Perinatal Lesioning and Lifelong Effects of the Noradrenergic Neurotoxin 6-Hydroxydopa

Richard M. Kostrzewa

Abstract 6-hydroxydopa (6-OHDOPA) was synthesized with the expectation that it would be able to cross the blood–brain barrier to be enzymatically decarboxylated to 6-hydroxydopamine (6-OHDA), the newly discovered neurotoxin for noradrenergic and dopaminergic neurons. In part, 6-OHDOPA fulfilled these criteria. When administered experimentally to rodents, 6-OHDOPA destroyed peripheral sympathetic noradrenergic nerves and did exert neurotoxicity to noradrenergic nerves in brain—in large part, from its conversion to 6-OHDA. However, the efficacy of 6-OHDOPA was less than that of 6-OHDA; also, 6-OHDOPA was relatively selective for noradrenergic neurons; near-lethal doses of 6-OHDOPA were required to damage dopaminergic nerves; and ultimately, 6-OHDOPA was found to be an agonist at AMPA receptors, thus accounting for more non-specificity. Nevertheless, 6-OHDOPA was found to be a particularly valuable tool in uncovering processes and mechanisms associated with noradrenergic nerve regeneration and sprouting, particularly when administered to perinatal rodents. Also, 6-OHDOPA was a good tool for selective mapping of noradrenergic nerve tracts in brain, since dopaminergic tracts were unaffected and did not interfere with the histofluorescent methodology used for this purpose in the early 1970s. As an experimental research tool, 6-OHDOPA was valuable in a short time-window, but its utility is largely limited because of newer research technologies that provide better means today for nerve tract mapping, and for experimental approaches engaged toward study of processes and mechanisms attending nerve regeneration. AMPA actions of 6-OHDOPA have not been extensively studied, so this avenue may enliven use of 6-OHDOPA in the future.

Keywords 6-hydroxydopa \cdot 6-hydroxydopamine \cdot Nerve regeneration \cdot Nerve sprouting · Noradrenergic nerves · Neuroteratogen · AMPA

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1 Introduction

The norepinephrine (NE) isomer 6-hydroxydopamine (6-OHDA) was initially found to produce long-lasting depletion of norepinephrine in heart (Porter et al. [1963,](#page-7-0) [1965\)](#page-7-0). Subsequently, 6-OHDA was shown by electron microscopy to overtly destroy noradrenergic nerves (Thoenen and Tranzer [1968a](#page-7-0), [b\)](#page-7-0) and dopaminergic nerves (Ungerstedt [1968\)](#page-7-0). One limitation of 6-OHDA, however, was its inability to cross the blood–brain barrier (BBB) (Kostrzewa and Jacobowitz [1974](#page-6-0)).

6-hydroxydopa (6-OHDOPA) was synthesized as an expected pro-toxin and, like levodopa, able to cross the BBB prior to its decarboxylation to 6-OHDA (Ong et al. [1969](#page-6-0); Berkowitz et al. [1970](#page-4-0); Evans and Cohen [1989](#page-5-0), [1993](#page-5-0)). As expected, 6-OHDOPA produced norepinephrine (NE) depletion in brain and peripheral tissues (Jonsson and Sachs [1973](#page-5-0); Kostrzewa and Jacobowitz [1972,](#page-6-0) [1973;](#page-6-0) Richardson and Jacobowitz [1973\)](#page-7-0) and subsequently was shown by loss of tyrosine hydroxylase activity and by histochemical, electronmicroscopic, and silver degeneration staining to overtly destroy noradrenergic nerves (Jacobowitz and Kostrzewa [1971;](#page-5-0) Kostrzewa and Harper [1974](#page-6-0), [1975;](#page-6-0) Sachs et al. [1973;](#page-7-0) Kostrzewa et al. [1978;](#page-5-0) Tohyama et al. [1974;](#page-7-0) Toyama et al. [1974\)](#page-7-0), with preference for noradrenergic perikarya in caudal locus coeruleus (LC) (Clark et al. [1979](#page-5-0)). Peripheral noradrenergic (i.e., sympathetic) nerves, in contrast to central noradrenergic nerves, appear to fully recover from 6-OHDOPA damage (Kostrzewa and Jacobowitz [1972](#page-6-0); Sachs and Jonsson [1972a,](#page-7-0) [b](#page-7-0)) and can be protected from damage by administering a peripherally acting dopa decarboxylase inhibitor, namely carbidopa (Kostrzewa et al. [2000](#page-6-0)).

2 Mechanism of Action of 6-OHDOPA

The destructive effects of 6-OHDOPA on noradrenergic and dopaminergic neurons are attributable to its conversion to 6-OHDA, which is known to auto-oxidize to ortho- and para-quinones, aminochromes, and hydroxyindoles (Adams et al. [1972;](#page-4-0)

Blank et al. [1972;](#page-4-0) Saner and Thoenen [1971;](#page-7-0) Senoh and Witkop [1959;](#page-7-0) Wehrli et al. [1972\)](#page-7-0)—reactive species leading to formation of intraneuronal peroxide (Heikkila and Cohen [1971,](#page-5-0) [1972a](#page-5-0), [b\)](#page-5-0), superoxide, and hydroxyl radical (Cohen and Heikkila [1974;](#page-5-0) Heikkila and Cohen [1973](#page-5-0)).

3 6-OHDOPA as a Neuroteratogen

When administered to perinatal rats, 6-OHDOPA (60 μ g/g at P0 + P2 + P4) produced lifelong alterations in noradrenergic innervation of brain (Kostrzewa [1975;](#page-5-0) Kostrzewa and Garey [1976\)](#page-6-0). The nucleus LC providing the major portion of noradrenergic innervation of dorsal brain was directly damaged, with there being loss of one-third of the approximately 1500 perikarya, and with half the numbers of cells in the caudal portion of the LC undergoing degeneration (Clark et al. [1979\)](#page-5-0). Hippocampal noradrenergic innervation was reduced by $>95\%$, and neocortex, by \sim 70 % (Kostrzewa and Harper [1974,](#page-6-0) [1975;](#page-6-0) Kostrzewa [1975](#page-5-0); Kostrzewa and Garey [1976,](#page-6-0) [1977\)](#page-6-0). As a consequence of damage to the dorsal bundle, the major ascending noradrenergic tract to forebrain, noradrenergic fibers projecting to regions near the LC per se sprouted and hyperinnervated midbrain, pons, medulla, and cerebellum (Jaim-Etcheverry and Zieher [1977](#page-5-0); Jaim-Etcheverry et al. [1975](#page-5-0); Kostrzewa and Harper [1974,](#page-6-0) [1975;](#page-6-0) Kostrzewa [1975;](#page-5-0) Kostrzewa and Garey [1976,](#page-6-0) [1977;](#page-6-0) Kostrzewa et al. [1978](#page-5-0); [1982;](#page-6-0) Zieher and Jaim-Etcheverry [1979\)](#page-7-0). This reactive sprouting resulted in as much as a twofold increase in numbers of fibers' innervation of caudal brain regions. In contrast, innervation to hypothalamus was slightly altered. The pairing of noradrenergic hypoinnervation of forebrain with noradrenergic hyperinnervation of hindbrain is replicated by knife cuts of the dorsal bundle shortly after birth, suggesting that hindbrain hyperinnervation is an outcome of forebrain noradrenergic hypoinnervation (Klisans-Fuenmayor et al. [1986](#page-5-0); Kostrzewa et al. [1988\)](#page-6-0).

This spectrum of effects is replicated (1) by single 6-OHDOPA treatment of rats at birth, also (2) by prenatal 6-OHDOPA, administered to pregnant rats at G14 or later, and (3) by prenatal 6-OHDOPA to pregnant mice at G13 or later (Jaim-Etcheverry et al. [1975](#page-5-0); Kostrzewa [2007;](#page-5-0) Kostrzewa et al. [1978;](#page-5-0) Zieher and Jaim-Etcheverry [1973](#page-7-0), [1975a](#page-7-0), [b\)](#page-7-0). When 6-OHDOPA is administered solely at P3, there is an absence of noradrenergic sprouting to cerebellum. Described perinatal 6-OHDOPA effects on noradrenergic neurons persist throughout life (Jaim-Etcheverry et al. [1975;](#page-5-0) McLean et al. [1976,](#page-6-0) [1980;](#page-6-0) Zieher and Jaim-Etcheverry [1973](#page-7-0), [1975a](#page-7-0), [b](#page-7-0)). A single 6-OHDOPA treatment at P5 fails to produce noradrenergic hyperinnervation of midbrain, while single 6-OHDOPA treatment as late as P14 still produces hyperinnervation of pons–medulla (Kostrzewa and Garey [1977\)](#page-6-0).

Agonists at µ-opioid receptors (i.e., morphine, met-/leu-enkephalin, beta-endorphin, and d-ala-enkephalinamide) appeared to enhance perinatal 6-OHDOPA neurotoxicity and thereby enhance noradrenergic hyperinnervation of hindbrain and cerebellum—the effect being attenuated by the opioid receptor antagonist naloxone (Harston et al. [1980](#page-5-0), [1981](#page-5-0); Kostrzewa and Klisans-Fuenmayor [1984\)](#page-6-0).

Despite the consequences of 6-OHDOPA on noradrenergic innervation, dopaminergic innervation to neostriatum is unaltered through the duration of postnatal ontogeny and for the life span by perinatal low-dose 6-OHDOPA (Kostrzewa and Garey [1976](#page-6-0)); at high-dose perinatal 6-OHDOPA, the tuberoinfundibular dopaminergic tract (Lin et al. [1993](#page-6-0)) is more susceptible to damage than the nigrostriatal dopaminergic tract (Nomura and Segawa [1979](#page-6-0)). Also, serotoninergic nerves are resistant to 6-OHDOPA neurotoxicity (Richardson et al. [1974](#page-7-0)).

Perinatal 6-OHDOPA treatment initially damaged sympathetic noradrenergic nerves innervating peripheral organs (i.e., heart, salivary glands), but by maturity all organs were fully innervated.

6-OHDOPA actions on noradrenergic neurons have been reviewed elsewhere (Kostrzewa [1988,](#page-5-0) [1988](#page-5-0), [2014](#page-5-0)).

4 6-OHDOPA Agonist Action at AMPA Receptors

Although 6-OHDOPA was shown, in 1976, to produce more of an excitatory action than glutamate on frog spinal neurons (Biscoe et al. [1976\)](#page-4-0), not until 1990 was it discovered that 6-OHDOPA exerts agonist action at alpha-amino-3 hydroxy-5-methylisoxazole-4-propionic acid receptors (AMPA-R) (Cha et al. [1991;](#page-4-0) Kunig et al. [1994a](#page-6-0), [b](#page-6-0); Olney et al. [1990](#page-6-0)). Actually, 6-OHDOPA-quinone is the suspected agonist (Aizenman et al. [1990,](#page-4-0) [1992;](#page-4-0) Rosenberg et al. [1991\)](#page-7-0). AMPA-R activity would represent a confound to the actions of 6-OHDOPA per se on noradrenergic neurons.

5 Non-specific Effects of Perinatal 6-OHDOPA

In rats treated with perinatal 6-OHDOPA, cholineacetyltransferase activity is reduced in brainstem (Jaim-Etcheverry et al. [1975\)](#page-5-0), and atropine-induced locomotor activity at P20 and P50 is enhanced, while pilocarpine catalepsy is abated (Nomura and Segawa [1979;](#page-6-0) Nomura et al. [1979](#page-6-0)). This cholinergic subsensitivity is reflected in the B_{max} for $[^{3}H]QNB$ (quinuclidinyl benzilate) binding at muscarinic receptor sites in mesolimbic and striatal brain regions and also in heart (Nomura et al. [1979\)](#page-6-0).

High-dose 6-OHDOPA treatment is associated with the production of methemoglobinemia (Corrodi et al. [1971\)](#page-5-0), which of itself promotes in vivo formation of 6-OHDOPA from tyrosine, a process enhanced by hydrogen peroxide formation (Agrup et al. [1983](#page-4-0)) and known to increase after 6-OHDOPA treatment.

6 Summary

6-OHDOPA was envisioned as an experimental tool, able to cross the BBB to be decarboxylated to 6-OHDA, and thus exert effects on noradrenergic and/or dopaminergic nerves in brain. By this means, 6-OHDA effects on brain could be realized without the necessity to otherwise inject 6-OHDA directly into brain since 6-OHDA does not cross the BBB. However, the neurotoxicity action (i.e., efficacy) of 6-OHDOPA is far less than that of 6-OHDA. Moreover, as a means of minimizing global effects, experimental 6-OHDA is generally applied to specific brain nuclei or specified tracts, not intraventricularly or intracisternally. Consequently, the overall utility of 6-OHDOPA is greatly restricted in biomedical research.

The non-specific effects of 6-OHDOPA, namely AMPA-R agonist activity by non-enzymatically formed 6-OHDOPA-quinone, and the methemoglobinemia arising from 6-OHDOPA and its quinone, further restrict the usefulness of 6-OHDOPA as an experimental tool.

Nevertheless, the early work with 6-OHDOPA did validate its role as a relatively selective noradrenergic neurotoxin, and the actions of 6-OHDOPA in perinates led to discovery of processes and mechanisms associated with nerve sprouting and nerve regeneration. Conceivably, action at the AMPA-R could still be advantageous in 6-OHDOPA use as a research tool.

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References

- Adams RN, Murrill E, McCreery R, Blank L, Karolczak M (1972) 6-Hydroxydopamine, a new oxidation mechanism. Eur J Pharmacol 17(2):287–292
- Agrup G, Hansson C, Rorsman H, Rosengren E, Tegner E (1983) Methaemoglobin-catalysed formation of dopa and 6-OH-dopa from tyrosine. Acta Derm Venereol 63(2):152–155
- Aizenman E, White WF, Loring RH, Rosenberg PA (1990) A 3,4-dihydroxyphenylalanine oxidation product is a non-N-methyl-D-aspartate glutamatergic agonist in rat cortical neurons. Neurosci Lett 116(1–2):168–171
- Aizenman E, Boeckman FA, Rosenberg PA (1992) Glutathione prevents 2,4,5-trihydroxyphenylalanine excitotoxicity by maintaining it in a reduced, non-active form. Neurosci Lett 144(1–2):233–236
- Berkowitz BA, Spector S, Brossi A, Focella A, Teitel S (1970) Preparation and biological properties of (−)- and (+)-6-hydroxydopa. Experientia 26(9):982–983
- Biscoe TJ, Evans RH, Headley PM, Martin MR, Watkins JC (1976) Structure-activity relations of excitatory amino acids on frog and rat spinal neurones. Br J Pharmacol 58(3):373–382
- Blank CL, Kissinger PT, Adams RN (1972) 5,6-Dihydroxyindole formation from oxidized 6-hydroxydopamine. Eur J Pharmacol 19(3):391–394
- Cha JH, Dure LS IV, Sakurai SY, Penney JB, Young AB (1991) 2,4,5-Trihydroxyphenylalanine (6-hydroxy-dopa) displaces [³H]AMPA binding in rat striatum. Neurosci Lett 132(1):55-58
- Clark MB, King JC, Kostrzewa RM (1979) Loss of nerve cell bodies in caudal locus coeruleus following treatment of neonates with 6-hydroxydopa. Neurosci Lett 13(3):331–336
- Cohen G, Heikkila RE (1974) The generation of hydrogen peroxide, superoxide radical, and hydroxyl radical by 6-hydroxydopamine, dialuric acid, and related cytotoxic agents. J Biol Chem 249(8):2447–2452
- Corrodi H, Clark WG, Masuoka DI (1971) The synthesis and effects of DL-6-hydroxydopa. In: Malmfors T, Thoenen H (eds) 6-Hydroxydopamine and catecholamine neurons. North Holland, Amsterdam, pp 187–192
- Evans JM, Cohen G (1989) Studies on the formation of 6-hydroxydopamine in mouse brain after administration of 2,4,5-trihydroxyphenylalanine (6-hydroxyDOPA). J Neurochem 52(5):1461– 1467
- Evans J, Cohen G (1993) Catecholamine uptake inhibitors elevate 6-hydroxydopamine in brain after administration of 6-hydroxydopa. Eur J Pharmacol 232(2–3):241–245
- Harston CT, Morrow A, Kostrzewa RM (1980) Enhancement of sprouting and putative regeneration of central noradrenergic fibers by morphine. Brain Res Bull 5(4):421–424
- Harston CT, Clark MB, Hardin JC, Kostrzewa RM (1981) Opiate-enhanced toxicity and noradrenergic sprouting in rats treated with 6-hydroxydopa. Eur J Pharmacol 71(4):365–373
- Heikkila R, Cohen G (1971) Inhibition of biogenic amine uptake by hydrogen peroxide: a mechanism for toxic effects of 6-hydroxydopamine. Science 172(3989):1257–1258
- Heikkila R, Cohen G (1972a) Further studies on the generation of hydrogen peroxide by 6-hydroxydopamine. Potentiation by ascorbic acid. Mol Pharmacol 8(2):241–248
- Heikkila RE, Cohen G (1972b) In vivo generation of hydrogen peroxide from 6-hydroxydopamine. Experientia 28(10):1197–1198
- Heikkila RE, Cohen G (1973) 6-Hydroxydopamine: evidence for superoxide radical as an oxidative intermediate. Science 181(4098):456–457
- Jacobowitz D, Kostrzewa R (1971) Selective action of 6-hydroxydopa on noradrenergic terminals: mapping of preterminal axons of the brain. Life Sci I 10(23):1329–1342
- Jaim-Etcheverry G, Zieher LM (1977) Differential effect of various 6-hydroxydopa treatments on the development of central and peripheral noradrenergic neurons. Eur J Pharmacol 45(2):105– 116
- Jaim-Etcheverry G, Teitelman G, Zieher LM (1975) Choline acetyltransferase activity increases in the brain stem of rats treated at birth with 6-hydroxydopa. Brain Res 100(3):699–704
- Jonsson G, Sachs C (1973) Pharmacological modifications of the 6-hydroxy-dopa induced degeneration of central noradrenaline neurons. Biochem Pharmacol 22(14):1709–1716
- Klisans-Fuenmayor D, Harston CT, Kostrzewa RM (1986) Alterations in noradrenergic innervation of the brain following dorsal bundle lesions in neonatal rats. Brain Res Bull 16 (1):47–54
- Kostrezewa RM, Klara JW, Robertson J, Walker LC (1978) Studies on the mechanism of sprouting of noradrenergic terminals in rat and mouse cerebellum after neonatal 6-hydroxydopa. Brain Res Bull 3(5):525–531
- Kostrzewa RM (1975) Effects of neonatal 6 hydroxydopa treatment on monamine content of rat brain and peripheral tissues. Res Commun Chem Pathol Pharmacol 11(4):567–579
- Kostrzewa RM (1988) Reorganization of noradrenergic neuronal systems following neonatal chemical and surgical injury. Prog Brain Res 73:405–423. Review. PMID: 3138742
- Kostrzewa RM (1998) 6-Hydroxydopa, a catecholamine neurotoxin and endogenous excitotoxin at non-NMDA receptors. In: Kostrzewa RM (ed) Highly selective neurotoxins: basic and clinical applications. Humana Press, Totowa NJ, pp 109–129
- Kostrzewa RM (2007) The blood-brain barrier for catecholamines—revisited. Neurotoxicity Res 11:261–271
- Kostrzewa RM (2014) Survey of selective neurotoxins, in Section on Selective Neurotoxins, In: Kostrzewa RM (ed) Handbook of neurotoxicity, Springer New York, Heidelberg, Dordrecht, London, pp 3–67. ISBN 978-1-4614-5835-7 (print); ISBN 978-1-4614-5836-4 (eBook); ISBN 978-1-4614-7458-6 (print and electronic bundle). doi[:10.1007/978-1-4614-5836-4_53](http://dx.doi.org/10.1007/978-1-4614-5836-4_53)
- Kostrzewa RM, Garey RE (1976) Effects of 6-hydroxydopa on noradrenergic neurons in developing rat brain. J Pharmacol Exp Ther 197:105–118
- Kostrzewa RM, Garey RE (1977) Sprouting of noradrenergic terminals in rat cerebellum following neonatal treatment with 6-hydroxydopa. Brain Res 124:385–391
- Kostrzewa RM, Harper JW (1974) Effect of 6-hydroxydopa on catecholamine-containing neurons in brains of newborn rats. Brain Res 69(1):174–181
- Kostrzewa RM, Harper JW (1975) Comparison of the neonatal effects of 6-hydroxydopa and 6-hydroxydopamine on growth and development of noradrenergic neurons in the central nervous system, In: Jonsson G, Malmfors T, Sachs C (eds) Chemical tools in catecholamine research, vol I. North Holland Publ Co, Amsterdam, The Netherlands, pp 181–188
- Kostrzewa R, Jacobowitz D (1972) The effect of 6-hydroxydopa on peripheral adrenergic neurons. J Pharmacol Exp Ther 183(2):284–297
- Kostrzewa R, Jacobwitz D (1973) Acute effects of 6-hydroxydopa on central monoaminergic neurons. Eur J Pharmacol 21(1):70–80
- Kostrzewa RM, Jacobowitz DM (1974) Pharmacological actions of 6-hydroxydopamine. Pharmacol Rev 26(3):199–288. Review. PMID: 4376244
- Kostrzewa RM, Klisans-Fuenmayor D (1984) Development of an opioid-specific action of morphine in modifying recovery of neonatally-damaged noradrenergic fibers in rat brain. Res Commun Chem Pathol Pharmacol 46(1):3–11
- Kostrzewa RM, Harston CT, Fukushima H, Brus R (1982) Noradrenergic fiber sprouting in the cerebellum. Brain Res Bull 9(1-6):509-517. PMID: 7172038
- Kostrzewa RM, Hardin JC, Jacobowitz DM (1988) Destruction of cells in the midportion of the locus coeruleus by a dorsal bundle lesion in neonatal rats. Brain Res 442(2):321–328
- Kostrzewa RM, Kostrzewa JP, Brus R (2000) Dopaminergic denervation enhances susceptibility to hydroxyl radicals in rat neostriatum. Amino Acids 19(1):183–199
- Künig G, Hartmann J, Niedermeyer B, Deckert J, Ransmayr G, Heinsen H, Beckmann H, Riederer P (1994a) Excitotoxins L-beta-oxalyl-amino-alanine (L-BOAA) and 3,4,6-trihydroxyphenylalanine (6-OH-DOPA) inhibit [3 $\mathsf{I}^3\mathsf{H}\mathsf{I}$ alpha-amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid (AMPA) binding in human hippocampus. Neurosci Lett 169(1-2):219-222
- Künig G, Niedermeyer B, Krause F, Hartmann J, Deckert J, Ransmayr G, Heinsen H, Beckmann H, Riederer P (1994b) Interactions of neurotoxins with non-NMDA glutamate receptors: an autoradiographic study. J Neural Transm Suppl 43:59–62
- Lin JY, Mai LM, Pan JT (1993) Effects of systemic administration of 6-hydroxydopamine, 6-hydroxydopa and 1-methyl-4-phenyl-1,2,3,6-tetrahydroxypyridine (MPTP) on tuberoinfundibular dopaminergic neurons in the rat. Brain Res 624(1–2):126–130
- McLean JH, Kostrzewa RM, May JG (1976) Behavioral and biochemical effects of neonatal treatment of rats with 6-hydroxydopa. Pharmacol Biochem Behav 4(5):601–607
- McLean JH, Glasser RS, Kostrzewa RM, May JG (1980) Effects of neonatal 6-hydroxydopa on behavior in female rats. Pharmacol Biochem Behav 13(6):863–868
- Morgan DN, McLean JH, Kostrzewa RM (1979) Effects of 6-hydroxydopamine and 6-hydroxydopa on development of behavior. Pharmacol Biochem Behav 11(3):309–312
- Nomura Y, Segawa T (1979) Striatal dopamine content reduced in developing rats treated with 6-hydroxydopa. Jpn J Pharmacol 29(2):306–309
- Nomura Y, Kajiyama H, Segawa T (1979) Decrease in muscarinic cholinergic response of the rat heart following treatment with 6-hydroxydopa. Eur J Pharmacol 60(4):323–327
- Olney JW, Zorumski CF, Stewart GR, Price MT, Wang GJ, Labruyere J (1990) Excitotoxicity of L-dopa and 6-OH-dopa: implications for Parkinson's and Huntington's diseases. Exp Neurol 108(3):269–272
- Ong HH, Creveling CR, Daly JW (1969) The synthesis of 2,4,5-trihydroxyphenylalanine (6-hydroxydopa). A centrally active norepinephrine-depleting agent. J Med Chem 12(3): 458–462
- Porter CC, Totaro JA, Stone CA (1963) Effect of 6-hydroxydopamine and some other compounds on the concentration of norepinephrine in the hearts of mice. J Pharmacol Exp Ther 140: 308–316
- Porter CC, Totaro JA, Burcin A (1965) The relationship between radioactivity and norepinephrine concentrations in the brains and hearts of mice following administration of labeled methyldopa or 6-hydroxydopamine. J Pharmacol Exp Ther 150(1):17–22
- Richardson JS, Jacobowitz DM (1973) Depletion of brain norepinephrine by intraventricular injection of 6-hydroxydopa: a biochemical, histochemical and behavioral study in rats. Brain Res 58(1):117–133
- Richardson JS, Cowan N, Hartman R, Jacobowitz DM (1974) On the behavioral and neurochemical actions of 6-hydroxydopa and 5,6-dihydroxytryptamine in rats. Res Commun Chem Pathol Pharmacol 8(1):29–44
- Rosenberg PA, Loring R, Xie Y, Zaleskas V, Aizenman E (1991) 2,4,5-trihydroxyphenylalanine in solution forms a non-N-methyl-D-aspartate glutamatergic agonist and neurotoxin. Proc Natl Acad Sci USA 88(11):4865–4869
- Sachs C, Jonsson G (1972a) Degeneration of central and peripheral noradrenaline neurons produced by 6-hydroxy-DOPA. J Neurochem 19(6):1561–1575
- Sachs C, Jonsson G (1972b) Selective 6-hydroxy-DOPA induced degeneration of central and peripheral noradrenaline neurons. Brain Res 40(2):563–568
- Sachs C, Jonsson G, Fuxe K (1973) Mapping of central noradrenaline pathways with 6-hydroxy-DOPA. Brain Res 63:249–261
- Saner A, Thoenen H (1971) Model experiments on the molecular mechanism of action of 6-hydroxydopamine. Mol Pharmacol 7(2):147–154
- Senoh S, Witkop B (1959) Formation and rearrangements of aminochromes from a new metabolite of dopamine and some of its derivatives. J Am Chem Soc 81:6231–6235
- Thoenen H, Tranzer JP (1968a) Chemical sympathectomy by selective destruction of adrenergic nerve endings with 6-Hydroxydopamine. Naunyn Schmiedebergs Arch Exp Pathol Pharmakol 261(3):271–288
- Thoenen H, Tranzer JP (1968b) On the possibility of chemical sympathectomy by selective destruction of adrenergic nerve endings with 6-hydroxydopamine (6-OH-DA). Naunyn Schmiedebergs Arch Exp Pathol Pharmakol 260(2):212–213. German. PMID: 4239240
- Tohyama M, Maeda T, Kashiba A, Shimizu N (1974) Fluorescence and electron microscopic analysis of axonal change of coerulo-cortical noradrenaline neuron system following destruction of locus coeruleus and administration of 6-hydroxydopa in the rat brain. Med J Osaka Univ 24(4):205–221
- Toyama M, Maeda T, Shimizu N (1974) Detailed noradrenaline pathways of locus coeruleus neuron to the cerebral cortex with use of 6-hydroxydopa. Brain Res 79(1):139–144
- Ungerstedt U (1968) 6-Hydroxy-dopamine induced degeneration of central monoamine neurons. Eur J Pharmacol 5(1):107–110
- Wehrli PA, Pigott F, Fischer U, Kaiser A (1972) Oxidation products of 6-hydroxy-dopamine. Helv Chim Acta 55(8):3057–3061 (Article in German)
- Zieher LM, Jaim-Etcheverry G (1973) Regional differences in the long-term effect of neonatal 6-hydroxydopa treatment on rat brain noradrenaline. Brain Res 60(1):199–207
- Zieher LM, Jaim-Etcheverry G (1975a) 6-hydroxydopa during development of central adrenergic neurons produces different long-term changes in rat brain noradrenaline. Brain Res 86(2): 271–281
- Zieher LM, Jaim-Etcheverry G (1975b) Different alterations in the development of the noradrenergic innervation of the cerebellum and the brain stem produced by neonatal 6-hydroxydopa. Life Sci 17(6):987–991
- Zieher LM, Jaim-Etcheverry G (1979) 6-Hydroxydopamine during development: relation between opposite regional changes in brain noradrenaline. Eur J Pharmacol 58(3):217-223. PMID: 510355