

Homeobox Genes and Homeodomain Proteins: New Insights into Cardiac Development, Degeneration and Regeneration

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Abstract

Cardiovascular diseases are the most common cause of human death in the developing world. Extensive evidence indicates that various toxic environmental factors and unhealthy lifestyle choices contribute to the risk, incidence and severity of cardiovascular diseases. Alterations in the genetic level of myocardium affects normal heart development and initiates pathological processes leading to various types of cardiac diseases. Homeobox genes are a large and highly specialized family of closely related genes that direct the formation of body structure, including cardiac development. Homeobox genes encode homeodomain proteins that function as transcription factors with characteristic structures that allow them to bind to DNA, regulate gene expression and subsequently control the proper physiological function of cells, tissues and organs. Mutations in homeobox genes are rare and usually lethal with evident alterations in cardiac function at or soon after the birth. Our understanding of homeobox gene family expression and function has expanded significantly during the recent years. However,

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the involvement of homeobox genes in the development of human and animal cardiac tissue requires further investigation. The phenotype of human congenital heart defects unveils only some aspects of human heart development. Therefore, mouse models are often used to gain a better understanding of human heart function, pathology and regeneration. In this review, we have focused on the role of homeobox genes in the development and pathology of human heart as potential tools for the future development of targeted regenerative strategies for various heart malfunctions.

Keywords

 $\begin{array}{l} \mbox{Cardiac development} \cdot \mbox{Cardiac regeneration} \cdot \\ \mbox{Heart disease} \cdot \mbox{Homeobox genes} \end{array}$

Abbreviations

AMHC1	atrial myosin heavy	chain-1	
ANTP	Antennapedia		
BMP	bone morphogenetic	e protein	
Cdh2	cadherin 2		
CDK	cyclin-dependent kinases		
Cited2	Cbp/P300	interacting	
	transactivator with	Glu/Asp Rich	
	Carboxy-Terminal I	Domain 2	
CNS	central nerve system	1	

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ESC embryonic stem cells FGF fibroblast growth factor FHF first heart field Flk1 fetal liver kinase 1 GJA5 gap junction protein alpha 5 GSC goosecoid H3K27me3 histone H3 methylation on the amino (N) terminal tail Hcn4 hyperpolarization-activated cyclic nucleotide-gated channel 4 gene HOXL homeobox transcription factor Hox-like Irx Iroquois family of homeobox genes ISL1 LIM-homeodomain transcription factor islet 1/insulin gene enhancer protein ISL-1 JMJD3 JmjC domain-containing protein 3 MEF2C myocyte-specific enhancer factor 2CMESP1 mesoderm posterior BHLH transcription factor 1 **MSCs** mesenchymal stem cells Myocd myocardin NKL NK-like homeobox protein NK-2 homolog Nkx2-5 E Nodal nodal growth differentiation factor Nppa natriuretic peptide A OFT outflow tract PCBP2 poly(rC)-binding protein 2 Pitx2 paired like homeodomain 2 Pitx2c paired-like homeodomain transcription factor 2 PROS prospero RA retinoic acid SAN sinoatrial node SHF second heart field Shox2 short stature homeobox 2 **SMAD** main signal transducers for receptors of the transforming growth factor beta $(TGF-\beta)$ superfamily; TALE three-amino-acid loop extension Tbx5 T-box transcription factor 5 TF transcription factors

TGF-β VCS ZEB2	transforming growth factor beta; ventricular conduction system zinc finger E-box binding homeo-		
2202	box 2		
ZF	zinc finger		
Ziro	zebrafish iroquois homeobox		
ZO-3	genes tight junction protein 3		

1 Introduction

Homeobox genes are a large family of genes that direct the formation of body structures along the head-tail axis in multicellular animal species (Innis 1997; Shashikant et al. 1991). It is also known that homeobox genes (Hox genes), as an ancient class of transcription factors, are important for the body patterning during embryo development (Innis 1997; Shashikant et al. 1991). Many of the homeobox genes play very important part in the spatiotemporal development of human heart (Lage et al. 2010). Likewise, some of these genes shape the human heart and control its multistep developmental process from simple crescent cells to a fully functional organ. For example, homeobox genes like homeobox protein NK-2 homolog E (Nkx2-5), LIM-homeodomain transcription factor islet 1 (Isl1), paired like homeodomain 2 (Pitx2) are widely known to be important for the proper development of human heart (Akazawa and Komuro 2005; Luo et al. 2014; Franco et al. 2017). However, there are many more homeobox genes that play substantial roles in cardiac function but thus far, there are less known and/or less investigated.

Specific inherited gene mutations cause congenital heart defects such as atrial or ventricle septal defects, abnormalities of outflow tract and etc. (Bao et al. 1999). Similarly, various pathological lifestyle factors like smoking, low physical activity, toxic and noxious agents and other environmental factors might also negatively affect cardiovascular function and promote heart failure (O'Toole et al. 2008; Nayor and Vasan 2015). Since it is impossible to exactly pinpoint how certain gene mutations influence development of human heart at the earliest stages, different mouse models have been created to better understand regulation of human heart development and its relation to various diseases (Camacho et al. 2016). Many of the genes studies in mouse models have similar vital roles in the development and function of human heart (Xu and Baldini 2007). Therefore, investigation of human disease and cues from mouse heart development models have revealed an important role of homeobox genes, including those that encode transcription factors.

Aside from already known homeobox genes, there are more homeobox genes that are essential for the formation of human and/or mouse myocardium. Some of these homeobox genes code transcription factors (TF), whereas other form a tight network regulating heart development and fate of heart progenitors. Several review articles have explored individual families of homeobox gene and their roles in embryo development. However, knowledge concerning the involvement of homeobox genes and homeodomain TF in the development of human heart referring mouse models are still lacking. Therefore, in this review we describe the role of more than 20 homeobox genes that are mainly involved in heart development and around 15 homeobox genes that are known to play minor or less investigated, but nonetheless important roles in cardiac development. Data summarized in this review will help to broaden the possible future applications of homeobox genes and their coded TF in targeted therapeutic strategies for cardiac regeneration and therapy.

2 Development of the Human Heart

Starting from the day first of fertilization, the zygote undergoes multiple cell divisions leading to the formation of third germ layer, known as the mesoderm (Moorman et al. 2003). Later

mesodermal cells migrate towards anterior part of embryo to form a distinct crescent-shaped epithelium, named the cardiac crescent (Buckingham et al. 2005). Cells situated in the distinct anterior-lateral territory within the cardiac crescent contribute to the formation of first heart field (FHF), distinguished by the expression of hyperpolarization-activated cyclic nucleotidegated channel 4 gene (Hcn4) (Liang et al. 2013). Cardiac progenitor cells also develop into second heart field (SHF), which is located medially to the cardiac crescent and extend posteriorly (Cai et al. 2003). Formation of SHF is marked by the expression of LIM-homeodomain transcription factor islet 1 (ISL1) (Cai et al. 2003). Sometimes progenitors of FHF and SHF are called cardiogenic or cardiac mesoderm (Dupays et al. 2015; Liu et al. 2014; Kitajima et al. 2000). These distinct heart fields fuse to form heart tube, which eventually develops into functional heart (Fig. 1) (Moorman et al. 2003; Nemer 2008). During this time, the primitive cardiac conduction system, including sinoatrial node (SAN), ventricular conduction system and other, starts to form (van Weerd and Christoffels 2016). The FHF cells develop into the left ventricle, as well as into the atrioventricular canal and part of the atria, whereas SHF cells develop into the right ventricle and outflow tract, with contribution to the formation of atria and inflow vessels (Buckingham et al. 2005). Once the heart fields are formed, they fuse into heart tube and undergo process called heart looping. During this phase the whole heart tube twists in the rightward direction eventually forming clearly visible, but still primitive, heart chambers (Santini et al. 2016). Later on, the heart undergoes septation to fully separated left and right sides of the heart.

There are many factors regulating human and mouse heart development, however only some of them may be considered to be core regulators of cardiogenesis. One of the most important TF is GATA4 which orchestrates expression of multiple transcriptions including other major determinants of cardiomyogenesis like Nkx2-5, T-box transcription factor 5 (Tbx5), heart- and



Fig. 1 Schematic representation of heart development in human and mouse. *FGF* first heart field, *SHF* second heart field, *RV* right ventricle, *LV* left ventricle, *RA* right atrium, *LA* left atrium. (Scheme adapted from (Nemer, 2008))

neural crest derivatives-expressed protein 1 (HAND1/2) and others (Bruneau et al. 2001a; Belaguli et al. 2000; Sepulveda et al. 1998). GATA4 integrates bone morphogenetic proteins (BMP) and SMAD, the main signal transducers for receptors of the transforming growth factor beta (TGF- β) superfamily, to ensure cardiac cell survival and stable lineage during cardiac development (Benchabane and Wrana 2003). Of course, there are other factors that promote development of various structures within the heart. For example, it is know that Tbx5 controls atrial gene expression, whereas myocyte-specific enhancer factor 2C (MEF2C) promotes development of ventricle and vasculogenesis (Bruneau et al. 2001a; Lin et al. 1997). Altogether, the development of human heart is a carefully controlled multistep process involving many genes, intracellular and extracellular signalling factors leading to proper cardiac function. The miss-controlled heart development process leads to various inherited or acquired cardiac disorders. This review focuses mostly on the homeodomain proteins, as one of the most important group of transcription factors regulating heart development, function and impairment.

3 Homeobox Genes

Homeodomain proteins are one of the most important group of proteins/transcription factors regulating plan of body structure and organogenesis in eukaryotes including heart development and disorders. DNA binding proteins have been extensively studied, but even today there are no established rules for predicting the specificity of DNA sequence based upon the amino acid sequence of the proteins. Homeodomain proteins are characterized by specific 60 amino acid long helix-turn-helix DNA binding homeodomain motif (Seifert et al. 2015). The homeodomain is a very highly conserved structure and consists of three helical regions folded into a tight globular structure that binds a 5'-TAAT-3' core motif. The high degree of conservation of homeodomain proteins is an ideal model to study specific protein-DNA interactions. The DNA sequence that encodes the homeodomain is called the "homeobox" and homeobox-containing genes are known as "hox" genes.

Most of the transcription factors belonging to this group are not only structurally but also evolutionary conserved and play crucial roles in embryonic patterning and differentiation (Pearson et al. 2005). The main role of homeodomain proteins *in vivo* is to control the genetic determination of development and implementation of the genetic body plan. There are 102 homeobox gene families that represent 235 active human homeodomain proteins, but only some homeodomain classes have close association with cardiac development and/or diseases (Bürglin and Affolter 2016). This review covers description of around 20 homeobox genes that up today are known to have a major



Fig. 2 Hierarchy of homeobox genes involved in heart development. Pink boxes indicate homeobox gene classes, yellow box indicates TALE gene superclass. Red boxes

impact in the regulation of heart development and functioning (Fig. 2).

4 Homeobox Genes in Mouse Heart Development and Human Disease

Humans have more than 67 genes that are important for cardiac hypertrophy and over 92 genes that control cardiovascular system, therefore it is reasonable to assume that some of the genes might be responsible for cardiac development and disease (van der Harst et al. 2016; Smith and Newton-Cheh 2015). Congenital heart disease (CHD) have structural heart anomaly, including atrial or ventricle septal defects, overriding aorta, right atrium isomerism and other structural changes in new born heart. Patients do usually display multiple symptoms, like, rapid breathing, bluish skin, poor weight

indicate genes with major involvement in heart development and diseases. Blue boxes indicate genes having less important role in heart development and diseases

gain and feeling tired in general (Sun et al. 2015). The main cause of these abnormalities is reduced blood oxygen levels in whole organism, which is a result of improper heart septation leading to the mixture of oxygenated and deoxygenated blood in systemic blood circulation. Additionally, CHD could be caused by genetical or environmental factors like infections during pregnancy such as Rubella, drugs and maternal illness (Sun et al. 2015). Since homeobox genes are important for the heart development, some mutation of the homeobox genes including *MEIS2*, *Nkx2-5* and others can potentially cause CHD (Zakariyah et al. 2017; Johansson et al. 2014). Of course, homeobox gene mutations is not a sole cause of heart defect present at human birth. For example, mutations in GATA4 and Tbx5 can also affect integrity of heart tissue (McCulley and Black 2012). In addition, impaired gene functioning might be also related to the other heart diseases like cardiomyopathy, hypertrophy, defects of the heart rhythm and other abnormalities (Kathiresan and Srivastava 2012). However, the impact of homeobox genes in heart disorders is still not clear and needs further investigation to clarify their role not only in the cardiac development but also in the physiological and pathophysiological conditions.

5 ANTP Class of Homeobox Genes

The Antennapedia (ANTP)-class of homeobox genes are involved in the determination of pattern formation along the anterior-posterior axis of the animal embryo. NK-like (NKL) and homeobox transcription factor Hox-like (HOXL) are an ancient subclasses of homeobox genes that belong to an ANTP homeobox gene class (Holland et al. 2007). It is likely that HOXL gene clusters originate from NKL, since NKL genes are widespread throughout the genome as tight clustered Hox genes (Bürglin and Affolter 2016). Both subclasses of the genes (NKL and HOXL) evolutionarily are conserved and play predetermined roles in heart patterning and disease.

6 Hox Gene Families of HOXL Subclass

Hox gene families belong to the HOXL subclass of ANTP class of homeobox. Hox genes code transcription factors which are important for whole body patterning and development (Pearson et al. 2005). It total, there are 39 human Hox genes grouped in HOXA, HOXB, HOXC and HOXD gene clusters. Hox genes are highly conserved, because they play a vital role in anterior-posterior formation of body axis (Pearson et al. 2005). The precise function of these genes is achieved by their specific temporal and spatial expression over the life course. During early mouse cardiac development, the retinoic acid (RA) might be responsible for the anterior-posterior patterning in SHF (Bertrand et al. 2011). Hoxb1, Hoxa1, and Hoxa3 act as downstream targets of RA and participate in forming outflow tract (OFT) and normal SHF development (Bertrand et al. 2011). Hoxb1-/or Hoxal - / -, Hoxbl + / - mouse embryos develop shortened OFT and display abnormal proliferation and premature differentiation of cardiac progenitors (Bertrand et al. 2011). This is probably related to the altered fibroblast growth factor (FGF) and BMP signaling pathways in developing mouse embryo (Roux et al. 2015). Clinical studies have revealed that Hoxa1 mutations might cause congenital human heart defects and other abnormalities like, mental retardation, deafness, horizontal gaze restriction and etc. (Bosley et al. 2008). Several other studies have shown that Hox genes might be also related to the human heart diseases, however more research is needed to unveil exact functions of these genes in human heart development (Gong et al. 2005; Haas et al. 2013).

7 Nk4 Gene Family of NKL Subclass

There are multiple NKL genes in mouse and humans regulating various developmental processes, however only some of them contribute to the development of the heart (Larroux et al. 2007). The *Nkx2-5* and *Nkx2-6* genes are the members of the NK4 homeobox gene family of NKL subclass and are closely related to the Drosophila *tinman* gene (Bürglin and Affolter 2016; Harvey 1996). To our knowledge, only *Nkx2-6* and *Nkx2-5* relate to the mouse and human heart development and disease, whereas *Nkx2-3*, *Nkx2-7*, *Nkx2-8* and *Nkx2-10* might be important for the heart development of zebrafish, frog or chicken (Newman and Krieg 1998; Wang et al. 2014; Tu et al. 2009; Allen et al. 2006; Brand et al. 1997).

During the early stages of embryogenesis, Nkx2-5 is expressed in myocardium and pharyngeal endoderm, whereas Nkx2-6 can be found in sinus venosus, pharyngeal endoderm and myocardium of the outflow tract (Lints et al. 1993). Moreover, during normal heart development, Nkx2-5 expression is essential for the looping of vertebrate embryonic heart, heart septation and formation of cardiac conduction system, whereas most of the *Nkx2-5* mutations are related to human congenital heart disease and conduction defects (Tanaka et al. 1999). Inactivation of *Nkx2-*5 arrested heart formation at the looping stage revealing its critical role in cardiac development (Lyons et al. 1995). However, targeted disruption of *Nkx-2.6* did not cause any abnormalities in the heart suggesting a possible compensatory function of Nkx-2.5 (Tanaka et al. 2000).

It is important to note that Nkx2-5 mutations lead to an altered spatiotemporal development of human heart, improper heart septation and formation of cardiac conduction system (Dupays et al. 2015; McCulley and Black 2012; McElhinney et al. 2003). Analysis of human Nkx2-5 mutants and gene truncations showed that most of the mutations affected Nkx2-5 binding to DNA or localization its but not protein-protein (McCulley 2012; interactions and Black Reamon-Buettner et al. 2004). Several different studies of mice Nkx2-5 knockout and human embryonic stem cells (ESC) revealed that Nkx2-5 mutations might alter gene expression of specific transcription factors like SP1, SRY, JUND, STAT6, MYCN, PRDM16, HEY2 and others (Anderson et al. 2018; Li et al. 2015). Also some studies support an idea that, Nkx2-5 mutant proteins might alter space and time specific human cardiac development by dysregulating BMP, Notch and Wnt signalling pathways (Anderson et al. 2018; Wang et al. 2011; Luxán et al. 2016; Cambier et al. 2014). There is a possibility that Nkx2-5 modulates these pathways by interacting with multiple transcription factors in time-dependent mode. For example, in mouse heart Nkx2-5 interacts with Hand2 transcriptions factor to activate Irx4, which is necessary for the ventricular identity (Yamagishi et al. 2001). Conversely, Nkx2-5 expression is also timely regulated since Nkx2-5 overexpression leads to an improper SAN formation in early mouse development (Roux et al. 2015). Mammalian

heart development is also regulated by the combination of cardiac transcription factors having specific DNA motifs in their centrally located DNA binding domains. It was also shown that Nkx2-5, GATA4 and Tbx5 can physically interact and synergistically regulate targeted genes (Hiroi et al. 2001; Pradhan et al. 2016). Since these genes are the master regulators of heart development, functional mutations in these genes are linked to various types of congenital heart diseases (Benson 2002; Hatcher et al. 2003). Taken together, Nkx2-5 and other transcription factors like Isl1, GATA4, Tbx5, Hand2, MEF2C, Irx4 form a core of transcription factors essential for the heart development and congenital heart disease (Fig. 3) (McCulley and Black 2012).

8 HHEX Gene Family of NKL Subclass

Proline rich homeodomain protein or homeobox protein (PRH/HHEX) expressed by hematopoietic system is a transcription factor belonging to the family of NKL subclass gene (Bedford et al. 1993). As the name implies, it is important for the development of hematopoietic cell, but not less is essential for the development of other systems, including heart (Bedford et al. 1993). Mouse double HHex mutants have multiple developmental issues, including defective vasculogenesis, hypoplasia of the right ventricle, aberrant development of the compact myocardium and other complications related to forebrain, thyroid and liver developmental disorders (Hallaq et al. 1998). Additional studies have revealed that HHex plays distinct role in mouse cardiac mesoderm specification and development. HHex expression is controlled by Sox17 transcription factor, which is known to be essential for the formation of mouse cardiac mesoderm (Liu et al. 2014). Several studies of human population have shown that common variants of HHex gene (rs7923837 and rs1111875) may also be associated with diabetes (Karns et al. 2013; Kelliny et al. 2009; Pechlivanis et al. 2010).



Fig. 3 Core transcription factors important for the heart development and congenital heart disease. (Scheme adapted from (McCulley and Black, 2012))

9 PRD and PRD-Like Homeobox Class

The PRD class is the second largest of the homeobox gene classes in animal genomes and, like the ANTP class, these genes have been found only in animals. The PRD class derives its name from the *Paired (Prd)* gene of *Drosophila*. Multiple gene families belong to the PRD homeobox class, including *Shox2*, *Hopx*, *GSC*, *Pitx2* and others, which are important for the heart development and disease.

10 Shox Gene Family of PRD Class

Short stature homeobox 2 (Shox2) is a homeobox gene belonging to the PRD class of homeobox. Shox2 is an essential for the development of limb and cardiac conduction systems, including formation of sinoatrial node (SAN) in mice and humans (Gu et al. 2008; Blaschke et al. 2007; Liu et al. 2011). Studies of Shox2 function during the mouse development revealed several cues how this homeodomain transcription factor in particular controls formation of SAN (Blaschke et al. 2007). Shox2 mice null mutants displayed severe cardiac conduction defects, such as low heart rhythm rate and drastically reduced cell proliferation (Espinoza-Lewis et al. 2009). This phenotype is probably related to the downregulation of HCN4, Tbx3 and the upregulation of natriuretic peptide A (Nppa), gap junction protein alpha 5 (GJA5) and Nkx2-5 gene expressions (Espinoza-Lewis et al. 2009). It is also known that HCN channels play a vital role in autonomic control of heart rate, so it is no surprise why Shox2 null mutants do not develop SAN (Alig et al. 2009). During the normal development of mouse cardiac expression of Shox2 is also tightly controlled by several transcription factors. For example, transcription factor Tbx5 activate Shox2 expression, however transcription factors like Pitx2c and NKX2-5 potentially silence Shox2 expression (Espinoza-Lewis et al. 2011; Puskaric et al. 2010). Paired-like homeodomain transcription factor 2 (Pitx2c) also can potentially inhibit left-sided pacemaker specification by suppressing Shox2 expression in left atrium, therefore SAN develops only in the region of right atrium (Wang et al. 2010). All these results indicate that Shox2 is essential for the maintaining pacemaker cell program during the heart development. On the other hand, in adulthood Nkx2-5



Fig. 4 Signaling networks governing pacemaker and atrial differentiation of cardiomyocyte in developing mouse heart. (Scheme adapted from (Liang et al. 2017))

antagonizes Shox2 and promotes cardiomyocyte formation (Fig. 4) (Liang et al. 2017).

Taken together, these studies indicate that Shox2 might be a good candidate to develop biological pacemaker. Results from mouse ESCs and canine mesenchymal stem cells (MSCs) have shown that cells overexpressing Shox2 induce expression of SAN markers such as HCN4, Cx45 and Tbx3 (Ionta et al. 2015; Feng et al. 2016). In addition, mouse embryonic bodies overexpressing Shox2 showed better contractile phenotype compared to the control group of embryonic bodies (Ionta et al. 2015). Moreover, human patients with an early-onset atrial fibrillation had significantly downregulated expression of Shox2 gene (Hoffmann et al. 2016). These results are promising for patients suffering from heart rhythm defects, however, more studies are needed to test functions of biological pacemaker in order to treat human arrhythmias.

11 HOPX Gene Family of PRD Class

HOPX is another PRD-class homeobox gene family important for the multiple organ

development and tissue homeostasis in adults (Chen et al. 2015; Schneider et al. 2015; Mariotto et al. 2016). Since it lacks a DNA binding domain, HOPX can only modulate gene expression by forming complexes with other regulatory proteins (Kook et al. 2006). In general HOPX acts as a cell proliferation inhibitor in humans cancer cells, however its function in mouse cardiac cell differentiation is not entirely clear (Chen et al. 2015; Waraya et al. 2012; Yap et al. 2016). Studies of mouse development have shown that HOPX plays a critical function in early formation of cardiomyocyte progenitors. HOPX integrates BMP and WNT signalling in developing mouse heart by interacting with SMAD proteins and inhibiting WNT signalling pathway leading to the formation and differentiation of cardiomyocyte progenitors (Jain et al. 2015).

On the other hand, there are some cues that HOPX might act as a negative regulator of cardiac differentiation in mice. HOPX interacts with HDAC2, thus reducing GATA4 transcriptional activity by deacetylation (Trivedi et al. 2010). These findings are consistent with previous reports that overexpression of HDAC2 inhibits the development of cardiomyocytes by downregulating the expression of *GATA4* and *Nkx2-5* genes (Kawamura et al. 2005; Karamboulas et al. 2006). All these results highlight the complex nature of HOPX and its partners in heart development. HOPX undeniably plays a critical role in early heart development, because some mouse mutants cannot develop a functional myocardium and display cardiac conduction defects (Chen et al. 2002; Ismat et al. 2005). HOPX also might be related to the human heart failure, since HOPX is downregulated in patients having cardiac hypertrophy (Güleç et al. 2014; Trivedi et al. 2011).

12 Goosecoid Gene Family of PRD Class

Goosecoid (GSC) is another protein that belongs to the bicoid related paired (PRD) homeobox class of genes. Goosecoid is often associated with limb, skeletal and craniofacial development, although, it might be important for cardiac mesoderm formation, since its expression is controlled by Mesp1 (Zhu et al. 1998). Mesoderm posterior BHLH transcription factor 1 (MESP1) preferentially binds to two variations of E-box sequences and activates critical mesoderm modulators, including Gata4, mix paired-like homeobox (Mixl1) and GSC homeobox (Soibam et al. 2015). In addition, mesoderm formation can be induced with 1-proline and trans-4-hydroxy-1-proline resulting in increased expression of Mix11 and GSC (Date et al. 2013). GSC also is important for the cell migration in early embryonic development, therefore the overexpression of goosecoid enhances oncogenic cell growth and metastasis (Kang et al. 2014).

13 Pitx Gene Family of PRD-Like Class

Paired like homeodomain 2 (*Pitx2*) is a PRD-like homeobox class gene which is important for the establishment of the left-right axis and for the asymmetrical development of the mouse and probably human heart, lung, and spleen, twisting of the gut and stomach, as well as the development of the eyes (Campione et al. 1999; Shiratori et al. 2006; Evans and Gage, 2005). There are several alternative *Pitx2* transcripts, however only Pitx2c isoform plays determined role in the asymmetric development of mouse heart (Liu et al. 2002). Higher vertebrates, at an early heart development stage and after the heart tube formation, undergo embryonic heart looping, which is the first visual evidence of embryo asymmetry (Harvey 2002). Transcription factors like nodal growth differentiation factor (Nodal) and Cbp/P300 interacting transactivator with Glu/Asp Rich Carboxy-Terminal Domain 2 (Cited2) activate Pitx2 transcription leading to the rightward twist of the heart tube and forming prospective embryonic atrial and ventricular chambers. Deletion of *Pitx2c* in mouse caused drastic alteration of looping process leading to various heart defects including the isomerism of right atrium and ventricle (Lin et al. 1999; Yu et al. 2001).

Humans with Pitx2c mutations develop various heart abnormalities, including an improper formation of ventricle and atrial chambers septa, atrial fibrillation and others. It is quite likely that septation defects caused are by the downregulation of transcription factors downstream of Pitx2, since certain Pitx2 mutants displayed reduced cardiac transcriptional activity in human patients (Wang et al. 2013; Wei et al. 2014). Surprisingly, the overexpression of Pitx2c in mouse R1-embryonic stem cells results in elevated gene expression of essential cardiac transcription factors like GATA4, MEF2C, Nkx2-5 and others (Lozano-Velasco et al. 2011). Consequently, Pitx2c might be a good candidate for heart regeneration, since it positively regulates multiple transcription factors important for cardiac development. Recently it was shown that mouse embryonic stem cells overexpressing Pitx2c could restore mouse heart function after a myocardium through infarct the multiple mechanisms including efficient terminal

differentiation, regulation of action potentials of cardiomyocytes and positive paracrine effects (Guddati et al. 2009). However, more studies need to be done to determine the utility of Pitx2c in human heart regeneration strategies.

14 TALE Homeobox Superclass

Three-amino-acid loop extension (TALE) is another superclass of homeobox genes, which for highly conserved transcription codes regulators essential for various developmental programs. These genes encode proteins with atypical homeodomain structure, defined by havthree additional amino acids ing in homeodomain. TALE homeobox gene superclass includes the main zinc finger (ZF), PBC and Meis homeobox 1 (Meis) classes. Out of 20 human homeodomains only Meis1, Meis2 and Iroquois homeobox proteins 1-6 (Irx1-6) have their clearly defined function in heart development and disease.

15 Meis Genes Family of Meis Class

Meis1 encodes the TALE superclass homeobox transcription factor implicated in cardiac, hematopoietic and neural development (Mariotto et al. 2013; Azcoitia et al. 2005; Hisa et al. 2004). Meis1 deficient mice have malformed cardiac outflow tracts with overriding aorta and ventricular septal defect (Stankunas et al. 2008). Downregulation of Meis1 leads to cardiac hypertrophy in humans and mice. Meis1 binds poly (rC)-binding protein 2 (PCBP2) gene promoter and activates its expression in order to suppress human or mouse heart hypertrophy (Zhang et al. 2016). In turn, PCBP2 represses angiotensin II, which enhances hypertrophic human or mouse cardiac growth (Zhang et al. 2015). There are around 79 cardiac specific genes that have Meis1 and NKX2-5 binding sites in developing mouse heart, some of them are associated with

cell signaling and cardiac progenitor differentiation, like Tbx20, myocardin, cadherin 2 (Cdh2), Wnt11, and Wnt2 (Dupays et al. 2015). Adult mouse cardiomyocytes with mutant MEIS1 exhibit increased proliferation and progression of the cell cycle (Mariotto et al. 2013). This function is emphasized in adult mouse hearts since Meis1 activates inhibitors of cyclindependent kinases (CDK) like p15, p16 and p21 (Mariotto et al. 2013). In humans non-synonymous Meis1 gene variants might be associated with congenital heart defects, whereas patients carrying 2p14 microdeletions show symptoms of deafness and cardiomyopathy (Mathieu et al. 2017; Arrington et al. 2012). It is likely that Meis1 is required for the control of spatiotemporal cell proliferation in early developing heart to prevent hypertrophy, however more studies need to be done to fully understand the role of Meis1 in cardiac development and disease.

Meis2 encodes TALE homeobox superclass transcription factor essential for the development of mouse cranial and cardiac neural crest (Machon et al. 2015). Recent findings indicate that Meis2 might be an important factor for the proliferation of fetal human cardiomyocyte cells (Wu et al. 2015). Reduction of Meis2 gene expression by miR-134 in slowed progression of human results cardiomyocyte progenitor cell cycle (Wu et al. 2015). A clinical and genetic study also revealed that small Meis2 deletion can negatively affect several developmental processes: human patients with small Meis2 non-frame shift deletion (c.998 1000del:p.Arg333del) had serious cleft palate and cardiac septal defects (Louw et al. 2015). It is known that Meis2 interacts with DNA and forms multimeric complexes with Hox and Pbx proteins (Louw et al. 2015). Single deletion of arginine residue affects the ability of Meis2 to bind DNA leading to serious developmental problems of human heart (Louw et al. 2015). Clinical studies have shown that patients having only one functioning Meis2 gene copy survive, however they have similar phenotype such as clefting and ventricular septal defects leading to delayed

motor development and learning disability (Johansson et al. 2014).

16 Irx Gene Family of IRX Class

Iroquois homeobox genes and their coded homeodomain proteins are another class of transcription factors belonging to TALE superclass of homeobox genes. Iroquois-class homeodomain TF (Irx) defining feature is atypical homeodomain structure and specific Iroquois (IRO) homeodomain family sequence motif, which is important for the recognition of DNA sequence (Gómez-Skarmeta and Modolell 2002; Cavodeassi et al. 2001). Humans and mice have six Irx proteins, which are important for the development of lung, nervous system, eye, pancreas, female gonad, early limb and, of course, heart patterning (Cavodeassi et al. 2001; Cheng et al. 2005; Schwab et al. 2006; van Tuyl et al. 2006; Ragvin et al. 2010; Jorgensen and Gao 2005; McDonald et al. 2010). Irx1 and Irx2 are expressed in interventricular septum from E14.5 onward, however, mouse Irx2 mutants are viable and display no notable phenotype defects in the developing heart (Christoffels et al. 2000; Lebel et al. 2003). Irx1 gene variants might be related to the congenital heart disease in humans (Guo et al. 2017).

Irx3 gene in mice seems to be very important for the ventricular conduction system (VCS) (Christoffels et al. 2000). Various studies suggest that Irx3 is required to maintain rapid electric conduction through the VCS for proper ventricular activation, via antithetical regulation of Cx40 and Cx43 expression (Zhang et al. 2011; Kasahara et al. 2003). Clinical studies have revealed that defects of *Irx3* gene can cause lethal cardiac arrhythmias in human patients (Koizumi et al. 2016). Irx3 function appears to be evolutionary conserved, since expression of Ziro3a, a Irx3 homologue in zebrafish, is detected in developing fish heart (Zhang et al. 2011).

Irx4 is associated with the formation of ventricular myocardium in mouse and humans (Christoffels et al. 2000; Cheng et al. 2011). Data from mouse and chicken indicate that Irx4 suppresses atrial gene expression by down regulating atrial myosin heavy chain-1 (AMHC1) (Bao et al. 1999; Bruneau et al. 2001b). Several *Irx4* mutations have been identified that might be associated with human congenital heart disease, particularly ventricular septal defect (Cheng et al. 2011).

Irx5 is expressed in adult mouse heart and maintains proper action potentials, particularly regulates T-wave seen in ECG (Costantini et al. 2005). Mice lacking Irx5 develop properly without any structural abnormalities in the heart (Costantini et al. 2005). This indicates that *Irx5* is not required for cardiac development or that other *Irx* genes can compensate for the loss of Irx5.

Irx6 is detectable in mouse developing heart, however its expression is relatively weak compared to other *Irx* genes (Christoffels et al. 2000).

17 Zeb Gene Family of ZF Class

ZEB2 or zinc finger E-box binding homeobox 2 is a gene coding transcription factor belonging to class of ZF homeobox gene and homeodomain class of ZN proteins (Bürglin and Affolter, 2016). It has multiple functional domains (E-box, Zinc finger, homeobox), so naturally it can control gene expression with a variety of transcription factors (Gheldof et al. 2012). The complex nature of Zeb2 shows that it drives multiple processes including the development of heart and neural systems, however, it usually acts as a transcription repressor rather than activator (Hegarty et al. 2015). Systematic study of mouse and human ESC transcriptome differentiation profiles revealed that Zeb2 might play important role in cardiac specialization. Human ESC with silenced Zeb2 gene proliferate more slowly and fail to differentiate into mature cardiomyocytes compared to the wild cells (Busser et al. 2015). In addition, cardiomyocytes with silenced Zeb2 do not show any contractile properties, although cardiac differentiation program is activated. More detailed analysis has revealed that silencing of *Zeb2* gene negatively affects human striated muscle contraction program, including genes related to calmodulin pathway, HCN and potassium channels (Busser et al. 2015). Targeted regulation of Zeb2 gene expression improves cardiomyogenic processes and heart regeneration.

Zeb2 mutation is also often associated with Mowat-Wilson syndrome (Garavelli and Mainardi 2007). Major signs of this disorder frequently include distinctive facial features, intellectual disability, delayed development, an intestinal disorder called Hirschsprung disease, Congenital Heart Disease and other types of birth defects (Garavelli and Mainardi 2007). All mentioned disorders are related to the improper heart development caused by the Zeb2 defective heart cells. In addition, Zeb2 repress epithelial genes (claudins, tight junction protein 3 (ZO-3), connexins, E-cadherin, plakophilin 2, desmoplakin, and crumbs3) in order to induce epithelial to mesenchymal transition (EMT), which is crucial for the developmental processes such as gastrulation, neural crest formation, heart morphogenesis, formation of the musculoskeletal system, and craniofacial structures (Vandewalle et al. 2009; Garavelli et al. 2017).

18 LIM Homeobox Class

LIM homeobox class genes encode two Lim domains and one homeodomain. Lim domain is a 50–60 amino acid length zinc finger motif, which is primarily involved in protein-protein interactions, so naturally LIM transcription factors can interact with multiple proteins in cell, thus regulating its phenotype.

19 Isl Gene Family of LIM Homeobox Class

Isl1 is a LIM homeobox class member that encodes a homeodomain transcription factor important for cell differentiation, fate determination and generation of cell diversity in multiple mouse and human tissues including central nerve system (CNS), pancreas and heart (Zhuang et al. 2013). During early cardiac development, Isl1, Nkx2-5 and fetal liver kinase 1 (Flk1) support the formation of SHF, which gives rise to the right ventricle, outflow tract and part of the atria (Dyer and Kirby, 2009). Isl1 promotes expansion, migration and proliferation of SHF progenitor cells during the development of the mouse heart (Witzel et al. 2012). Additionally, Isl1+ mouse heart cells have potential to differentiate into multiple cell types within the heart, including cardiomyocytes, smooth muscle, pacemaker and endothelial cells (Laugwitz et al. 2007).

There are multiple mechanisms explaining how Isl1 can promote expression of target genes, which suggests the expression of Isl1 is tightly controlled during the mouse heart development. For example, Nkx2-5 homeodomain transcription factor downregulates Isl1 expression in order to promote ventricular development in mouse heart (Witzel et al. 2012; Prall et al. 2007). The newest studies indicate, that Isl1 may repress development of mouse heart ventricle in order to promote the development of SAN (Dorn et al. 2015). Mouse embryos overexpressing Isl1 develop SAN-like cells instead of ventricle myocardium (Dorn et al. 2015). It is likely that the expression of Isl1 activates Nkx2-5 expression in SHF progenitor cells, however, in later staged of heart development Nkx2-5 shuts down ISL1 expression to promote ventricular development (Dorn et al. 2015). Isl1 orchestrates the expression of hundreds of potential genes implicated in cardiac differentiation, mainly through epigenetic mechanisms (Wang et al. 2016). Isl1 in mouse ESCs acts together with JmjC domain-containing protein 3 (JMJD3) histone demethylase to promote the demethylation or tri-methylation of core histone H3 on the amino (N) terminal tail (H3K27me3) at the enhancer's place of key downstream target genes, such as myocardin (Myocd), MEF2C and others (Wang et al. 2016). In addition, Isl1 may reduce histone methylation near GATA4 and Nkx2-5 genes after the expression of Isl1 lentiviral gene, and can also recruit p300 histone acetyltransferase to the promoter of MEF2C gene in order to promote Mef2c expression in developing mouse embryo (Yu et al. 2013). Other data suggest that lentiviral-induced overexpression of Isll gene promotes not only MEF2C gene acetylation, but also GATA4 and Nkx2-5 in C3H10T1/2 mouse cell line (Xu et al. 2016). The tight control of *Ils1* gene expression is required, since it acts positive as а cardiomyogenic gene regulator reducing methylation and increasing acetylation levels of genes and histones by direct and indirect methods. Most of the published data concerning Isl1 function have come from the studies of mouse development, however there are some studies that link Isl1 gene expression with the susceptibility to human congenital heart disease (Luo et al. 2014; Stevens et al. 2010). Development of mechanisms that could control expression of Isl1 might be important target in further regulation of heart regeneration.

20 PROS Homeobox Class

Homeobox prospero (PROS) genes code atypical C terminal prospero domain and belongs to a distinctive class of Prospero homeodomain proteins (Yousef and Matthews, 2005). The PROS domain is a DNA binding domain of approximately 100 amino acids. In addition, PROS homeobox genes code additional three amino acids in their HD domain (Yousef and Matthews 2005).

21 PROX Gene Family of PROS Homeobox Class

Prospero homeobox 1 or Prox1 is a gene coding a transcription factor that plays important role in the development of mouse heart, CNS, eye, liver and lymphatic system (Elsir et al. 2012). Firstly, it was discovered in Drosophila as an important player in the development of central nervous system in insects. However, later Prox1 homologues were found in vertebrates and mammals (Elsir

et al. 2012). In mouse heart development of Prox1 is important for the sarcomere formation and muscle contraction (Risebro et al. 2009). Mouse Prox1 conditional mutants show increased number of fast twitch fibers compared to slow twitch fibers. Prox1 mutant mice develop fatal dilated cardiomyopathy and die around 7-14th week (Petchey et al. 2014). It was shown that in mice Prox1 acts as a transcriptional repressor of genes like Tnnt3, Tnni2 and Myl1 that are essential for the formation of fast twitch fibbers (Petchey et al. 2014). Prox1 might also be important for the maintenance of cardiac conduction system in adult mice. It was also shown that uncontrolled Nkx2-5 expression led to cardiac conduction defects, surprisingly suggesting that Prox1 might act as a direct upstream modifier of *Nkx2-5* gene expression (Risebro et al. 2012). In humans dysregulation of *Prox1* gene expression might also lead to congenital heart disease, like, hypoplastic left heart (Gill et al. 2009). Thus, the close connection of Prox1 with Nkx2-5 and other heart development and diseases regulating genes makes it an attractive target in cardiac regeneration field.

22 Role of Homeobox Genes in Cardiomyogenesis

The summarized and reviewed data of estimated involvement of homeobox genes in the heart development, diseases and/or regeneration processes suggest that some homeobox genes play more important role than the other. Data summarized in Table 1 show the homeobox have genes that been most commonly investigated with important roles in cardiomyogenesis.

It is quite evident that dozens of homeobox genes are required for early cardiomyogenesis, heart septation, formation of pacemaker cell, cardiomyocyte and etc. Some of the homeobox genes are directly related to the development of CHD, atrial fibrillations and other cardiac pathologies. However, there are much more

Gene	Development	Disease	Reference	
Hox	Hoxa1, Hoxb2 and Hoxb2 is important for anterior-posterior patterning in SHF. Integrating FGF and BMP signalling.	Hoxa1 mutations might cause CHD. HOXB13, and HOXC5 mutations might be related to heart disease.	Pearson et al. (2005), Bertrand et al. (2011), Bosley et al. (2008), Gong et al. (2005) and Haas et al. (2013)	
Nkx2–5	Heart looping, heart septation and cardiac conduction system formation. Integrates BMP, notch and WNT signaling during development.	Multiple gene variants and truncations are related to CHD.	McCulley and Black (2012), Tanaka et al. (1999), McElhinney et al. (2003), Anderson et al. (2018), Wang et al. (2011), Luxán et al. (2016) and Cambier et al. (2014)	
Hhex	Cardiac mesoderm specification.	HHex gene variants might be associated with diabetes.	Liu et al. (2014), Karns et al. (2013), Kelliny et al. (2009) and Pechlivanis et al. (2010)	
Shox2	Cardiac conduction system development.	Downregulation during early-onset atrial fibrillation.	Blaschke et al. (2007) and Hoffmann et al. (2016)	
Норх	Cardiomyocyte progenitor formation in mouse early heart. Negative regulator of GATA4 expression.	Downregulated in patients having cardiac hypertrophy.	Jain et al. (2015), Trivedi et al. (2010) and Trivedi et al. (2011)	
GSC	Cardiac mesoderm specification.		Zhu et al. (1998)	
Pitx2c	Establishment of the left-right axis in heart development. Heart looping and chamber septation.	Mutations cause improper ventricle and atrial chambers septa formation, atrial fibrillation.	Liu et al. (2002), Wang et al. (2013) and Wei et al. (2014)	
Meis1 and Meis2	Meis1 and Meis2 control of cell cycle progression during heart development.	Meis1 and Meis2 gene variants might be associated with CHD.	Mariotto et al. (2013), Arrington et al. (2012), Wu et al. (2015) and Louw et al. (2015)	
Irx1-6	Irx3 very important for ventricular conduction system. Irx4 is associated with the formation of ventricular myocardium in mouse and humans	Irx1 gene variants might be associated with CHD. Irx3 gene defects can cause lethal cardiac arrhythmias in human patients	Christoffels et al. (2000), Guo et al. (2017), Koizumi et al. (2016) and Cheng et al. (2011)	
ZEB2	Controls striated muscle development and contraction.	Gene variants cause Mowat- Wilson syndrome. Patients display CHD and other defects.	Busser et al. (2015) and Garavelli and Mainardi (2007)	
Islet1	Cell expansion, migration and proliferation. Marks formation of SHF. Repress ventricular fate in order to promote sinoatrial node development. Positive gene regulator, which reduces gene and histone methylation levels and increase acetylation.	Gene variants might be related to CHD.	Witzel et al. (2012), Dorn et al. (2015), Wang et al. (2016) and Stevens et al. (2010)	
Proxl	Important for sarcomere formation and muscle contraction.	Dysregulation of Prox1 might lead to CHD.	Elsir et al. (2012) and Petchey et al. (2014)	

Table 1 Homeobox genes with major involvement in heart development and diseases

homeobox genes related to the heart development, that so far have been less investigated or in one or another model system showed less direct involvement in cardiomyogenic processes (Table 2). Data summarized in Table 2 also highlight the fact that many more studies are needed to understand regulation of homeobox genes and their role in cardiomyogenic processes.

Class	Subclass	Gene	Development and disease	Reference
ANTP	HOXL	MEOX1 and MEOX2	Control of vascular endothelial cells proliferation in mice. Dysregulation might be associated with heart disease in mouse.	Lu et al. (2018), Douville et al. (2011)
ANTP	NKL	Msx1 and Msx2	Regulate survival of secondary heart field precursors and post-migratory proliferation of cardiac neural crest in the outflow tract	Chen et al. (2007)
ANTP	NKL	Lbx1	Specification of a subpopulation of cardiac neural crest necessary for normal heart development.	Schäfer et al. (2003)
ANTP	NKL	Nkx2-6	NKX2-6 mutation predisposes to familial atrial fibrillation.	Wang et al. (2014)
PRD		Prrx1 and Prrx2	Formation of cardiovascular system and connective tissues of the heart and in the great arteries and veins.	Bergwerff et al. (2000)
PRD		Pax3, Pax7	Involved in neural crest and cardiac development	JA (1996)
ZF (TALE)		ZFHX3	Genetic polymorphisms in are associated with atrial fibrillation in a Chinese Han population.	Liu et al. (2014)
PBC (TALE)		Pbx1	Patterning of the great arteries and cardiac outflow tract. Pbx acts with Hand2 in early myocardial differentiation in zebrafish. Non-synonymous variants in PBX genes are associated with congenital heart defects.	Stankunas et al. (2008) and Arrington et al. (2012), Chang et al. (2008)
TGIF (TALE)		Tgif1 and $Tgif2$	Left-right asymmetry formation and embryonic heart looping.	Powers et al. (2010)
SIX/SO		Six2	Six2 marks a dynamic subset of second heart field progenitors.	Zhou et al. (2017)

 Table 2
 Homeobox genes having less important role in heart development and diseases

23 Concluding Remarks

Heart development is a complex process requiring strict spatiotemporal development to form a healthy organ providing properly functioning organism. Multiple transcription factors, signalling pathways, morphogens and other stimuli govern the heart development process. However, it is possible to state that multiple homeobox genes and their coded transcription factors come into the heart developmental stages when their function is needed (Tables 1 and 2). None of these homeodomain transcription factors can be separated from each other, since their ability to bind DNA affects patterns of multiple gene thus resulting in changed transcriptome level of multiple cells.

Homeodomain proteins are not the only transcription factors important for cardiac development. The transcription factors of other gene families also significantly contribute heart development. For example, GATA, Tbx, HAND, Mef2c and other accompany homeodomain factors like Nkx2-5, Isl1, etc. (Hiroi et al. 2001; Gao et al. 2011; Maves et al. 2009; Skerjanc et al. 1998). Most of these homeodomain transcription factors are conserved and display coexistence and codependence in heart development of human as well as simple invertebrates like fruit fly or ascidians (Jensen et al. 2013a; Olson 2006). Only birds and mammals display fully separated heart, however reptilians still have no septum between right and left ventricles (Jensen et al. 2013b). Deeper further insights into septum formation of lower vertebrates like snakes, lizards and turtles could also help to understand signalling networks of human congenital heart diseases. Maybe in the future will be possible to engineer a reptile with four chambered heart, thus leading to better understanding of cardiac regeneration process and allowing to develop new therapeutic strategies for human cardiac congenital and other types of diseases.

Acknowledgements The study is funded by the Lithuanian Research council, project No. S-MIP-17-13.

Ethics Approval and Consent to Participate Not applicable.

Consent for Publication All authors agree to the publication of this manuscript.

Availability of Data and Material Not applicable.

Competing Interests The authors declare that they have no competing interests.

Authors' Contributions RM wrote the manuscript draft. DB revised the manuscript. AM read, corrected and approved the final manuscript.

Funding The study is funded by the Lithuanian Research council, project No. S-MIP-17-13.

References

- Akazawa H, Komuro I (2005) Cardiac transcription factor Csx/Nkx2-5: its role in cardiac development and diseases. Pharmacol Ther 107:252–268. https://doi. org/10.1016/j.pharmthera.2005.03.005
- Alig J, Marger L, Mesirca P, Ehmke H, Mangoni ME, Isbrandt D (2009) Control of heart rate by cAMP sensitivity of HCN channels. Proc Natl Acad Sci U S A 106:12189–12194. https://doi.org/10.1073/pnas. 0810332106
- Allen BG, Allen-Brady K, Weeks DL (2006) Reduction of XNkx2-10 expression leads to anterior defects and malformation of the embryonic heart. Mech Dev 123:719–729. https://doi.org/10.1016/j.mod.2006.07. 008
- Anderson DJ, Kaplan DI, Bell KM, Koutsis K, Haynes JM, Mills RJ, Phelan DG, Qian EL, Leitoguinho AR, Arasaratnam D, Labonne T, Ng ES, Davis RP, Casini S, Passier R, Hudson JE, Porrello ER, Costa MW, Rafii A, Curl CL, Delbridge LM et al (2018) NKX2-5 regulates human cardiomyogenesis via a HEY2 dependent transcriptional network. Nat Commun 9:1–13. https://doi.org/10.1038/s41467-018-03714-x
- Arrington CB, Dowse BR, Bleyl SB, Bowles NE (2012) Non-synonymous variants in pre-B cell leukemia homeobox (PBX) genes are associated with congenital heart defects. Eur J Med Genet 55:235–237. https://

doi.org/10.1016/j.ejmg.2012.02.002.Nonsynonymous

- Azcoitia V, Aracil M, Martínez-A C, Torres M (2005) The homeodomain protein Meis1 is essential for definitive hematopoiesis and vascular patterning in the mouse embryo. Dev Biol 280:307–320. https://doi.org/10. 1016/j.ydbio.2005.01.004
- Bao ZZ, Bruneau BG, Seidman JG, Seidman CE, Cepko CL (1999) Regulation of chamber-specific gene expression in the developing heart by irx4. Science 283:1161–1164
- Bedford FK, Ashworth A, Enver T, Wiedemann LM (1993) HEX: a novel homeobox gene expressed during haematopoiesis and conserved between mouse and human. Nucleic Acids Res 21:1245–1249
- Belaguli NS, Sepulveda JL, Nigam V, Charron F, Nemer M, Schwartz RJ (2000) Cardiac tissue enriched factors serum response factor and GATA-4 are mutual coregulators. Mol Cell Biol 20:7550–7558
- Benchabane H, Wrana JL (2003) GATA- and Smad1dependent enhancers in the Smad7 gene differentially interpret bone morphogenetic protein concentrations. Mol Cell Biol 23:6646–6661
- Benson DW (2002) The genetics of congenital heart disease: a point in the revolution. Cardiol Clin 20:385–394
- Bergwerff M, Gittenberger-de Groot AC, Wisse LJ, DeRuiter MC, Wessels A, Martin JF, Olson EN, Kern MJ (2000) Loss of function of the Prx1 and Prx2 homeobox genes alters architecture of the great elastic arteries and ductus arteriosus. Virchows Arch 436:12– 19
- Bertrand N, Roux M, Ryckebüsch L, Niederreither K, Dollé P, Moon A, Capecchi M, Zaffran S (2011) Hox genes define distinct progenitor sub-domains within the second heart field. Dev Biol 353:266–274. https:// doi.org/10.1016/j.ydbio.2011.02.029
- Blaschke RJ, Hahurij ND, Kuijper S, Just S, Wisse LJ, Deissler K, Maxelon T, Anastassiadis K, Spitzer J, Hardt SE, Schöler H, Feitsma H, Rottbauer W, Blum M, Meijlink F, Rappold G, Gittenberger-de Groot AC (2007) Targeted mutation reveals essential functions of the homeodomain transcription factor Shox2 in sinoatrial and pacemaking development. Circulation 115:1830–1838. https://doi.org/10.1161/ CIRCULATIONAHA.106.637819
- Bosley TM, Alorainy IA, Salih MA, Aldhalaan HM, Abu-Amero KK, Oystreck DT, Tischfield MA, Engle EC, Erickson RP (2008) The clinical spectrum of homozygous HOXA1 mutations. Am J Med Genet A 146:1235–1240. https://doi.org/10.1002/ajmg.a.32262
- Brand T, Andrée B, Schneider A, Buchberger A, Arnold H (1997) Chicken NKx2-8, a novel homeobox gene expressed during early heart and foregut development. Mech Dev 64:53–59
- Bruneau BG, Nemer G, Schmitt JP, Charron F, Robitaille L, Caron S, Conner DA, Gessler M, Nemer M, Seidman CE, Seidman JG (2001a) A murine model of Holt-Oram syndrome defines roles of the

T-box transcription factor Tbx5 in cardiogenesis and disease. Cell 106:709–721

- Bruneau BG, Bao ZZ, Fatkin D, Xavier-Neto JGD, Maguire CT, Berul CI, Kass DA, Kuroski-de Bold ML, de Bold AJ, Conner DA, Rosenthal N, Cepko CL, Seidman CE, Seidman JG (2001b) Cardiomyopathy in irx4-deficient mice is preceded by abnormal ventricular gene expression. Mol Cell Biol 21:1730–1736. https://doi.org/10.1128/MCB.21.5. 1730-1736.2001
- Buckingham M, Meilhac S, Zaffran S (2005) Building the mammalian heart from two sources of myocardial cells. Nat Rev Genet 6:826–837. https://doi.org/10. 1038/nrg1710
- Bürglin TR, Affolter M (2016) Homeodomain proteins: an update. Chromosoma 125:497–521. https://doi.org/10. 1007/s00412-015-0543-8
- Busser BW, Lin Y, Yang Y et al (2015) An orthologous epigenetic gene expression signature derived from differentiating embryonic stem cells identifies regulators of cardiogenesis. PLoS One 10:e0141066. https://doi.org/10.1371/journal.pone.0141066
- Cai CL, Liang X, Shi Y, Chu PH, Pfaff SL, Chen J, Evans S (2003) Isl1 identifies a cardiac progenitor population that proliferates prior to differentiation and contributes a majority of cells to the heart. Dev Cell 5:877–889
- Camacho P, Fan H, Liu Z, He J (2016) Small mammalian animal models of heart disease. Am J Cardiovasc Dis 6:70–80
- Cambier L, Plate M, Sucov HM, Pashmforoush M (2014) Nkx2-5 regulates cardiac growth through modulation of Wnt signaling by R-spondin3. Development 141:2959–2971. https://doi.org/10.1242/dev.103416
- Campione M, Steinbeisser H, Schweickert A, Deissler K, van Bebber F, Lowe LA, Nowotschin S, Viebahn C, Haffter P, Kuehn MR, Blum M (1999) The homeobox gene Pitx2: mediator of asymmetric left-right signaling in vertebrate heart and gut looping. Development 126:1225–1234
- Cavodeassi F, Modolell J, Gómez-Skarmeta JL (2001) The Iroquois family of genes: from body building to neural patterning. Development 128:2847–2855
- Chang CP, Stankunas K, Shang C, Kao SC, Twu KY, Cleary ML (2008) Pbx1 functions in distinct regulatory networks to pattern the great arteries and cardiac outflow tract. Development 135:3577–3586. https://doi. org/10.1242/dev.022350
- Chen F, Kook H, Milewski R, Gitler AD, Lu MM, Li J, Nazarian R, Schnepp R, Jen K, Biben C, Runke G, Mackay JP, Novotny J, Schwartz RJ, Harvey RP, Mullins MC, Epstein JA (2002) Hop is an unusual homeobox gene that modulates cardiac development. Cell 110:713–723
- Chen YH, Ishii M, Sun J, Sucov HM, Maxson RE Jr (2007) Msx1 and Msx2 regulate survival of secondary heart field precursors and post-migratory proliferation of cardiac neural crest in the outflow tract. Dev Biol 308:421–437. https://doi.org/10.1016/j.ydbio.2007.05. 037

- Chen Y, Yang L, Cui T, Pacyna-Gengelbach M, Petersen I (2015) Hopx is methylated and exerts tumoursuppressive function through ras-induced senescence in human lung cancer. J Pathol 235:397–407. https:// doi.org/10.1002/path.4469
- Cheng CW, Chow RL, Lebel M, Sakuma R, Cheung HO, Thanabalasingham V, Zhang X, Bruneau BG, Birch DG, Hui CC, McInnes RR, Cheng S (2005) The Iroquois homeobox gene, Irx5, is required for retinal cone bipolar cell development. Dev Biol 287:48–60. https:// doi.org/10.1016/j.ydbio.2005.08.029
- Cheng Z, Wang J, Su D, Pan H, Huang G, Li X, Li Z, Shen A, Xie X, Wang B, Ma X (2011) Two novel mutations of the IRX4 gene in patients with congenital heart disease. Hum Genet 130:657–662. https://doi. org/10.1007/s00439-011-0996-7
- Christoffels VM, Keijser AG, Houweling AC, Clout DE, Moorman AFM (2000) Patterning the embryonic heart: identification of five mouse Iroquois homeobox genes in the developing heart. Dev Biol 224:263–274. https:// doi.org/10.1006/dbio.2000.9801
- Costantini DL, Arruda EP, Agarwal P, Kim KH, Zhu Y, Zhu W, Lebel M, Cheng CW, Park CY, Pierce SA, Guerchicoff A, Pollevick GD, Chan TY, Kabir MG, Cheng SH, Husain M, Antzelevitch C, Srivastava D, Gross GJ, Hui CC, Backx PH, Bruneau BG (2005) The homeodomain transcription factor Irx5 establishes the mouse cardiac ventricular repolarization gradient. Cell 123:347–358. https://doi.org/10.1016/j.cell.2005.08. 004
- Date Y, Hasegawa S, Yamada T, Inoue Y, Mizutani H, Nakata S, Akamatsu H (2013) Major amino acids in collagen hydrolysate regulate the differentiation of mouse embryoid bodies. J Biosci Bioeng 116:386–390. https://doi.org/10.1016/j.jbiosc.2013. 03.014
- Dorn T, Goedel A, Lam JT, Haas J, Tian Q, Herrmann F, Bundschu K, Dobreva G, Schiemann M, Dirschinger R, Guo Y, Kühl SJ, Sinnecker D, Lipp P, Laugwitz K-L, Kühl M, Moretti A (2015) Direct Nkx2-5 transcriptional repression of isl1 controls cardiomyocyte subtype identity. Stem Cells 33:1113–1129. https://doi.org/10.1002/stem.1923
- Douville JM, Cheung DY, Herbert KL, Moffatt T, Wigle JT (2011) Mechanisms of MEOX1 and MEOX2 regulation of the cyclin dependent kinase inhibitors p21 and p16 in vascular endothelial cells. PLoS One 6:e29099. https://doi.org/10.1371/journal.pone.0029099
- Dupays L, Shang C, Wilson R, Kotecha S, Wood S, Towers N, Mohun T (2015) Sequential binding of MEIS1 and NKX2-5 on the Popdc2 gene: a mechanism for spatiotemporal regulation of enhancers during cardiogenesis. Cell Rep 13:183–195. https://doi.org/ 10.1016/j.celrep.2015.08.065
- Dyer LA, Kirby ML (2009) The role of secondary heart field in cardiac development. Dev Biol 336:137–144. https://doi.org/10.1016/j.ydbio.2009.10.009
- Elsir T, Smits A, Lindström MS, Nister M (2012) Transcription factor PROX1: its role in development and

cancer. Cancer Metastasis Rev 31:793–805. https://doi. org/10.1007/s10555-012-9390-8

- Espinoza-Lewis RA, Yu L, He F, Liu H, Tang R, Shi J, Sun X, Martin JF, Wang D, Yang J, Chen Y (2009) Shox2 is essential for the differentiation of cardiac pacemaker cells by repressing Nkx2-5. Dev Biol 327:376–385. https://doi.org/10.1021/nl061786n. Core-Shell
- Espinoza-Lewis RA, Liu H, Sun C, Chen C, Jiao K, Chen Y (2011) Ectopic expression of Nkx2.5 suppresses the formation of the sinoatrial node in mice. Dev Biol 356:359–369. https://doi.org/10.1016/j.ydbio.2011. 05.663
- Epstein JA (1996) Pax3, neural crest and cardiovascular development. Trends Cardiovasc Med 6:255–260. https://doi.org/10.1016/S1050-1738(96)00110-7
- Evans AL, Gage PJ (2005) Expression of the homeobox gene Pitx2 in neural crest is required for optic stalk and ocular anterior segment development. Hum Mol Genet 14:3347–3359. https://doi.org/10.1093/hmg/ddi365
- Feng Y, Yang P, Luo S, Zhang Z, Li H, Zhu P, Song Z (2016) Shox2 influences mesenchymal stem cell fate in a co-culture model in vitro. Mol Med Rep 14:637–642. https://doi.org/10.3892/mmr.2016.5306
- Franco D, Sedmera D, Lozano-Velasco E (2017) Multiple roles of Pitx2 in cardiac development and disease. J Cardiovasc Dev Dis 4:16. https://doi.org/10.3390/ jcdd4040016
- Gao XR, Tan YZ, Wang HJ (2011) Overexpression of Csx/Nkx2.5 and GATA-4 enhances the efficacy of mesenchymal stem cell transplantation after myocardial infarction. Circ J 75:2683–2691. https:// doi.org/10.1253/circj.CJ-11-0238
- Garavelli L, Mainardi PC (2007) Mowat-Wilson syndrome. Orphanet J Rare Dis 12:1–12. https://doi.org/ 10.1186/1750-1172-2-42
- Garavelli L, Ivanovski I, Caraffi SG, Santodirocco D, Pollazzon M, Cordelli DM, Abdalla E, Accorsi P, Adam MP, Baldo C, Bayat A, Belligni E, Bonvicini F, Breckpot J, Callewaert B, Cocchi G, Cuturilo G, Devriendt K, Dinulos MB, Djuric O et al (2017) Neuroimaging findings in Mowat – Wilson syndrome: a study of 54 patients. Genet Med 19:691–700. https://doi.org/10.1038/gim.2016.176
- Gheldof A, Hulpiau P, van Roy F, De Craene B, Berx G (2012) Evolutionary functional analysis and molecular regulation of the ZEB transcription factors. Cell Mol Life Sci 69:2527–2541. https://doi.org/10.1007/ s00018-012-0935-3
- Gill HK, Parsons SR, Spalluto C, Davies AF, Knorz VJ, Burlinson CE, Ng B, Carter NP, Ogilvie CM, Wilson DI, Roberts RG (2009) Separation of the PROX1 gene from upstream conserved elements in a complex inversion/translocation patient with hypoplastic left heart. Eur J Hum Genet 17:1423–1431. https://doi.org/10. 1038/ejhg.2009.91
- Gómez-Skarmeta JL, Modolell J (2002) Iroquois genes: genomic organization and function in vertebrate neural development. Curr Opin Genet Dev 12:403–408

- Gong LG, Qiu GR, Jiang H, Xu XY, Zhu HY, Sun KL (2005) Analysis of single nucleotide polymorphisms and haplotypes in HOXC gene cluster within susceptible region 12q13 of simple congenital heart disease. Zhonghua Yi Xue Yi Chuan Xue Za Zhi 22:497–501
- Gu S, Wei N, Yu L, Fei J, Chen Y (2008) Shox2deficiency leads to dysplasia and ankylosis of the temporomandibular joint in mice. Mech Dev 125:729–742. https://doi.org/10.1016/j.mod.2008.04. 003
- Guddati AK, Otero JJ, Kessler E, Aistrup G, Wasserstrom JA, Han X, Lomasney JW, Kessler JA (2009) Embryonic stem cells overexpressing Pitx2 engraft in infarcted myocardium and improve cardiac function. Int Heart J 50:783–799
- Güleç Ç, Abacı N, Bayrak F, Kömürcü Bayrak E, Kahveci G, Güven C, Ünaltuna NE (2014) Association between non-coding polymorphisms of HOPX gene and syncope in hypertrophic cardiomyopathy. Anadolu Kardiyol Derg 14:617–624. https://doi.org/10.5152/ akd.2014.4972
- Guo C, Wang Q, Wang Y, Yang L, Luo H, Cao XF, An L, Qiu Y, Du M, Ma X, Hui L, Lu C (2017) Exome sequencing reveals novel IRXI mutation in congenital heart disease. Mol Med Rep 15:3193–3197. https://doi. org/10.3892/mmr.2017.6410
- Haas J, Frese KS, Park YJ, Keller A, Vogel B, Lindroth AM, Weichenhan D, Franke J, Fischer S, Bauer A, Marquart S, Sedaghat-Hamedani FKE, Köhler D, Wolf NM, Hassel S, Nietsch R, Wieland T, Ehlermann P, Schultz JH, Dösch A, Mereles D, Hardt S, Backs J, Hoheisel JD, Plass C, Katus HA, Meder B (2013) Alterations in cardiac DNA methylation in human dilated cardiomyopathy. EMBO Mol Med 5:413–429. https://doi.org/10.1002/emmm. 201201553
- Hallaq H, Pinter E, Enciso J et al (1998) A null mutation of Hhex results in abnormal cardiac development, defective vasculogenesis and elevated Vegfa levels. Development 131:5197–5209. https://doi.org/10.1242/dev. 01393
- Harvey RP (1996) NK-2 homeobox genes and heart development. Dev Biol 178:203–216
- Harvey RP (2002) Patterning the vertebrate heart. Nat Rev Genet 3:544–556. https://doi.org/10.1038/nrg843
- Hatcher CJ, Diman NY, McDermott DA, Basson CT (2003) Transcription factor cascades in congenital heart malformation. Trends Mol Med 9:512–515
- Hegarty SV, Sullivan AM, Keeffe GWO (2015) Progress in Neurobiology Zeb2: a multifunctional regulator of nervous system development. Prog Neurobiol 132:81–95. https://doi.org/10.1016/j.pneurobio.2015. 07.001
- Hiroi Y, Kudoh S, Monzen K, Ikeda Y, Yazaki Y, Nagai R, Komuro I (2001) Tbx5 associates with Nkx2-5 and synergistically promotes cardiomyocyte differentiation. Nat Genet 28:276–280. https://doi.org/ 10.1038/90123

- Hisa T, Spence SE, Rachel RA, Fujita M, Nakamura T, Ward JM, Devor-Henneman DE, Saiki Y, Kutsuna H, Tessarollo L, Jenkins NA, Copeland NG (2004) Hematopoietic, angiogenic and eye defects in Meis1 mutant animals. EMBO J 23:450–459. https://doi.org/ 10.1038/sj.emboj.7600038
- Hoffmann S, Clauss S, Berger IM, Weiß B, Montalbano A, Röth R, Bucher M, Klier I, Wakili R, Seitz H, Schulze-Bahr E, Katus HA, Flachsbart F, Nebel A, Guenther SP, Bagaev E, Rottbauer W, Kääb S, Just S, Rappold G (2016) Coding and non-coding variants in the SHOX2 gene in patients with early-onset atrial fibrillation. Basic Res Cardiol 111:36. https://doi.org/10.1007/s00395-016-0557-2
- Holland PWH, Booth HAF, Bruford EA (2007) Classification and nomenclature of all human homeobox genes. BMC Biol 5:1–29. https://doi.org/10.1186/ 1741-7007-5-47
- Innis JW (1997) Role of HOX genes in human development. Curr Opin Pediatr 9:617–622
- Ionta V, Liang W, Kim EH, Rafie R, Giacomello A, Marbán E, Cho C (2015) SHOX2 overexpression favors differentiation of embryonic stem cells into cardiac pacemaker cells, improving biological pacing ability. Stem Cell Reports 4:129–142. https://doi.org/ 10.1016/j.stemcr.2014.11.004
- Ismat FA, Zhang M, Kook H, Huang B, Zhou R, Ferrari VA, Epstein JA, Patel VV (2005) Homeobox protein Hop functions in the adult cardiac conduction system. Circ Res 96:898–903. https://doi.org/10.1161/01.RES. 0000163108.47258.f3
- JA E (1996) Pax3, neural crest and cardiovascular development. Trends Cardiovasc Med 6:255–60. https://doi. org/10.1016/S1050-1738(96)00110-7
- Jain R, Li D, Gupta M, Manderfield LJ, Ifkovits JL, Wang Q, Liu F, Liu Y, Poleshko A, Padmanabhan A, Raum JC, Li L, Morrisey EE, Lu MM, Won KJ, Epstein JA (2015) Integration of Bmp and Wnt signaling by Hopx specifies commitment of cardiomyoblasts. Science 348:aaa6071. https://doi.org/10.1016/j.joca. 2015.05.020.Osteoarthritic
- Jensen B, Wang T, Christoffels VM, Moorman AFM (2013a) Evolution and development of the building plan of the vertebrate heart. Biochim Biophys Acta 1833:783–794. https://doi.org/10.1016/j.bbamcr.2012. 10.004
- Jensen B, van den Berg G, van den Doel R, Oostra RJ, Wang T, Moorman AFM (2013b) Development of the hearts of lizards and snakes and perspectives to cardiac evolution. PLoS One 8:e63651. https://doi.org/10. 1371/journal.pone.0063651
- Johansson S, Berland S, Gradek GA, Bongers E, de Leeuw N, Pfundt R, Fannemel M, Rødningen O, Brendehaug A, Haukanes BI, Hovland R, Helland G, Houge G (2014) Haploinsufficiency of MEIS2 is associated with orofacial clefting and learning disability. Am J Med Genet A 164A:1622–1626. https://doi. org/10.1002/ajmg.a.36498

- Jorgensen JS, Gao L (2005) Irx3 is differentially up-regulated in female gonads during sex determination. Gene Expr Pattern 5:756–762. https://doi.org/10. 1016/j.modgep.2005.04.011
- Kang KW, Lee MJ, Song JA, Jeong JY, Kim YK, Lee C, Kim TH, Kwak KB, Kim OJ, An HJ (2014) Overexpression of goosecoid homeobox is associated with chemoresistance and poor prognosis in ovarian carcinoma. Oncol Rep 32:189–198. https://doi.org/10. 3892/or.2014.3203
- Karamboulas C, Swedani A, Ward C, Al-Madhoun AS, Wilton S, Boisvenue S, Ridgeway AG, Skerjanc IS (2006) HDAC activity regulates entry of mesoderm cells into the cardiac muscle lineage. J Cell Sci 119:4305–4314. https://doi.org/10.1242/jcs.03185
- Karns R, Succop P, Zhang G, Sun G, Indugula SR, Havas-Augustin D, Novokmet N, Durakovic Z, Milanovic SM, Missoni S, Vuletic S, Chakraborty R, Rudan P, Deka R (2013) Modeling metabolic syndrome through structural equations of metabolic traits, comorbid diseases, and GWAS variants. Obesity 21:745–754. https://doi.org/10.1002/oby.20445
- Kasahara H, Ueyama T, Wakimoto H, Liu MK, Maguire CT, Converso KL, Kang PM, Manning WJ, Lawitts J, Paul DL, Berul CI, Izumo S (2003) Nkx2.5 homeoprotein regulates expression of gap junction protein connexin 43 and sarcomere organization in postnatal cardiomyocytes. J Mol Cell Cardiol 35:243–256. https://doi.org/10.1016/S0022-2828(03) 00002-6
- Kathiresan S, Srivastava D (2012) Genetics of human cardiovascular disease. Cell 148:1242–1257. https:// doi.org/10.1016/j.cell.2012.03.001
- Kawamura T, Ono K, Morimoto T et al (2005) Acetylation of GATA-4 is involved in the differentiation of embryonic stem cells into cardiac myocytes. J Biol Chem 280:19682–19688. https://doi.org/10.1074/jbc. M412428200
- Kelliny C, Ekelund U, Andersen LB, Brage S, Loos RJ, Wareham NJ, Langenberg C (2009) Common genetic determinants of glucose homeostasis in healthy children: the European Youth Heart Study. Diabetes 58:2939–2945. https://doi.org/10.2337/db09-0374
- Kitajima S, Takagi A, Inoue T, Saga Y (2000) MesP1 and MesP2 are essential for the development of cardiac mesoderm. Development 127:3215–3226
- Koizumi A, Sasano T, Kimura W, Miyamoto Y, Aiba T, Ishikawa T, Nogami A, Fukamizu S, Sakurada H, Takahashi Y, Nakamura H, Ishikura T, Koseki H, Arimura T, Kimura A, Hirao K, Isobe M, Shimizu W, Miura N, Furukawa T (2016) Genetic defects in a His-Purkinje system transcription factor, IRX3, cause lethal cardiac arrhythmias. Eur Heart J 37:1469–1475. https://doi.org/10.1093/eurheartj/ ehv449
- Kook H, Yung WW, Simpson RJ, Kee HJ, Shin S, Lowry JA, Loughlin FE, Yin Z, Epstein JA, Mackay J (2006) Analysis of the structure and function of the

transcriptional coregulator HOP. Biochemistry 45:10584–10590. https://doi.org/10.1021/bi060641s

- Lage K, Møllgård K, Greenway S, Wakimoto H, Gorham JM, Workman CT, Bendsen E, Hansen NT, Rigina O, Roque FS, Wiese C, Christoffels VM, Roberts AE, Smoot LB, Pu WT, Donahoe PK, Tommerup N, Brunak S, Seidman CE, Seidman JG, Larsen LA (2010) Dissecting spatio-temporal protein networks driving human heart development and related disorders. Mol Syst Biol 6:381. https://doi.org/10. 1038/msb.2010.36
- Larroux C, Fahey B, Degnan SM, Adamski M, Rokhsar DS, Degnan BM (2007) The NK homeobox gene cluster predates the origin of Hox genes. Curr Biol 17:706–710. https://doi.org/10.1016/j.cub.2007.03. 008
- Laugwitz KL, Moretti A, Caron L, Nakano A, Chien KR (2007) Islet1 cardiovascular progenitors: a single source for heart lineages. Development 135:193–205. https://doi.org/10.1242/dev.001883
- Lebel M, Agarwal P, Cheng CW, Kabir MG, Chan TY, Thanabalasingham V, Zhang X, Cohen DR, Husain M, Cheng SH, Bruneau BG, Cheng SH (2003) The Iroquois homeobox gene irx2 is not essential for normal development of the heart and midbrain-hindbrain boundary in mice. Mol Cell Biol 23:8216–8225
- Li J, Cao Y, Wu Y, Chen W, Yuan Y, Ma X, Huang G (2015) The expression profile analysis of NKX2-5 knock-out embryonic mice to explore the pathogenesis of congenital heart disease. J Cardiol 66:527–531. https://doi.org/10.1016/j.jjcc.2014.12.022
- Liang X, Wang G, Lin L, Lowe J, Zhang Q, Bu L, Chen Y, Chen J, Sun Y, Evans SM (2013) HCN4 dynamically marks the first heart field and conduction system precursors. Circ Res 113:399–407. https://doi.org/10. 1161/CIRCRESAHA.113.301588
- Liang X, Evans SM, Sun Y (2017) Development of the cardiac pacemaker. Cell Mol Life Sci 74:1247–1259. https://doi.org/10.1007/s00018-016-2400-1
- Lin Q, Schwarz J, Bucana C, Olson EN (1997) Control of mouse cardiac morphogenesis and myogenesis by transcription factor MEF2C. Science 276:1404–1407
- Lin CR, Kioussi C, O'Connell S et al (1999) Pitx2 regulates lung asymmetry, cardiac positioning and pituitary and tooth morphogenesis. Nature 401:279–282. https://doi.org/10.1038/45803
- Lints TJ, Parsons LM, Hartley L, Lyons I, Harvey RP (1993) Nkx-2.5: a novel murine homeobox gene expressed in early heart progenitor cells and their myogenic descendants. Development 119:419–431
- Liu CC, Liu WW, Palie JJ et al (2002) Pitx2c patterns anterior myocardium and aortic arch vessels and is required for local cell movement into atrioventricular cushions. Development 129:5081–5091. https://doi. org/10.1242/dev.00173
- Liu H, Chen CH, Espinoza-Lewis RA, Jiao Z, Sheu I, Hu X, Lin M, Zhang Y, Chen Y (2011) Functional redundancy between human SHOX and mouse Shox2 genes in the regulation of sinoatrial node formation and Pacemaking function *. J Biol Chem

286:17029–17038. https://doi.org/10.1074/jbc.M111. 234252

- Liu Y, Kaneda R, Leja TW, Subkhankulova T, Tolmachov O, Minchiotti G, Schwartz RJ, Barahona M, Schneider M (2014) Hhex and Cer1 mediate the Sox17 pathway for cardiac mesoderm formation in embryonic stem. Stem Cells 32:1515–1526. https://doi.org/10.1002/stem.1695
- Liu Y, Ni B, Lin Y, Chen XG, Fang Z, Zhao L, Hu Z, Zhang F, Ai X (2014) Genetic polymorphisms in ZFHX3 are associated with atrial fibrillation in a Chinese Han population. PLoS One 9:e101318. https:// doi.org/10.1371/journal.pone.0101318
- Louw JJ, Corveleyn A, Jia Y, Hens G, Gewillig M, Devriendt K (2015) MEIS2 involvement in cardiac development, cleft palate, and intellectual disability. Am J Med Genet A 167A:1142–1146. https://doi.org/ 10.1002/ajmg.a.36989
- Lozano-Velasco E, Chinchilla A, Martínez-Fernández S et al (2011) Pitx2c modulates cardiac-specific transcription factors networks in differentiating cardiomyocytes from murine embryonic stem cells. Cells Tissues Organs 194:349–362. https://doi.org/10. 1159/000323533
- Lu D, Wang J, Li J, Guan F, Zhang X, Dong W, Liu N, Gao S, Zhang L (2018) Meox1 accelerates myocardial hypertrophic decompensation through Gata4. Cardiovasc Res 114:300–311. https://doi.org/10. 1093/cvr/cvx222
- Luo ZL, Sun H, Yang ZQ, Ma YH, Gu Y, He YQ, Wei D, Xia LB, Yang BH, Guo T (2014) Genetic variations of ISL1 associated with human congenital heart disease in Chinese Han people. Genet Mol Res 13:1329–1338. https://doi.org/10.4238/2014.February.28.5
- Luxán G, D'Amato G, de la Pompa JL (2016) Endocardial notch signaling in cardiac development and disease. Circ Res 118:1–18. https://doi.org/10.1161/ CIRCRESAHA.115.305350
- Lyons I, Parsons LM, Hartley L, Li R, Andrews JE, Robb L, Harvey RP (1995) Myogenic and morphogenetic defects in the heart tubes of murine embryos lacking the homeo box gene Nkx2–5. Genes Dev 9:1654–1666
- Machon O, Masek J, Machonova O, Krauss S, Kozmik Z (2015) Meis2 is essential for cranial and cardiac neural crest development. BMC Dev Biol 15:40. https://doi. org/10.1186/s12861-015-0093-6
- Mariotto A, Pavlova O, Park HS, Huber M, Sadek HA (2013) Meis1 regulates postnatal cardiomyocyte cell cycle arrest. Nature 497:249–253. https://doi.org/10. 1038/jid.2014.371
- Mariotto A, Pavlova O, Park HS et al (2016) HOPX: the unusual homeodomain-containing protein. J Invest Dermatol 136:905–911. https://doi.org/10.1016/j.jid. 2016.01.032
- Mathieu ML, Demily C, Chantot-Bastaraud S, Afenjar A, Mignot C, Andrieux J, Gerard M, Catala-Mora J, Jouk PS, Labalme A, Edery P, Sanlaville D, Rossi M (2017) Clinical and molecular cytogenetic characterization of four unrelated patients carrying 2p14 microdeletions.

Am J Med Genet A 173:2268–2274. https://doi.org/10. 1002/ajmg.a.38307

- Maves L, Tyler A, Moens CB, Tapscott SJ (2009) Pbx acts with Hand2 in early myocardial differentiation. Dev Biol 333:409–418. https://doi.org/10.1016/j.ydbio. 2009.07.004
- McCulley DJ, Black BL (2012) Transcription factor pathways and congenital heart disease. Curr Top Dev Biol 100:253–277. https://doi.org/10.1016/B978-0-12-387786-4.00008-7.Transcription
- McDonald LA, Gerrelli D, Fok Y, Hurst LD, Tickle C (2010) Comparison of Iroquois gene expression in limbs/fins of vertebrate embryos. J Anat 216:683–691. https://doi.org/10.1111/j.1469-7580. 2010.01233.x
- McElhinney DB, Geiger E, Blinder J, Benson DW, Goldmuntz E (2003) NKX2.5 mutations in patients with congenital heart disease. J Am Coll Cardiol 42:1650–1655
- Moorman A, Webb S, Brown NA et al (2003) Development of the heart: (1) formation of the cardiac chambers and arterial trunks. Heart 89:806–814
- Nayor M, Vasan RS (2015) Preventing heart failure: the role of physical activity. Curr Opin Cardiol 30:543–550. https://doi.org/10.1097/HCO. 000000000000206
- Nemer M (2008) Genetic insights into normal and abnormal heart development. Cardiovasc Pathol 17:48–54. https://doi.org/10.1016/j.carpath.2007.06.005
- Newman CS, Krieg PA (1998) Tinman-related genes expressed during heart development in Xenopus. Dev Genet 22:230–238. https://doi.org/10.1002/(SICI) 1520-6408(1998)22:3<230::AID-DVG5>3.0.CO;2-7
- O'Toole TE, Conklin DJ, Bhatnagar A (2008) Environmental risk factors for heart disease. Rev Environ Health 23:167–202. https://doi.org/10.1038/nrcardio. 2015.152
- Olson EN (2006) Gene regulatory networks in the evolution and development of the heart. Science 313:1922–1927. https://doi.org/10.1126/science. 1132292
- Pearson JC, Lemons D, McGinnis W (2005) Modulating Hox gene functions during animal body patterning. Nat Rev Genet 6:893–904. https://doi.org/10.1038/ nrg1726
- Pechlivanis S, Scherag A, Mühleisen TW, Möhlenkamp S, Horsthemke B, Boes T, Bröcker-Preuss M, Mann K, Erbel R, Jöckel KH, Nöthen MM, Moebus S (2010) Coronary artery calcification and its relationship to validated genetic variants for diabetes mellitus assessed in the Heinz Nixdorf recall cohort. Arterioscler Thromb Vasc Biol 30:1867–1872. https://doi.org/10.1161/ATVBAHA.110.208496
- Petchey LK, Risebro CA, Vieira JM, Roberts T, Bryson JB, Greensmith L, Lythgoe MF, Riley PR (2014) Loss of Prox1 in striated muscle causes slow to fast skeletal muscle fiber conversion and dilated cardiomyopathy. Proc Natl Acad Sci U S A 111:9515–9520. https://doi.org/10.1073/pnas.1406191111

- Powers SE, Taniguchi K, Yen W, Melhuish TA, Shen J, Walsh CA, Sutherland AE, Wotton D (2010) Tgif1 and Tgif2 regulate Nodal signaling and are required for gastrulation. Development 137:249–259. https://doi. org/10.1242/dev.040782
- Pradhan L, Gopal S, Li S, Ashur S, Suryanarayanan S, Kasahara H, Nam H (2016) Intermolecular interactions of cardiac transcription factors NKX2.5 and TBX5. Biochemistry 55:1702–1710. https://doi.org/10.1021/ acs.biochem.6b00171
- Prall OW, Menon MK, Solloway MJ, Watanabe Y, Zaffran S, Bajolle F, Biben C, McBride JJ, Robertson BR, Chaulet H, Stennard FA, Wise N, Schaft D, Wolstein O, Furtado MB, Shiratori H, Chien KR, Hamada H, Black BL, Saga Y, Robertson EJ, Buckingham ME, Harvey RP (2007) An Nkx2-5/ Bmp2/Smad1 negative feedback loop controls heart progenitor specification and proliferation. Cell 128:947–959
- Puskaric S, Schmitteckert S, Mori AD, Glaser A, Schneider KU, Bruneau BG, Blaschke RJ, Steinbeisser H, Rappold G (2010) Shox2 mediates Tbx5 activity by regulating Bmp4 in the pacemaker region of the developing heart. Hum Mol Genet 19:4625–4633. https://doi.org/10.1093/hmg/ddq393
- Ragvin A, Moro E, Fredman D, Navratilova P, Drivenes Ø, Engström PG, Alonso ME, de la Calle Mustienes E, Gómez Skarmeta JL, Tavares MJ, Casares F, Manzanares M, van Heyningen V, Molven A, Njølstad PR, Argenton F, Lenhard B, Becker TS (2010) Longrange gene regulation links genomic type 2 diabetes and obesity risk regions to HHEX, SOX4, and IRX3. Proc Natl Acad Sci U S A 107:775–780. https://doi. org/10.1073/pnas.0911591107
- Reamon-Buettner SM, Hecker H, Spanel-Borowski K, Craatz S, Kuenzel E, Borlak J (2004) Novel NKX2-5 mutations in diseased heart tissues of patients with cardiac malformations. Am J Pathol 164:2117–2125. https://doi.org/10.1016/S0002-9440(10)63770-4
- Risebro CA, Searles RG, Melville A, Athalie AD et al (2009) Prox1 maintains muscle structure and growth in the developing heart. Development 136:495–505. https://doi.org/10.1242/dev.030007
- Risebro CA, Petchey LK, Smart N et al (2012) Epistatic rescue of Nkx2.5 adult cardiac conduction disease phenotypes by prospero-related homeobox protein 1 and HDAC3. Circ Res 111:e19-31. https://doi.org/ 10.1161/CIRCRESAHA.111.260695
- Roux M, Laforest B, Capecchi M, Bertrand N, Zaffran S (2015) Hoxb1 regulates proliferation and differentiation of second heart field progenitors in pharyngeal mesoderm and genetically interacts with Hoxa1 during cardiac outflow tract development. Dev Biol 406:247–258. https://doi.org/10.1016/j.ydbio.2015. 08.015
- Santini MP, Forte E, Harvey RP, Kovacic JC (2016) Developmental origin and lineage plasticity of endogenous cardiac stem cells. Development 4:1242–1258. https://doi.org/10.1242/dev.111591

- Schäfer K, Neuhaus P, Kruse J, Braun T (2003) The homeobox gene Lbx1 specifies a subpopulation of cardiac neural crest necessary for normal heart development. Circ Res 92:73–80
- Schneider MD, Baker AH, Riley P (2015) Hopx and the cardiomyocyte parentage. Mol Ther 23:1420–1422. https://doi.org/10.1038/mt.2015.140
- Schwab K, Hartman HA, Liang HC, Aronow BJ, Patterson LT, Potter SS (2006) Comprehensive microarray analysis of hoxa11/hoxd11 mutant kidney development. Dev Biol 293:540–554. https://doi.org/10.1016/j. ydbio.2006.02.023
- Seifert A, Werheid DF, Knapp SM, Tobiasch E (2015) Role of Hox genes in stem cell differentiation. World J Stem Cells 7:583–595. https://doi.org/10.4252/wjsc. v7.i3.583
- Sepulveda JL, Belaguli N, Nigam V, Chen CY, Nemer M, Schwartz RJ (1998) GATA-4 and Nkx-2.5 coactivate Nkx-2 DNA binding targets: role for regulating early cardiac gene expression. Mol Cell Biol 18:3405–3415
- Shashikant CS, Utset MF, Violette SM, Wise TL, Einat P, Einat MPJ, Schughart K, Ruddle FH (1991) Homeobox genes in mouse development. Crit Rev Eukaryot Gene Expr 1:207–245
- Shiratori H, Yashiro K, Shen MM, Hamada H (2006) Conserved regulation and role of Pitx2 in situs-specific morphogenesis of visceral organs. Development 133:3015–3025. https://doi.org/10.1242/dev.02470
- Skerjanc IS, Petropoulos H, Ridgeway AG, Wilton S (1998) Myocyte enhancer factor 2C and Nkx2-5 up-regulate each other's expression and initiate cardiomyogenesis in P19 cells. J Biol Chem 273:34904–34910. https://doi.org/10.1074/jbc.273.52. 34904
- Smith JG, Newton-Cheh C (2015) Genome-wide association studies of late-onset cardiovascular disease. J Mol Cell Cardiol 83:131–141. https://doi.org/10.1016/j. yjmcc.2015.04.004
- Soibam B, Benham A, Kim J, Weng KC, Yang L, Xu X, Robertson M, Azares A, Cooney AJ, Schwartz RJ, Liu Y (2015) Genome-wide identification of MESP1 targets demonstrates primary regulation over. Stem Cells 33:3254–3265. https://doi.org/10.1002/stem. 2111
- Stankunas K, Shang C, Twu KY, Kao SC, Jenkins NA, Copeland NG, Sanyal M, Selleri L, Cleary ML, Chang C (2008) Pbx/Meis deficiencies demonstrate multigenetic origins of congenital heart disease. Circ Res 103:702–709. https://doi.org/10.1161/ CIRCRESAHA.108.175489
- Stevens KN, Hakonarson H, Kim CE, Doevendans PA, Koeleman BP, Mital S, Raue J, Glessner JT, Coles JG, Moreno V, Granger A, Gruber SB, Gruber PJ (2010) Common variation in ISL1 confers genetic susceptibility for human congenital heart disease. PLoS One 26: e10855. https://doi.org/10.1371/journal.pone.0010855
- Sun R, Liu M, Lu L, Zheng Y, Zhang P (2015) Congenital heart disease: causes, diagnosis, symptoms, and treatments. Cell Biochem Biophys 72:857–860. https://doi.org/10.1007/s12013-015-0551-6

- Tanaka M, Chen Z, Bartunkova S, Yamasaki N, Izumo S (1999) The cardiac homeobox gene Csx/Nkx2.5 lies genetically upstream of multiple genes essential for heart development. Development 126:1269–1280
- Tanaka M, Yamasaki N, Izumo S (2000) Phenotypic characterization of the murine Nkx2.6 homeobox gene by gene targeting. Mol Cell Biol 20:2874–2879
- Trivedi CM, Zhu W, Wang Q et al (2010) Hopx and Hdac2 interact to modulate Gata4 acetylation and embryonic cardiac myocyte proliferation. Dev Cell 19:450–459. https://doi.org/10.1016/j.devcel.2010.08. 012
- Trivedi CM, Cappola TP, Margulies KB, Epstein JA (2011) Homeodomain only protein x is downregulated in human heart failure. J Mol Cell Cardiol 50:1056–1058. https://doi.org/10.1021/nl061786n. Core-Shell
- Tu CT, Yang TC, Tsai HJ (2009) Nkx2.7 and Nkx2.5 function redundantly and are required for cardiac morphogenesis of zebrafish embryos. PLoS One 4:e4249. https://doi.org/10.1371/journal.pone.0004249
- van der Harst P, van Setten J, Verweij N, Vogler G, Franke L, Maurano MT, Wang X, Mateo Leach I, Eijgelsheim M, Sotoodehnia N, Hayward C, Sorice R, Meirelles O, Lyytikäinen LP, Polašek O, Tanaka T, Arking DE, Ulivi S, Trompet S et al (2016) 52 genetic loci influencing myocardial mass. J Am Coll Cardiol 68:1435–1448. https://doi.org/10. 1016/j.jacc.2016.07.729
- van Tuyl M, Liu J, Groenman F, Ridsdale R, Han RN, Venkatesh V, Tibboel D, Post M (2006) Iroquois genes influence proximo-distal morphogenesis during rat lung development. Am J Physiol Lung Cell Mol Physiol 290:L777–L789. https://doi.org/10.1152/ ajplung.00293.2005
- van Weerd JH, Christoffels VM (2016) The formation and function of the cardiac conduction system. Development 143:197–210. https://doi.org/10.1242/dev. 124883
- Vandewalle C, Van Roy F, Berx G (2009) The role of the ZEB family of transcription factors in development and disease. Cell Mol Life Sci 66:773–787. https://doi.org/ 10.1007/s00018-008-8465-8
- Wang J, Klysik E, Sood S, Johnson RL, Wehrens XH, Martin JF (2010) Pitx2 prevents susceptibility to atrial arrhythmias by inhibiting left-sided pacemaker specification. Proc Natl Acad Sci U S A 107:9753–9758. https://doi.org/10.1073/pnas.0912585107
- Wang J, Greene SB, Martin JF (2011) BMP signaling in congenital heart disease: new developments and future directions. Birth Defects Res A Clin Mol Teratol 91:441–448. https://doi.org/10.1002/bdra.20785
- Wang J, Xin YF, Xu WJ, Liu ZM, Qiu XB, Qu XK, Xu L, Li X, Yang Y (2013) Prevalence and spectrum of PITX2c mutations associated with congenital heart disease. DNA Cell Biol 32:708–716. https://doi.org/ 10.1089/dna.2013.2185
- Wang J, Zhang DF, Sun YM, Li RG, Qiu XB, Qu XK, Liu X, Fang WY, Yang YQ (2014) NKX2-6 mutation predisposes to familial atrial fibrillation. Int J Mol Med

34:1581–1590. https://doi.org/10.3892/ijmm.2014.

- Wang Y, Li Y, Guo C, Lu Q, Wang W, Jia Z, Chen P, Ma K, Reinberg D, Zhou C (2016) ISL1 and JMJD3 synergistically control cardiac differentiation of embryonic stem cells. Nucl Acids Res 44:6741–6755. https://doi.org/10.1093/nar/gkw301
- Waraya M, Yamashita K, Katoh H, Ooki A, Kawamata H, Nishimiya HNK, Ema A, Watanabe M (2012) Cancer specific promoter CpG Islands hypermethylation of HOP homeobox (HOPX) gene and its potential tumor suppressive role in pancreatic carcinogenesis. BMC Cancer 12:397. https://doi.org/10.1186/1471-2407-12-397
- Wei D, Gong XH, Qiu G, Wang J, Yang Y (2014) Novel PITX2c loss-of-function mutations associated with complex congenital heart disease. Int J Mol Med 33:1201–1208. https://doi.org/10.3892/ijmm.2014. 1689
- Witzel HR, Jungblut B, Choe CP, Crump JG, Braun T, Crump JG (2012) The LIM protein Ajuba restricts the second heart field progenitor pool by regulating Isl1 activity. Dev Cell 23:58–70. https://doi.org/10.1016/j. devcel.2012.06.005
- Wu YH, Zhao H, Zhou LP, Zhao CX, Wu YF, Zhen LX, Li J, Ge DX, Xu L, Lin L, Liu Y, Liang DD, Chen Y (2015) miR-134 modulates the proliferation of human cardiomyocyte progenitor cells by targeting Meis2. Int J Mol Sci 6224:25199–25213. https://doi.org/10.3390/ ijms161025199
- Xu H, Baldini A (2007) Genetic pathways to mammalian heart development: recent progress from manipulation of the mouse genome. Semin Cell Dev Biol 18:77–83. https://doi.org/10.1016/j.semcdb.2006.11.011
- Xu H, Yi Q, Yang C, Wang Y, Tian J, Zhu J (2016) Histone modifications interact with DNA methylation at the GATA4 promoter during differentiation of mesenchymal stem cells into cardiomyocyte-like cells. Cell Prolif 49:315–329. https://doi.org/10.1111/cpr.12253
- Yamagishi H, Yamagishi C, Nakagawa O, Harvey RP, Olson EN, Srivastava D (2001) The combinatorial activities of Nkx2.5 and dHAND are essential for cardiac ventricle formation. Dev Biol 239:190–203. https://doi.org/10.1006/dbio.2001.0417
- Yap LF, Lai SL, Patmanathan SN et al (2016) HOPX functions as a tumour suppressor in head and neck cancer. Sci Rep 6:38758. https://doi.org/10.1038/ srep38758
- Yousef MS, Matthews BW (2005) Structural basis of Prospero-DNA interaction: implications for transcription regulation in developing cells. Structure 13:601–607. https://doi.org/10.1016/j.str.2005.01.023

- Yu X, St Amand TR, Wang S, Li G, Zhang Y, Hu YP, Nguyen L, Qiu MS, Chen Y (2001) Differential expression and functional analysis of Pitx2 isoforms in regulation of heart looping in the chick. Development 1013:1005–1013
- Yu Z, Kong J, Pan B, Sun H, Lv T, Zhu J, Huang G, Tian J (2013) Islet-1 may function as an assistant factor for histone acetylation and regulation of cardiac development-related transcription factor Mef2c expression. PLoS One 8:e77690. https://doi.org/10. 1371/journal.pone.0077690
- Zakariyah AF, Rajgara RF, Veinot JP, Skerjanc IS, Burgon PG (2017) Congenital heart defect causing mutation in Nkx2.5 displays in vivo functional deficit. J Mol Cell Cardiol 105:89–98. https://doi.org/10.1016/ j.yjmcc.2017.03.003
- Zhang SS, Kim KH, Rosen A, Smyth JW, Sakuma R, Delgado-Olguín P, Davis M, Chi NC, Puviindran V, Gaborit N, Sukonnik T, Wylie JN, Brand-Arzamendi-K, Farman GP, Kim J, Rose RA, Marsden PA, Zhu Y, Zhou YQ, Miquerol L, Henkelman RM, Stainier DY, Shaw RM, Hui CC, Bruneau BG, Backx PH (2011) Iroquois homeobox gene 3 establishes fast conduction in the cardiac His-Purkinje network. Proc Natl Acad Sci U S A 108:13576–13581. https://doi.org/10.1073/ pnas.1106911108
- Zhang Y, Si Y, Ma N, Mei J (2015) The RNA-binding protein PCBP2 inhibits Ang II-induced hypertrophy of cardiomyocytes though promoting GPR56 mRNA degeneration. Biochem Biophys Res Commun 464:679–684. https://doi.org/10.1016/j.bbrc.2015.06.139
- Zhang Y, Si Y, Ma N (2016) Meis1 promotes poly (rC)binding protein 2 expression and inhibits angiotensin II-induced cardiomyocyte hypertrophy. IUBMB Life 68:13–22. https://doi.org/10.1002/iub.1456
- Zhou Z, Wang J, Guo C, Chang W, Zhuang J, Zhu P, Li X (2017) Temporally distinct Six2-positive second heart field progenitors regulate mammalian heart development and disease. Cell Rep 18:1019–1032. https://doi. org/10.1016/j.celrep.2017.01.002
- Zhu CC, Yamada G, Nakamura S, Terashi T, Schweickert A, Blum M (1998) Malformation of trachea and pelvic region in goosecoid mutant mice. Dev Dyn 211:374–381. https://doi.org/10.1002/(SICI) 1097-0177(199804)211:4<374::AID-AJA8>3.0. CO:2-E
- Zhuang S, Zhang Q, Zhuang T, Evans SM, Liang X, Sun Y (2013) Expression of Isl1 during mouse development. Gene Expr Patterns 13:407–412. https://doi.org/ 10.1016/j.gep.2013.07.001.Expression