Targeting TNF for Treatment of Cancer and Autoimmunity

Gautam Sethi, Bokyung Sung, Ajaikumar B. Kunnumakkara and Bharat B. Aggarwal*

Abstract

Important necrosis factor- α (TNF- α) was first isolated two decades ago as a macrophage-produced protein that can effectively kill tumor cells. TNF- α is also an essential component of the immune system and is required for hematopoiesis, for protection from bacterial infection and for immune cell-mediated cytotoxicity. Extensive research, however, has revealed that TNF- α is one of the major players in tumor initiation, proliferation, invasion, angiogenesis and metastasis. The proinflammatory activities link TNF- α with a wide variety of autoimmune diseases, including psoriasis, inflammatory bowel disease, rheumatoid arthritis, systemic sclerosis, systemic lupus erythematosus, multiple sclerosis, diabetes and ankylosing spondylitis. Systemic inhibitors of TNF such as etanercept (Enbrel) (a soluble TNF receptor) and infliximab (Remicade) and adalimumab (Humira) (anti-TNF antibodies) have been approved for the treatment inflammatory bowel disease, psoriasis and rheumatoid arthritis. These drugs, however, exhibit severe side effects and are expensive. Hence orally active blockers of TNF- α that are safe, efficacious and inexpensive are urgently needed. Numerous products from fruits, vegetable and traditional medicinal plants have been described which can suppress TNF expression and TNF signaling but their clinical potential is yet uncertain.

Discovery of TNF

Tumor necrosis factor (TNF), an activity in the serum of endotoxin-injected animals, was first identified in 1944, rediscovered in the mid-1970s and chemically isolated from macrophage-conditioned medium as a cytokine that kills tumor cells in culture in 1984. ^{1,2} Two distinct factors were identified in macrophages and lymphocytes: TNF- α and TNF- β , respectively. The identification of their primary amino acid sequences led to the cloning of their genes and the availability of large amounts of pure cytokines for preclinical and clinical evaluation. Intravenous administration of TNF to cancer patients produced numerous toxic reactions including fever. ³ In animal studies, TNF- α has been shown to mediate endotoxin-mediated septic shock. ⁴ Several reports over the past years have indicated that dysregulation of TNF- α synthesis mediates a wide variety of autoimmune diseases and cancer. ²

Signaling Mechanism(s) by TNF- α

TNF- α mediates its effects through two different receptors: TNF receptor I (also known as p55 or p60) and TNF receptor II (also known as p75 or p80). Whereas TNF receptor I is expressed

*Corresponding Author: Bharat B. Aggarwal—Cytokine Research Laboratory, Department of Experimental Therapeutics, The University of Texas MD Anderson Cancer Center, Unit 143, 1515 Holcombe Boulevard, Houston, Texas 77030, USA. Email: aggarwal@mdanderson.org

Therapeutic Targets of the TNF Superfamily, edited by Iqbal S. Grewal. ©2009 Landes Bioscience and Springer Science+Business Media.

on all cell types in the body, TNF receptor II is expressed selectively on endothelial cells and on cells of the immune system. $^{2.5}$ The cytoplasmic domain of the TNF receptor I has a death domain, which has been shown to sequentially recruit TNF receptor-associated death domain (TRADD), Fas-associated death domain (FADD) and FADD-like ICE (FLICE) (also called caspase-8) lead to caspase-3 activation, which in turn induces apoptosis by inducing degradation of multiple proteins. TRADD also recruits TNF receptor-associated factor (TRAF2), which through receptor-interacting protein (RIP) activates IkBa kinase (IKK) leading to IkBa phosphorylation, ubiquitination and degradation, which finally leads to NF-kB activation. Through recruitment of TRAF2, TNF also activates various mitogen-activated protein kinases (MAPK), including the c-jun N-terminal kinases (JNK) p38 MAPK and p42/p44 MAPK. TRAF2 is also essential for the TNF-induced activation of AKT, another cell-survival signaling pathway. Thus TNFRI activates both apoptosis and cell survival signaling pathways simultaneously. The survival signaling pathways simultaneously.

Gene-deletion studies have shown that TNFR2 can also activate NF- κ B, JNK, p38 MAPK and p42/p44 MAPK. 10 TNFR2 can also mediate TNF-induced apoptosis. 11 Because TNFR2 cannot recruit TRADD-FADD-FLICE, how TNFR2 mediates apoptosis is not understood. However, the true physiological role of TNF, its receptors and associated proteins has been explored through gene-deletion experiments. It was found that animals with homologous gene deletion are fully viable but are more susceptible to infection $^{12-24}$ (Table 1). Overall the deletion of TNF, its receptors and associated proteins indicates the critical role of this cytokine in protection from microorganisms, the formation of lymph nodes and the development of the immune system.

Role of TNF-a in Cancer

TNF- α , initially discovered as a result of its antitumor activity, has now been shown to mediate all steps involved in tumorigenesis, including cellular transformation, promotion, survival, proliferation, invasion, angiogenesis and metastasis²⁵ (Fig. 1). These are discussed in detail as follows.

TNF-α Can Induce Tumor Initiation and Promotion

A number of reports indicate that TNF- α induces tumor initiation and tumor promotion 5.26,27 Komori's group reported that human TNF- α is 1000 times more effective than the chemical tumor promoters okadaic acid and 12-O-tetradecanoylphorbol-13-acetate in inducing cancer. Once initiated with these chemical carcinogens and exposed for 2 weeks to TNF- α , BALB/3T3 cells underwent transformation and yielded tumors in nude mice. ²⁸ The essential role of TNF- α in tumor promotion has also been demonstrated using TNF- α -deficient mice. Specifically, okadaic acid did not show any tumor-promoting activity in TNF- f mice after up to 19 weeks of tumor promotion, whereas okadaic acid induced strong tumor-promoting activity in TNF+ f mice. Tumor development in TPA-treated TNF- f mice was delayed and both the average number of tumors per mouse and the tumor size were dramatically reduced compared with results for TNF+ f CD-1 mice. ²⁹ Similarly, in a model of chemically induced liver cancer, TNF- α production by hepatocytes was implicated in tumor development. ³⁰ All these reports establish that TNF- α plays a critical role in tumor promotion.

Tumor Cells Produce TNF-\alpha and Mediate Proliferation

TNF- α is also produced by a wide variety of tumor cells, including B-cell lymphoma, ^{31,32} cutaneous T-cell lymphoma, ³³ megakaryoblastic leukemia, ³⁴ adult T-cell leukemia, ³⁵ CLL, ³⁶ ALL, ³⁷ breast carcinoma, ³⁸ lung carcinoma, ³⁹ pancreatic cancer, ⁴⁰ ovarian carcinoma, ⁴¹ cervical epithelial cancer, ⁴² glioblastoma ⁴³ and neuroblastoma. ⁴⁴ In most of these cells, TNF- α acts as an autocrine growth factor; however; in some cell types TNF- α induces the expression of other growth factors that mediate proliferation of tumors. For instance, in cervical cells TNF- α induces amphiregulin, which induces the proliferation of cells, ⁴² whereas in pancreatic cells TNF- α induces the expression of epidermal growth factor receptor (EGFR) and transforming growth factor (TGF- α), which mediate proliferation. ⁴⁰

Table 1. Phenotype of mice with gene deletion for TNF, TNF receptor and receptor-associated proteins

Gene	Phenotype	Ref.
TNF	Homozygous mutants viable	12-14
	 Readily succumb to Listeria monocytogenes infection 	
	 Show reduced contact hypersensitivity responses 	
	 Resistant to lipopolysaccharide toxicity 	
	 Lack splenic primary B-cell follicle follicular dendritic cell network 	
	Exhibit resistance to skin carcinogenesis	
TNFR1	 Resistant to low levels of lipopolysaccharide 	15
	 Increased susceptibility to Listeria monocytogenes infection 	
TNFR2	 Resistant to low levels of lipopolysaccharide 	16
	Impaired T-cell development	
	 Reduced cytotoxic T-lymphocyte proliferation 	
	 Increased resistance to TNF-induced necrotic cell death 	
TRAF1	 Exhibit stronger proliferation than wild-type T-cell to anti-CD3 	17
	 Respond to TNF-induced NF-κB and AP-1 signaling pathways 	
	 Skin hypersensitive to TNF-induced necrosis 	
TRAF2	 Defective Th-dependent antibody response 	18, 19
	 CD40-mediated proliferation and NF-κB activation 	
	 Thymus and spleen atrophied and B-cell precursors depleted 	
	 Thymocytes and hematopoietic progenitors sensitive to 	
	TNF-induced apoptosis	
	Serum TNF levels elevated	
	 Reduced TNF-mediated JNK/SAPK activation 	
	Mild effect on NF-κB activation	
RIP1	 Appear normal at birth but fail to thrive 	20
	• Die at 1-3 days of age	
	 Extensive apoptosis in lymphoid and adipose tissue 	
	 RIP^{-/-} cells highly sensitive to TNFα-induced cell death 	
	 No NF-κB activation 	
FADD	 Do not survive beyond day 11.5 of embryogenesis 	21, 22
	Cardiac failure and abdominal hemorrhage	
	Chimeric embryos showing a high contribution of FADD	
	null mutantcells to the heart reproduce the phenotype	
	of FADD ^{-/-} mutants	
	 Activates rearrangement of the immunoglobulin and TCR genes 	
	Fas-induced apoptosis completely blocked	
	Fas-mediated activation-induced proliferation impaired	
Caspase-8		23
	Congested accumulation of erythrocytes in embryos	
	TNF receptors, Fas/Apo1 and DR3 fail to induce cell death	
ΙΚΚβ	Die at mid-gestation from uncontrolled liver apoptosis	24
	 IKKβ-deficientcells lack activation of IKK and NF-κB in 	
	response to TNF- α or IL-1 β	

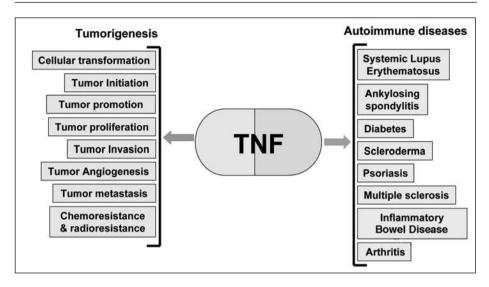


Figure 1. Pro-inflammatory effects of TNF are involved in tumorigenesis and in autoimmune diseases.

TNF- α Can Induce Invasion and Angiogenesis of Tumor Cells

That TNF- α can induce invasion and angiogenesis of tumor cells is well documented. TNF- α has been shown to confer an invasive, transformed phenotype on mammary epithelial cells. ³⁸ TNF- α has been reported to induce angiogenic factor upregulation in malignant glioma cells. ⁴⁵ This upregulation in turn promotes angiogenesis and tumor progression. TNF- α also stimulates epithelial tumor cell motility, which is a critical function in embryonic development, tissue repair and tumor invasion. ⁴⁶ TNF- α has been even reported to mediate macrophage-induced angiogenesis. ⁴⁷

Role of TNF- α in Tumor Metastasis

TNF- α also plays a role in the metastasis of cancer cells. In a model of experimental lung metastasis of colon adenocarcinoma, injection of LPS into mice enhanced the development of metastatic lesions. The increased metastasis was dependent on TNF- α production by host hematopoietic cells. This TNF- α activated NF- κ B in the tumor cells, increasing their proliferation and survival. Moreover, endogenous and exogenous TNF- α administration enhanced metastasis in an experimental fibrosarcoma metastasis model. Mice injected with fibrosarcoma cells showed enhanced metastasis to the lungs in the presence of exogenous TNF. Neutralization of endogenous tumor-induced TNF led to a significant decrease of the number of pulmonary metastases. An essential role of TNFR p55 has been found in liver metastases following intrasplenic administration of colon 26 cells. Malik et al described found that overexpression of TNF- α conferred invasive properties on xenograft tumors. Neutralization of endogenous TNF- α reversed the hepatic metastases and prolonged survival in mouse models.

Role of TNF-α in the Immune System

TNF- α is a critical component of effective immune surveillance and is required for proper proliferation and function of natural killer cells (NK-cells), T-cells, B-cells, macrophages and dendritic cells. TNF- α can influence inflammation and innate immunity, lymphoid organization and activation of APCs and can provide direct signals to T-cells. TNFR2 can augment T-cell proliferation and thus may also provide a costimulatory signal for T-cells. Mice strains in which the TNF- α gene or its p55 receptor has been deleted (TNF-KO or TNFR1-KO mice) have severe defects in lymph node follicle and germinal center formation. TNF- α acting through TNF

receptor p55 is involved in the development/maturation of dendritic cells (DCs) in bone marrow progenitor cultures. 60 Moreover, the microenvironment in peripheral lymphoid organs is associated with TNF-α signaling and chemokine production is critical for recruitment efficiency of DCs. 60 Follicular DCs are specialized mesenchymal cells that collect antigens in draining lymph nodes, interact with clonally expanding B-cells and form networks in the follicle under the influence of TNF-α and TNF- 60 33

TNFR1 is a costimulator of T-cell activation and is expressed by activated T-cells. The initiation of an immune response by dendritic cells originating in epithelial barriers and stimulating naive T-cells in draining lymph nodes involves active involvement of TNF/TNFR1. Moreover, TNF- α regulates the expansion and survival of CD4+ and CD8+ T-cells. ^{53,54} T-cell-derived TNF- α is important for protection against high bacterial load, whereas mastcell-derived TNF- α is a critical and early component of the allergic response. ⁶¹

TNF- α also plays a central role in initiating the inflammatory reactions of the innate immune system. Bacterial pathogens and several other proinflammatory and environmental stimuli induce TNF- α and NF- κ B signaling cascade via Toll-like receptors and also enhance its translational efficiency. Early production of TNF- α is prominent in the subsequent initiation of a highly complex biological cascade involving chemokines, cytokines and endothelial adhesions that recruits and activates neutrophils, macrophages and lymphocytes at the sites of infections. Release of preformed TNF- α acts as a positive autocrine feedback signal to activate NF- κ B and to induce further TNF- α and other cytokines such as granulocte-monocyte colony-stimulating factor (GM-CSF) and IL-8. In TNF- α exerts a global regulatory effect on the immune system.

Role of TNF-α in Autoimmune Diseases

Dysregulation of TNF- α has been implicated in a wide variety of autoimmune diseases, including rheumatoid arthritis, Crohn's disease, multiple sclerosis, psoriasis, scleroderma, systemic lupus erythromatosus, ankylosing spondylitis and diabetes (Fig. 1). How TNF- α mediates disease-causing effects is incompletely understood. The induction of proinflammatory genes by TNF- α has been linked to most diseases. The proinflammatory effects of TNF- α are primarily due to its ability to activate NF- κ B. ⁶³ Almost all cell types, when exposed to TNF- α , activate NF- κ B, leading to the expression of inflammatory genes. The role of TNF- α in some of the autoimmune diseases is discussed in detail below.

Psoriasis

Psoriasis is a chronic inflammatory disease of the skin, affecting 2-3% of the world's population. Histopathologically, psoriasis is characterized by hyperproliferation of epidermal keratinocytes and hyperkeratosis, as well as infiltration of immunocytes along with angiogenesis. ⁶⁴ T-cells play a major role in the initiation of psoriatic lesions. Activated T-cells in the region of the dermal epidermal junction promote the hyperplastic proliferative response through increased production of Th1 cytokines, among which TNF- α is the major player. ⁶⁵ In psoriatic lesions, levels of TNF- α -induced genes, such as IL-1 β , IL-8 and IL-6, are greatly increased. ⁶⁶⁻⁶⁸ Furthermore, in psoriatic plaques, there is a significant upregulation of activated phosphorylated NF- κ B compared with normal epidermis and uninvolved epidermis from psoriasis patients. ⁶⁹ TNF blockers have been shown to reverse the epidermal hyperplasia and cutaneous inflammation characteristic of psoriatic plaques. ⁷⁰ All these findings together suggest a major role for TNF- α in both initiation and progression of psoriasis.

Inflammatory Bowel Disease

Inflammatory bowel disease (IBD) is characterized by a chronic relapsing inflammation of the gastrointestinal tract and is divided into two primary forms: Crohn's disease and ulcerative colitis. ⁷¹ IBD is associated with the activation of local intestinal and systemic immune responses and is caused by the loss of tolerance against intestinal antigens. ⁷² TNF- α levels are elevated in the serum, mucosa and stool of IBD patients and $TNF^{-/-}$ mice show a marked reduction in chemically induced intestinal inflammation. ⁷³⁻⁷⁶ Increased nuclear translocation of NF- κ B has also been shown

in lamina propria mononuclear cells derived from IBD patients.⁷⁷ Hence TNF-α is considered to be an attractive target for the treatment of IBD and several antiTNF reagents have been developed, but most of them have not proven safe and efficacious in the treatment of IBD.

Arthritis

As a proinflammatory cytokine, TNF- α has perhaps the most dominant role in the etiology of rheumatoid arthritis. ⁷⁸ Patients with rheumatoid arthritis have high concentrations of TNF- α in the synovial fluid and at the cartilage-pannus junction, which leads to the erosion of bone. ^{79,80} In cultures of synovial cells from patients with rheumatoid arthritis, blocking TNF- α with antibodies significantly reduced the production of IL-1 β , IL-6, IL-8 and GM-CSF. ⁸¹ Hence, the inhibition of TNF- α has a more global effect on inflammation than the suppression of other cytokines present in high concentrations in synovial fluids, such as IL-1 β . The results of studies in animals provide further evidence of the importance of TNF- α in rheumatoid arthritis. In transgenic mice that expressed a deregulated human TNF- α gene, an inflammatory and destructive polyarthritis similar to rheumatoid arthritis spontaneously developed. ⁸² AntiTNF- α therapies are being used for the treatment of rheumatoid arthritis, but these agents are associated with side effects, some of them quite serious. ⁸³ Hence novel agents are needed for the management of rheumatoid arthritis.

Systemic Sclerosis (Scleroderma)

Systemic sclerosis (scleroderma) is a generalized connective tissue disorder, characterized by a wide spectrum of microvascular and immunological abnormalities, leading to progressive thickening and fibrosis of the skin and other visceral organs, such as the lungs, gastrointestinal tract, heart and kidneys. ^{84,85} Compelling evidence indicates that the increased production of TNF- α is involved in the pathogenesis of scleroderma. ⁸⁶ Patients with systemic sclerosis exhibit a systemic and local rise in TNF- α levels that leads to pulmonary fibrosis. ⁸⁷ The serum levels of TNFR1 are directly correlated to the severity of the disease. ⁸⁸ TNF- α gene polymorphism is also associated with scleroderma. ⁸⁹ Thus dysregulation of TNF- α plays a critical role in the development of systemic sclerosis in normal human subjects.

Systemic Lupus Erythematosus

Systemic lupus erythematosus (SLE) is a multifactorial autoimmune disease characterized by the breakdown of self-tolerance, B-cell hyperactivity, autoantibody production, aberrant formation of immune complexes and inflammation of multiple organs. The TNF- α level is increased and seems to be bioactive in the serum of patients with active SLE. The levels of TNF- α have been shown to correlate with SLE disease activity. Various antiTNF- α agents are currently being used for the treatment of SLE.

Ankylosing Spondylitis

Ankylosing spondylitis (AS) is an autoimmune disease characterized by prominent inflammation of the spinal joints and adjacent structures leading to progressive bony fusion of the spine. Pathophysiologically, TNF- α appears to play a role in promoting the inflammatory pattern associated with AS. Increased TNF- α protein is found in the sacroiliac joints and peripheral synovium sa well as the serum 79.98 of patients with active AS. While disease activity cannot be predicted from levels of TNF- α , blockade of this protein has been shown to have benefits in animal models and human studies of AS. Considering the critical role of TNF- α in the pathogenesis of AS, the molecules targeted at blocking the effects of TNF- α are likely to play a crucial role in the management of this disease.

Diabetes Mellitus

Autoimmune diabetes, or insulin-dependent diabetes mellitus (IDDM), is characterized by selective destruction of insulin-producing cells. ⁹⁹ The role of TNF- α in the pathogenesis of autoimmune diabetes has received increasing attention recently. ¹⁰⁰ It was shown that TNF- α in combination with IFN- γ could induce the aberrant expression of class II major histocompatibility complex

(MHC) molecules on pancreatic beta cells, suggesting a role for these cytokines in the induction of the autoimmune process in diabetes. 101 A different group of investigators has suggested that IL-1 β is toxic to pancreatic beta cells and that TNF- α significantly enhances this toxicity. 102 Transgenic mice, expressing constitutively active IKK- β , a kinase required for activation of NF- κ B, exhibited type 2 diabetes phenotype and increased hepatic production of TNF- α . Hepatic expression of the I κ B α super repressor reversed this diabetic phenotype in transgenic mice as well as wild-type mice fed a high-fat diet. 103 These findings indicate that lipid accumulation in the liver leads to subacute hepatic 'inflammation' through NF- κ B activation and downstream cytokine production. This causes insulin resistance both locally in liver and systemically. Thus novel blockers of TNF- α have significant implications for future new therapeutic strategies for insulin-dependent diabetes mellitus.

Multiple Sclerosis

Multiple sclerosis (MS) is an inflammatory disease of the central nervous system characterized by localized areas of demyelination. 104 TNF- α plays an important role in the pathogenesis of MS and its animal model, experimental autoimmune encephalomyelitis. 105,106 TNF- α has been detected in MS plaques 107,108 and circulating levels of TNF- α and its receptor have been found in cerebro-spinal fluid of MS patients. 109,110 All these findings support an enormous role for TNF- α inhibitors in the treatment of multiple sclerosis.

TNF Inhibitors

On the basis of the above descriptions, TNF blockers have tremendous potential for the treatment of various cancers and autoimmune diseases. Several classes of TNF- α inhibitors are available and these are discussed below.

TNF Antibodies

The best studied of the monoclonal TNF- α antibodies is infliximab (Remicade), originally referred to as cA2. Infliximab binds with high specificity and affinity to free and membrane-bound TNF- α , which is expressed at the cell surface by activated T-cells and macrophages. ¹¹¹ Adalimumab (Humira) is a human monoclonal IgG₁ antibody containing only human peptide sequences. It binds with high specificity and affinity to soluble and membrane-bound TNF- α and blocks its interaction with the p55 and p75 cell surface TNF receptors, thereby neutralizing the biological activities of this cytokine. ¹¹² However, these antibodies have demonstrated several potentially serious adverse effects that include greater predisposition towards infection, congestive heart failure, neurologic changes (e.g., demyelination), lymphomas, re-exacerbation of latent tuberculosis and problems related to autoimmunity, for example lupus-like syndrome. ¹¹³

Soluble TNF Receptors

In the second approach to TNF- α inhibition, soluble TNF receptors have been engineered as fusion proteins in which the extracellular ligand-binding portion of TNFRI or TNFR2 is coupled to a human immunoglobulin-like molecule. Etanercept (Enbrel) is a recombinant human fusion protein that consists of two soluble p75 TNF receptors and the F_c portion of human IgG_1 . Etanercept possesses a dimeric structure with high affinity to TNF- α and the linkage to the F_c portion of human IgG produces a longer half-life. Etanercept is better at neutralizing TNF- α than is the monomeric soluble p75 receptor. The various side effects observed include lymphomas, re-exacerbation of latent tuberculosis and problems related to autoimmunity. Recent studies indicate that administration of TNF- α inhibitors can even lead to psoriasis and contribute to the severity of the disease in paracoccidioidomycosis.

Besides p75, TNF has been shown to bind to p55 receptor with an affinity either equal or even greater than p75. ¹¹⁷ Although soluble p75 receptors clearly can sequester TNF, very little is known about the ability of the soluble form of the p55 receptor to sequester TNF in vivo.

Inhibitors of TNF Expression

Several compounds that can inhibit both TNF- α expression and synthesis are also available. These include thalidomide ([+]-alpha-phthalimidoglutarimide), which is currently being used for treatment of multiple myeloma^{118,119} and pentoxifylline, used to treat leg pain caused by poor blood circulation.¹²⁰ Thus these agents may be useful for the treatment of various cancers and autoimmune diseases mediated by TNF.

Inhibitors of TNF Oligomerization

Some inhibitors that can suppress oligomerization of TNF are also known. Steed and coworkers less designed a novel dominant-negative variant TNF protein that rapidly forms heterotrimers with native TNF to give complexes that neither bind to nor stimulate signaling through TNF receptors and thus inactivate TNF by sequestration. He et al less identified another small-molecule inhibitor that promotes subunit disassembly of trimeric TNF. This compound inhibited TNF activity in biochemical and cell-based assays, with median inhibitory concentrations of 22 and 4.6 micromolar, respectively. Formation of an intermediate complex between the compound and the intact trimer resulted in a 600-fold accelerated subunit dissociation rate that led to trimer dissociation.

Inhibitors of TNF- α -Induced Signaling Pathways

TNF- α activates cell survival signaling pathways, i.e., NF- κ B, Akt and MAPK pathways, as well as apoptotic pathways such as JNK, p38 and AP-1. Hence, inhibitors that target these pathways also have potential against various proinflammatory conditions mediated by TNF- α . For example, TNF- α activates NF- κ B, which in turn regulates TNF- α production. Hence various NF- κ B blockers (both synthetic and natural) are currently available on the market and effective against a wide variety of inflammatory conditions.

Natural Products as Inhibitors of TNF

Numerous plant-derived products have been identified that can suppress TNF- α expression from macrophages activated by numerous inflammatory stimuli (129-165, see Table 2). These include curcumin, resveratrol, emodin, silymarin and others. Thus these products are likely to be useful for the treatment of cancer and autoimmune diseases mediated by TNF.

Table 2. A list of natural products that inhibit the expression of TNF

- 1'-acetoxychavicol acetate123
- 1'-acetoxyeugenol acetate¹²⁴
- Allium sativum¹²⁵
- Aloe vera¹²⁶
- Aloe barbadensis¹²⁷
- Asparagus cochinchinensis¹²⁸
- Bisdemethoxycurcumin¹²⁹
- Butein¹³⁰
- Cardamomin¹³¹
- Curcumin^{132, 133}
- Diphenyl dimethyl bicarboxylate134
- Emodin inhibits IL-1β and IL-6¹³⁵
- Epigallocatechin gallate¹³⁶
- F022¹³⁷
- Ginkgolide B^{138, 139}
- 2'-Hydroxychalcone^{140, 141}
- Hypoestoxide¹⁴²

- Lonicera japonica¹⁴³
- Neolignans and lignans¹⁴⁴
- Patridoids I, II and IIA145
- Phthalide lactone¹⁴⁶
- Phloroglucinol derivatives¹⁴⁷
- Platycodin D and D3¹⁴⁸
- Phlebodium decumanum¹⁴⁹
- Phyllanthus amarus¹⁵⁰
- Resveratrol¹⁵¹
- 14,15-Secopregnanederivatives argelosides¹⁵²
- Silymarin¹⁵³
- Tanacetum microphyllum¹⁵⁴
- Taraxacum officinale¹⁵⁵
- Δ⁹-Tetrahydrocannabinoid¹⁵⁶
- Theobroma cacao¹⁵⁷
- Uncaria guianensis¹⁵⁸
- Zostera japonica¹⁵⁹

Conclusion

TNF clearly plays a major role in cancer and in autoimmune diseases. Because TNF is also needed for the proper functioning of the immune system, complete suppression of TNF over a long period is likely to prove harmful. The potential of TNF inhibitors in the treatment of autoimmune diseases as employed currently is just "the tip of the iceberg." Any chronic inflammatory condition, linked to majority of the inflammatory diseases, could be a potential target for antiTNF therapy. Thus the development of inhibitors that are orally active, safe and inexpensive would have major potential. Because of long-term safety and cost, nutraceuticals derived from fruits and vegetables, that can suppress TNF expression and TNF signaling, should be explored clinically for efficacy.

References

- 1. Pennica D, Nedwin GE, Hayflick JS et al. Human tumour necrosis factor: precursor structure, expression and homology to lymphotoxin. Nature 1984; 312:724-729.
- 2. Aggarwal BB. Signalling pathways of the TNF superfamily: a double-edged sword. Nat Rev Immunol 2003; 3:745-756.
- 3. Kurzrock R, Rosenblum MG, Sherwin SA et al. Pharmacokinetics, single-dose tolerance and biological activity of recombinant gamma-interferon in cancer patients. Cancer Res 1985; 45:2866-2872.
- Beutler B, Milsark IW, Cerami AC. Passive immunization against cachectin/tumor necrosis factor protects mice from lethal effect of endotoxin. Science 1985; 229:869-871.
- 5. Aggarwal BB, Shishodia S, Ashikawa K et al. The role of TNF and its family members in inflammation and cancer: lessons from gene deletion. Curr Drug Targets Inflamm Allergy 2002; 1:327-341.
- 6. Nagata S and Golstein P. The Fas death factor. Science 1995; 267:1449-1456.
- 7. Darnay BG Aggarwal BB. Early events in TNF signaling: a story of associations and dissociations. J Leukoc Biol 1997; 61:559-566.
- 8. Bhardwaj A and Aggarwal BB. Receptor-mediated choreography of life and death. J Clin Immunol 2003; 23:317-332.
- 9. Aggarwal BB and Takada Y. Pro-apototic and anti-apoptotic effects of tumor necrosis factor in tumor cells. Role of nuclear transcription factor NF-kappaB. Cancer Treat Res 2005; 126:103-127.
- 10. Mukhopadhyay A, Suttles J, Stout RD et al. Genetic deletion of the tumor necrosis factor receptor p60 or p80 abrogates ligand-mediated activation of nuclear factor-kappa B and of mitogen-activated protein kinases in macrophages. J Biol Chem 2001; 276:31906-31912.
- 11. Haridas V, Darnay BG, Natarajan K et al. Overexpression of the p80 TNF receptor leads to TNF-dependent apoptosis, nuclear factor-kappa B activation and c-Jun kinase activation. J Immunol 1998; 160:3152-3162.
- 12. Pasparakis M, Alexopoulou L, Episkopou V et al. Immune and inflammatory responses in TNF-alpha-deficient mice: a critical requirement for TNF-alpha in the formation of primary B-cell follicles, follicular dendritic cell networks and germinal centers and in the maturation of the humoral immune response. J Exp Med 1996; 184:1397-1411.
- 13. Marino MW, Dunn A, Grail D et al. Characterization of tumor necrosis factor-deficient mice. Proc Natl Acad Sci USA 1997; 94:8093-8098.
- 14. Moore RJ, Owens DM, Stamp G et al. Mice deficient in tumor necrosis factor-alpha are resistant to skin carcinogenesis. Nat Med 1999; 5:828-831.
- 15. Rothe J, Lesslauer W, Lotscher H et al. Mice lacking the tumour necrosis factor receptor 1 are resistant to TNF-mediated toxicity but highly susceptible to infection by Listeria monocytogenes. Nature 1993; 364:798-802.
- 16. Erickson SL, de Sauvage FJ, Kikly K et al. Decreased sensitivity to tumour-necrosis factor but normal T-cell development in TNF receptor-2-deficient mice. Nature 1994; 372:560-563.
- 17. Tsitsikov EN, Laouini D, Dunn IF et al. TRAF1 is a negative regulator of TNF signaling. enhanced TNF signaling in TRAF1-deficient mice. Immunity 2001; 15:647-657.
- 18. Yeh WC, Shahinian A, Speiser D et al. Early lethality, functional NF-kappaB activation and increased sensitivity to TNF-induced cell death in TRAF2-deficient mice. Immunity 1997; 7:715-725.
- 19. Nguyen LT, Duncan GS, Mirtsos C et al. TRAF2 deficiency results in hyperactivity of certain TNFR1 signals and impairment of CD40-mediated responses. Immunity 1999; 11:379-389.
- 20. Kelliher MA, Grimm S, Ishida Y et al. The death domain kinase RIP mediates the TNF-induced NF-kappaB signal. Immunity 1998; 8:297-303.
- 21. Yeh JH, Hsu SC, Han SH et al. Mitogen-activated protein kinase kinase antagonized fas-associated death domain protein-mediated apoptosis by induced FLICE-inhibitory protein expression. J Exp Med 1998; 188:1795-1802.

- 22. Zhang J, Cado D, Chen A et al. Fas-mediated apoptosis and activation-induced T-cell proliferation are defective in mice lacking FADD/Mort.1 Nature 1998; 392:296-300.
- Varfolomeev EE, Schuchmann M, Luria V et al. Targeted disruption of the mouse Caspase 8 gene ablates cell death induction by the TNF receptors, Fas/Apo,1 and DR3 and is lethal prenatally. Immunity 1998; 9:267-276.
- 24. Li ZW, Chu W, Hu Y et al. The IKKbeta subunit of IkappaB kinase (IKK) is essential for nuclear factor kappaB activation and prevention of apoptosis. J Exp Med 1999; 189:1839-1845.
- Balkwill F. Tumor necrosis factor or tumor promoting factor? Cytokine Growth Factor Rev 2002; 13:135-141.
- Aggarwal BB, Shishodia S, Takada Y et al. TNF blockade: an inflammatory issue. Ernst Schering Res Found Workshop, 2006:161-186.
- 27. Aggarwal BB, Shishodia S, Sandur SK et al. Inflammation and cancer: how hot is the link? Biochem Pharmacol 2006; 72:1605-1621.
- 28. Komori A, Yatsunami J, Suganuma M et al. Tumor necrosis factor acts as a tumor promoter in BALB/3T3 cell transformation. Cancer Res 1993; 53:1982-1985.
- Suganuma M, Okabe S, Marino MW et al. Essential role of tumor necrosis factor alpha (TNF-alpha) in tumor promotion as revealed by TNF-alpha-deficient mice. Cancer Res 1999; 59:4516-4518.
- Knight B, Yeoh GC, Husk KL et al. Impaired preneoplastic changes and liver tumor formation in tumor necrosis factor receptor type 1 knockout mice. J Exp Med 2000; 192:1809-1818.
- 31. Digel W, Stefanic M, Schoniger W et al. Tumor necrosis factor induces proliferation of neoplastic B-cells from chronic lymphocytic leukemia. Blood 1989; 73:1242-1246.
- 32. Digel W, Schoniger W, Stefanic M et al. Receptors for tumor necrosis factor on neoplastic B-cells from chronic lymphocytic leukemia are expressed in vitro but not in vivo. Blood 1990; 76:1607-1613.
- 33. Giri DK and Aggarwal BB. Constitutive activation of NF-kappaB causes resistance to apoptosis in human cutaneous T-cell lymphoma HuT-78 cells. Autocrine role of tumor necrosis factor and reactive oxygen intermediates. J Biol Chem 1998; 273:14008-14014.
- 34. Liu RY, Fan C, Mitchell S et al. The role of type I and type II tumor necrosis factor (TNF) receptors in the ability of TNF-alpha to transduce a proliferative signal in the human megakaryoblastic leukemic cell line Mo7e. Cancer Res 1998; 58:2217-2223.
- 35. Tsukasaki K, Miller CW, Kubota T et al. Tumor necrosis factor alpha polymorphism associated with increased susceptibility to development of adult T-cell leukemia/lymphoma in human T-lymphotropic virus type 1 carriers. Cancer Res 2001; 61:3770-3774.
- 36. Duncombe AS, Heslop HE, Turner M et al. Tumor necrosis factor mediates autocrine growth inhibition in a chronic leukemia. I Immunol 1989; 143:3828-3834.
- Elbaz O and Mahmoud LA. Tumor necrosis factor and human acute leukemia. Leuk Lymphoma 1994; 12:191-195.
- 38. Montesano R, Soulie P, Eble JA et al. Tumour necrosis factor lpha confers an invasive, transformed phenotype on mammary epithelial cells. J Cell Sci 2005; 118:3487-3500.
- 39. Kalthoff H, Roeder C, Gieseking J et al. Inverse regulation of human ERBB2 and epidermal growth factor receptors by tumor necrosis factor alpha. Proc Natl Acad Sci USA 1993; 90:8972-8976.
- 40. Schmiegel W, Roeder C, Schmielau J et al. Tumor necrosis factor alpha induces the expression of transforming growth factor alpha and the epidermal growth factor receptor in human pancreatic cancer cells. Proc Natl Acad Sci USA 1993; 90:863-867.
- 41. Wu S, Boyer CM, Whitaker RS et al. Tumor necrosis factor alpha as an autocrine and paracrine growth factor for ovarian cancer: monokine induction of tumor cell proliferation and tumor necrosis factor alpha expression. Cancer Res 1993; 53:1939-1944.
- 42. Woodworth CD, McMullin E, Iglesias M et al. Interleukin 1 alpha and tumor necrosis factor alpha stimulate autocrine amphiregulin expression and proliferation of human papillomavirus-immortalized and carcinoma-derived cervical epithelial cells. Proc Natl Acad Sci USA 1995; 92:2840-2844.
- 43. Aggarwal BB, Schwarz L, Hogan ME et al. Triple helix-forming oligodeoxyribonucleotides targeted to the human tumor necrosis factor (TNF) gene inhibit TNF production and block the TNF-dependent growth of human glioblastoma tumor cells. Cancer Res 1996; 56:5156-5164.
- 44. Goillot E, Combaret V, Ladenstein R et al. Tumor necrosis factor as an autocrine growth factor for neuroblastoma. Cancer Res 1992; 52:3194-3200.
- 45. Nabors LB, Suswam E, Huang Y et al. Tumor necrosis factor alpha induces angiogenic factor up-regulation in malignant glioma cells: a role for RNA stabilization and HuR. Cancer Res 2003; 63:4181-4187.
- Rosen EM, Goldberg ID, Liu D et al. Tumor necrosis factor stimulates epithelial tumor cell motility. Cancer Res 1991; 51:5315-5321.
- 47. Leibovich SJ, Polverini PJ, Shepard HM et al. Macrophage-induced angiogenesis is mediated by tumour necrosis factor-alpha. Nature 1987; 329:630-632.

- 48. Luo JL, Maeda S, Hsu LC et al. Inhibition of NF-kappaB in cancer cells converts inflammationinduced tumor growth mediated by TNF-alpha to TRAIL-mediated tumor regression. Cancer Cell 2004; 6:297-305.
- 49. Orosz P, Echtenacher B, Falk W et al. Enhancement of experimental metastasis by tumor necrosis factor. J Exp Med 1993; 177:1391-1398.
- 50. Kitakata H, Nemoto-Sasaki Y, Takahashi Y et al. Essential roles of tumor necrosis factor receptor p55 in liver metastasis of intrasplenic administration of colon 26 cells. Cancer Res 2002; 62:6682-6687.
- 51. Malik ST, Naylor MS, East N et al. Cells secreting tumour necrosis factor show enhanced metastasis in nude mice. Eur J Cancer 1990; 26:1031-1034.
- 52. Orosz P, Kruger A, Hubbe M et al. Promotion of experimental liver metastasis by tumor necrosis factor. Int J Cancer 1995; 60:867-871.
- 53. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. Cell 2001; 104:487-501.
- 54. Pfeffer K. Biological functions of tumor necrosis factor cytokines and their receptors. Cytokine Growth Factor Rev 2003; 14:185-191.
- 55. Croft M. Costimulatory members of the TNFR family: keys to effective T-cell immunity? Nat Rev Immunol 2003; 3:609-620.
- 56. Kim EY and Te HS. TNF type 2 receptor (p75) lowers the threshold of T-cell activation. J Immunol 2001; 167:6812-6820.
- 57. Fu YX and Chaplin DD. Development and maturation of secondary lymphoid tissues. Annu Rev Immunol 1999; 17:399-433.
- 58. Pasparakis M, Alexopoulou L, Grell M et al. Peyer's patch organogenesis is intact yet formation of B lymphocyte follicles is defective in peripheral lymphoid organs of mice deficient for tumor necrosis factor and its 55-kDa receptor. Proc Natl Acad Sci USA 1997; 94:6319-6323.
- 59. Kuprash DV, Tumanov AV, Liepinsh DJ et al. Novel tumor necrosis factor-knockout mice that lack Peyer's patches. Eur J Immunol 2005; 35:1592-1600.
- 60. Abe K, Yarovinsky FO, Murakami T et al. Distinct contributions of TNF and LT cytokines to the development of dendritic cells in vitro and their recruitment in vivo. Blood 2003; 101:1477-1483.
- Grivennikov SI, Tumanov AV, Liepinsh DJ et al. Distinct and nonredundant in vivo functions of TNF produced by T-cells and macrophages/neutrophils: protective and deleterious effects. Immunity 2005; 22:93-104.
- 62. Akira S, Takeda K. Toll-like receptor signalling. Nat Rev Immunol 2004; 4:499-511.
- 63. Aggarwal BB. Nuclear factor-kappaB: the enemy within. Cancer Cell 2004; 6:203-208.
- 64. Schon MP and Boehncke WH. Psoriasis. N Engl J Med 2005; 352:1899-1912.
- 65. Okubo Y and Koga M. Peripheral blood monocytes in psoriatic patients overproduce cytokines. J Dermatol Sci 1998; 17:223-232.
- 66. Nickoloff BJ, Karabin GD, Barker JN et al. Cellular localization of interleukin-8 and its inducer, tumor necrosis factor-alpha in psoriasis. Am J Pathol 1991; 138:129-140.
- 67. Gomi T, Shiohara T, Munakata T et al. Interleukin 1 alpha, tumor necrosis factor alpha and interferon gamma in psoriasis. Arch Dermatol 1991; 127:827-830.
- 68. Ettehadi P, Greaves MW, Wallach D et al. Elevated tumour necrosis factor-alpha (TNF-alpha) biological activity in psoriatic skin lesions. Clin Exp Immunol 1994; 96:146-151.
- 69. Lizzul PF, Aphale A, Malaviya R et al. Differential expression of phosphorylated NF-kappaB/RelA in normal and psoriatic epidermis and downregulation of NF-kappaB in response to treatment with etanercept. J Invest Dermatol 2005; 124:1275-1283.
- 70. Gottlieb AB, Chamian F, Masud S et al. TNF inhibition rapidly down-regulates multiple proinflammatory pathways in psoriasis plaques. J Immunol 2005; 175:2721-2729.
- 71. Bouma G, Strober W. The immunological and genetic basis of inflammatory bowel disease. Nat Rev Immunol 2003; 3:521-533.
- 72. Podolsky DK. Inflammatory bowel disease. N Engl J Med 2002; 347:417-429.
- 73. Komatsu M, Kobayashi D, Saito K et al. Tumor necrosis factor-alpha in serum of patients with inflammatory bowel disease as measured by a highly sensitive immuno-PCR. Clin Chem 2001; 47:1297-1301.
- 74. Braegger CP, Nicholls S, Murch SH et al. Tumour necrosis factor alpha in stool as a marker of intestinal inflammation. Lancet 1992; 339:89-91.
- 75. Breese EJ, Michie CA, Nicholls SW et al. Tumor necrosis factor alpha-producing cells in the intestinal mucosa of children with inflammatory bowel disease. Gastroenterology 1994; 106:1455-1466.
- 76. Neurath MF, Fuss I, Pasparakis M et al. Predominant pathogenic role of tumor necrosis factor in experimental colitis in mice. Eur J Immunol 1997; 27:1743-1750.
- 77. Schreiber S, Nikolaus S, Hampe J. Activation of nuclear factor kappa B inflammatory bowel disease. Gut 1998; 42:477-484.
- 78. Firestein GS. Evolving concepts of rheumatoid arthritis. Nature 2003; 423:356-361.

- 79. Saxne T, Palladino MA Jr, Heinegard D et al. Detection of tumor necrosis factor alpha but not tumor necrosis factor beta in rheumatoid arthritis synovial fluid and serum. Arthritis Rheum 1988; 31:1041-1045.
- 80. Chu CQ, Field M, Feldmann M et al. Localization of tumor necrosis factor alpha in synovial tissues and at the cartilage-pannus junction in patients with rheumatoid arthritis. Arthritis Rheum 1991; 34:1125-1132.
- 81. Butler DM, Maini RN, Feldmann M et al. Modulation of proinflammatory cytokine release in rheumatoid synovial membrane cell cultures. Comparison of monoclonal anti TNF-alpha antibody with the interleukin-1 receptor antagonist. Eur Cytokine Netw 1995; 6:225-230.
- 82. Keffer J, Probert L, Cazlaris H et al. Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. EMBO J 1991; 10:4025-4031.
- Roos JC and Ostor AJ. Tumor necrosis factor inhibitors for rheumatoid arthritis. N Engl J Med 2006; 355:2046-2047; author reply 2048.
- 84. Johnson RW, Tew MB, Arnett FC. The genetics of systemic sclerosis. Curr Rheumatol Rep 2002; 4:99-107.
- 85. Medsger TA, Jr. Assessment of damage and activity in systemic sclerosis. Curr Opin Rheumatol 2000; 12:545-548.
- 86. Gruschwitz MS, Albrecht M, Vieth G et al. In situ expression and serum levels of tumor necrosis factoralpha receptors in patients with early stages of systemic sclerosis. J Rheumatol 1997; 24:1936-1943.
- 87. Hasegawa M, Fujimoto M, Kikuchi K et al. Elevated serum tumor necrosis factor-alpha levels in patients with systemic sclerosis: association with pulmonary fibrosis. J Rheumatol 1997; 24:663-665.
- 88. Majewski S, Wojas-Pelc A, Malejczyk M et al. Serum levels of soluble TNF-alpha receptor type I and the severity of systemic sclerosis. Acta Derm Venereol 1999; 79:207-210.
- 89. Pandey JP and Takeuchi F. TNF-alpha and TNF-beta gene polymorphisms in systemic sclerosis. Hum Immunol 1999; 60:1128-1130.
- 90. Manson JJ and Isenberg DA. The pathogenesis of systemic lupus erythematosus. Neth J Med 2003; 61:343-346.
- 91. Aringer M, Feierl E, Steiner G et al. Increased bioactive TNF in human systemic lupus erythematosus: associations with cell death. Lupus 2002; 11:102-108.
- 92. Gabay C, Cakir N, Moral F et al. Circulating levels of tumor necrosis factor soluble receptors in systemic lupus erythematosus are significantly higher than in other rheumatic diseases and correlate with disease activity. J Rheumatol 1997; 24:303-308.
- 93. Sieper J, Braun J, Rudwaleit M et al. Ankylosing spondylitis: an overview. Ann Rheum Dis 2002; 61 Suppl 3:iii8-18.
- 94. Braun J, Bollow M, Neure L et al. Use of immunohistologic and in situ hybridization techniques in the examination of sacroiliac joint biopsy specimens from patients with ankylosing spondylitis. Arthritis Rheum 1995; 38:499-505.
- 95. Grom AA, Murray KJ, Luyrink L et al. Patterns of expression of tumor necrosis factor alpha, tumor necrosis factor beta and their receptors in synovia of patients with juvenile rheumatoid arthritis and juvenile spondylarthropathy. Arthritis Rheum 1996; 39:1703-1710.
- 96. Canete JD, Llena J, Collado A et al. Comparative cytokine gene expression in synovial tissue of early rheumatoid arthritis and seronegative spondyloarthropathies. Br J Rheumatol 1997; 36:38-42.
- 97. Gratacos J, Collado A, Filella X et al. Serum cytokines (IL-6, TNF-alpha, IL-1 beta and IFN-gamma) in ankylosing spondylitis: a close correlation between serum IL-6 and disease activity and severity. Br J Rheumatol 1994; 33:927-931.
- 98. Toussirot E, Lafforgue P, Boucraut J et al. Serum levels of interleukin 1-beta, tumor necrosis factor-alpha, soluble interleukin 2 receptor and soluble CD8 in seronegative spondylarthropathies. Rheumatol Int 1994; 13:175-180.
- 99. Moller DE. Potential role of TNF-alpha in the pathogenesis of insulin resistance and type 2 diabetes. Trends Endocrinol Metab 2000; 11:212-217.
- 100. Zoppini G, Faccini G, Muggeo M et al. Elevated plasma levels of soluble receptors of TNF-alpha and their association with smoking and microvascular complications in young adults with type 1 diabetes. J Clin Endocrinol Metab 2001; 86:3805-3808.
- 101. Pujol-Borrell R, Todd I, Doshi M et al. HLA class II induction in human isletcells by interferon-gamma plus tumour necrosis factor or lymphotoxin. Nature 1987; 326:304-306.
- 102. Mandrup-Poulsen T, Bendtzen K, Dinarello CA et al. Human tumor necrosis factor potentiates human interleukin 1-mediated rat pancreatic beta-cell cytotoxicity. J Immunol 1987; 139:4077-4082.
- 103. Cai D, Yuan M, Frantz DF et al. Local and systemic insulin resistance resulting from hepatic activation of IKK-beta and NF-kappaB. Nat Med 2005; 11:183-190.
- 104. Bernard CC, Kerlero de Rosbo N. Multiple sclerosis: an autoimmune disease of multifactorial etiology. Curr Opin Immunol 1992; 4:760-765.

- Navikas V, Link H. Review: cytokines and the pathogenesis of multiple sclerosis. J Neurosci Res 1996; 45:322-333.
- 106. Selmaj K, Papierz W, Glabinski A et al. Prevention of chronic relapsing experimental autoimmune encephalomyelitis by soluble tumor necrosis factor receptor I. J Neuroimmunol 1995; 56:135-141.
- 107. Hofman FM, Hinton DR, Johnson K et al. Tumor necrosis factor identified in multiple sclerosis brain. J Exp Med 1989; 170:607-612.
- 108. Selmaj K, Raine CS, Cannella B et al. Identification of lymphotoxin and tumor necrosis factor in multiple sclerosis lesions. J Clin Invest 1991; 87:949-954.
- 109. Sharief MK, Hentges R. Association between tumor necrosis factor-alpha and disease progression in patients with multiple sclerosis. N Engl J Med 1991; 325:467-472.
- 110. Tsukada N, Matsuda M, Miyagi K et al. Increased levels of intercellular adhesion molecule-1 (ICAM-1) and tumor necrosis factor receptor in the cerebrospinal fluid of patients with multiple sclerosis. Neurology 1993; 43:2679-2682.
- 111. Maini R, St Clair EW, Breedveld F et al. Infliximab (chimeric antitumour necrosis factor alpha monoclonal antibody) versus placebo in rheumatoid arthritis patients receiving concomitant methotrexate: a randomised phase III trial. ATTRACT Study Group. Lancet 1999; 354:1932-1939.
- 112. Keystone EC, Kavanaugh AF, Sharp JT et al. Radiographic, clinical and functional outcomes of treatment with adalimumab (a human antitumor necrosis factor monoclonal antibody) in patients with active rheumatoid arthritis receiving concomitant methotrexate therapy: a randomized, placebo-controlled, 52-week trial. Arthritis Rheum 2004; 50:1400-1411.
- 113. Hasan U. Tumour necrosis factor inhibitors—what we need to know. N Z Med J 2006; 119:U2336.
- 114. Bathon JM, Martin RW, Fleischmann RM et al. A comparison of etanercept and methotrexate in patients with early rheumatoid arthritis. N Engl J Med 2000; 343:1586-1593.
- 115. Ubriani R, Van Voorhees AS. Onset of psoriasis during treatment with TNF-{alpha} antagonists: a report of 3 cases. Arch Dermatol 2007; 143:270-272.
- 116. Corvino CL, Mamoni RL, Fagundes GZ et al. Serum interleukin-18 and soluble tumour necrosis factor receptor 2 are associated with disease severity in patients with paracoccidioidomycosis. Clin Exp Immunol 2007; 147:483-490.
- 117. Higuchi M, Aggarwal BB. Modulation of two forms of tumor necrosis factor receptors and their cellular response by soluble receptors and their monoclonal antibodies. J Biol Chem 1992; 267:20892-20899.
- 118. Majumdar S, Lamothe B, Aggarwal BB. Thalidomide suppresses NF-kappa B activation induced by TNF and H₂O₂, but not that activated by ceramide, lipopolysaccharides, or phorbol ester. J Immunol 2002; 168:2644-2651.
- Harousseau JL. Thalidomide in multiple myeloma: past, present and future. Future Oncol 2006;
 2:577-589.
- 120. Zabel P, Schade FU, Schlaak M. Inhibition of endogenous TNF formation by pentoxifylline. Immunobiology 1993; 187:447-463.
- 121. Steed PM, Tansey MG, Zalevsky J et al. Inactivation of TNF signaling by rationally designed dominant-negative TNF variants. Science 2003; 301:1895-1898.
- 122. He MM, Smith AS, Oslob JD et al. Small-molecule inhibition of TNF-alpha. Science 2005; 310:1022-1025.
- 123. Grzanna R, Phan P, Polotsky A et al. Ginger extract inhibits beta-amyloid peptide-induced cyto-kine and chemokine expression in cultured THP-1 monocytes. J Altern Complement Med 2004; 10:1009-1013.
- 124. Matsuda H, Morikawa T, Managi H et al. Antiallergic principles from Alpinia galanga: structural requirements of phenylpropanoids for inhibition of degranulation and release of TNF-alpha and IL-4 in RBL-2H3 cells. Bioorg Med Chem Lett 2003; 13:3197-3202.
- 125. Makris A, Thornton CE, Xu B et al. Garlic increases IL-10 and inhibits TNF-alpha and IL-6 production in endotoxin-stimulated human placental explants. Placenta 2005; 26:828-834.
- 126. Duansak D, Somboonwong J, Patumraj S. Effects of Aloe vera on leukocyte adhesion and TNF-alpha and IL-6 levels in burn wounded rats. Clin Hemorheol Microcirc 2003; 29:239-246.
- 127. Qiu Z, Jones K, Wylie M et al. Modified Aloe barbadensis polysaccharide with immunoregulatory activity. Planta Med 2000; 66:152-156.
- 128. Kim H, Lee E, Lim T et al. Inhibitory effect of Asparagus cochinchinensis on tumor necrosis factor-alpha secretion from astrocytes. Int J Immunopharmacol 1998; 20:153-162.
- 129. Matsuda H, Tewtrakul S, Morikawa T et al. Anti-allergic principles from Thai zedoary: structural requirements of curcuminoids for inhibition of degranulation and effect on the release of TNF-alpha and IL-4 in RBL-2H3 cells. Bioorg Med Chem 2004; 12:5891-5898.
- 130. Lee SH, Seo GS, Sohn DH. Inhibition of lipopolysaccharide-induced expression of inducible nitric oxide synthase by butein in RAW 264.7 cells. Biochem Biophys Res Commun 2004; 323:125-132.

- 131. Lee JH, Jung HS, Giang PM et al. Blockade of nuclear factor-kappaB signaling pathway and antiinflammatory activity of cardamomin, a chalcone analog from Alpinia conchigera. J Pharmacol Exp Ther 2006; 316:271-278.
- 132. Chan MM. Inhibition of tumor necrosis factor by curcumin, a phytochemical. Biochem Pharmacol 1995; 49:1551-1556.
- 133. Shishodia S, Amin HM, Lai R et al. Curcumin (diferuloylmethane) inhibits constitutive NF-kappaB activation, induces G1/S arrest, suppresses proliferation and induces apoptosis in mantle cell lymphoma. Biochem Pharmacol 2005; 70:700-713.
- 134. Gao M, Zhang J, Liu G. Effect of diphenyl dimethyl bicarboxylate on concanavalin A-induced liver injury in mice. Liver Int 2005; 25:904-912.
- 135. Kuo YC, Tsai WJ, Meng HC et al. Immune reponses in human mesangial cells regulated by emodin from Polygonum hypoleucum Ohwi. Life Sci 2001; 68:1271-1286.
- 136. Matsunaga K, Klein TW, Friedman H et al. Epigallocatechin gallate, a potential immunomodulatory agent of tea components, diminishes cigarette smoke condensate-induced suppression of antiLegionella pneumophila activity and cytokine responses of alveolar macrophages. Clin Diagn Lab Immunol 2002; 9:864-871.
- 137. Lin AH, Fang SX, Fang JG et al. [Studies on anti-endotoxin activity of F022 from Radix Isatidis]. Zhongguo Zhong Yao Za Zhi 2002; 27:439-442.
- 138. Nie ZG, Peng SY, Wang WJ. [Effects of ginkgolide B on lipopolysaccharide-induced TNF-alpha production in mouse peritoneal macrophages and NF-kappaB activation in rat pleural polymorphonuclear leukocytes]. Yao Xue Xue Bao 2004; 39:415-418.
- 139. Wadsworth TL, McDonald TL, Koop DR. Effects of Ginkgo biloba extract (EGb 761) and quercetin on lipopolysaccharide-induced signaling pathways involved in the release of tumor necrosis factor-alpha. Biochem Pharmacol 2001; 62:963-974.
- 140. Abuarqoub H, Foresti R, Green CJ et al. Heme oxygenase-1 mediates the anti-inflammatory actions of 2'-hydroxychalcone in RAW 264.7 murine macrophages. Am J Physiol Cell Physiol 2006; 290: C1092-1099.
- 141. Ban HS, Suzuki K, Lim SS et al. Inhibition of lipopolysaccharide-induced expression of inducible nitric oxide synthase and tumor necrosis factor-alpha by 2'-hydroxychalcone derivatives in RAW 264.7 cells. Biochem Pharmacol 2004; 67:1549-1557.
- 142. Ojo-Amaize EA, Kapahi P, Kakkanaiah VN et al. Hypoestoxide, a novel anti-inflammatory natural diterpene, inhibits the activity of IkappaB kinase. Cell Immunol 2001; 209:149-157.
- 143. Kang OH, Choi YA, Park HJ et al. Inhibition of trypsin-induced mastcell activation by water fraction of Lonicera japonica. Arch Pharm Res 2004; 27:1141-1146.
- 144. Cho JY, Park J, Yoo ES et al. Inhibitory effect of lignans from the rhizomes of Coptis japonica var. dissecta on tumor necrosis factor-alpha production in lipopolysaccharide-stimulated RAW264.7 cells. Arch Pharm Res 1998; 21:12-16.
- 145. Ju HK, Moon TC, Lee E et al. Inhibitory effects of a new iridoid, patridoid II and its isomers, on nitric oxide and TNF-alpha production in cultured murine macrophages. Planta Med 2003; 69:950-953.
- 146. Liu L, Ning ZQ, Shan S et al. Phthalide Lactones from Ligusticum chuanxiong inhibit lipopolysaccharide-induced TNF-alpha production and TNF-alpha-mediated NF-kappaB Activation. Planta Med 2005; 71:808-813.
- 147. Ishii R, Horie M, Saito K et al. Inhibition of lipopolysaccharide-induced pro-inflammatory cytokine expression via suppression of nuclear factor-kappaB activation by Mallotus japonicus phloroglucinol derivatives. Biochim Biophys Acta 2003; 1620:108-118.
- 148. Wang C, Schuller Levis GB, Lee EB et al. Platycodin D and D3 isolated from the root of Platycodon grandiflorum modulate the production of nitric oxide and secretion of TNF-alpha in activated RAW 264.7 cells. Int Immunopharmacol 2004; 4:1039-1049.
- Punzon C, Alcaide A, Fresno M. In vitro anti-inflammatory activity of Phlebodium decumanum. Modulation of tumor necrosis factor and soluble TNF receptors. Int Immunopharmacol 2003; 3:1293-1299.
- 150. Kiemer AK, Hartung T, Huber C et al. Phyllanthus amarus has anti-inflammatory potential by inhibition of iNOS, COX-2 and cytokines via the NF-kappaB pathway. J Hepatol 2003; 38:289-297.
- 151. Bi XL, Yang JY, Dong YX et al. Resveratrol inhibits nitric oxide and TNF-alpha production by lipopolysaccharide-activated microglia. Int Immunopharmacol 2005; 5:185-193.
- 152. Perrone A, Plaza A, Ercolino SF et al. 14,15-secopregnane derivatives from the leaves of Solenostemma argel. J Nat Prod 2006; 69:50-54.
- 153. Zi X, Mukhtar H, Agarwal R. Novel cancer chemopreventive effects of a flavonoid antioxidant silymarin: inhibition of mRNA expression of an endogenous tumor promoter TNF-alpha. Biochem Biophys Res Commun 1997; 239:334-339.

- 154. Abad MJ, Bermejo P, Alvarez M et al. Flavonoids and a sesquiterpene lactone from Tanacetum microphyllum inhibit anti-inflammatory mediators in LPS-stimulated mouse peritoneal macrophages. Planta Med 2004; 70:34-38.
- 155. Kim HM, Shin HY, Lim KH et al. Taraxacum officinale inhibits tumor necrosis factor-alpha production from rat astrocytes. Immunopharmacol Immunotoxicol 2000; 22:519-530.
- 156. Verhoeckx KC, Korthout HA, van Meeteren-Kreikamp AP et al. Unheated Cannabis sativa extracts and its major compound THC-acid have potential immuno-modulating properties not mediated by CB1 and CB2 receptor coupled pathways. Int Immunopharmacol 2006; 6:656-665.
- 157. Ramiro E, Franch A, Castellote C et al. Effect of Theobroma cacao flavonoids on immune activation of a lymphoid cell line. Br J Nutr 2005; 93:859-866.
- 158. Piscoya J, Rodriguez Z, Bustamante SA et al. Efficacy and safety of freeze-dried car's claw in osteoarthritis of the knee: mechanisms of action of the species Uncaria guianensis. Inflamm Res 2001; 50:442-448.
- 159. Hua KF, Hsu HY, Su YC et al. Study on the antiinflammatory activity of methanol extract from seagrass Zostera japonica. J Agric Food Chem 2006; 54:306-311.