

Neutrophils: The Role of Oxidative and Nitrosative Stress in Health and Disease

Aneta Manda-Handzlik and Urszula Demkow

Abstract

Neutrophils constitute the first line of the innate immunity in humans. They employ several strategies to trap and kill microorganisms, such as phagocytosis, degranulation, and the formation of extracellular traps (NETs). It has been well documented, that generation of reactive oxygen and nitrogen species (ROS and RNS) is crucial in the life cycle of a polymorphonuclear phagocyte. These compounds due to high reactivity act as powerful antimicrobial factors in the process of pathogens clearance and can also modulate immunological response. On the other hand, excessive amount of free radicals may have detrimental effect on host tissues and markers of oxidative and nitrosative stress are detectable in many diseases. It is necessary to maintain the balance between ROS/RNS formation and removal. The review highlights our current understanding of the role of ROS and RNS produced by neutrophils in health and disease.

Keywords

Cell signaling • Nitrosative stress • Neutrophils • Oxidative stress • Pathology

A. Manda-Handzlik
Department of Laboratory Diagnostics and Clinical Immunology of Developmental Age, Warsaw Medical University, 24 Marszałkowska St., 00-576 Warsaw, Poland

Postgraduate School of Molecular Medicine, Warsaw Medical University, Warsaw, Poland

U. Demkow (✉)
Department of Laboratory Diagnostics and Clinical Immunology of Developmental Age, Warsaw Medical University, 24 Marszałkowska St., 00-576 Warsaw, Poland
e-mail: urszula.demkow@litewska.edu.pl

1 Introduction

Neutrophils represent the first line of defense against pathogens and they are recruited as firsts to sites of infection and tissue damage. This most abundant subpopulation of leukocytes is released to venous sinuses of the bone marrow and the marrow reserve is estimated at 6×10^{11} cells. Neutrophils are released into the blood stream, where they constitute the circulating and marginating pools of granulocytes, both of approximately equal size. Neutrophils have

short circulating half-life (6–8 h) and they quickly migrate to tissues where they perform immune functions (Brinkmann et al. 2004; Summers et al. 2010).

Neutrophils react with pathogens in a non-specific way and they employ several strategies to combat invading microorganisms. Mature neutrophils can phagocyte microbes. Whereas macrophages efficiently engulf pathogens in body fluids and on tissue surfaces, polymorphonuclear leukocytes engulf only surface-associated and not fluid-borne bacteria (Colucci-Guyon et al. 2011). The repertoire of neutrophil mechanisms to fight infections includes degranulation and the generation of reactive oxygen species in the process called oxidative burst. During degranulation, a neutrophil releases potent antimicrobial factors normally stored within intracellular granules, such as elastase or gelatinase (Simard et al. 2010). In 2004 a novel mechanism employed by neutrophils was described, consisting of releasing structures resembling nets. These structures are called neutrophil extracellular traps (NET) and are formed by nuclear DNA, histones, and granular and cytoplasmic antimicrobial proteins (Brinkmann et al. 2004; Urban et al. 2009). NETs physically trap bacteria, prevent them from spreading, and provide a high local concentration of antimicrobial factors which can kill pathogens or downgrade their virulence (Brinkmann et al. 2004). Nonetheless, it has been pointed out that the formation of NETs may have not only beneficial but also detrimental consequences as they participate in the pathogenesis of several inflammatory and autoimmune diseases (Brill et al. 2012; Cools-Lartigue et al. 2013; Gupta et al. 2007; Papayannopoulos et al. 2011). It seems that generation of reactive oxygen species (ROS) plays a crucial role in the formation of NETs. Thus, ROS's involvement in NETosis has been an area of extensive study of late.

Among many substances produced by polymorphonuclear granulocytes, a special role is attributed to ROS and reactive nitrogen species (RNS), released during oxidative burst, being not

only antimicrobial factors but also important regulators of immune response. Reactive species have ideal characteristics to fulfill this function, as their small molecules can easily diffuse through cell membranes. Additionally, they are rapidly produced and degraded (Tonks 2005). It has been shown that these molecules can regulate immune response on a very basic, molecular level. On the other hand, ROS and RNS produced by neutrophils were shown to play a crucial role in the pathogenesis of several diseases. Further studies are warranted to understand the link between ROS and RNS and immunity, to develop new therapies targeting these specific cell signaling pathways.

2 Oxidative Burst

Respiratory or oxidative burst is the mechanism in which activated neutrophils (or other cell types, as these mechanisms is not limited to these phagocytes) generate and release reactive oxygen intermediates. Reactive oxygen species can be both radicals, like superoxide ($O_2^{\bullet-}$), hydroxyl ($\bullet OH$) radical, and nonradicals, like hydrogen peroxide or hypochlorous acid. ROS formation is a critical step in phagocytosis, as these highly reactive compounds can kill pathogens within phagolysosomes.

A key enzyme in this process is the nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase, NOX) (Fig. 1). It consists of six subunits, which are separated under physiological conditions to prevent ROS generation and consequent cell damage. Before the complex is activated and produces large amounts of oxidative agents, the subunits have to assemble to form multicomponent enzyme. Two of these proteins, $p22^{phox}$ and $p91^{phox}$, are located within the cell membrane and together build cytochrome b558. Cytochrome b558 is the catalytic core of the enzyme, as electron transfer occurs through $gp91^{phox}$. After neutrophil activation, three regulatory subunits ($p67^{phox}$, $p47^{phox}$, and $p40^{phox}$) are phosphorylated and along with GTPase Rac2 are dislocated from cytosol to phagolysosome membrane, where active NADPH oxidase

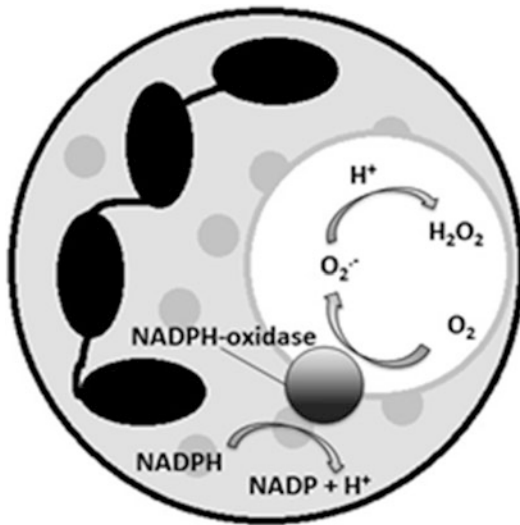


Fig. 1 Function of NADPH oxidase in oxidative burst of neutrophils

is formed (Knaus et al. 1991). NOX is capable of transferring electrons from NADPH to O₂ to form superoxide. Patients bearing mutations in genes for NOX subunits suffer from recurrent infections due to inability of neutrophils to generate oxidative stress during immunological response. The most common mutation in patients with this condition, chronic granulomatous disease (CGD), is a mutation in the gene encoding the protein p91^{phox}, observed in two thirds of all the cases (Patino et al. 1999).

Superoxide is not the only reactive oxygen species formed during oxidative burst, as further cascade of chemical reactions leads to the generation of other oxidative agents. Superoxide may dismutate in the reaction either catalyzed by superoxide dismutase (SOD) or spontaneously into hydrogen peroxide. It is possible to generate hydroxyl radical from hydrogen peroxide in the Fenton reaction, catalyzed by iron, although this mechanism is deficient in neutrophils as they lack ferrous ions (Britigan et al. 1990). After neutrophils' activation azurophilic granules release myeloperoxidase (MPO) into the phagosome, where MPO catalyzes the synthesis of hypochlorous acid from hydrogen peroxide as a substrate. HOCl then reacts with amino acids, giving chloramines. Furthermore,

superoxide may react with hypochlorous acid, which results in the formation of hydroxyl radical (Ramos et al. 1992). In the reaction between hydrogen peroxide and hypochlorous acid singlet oxygen is formed. Another source of ROS is the reaction between hydrogen peroxide and superoxide, but low amounts of synthesized singlet oxygen and hydroxyl radical seem not to be of high importance in biological systems (MacManus-Spencer and McNeill 2005).

Generation of ROS is supported by the production of RNS, which are also involved in the clearance of pathogens. Enhanced synthesis of nitric oxide (NO) is associated with the process of inflammation, when mRNA for inducible form of nitric oxide synthase (iNOS) is being transcribed. The substrates for NO synthesis are L-arginine, NADPH, and oxygen, with the tetrahydrobiopterin as a cofactor. NO diffuses easily across cellular membranes and its antimicrobial properties are significantly enhanced when it reacts with superoxide, forming peroxynitrite, a potent pro-inflammatory and cytotoxic molecule (Cuzzocrea et al. 2001). Other RNS that might be formed during reactions of endogenous NO with oxidants are nitrogen dioxide and dinitrogen trioxide. Both ROS and RNS are strong oxidants and these compounds can react with many biomolecules including DNA, proteins, lipids, and thiols. Reactions of oxidation and nitration/nitrosylation result in the formation of toxic products and, in turn, they facilitate clearance of pathogens (Eiserich et al. 1998).

3 ROS and RNS in Chemotaxis, Neutrophil Adhesion, Rolling, and Phagocytosis

It has been observed that ROS are crucial regulators of neutrophil chemotaxis. In the process of chemotaxis, a neutrophil forms pseudopods. The well-directed pseudopods are maintained, whereas pseudopods facing the wrong direction are destroyed enabling migration toward chemoattractants. It has been proposed that disruption of misoriented

pseudopods may be, in large part, mediated by ROS production (Hattori et al. 2010).

As mentioned above, compounds released during oxidative burst are able to move across cell membranes. Therefore, even if produced within cell, they may affect processes occurring extracellularly. Neutrophils migrating to tissues release ROS, which, in turn, activate endothelial cells (Fialkow et al. 2007). It creates a specific loop, as ROS induce the prolonged expression of the glycoprotein ligand for polymorphonuclear adhesion GMP140 on endothelial cell surface. ROS are able to regulate the expression of endothelial cell adhesion molecules (CAM) by a direct activation of CAMs on the surface and also by a transcription-dependent mechanism which involves transcription factors sensitive to oxidation (Patel et al. 1991).

Not only reactive oxygen species but also nitrogen species may regulate endothelium – neutrophil adhesion. Depletion of NO causes increased oxidative stress within endothelium and mast cells. It results in a promotion of neutrophil adhesion to the blood vessel wall (Niu et al. 1996). Other studies revealed that treatment of neutrophils with NOS inhibitors enhanced carrageenan-induced rolling/adhesion of neutrophils to endothelial cells. Additionally, NOS inhibitors block the apoptosis process in migrating neutrophils (Dal Secco et al. 2003). Moreover, NO can modulate migration of neutrophils by inducing neutrophil-derived microparticles (Nolan et al. 2008).

On the surface of neutrophils there are two types of Fc receptors: FcγRIIa and FcγRIIIb. They are responsible for phagocytosis and killing of opsonized pathogens within the cell. Activation of FcγRIIa may be induced by crosslinking of FcγRIIIb, which, in turn, leads to neutrophil degranulation and generation of reactive oxygen intermediates (Salmon et al. 1995). Under physiological conditions, respiratory burst is involved in amplification of Fc receptor-mediated phagocytic function and in patients suffering from chronic granulomatous disease Fc receptor-mediated phagocytosis is abrogated. This pathway requires hydrogen peroxide, lactoferrin, and superoxide anion, but not hydrogen peroxide-

MPO-halide system, which underscores the pivotal role of ROS in the amplification of phagocytosis (Gresham et al. 1988). Interestingly, it has been recently shown that inducible nitric oxide synthase interacts with Rac2 and is translocated to phagosomes after phagocytosis. As a consequence, ROS and RNS are produced to kill microbes within this cellular compartment (Jyoti et al. 2014).

4 Role of Oxidative Stress in NETs Formation

Convincing evidence on the role of ROS in NETosis has come from studies on neutrophils isolated from CGD-patients. Lack of NADPH-oxidase in cell membrane impairs the production and release of NETs in response to different stimuli (Fuchs et al. 2007). In line with those findings, murine model of CGD confirmed this observations as neutrophils isolated from gp91 –/– mice do not release extracellular traps (Ermert et al. 2009). In the same study, NET formation interrelated with the amount of ROS produced. It has been shown that the mechanism underlying ROS-induced NETosis is based upon translocation of neutrophil elastase (NE) to the nucleus, where it degrades histones and facilitates chromatin decondensation. Myeloperoxidase acts synergistically with NE in this process. Further support for the role of ROS in NETosis comes from the fact that phorbol myristate acetate (PMA) – a potent stimulant of NETosis becomes inactive when exposed to diphenyleneiodonium chloride (DPI), an NADPH oxidase inhibitor (Papayannopoulos et al. 2010). Some studies were undertaken to establish the clear role of different oxidants in NETosis. Due to difficulties with targeting appropriate cell compartment using oxidant scavengers and enzymes inhibitors, and multiple reactions involved in the generation of ROS, there are no apparent conclusions (Parker and Winterbourn 2012). Yet, it has been shown that mitochondrial ROS are not involved in NETosis (Kirchner et al. 2012).

Notably, in some cases of NETs formation, ROS are not involved. For instance, in cystic fibrosis airway inflammation traps may be released upon activation of G protein-coupled receptor CXCR2 and this process is independent of NADPH oxidase (Marcos et al. 2010). Furthermore, early formation of NETs observed after exposure to *S. aureus* is also independent of ROS produced by NADPH oxidase (Pilszczek et al. 2010). Accordingly, it is possible that ROS might account for most but not all NETs.

To date, the role of NO and nitrosative stress in NETs formation has not been largely studied. However, it has been recently discovered that NO is an inducer of NETosis and acts *via* augmenting free radical generation. It has been suggested that oxidative stress and iNOS induction may act in concert during inflammation to promote NETosis (Patel et al. 2010).

5 Reactive Oxygen and Nitrogen Species in Cell Signaling Pathways

Nuclear factor kappa B (NF- κ B) is a transcription factor that once activated regulates the process of transcription of numerous genes. Their products (e.g., cytokines, protein p53, growth factors, and cell adhesion molecules) are involved in cell cycle regulation, inflammation, and immune response (Celec 2004). The influence of ROS on NF- κ B regulation varies in different compartment of a cell. ROS in the cytoplasm stimulate the signal transduction pathways for NF- κ B activation and translocation into the nucleus. On the other hand, ROS inhibit the activity of this transcription factor in the nucleus. Oxidative and nitrosative stress may regulate NF- κ B pathway in many ways. The sulphhydryl group of Cys62 in NF- κ B molecule is a determinant of DNA recognition by the p50 subunit and it is especially sensitive to oxidation (Matthews et al. 1993). It cannot be excluded that oxidation is followed by the S-glutathiolation, as changes in the GSH/GSSG ratio are responsible for inhibition of binding of p50 subunit to DNA (Klatt and Lamas 2000). It has also been shown

that NF- κ B/RelA Ser(276) phosphorylation induced by tumor necrosis factor- α (TNF- α), a modification critical for NF- κ B transcriptional activity, is mediated by the ROS signaling pathway (Jamaluddin et al. 2007).

Several studies highlighted that ROS affect the phosphorylation of I- κ B α and independently induce NF- κ B activation and translocation to the nucleus (Schoonbroodt et al. 2000; Takada et al. 2003). Contradictory findings on the influence of ROS on IKK have been reported. Some studies show that ROS activate IKK kinase complex, the core element of the NF- κ B cascade, while others found that ROS may have an inhibitory effect on this kinase (Byun et al. 2002; Herscovitch et al. 2008). Moreover, catalytically active subunit of IKK is a site for S-nitrosylation by nitric oxide. Following inhibition of NF- κ B provides an explanation for anti-inflammatory effect of NO (Reynaert et al. 2004). As mentioned above, release of NO down-regulates neutrophil migration, as it decreases rolling and adhesion of neutrophils to endothelium and induces apoptosis (Dal Secco et al. 2003).

Oxidants may also modulate cell signaling pathway involving the mitogen-activated protein kinase (MAPK) family. They are activated to induce the production of cytokines, like IL-6, and inflammatory mediators (Craig et al. 2000; Lang et al. 2006). Exogenous oxidants induce tyrosine phosphorylation and activation of ERK, which is a member of MAPK. This effect is enhanced by the inhibition of CD45, which also is a result of oxidative influence. Oxidized CD45 cannot efficiently dephosphorylate and inactivate MAP kinase (Fialkow et al. 1994). It has been recently demonstrated that PMA activates both extracellular signal-regulated kinases (ERK) and p38 MAPK and the process of phosphorylation of these two kinases is ROS-dependent (Keshari et al. 2013).

ROS are capable of regulating the activity of protein tyrosine kinases and phosphatases. The balance between these two activities is responsible for the level of tyrosine phosphorylation. Signaling pathways involving these enzymes are of great importance for neutrophil inflammatory response and its components, such

as adherence, chemotaxis, and phagocytosis (Kobayashi et al. 1995). Invariant cysteine residue in protein tyrosine phosphatase superfamily (PTP) functions as a nucleophile during catalysis. The active site Cys may be oxidized, which abrogates its nucleophilic properties and inhibits PTP activity. When oxidation by ROS occurs upon activation, it leads to inactivation of PTP and facilitates the response to the activating stimulus (Rhee et al. 2003). It has also been found that oxidative burst can inhibit expression of CD45, a tyrosine phosphatase commonly found on the surface of leukocytes. This inhibition is, at least partially, mediated by NOX (Fialkow et al. 1997). Cysteine residue is also a target for oxidants in protein kinase C (PKC). This family of proteins is responsible for regulation of cell growth, differentiation, apoptosis, transformation, and tumorigenesis. Cysteine is located within the zinc-binding domain of autoinhibitory function and when oxidized it leads to the activation of PKC (Cosentino-Gomes et al. 2012).

6 Reactive Oxygen/Nitrogen Species and Neutrophils in Pathology

It is known that patients with human immunodeficiency virus (HIV) suffer from systemic oxidative stress. It leads to the activation of NF- κ B and, as a consequence, increases the HIV replication (Nakamura et al. 2002). Increased ROS production is due to changes in the expression of Bcl-2 and thioredoxin, antiapoptotic and antioxidant molecules. Spontaneously increased production of ROS by phagocytes (both macrophages and neutrophils generate high amounts of H₂O₂) can participate in the oxidative injury in the course of HIV infection. Despite the production of hydrogen peroxide by neutrophils, these cells show decreased release of ROS after the specific stimulation when exposed to oxidative burst inducers such as TNF- α , IL-8, or bacterial N-formyl peptides. That may contribute to the enhanced susceptibility to bacterial infections in HIV-infected patients (Elbim et al. 2001).

In patients suffering from acute respiratory distress syndrome, bronchoalveolar lavage contains a high number of neutrophils, which are a source of inflammatory mediators and superoxide anion. Peroxynitrite generated from NO and superoxide, nitrates, and oxidized amino acids inhibit function of proteins, such as surfactant protein A. Improper balance between antioxidants and reactive oxygen and nitrogen species aggravates the disease (Lang et al. 2002). Oxidant/antioxidant imbalance is observed also in patients with chronic obstructive pulmonary disease (COPD). Numerous neutrophils in the airways of these patients produce oxidants and the amount of free radicals is additionally increased in patients smoking cigarettes. Evidence for enhanced oxidative stress in smoking patients are also supported by the presence of nitrated proteins: ceruloplasmin, plasminogen, fibrinogen and transferrin, as well as elevated levels of lipid peroxidation products in blood (Morrison et al. 1999; Pignatelli et al. 2001). Oxidants can activate NF- κ B, which, in turn, causes transcription of inflammation-related mediators. That results in increased sputum concentration of TNF- α , nitric oxide, and IL-8. Oxidation also silences α -1-antitrypsin, disturbing the balance between proteases and antiproteases, which causes tissue injury (Dalle-Donne et al. 2005). It has been shown that during acute exacerbation of COPD the production of superoxide by neutrophils is increased (Rahman et al. 1997).

In rheumatoid arthritis, neutrophils present in inflamed joints produce both ROS and RNS, which can exacerbate the disease. Nitrotyrosine, which is a marker of peroxynitrate production, is elevated in serum and synovial fluid collected from patients with the disease and the level of 3-nitrotyrosine correlates directly with disease activity (Kaur and Halliwell 1994; Khan and Siddiqui 2006). The level of S-nitrosoproteins in synovial fluid is especially high when compared with plasma concentrations (Hilliquin et al. 1997). This supports a notion of NO being formed by inflammatory cells within the joint. Superoxide and hydroxyl radicals are also significantly

raised in peripheral blood and synovial infiltrate collected from rheumatoid arthritis patients. ROS can serve as an indirect measure of the intensity of inflammation (Kundu et al. 2012). Similarly, superoxide dismutase activity is higher in the joint fluid of these patients than in healthy subjects and its activity may serve as a valid index of articular destruction and repair (Sumii et al. 1996).

Patients with ulcerative colitis have an elevated neutrophil count when compared with healthy subjects (Hanai et al. 2004). Moreover, polymorphonuclear granulocytes accumulate in the epithelial crypts of the intestinal mucosa in inflamed bowel. Increased production of ROS by neutrophils leads to tissue destruction (Ramonaitė et al. 2013). It has been suggested that oxidative stress damages DNA in a damage-regeneration cycle and enhances the risk of carcinogenesis in ulcerative colitis patients (Seril et al. 2003). In patients with Crohn's disease, neutrophils respond to stimulation less than in healthy subjects. Superoxide and lysozyme release by neutrophils, a possible mechanism limiting the extent of inflammation in the intestinal wall, is significantly reduced in patients with active Crohn's disease (Maor et al. 2008).

ROS may also participate in the pathogenesis of preeclampsia, where oxidative stress contributes to endothelial dysfunction. It is suggested that generation of ROS and RNS is caused by reduced blood flow through the placenta. Neutrophils, in line with macrophages, can be activated by oxidative stress, inflammatory agents, or hypoxic conditions during their passage through placental blood vessels. Once activated, polymorphonuclear granulocytes are a source of ROS (Dalle-Donne et al. 2005). Neutrophils are more intensively adhering to the endothelium and infiltrating the intimal space of systemic blood vessels in preeclamptic compared to normal pregnant and non-pregnant women (Cadden and Walsh 2008). It has been shown that supplementation with vitamins C and E in women at increased risk of preeclampsia may be of benefit (Chappell et al. 1999).

7 Conclusions

The role of reactive oxygen and nitrogen species in the physiology of neutrophils is complex. These compounds are potent antimicrobial factors killing bacteria *via* oxidation, nitration, and nitrosylation. It is also evident that these cells regulate the immune response and the intercellular communication. Nonetheless, production of oxidants by polymorphonuclear phagocytes has been shown to exacerbate several diseases. Thus, further investigation is warranted to discern pathologies that dependent on nitrosative and oxidative insults.

Conflicts of Interest The authors declare no conflicts of interest in relation to this article.

References

- Brill A, Fuchs TA, Savchenko AS, Thomas GM, Martinod K, De Meyer SF, Bhandari AA, Wagner DD (2012) Neutrophil extracellular traps promote deep vein thrombosis in mice. *J Thromb Haemost* 10:136–144
- Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y, Zychlinsky A (2004) Neutrophil extracellular traps kill bacteria. *Science* 303:1532–1535
- Britigan BE, Coffman TJ, Buettner GR (1990) Spin trapping evidence for the lack of significant hydroxyl radical production during the respiration burst of human phagocytes using a spin adduct resistant to superoxide-mediated destruction. *J Biol Chem* 265:2650–2656
- Byun MS, Jeon KI, Choi JW, Shim JY, Jue DM (2002) Dual effect of oxidative stress on NF-kappaB activation in HeLa cells. *Exp Mol Med* 34:332–339
- Cadden KA, Walsh SW (2008) Neutrophils, but not lymphocytes or monocytes, infiltrate maternal systemic vasculature in women with preeclampsia. *Hypertens Pregnancy* 27:396–405
- Celec P (2004) Nuclear factor kappa B-molecular biomedicine: the next generation. *Biomed Pharmacother* 58:365–371
- Chappell LC, Seed PT, Briley AL, Kelly FJ, Lee R, Hunt BJ, Parmar K, Bewley SJ, Shennan AH, Steer PJ, Poston L (1999) Effect of antioxidants on the occurrence of pre-eclampsia in women at increased risk: a randomised trial. *Lancet* 354:810–816
- Colucci-Guyon E, Tinevez JY, Renshaw SA, Herbomel P (2011) Strategies of professional phagocytes in vivo: unlike macrophages, neutrophils engulf only surface-associated microbes. *J Cell Sci* 124:3053–3059

- Cools-Lartigue J, Spicer J, McDonald B, Gowing S, Chow S, Giannias B, Bourdeau F, Kubes P, Ferri L (2013) Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis. *J Clin Invest* 123:3446–3458
- Cosentino-Gomes D, Rocco-Machado N, Meyer-Fernandes JR (2012) Cell signaling through protein kinase C oxidation and activation. *Int J Mol Sci* 13:10697–10721
- Craig R, Larkin A, Mingo AM, Thuerlauf DJ, Andrews C, McDonough PM, Glembotski CC (2000) p38 MAPK and NF-kappa B collaborate to induce interleukin-6 gene expression and release. Evidence for a cytoprotective autocrine signaling pathway in a cardiac myocyte model system. *J Biol Chem* 275:23814–23824
- Cuzzocrea S, Mazzon E, Dugo L, Caputi AP, Aston K, Riley DP, Salvemini D (2001) Protective effects of a new stable, highly active SOD mimetic, M40401 in splanchnic artery occlusion and reperfusion. *Br J Pharmacol* 132:19–29
- Dal Secco D, Paron JA, de Oliveira SH, Ferreira SH, Silva JS, Cunha Fde Q (2003) Neutrophil migration in inflammation: nitric oxide inhibits rolling, adhesion and induces apoptosis. *Nitric Oxide* 9:153–164
- Dalle-Donne I, Scaloni A, Giustarini D, Cavarra E, Tell G, Lungarella G, Colombo R, Rossi R, Milzani A (2005) Proteins as biomarkers of oxidative/nitrosative stress in diseases: the contribution of redox proteomics. *Mass Spectrom Rev* 24:55–99
- Eiserich JP, Patel RP, O'Donnell VB (1998) Pathophysiology of nitric oxide and related species: free radical reactions and modification of biomolecules. *Mol Aspects Med* 19:221–357
- Elbim C, Pillet S, Prevost MH, Preira A, Girard PM, Rogine N, Hakim J, Israel N, Gougerot-Pocidallo MA (2001) The role of phagocytes in HIV-related oxidative stress. *J Clin Virol* 20:99–109
- Ermert D, Urban CF, Laube B, Goosmann C, Zychlinsky A, Brinkmann V (2009) Mouse neutrophil extracellular traps in microbial infections. *J Innate Immun* 1:181–193
- Fialkow L, Chan CK, Rotin D, Grinstein S, Downey GP (1994) Activation of the mitogen-activated protein kinase signaling pathway in neutrophils. Role of oxidants. *J Biol Chem* 269:31234–31242
- Fialkow L, Chan CK, Downey GP (1997) Inhibition of CD45 during neutrophil activation. *J Immunol* 158:5409–5417
- Fialkow L, Wang Y, Downey GP (2007) Reactive oxygen and nitrogen species as signaling molecules regulating neutrophil function. *Free Radic Biol Med* 42:153–164
- Fuchs TA, Abed U, Goosmann C, Hurwitz R, Schulze I, Wahn V, Weinrauch Y, Brinkmann V, Zychlinsky A (2007) Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol* 176:231–241
- Gresham HD, McGarr JA, Shackelford PG, Brown EJ (1988) Studies on the molecular mechanisms of human Fc receptor-mediated phagocytosis. Amplification of ingestion is dependent on the generation of reactive oxygen metabolites and is deficient in polymorphonuclear leukocytes from patients with chronic granulomatous disease. *J Clin Invest* 82:1192–1201
- Gupta AK, Hasler P, Holzgreve W, Hahn S (2007) Neutrophil NETs: a novel contributor to preeclampsia-associated placental hypoxia? *Semin Immunopathol* 29:163–167
- Hanai H, Takeuchi K, Iida T, Kashiwagi N, Saniabadi AR, Matsushita I, Sato Y, Kasuga N, Nakamura T (2004) Relationship between fecal calprotectin, intestinal inflammation, and peripheral blood neutrophils in patients with active ulcerative colitis. *Dig Dis Sci* 49:1438–1443
- Hattori H, Subramanian KK, Sakai J, Luo HR (2010) Reactive oxygen species as signaling molecules in neutrophil chemotaxis. *Commun Integr Biol* 3:278–281
- Herscovitch M, Comb W, Ennis T, Coleman K, Yong S, Armstead B, Kalaitzidis D, Chandani S, Gilmore TD (2008) Intermolecular disulfide bond formation in the NEMO dimer requires Cys54 and Cys347. *Biochem Biophys Res Commun* 367:103–108
- Hilliquin P, Borderie D, Hervann A, Menkes CJ, Ekindjian OG (1997) Nitric oxide as S-nitrosoproteins in rheumatoid arthritis. *Arthritis Rheum* 40:1512–1517
- Jamaluddin M, Wang S, Boldogh I, Tian B, Brasier AR (2007) TNF-alpha-induced NF-kappaB/RelA Ser (276) phosphorylation and enhanceosome formation is mediated by an ROS-dependent PKAc pathway. *Cell Signal* 19:1419–1433
- Jyoti A, Singh AK, Dubey M, Kumar S, Saluja R, Keshari RS, Verma A, Chandra T, Kumar A, Bajpai VK, Barthwal MK, Dikshit M (2014) Interaction of inducible nitric oxide synthase with rac2 regulates reactive oxygen and nitrogen species generation in the human neutrophil phagosomes: implication in microbial killing. *Antioxid Redox Signal* 20:417–431
- Kaur H, Halliwell B (1994) Evidence for nitric oxide-mediated oxidative damage in chronic inflammation. Nitrotyrosine in serum and synovial fluid from rheumatoid patients. *FEBS Lett* 350:9–12
- Keshari RS, Verma A, Barthwal MK, Dikshit M (2013) Reactive oxygen species-induced activation of ERK and p38 MAPK mediates PMA-induced NETs release from human neutrophils. *J Cell Biochem* 114:532–540
- Khan F, Siddiqui AA (2006) Prevalence of anti-3-nitrotyrosine antibodies in the joint synovial fluid of patients with rheumatoid arthritis, osteoarthritis and systemic lupus erythematosus. *Clin Chim Acta* 370:100–107
- Kirchner T, Moller S, Klinger M, Solbach W, Laskay T, Behnen M (2012) The impact of various reactive oxygen species on the formation of neutrophil extracellular traps. *Mediators Inflamm* 2012:849136
- Klatt P, Lamas S (2000) Regulation of protein function by S-glutathiolation in response to oxidative and nitrosative stress. *Eur J Biochem* 267:4928–4944

- Knaus UG, Heyworth PG, Evans T, Curnutte JT, Bokoch GM (1991) Regulation of phagocyte oxygen radical production by the GTP-binding protein Rac 2. *Science* 254:1512–1515
- Kobayashi K, Takahashi K, Nagasawa S (1995) The role of tyrosine phosphorylation and Ca²⁺ accumulation in Fc gamma-receptor-mediated phagocytosis of human neutrophils. *J Biochem* 117:1156–1161
- Kundu S, Ghosh P, Datta S, Ghosh A, Chattopadhyay S, Chatterjee M (2012) Oxidative stress as a potential biomarker for determining disease activity in patients with rheumatoid arthritis. *Free Radic Res* 46:1482–1489
- Lang JD, McArdle PJ, O'Reilly PJ, Matalon S (2002) Oxidant-antioxidant balance in acute lung injury. *Chest* 122:314S–320S
- Lang R, Hammer M, Mages J (2006) DUSP meet immunology: dual specificity MAPK phosphatases in control of the inflammatory response. *J Immunol* 177:7497–7504
- MacManus-Spencer LA, McNeill K (2005) Quantification of singlet oxygen production in the reaction of superoxide with hydrogen peroxide using a selective chemiluminescent probe. *J Am Chem Soc* 127:8954–8955
- Maor I, Rainis T, Lanir A, Lavy A (2008) Oxidative stress, inflammation and neutrophil superoxide release in patients with Crohn's disease: distinction between active and non-active disease. *Dig Dis Sci* 53:2208–2214
- Marcos V, Zhou Z, Yildirim AO, Bohla A, Hector A, Vitkov L, Wiedenbauer EM, Krautgartner WD, Stoiber W, Belohradsky BH, Rieber N, Kormann M, Koller B, Roscher A, Roos D, Griese M, Eickelberg O, Doring G, Mall MA, Hartl D (2010) CXCR2 mediates NADPH oxidase-independent neutrophil extracellular trap formation in cystic fibrosis airway inflammation. *Nat Med* 16:1018–1023
- Matthews JR, Kaszubska W, Turcatti G, Wells TN, Hay RT (1993) Role of cysteine62 in DNA recognition by the P50 subunit of NF-kappa B. *Nucleic Acids Res* 21:1727–1734
- Morrison D, Rahman I, Lannan S, MacNee W (1999) Epithelial permeability, inflammation, and oxidant stress in the air spaces of smokers. *Am J Respir Crit Care Med* 159:473–479
- Nakamura H, Masutani H, Yodoi J (2002) Redox imbalance and its control in HIV infection. *Antioxid Redox Signal* 4:455–464
- Niu XF, Ibbotson G, Kubes P (1996) A balance between nitric oxide and oxidants regulates mast cell-dependent neutrophil-endothelial cell interactions. *Circ Res* 79:992–999
- Nolan S, Dixon R, Norman K, Hellewell P, Ridger V (2008) Nitric oxide regulates neutrophil migration through microparticle formation. *Am J Pathol* 172:265–273
- Papayannopoulos V, Metzler KD, Hakkim A, Zychlinsky A (2010) Neutrophil elastase and myeloperoxidase regulate the formation of neutrophil extracellular traps. *J Cell Biol* 191:677–691
- Papayannopoulos V, Staab D, Zychlinsky A (2011) Neutrophil elastase enhances sputum solubilization in cystic fibrosis patients receiving DNase therapy. *PLoS One* 6:e28526
- Parker H, Winterbourn CC (2012) Reactive oxidants and myeloperoxidase and their involvement in neutrophil extracellular traps. *Front Immunol* 3:424
- Patel KD, Zimmerman GA, Prescott SM, McEver RP, McIntyre TM (1991) Oxygen radicals induce human endothelial cells to express GMP-140 and bind neutrophils. *J Cell Biol* 112:749–759
- Patel S, Kumar S, Jyoti A, Srinag BS, Keshari RS, Saluja R, Verma A, Mitra K, Barthwal MK, Krishnamurthy H, Bajpai VK, Dikshit M (2010) Nitric oxide donors release extracellular traps from human neutrophils by augmenting free radical generation. *Nitric Oxide* 22:226–234
- Patino PJ, Perez JE, Lopez JA, Condino-Neto A, Grumach AS, Botero JH, Curnutte JT, Garcia de Olate D (1999) Molecular analysis of chronic granulomatous disease caused by defects in gp91-phox. *Hum Mutat* 13:29–37
- Pignatelli B, Li CQ, Boffetta P, Chen Q, Ahrens W, Nyberg F, Mukeria A, Bruske-Hohlfeld I, Fortes C, Constantinescu V, Ischiropoulos H, Ohshima H (2001) Nitrated and oxidized plasma proteins in smokers and lung cancer patients. *Cancer Res* 61:778–784
- Pilszczek FH, Salina D, Poon KK, Fahey C, Yipp BG, Sibley CD, Robbins SM, Green FH, Surette MG, Sugai M, Bowden MG, Hussain M, Zhang K, Kubes P (2010) A novel mechanism of rapid nuclear neutrophil extracellular trap formation in response to *Staphylococcus aureus*. *J Immunol* 185:7413–7425
- Rahman I, Skwarska E, MacNee W (1997) Attenuation of oxidant/antioxidant imbalance during treatment of exacerbations of chronic obstructive pulmonary disease. *Thorax* 52:565–568
- Ramonaite R, Skieceviciene J, Kiudelis G, Jonaitis L, Tamelis A, Cizas P, Borutaite V, Kupcinskas L (2013) Influence of NADPH oxidase on inflammatory response in primary intestinal epithelial cells in patients with ulcerative colitis. *BMC Gastroenterol* 13:159
- Ramos CL, Pou S, Britigan BE, Cohen MS, Rosen GM (1992) Spin trapping evidence for myeloperoxidase-dependent hydroxyl radical formation by human neutrophils and monocytes. *J Biol Chem* 267:8307–8312
- Reynaert NL, Ckless K, Korn SH, Vos N, Guala AS, Wouters EF, van der Vliet A, Janssen-Heininger YM (2004) Nitric oxide represses inhibitory kappaB kinase through S-nitrosylation. *Proc Natl Acad Sci U S A* 101:8945–8950
- Rhee SG, Chang TS, Bae YS, Lee SR, Kang SW (2003) Cellular regulation by hydrogen peroxide. *J Am Soc Nephrol* 14:S211–S215

- Salmon JE, Millard SS, Brogle NL, Kimberly RP (1995) Fc gamma receptor IIIb enhances Fc gamma receptor IIa function in an oxidant-dependent and allele-sensitive manner. *J Clin Investig* 95:2877–2885
- Schoonbroodt S, Ferreira V, Best-Belpomme M, Boelaert JR, Legrand-Poels S, Korner M, Piette J (2000) Crucial role of the amino-terminal tyrosine residue 42 and the carboxyl-terminal PEST domain of I kappa B alpha in NF-kappa B activation by an oxidative stress. *J Immunol* 164:4292–4300
- Seril DN, Liao J, Yang GY, Yang CS (2003) Oxidative stress and ulcerative colitis-associated carcinogenesis: studies in humans and animal models. *Carcinogenesis* 24:353–362
- Simard JC, Girard D, Tessier PA (2010) Induction of neutrophil degranulation by S100A9 via a MAPK-dependent mechanism. *J Leukoc Biol* 87:905–914
- Sumii H, Inoue H, Onoue J, Mori A, Oda T, Tsubokura T (1996) Superoxide dismutase activity in arthropathy: its role and measurement in the joints. *Hiroshima J Med Sci* 45:51–55
- Summers C, Rankin SM, Condliffe AM, Singh N, Peters AM, Chilvers ER (2010) Neutrophil kinetics in health and disease. *Trends Immunol* 31:318–324
- Takada Y, Mukhopadhyay A, Kundu GC, Mahabeleshwar GH, Singh S, Aggarwal BB (2003) Hydrogen peroxide activates NF-kappa B through tyrosine phosphorylation of I kappa B alpha and serine phosphorylation of p65: evidence for the involvement of I kappa B alpha kinase and Syk protein-tyrosine kinase. *J Biol Chem* 278:24233–24241
- Tonks NK (2005) Redox redux: revisiting PTPs and the control of cell signaling. *Cell* 121:667–670
- Urban CF, Ermert D, Schmid M, Abu-Abed U, Goosmann C, Nacken W, Brinkmann V, Jungblut PR, Zychlinsky A (2009) Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against *Candida albicans*. *PLoS Pathog* 5:e1000639