

Chapter 15

Genetics of Behçet's Disease

Ahmet Gül and Shigeaki Ohno

Keywords Behçet's disease • Familial aggregation • Sibling recurrence risk ratio (λ_s) • HLA-B51 • MICA • HLA-A26 • Linkage study • Genomewide association study

Behçet's disease (BD) is a systemic inflammatory disorder of unknown etiology. It is generally accepted as a multifactorial disease with a strong genetic background, and the disease manifestations are considered to be triggered by various environmental factors in genetically susceptible individuals [1, 2].

There are several clues supporting involvement of genetic factors in the pathogenesis of BD, which include familial aggregation, distinct geographic distribution, and its association with the HLA-B51 antigen.

Familial Aggregation

Although majority of BD patients are seen as sporadic cases, increased frequency of BD has long been noted among the relatives [3–16]. Varying frequencies of patients with a positive family history for BD were described in large series of patients, with a tendency for higher figures in the Middle Eastern patients compared to the patients from Asian and European countries [16, 17].

Gül and colleagues analyzed the sibling recurrence risk ratio (λ_s) for quantifying the familial aggregation in BD [17]. They calculated the sibling recurrence rate as 4.2 by taking into account only the immediately older sibling, or if an older sibling is not available, immediately younger sibling for evaluation. By using the prevalence rates of BD in Turkey, λ_s value was found to be 11.4–52.5 for BD [17].

A. Gül (✉)

Department of Internal Medicine, Division of Rheumatology, Istanbul
Faculty of Medicine, Istanbul University, Istanbul, Turkey
e-mail: agul@istanbul.edu.tr

This λ_s value was considered as strongly supporting the contribution of genetics to the multifactorial pathogenesis of BD.

Familial clustering was more frequently observed among juvenile-onset (<16) BD patients [18, 19]. Molinary and colleagues conducted a segregation analysis using the pedigree data of 106 BD cases. They included “possible” BD patients who had only two of the classical disease manifestations into the analysis, and they found a pattern compatible with autosomal recessive inheritance in pediatric BD subgroup, and no Mendelian pattern in adult-onset patients [19]. This study suggested a genetic heterogeneity with a higher impact of genetic load in juvenile BD cases [19].

Frequency of HLA-B51 was found to be higher in familial patients [11, 12]. However, presence of unaffected siblings with risk alleles also showed the complex nature of the disease indicating the contribution of other genes and/or environmental factors [11, 20]. A comparison of related pairs of patients according to their age at onset also supports involvement of both genetic and environmental factors in the pathogenesis [21, 22].

Another study from Turkey documented clues for genetic anticipation in the form of earlier disease onset in the second generation compared with their affected parents in 15 out of 18 familial cases studied [23]. However, no trinucleotide repeat expansion data are yet available to further support this observation.

No large series of twins concordant or discordant for BD were reported so far [24–26]. Therefore, large series of monozygotic and dizygotic twins are being awaited for heritability analysis to assess the relative contribution of genes and environment to the pathogenesis of BD.

Geographic Distribution

Epidemiology of BD has a distinct feature in terms of its geographic distribution. Prevalence of BD is much higher in an area extending from the Mediterranean basin to Japan, between 30° and 45° latitudes North, which overlaps with the ancient Silk Road [27]. There is no known specific environmental factor common along this route, but shared genetic factors may explain the clustering of BD cases. The frequency of BD-related HLA-B51 allele is higher in the healthy population living along this region, and distribution of HLA-B51 allele is suggested to play a role in the disease clustering [27, 28].

HLA-B51 and Other MHC Associations

BD is strongly associated with a class I major histocompatibility complex (MHC) allele, HLA-B51. This association was first reported in Japanese BD patients [28–30]. Association of HLA-B51 with BD was later confirmed in other ethnic groups, including those in which BD is seen very rarely [1, 2, 16, 27, 31–34].

No disease specific differences were observed in the sequence of HLA-B51 alleles between BD patients and healthy controls, neither in the coding region nor in the regulatory sequences [35, 36]. HLA-B51 is a split antigen of HLA-B5, and the other split antigen HLA-B52 has not been associated with BD despite some exceptional reports [37, 38]. HLA-B51 differs from HLA-B52 only by two aminoacids in the $\alpha 1$ helix. Asparagine and phenylalanine at positions 63 and 67 of the HLA-B51 molecule are replaced with glutamic acid and serine in the HLA-B52 at the same positions [39]. These two aminoacids are located at the B pocket of the antigen binding groove (Fig. 15.1). HLA-B51 allele can bind peptides with eight or nine aminoacids and a hydrophobic C-terminus [40]. Later studies suggested that B pocket can be occupied by small aminoacids alanine and proline, and changes in the B pocket can affect the motif of the peptides that can bind to HLA molecule [41]. Isoleucine and valine were identified as dominant anchor residues in the C-terminus of the refined peptide motif which binds to relatively small F pocket, and aminoacids making the F pocket are conserved in all HLA-B51 alleles [41].

HLA-B51 allele has 73 different subtypes (HLA-B*5101–B*5173), and they all share the same aminoacid sequence at the B pocket of the antigen binding groove except for B*5107 and B*5122. HLA-B*5101 is the dominant subtype of the B51 molecule, and molecular HLA-B51 typing in different ethnic groups suggests that HLA-B51 subtypes in BD patients are not different from those in healthy controls, with HLA-B*5101 and -B*5108 as the main subtypes [42–46].

Molecular typing of HLA-B51 molecules suggests that presentation of certain BD-associated peptides with its specific B and F pocket features might be one of

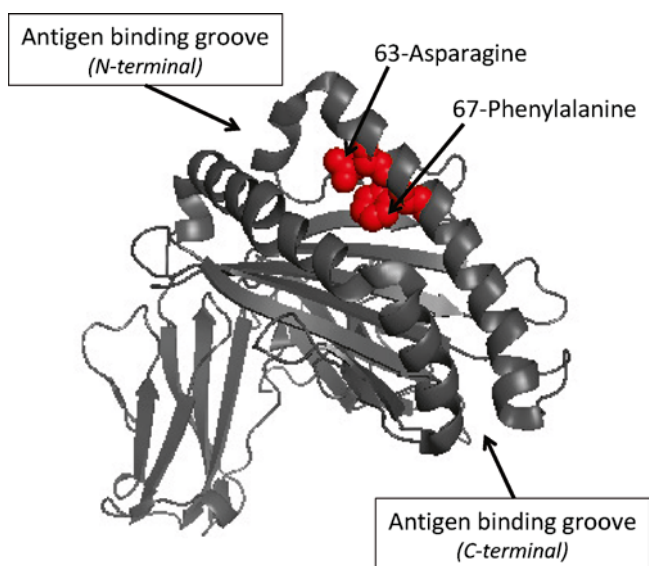


Fig. 15.1 A model of HLA-B51 molecule (1E28) showing the critical asparagine and phenylalanine at positions 63 and 67 in its antigen binding groove (drawn by PyMOL)

the pathogenic mechanisms behind the susceptibility to BD. So far, only major histocompatibility complex class I chain-related gene A (MICA)-derived nonamer peptide (AAAAAIFVI) was shown to induce T cells in less than one-third of active HLA-B51 positive BD patients compared to none of the healthy controls [47].

HLA-B51, as a class I molecule, also interacts with a group of receptors expressed on natural killer (NK) cells, CD8+ and $\gamma\delta$ T cells [48]. The killer immunoglobulin-like receptors (KIR), bind to conserved Bw4 epitopes at residues 77–83 of the α 1-helix, which are shared by different allelic groups of HLA class I molecules. Engagement of these receptors can result in selective inhibition of NK or T cell mediated cytotoxicity. A relative predispositional effects analysis, conducted to search for weaker HLA-B associations with BD masked by strong HLA-B51 association, revealed a weak association of HLA-B*2702 with BD, which shares the same Bw4 motif with HLA-B51 [49]. Investigation of HLA-B51 interacting KIR3DL1/DS1 polymorphism documented the association of DL1/DL1 genotype with BD in Bw4-motif positive patients [50]. These preliminary studies support an alternative hypothesis that the pathogenic role of HLA-B51 may also include its interaction with KIR3DL1 molecules expressed on inflammatory cells.

HLA-B51-derived peptides can be presented by HLA class II molecules. HLA class I heavy chain misfolding as well as enhanced expression due to up-regulated immune response increase the possibility of class I-derived peptide presentation. Wildner and Thurau identified a polymorphic HLA-B sequence common in HLA-B27, -B51, and several other HLA-B alleles (B27PD), which shares aminoacid homologies with retinal soluble antigen (S-Ag)-derived peptide [51]. Kurhan-Yavuz and colleagues demonstrated increased T cell response against retinal S-Ag, retinal S-Ag derived peptide, and B27PD peptide in BD patients with posterior uveitis compared with those BD patients without eye disease or patients with non-BD anterior uveitis [52].

HLA-B51 is one of the slow folding MHC molecules [53]. However, there is no data showing the role of HLA-B51 folding problems and unfolded protein response in BD pathogenesis similar to the observations on HLA-B27 in ankylosing spondylitis animal models [54].

There is only one HLA-B*5101 heavy chain transgenic mouse model developed so far in investigating the direct role of HLA-B51 molecules in BD [55]. No manifestation typical for BD was observed in these transgenic animals. HLA-B51 transgenic animals showed an increased neutrophil activity following f-Met-Leu-Phe (fMLP) stimulation compared to HLA-B35 and nontransgenic mice [55]. A similar enhanced neutrophil activity was reported in HLA-B51 positive healthy individuals [11, 55, 56]. Extrapolating from the experience with HLA-B27 animal models, it is still needed to have a high heavy chain copy number transgenic animal models with and without human β 2-microglobulin in different strains of mice and rats to explore the role of HLA-B51 in BD [57].

In addition to association studies, analysis of 12 multicase families confirmed the genetic linkage of the HLA-B locus to BD by using the transmission disequilibrium test [58]. Contribution of the HLA-B locus to the overall genetic susceptibility

to BD was estimated to be 19% assuming multiplicative interaction between disease susceptibility loci [58]. This result supports the need for studies to look for other susceptibility loci.

Other MHC Associations

Linkage disequilibrium (LD) is high in the MHC, especially in the class I region with larger haplotype blocks [59]. It has long been discussed whether HLA-B51 has a direct role in the BD pathogenesis, or whether this strong association reflects LD with one or more susceptibility genes located close to the HLA-B locus (Fig. 15.2). The tumor necrosis factor (TNF) and lymphotoxin genes, which are located centromeric to HLA-B, were investigated first as possible candidate susceptibility genes. The analysis of the genomic segment between the TNF and HLA-B loci revealed a strong association of MICA gene with BD, which is located 46-kb centromeric to HLA-B [60]. The MICA gene *009 allele and its transmembrane region microsatellite polymorphism A6 allele were found to be significantly increased in BD patients [60–62]. Fine mapping of the region in different ethnic groups revealed HLA-B as the gene providing strongest association with BD, and all other associations including the MICA were resulting from strong LD with HLA-B51 [63]. However, it is still hard to rule out individual contribution of the MICA gene on an HLA-B51 haplotype to the BD susceptibility through its interaction with NK and $\gamma\delta$ T cells.

Within the MHC region, no association with class II antigens was observed [64], but HLA-B51-associated LD extends to telomeric part of class I region. Weaker associations with HLA-Cw14, Cw15, and C*16 alleles [65, 66] and a negative association with nonclassical HLA-E*0101 and HLA-G*010101 alleles [67] were reported. Recent studies suggest a second HLA class I region association independent of HLA-B51 [68]. Meguro and colleagues reported the association of HLA-A26 allele and HLA-A*26-F*010101-G*010102 haplotype with BD even in HLA-B51 negative patients in Japan [68]. Association of HLA-A26 allele with BD was also observed in Taiwanese and Greek patients. These observations suggest that contribution of the MHC region to the BD susceptibility includes both HLA-B51 and other classical or nonclassical HLA associations with possible different pathogenic mechanisms.

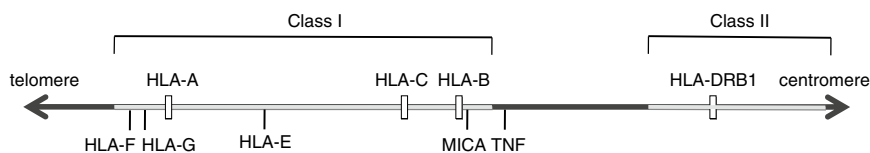


Fig. 15.2 Genetic map of the major histocompatibility complex (MHC) in short arm of chromosome 6 showing the Behçet's disease-associated loci in class I region

Non-HLA Genes and Behçet's Disease

As a complex disease, non-HLA genetic polymorphisms can also contribute to the BD susceptibility. For investigation of these susceptibility genes, a candidate gene approach was frequently preferred by investigators despite no clear evidence for

Table 15.1 List of non-HLA genes reported to be associated with Behçet's disease

Name	Ethnic groups
<i>Cytokines, chemokines and their receptors</i>	
Tumor necrosis factor (TNF)	British Caucasian, Turkish, Korean, Tunisian [69–72]
Interleukin 1 gene cluster	Turkish [73]
Interleukin 6 (IL6)	Korean [74]
Interleukin 8 (IL8/CXCL8)	Korean [75]
Interleukin 10 (IL10)	British Caucasian [76]
Interleukin 12 p40 (IL12B)	Japanese [77]
Interleukin-17F (IL17F)	Korean [78]
Interleukin 18 (IL18)	Korean [79]
Chemokine receptor 5 (CCR5)	Iranian [80]
<i>Other immune response genes</i>	
Natural resistance associated macrophage protein 1 (NRAMP1)	Turkish, Korean [81, 82]
CTLA4	Tunisian, Korean [83, 84]
Mannose binding lectin	Japanese, Korean [85, 86]
Ficolin 2 (FCN2)	Japanese [87]
Small ubiquitin-like modifier 4 (SUMO4)	Chinese [88]
Fc receptor-like 3 gene (FCRL3)	Chinese [89]
CD94/NKG2A	Korean [90]
PTPN22 (negative association)	British Caucasian, Middle Eastern [91]
CD28	Turkish [92]
<i>Pathogen associated molecular pattern receptors</i>	
Toll-like receptor 4 (TLR4)	Japanese, Korean [93, 94]
<i>Autoinflammatory polymorphisms</i>	
Pyrin (MEFV)	Turkish, European, Jewish, Palestinian [95–99]
TNF receptor p55 (TNFRSF1A)	European [100]
<i>Vascular and procoagulant polymorphisms</i>	
Intercellular adhesion molecule-1 (ICAM1)	Palestinian/Jordanian, Italian, Korean, Lebanese [101–104]
Endothelial nitric oxide synthase (eNOS)	Italian, Korean, Turkish, Tunisian [105–108]
Factor V Leiden	Turkish, Arabian [109–111]
Prothrombin	Turkish, Spanish [112, 113]
Manganese superoxide dismutase (SOD)	Japanese [114]
<i>Others</i>	
Cytochrome P450	Turkish, Taiwanese [115, 116]
N-acetyltransferase 2	Turkish [117]

utilizing this method in deciphering the pathogenic mechanisms of BD. Most of these association studies were carried out using small numbers of cases and controls with limited power. The list of non-HLA genes reported to be associated with BD are given in Table 15.1 [69–117]. Among the reported associations, only a few were replicated in different ethnic groups, including polymorphisms in the TNF, MEFV, ICAM1, and eNOS genes. None of these polymorphisms are disease specific, and they are considered to be contributing to a disease-specific inflammatory reaction.

Another approach for investigating complex disease susceptibility genes is screening of whole genome without a priori hypothesis about disease pathogenesis. A genomewide linkage screen using 193 individuals from 28 multicase BD families of Turkish origin with 83 affecteds revealed evidence for linkage to 15 non-HLA chromosomal regions: 1p36, 4p15, 5q12, 5q23, 6q16, 6q25–26, 7p21, 10q24, 12p12–13, 12q13, 16q12, 16q21–23, 17p13, 20q12–13, and Xq26–28 [118]. The linkage peak in the short arm of chromosome 6 (the maximum nonparametric linkage score 3.7) confirmed the strong association of HLA-B locus and also suggested another telomeric susceptibility loci [118, 119]. After the addition of further markers, high maximum nonparametric linkage scores were observed at chromosome 12p12-13 (3.94) and 6q25-26 (3.14).

Linkage studies in families are expected to identify rare, but penetrant genetic variations. However, genomewide association studies (GWAS) in large number of cases and controls can reveal common, but less penetrant polymorphisms affecting the disease susceptibility. A recent GWAS investigated 300 Japanese BD patients and 300 healthy controls with 23,465 microsatellite markers. This study identified six possible genomic regions, including two from the MHC region, one corresponding to HLA-B and the other to HLA-A [68]. Other non-HLA microsatellite markers suggested chromosomal regions 3p12 (D3S0186i), 6q25.1 (536G12Aa), 12p12.1 (D12S0645i), and 22q11.22 (D22S0104im) as possible genomic segments harboring disease susceptibility loci, two of which overlap with the findings of the previous linkage study [68]. Current GWAS approach enables us to analyze thousands of samples using chips for >300,000 single nucleotide polymorphisms in a relatively short time. Results of GWAS from different ethnic groups are eagerly being awaited to clarify the genetics of BD further.

References

1. Gül A (2001) Behçet's disease: an update on the pathogenesis. *Clin Exp Rheumatol* 19(Suppl 24):S6–S12
2. Zierhut M, Mizuki N, Ohno S et al (2003) Immunology and functional genomics of Behçet's disease. *Cell Mol Life Sci* 60:1903–1922
3. Fowler TJ, Humpston DJ, Nussey AM, Small M (1968) Behçet's syndrome with neurological manifestations in two sisters. *Br Med J* 2:473–474
4. Mason RM, Barnes CG (1969) Behçet's syndrome with arthritis. *Ann Rheum Dis* 28:95–103
5. Fadli ME, Youssef MM (1973) Neuro-Behçet's syndrome in the United Arab Republic. *Eur Neurol* 9:76–89

6. Chajek T, Fainaru M (1975) Behçet's disease: report of 41 cases and a review of the literature. *Medicine (Baltimore)* 54:179–196
7. Goolamali SK, Comaish JS, Hassanyeh F (1976) Familial Behçet's syndrome. *Br J Dermatol* 95:637–642
8. Nahir M, Scharf Y, Gidoni O et al (1978) HL-A antigens in Behçet's disease. A family study. *Dermatologica* 156:205–208
9. Abdel-Aziz AH, Fairburn EA (1978) Familial Behçet's syndrome. *Cutis* 21:649–652
10. Dündar SV, Gencalp U, Simsek H (1985) Familial cases of Behçet's disease. *Br J Dermatol* 113:319–321
11. Chajek-Shaul T, Pisanty S, Knobler H et al (1987) HLA-B51 may serve as an immunogenetic marker for a subgroup of patients with Behçet's syndrome. *Am J Med* 83:666–672
12. Akpolat T, Koc Y, Yeniay I et al (1992) Familial Behçet's disease. *Eur J Med* 1:391–395
13. Villanueva JL, Gonzalez-Dominguez J, Gonzalez-Fernandez R et al (1993) HLA antigen familial study in complete Behçet's syndrome affecting three sisters. *Ann Rheum Dis* 52:155–157
14. Nishiura K, Kotake S, Ichiishi A, Matsuda H (1996) Familial occurrence of Behçet's disease. *Jpn J Ophthalmol* 40:255–259
15. Nishiyama M, Nakae K, Umehara T (2001) A study of familial occurrence of Behçet's disease with and without ocular lesions. *Jpn J Ophthalmol* 45:313–316
16. Fietta P (2005) Behçet's disease: familial clustering and immunogenetics. *Clin Exp Rheumatol* 23(Suppl 38):S96–S105
17. Gül A, Inanc M, Ocal L et al (2000) Familial aggregation of Behçet's disease in Turkey. *Ann Rheum Dis* 59:622–625
18. Treudler R, Orfanos CE, Zouboulis CC (1999) Twenty-eight cases of juvenile-onset Adamantiades-Behçet's disease in Germany. *Dermatology* 199:15–19
19. Koné-Paut I, Geisler I, Wechsler B et al (1999) Familial aggregation in Behçet's disease: high frequency in siblings and parents of pediatric probands. *J Pediatr* 135:89–93
20. Hayasaka S, Kurome H, Noda S (1994) HLA antigens in a Japanese family with Behçet's disease. *Graefes Arch Clin Exp Ophthalmol* 232:589–590
21. Nishiyama M, Nakae K, Kuriyama T et al (2002) A study among related pairs of Japanese patients with familial Behçet's disease: group comparisons by interval of disease onsets. *J Rheumatol* 29:743–747
22. Aronsson A, Tegner E (1983) Behçet's syndrome in two brothers. *Acta Derm Venereol* 63:73–74
23. Fresko I, Soy M, Hamuryudan V et al (1998) Genetic anticipation in Behçet's syndrome. *Ann Rheum Dis* 57:45–48
24. Hamuryudan V, Yurdakul S, Ozbakir F et al (1991) Monozygotic twins concordant for Behçet's syndrome. *Arthritis Rheum* 34:1071–1072
25. Gül A, Inanc M, Ocal L et al (1997) HLA-B51 negative monozygotic twins discordant for Behçet's disease. *Br J Rheumatol* 36:922–923
26. Kobayashi T, Sudo Y, Okamura S et al (2005) Monozygotic twins concordant for intestinal Behçet's disease. *J Gastroenterol* 40:421–425
27. Verity DH, Marr JE, Ohno S et al (1999) Behçet's disease, the Silk Road and HLA-B51: historical and geographical perspectives. *Tissue Antigens* 54:213–220
28. Ohno S, Ohguchi M, Hirose S et al (1982) Close association of HLA-Bw51 with Behçet's disease. *Arch Ophthalmol* 100:1455–1458
29. Ono S, Aoki K, Sugiura S et al (1973) HL-A5 and Behçet's disease. *Lancet* 2:1383–1384
30. Ono S, Nakayama E, Sugiura S et al (1975) Specific histocompatibility antigens associated with Behçet's disease. *Am J Ophthalmol* 80:636–641
31. Kilmartin DJ, Finch A, Acheson RW (1997) Primary association of HLA-B51 with Behçet's disease in Ireland. *Br J Ophthalmol* 81:649–653
32. Ambresin A, Tran T, Spertini F, Herbort C (2002) Behçet's disease in Western Switzerland: epidemiology and analysis of ocular involvement. *Ocul Immunol Inflamm* 10:53–63
33. Pipitone N, Boiardi L, Olivieri I et al (2004) Clinical manifestations of Behçet's disease in 137 Italian patients: results of a multicenter study. *Clin Exp Rheumatol* 22(Suppl 36):S46–S51

34. Bettencourt A, Pereira C, Carvalho L et al (2008) New insights of HLA class I association to Behçet's disease in Portuguese patients. *Tissue Antigens* 72:379–382
35. Sano K, Yabuki K, Imagawa Y et al (2001) The absence of disease-specific polymorphisms within the HLA-B51 gene that is the susceptible locus for Behçet's disease. *Tissue Antigens* 58:77–82
36. Takemoto Y, Naruse T, Namba K et al (2008) Re-evaluation of heterogeneity in HLA-B*510101 associated with Behçet's disease. *Tissue Antigens* 72:347–353
37. Arber N, Klein T, Meiner Z et al (1991) Close association of HLA-B51 and B52 in Israeli patients with Behçet's syndrome. *Ann Rheum Dis* 50:351–353
38. Sugisaki K, Saito R, Takagi T et al (2005) HLA-B52-positive vasculo-Behçet's disease: usefulness of magnetic resonance angiography, ultrasound study, and computed tomographic angiography for the early evaluation of multiarterial lesions. *Mod Rheumatol* 15:56–61
39. Falk K, Röttschke O, Takiguchi M et al (1995) Peptide motifs of HLA-B51, -B52 and -B78 molecules, and implications for Behçet's disease. *Int Immunol* 7:223–228
40. Sakaguchi T, Ibe M, Miwa K et al (1997) Predominant role of N-terminal residue of nonamer peptides in their binding to HLA-B* 5101 molecules. *Immunogenetics* 46:245–248
41. Lemmel C, Rammensee H-G, Stevanovic S (2003) Peptide motif of HLA-B*5101 and the linkage to Behçet's disease. In: Zierhut M, Ohno S (eds) *Immunology of Behçet's disease*. Swets & Zeitlinger, Lisse, pp 127–137
42. Mizuki N, Inoko H, Ando H et al (1993) Behçet's disease associated with one of the HLA-B51 subantigens, HLA-B* 5101. *Am J Ophthalmol* 116:406–409
43. Mizuki N, Ota M, Katsuyama Y et al (2002) Sequencing-based typing of HLA-B*51 alleles and the significant association of HLA-B*5101 and -B*5108 with Behçet's disease in Greek patients. *Tissue Antigens* 59:118–121
44. Pirim I, Atasoy M, Ikbal M et al (2004) HLA class I and class II genotyping in patients with Behçet's disease: a regional study of eastern part of Turkey. *Tissue Antigens* 64:293–297
45. Kera J, Mizuki N, Ota M et al (1999) Significant associations of HLA-B*5101 and B*5108, and lack of association of class II alleles with Behçet's disease in Italian patients. *Tissue Antigens* 54:565–571
46. Yabuki K, Ohno S, Mizuki N et al (1999) HLA class I and II typing of the patients with Behçet's disease in Saudi Arabia. *Tissue Antigens* 54:273–277
47. Yasuoka H, Okazaki Y, Kawakami Y et al (2004) Autoreactive CD8+ cytotoxic T lymphocytes to major histocompatibility complex class I chain-related gene A in patients with Behçet's disease. *Arthritis Rheum* 50:3658–3662
48. Martin MP, Gao X, Lee J-H et al (2002) Epistatic interaction between *KIR3DS1* and *HLA-B* delays the progression to AIDS. *Nat Genet* 31:429–434
49. Gül A, Uyar FA, Inanç M et al (2002) A weak association of HLA-B*2702 with Behçet's disease. *Genes Immun* 3:368–372
50. Duymaz-Tozkir J, Uyar A, Norman PJ et al (2008) Distribution of killer immunoglobulin-like receptor 3DL1/3DS1 alleles in Behçet's disease. *Arthritis Rheum* 58(Suppl):S855
51. Wildner G, Thureau SR (1994) Cross-reactivity between an HLA-B27-derived peptide and a retinal autoantigen peptide: a clue to major histocompatibility complex association with autoimmune disease. *Eur J Immunol* 24:2579–2585
52. Kurhan-Yavuz S, Direskeneli H, Bozkurt N et al (2000) Anti-MHC autoimmunity in Behçet's disease: T cell responses to an HLA-B-derived peptide cross-reactive with retinal-S antigen in patients with uveitis. *Clin Exp Immunol* 120:162–166
53. Hill A, Takiguchi M, McMichael A (1993) Different rates of HLA class I molecule assembly which are determined by amino acid sequence in the alpha 2 domain. *Immunogenetics* 37:95–101
54. Turner MJ, Sowders DP, DeLay ML et al (2005) HLA-B27 misfolding in transgenic rats is associated with activation of the unfolded protein response. *J Immunol* 175:2438–2448
55. Takeno M, Kariyone A, Yamashita N et al (1995) Excessive function of peripheral blood neutrophils from patients with Behçet's disease and from HLA-B51 transgenic mice. *Arthritis Rheum* 38:426–433

56. Sensi A, Gavioli R, Spisani S et al (1991) HLA B51 antigen associated with neutrophil hyper-reactivity. *Dis Markers* 9:327–331
57. Taurog JD, Maika SD, Satumira N et al (1999) Inflammatory disease in HLA-B27 transgenic rats. *Immunol Rev* 169:209–223
58. Gül A, Hajeer AH, Worthington J et al (2001) Evidence for linkage of the HLA-B locus in Behçet's disease, obtained using the transmission disequilibrium test. *Arthritis Rheum* 44(1):239–240
59. Miretti MM, Walsh EC, Ke X et al (2005) A high-resolution linkage-disequilibrium map of the human major histocompatibility complex and first generation of tag single-nucleotide polymorphisms. *Am J Hum Genet* 76:634–646
60. Mizuki N, Ota M, Kimura M et al (1997) Triplet repeat polymorphism in the transmembrane region of the MICA gene: a strong association of six GCT repetitions with Behçet's disease. *Proc Natl Acad Sci U S A* 94:1298–1303
61. Hughes EH, Collins RW, Kondeatis E et al (2005) Associations of major histocompatibility complex class I chain-related molecule polymorphisms with Behçet's disease in Caucasian patients. *Tissue Antigens* 66:195–199
62. Mizuki N, Meguro A, Tohnai I et al (2007) Association of major histocompatibility complex Class I chain-related Gene A and HLA-B alleles with Behçet's disease in Turkey. *Jpn J Ophthalmol* 51:431–436
63. Mizuki N, Ota M, Yabuki K et al (2000) Localization of the pathogenic gene of Behçet's disease by microsatellite analysis of three different populations. *Invest Ophthalmol Vis Sci* 41:3702–3708
64. Mizuki N, Ohno S, Tanaka H et al (1992) Association of HLA-B51 and lack of association of class II alleles with Behçet's disease. *Tissue Antigens* 40:22–30
65. Mizuki N, Ohno S, Ando H et al (1996) HLA-C genotyping of patient with Behçet's disease in the Japanese population. *Hum Immunol* 50:47–53
66. Sanz L, González-Escribano F, de Pablo R et al (1998) HLA-Cw*1602: a new susceptibility marker of Behçet's disease in southern Spain. *Tissue Antigens* 51:111–114
67. Park KS, Park JS, Nam JH et al (2007) HLA-E*0101 and HLA-G*010101 reduce the risk of Behçet's disease. *Tissue Antigens* 69:139–144
68. Meguro A, Inoko H, Ota M, et al (2009) Genetics of Behçet's disease inside and outside the MHC. *Ann Rheum Dis* 69:747–754
69. Ahmad T, Wallace GR, James T et al (2003) Mapping the HLA association in Behçet's disease: a role for tumor necrosis factor polymorphisms? *Arthritis Rheum* 48:807–813
70. Akman A, Sallakci N, Coskun M et al (2006) TNF-alpha gene 1031 T/C polymorphism in Turkish patients with Behçet's disease. *Br J Dermatol* 155:350–356
71. Park K, Kim N, Nam J et al (2006) Association of TNFA promoter region haplotype in Behçet's disease. *J Korean Med Sci* 21:596–601
72. Kamoun M, Chelbi H, Houman MH et al (2007) Tumor necrosis factor gene polymorphisms in Tunisian patients with Behçet's disease. *Hum Immunol* 68:201–205
73. Karasneh J, Hajeer AH, Barrett J et al (2003) Association of specific interleukin 1 gene cluster polymorphisms with increased susceptibility for Behçet's disease. *Rheumatology (Oxford)* 42:860–864
74. Chang HK, Jang WC, Park SB et al (2005) Association between interleukin 6 gene polymorphisms and Behçet's disease in Korean people. *Ann Rheum Dis* 64:339–340
75. Lee EB, Kim JY, Zhao J et al (2007) Haplotype association of IL-8 gene with Behçet's disease. *Tissue Antigens* 69:128–132
76. Wallace GR, Kondeatis E, Vaughan RW et al (2007) IL-10 genotype analysis in patients with Behçet's disease. *Hum Immunol* 68:122–127
77. Yanagihori H, Oyama N, Nakamura K et al (2006) Role of IL-12B promoter polymorphism in Adamantiades-Behçet's disease susceptibility: an involvement of Th1 immunoreactivity against *Streptococcus Sanguinis* antigen. *J Invest Dermatol* 126:1534–1540
78. Jang WC, Nam YH, Ahn YC et al (2008) Interleukin-17F gene polymorphisms in Korean patients with Behçet's disease. *Rheumatol Int* 29:173–178

79. Lee YJ, Kang SW, Park JJ et al (2006) Interleukin-18 promoter polymorphisms in patients with Behçet's disease. *Hum Immunol* 67:812–818
80. Mojtaheidi Z, Ahmadi SB, Razmkhah M et al (2006) Association of chemokine receptor 5 (CCR5) delta32 mutation with Behçet's disease is dependent on gender in Iranian patients. *Clin Exp Rheumatol* 24(Suppl 42):S91–S94
81. Ateş O, Dalyan L, Hatemi G et al (2009) Genetic susceptibility to Behçet's syndrome is associated with NRAMP1 (SLC11A1) polymorphism in Turkish patients. *Rheumatol Int* 29:787–791
82. Kim SK, Jang WC, Park SB et al (2006) SLC11A1 gene polymorphisms in Korean patients with Behçet's disease. *Scand J Rheumatol* 35:398–401
83. Ben Dhifallah I, Chelbi H, Braham A et al (2009) CTLA-4 +49A/G polymorphism is associated with Behçet's disease in a Tunisian population. *Tissue Antigens* 73(3):213–217
84. Park KS, Baek JA, Do JE et al (2009) CTLA4 gene polymorphisms and soluble CTLA4 protein in Behçet's disease. *Tissue Antigens* 74:222–227
85. Wang H, Nakamura K, Inoue T et al (2004) Mannose-binding lectin polymorphisms in patients with Behçet's disease. *J Dermatol Sci* 36:115–117
86. Park KS, Min K, Nam JH et al (2005) Association of HYPA haplotype in the mannose-binding lectin gene-2 with Behçet's disease. *Tissue Antigens* 65:260–265
87. Chen X, Katoh Y, Nakamura K et al (2006) Single nucleotide polymorphisms of Ficolin 2 gene in Behçet's disease. *J Dermatol Sci* 43:201–205
88. Hou S, Yang P, Du L et al (2008) SUMO4 gene polymorphisms in Chinese Han patients with Behçet's disease. *Clin Immunol* 129:170–175
89. Li K, Zhao M, Hou S, Du L et al (2008) Association between polymorphisms of FCRL3, a non-HLA gene, and Behçet's disease in a Chinese population with ophthalmic manifestations. *Mol Vis* 14:2136–2142
90. Seo J, Park JS, Nam JH et al (2007) Association of CD94/NKG2A, CD94/NKG2C, and its ligand HLA-E polymorphisms with Behçet's disease. *Tissue Antigens* 70:307–313
91. Baranathan V, Stanford MR, Vaughan RW et al (2007) The association of the PTPN22 620W polymorphism with Behçet's disease. *Ann Rheum Dis* 66:1531–1533
92. Gunesacar R, Erken E, Bozkurt B et al (2007) Analysis of CD28 and CTLA-4 gene polymorphisms in Turkish patients with Behçet's disease. *Int J Immunogenet* 34:45–49
93. Meguro A, Ota M, Katsuyama Y et al (2008) Association of the toll-like receptor 4 gene polymorphisms with Behçet's disease. *Ann Rheum Dis* 67:725–727
94. Horie Y, Meguro A, Ota M et al (2009) Association of TLR4 polymorphisms with Behçet's disease in a Korean population. *Rheumatology (Oxford)* 48:638–642
95. Touitou I, Magne X, Molinari N et al (2000) MEFV mutations in Behçet's disease. *Hum Mutat* 16:271–272
96. Atagunduz P, Ergun T, Direskeneli H (2003) MEFV mutations are increased in Behçet's disease (BD) and are associated with vascular involvement. *Clin Exp Rheumatol* 21(Suppl 30):S35–S37
97. Imirzalioglu N, Dursun A, Tastan B et al (2005) MEFV gene is a probable susceptibility gene for Behçet's disease. *Scand J Rheumatol* 34:56–58
98. Rabinovich E, Shinar Y, Leiba M et al (2007) Common FMF alleles may predispose to development of Behçet's disease with increased risk for venous thrombosis. *Scand J Rheumatol* 36:48–52
99. Ayesh S, Abu-Rmaileh H, Nassar S et al (2008) Molecular analysis of MEFV gene mutations among Palestinian patients with Behçet's disease. *Scand J Rheumatol* 37:370–374
100. Amoura Z, Dodé C, Hue S et al (2005) Association of the R92Q TNFRSF1A mutation and extracranial deep vein thrombosis in patients with Behçet's disease. *Arthritis Rheum* 52:608–611
101. Verity DH, Vaughan RW, Kondeatis E et al (2000) Intercellular adhesion molecule-1 gene polymorphisms in Behçet's disease. *Eur J Immunogenet* 27:73–76
102. Boiardi L, Salvarani C, Casali B et al (2001) Intercellular adhesion molecule-1 gene polymorphisms in Behçet's disease. *J Rheumatol* 28:1283–1287
103. Kim EH, Mok JW, Bang DS et al (2003) Intercellular adhesion molecule-1 polymorphisms in Korean patients with Behçet's disease. *J Korean Med Sci* 18:415–418

104. Chmaisse HN, Fakhoury HA, Salti NN, Makki RF (2006) The ICAM-1 469 T/C gene polymorphism but not 241 G/A is associated with Behçet's disease in the Lebanese population. *Saudi Med J* 27:604–607
105. Salvarani C, Boiardi L, Casali B et al (2002) Endothelial nitric oxide synthase gene polymorphisms in Behçet's disease. *J Rheumatol* 29:535–540
106. Kim JU, Chang HK, Lee SS et al (2003) Endothelial nitric oxide synthase gene polymorphisms in Behçet's disease and rheumatic diseases with vasculitis. *Ann Rheum Dis* 62:1083–1087
107. Karasneh JA, Hajeer AH, Silman A et al (2005) Polymorphisms in the endothelial nitric oxide synthase gene are associated with Behçet's disease. *Rheumatology (Oxford)* 44:614–617
108. Ben Dhifallah I, Houman H, Khanfir M, Hamzaoui K (2008) Endothelial nitric oxide synthase gene polymorphism is associated with Behçet's disease in Tunisian population. *Hum Immunol* 69:661–665
109. Gül A, Ozbek U, Oztürk C et al (1996) Coagulation factor V gene mutation increases the risk of venous thrombosis in Behçet's disease. *Br J Rheumatol* 35:1178–1180
110. Verity DH, Vaughan RW, Madanat W et al (1999) Factor V Leiden mutation is associated with ocular involvement in Behçet's disease. *Am J Ophthalmol* 128(3):352–356
111. Mammo L, Al-Dalaan A, Bahabri SS, Saour JN (1997) Association of factor V Leiden with Behçet's disease. *J Rheumatol* 24:2196–2198
112. Gül A, Aslantas AB, Tekinay T et al (1999) Procoagulant mutations and venous thrombosis in Behçet's disease. *Rheumatology (Oxford)* 38:1298–1299
113. Ricart JM, Vayá A, Todolí J et al (2006) Thrombophilic risk factors and homocysteine levels in Behçet's disease in eastern Spain and their association with thrombotic events. *Thromb Haemost* 95(4):618–624
114. Nakao K, Isashiki Y, Sonoda S et al (2007) Nitric oxide synthase and superoxide dismutase gene polymorphisms in Behçet's disease. *Arch Ophthalmol* 125:246–251
115. Tursen U, Tamer L, Api H et al (2007) Cytochrome P450 polymorphisms in patients with Behçet's disease. *Int J Dermatol* 46:153–156
116. Yen JH, Tsai WC, Lin CH et al (2004) Cytochrome P450 1A1 and manganese superoxide dismutase gene polymorphisms in Behçet's disease. *J Rheumatol* 31:736–740
117. Tamer L, Tursen U, Eskandari G et al (2005) N-acetyltransferase 2 polymorphisms in patients with Behçet's disease. *Clin Exp Dermatol* 30:56–60
118. Karasneh J, Gül A, Ollier WE et al (2005) Whole-genome screening for susceptibility genes in multicase families with Behçet's disease. *Arthritis Rheum* 52:1836–1842
119. Gül A, Hajeer AH, Worthington J et al (2001) Linkage mapping of a novel susceptibility locus for Behçet's disease to chromosome 6p22–23. *Arthritis Rheum* 44:2693–2696