Chapter 2 Primary Myelofibrosis

Ayalew Tefferi

2.1 Introduction

Myelofibrosis (MF) in the context of a myeloproliferative disorder is a clinicopathologically defined entity characterized by anemia, marked splenomegaly, constitutional symptoms, leukoerythroblastosis (i.e., the presence of immature granulocytes and nucleated red blood cells), dacryocytosis (i.e., presence of teardrop-shaped red blood cells), and a bone marrow that displays dysplastic megakaryocyte hyperplasia, granulocyte proliferation, and reticulin and/or collagen fibrosis (1). Disease presentation could be either *de novo* (primary MF; PMF) or preceded by either polycythemia vera (post-PV MF) or essential thrombocythemia (post-ET MF). PMF is also known by many other names (Table 2.1), including chronic idiopathic myelofibrosis (CIMF), the term used by the World Health Organization (WHO) system for classification of myeloid neoplasms (2). However, the use of the term "PMF" was recently endorsed by the International Working Group for Myelofibrosis Research and Treatment (IWG-MRT) (3).

2.2 Historical Perspective and Disease Classification

The first description of PMF is credited to Heuck (1879) (4). He described two cases, which he referred to at the time as "splenic-medullary leukemia" and "pure splenic leukemia" (4). The term "osteosclerosis," which refers to the new bone formation that often accompanies bone marrow fibrosis in PMF, was first introduced by Assman in 1907 (5). Post-PV MF was recognized as early as 1935 by Hirsch-. (6). Similarly, the typical histological characteristics of the disease, including hepatosplenic extramedullary hematopoiesis (1908) (7, 8), bone marrow fibrosis (8), and leukoerythroblastosis (1939) (9), were all recognized in the first half of the 20th century. PMF was referred to as agnogenic myeloid metaplasia of the spleen (AMM), a term first used in 1940 (10) and later endorsed by Dameshek in 1951 (11).

Dameshek classified PMF as a myeloproliferative disorder (MPD), along with chronic myeloid leukemia (CML), ET, and PV (11). In 1960, Nowell and

 Table 2.1
 Terms used to refer to primary myelofibrosis

Agnogenic myeloid metaplasia Atypical myeloid leukemia Atypical myelosis Aleukemic myelosis with osteosclerosis Chronic idiopathic myelofibrosis Chronic nonleukemic myelosis Chronic erythroblastosis Chronic megakaryocytic leukemia Chronic megakaryocytic-granulocytic myelosis Heuck-Assman disease Idiopathic myelofibrosis Idiopathic myeloid metaplasia Leukoerythroblastic anemia Leukanemia Megakaryocytic myelosis with osteosclerosis Megakaryocytic splenomegaly Myelosis Myelofibrosis with myeloid metaplasia Myeloid metaplasia with myelofibrosis Megakaryocytic myelosis Myelosclerosis Osteosclerotic pseudoleukemia Osteomyeloreticulosis Osteomyelosclerosis Osteosclerotic anemia Osteomyelofibrosis Primary myelofibrosis

Hungerford described the Philadelphia chromosome in CML (12), which was later shown to harbor first the t(9;22)(q32;q13) (13) and subsequently the *BCR-ABL* disease-causing mutation (14). Accordingly, modern classification systems list PMF, PV, and ET as *BCR-ABL*-negative classic MPDs (15). In 1978, G6PD-based clonality studies established PMF as a stem-cell-derived clonal myeloproliferation (16). In 2005, a novel gain-of-function (GOF) mutation involving the JAK2 tyrosine kinase (*JAK2*V617F) was described in approximately 50% of PMF patients but also in the majority of those with PV as well as ET (17). In 2006, another GOF mutation involving MPL (*MPL*W515L/K) was described in approximately 5% of patients with PMF (18).

In 1967, an International Polycythemia Vera Study Group (PVSG) was created under the auspices of the National Cancer Institute and the group provided, for the first time, formal criteria for the diagnosis of each one of the *BCR-ABL*negative classic MPDs, including PMF (19). Subsequently, a WHO-sponsored committee on the classification of hematological malignancies revised the PVSG diagnostic criteria for PMF and reorganized the overall classification system for myeloid neoplasms (2). The WHO system considers two broad categories of myeloid malignancies: acute myeloid leukemia (AML) and chronic myeloid disorders (CMDs) (20). AML is defined by the presence of 20% or more "blasts" in either the bone marrow or blood and/or certain recurring cytogenetic abnormalities including t(8;21)(q22;q22), t(15;17)(q22;q12), inv(16)(p13;q22), and t(16;16)(p13;q22) (20). Table 2.2 presents the current WHO classification scheme for CMD. Most recently, a semimolecular classification system has been proposed (Table 2.3) (21).

2.3 Epidemiology

The prevalence of PMF is similar in men and women (M:F = 1.6:1) and overall reported incidence figures range from 0.4 to 1.5/100,000 (22–26). A higher incidence has been suggested in persons of Jewish ancestry (27). Median age at diagnosis is estimated between 55 and 60 years and approximately 2%, 10%, and 30% of patients are diagnosed before age 30, 40, and 50 years, respectively (28). In one study of 323 patients, 9 (2.8%; 6 females) were age 30 years or younger (range: 17–30). The clinical course in these nine young patients was more indolent compared to that seen in older adults and more like that seen in children, where disease occurrence is very rare (29, 30). In general, there is little evidence that links PMF to environmental toxins. However, the possibility of some association with exposure to benzene, other industrial solvents, thorotrast injections, and radiation accidents has been suggested in the past (31–35).

| Major categories | Subcategories |
|--------------------------------------|--------------------------------------|
| 1. Myelodysplastic syndrome (MDS) | |
| 2. Myeloproliferative disorder (MPD) | i. Chronic myeloid leukemia (CML) |
| | ii. Polycythemia vera |
| | iii. Essential thrombocythemia |
| | iv. Primary myelofibrosis |
| | v. Chronic neutrophilic leukemia |
| | vi. Chronic eosinophilic leukemia |
| | viii. Hypereosinophilic syndrome |
| | ix. Unclassified MPD |
| 3. MDS/MPD | i. Chronic myelomonocytic leukemia |
| | ii. Juvenile myelomonocytic leukemia |
| | iii. Atypical CML |
| 4. Systemic mastocytosis (SM) | |

Table 2.2 The World health Organization classification system for chronic myeloid disorders

| Mai | in | Clinicopathologic | Molecular |
|------|---|--|-------------------------------------|
| cate | egories | subcategories | subcategories |
| I. | Myelodysplastic syndrome | According to WHO classification system | |
| II. | Classic myeloproliferative disorders | 1. Chronic myeloid leukemia | 100% BCR-ABL ⁽⁺⁾ |
| | | 2. Polycythemia vera | ~100% JAK2V617F ⁽⁺⁾ |
| | | 3. Essential thrombocythemia | ~50% JAK2V617F ⁽⁺⁾ |
| | | | ~1% MPLW515L/K ⁽⁺⁾ |
| | | 4. Primary Myelofibrosis | ~50% JAK2V617F ⁽⁺⁾ |
| | | | ~5% MPLW515L/K ⁽⁺⁾ |
| III. | Atypical myeloproliferative disorders | 1. Chronic myelomonocytic leukemia | ~3% JAK2V617F ⁽⁺⁾ |
| | | 2. Juvenile myelomonocytic leukemia | ~30% PTPN11 mutation ⁽⁺⁾ |
| | | | ~15% NF1 mutation ⁽⁺⁾ |
| | | | ~15% RAS mutation ⁽⁺⁾ |
| | | 3. Chronic neutrophilic leukemia | ~20% JAK2V617F ⁽⁺⁾ |
| | | 4. Chronic eosinophilic leukemia/eosinophilic MPD | A. PDGFRA-rearranged |
| | | _ | B. PDGFRB-rearranged |
| | | | C. FGFR1-rearranged |
| | | | D. Molecularly undefine |
| | | 5. Hypereosinophilic syndrome | |
| | | 6. Chronic basophilic leukemia | |
| | | 7. Systemic mastocytosis | A. KITD816V ⁽⁺⁾ |
| | | | B. Other KIT mutation |
| | | | C. FIP1L1-PDGFRA ⁽⁺⁾ |
| | | | D. Molecularly undefine |
| | | 8. Unclassified MPD | $\sim 20\% JAK2V617F^{(+)}$ |
| | | i. Mixed/overlap MDS/MPD | |
| | | ii. CML-like but <i>BCR-ABL</i> ⁽⁻⁾ | |

 Table 2.3
 A semimolecular classification of chronic myeloid disorders

2.4 Pathogenesis

The central pathogenetic process in PMF is stem-cell-derived clonal myeloproliferation (16, 36). Unlike the case with CML and *BCR-ABL*, the primary oncogenic event in PMF has not been characterized. However, activating mutations of the JAK2 tyrosine kinase (*JAK2*V617F) and thrombopoietin receptor (*MPL*W515L/K) have recently been reported in approximately 50% and 5% of patients, respectively (17, 18). *JAK2*V617F is an exon 14 *JAK2* mutation at nucleotide position 1849 representing a G to T somatic point mutation. The mutation results in the substitution of value to phenylalanine at

codon 617. *MPL*W515L mutation represents a G to T transition at nucleotide 1544, resulting in a tryptophan to leucine substitution at codon 515 of the transmembrane region of the MPL receptor (18). *JAK2*V617F has also been described in ET at a similar mutational frequency and in PV, where almost all patients carry the mutation (17, 37–39). Similarly, *MPL*W515L/K also occurs in approximately 1% of ET patients (40). Both mutations induce an MPD phenotype in mice, the former a PV-like disease (17, 41, 42) and the latter a PMF-like disease (43). Regardless, about half of the patients with PMF do not display either mutation and the precise pathogenetic role of these mutations, when they are present, remains to be clarified.

The bone marrow stromal reaction in PMF, including reticulin/collagen fibrosis, osteosclerosis, and angiogenesis is currently believed to be reactive in nature and cytokine mediated. In mice, for example, PMF-associated bone marrow stromal changes have been induced by either systemic overexpression of thrombopoietin (TPO^{high} mice) or by megakaryocyte lineage restricted underexpression of the transcription factor GATA-1 (GATA-1^{low} mice). In both instances, the megakaryocytes display abnormal distribution of P-selectin that is believed to promote a pathologic interaction between megakaryocytes and neutrophils (emperipolesis), resulting in the release of both fibrogenic and

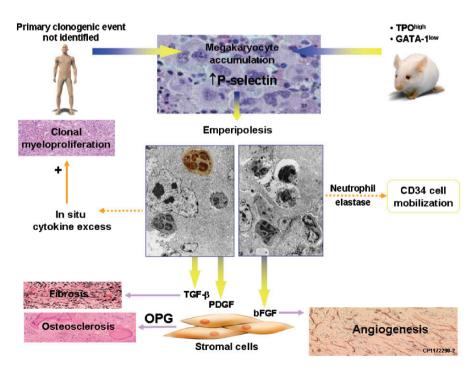


Figure 2.1 Pathogenesis of primary myelofibrosis. TPO, thrombopoietin; TGF, transforming growth factor; PDGF, platelet-derived growth factor; FGF, fibroblast growth factor; OPG, osteo-protegerin. (From Ref. 51; published with permission)

angiogenic cytokines, including transforming growth factor- $\beta 1$ (TGF- β), plateletderived growth factor (PDGF), basic fibroblast growth factor (bFGF), vascular endothelial growth factor (VEGF), tissue inhibitors of matrix metalloproteinases, and neutrophil-derived elastase and other proteases (44, 45). Among the latter, megakaryocyte-derived TGF- $\beta 1$ might be the most important in the pathogenesis of the stromal reaction in PMF (46–48). Peripheral blood expansion of both CD34-positive myeloid progenitors and endothelial cells provide additional evidence for aberrant bone microenvironment in PMF (49, 50). Figure 2.1 summarizes the current speculation regarding the mechanisms of stromal reaction in PMF (51, 52).

2.5 Clinical Features and Diagnosis

Most, but not all, patients with PMF are symptomatic at diagnosis. The typical presentation includes anemia, marked splenomegaly, and profound constitu tional symptoms, including fatigue and night sweats. Other manifestations, either at diagnosis or during the course of the disease, include left upper quadrant discomfort, including recurrent pain from splenic infarcts (may be referred to the left shoulder), early satiety and change in bowel habits, pruritus, easy bruising, peripheral edema, lymphadenopathy, ascites, bleeding, and thrombosis (53–55). The spleen is palpably enlarged in approximately 80% of patients at diagnosis (marked splenomegaly in half of the cases) and the liver in 50% (56). Organomegaly in PMF is secondary to extramedullary hematopoeisis (EMH) that might also involve other organs: lymph nodes (lymphadenopathy), pleura (effusion), peritoneum (ascites), lung (interstitial process), and the paraspinal and epidural spaces (spinal cord and nerve root compression) (57–60).

The peripheral blood smear in PMF often shows leukoerythroblastosis (presence of nucleated red blood cells and immature granulocytes) and teardrop-shaped red blood cells (Figure 2.2). Anemia is present at diagnosis in the majority of the patients and approximately 20% might be red blood cell transfusion-dependent at presentation (59, 61–63). Other laboratory abnormalities at diagnosis include leukocytosis (41–49% incidence), leukopenia (7–22%), thrombocytosis (13–31%), thrombocytopenia (21–37%), presence of circulating blasts (33–53%), increased serum levels of lactate dehydrogenase (LDH; 83%), and low cholesterol levels (32%) (59, 60, 62, 63).

Bone marrow examination reveals both "cellular phase" and "overtly fibrotic" stages of the disease (Figure 2.3) (64). In cellular-phase disease, reticulin fibrosis, might be absent (i.e., prefibrotic stage). Therefore, the most helpful diagnostic feature in the bone marrow is the presence of dense megakaryocyte clusters with atypical megakaryocyte morphology (cloudlike nuclear morphology) that is accompanied by increased granulocyte proliferation and reduced erythropoiesis (64). Additional histological features of advanced disease include osteosclerosis, dilated sinuses, and intrasinusoidal hematopoiesis (Figure 2.4).

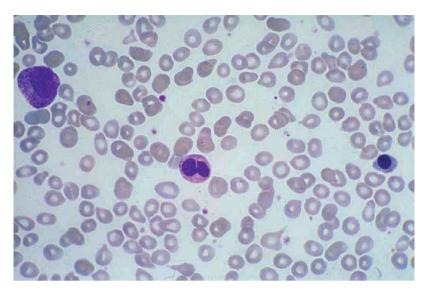


Figure 2.2 Peripheral blood smear in primary myelofibrosis showing myelophthisis; presence of nucleated red blood cells, immature granulocytes, and dacryocytes

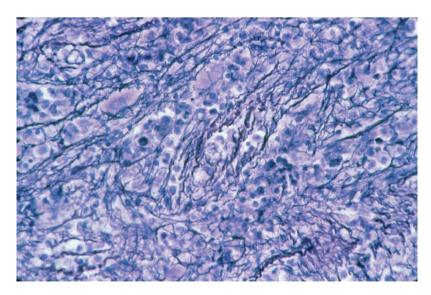


Figure 2.3 Reticulin fibrosis in primary myelofibrosis

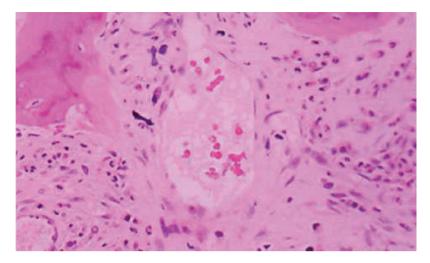


Figure 2.4 Osteosclerosis and intrasinusoidal hematopoiesis in primary myelofibrosis

2.6 Differential Diagnosis

Neither leukoerythroblastosis nor bone marrow fibrosis is specific to PMF. Bone marrow fibrosis might accompany a number of hematologic and nonhematologic conditions, as listed in Table 2.4. In most situations, mutation screening for *BCR-ABL* (to exclude the diagnostic possibility of CML) and *JAK2*V617F (to exclude the possibility of bone marrow fibrosis associated with nonmalignant condition, lymphoid disorder, or metastatic cancer) is highly recommended. It should be noted, however, that *JAK2*V617F cannot distinguish PMF from other myeloid disorders such as MDS, ET, PV, or atypical MPD. Therefore, accurate diagnosis requires careful morphological evaluation of the bone marrow.

Primary MF is typically characterized by the presence of morphologically abnormal megakaryocytes (bulbous and hyperchromatic nuclei) in dense clusters. MDS is characterized by the presence of erythroid and/or granulocytic dysplasia. As mentioned earlier, reticulin fibrosis can be absent in cellular-phase PMF and it is thus possible to confuse cellular-phase PMF with ET. Unlike ET, however, bone marrow in PMF is markedly hypercellular with both granulocytic and megakaryocytic proliferation in PMF as opposed to often normocellular bone marrow with only megakaryocytic hyperplasia in ET (65). Other distinguishing features between cellular-phase PMF and ET include the presence of myelophthisis and/or increased LDH in the former but not the latter.

Acute myelofibrosis, considered a variant of AML, can sometimes be confused with PMF. In general, patients with acute myelofibrosis usually present with severe constitutional symptoms, pancytopenia, mild or no splenomegaly, and circulating blasts. Both immunohistochemistry and cytogenetic studies are helpful in

| Hematologic disorders | | |
|---------------------------------|--------------------|---|
| Myeloid disorders | Lymphoid disorders | Nonhematologic disorders |
| | | Nonhematologic disorders Metastatic cancer (126) Autoimmune myelofibrosis (127) Systemic lupus erythematosus (128) Kala-Azar (leishmaniasis) (129) Tuberculosis (130) Paget's disease (131) HIV infection (132) Vitamin D-deficient rickets (133) Renal osteodystrophy (134) Hyperparathyroidism (135) Gray platelet |
| • Malignant histiocytosis (121) | | Familial infantile myelofibrosis (137) Idiopathic pulmonary hypertension (138) |

Table 2.4 Causes of bone marrow fibrosis

distinguishing PMF from both acute myelofibrosis and MDS with fibrosis. For example, CD34 and CD61 immunoperoxidase staining provides a better estimate of the marrow blast and megakaryocyte content, respectively. Similarly, although cytogenetic abnormalities that occur in approximately half of the patients with PMF are mostly not specific to the disease [e.g., del(20)(q11;q13), del(13)(q12;q22), trisomy 8, trisomy 9, del(12)(p11;p13), monosomy or long arm deletions involving chromosome 7, and partial trisomy 1q] (66). the presence of either del(13)(q12;q22) or der(6)t(1;6)(q21–23;p21–23) is strongly suggestive of PMF diagnosis (67).

2.7 Clinical Course and Prognosis

Primary MF displays a progressive course in the majority of cases, and disease complications include cachexia, peripheral edema, severe fatigue, excessive night sweats, low-grade fever, symptomatic portal hypertension, variceal bleeding, ascites, debilitating diffuse and/or extremity bone pain, and "idiopathic" pulmonary hypertension (68). Causes of death includes development of blast-phase PMF,

which occurs in approximately 10% of patients during the first decade of their disease (60, 63, 69, 70), infections (26–29%), bleeding (11–22%), heart failure (7–15%), liver failure (3–8%), solid tumor (3%), respiratory failure (3%), and portal hypertension (6%) (63, 69).

Survival in PMF is estimated by the use of one of several prognostic scoring systems (PSSs) that rely on the presence or absence of well-established adverse prognostic features (Table 2.5) (62, 63, 69, 71). Among the latter, the Mayo Clinic PSS has been reported to be superior, compared to other PSSs, in delineating both low-risk and intermediate-risk disease categories. According to the Mayo PSS (Table 2.5), median survival for low-risk young patients (age <60 years) approaches 15 years compared to approximately 5 years in intermediate-risk patients and less than 3 years in high-risk patients. Additional risk factors for inferior survival, in addition to those listed in Table 2.5, include circulating immature granulocytes of $\geq 10\%$ (59), circulating blast count of $\geq 3\%$ (69), advanced age (61, 69, 72), male sex (69), and cytogenetic abnormalities other than 13q- or 20q- (66, 73, 74).

2.8 Management

Unfortunately, current therapy for PMF is inadequate and often palliative at best. Among the several treatment modalities that are currently employed, allogeneic stem cell transplantation (ASCT) is the only one with a potential for prolonging survival. However, ASCT is associated with substantial mortality and morbidity and is currently utilized in a select group of patients with high-risk disease. Drug therapy in PMF is used to alleviate symptomatic cytopenias, organomegaly, or marked thrombocytosis and/or leukocytosis. Other treatment modalities are also palliative and include involved field radiation, splenectomy, and blood component transfusions. Therefore, in the asymptomatic patient with low-risk PMF, it is currently reasonable to defer therapy (i.e., watchful waiting), regardless of age. In the presence of symptoms, either conventional or experimental drug therapy is advised in older patients as well as in younger low-risk patients. The risk associated with ASCT might be justified in young patients with high-risk disease and in some with intermediate-risk disease (Table 2.6). The choice between myeloablative versus reduced intensity conditioning (RIC) ASCT is made taking age and the presence of other comorbid conditions into consideration (Table 2.6).

2.8.1 Drug Therapy

The primary reason for using drug therapy in PMF is the presence of either anemia or splenomegaly that is symptomatic. Drug options for the former include subcutaneous (SC) erythropoietin (Epo) or oral drugs, including androgen preparations, corticosteroids, danazol, thalidomide, and lenalidomide. The starting dose for SC

| Table 2.5 Prognostic models in primary myelofibrosis | odels in primary | myelofibı | rosis | | | | | | |
|--|-------------------|------------|----------------|--------------|---------------------------|-------------------------|--------------------------|-----------|------------------|
| | | | Median | Score | Score | Score | Score | Score | Score |
| Prognostic | Risk | Score | survival | for Hgb | for WBC < 4 | for Plt | for AMC | for | for circulating |
| scoring system | category | sum | (months) | < 10g/dL | or $> 30 \times 10^{9}/L$ | $< 100 \times 10^{9}/L$ | $\geq 1 \times 10^{9}/L$ | symptoms* | $blasts \ge 1\%$ |
| Elliott et al. | Low | 0 | 173 | 1 | 1 | 1 | 1 | N/A | N/A |
| (139) $(n = 129)$ | Intermediate | 1 | 61 | | | | | | |
| (ages < 60 years; median: 52) | High | ≥2 | 26 | | | | | | |
| (Mayo prognostic model) | | | | | | | | | |
| Dingli et al. | Low | 0 | 155 | 1 | 1 | 1 | N/A | N/A | N/A |
| (71) (n = 160) | Intermediate | 1 | 69 | | | | | | |
| (ages < 60 years; median: 52) | High | ≥2 | 24 | | | | | | |
| Cervantes et al. | Low | 0 or 1 | 176 | 1 | N/A | N/A | N/A | 1 | 1 |
| (140) (n = 116) | High | ≥2 | 33 | | | | | | |
| (ages ≤ 55 years; median: 46) | I | | | | | | | | |
| Cervantes et al.63 | Low | 0 or 1 | 66 | 1 | N/A | N/A | N/A | 1 | 1 |
| (n = 106) (all ages; | High | ≥2 | 21 | | | | | | |
| median: 64 years) | | | | | | | | | |
| Dupriez et al.62 | Low | 0 | 93 | 1 | 1 | N/A | N/A | N/A | N/A |
| (n = 195) (all ages; | Intermediate | 1 | 26 | | | | | | |
| median: 65 years) | High | 2 | 13 | | | | | | |
| Hgb, hemoglobin; WBC, white blood cell count; Plt, platelet count; AMC, absolute monocyte count. | , white blood cel | l count; P | lt, platelet c | ount; AMC, a | bsolute monocyte c | ount. | | | |

| Risk stratification | Age <45 years | Age 45-60 years | Age >60 years |
|---------------------------------|--------------------|------------------|------------------|
| Low risk | Watchful waiting | Watchful waiting | Watchful waiting |
| (no risk factors) ^a | or | or | or |
| | Experimental | Experimental | Experimental |
| | drug therapy | drug therapy | drug therapy |
| Intermediate risk | Experimental | Experimental | Experimental |
| | drug therapy | drug therapy | drug therapy |
| (one risk factor) | or | | |
| | RIC ASCT | | |
| High risk | Myeloablative ASCT | RIC ASCT | Experimental |
| $(\geq 2 \text{ risk factors})$ | • | | drug therapy |

 Table 2.6
 Suggested treatment algorithm in primary myelofibrosis

RIC, reduced-intensity conditioning; ASCT, allogeneic stem cell transplant.

^aAccording to Mayo prognostic scoring system; hemoglobin <10g/dL, platelet count < 100×10^{9} /L, monocyte count ≥ 1 × 10⁹/L, leukocyte count > 30 × 10⁹/L or < 4 × 10⁹.

Epo injection is 40,000 units weekly and such therapy is most appropriate in the presence of an endogenous serum Epo level below 100 U/L, where an approximately 50% response rate is expected (75). Some patients under Epo therapy experience further enlargement of their spleen. Several androgen preparations, including testosterone enanthate (400–600 mg IM weekly) and oral fluoxymesterone (10 mg TID) have been shown to improve anemia in a third of treated patients (76). The response rate from androgen therapy is improved by the concomitant use of corticosteroids (e.g., prednisone 30 mg/day) and compromised by the presence of cytogenetic abnormalities (76, 77). Danazol (600 mg/day) is a synthetically modified testosterone and produces response rates in PMF that is similar to that seen with other androgen preparations (78).

Thalidomide and lenalidomide have recently been shown to have therapeutic activity in PMF (79, 80). The mechanism of action for both drugs is not clearly understood but believed to be related to their anticytokine and immunomodulatory properties. The anticytokine treatment approach in PMF is based on both circumstantial evidence from affected patients and experimental myelofibrosis in mice. Thalidomide displays both antiangiogenic (81) and anti-tumor necrosis factor $(TNF)-\alpha$ (82) activity. There are currently two classes of thalidomide analogs: the selective cytokine inhibitory drugs (SelCIDs) and the immunomodulatory drugs (ImiDs) (83). Like thalidomide, both drug classes have anti-TNF- α , antiangiogenic, and anti-inflammatory activity (84). The activity of SelCIDs is mostly tied to phosphodiesterase 4 inhibition. The ImiDs, including CC-5013 (lenalidomide[™]) and CC-4047 (actimidTM), do not inhibit phosphodiesterase 4 and have a broader cytokine inhibitory activity [inhibit TNF- α , interleukin (IL)-1 β , IL-6, and IL-12]. In addition, they costimulate T-cells with upregulation of IL-2 and interferon (IFN)-y production by T helper-1 cells and IL-5 and IL-10 production by T helper-2 cells (85). Lenalidomide (CC-5013) is the lead compound among the ImiDs and its ex vivo antiangiogenic as well as anti-TNF property is estimated to be at least 50-fold higher than that of thalidomide (83, 84). In PMF, thalidomide works best at low doses (50 mg/day) and in combination with corticosteroids (prednisone 15-30 mg/day) (86) and lenalidomide in the presence of del(5)(q31) (80). Singleagent therapy in unselected patients with either thalidomide or lenalidomide produces 15% and 20% response rates in anemia, respectively. The addition of corticosteroids doubles the response rate with thalidomide and the presence of del(5)(q31) is associated with complete hematologic remission in the majority of patients treated with lenalidomide. In addition, both drugs have been shown to improve thrombocytopenia (approximately 50% response rates) and splenomegaly (approximately 30% response rate) (79, 80).

The drug of choice for symptomatic splenomegaly in PMF is hydroxyurea (starting dose 500 mg TID). The drug is also used for controlling symptomatic thrombocytosis and/or leukocytosis. Hydroxyurea-refractory cases are sometimes managed by the use of alternative myelosuppressive agents, including intravenous cladribine (5 mg/m²/day in a 2-h infusion for 5 consecutive days to be repeated monthly for four to six cycles) (87), oral melphalan (2.5 mg three times a week) (88), and oral busulfan (2–6 mg/day with close monitoring of blood counts) (89, 90). In contrast, interferon- α therapy is poorly tolerated and has limited efficacy in the treatment of PMF (91–96).

It is expected that all clinicians disclose the side effects of the above-mentioned drugs before prescribing them. In addition, one must always look out for the presence of contraindications to the use of these drugs. For example, androgen use requires monitoring of serum prostate-specific antigen in men, liver function tests in both men and women, and underscoring the possibility of masculinizing side effects in women. Similarly, the use of thalidomide requires strict supervision and any possibility of pregnancy during its use must be prevented. Other side effects of thalidomide include somnolence, constipation, rash, and neuropathy. Lenalidomide is myelosuppressive and can result in neutropenic fever and sepsis. Therefore, one has to follow CBC closely and intervene with myeloid growth factors if the absolute neutrophil count drops to below 1×10^{9} /L. Other notable side effects of drugs used in PMF include mucocutaneous ulcers and skin/nail pigmentations associated with hydroxyurea use and the usual complications of corticosteroid use.

2.8.2 Splenectomy

Splenectomy is a strictly palliative treatment modality in PMF and does not alter the natural history of the disease. The procedure is associated with approximately 10% mortality and a higher incidence of morbidity that includes thrombosis, bleeding, postsplenectomy enlargement of the liver, and exacerbation of thrombocytosis/ leukocytosis. Current indications for splenectomy in PMF include complications of portal hypertension, including ascites and variceal bleeding, drug-refractory symptomatic splenomegaly, or very frequent red blood cell transfusions (97). Severe thrombocytopenia in PMF is a marker of impending leukemic transformation and overall outcome in its presence might not be favorably affected with splenectomy.

In preparation for splenectomy, prophylactic therapy with hydroxyurea is advised in patients with leukocyte count of $>5 \times 10^{9}$ /L and/or platelet count $>150 \times 10^{9}$ /L in order to prevent postsplenectomy thrombocytosis and/or leukocytosis that might facilitate thrombotic complications (97). In addition, there is some evidence that suggests an increased incidence of bleeding in patients displaying laboratory evidence of DIC (i.e., presence of markedly increased d-dimer). Anecdotal evidence supports the use of low-dose prednisone (20 mg/day) in preparation for surgery. In addition, short-term (4–8 weeks) systemic anticoagulation, once hemostasis is secured after surgery, might reduce the risk of postoperative thrombotic complications.

2.8.3 Radiation Therapy

Involved field radiotherapy provides transient (median response duration of 3–6 months) relief of mechanical discomfort from hepatosplenomegaly (98, 99). However, such therapy is often complicated by protracted pancytopenia and drug therapy is instead preferred. In contrast, irradiation therapy is very useful in patients with nonhepatosplenic EMH; most frequent sites include vertebral column, lungs, pleura, and peritoneum. When symptomatic, nonhepatosplenic EMH is effectively treated with low-dose radiation therapy (0.1–1 Gy in 5–10 fractions) (58). Sometimes, occult pulmonary EMH presents with "idiopathic" pulmonary hypertension and a technetium 99m sulfur colloid scintigraphy is recommended if such an occurrence is suspected and treatment with single-fraction (0.1 Gy) whole-lung irradiation has been shown to be effective (68, 100).

2.8.4 Allogeneic Stem Cell Transplant

An increasing amount of information is being gathered regarding the use of ASCT in PMF, in the context of both myeloablative (101–104) and RIC (105, 106) transplant. Early engraftment rate is acceptable in both instances regardless of whether a related or matched unrelated donor is used. The experience so far with myeloablative ASCT is encouraging in very young patients (age <45 years), but posttransplant long-term survival in older patients is less than 20% (103, 104). Furthermore, the majority of survivors after ASCT experience reduced quality of life because of chronic graft versus host disease (GVHD) (101). A recent multivariable analysis involving 320 patients with PMF registered to an international transplant database identified young age, HLA-matched sibling transplant, excellent performance status, absence of circulating blasts, and more recent transplant date as independent indicators of favorable transplant outcome (107). To date, the advantage of RIC transplant over myeloablative ASCT has not been examined in a controlled setting, although single-cohort studies suggest better outcome in terms of both 1-year mortality (0–33%) and morbidity (0–50% rate of acute GVHD) (108).

2.9 Conclusions

Over the last two decades, many drugs have been investigated for their therapeutic value in PMF. Negative studies have included drugs such as IFN- α , anagrelide, suramin, pirfenidone, imatinib mesylate, farnesyl transferase inhibitors such as R115777, and certain VEGF receptor inhibitors, including PTK-787 and SU5416 (109). In contrast, promising results were obtained with cladribine (87), etanercept (110), thalidomide (111), and lenalidomide (80). Despite such progress, treatment in PMF remains suboptimal in terms of both survival and quality of life. At present, it is reasonable to consider all high-risk patients for either ASCT (if transplant-eligible) or experimental drug therapy. It is equally reasonable to undertake a "watchful waiting" approach in low-risk patients. Management in intermediate-risk patients should be individualized and is often dictated by age, performance status, and patient preference. In all patients, the presence of del(5)(q31–32) warrants a therapeutic trial with lenalidomide. The recent discovery of PMF-associated activating mutations involving *JAK2* and *MPL* has raised the prospect of small molecule drug therapy that targets JAK2.

References

- 1. Tefferi A. Myelofibrosis with myeloid metaplasia. N Engl J Med 2000;342:1255-1265.
- Thiele J, Vardiman JW, Pierre R, et al. Chronic idiopathic myelofibrosis. In: Jaffe ES, Harris NL, Stein H, et al., editors. World Health Organization classification of tumors: Tumours of the haematopoietic and lymphoid tissues. Lyon: International Agency for Research on Cancer (IARC) Press; 2001. p. 35–38.
- 3. Mesa R, Verstovsek S, Cervantes F, et al. Primary myelofibrosis (PMF), post polycythemia vera myelofibrosis (post-PV MF), post essential thrombocythemia myelofibrosis (post-ET MF), blast phase PMF (PMF-BP): Consensus on terminology by the International Working Group for Myelofibrosis Research and Treatment (IWG-MRT). Leuk Res 2007; in press.
- Heuck G. Zwei Falle von Leukamie mit eigenthumlichem Blut- resp. Knochenmarksbefund [Two cases of leukemia with peculiar blood and bone marrow findings, respectively]. Arch Pathol Anat Physiol Virchows 1879;78:475–496.
- 5. Assmann H. Beitrage zur osteosklerotischen anamie. Beitr Pathol Anat Allgemeinen Pathol (Jena) 1907;41:565–595.
- 6. Hirsch R. Generalized osteosclerosis with chronic polycythemia vera. Arch Pathol 1935;19:91–97.
- 7. Donhauser J. The human spleen as an haematoplastic organ, as exemplified in a case of splenomegaly with sclerosis of the bone-marrow. J Exp Med 1908;10:559–574.
- Askanazy M. Ueber extrauterine Bildung von Blutzellen in der Leber. Verh Dtsch Pathol Ges 1904;7:58–65.
- 9. Vaughan JM HC. Leuco-erythrobalstic anaemia and myelosclerosis. J Pathol Bacteriol 1939;48:339–352.
- Jackson H Jr, PFJ, Lemon HM. Agnogenic myeloid metaplasia of the spleen: a syndrome simulating other more definite hematological disorders. N Engl J Med 1940;222:985–994.
- 11. Dameshek W. Some speculations on the myeloproliferative syndromes. Blood 1951;6:372-375.
- Nowell PC, Hungerford DA. A minute chromosome in human chronic granulocytic leukemia. J Nat Cancer Inst 1960;25:85.

- Rowley JD. Letter: A new consistent chromosomal abnormality in chronic myelogenous leukaemia identified by quinacrine fluorescence and Giemsa staining. Nature 1973;243:290–293.
- 14. Daley GQ, Van Etten RA, Baltimore D. Induction of chronic myelogenous leukemia in mice by the P210bcr/abl gene of the Philadelphia chromosome. Science 1990;247:824–830.
- 15. Tefferi A, Gilliland DG. Classification of myeloproliferative disorders: From Dameshek towards a semi-molecular system. Best Pract Res Clin Haematol 2005; in press.
- Jacobson RJ, Salo A, Fialkow PJ. Agnogenic myeloid metaplasia: a clonal proliferation of hematopoietic stem cells with secondary myelofibrosis. Blood 1978;51:189–194.
- James C, Ugo V, Le Couedic JP, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. Nature 2005;434:1144–1148.
- Pikman Y, Lee BH, Mercher T, et al. MPLW515L is a novel somatic activating mutation in myelofibrosis with myeloid metaplasia. PLoS Med 2006;3:e270.
- 19. Wasserman LR. The treatment of polycythemia. A panel discussion. Blood 1968;32:483-487.
- Vardiman JW, Harris NL, Brunning RD. The World Health Organization (WHO) classification of the myeloid neoplasms. Blood 2002;100:2292–2302.
- 21. Tefferi A, Gilliland DG. Classification of myeloproliferative disorders: From Dameshek towards a semi-molecular system. Best Pract Res Clin Haematol 2006;19:361–364.
- 22. Mesa RA, Silverstein MN, Jacobsen SJ, et al. Population-based incidence and survival figures in essential thrombocythemia and agnogenic myeloid metaplasia: An Olmsted County study, 1976–1995. Am J Hematol 1999;61:10–15.
- 23. Ridell B, Carneskog J, Wedel H, et al. Incidence of chronic myeloproliferative disorders in the city of Goteborg, Sweden 1983–1992. Eur J Haematol 2000;65:267–271.
- Woodliff HJ, Dougan L. Myelofibrosis in Western Australia: an epidemiological study of 29 cases. Med J Aust 1976;1:523–525.
- Heudes D, Carli PM, Bailly F, et al.. Myeloproliferative disorders in the department of Cote d'Or between 1980 and 1986. Nouv Rev Francaise Hematol 1989;31:375–378.
- McNally RJ, Rowland D, Roman E, et al. Age and sex distributions of hematological malignancies in the U.K. Hematol Oncol 1997;15:173–189.
- 27. Chaiter Y, Brenner B, Aghai E, et al. High incidence of myeloproliferative disorders in Ashkenazi Jews in northern Israel. Leuk Lymphoma 1992;7:251–255.
- Cervantes F, Barosi G, Hernandez-Boluda JC, et al. Myelofibrosis with myeloid metaplasia in adult individuals 30 years old or younger: presenting features, evolution and survival. Eur J Haematol 2001;66:324–327.
- 29. Sekhar M, Prentice HG, Popat U, et al. Idiopathic myelofibrosis in children. Br J Haematol 1996;93:394–397.
- Altura RA, Head DR, Wang WC. Long-term survival of infants with idiopathic myelofibrosis. Br J Haematol 2000;109:459–462.
- Tondel M, Persson B, Carstensen J. Myelofibrosis and benzene exposure. Occup Med (Lond) 1995;45:51–52.
- 32. Honda Y, Delzell E, Cole P. An updated study of mortality among workers at a petroleum manufacturing plant. J Occup Environ Med 1995;37:194–200.
- Mueller K. [Panmyelopathy and myelofibrosis after therapy with thorium X (peteosthor)]. Med Monatsschr 1960;14:241–243.
- 34. Bastrup-Madsen P, Jensen BN. Myelofibrosis with myeloid metaplasia and pancytopenia after thorotrast injection. Acta Med Scand 1971;189:355–358.
- 35. Anderson RE, Hoshino T, Yamamoto T. Myelofibrosis with myeloid metaplasia in survivors of the atomic bomb in Hiroshima. Ann Intern Med 1964;60:1–18.
- 36. Reeder TL, Bailey RJ, Dewald GW, et al. Both B and T lymphocytes may be clonally involved in myelofibrosis with myeloid metaplasia. Blood 2003;101:1981–1983.
- Baxter EJ, Scott LM, Campbell PJ, et al. Acquired mutation of the tyrosine kinase JAK2 in human myeloproliferative disorders. Lancet 2005;365:1054–1061.
- Levine RL, Wadleigh M, Cools J, et al. Activating mutation in the tyrosine kinase JAK2 in polycythemia vera, essential thrombocythemia, and myeloid metaplasia with myelofibrosis. Cancer Cell 2005;7:387–397.

- Kralovics R, Passamonti F, Buser AS, et al. A gain of function mutation in Jak2 is frequently found in patients with myeloproliferative disorders. New Engl J Med 2005;352:1779–1790.
- Pardanani AD, Levine RL, Lasho T, et al. MPL515 mutations in myeloproliferative and other myeloid disorders: a study of 1182 patients. Blood First Edition Paper, prepublished online July 25, 2006; doi 101182/blood-2006-04-018879. 2006.
- 41. Wernig G, Mercher T, Okabe R, Levine RL, Lee BH, Gilliland DG. Expression of Jak2V617F causes a polycythemia vera-like disease with associated myelofibrosis in a murine bone marrow transplant model. Blood. 2006;107:4274–4281.
- 42. Lacout C, Pisani DF, Tulliez M, et al. JAK2V617F expression in murine hematopoietic cells leads to MPD mimicking human PV with secondary myelofibrosis. Blood First Edition Paper, prepublished online May 2, 2006; doi 101182/blood-2006-02-002030. 2006.
- 43. Pikman Y, Lee BH, Mercher T, et al. MPLW515L is a novel somatic activating mutation in myelofibrosis with myeloid metaplasia. PLoS. 2006; in press.
- 44. Schmitt A, Jouault H, Guichard J, et al. Pathologic interaction between megakaryocytes and polymorphonuclear leukocytes in myelofibrosis. Blood 2000;96:1342–1347.
- 45. Xu M, Bruno E, Chao J, et al. Constitutive mobilization of CD34+ cells into the peripheral blood in idiopathic myelofibrosis may be due to the action of a number of proteases. Blood 2005;105:4508–4515.
- Chagraoui H, Komura E, Tulliez M, et al. Prominent role of TGF-beta 1 in thrombopoietininduced myelofibrosis in mice. Blood 2002;100:3495–3503.
- 47. Chagraoui H, Tulliez M, Smayra T, et al. Stimulation of osteoprotegerin production is responsible for osteosclerosis in mice overexpressing TPO. Blood 2003;101:2983–2989.
- 48. Vannucchi AM, Bianchi L, Cellai C, et al. Development of myelofibrosis in mice genetically impaired for GATA-1 expression (GATA-1(low) mice). Blood 2002;100:1123–1132.
- Massa M, Rosti V, Ramajoli I, et al. Circulating CD34+, CD133+, and vascular endothelial growth factor receptor 2-positive endothelial progenitor cells in myelofibrosis with myeloid metaplasia. J Clin Oncol 2005;23:5688–5695.
- Barosi G, Viarengo G, Pecci A, et al. Diagnostic and clinical relevance of the number of circulating CD34(+) cells in myelofibrosis with myeloid metaplasia. Blood 2001;98:3249–3255.
- 51. Tefferi A. New insights into the pathogenesis and drug treatment of myelofibrosis. Curr Opin Hematol 2006;13:87–92.
- 52. Tefferi A. Pathogenesis of myelofibrosis with myeloid metaplasia. J Clin Oncol. 2005;23:8520-8530.
- 53. Ward HP, Block MH. The natural history of agnogenic myeloid metaplasia (AMM) and a critical evaluation of its relationship with the myeloproliferative syndrome. Medicine 1971;50:357–420.
- 54. Cervantes F, Alvarez-Larran A, Arellano-Rodrigo E, et al. Frequency and risk factors for thrombosis in idiopathic myelofibrosis: analysis in a series of 155 patients from a single institution. Leukemia 2006;20:55–60.
- 55. Jaroch MT, Broughan TA, Hermann RE. The natural history of splenic infarction. Surgery 1986;100:743–750.
- 56. Cervantes F, Pereira A, Esteve J, et al. The changing profile of idiopathic myelofibrosis: a comparison of the presenting features of patients diagnosed in two different decades. European Journal of Haematology 1998;60:101–105.
- 57. Mesa RA, Li CY, Schroeder G, et al. Clinical correlates of splenic histopathology and splenic karyotype in myelofibrosis with myeloid metaplasia. Blood 2001;97:3665–3667.
- Koch CA, Li CY, Mesa RA, et al. Nonhepatosplenic extramedullary hematopoiesis: associated diseases, pathology, clinical course, and treatment. Mayo Clinic Proc 2003;78: 1223–1233.
- 59. Visani G, Finelli C, Castelli U, et al. Myelofibrosis with myeloid metaplasia: clinical and haematological parameters predicting survival in a series of 133 patients. Br J Haematol 1990;75:4–9.
- 60. Rupoli S, Da Lio L, Sisti S, et al. Primary myelofibrosis: a detailed statistical analysis of the clinicopathological variables influencing survival. Ann Hematol 1994;68:205–212.

- 61. Strasser-Weippl K, Steurer M, Kees M, et al. Age and hemoglobin level emerge as most important clinical prognostic parameters in patients with osteomyelofibrosis: introduction of a simplified prognostic score. Leuk Lymphoma 2006;47:441–450.
- 62. Dupriez B, Morel P, Demory JL, et al. Prognostic factors in agnogenic myeloid metaplasia: a report on 195 cases with a new scoring system. Blood 1996;88:1013–1018.
- 63. Cervantes F, Pereira A, Esteve J, et al. Identification of 'short-lived' and 'long-lived' patients at presentation of idiopathic myelofibrosis. Br J Haematol 1997;97:635–640.
- 64. Thiele J, Kvasnicka HM. Hematopathologic findings in chronic idiopathic myelofibrosis. Semin Oncol 2005;32:380–394.
- 65. Thiele J, Kvasnicka HM. A critical reappraisal of the WHO classification of the chronic myeloproliferative disorders. Leuk Lymphoma 2006;47:381–396.
- 66. Tefferi A, Mesa RA, Schroeder G, et al. Cytogenetic findings and their clinical relevance in myelofibrosis with myeloid metaplasia. Br J Haematol 2001;113:763–771.
- 67. Dingli D, Grand FH, Mahaffey V, et al. Der(6)t(1;6)(q21–23;p21.3): a specific cytogenetic abnormality in myelofibrosis with myeloid metaplasia. Br J Haematol 2005;130:229–232.
- Dingli D, Utz JP, Krowka MJ, et al. Unexplained pulmonary hypertension in chronic myeloproliferative disorders. Chest 2001;120:801–808.
- 69. Okamura T, Kinukawa N, Niho Y, et al. Primary chronic myelofibrosis: clinical and prognostic evaluation in 336 Japanese patients. Int J Hematol 2001;73:194–198.
- Mesa RA, Tefferi A. Survival and outcomes to therapy in leukemic transformation of myelofibrosis with myeloid metaplasia; a single institution experience with 91 patients. Blood 2003;102:917a–918a.
- 71. Dingli D, Schwager SM, Mesa RA, et al. Prognosis in transplant-eligible patients with agnogenic myeloid metaplasia: A simple CBC-based scoring system. Cancer 2005;in press.,
- Kvasnicka HM, Thiele J, Werden C, et al. Prognostic factors in idiopathic (primary) osteomyelofibrosis. Cancer 1997;80:708–719.
- 73. Reilly JT, Snowden JA, Spearing RL, et al. Cytogenetic abnormalities and their prognostic significance in idiopathic myelofibrosis: a study of 106 cases. Br J Haematol 1997;98:96–102.
- Tefferi A, Dingli D, Li CY, et al. Prognostic diversity among cytogenetic abnormalities in myelofibrosis with myeloid metaplasia. Cancer 2005;104:1656–1660.
- Cervantes F, Alvarez-Larran A, Hernandez-Boluda JC, et al.. Erythropoietin treatment of the anaemia of myelofibrosis with myeloid metaplasia: results in 20 patients and review of the literature. Br J Haematol 2004;127:399–403.
- 76. Silverstein MN. Agnogenic myeloid metaplasia. Acton, MA Publishing Science Group; 1975, p. 126.
- 77. Besa EC, Nowell PC, Geller NL, et al. Analysis of the androgen response of 23 patients with agnogenic myeloid metaplasia: the value of chromosomal studies in predicting response and survival. Cancer 1982;49:308–313.
- 78. Cervantes F, Alvarez-Larran A, Domingo A, et al. Efficacy and tolerability of danazol as a treatment for the anaemia of myelofibrosis with myeloid metaplasia: long-term results in 30 patients. Br J Haematol 2005;129:771–775.
- Elliott MA, Mesa RA, Li CY, et al. Thalidomide treatment in myelofibrosis with myeloid metaplasia. Br J Haematol 2002;117:288–296.
- Tefferi A, Cortes J, Verstovsek S, et al. Lenalidomide therapy in myelofibrosis with myeloid metaplasia. Blood 2006;108:1158–1164.
- D'Amato RJ, Loughnan MS, Flynn E, et al. Thalidomide is an inhibitor of angiogenesis. Proc Natl Acad Sci USA 1994;91:4082–4085.
- Moreira AL, Sampaio EP, Zmuidzinas A, et al. Thalidomide exerts its inhibitory action on tumor necrosis factor alpha by enhancing mRNA degradation. J Exp Med 1993;177: 1675–1680.
- Corral LG, Haslett PA, Muller GW, et al. Differential cytokine modulation and T cell activation by two distinct classes of thalidomide analogues that are potent inhibitors of TNF- alpha. J Immunol 1999;163:380–386.

- Dredge K, Marriott JB, Macdonald CD, et al. Novel thalidomide analogues display antiangiogenic activity independently of immunomodulatory effects. Br J Cancer 2002;87: 1166–1172.
- Schafer PH, Gandhi AK, Loveland MA, et al. Enhancement of cytokine production and AP-1 transcriptional activity in T cells by thalidomide-related immunomodulatory drugs. J Pharmacol Exp Ther 2003;20:20.
- 86. Mesa RA, Steensma DP, Pardanani A, et al. A phase 2 trial of combination low-dose thalidomide and prednisone for the treatment of myelofibrosis with myeloid metaplasia. Blood 2003;101:2534–2541.
- Tefferi A, Silverstein MN, Li CY. 2-Chlorodeoxyadenosine treatment after splenectomy in patients who have myelofibrosis with myeloid metaplasia. Br J Haematol 1997;99: 352–357.
- Petti MC, Latagliata R, Spadea T, et al. Melphalan treatment in patients with myelofibrosis with myeloid metaplasia. Br J Haematol 2002;116:576–581.
- 89. Shojania AM. Reversion of post polycythemia vera (PV) myelofibrosis (MF) to PV following busulfan therapy. Blood 2002;100:343b–343b.
- Naqvi T, Baumann MA. Myelofibrosis: response to busulfan after hydroxyurea failure. Int J Clin Pract 2002;56:312–313.
- Parmeggiani L, Ferrant A, Rodhain J, et al. Alpha interferon in the treatment of symptomatic myelofibrosis with myeloid metaplasia. Eur J Haematol 1987;39:228–232.
- 92. Seewann HL, Gastl G, Lang A, et al. Interferon-alpha-2 in the treatment of idiopathic myelofibrosis. Blut 1988;56:161–163.
- 93. Barosi G, Liberato LN, Costa A, et al. Cytoreductive effect of recombinant alpha interferon in patients with myelofibrosis with myeloid metaplasia. Blut 1989;58:271–274.
- 94. Barosi G, Liberato LN, Costa A, et al. Induction and maintenance alpha-interferon therapy in myelofibrosis with myeloid metaplasia. Eur J Haematol 1990;52(Suppl):12–14.
- 95. Gilbert HS. Long term treatment of myeloproliferative disease with interferon-alpha-2b feasibility and efficacy. Cancer 1998;83:1205–1213.
- Tefferi A, Elliot MA, Yoon SY, et al. Clinical and bone marrow effects of interferon alfa therapy in myelofibrosis with myeloid metaplasia. Blood 2001;97:1896.
- Tefferi A, Mesa RA, Nagorney DM, et al. Splenectomy in myelofibrosis with myeloid metaplasia: a single-institution experience with 223 patients. Blood 2000;95:2226–2233.
- Elliott MA, Chen MG, Silverstein MN, et al. Splenic irradiation for symptomatic splenomegaly associated with myelofibrosis with myeloid metaplasia. Br J Haematol. 1998;103: 505–511.
- 99. Tefferi A, Jimenez T, Gray LA, et al. Radiation therapy for symptomatic hepatomegaly in myelofibrosis with myeloid metaplasia. Eur J Haematol 2001;66:37–42.
- 100. Steensma DP, Hook CC, Stafford SL, et al. Low-dose, single-fraction, whole-lung radiotherapy for pulmonary hypertension associated with myelofibrosis with myeloid metaplasia. Br J Haematol 2002;118:813–816.
- 101. Deeg HJ, Gooley TA, Flowers ME, et al. Allogeneic hematopoietic stem cell transplantation for myelofibrosis. Blood 2003;102:3912–3918.
- 102. Ditschkowski M, Beelen DW, Trenschel R, et al. Outcome of allogeneic stem cell transplantation in patients with myelofibrosis. Bone Marrow Transplant 2004;34:807–813.
- 103. Daly A, Song K, Nevill T, et al. Stem cell transplantation for myelofibrosis: a report from two Canadian centers. Bone Marrow Transplant 2003;32:35–40.
- 104. Guardiola P, Anderson JE, Gluckman E. Myelofibrosis with myeloid metaplasia. N Engl J Med 2000;343:659; discussion 659–660.
- 105. Kroger N, Zabelina T, Schieder H, et al. Pilot study of reduced-intensity conditioning followed by allogeneic stem cell transplantation from related and unrelated donors in patients with myelofibrosis. Br J Haematol 2005;128:690–697.
- 106. Rondelli D, Barosi G, Bacigalupo A, et al. Allogeneic hematopoietic stem-cell transplantation with reduced-intensity conditioning in intermediate- or high-risk patients with myelofibrosis with myeloid metaplasia. Blood 2005;105:4115–4119.

- Ballen K, Sobocinski KA, Zhang MJ, et al. Outcome of bone marrow transplantation for myelofibrosis. Blood 2005;106:53a–53a.
- Arana-Yi C, Quintas-Cardama A, Giles F, et al. Advances in the therapy of chronic idiopathic myelofibrosis. Oncologist 2006;11:929–943.
- Hennessy BT, Thomas DA, Giles FJ, et al. New approaches in the treatment of myelofibrosis. Cancer 2005;103:32–43.
- 110. Steensma DP, Mesa RA, Li CY, et al. Etanercept, a soluble tumor necrosis factor receptor, palliates constitutional symptoms in patients with myelofibrosis with myeloid metaplasia: results of a pilot study. Blood 2002;99:2252–2254.
- 111. Tefferi A, Elliot MA. Serious myeloproliferative reactions associated with the use of thalidomide in myelofibrosis with myeloid metaplasia. Blood 2000;96:4007.
- 112. Buesche G, Georgii A, Duensing A, et al. Evaluating the volume ratio of bone marrow affected by fibrosis: a parameter crucial for the prognostic significance of marrow fibrosis in chronic myeloid leukemia. Hum Pathol 2003;34:391–401.
- 113. Steensma DP, Hanson CA, et al. Myelodysplasia with fibrosis: a distinct entity? Leuk Res 2001;25:829–838.
- 114. Tefferi A, Hoagland HC, Therneau TM, et al. Chronic myelomonocytic leukemia: natural history and prognostic determinants. Mayo Clin Proc 1989;64:1246–1254.
- 115. Michel G, Thuret I, Capodano AM, et al. Myelofibrosis in a child suffering from a hypereosinophilic syndrome with trisomy 8: response to corticotherapy. Med Pediatr Oncol 1991;19:62–65.
- 116. Baek JY, Li CY, Pardanani A, et al. Bone marrow angiogenesis in systemic mast cell disease. J Hematother Stem Cell Res 2002;11:139–146.
- 117. Ruiz-Arguelles GJ, Marin-Lopez A, Lobato-Mendizabal E, et al. Acute megakaryoblastic leukaemia: a prospective study of its identification and treatment. Br J Haematol 1986;62: 55–63.
- 118. Mori A, Wada H, Okada M, et al. Acute promyelocytic leukemia with marrow fibrosis at initial presentation: possible involvement of transforming growth factor-beta(1). Acta Haematol 2000;103:220–223.
- Wallis JP, Reid MM. Bone marrow fibrosis in childhood acute lymphoblastic leukaemia. J Clin Pathol 1989;42:1253–1254.
- 120. Thiele J, Krech R, Vykoupil KF, Georgii A. Malignant (acute) myelosclerosis: a clinical and pathological study in 6 patients. Scand J Haematol 1984;33:95–109.
- 121. Hasselbalch H. Idiopathic myelofibrosis: a clinical study of 80 patients. Am J Hematol 1990;34:291–300.
- 122. Shehata M, Schwarzmeier JD, Hilgarth M, et al. TGF-beta1 induces bone marrow reticulin fibrosis in hairy cell leukemia. J Clin Invest 2004;113:676–685.
- 123. Meadows LM, Rosse WR, Moore JO, et al. Hodgkin's disease presenting as myelofibrosis. Cancer 1989;64:1720–1726.
- 124. Matsunaga T, Takemoto N, Miyajima N, et al. Splenic marginal zone lymphoma presenting as myelofibrosis associated with bone marrow involvement of lymphoma cells which secrete a large amount of TGF-beta. Ann Hematol 2004;83:322–325.
- 125. Meerkin D, Ashkenazi Y, Gottschalk-Sabag S, et al. Plasma cell dyscrasia with marrow fibrosis. A reversible syndrome mimicking agnogenic myeloid metaplasia. Cancer 1994;73: 625–628.
- 126. Kiely JM, Silverstein MN. Metastatic carcinoma simulating agnogenic myeloid metaplasia and myelofibrosis. Cancer 1969;24:1041–1044.
- 127. Paquette RL, Meshkinpour A, Rosen PJ. Autoimmune myelofibrosis. A steroid-responsive cause of bone marrow fibrosis associated with systemic lupus erythematosus. Medicine 1994;73:145–152.
- 128. Inoue Y, Matsubara A, Okuya S, et al. Myelofibrosis and systemic lupus erythematosus: reversal of fibrosis with high-dose corticosteroid therapy. Acta Haematol. 1992;88:32–36.
- 129. Rocha Filho FD, Ferreira FV, Mendes FdO, et al. Bone marrow fibrosis (pseudo-myelofibrosis) in human kala-azar. Rev Soc Bras Med Trop 2000;33:363–366.

- Viallard JF, Parrens M, Boiron JM, et al. Reversible myelofibrosis induced by tuberculosis. Clin Infect Dis 2002;34:1641–1643.
- 131. Murrin RJ, Harrison P. Abnormal osteoclasts and bone marrow fibrosis in Paget's disease of the bone. Br J Haematol 2004;124:3.
- Sitalakshmi S, Srikrishna A, Damodar P. Haematological changes in HIV infection. Indian J Pathol Microbiol 2003;46:180–183.
- Stephan JL, Galambrun C, Dutour A, et al. Myelofibrosis: an unusual presentation of vitamin D-deficient rickets. Eur J Pediatr 1999;158:828–829.
- 134. Nomura S, Ogawa Y, Osawa G, et al. Myelofibrosis secondary to renal osteodystrophy. Nephron 1996;72:683–687.
- Kumbasar B, Taylan I, Kazancioglu R, et al. Myelofibrosis secondary to hyperparathyroidism. Exp Clin Endocrinol Diabetes 2004;112:127–130.
- 136. Falik-Zaccai TC, Anikster Y, Rivera CE, et al. A new genetic isolate of gray platelet syndrome (GPS): clinical, cellular, and hematologic characteristics. Mol Genet Metab 2001;74: 303–313.
- 137. Sheikha A. Fatal familial infantile myelofibrosis. J Pediatr Hematol Oncol 2004;26:164-168.
- 138. Popat U, Frost A, Liu E, et al. New onset of myelofibrosis in association with pulmonary arterial hypertension. Ann Intern Med 2005;143:466–467.
- 139. Elliott M, Dingli D, Schwager S, et al. Absolute monocyte count is an independent prognostic factor for survival in agnogenic myeloid metaplasia. Blood 2006: Abstract.
- 140. Cervantes F, Barosi G, Demory JL, et al. Myelofibrosis with myeloid metaplasia in young individuals: disease characteristics, prognostic factors and identification of risk groups. Br J Haematol 1998;102:684–690.