentiate along a default pathway, or undergo apoptosis. Yet it is also essential that the stem cell numbers are maintained. Loss of e.g. colon or hematopoietic stem cells would result in death in days to weeks. Stem cell homeostasis (Fig. 30.3) requires that under steady state conditions, following stem cell division, on average one of the daughter cells remains a stem cell, while the other cell either commits to differentiation, or undergoes apoptosis. The place where



**Fig. 30.4** The concept of the stem cell niche. Shown are two examples of stem cell niches, the Drosophila spermatogonium, and the mammalian hematopoietic stem cell. (**a**) In this niche concept the germ and somatic cells in the apical tip of the testis that are in direct contact with the hub cells remain stem cells. Once this contact is lost, through oriented cell division, the cells start their differentiation toward spermatozoa, which takes many steps and several more cell division. (**b**) In the hematopoietic system the organization is more flexible in that HSC have the ability to move from their resting niches near the bone to the vasculature, and back. Both environments differ significantly, e.g. in  $O_2$  levels. Most differentiation (including the accompanying proliferation) also takes place in the bone marrow. The resulting mature cells enter the blood to leave the bone marrow

stem cells receive these signals is often referred to as the stem cell niche (Fig. 30.4). This can be a very defined, physical environment. Figure 30.4 shows two examples of stem cell niches. In the *Drosophila* spermatogonium the location of the stem cells that produce the spermatozoa is exactly defined. They are at the tip of the gonad, in direct contact with other cells known as hub-cells. These hub cells provide the signals necessary for the stem cells to remain stem cells. Once the stem cells divide the cell division is oriented such that one daughter cell remains in contact with the hub

cells, and the other daughter loses contact. The daughter cell that has lost contact commits to differentiation, resulting in an obligatory asymmetric cell division. A similar asymmetric division occurs for the accompanying cyst cells that surround the germ cell throughout the differentiation process. The critical signal, through the Jak/Stat pathway, provided by the hub cells is called Unpaired (Upd), which is a ligand for the receptor Domeless (Dome) and the associated Jak kinase Hopscotch (Hop). Upon binding of Upd the receptor is phosphorylated by Hop, followed by Stat binding, phosphorylation, dimerization, and translocation to the nucleus. Mutants in which the cyst cells overexpress the ligand Upd never form functional sperm. Instead the spermatogonium fills up with stem cells. Mutants without functional Stat do not maintain stem cells, but have a single early wave of spermatogenesis [26, 27].

In the mammalian bone marrow the location of the hematopoietic stem cells has been less clear. One reason is that bone marrow is typically studied as a cell suspension by flow cytometry and colony- or reconstitution assays after it has been removed from the marrow spaces, rather than by histology in situ. Also, regulation is more complex in that hematopoietic stem cells can leave their niches in the bone marrow and travel through the vasculature to new locations such as spleen and liver. They do so in a regulated fashion several times during development [28], but retain the ability to migrate in adult life as a response to various stimuli [29, 30], with  $CXCR4/SDF-1\alpha$  signaling as a central component. Clinically, this is used to harvest them: Rather than harvest bone marrow directly hematopoietic stem cells are harvested as Mobilized Peripheral Blood (MPB) by leukapheresis after treatment of the donor with the growth factor G-CSF and the chemotherapeutic cyclophosphamide. Nevertheless, much has been learned in recent years about the important cells in the niche, and the signals governing stem cell behavior. An in-depth discussion is outside of the scope of this more general chapter, and many reviews can be used as starting points to further probe the literature, see e.g. [31–36]. A variety of cells have been reported to be part of this niche, including osteoblasts [37-39], sinusoidal endothelium [40], CXCL12 expressing reticular cells [41], adipocytes [42] and mesenchymal stem cells [43]. A number of regulators have been postulated to be involved in stem cell maintenance in this niche. These include Wnt proteins [44-47], the Notch pathway [48–50], insulin-like growth factor (IGF-2) and angiopoietinlike proteins [51–53]. Important regulators of cell cycle progression include the p16<sup>Ink4a</sup>–CDk4/6Rb and p19<sup>Arf</sup>-P53-P21<sup>Cip1</sup> pathways [54, 55]. Another important factor for stem cells is the ability to maintain telomere length during successive cell divisions. To accomplish this stem cells express telomerase. Loss of telomerase can limit stem cell self-renewal during serial transplantation [56] and aging [57]. It has recently been reported that telomerase activity is regulated by Wnt/β-catenin in stem cells [58]. Despite all of the progress that has been made in characterizing the regulation of hematopoietic stem cells no clear conditions have been defined yet that allow for robust expansion of these cells outside of the body [59], something that would be extremely useful in broadening their therapeutic potential. This indicates that our understanding of HSC self-renewal, and the molecular players involved, is still lacking.

Other types of stem cells, in particular Embryonic Stem cells, are capable of extensive expansion outside of the body using defined conditions without reduction of their pluripotency, their ability to differentiate into all tissues of the body. Much has been learned about the factors governing unlimited self-renewal and developmental potential [55, 60–62]. Central components include the transcription factors Oct4, Sox2 and Nanog. These transcription factors, which interact physically, form a finely tuned network, disturbance of which can lead to loss of pluripotency. Additional proteins, like the zinc-finger DNA binding protein Ronin, are also involved and can prevent differentiation [63]. In addition there is regulation at the epigenetic level, with Polycomp complexes playing an important role [62]. Ultimately demonstrating the importance of these pathways has been the ability to use forced expression of several of these proteins to convert somatic cells into pluripotent stem cells [61] (See iPSC section below.)

Overall, these examples illustrate that niches are important and specific, and their presence can limit and regulate stem cell presence. This regulation is especially critical for cells that continuously produce large numbers of mature cells. Loss of control of expansion would quickly result in a proliferative disease, or even cancer.

#### Stem Cells: Clinical Use

The specifics of the use of stem cells in the heart are discussed elsewhere in this book. This section will be limited to a more general overview of the current and potential use of stem cells in medicine. As mentioned earlier, hematopoietic stem cell transplantation is currently in widespread use, albeit that this involves transfer of stem cell containing cell preparations, and not highly purified HSC. This is an important distinction, as the main indication for use is the treatment of malignancies. Autologous transplants in which patients receive their own cells, harvested prior to intensive chemotherapy, thus carry a risk of reseeding the patient with cancer cells that contaminate the graft. Small-scale studies with highly purified autologous hematopoietic stem cells indicate that high level purification indeed improves outcome [64, 65]. Allogeneic transplants, in which the patient receives cells from a different individual, do not carry this risk, but, specific for hematopoietic transplants, may result in adaptive immune cells in the graft reacting against the host body. This is known as graft-versus host disease, and is a potentially lethal complication of allogeneic hematopoietic cell transplantation [66–68].

Hematopoietic stem cells have the potential, and are starting to be used for many other therapies [69], including the treatment of autoimmune diseases [70], the treatment of inherited metabolic diseases [71] and the induction of tolerance for solid organ transplantation [69, 72–74].

Skin is another tissue that is transplanted in routine clinical use, and that depends for functioning on the transplanted stem cells [75–77]. Skin, like blood, is continuously regenerated from stem cells. Old cells slough off and are discarded. Autologous

transplants, in which skin is harvested at one place of the body and transplanted to another, e.g. to cover a burn, results in lasting engraftment. Allogeneic skin transplants, typically harvested from a deceased donor, are used as temporary cover, as they will be rejected by the immune system.

Other types of stem cells, especially mesenchymal stem cells, are being tested in clinical trials [78–80], including trials addressing cardiac failures [81–83], but are not yet in routine use. Mesenchymal stem cells can be harvested relatively easily from various sources, including bone marrow and adipose tissue, expanded in culture and can differentiate into many tissues, including bone, adipose, myogenic and hepatic cells [84]. In addition, mesenchymal stem cells can modulate immune responses [78, 80]. Neuronal stem cells are also being tested in initial clinical trials, e.g. [85–87]. Other cells, including ES cells [88, 89] and iPS cells [21, 90–93], are being developed for eventual clinical use.

## Stem Cells and Cancer

Stem cells, through their ability to persist for long periods of time while cycling, even if slowly, are prime candidates to collect the mutations necessary to allow escape from their normal controls and become transformed. Like normal, non-transformed, stem cells cancer stem cells may remain dependent on a stem cell niche for some of their regulatory (pro-mitogenic) signals [35, 94], even though they may have lost other parts of the normal regulatory circuitry, like essential tumor suppressor genes that should prevent these cells from pro-liferating [55, 95].

Interestingly, it has been recognized recently that cancers themselves tend to be organized in cancer stem cells, which may only make up a small part of the tumor and the bulk of the tumor cells which are derived from these stem cells, analogous to differentiated cells in normal tissues [96–101]. In this model the cancer stem cells are both necessary to maintain the tumor, and sufficient, if not eradicated completely, to regrow the tumor (relapse) after treatment.

#### Induced Pluripotent Stem Cells (IPSC)

One of the most exciting developments in stem cell biology in the last decade has been the discovery that a stem cell phenotype, even that of pluripotent stem cells, can be induced in somatic cells following transduction with a limited set of genes. In the initial landmark studies mouse fibroblasts, transduced with *Oct4, Sox2, Klf4* and *c-Myc*, were reprogrammed into cells very similar to Embryonic Stem cells [19]. These results were rapidly confirmed with human cells, and a slightly different gene combination (*Oct4, Sox2, Nanog* and *Lin28*) [18, 20]. Many different types of

cells are open to reprogramming, including fibroblasts (above), keratinocytes [102], neural stem cells, hepatocytes and gastric epithelial cells [103], adipocytes and hematopoietic cells [104].

IPS cells have a number of obvious advantages over ES cells. Unlike ES cells, their origin is not ethically controversial. Furthermore, the ability to generate pluripotent cells from any donor holds great promises for the development of specific disease models. However, it remains to be established how closely iPS cells resemble ES cells, and the presence, at least in the initial experiments, of the oncogene c-Myc in the transformation mix gives pause when contemplating clinical use [61]. The optimal gene combination for induction of iPS cells remains a subject of interest [61], complicated by the more recent observations that show that it is possible to reprogram cells to a different fate without passing through the pluripotent intermediate stage [105, 106]. It is possible, e.g. to reprogram fibroblasts directly into neurons [107] or cardiomyocytes [108]. While in early stages, this clearly further increases the possibilities for creating genetically matched tissues for research and therapy.

#### Stem Cells and the Heart

When discussing stem cells in the context of the heart in general, or heart failure in particular, there are several different points of view that can be considered. Stem cells play a role in generating the structures of the heart [109]. Heart specification starts with the so-called first heart field, which will initially form a tube with endocardial layer on the inside, and a myocardial layer on the outside. Through differential growth and folding this will eventually result in a multi-chambered heart, with cells from the first heart field forming the left side of the heart, while cells from a second heart field mostly form the right side and outflow tract [110, 111]. To some extent, progenitors remain present once the heart is formed and provide a certain level of regenerative potential. There may be other stem cells as well that can be used to ameliorate the function of damaged heart tissue [112–114]. In addition to endogenous heart cells [115], there are other somatic stem cells (like mesenchymal stem cells) [116] and pluripotent stem cells (ES cells or iPS cells) [21, 110]. The therapeutic use of stem cells for cardiac repair is discussed in more detail elsewhere in this book.

Interestingly, while regeneration of functional myocardial tissue is limited in heart failure following myocardial infarct or other damage, cardiac progenitor cells are present in the myocardium, e.g. [117–119]. It may be possible to use these cells to improve function in failing hearts. In addition to the presence of cardiac progenitors, it has recently also been shown that at least some existing cardiomyocytes retain the potential to divide and replace cells [120]. This potential of cardiomyocyte proliferation can be greatly stimulated with exogenous administration of select micro RNA's (miRNA's) [121].

# Conclusion

Stem cells play an essential role, during development and continuing in adult life. While much has been learned about the biology of stem cells in different organs and organisms, much more remains to be learned. The clinical promise is enormous, and current use barely scratches the surface. To develop this promise into eventual clinical reality it will be essential to follow the winding path, despite its frequent switchbacks, carefully. As tempting as shortcuts can be, in systems as complex as those discussed here, a methodical approach is the only reliable method of distinguishing what does and does not work when translated to the clinic. The biggest danger in attempting and failing at shortcuts to the clinic is establishing as common knowledge that something "does not work" before it has been tested. This can prevent promising avenues from being explored and developed.

# References

- 1. Jacobsen LO, Marks EK, Robson MJ, Gaston EO, Zirkle RE. Effect of spleen protection on mortality following x-irradiation. J Lab Clin Med. 1949;34:1538.
- 2. Lorenz E, Uphoff D, Reid TR, Shelton E. Modification of irradiation injury in mice and guinea pigs by bone marrow injections. J Natl Cancer Inst. 1951;12:197–201.
- Becker AJ, Mc CE, Till JE. Cytological demonstration of the clonal nature of spleen colonies derived from transplanted mouse marrow cells. Nature. 1963;197:452–4.
- Siminovitch L, McCulloch EA, Till JE. The distribution of colony-forming cells among spleen colonies. J Cell Physiol. 1963;62:327–36.
- Wu AM, Till JE, Siminovitch L, McCulloch EA. Cytological evidence for a relationship between normal hemotopoietic colony-forming cells and cells of the lymphoid system. J Exp Med. 1968;127(3):455–64.
- 6. Spooncer E, Lord BI, Dexter TM. Defective ability to self-renew in vitro of highly purified primitive haematopoietic cells. Nature. 1985;316(6023):62–4.
- Spangrude GJ, Heimfeld S, Weissman IL. Purification and characterization of mouse hematopoietic stem cells. Science. 1988;241(4861):58–62.
- Gratwohl A, Baldomero H, Aljurf M, Pasquini MC, Bouzas LF, Yoshimi A, et al. Hematopoietic stem cell transplantation: a global perspective. JAMA. 2010;303(16): 1617–24.
- Harrison DE, Astle CM. Loss of stem cell repopulating ability upon transplantation. Effects of donor age, cell number, and transplantation procedure. J Exp Med. 1982;156(6): 1767–79.
- 10. Pawliuk R, Eaves C, Humphries RK. Evidence of both ontogeny and transplant doseregulated expansion of hematopoietic stem cells in vivo. Blood. 1996;88(8):2852–8.
- 11. Iscove NN, Nawa K. Hematopoietic stem cells expand during serial transplantation in vivo without apparent exhaustion. Curr Biol. 1997;7(10):805–8.
- Lagasse E, Connors H, Al-Dhalimy M, Reitsma M, Dohse M, Osborne L, et al. Purified hematopoietic stem cells can differentiate into hepatocytes in vivo. Nat Med. 2000;6(11):1229–34.
- Mezey. E, Chandross KJ, Harta G, Maki RA, McKercher SR. Turning blood into brain: cells bearing neuronal antigens generated in vivo from bone marrow. Science. 2000;290(5497): 1779–82.
- 14. Wagers AJ, Weissman IL. Plasticity of adult stem cells. Cell. 2004;116(5):639-48.

- Camargo FD, Chambers SM, Goodell MA. Stem cell plasticity: from transdifferentiation to macrophage fusion. Cell Prolif. 2004;37(1):55–65.
- 16. Raff M. Adult stem cell plasticity: fact or artifact? Annu Rev Cell Dev Biol. 2003;19:1-22.
- Ho PJ, Yen ML, Yet SF, Yen BL. Current applications of human pluripotent stem cells: possibilities and challenges. Cell Transplant. 2012;21(5):801–14.
- Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. Cell. 2007;131(5):861–72.
- 19. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell. 2006;126(4):663–76.
- Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, et al. Induced pluripotent stem cell lines derived from human somatic cells. Science. 2007;318(5858): 1917–20.
- Zwi-Dantsis L, Gepstein L. Induced pluripotent stem cells for cardiac repair. Cell Mol Life Sci. 2012;69(19):3285–99.
- 22. Benitah SA, Frye M. Stem cells in ectodermal development. J Mol Med (Berl). 2012;90(7):783–90.
- Domen J, Wagers AJ, Weissman IL. Bone marrow (hematopoietic stem cells). Regen Med. 2006;2006:13–34. Available from: http://stemcells.nih.gov/info/scireport/2006report.htm.
- Lensch MW. An evolving model of hematopoietic stem cell functional identity. Stem Cell Rev. 2012;8(2):551–60.
- Takizawa H, Manz MG. In vivo divisional tracking of hematopoietic stem cells. Ann N Y Acad Sci. 2012;1266:40–6.
- Hombria JC, Brown S. The fertile field of Drosophila Jak/STAT signalling. Curr Biol. 2002;12(16):R569–75.
- Resende LP, Jones DL. Local signaling within stem cell niches: insights from Drosophila. Curr Opin Cell Biol. 2012;24(2):225–31.
- Christensen JL, Wright DE, Wagers AJ, Weissman IL. Circulation and chemotaxis of fetal hematopoietic stem cells. PLoS Biol. 2004;2(3):E75.
- 29. Hoggatt J, Pelus LM. Mobilization of hematopoietic stem cells from the bone marrow niche to the blood compartment. Stem Cell Res Ther. 2011;2(2):13.
- Greenbaum AM, Link DC. Mechanisms of G-CSF-mediated hematopoietic stem and progenitor mobilization. Leukemia. 2011;25(2):211–7.
- Shiozawa Y, Havens AM, Pienta KJ, Taichman RS. The bone marrow niche: habitat to hematopoietic and mesenchymal stem cells, and unwitting host to molecular parasites. Leukemia. 2008;22(5):941–50.
- 32. Garrett RW, Emerson SG. Bone and blood vessels: the hard and the soft of hematopoietic stem cell niches. Cell Stem Cell. 2009;4(6):503–6.
- Lilly AJ, Johnson WE, Bunce CM. The haematopoietic stem cell niche: new insights into the mechanisms regulating haematopoietic stem cell behaviour. Stem Cells Int. 2011;2011:274564.
- Mercier FE, Ragu C, Scadden DT. The bone marrow at the crossroads of blood and immunity. Nat Rev Immunol. 2012;12(1):49–60.
- Tieu KS, Tieu RS, Martinez-Agosto JA, Sehl ME. Stem cell niche dynamics: from homeostasis to carcinogenesis. Stem Cells Int. 2012;2012:367567.
- 36. Shiozawa Y, Taichman RS. Getting blood from bone: an emerging understanding of the role that osteoblasts play in regulating hematopoietic stem cells within their niche. Exp Hematol. 2012;40(9):685–94.
- 37. Arai F, Hirao A, Ohmura M, Sato H, Matsuoka S, Takubo K, et al. Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche. Cell. 2004;118(2):149–61.
- Calvi LM, Adams GB, Weibrecht KW, Weber JM, Olson DP, Knight MC, et al. Osteoblastic cells regulate the haematopoietic stem cell niche. Nature. 2003;425(6960):841–6.
- Zhang J, Niu C, Ye L, Huang H, He X, Tong WG, et al. Identification of the haematopoietic stem cell niche and control of the niche size. Nature. 2003;425(6960):836–41.

- Ding L, Saunders TL, Enikolopov G, Morrison SJ. Endothelial and perivascular cells maintain haematopoietic stem cells. Nature. 2012;481(7382):457–62.
- Sugiyama T, Kohara H, Noda M, Nagasawa T. Maintenance of the hematopoietic stem cell pool by CXCL12-CXCR4 chemokine signaling in bone marrow stromal cell niches. Immunity. 2006;25(6):977–88.
- Naveiras O, Nardi V, Wenzel PL, Hauschka PV, Fahey F, Daley GQ. Bone-marrow adipocytes as negative regulators of the haematopoietic microenvironment. Nature. 2009;460(7252):259–63.
- Mendez-Ferrer S, Michurina TV, Ferraro F, Mazloom AR, Macarthur BD, Lira SA, et al. Mesenchymal and haematopoietic stem cells form a unique bone marrow niche. Nature. 2010;466(7308):829–34.
- 44. Reya T, Duncan AW, Ailles L, Domen J, Scherer DC, Willert K, et al. A role for Wnt signalling in self-renewal of haematopoietic stem cells. Nature. 2003;423(6938):409–14.
- Willert K, Brown JD, Danenberg E, Duncan AW, Weissman IL, Reya T, et al. Wnt proteins are lipid-modified and can act as stem cell growth factors. Nature. 2003;423(6938):448–52.
- 46. Murdoch B, Chadwick K, Martin M, Shojaei F, Shah KV, Gallacher L, et al. Wnt-5 A augments repopulating capacity and primitive hematopoietic development of human blood stem cells in vivo. Proc Natl Acad Sci U S A. 2003;100(6):3422–7.
- 47. Luis TC, Naber BA, Roozen PP, Brugman MH, de Haas EF, Ghazvini M, et al. Canonical wnt signaling regulates hematopoiesis in a dosage-dependent fashion. Cell Stem Cell. 2011;9(4):345–56.
- Varnum-Finney B, Xu L, Brashem-Stein C, Nourigat C, Flowers D, Bakkour S, et al. Pluripotent, cytokine-dependent, hematopoietic stem cells are immortalized by constitutive Notch1 signaling. Nat Med. 2000;6(11):1278–81.
- 49. Varnum-Finney B, Brashem-Stein C, Bernstein ID. Combined effects of Notch signaling and cytokines induce a multiple log increase in precursors with lymphoid and myeloid reconstituting ability. Blood. 2003;101(5):1784–9.
- Delaney C, Heimfeld S, Brashem-Stein C, Voorhies H, Manger RL, Bernstein ID. Notchmediated expansion of human cord blood progenitor cells capable of rapid myeloid reconstitution. Nat Med. 2010;16(2):232–6.
- 51. Zhang CC, Lodish HF. Insulin-like growth factor 2 expressed in a novel fetal liver cell population is a growth factor for hematopoietic stem cells. Blood. 2004;103(7):2513–21.
- 52. Huynh H, Iizuka S, Kaba M, Kirak O, Zheng J, Lodish HF, et al. Insulin-like growth factorbinding protein 2 secreted by a tumorigenic cell line supports ex vivo expansion of mouse hematopoietic stem cells. Stem Cells. 2008;26(6):1628–35.
- 53. Zhang CC, Kaba M, Iizuka S, Huynh H, Lodish HF. Angiopoietin-like 5 and IGFBP2 stimulate ex vivo expansion of human cord blood hematopoietic stem cells as assayed by NOD/ SCID transplantation. Blood. 2008;111(7):3415–23.
- 54. Sherr CJ. Principles of tumor suppression. Cell. 2004;116(2):235-46.
- He S, Nakada D, Morrison SJ. Mechanisms of stem cell self-renewal. Annu Rev Cell Dev Biol. 2009;25:377–406.
- Allsopp RC, Morin GB, DePinho R, Harley CB, Weissman IL. Telomerase is required to slow telomere shortening and extend replicative lifespan of HSCs during serial transplantation. Blood. 2003;102(2):517–20.
- 57. Rossi DJ, Bryder D, Seita J, Nussenzweig A, Hoeijmakers J, Weissman IL. Deficiencies in DNA damage repair limit the function of haematopoietic stem cells with age. Nature. 2007;447(7145):725–9.
- Hoffmeyer K, Raggioli A, Rudloff S, Anton R, Hierholzer A, Del Valle I, et al. Wnt/betacatenin signaling regulates telomerase in stem cells and cancer cells. Science. 2012;336(6088):1549–54.
- 59. Walasek MA, van Os R, de Haan G. Hematopoietic stem cell expansion: challenges and opportunities. Ann N Y Acad Sci. 2012;1266:138–50.
- Jaenisch R, Young R. Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming. Cell. 2008;132(4):567–82.

- De Los Angeles A, Loh YH, Tesar PJ, Daley GQ. Accessing naive human pluripotency. Curr Opin Genet Dev. 2012;22(3):272–82.
- 62. Li M, Liu GH, Izpisua Belmonte JC. Navigating the epigenetic landscape of pluripotent stem cells. Nat Rev Mol Cell Biol. 2012;13(8):524–35.
- Dejosez M, Krumenacker JS, Zitur LJ, Passeri M, Chu LF, Songyang Z, et al. Ronin is essential for embryogenesis and the pluripotency of mouse embryonic stem cells. Cell. 2008;133(7):1162–74.
- 64. Negrin RS, Atkinson K, Leemhuis T, Hanania E, Juttner C, Tierney K, et al. Transplantation of highly purified CD34+Thy-1+ hematopoietic stem cells in patients with metastatic breast cancer. Biol Blood Marrow Transplant. 2000;6(3):262–71.
- 65. Muller AM, Kohrt HE, Cha S, Laport G, Klein J, Guardino AE, et al. Long-term outcome of patients with metastatic breast cancer treated with high-dose chemotherapy and transplantation of purified autologous hematopoietic stem cells. Biol Blood Marrow Transplant. 2012;18(1):125–33.
- 66. Harris AC, Ferrara JL, Levine JE. Advances in predicting acute GVHD. Br J Haematol. 2013;160(3):288–302 . Published online 2012 Dec 4. doi:10.1111/bjh.12142.
- 67. Harris AC, Levine JE, Ferrara JL. Have we made progress in the treatment of GVHD? Best Pract Res Clin Haematol. 2012;25(4):473–8.
- Petersdorf EW. Genetics of graft-versus-host disease: The major histocompatibility complex. Blood Rev. 2013;27(1):1–12. doi:10.1016/j.blre.2012.10.001. Epub 2012 Nov 20.
- Domen J, Gandy K, Dalal J. Emerging uses for pediatric hematopoietic stem cells. Pediatr Res. 2012;71(4–2):411–7.
- Sullivan KM, Muraro P, Tyndall A. Hematopoietic cell transplantation for autoimmune disease: updates from Europe and the United States. Biol Blood Marrow Transplant. 2010;16(1 Suppl):S48–56.
- Prasad VK, Kurtzberg J. Cord blood and bone marrow transplantation in inherited metabolic diseases: scientific basis, current status and future directions. Br J Haematol. 2010;148(3):356–72.
- 72. Strober S. Protective conditioning against GVHD and graft rejection after combined organ and hematopoietic cell transplantation. Blood Cells Mol Dis. 2008;40(1):48–54.
- Strober S, Spitzer TR, Lowsky R, Sykes M. Translational studies in hematopoietic cell transplantation: Treatment of hematologic malignancies as a stepping stone to tolerance induction. Semin Immunol. 2011;23(4):273–81.
- Sachs DH, Sykes M, Kawai T, Cosimi AB. Immuno-intervention for the induction of transplantation tolerance through mixed chimerism. Semin Immunol. 2011;23(3):165–73.
- 75. Ghadially R. 25 years of epidermal stem cell research. J Invest Dermatol. 2012;132(3 Pt 2):797–810.
- Goldstein J, Horsley V. Home sweet home: skin stem cell niches. Cell Mol Life Sci. 2012;69(15):2573–82.
- Yan X, Owens DM. The skin: a home to multiple classes of epithelial progenitor cells. Stem Cell Rev. 2008;4(2):113–8.
- Dalal J, Gandy K, Domen J. Role of mesenchymal stem cell therapy in Crohn's disease. Pediatr Res. 2012;71(4–2):445–51.
- Harrop JS, Hashimoto R, Norvell D, Raich A, Aarabi B, Grossman RG, et al. Evaluation of clinical experience using cell-based therapies in patients with spinal cord injury: a systematic review. J Neurosurg Spine. 2012;17(1 Suppl):230–46.
- 80. Keating A. Mesenchymal stromal cells: new directions. Cell Stem Cell. 2012;10(6):709-16.
- 81. Hare JM, Fishman JE, Gerstenblith G, DiFede Velazquez DL, Zambrano JP, Suncion VY, et al. Comparison of allogeneic vs autologous bone marrow-derived mesenchymal stem cells delivered by transendocardial injection in patients with ischemic cardiomyopathy: the POSEIDON randomized trial. JAMA. 2012;308(22):2369–79.
- Miettinen JA, Salonen RJ, Ylitalo K, Niemela M, Kervinen K, Saily M, et al. The effect of bone marrow microenvironment on the functional properties of the therapeutic bone marrowderived cells in patients with acute myocardial infarction. J Transl Med. 2012;10:66.

- 83. Mathiasen AB, Jorgensen E, Qayyum AA, Haack-Sorensen M, Ekblond A, Kastrup J. Rationale and design of the first randomized, double-blind, placebo-controlled trial of intramyocardial injection of autologous bone-marrow derived Mesenchymal Stromal Cells in chronic ischemic Heart Failure (MSC-HF Trial). Am Heart J. 2012;164(3):285–91.
- Ishikawa T, Banas A, Teratani T, Iwaguro H, Ochiya T. Regenerative cells for transplantation in hepatic failure. Cell Transplant. 2012;21(2–3):387–99.
- Kondziolka D, Wechsler L, Goldstein S, Meltzer C, Thulborn KR, Gebel J, et al. Transplantation of cultured human neuronal cells for patients with stroke. Neurology. 2000;55(4):565–9.
- Luan Z, Liu W, Qu S, Du K, He S, Wang Z, et al. Effects of neural progenitor cell transplantation in children with severe cerebral palsy. Cell Transplant. 2012;21(Suppl 1):S91–8.
- Riley J, Federici T, Polak M, Kelly C, Glass J, Raore B, et al. Intraspinal stem cell transplantation in amyotrophic lateral sclerosis: a phase I safety trial, technical note, and lumbar safety outcomes. Neurosurgery. 2012;71(2):405–16; discussion 416.
- Ben-David U, Kopper O, Benvenisty N. Expanding the boundaries of embryonic stem cells. Cell Stem Cell. 2012;10(6):666–77.
- Serra M, Brito C, Correia C, Alves PM. Process engineering of human pluripotent stem cells for clinical application. Trends Biotechnol. 2012;30(6):350–9.
- 90. Zhu Y, Wan S, Zhan RY. Inducible pluripotent stem cells for the treatment of ischemic stroke: current status and problems. Rev Neurosci. 2012;23(4):393–402.
- Wang H, Doering LC. Induced pluripotent stem cells to model and treat neurogenetic disorders. Neural Plast. 2012;2012:346053.
- 92. van Bekkum DW, Mikkers HM. Prospects and challenges of induced pluripotent stem cells as a source of hematopoietic stem cells. Ann N Y Acad Sci. 2012;1266:179–88.
- Kao DI, Chen S. Pluripotent stem cell-derived pancreatic beta-cells: potential for regenerative medicine in diabetes. Regen Med. 2012;7(4):583–93.
- Boral D, Nie D. Cancer stem cells and niche mircoenvironments. Front Biosci (Elite Ed). 2012;4:2502–14.
- Verga Falzacappa MV, Ronchini C, Reavie LB, Pelicci PG. Regulation of self-renewal in normal and cancer stem cells. FEBS J. 2012;279(19):3559–72.
- Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. Nature. 2001;414(6859):105–11.
- Al-Hajj M, Becker MW, Wicha M, Weissman I, Clarke MF. Therapeutic implications of cancer stem cells. Curr Opin Genet Dev. 2004;14(1):43–7.
- 98. Dick JE. Acute myeloid leukemia stem cells. Ann N Y Acad Sci. 2005;1044:1-5.
- Alison MR, Lin WR, Lim SM, Nicholson LJ. Cancer stem cells: in the line of fire. Cancer Treat Rev. 2012;38(6):589–98.
- Magee JA, Piskounova E, Morrison SJ. Cancer stem cells: impact, heterogeneity, and uncertainty. Cancer Cell. 2012;21(3):83–96.
- 101. Vermeulen L. de Sousa e Melo F, Richel DJ, Medema JP. The developing cancer stem-cell model: clinical challenges and opportunities. Lancet Oncol. 2012;13(2):e83–9.
- 102. Aasen T, Raya A, Barrero MJ, Garreta E, Consiglio A, Gonzalez F, et al. Efficient and rapid generation of induced pluripotent stem cells from human keratinocytes. Nat Biotechnol. 2008;26(11):1276–84.
- 103. Aoi T, Yae K, Nakagawa M, Ichisaka T, Okita K, Takahashi K, et al. Generation of pluripotent stem cells from adult mouse liver and stomach cells. Science. 2008;321(5889):699–702.
- 104. Hanna J, Markoulaki S, Schorderet P, Carey BW, Beard C, Wernig M, et al. Direct reprogramming of terminally differentiated mature B lymphocytes to pluripotency. Cell. 2008;133(2):250–64.
- Chambers SM, Studer L. Cell fate plug and play: direct reprogramming and induced pluripotency. Cell. 2011;145(6):827–30.
- Stadtfeld M, Hochedlinger K. Induced pluripotency: history, mechanisms, and applications. Genes Dev. 2010;24(20):2239–63.

- Vierbuchen T, Ostermeier A, Pang ZP, Kokubu Y, Sudhof TC, Wernig M. Direct conversion of fibroblasts to functional neurons by defined factors. Nature. 2010;463(7284):1035–41.
- Ieda M, Fu JD, Delgado-Olguin P, Vedantham V, Hayashi Y, Bruneau BG, et al. Direct reprogramming of fibroblasts into functional cardiomyocytes by defined factors. Cell. 2010;142(3):375–86.
- 109. Riley PR. An epicardial floor plan for building and rebuilding the mammalian heart. Curr Top Dev Biol. 2012;100:233–51.
- 110. Lui KO, Bu L, Li RA, Chan CW. Pluripotent stem cell-based heart regeneration: from the developmental and immunological perspectives. Birth Defects Res C Embryo Today. 2012;96(1):98–108.
- 111. Buckingham M, Meilhac S, Zaffran S. Building the mammalian heart from two sources of myocardial cells. Nat Rev Genet. 2005;6(11):826–35.
- 112. Bernstein HS, Srivastava D. Stem cell therapy for cardiac disease. Pediatr Res. 2012;71(4 Pt 2):491–9.
- 113. Ptaszek LM, Mansour M, Ruskin JN, Chien KR. Towards regenerative therapy for cardiac disease. Lancet. 2012;379(9819):933–42.
- 114. Hotkar AJ, Balinsky W. Stem cells in the treatment of cardiovascular disease--an overview. Stem Cell Rev. 2012;8(2):494–502.
- 115. Dimarakis I, Menasche P, Habib NA, Gordon MY, editors. Handbook of cardiac stem cell therapy. London: Imperial College Press; 2009. p. 1–285.
- 116. Mullenix PS, Huddleston SJ, Stojadinovic A, Trachiotis GD, Alexander EP. A new heart: somatic stem cells and myocardial regeneration. J Surg Oncol. 2012;105(5):475–80.
- 117. Takamiya M, Haider KH, Ashraf M. Identification and characterization of a novel multipotent sub-population of Sca-1(+) cardiac progenitor cells for myocardial regeneration. PLoS One. 2011;6(9):e25265.
- 118. Ye J, Boyle A, Shih H, Sievers RE, Zhang Y, Prasad M, et al. Sca-1+ cardiosphere-derived cells are enriched for Isl1-expressing cardiac precursors and improve cardiac function after myocardial injury. PLoS One. 2012;7(1):e30329.
- 119. van Wijk B, Gunst QD, Moorman AF, van den Hoff MJ. Cardiac regeneration from activated epicardium. PLoS One. 2012;7(9):e44692.
- 120. Senyo SE, Steinhauser ML, Pizzimenti CL, Yang VK, Cai L, Wang M, et al. Mammalian heart renewal by pre-existing cardiomyocytes. Nature. 2013;493(7432):433–6. doi:10.1038/ nature11682. Epub 2012 Dec 5.
- 121. Eulalio A, Mano M, Ferro MD, Zentilin L, Sinagra G, Zacchigna S, et al. Functional screening identifies miRNAs inducing cardiac regeneration. Nature. 2012;492(7429):376–81. doi:10.1038/nature11739. Epub 2012 Dec 5.