# **1** Histopathological Classification

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# 1.1 Introduction

Among all benign and malignant tumors, few are the result of kidney cancer. Cancer of the kidney accounts for only about 3% of all cancers in adults, with about 36,000 cases expected in the United States in 2004 (JEMAL et al. 2004). The first description of a renal cell carcinoma, in 1883, appears to have been hypernephroma (GRAWITZ 1883). More recently, investigators have developed a classification system for renal tumors based on cytological appearance, architectural features and presumed cellular origin in the renal tubular system (COLVIN 2003; DELAHUNT and EBLE 2002; KOVACS et al. 1997). Recently, a revised pathological classification of kidney tumors has been proposed (TAKAHASHI et al. 2003; ZAMBRANO et al. 1999). Hereditary kidney tumors are associated with genetic disorders, and the same genetic abnormalities are thought to be involved in sporadic kidney cancers (NAGASHIMA et al. 2004).

Herein we describe the World Health Organization (WHO) classification of renal tumors based on pathology (EBLE et al. 2004), and discuss relevant issues in arriving at the correct differential diagnosis based on morphological findings.

### 1.2 Histopathological Classification

Kidney tumors are classified as benign or malignant depending on their histopathological features. Papillary/tubulopapillary adenoma, oncocytic adenoma, and metanephric adenoma are classified as benign. Conventional (clear cell) renal cell carcinoma, papillary renal cell carcinoma, chromophobe renal cell carcinoma, collecting duct carcinoma, cyst-associated renal cell carcinoma, sarcomatoid renal cell carcinoma, and renal medullary carcinoma are classified as malignant (Table 1.1).

Early attempts to develop a classification system of kidney cancers met with limited success (DELAHUNT and EBLE 1998). The first worldwide classification of kidney cancer, the Mainz classification, was based solely on morphological criteria (THOENES et al. 1986). The WHO has developed a new classification system for kidney cancer based on morphology and molecular findings (EBLE et al. 2004).

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Malign	ant tumors	
Conve	ntional (clear cell) RCC	
Papill	ary renal cell carcinoma	
Chron	nophobe RCC	
Collec	ting duct RCC	
Renal	cell carcinoma, unclassified	
Other	: Cyst-associated carcinoma RCC	
	Sarcomatoid RCC (Spindle RCC)	
	Granular cell carcinoma	
	Renal medullary carcinoma	
Benign	tumors	
Papill	ary/tubulopapillary adenoma	
Renal	oncocytoma	
Metan	ephric adenoma	

ors. RCC

### 1.2.1 **Conventional Clear Cell Carcinoma**

Renal cell carcinoma (RCC) accounts for 2% of all cancers. Clear cell carcinoma accounts for about 80-90% of all RCC. Generally, patients with RCC are men (by a ratio of 4:1) diagnosed in their sixth decade. Renal cell carcinoma usually presents with micro/macro-hematuria, costal vertebral pain, and abdominal mass (MOTZER et al. 1996); however, patients rarely present with this classical triad. Weight loss, anemia, thrombocytosis (INOUE et al. 2004), and abnormal hormone production (ALTAFFER and CHENAULT 1979) are common. Renal cell carcinoma may present with hypertension caused by rennin secretion (HOLLIFIELD et al. 1975), and polycythemia as the result of erythropoietin secretion (OKABE et al. 1985).

Macroscopic findings of typical clear cell RCC show a rounded, whitish-yellow, sometimes tanbrown or gray, solid, well-circumscribed tumor with focal hemorrhage and necrosis (Fig. 1.1a). A pseudocapsule is often formed by compressed normal parenchyma. Multiple nodules may be present. Cystic change can be marked, and occasionally tumors are seen as small yellow nodules on the inner surface of benign cyst. The size may range from less than 1 cm to more than 25 cm.

Microscopic findings show the usual pattern is predominantly clear cells in an alveolar architecture (Fig. 1.1b). The tumor cells are generally homogenous and characteristically possess clear cytoplasm due to abundant lipid and glycogen (Fig. 1.2). Some of the tumor cells may be eosinophilic or contain granular cytoplasm. Focal dysplastic areas can be seen in adjacent normal tubules. The tumor frequently causes hyalinization and hemorrhage. Immunohistochemically, RCC shows reactivity to keratin (CK8, 18, and 19; BANNER et al. 1990; LANGNER et al. 2004), epithelial membrane antigen (EMA), and vimentin (WALDHERR and SCHWECH-HEIMER 1985). The tumor cells are negative for col-



Fig. 1.1a,b. Conventional clear cell carcinoma in a 55-year-old man. a Macroscopic view shows a rounded, whitish-yellow, solid, well-circumscribed tumor with focal hemorrhage and necrosis. b Microscopic examination shows typical pattern is predominantly proliferation of clear cells, due to abundant lipid and glycogen in an alveolar architecture (arrows) (Hematoxylin and eosin stain; original magnification,  $\times 100$ ).



**Fig. 1.2.** Small conventional clear cell carcinoma in a 42-yearold woman. Microscopic examination shows positive stain for neutral lipid in frozen section (oil red O stain; original magnification, ×200).

loidal iron, HMB45, and lectin. The tumor tends to invade the renal vein and the inferior vena cava with metastases especially to the lung, bone, liver, and brain (Fig. 1.3; BUKOWSKI et al. 2004).

It is usually not difficult to diagnose clear cell carcinoma, but the possibility of an adrenal rest should be excluded. Distinguishing clear cell carcinoma from adrenal cortical tumor may be difficult; immunohistochemical investigation with CD10 or RCC antigen expression can help. It is important to distinguish between primary and secondary clear cell tumor with vimentin, CD10, and RCC antigen (McGREGOR et al. 2001).

Prognoses vary depending on tumor size, tumor stage, nuclear grade, DNA content, and proliferation markers. The size of the primary tumor correlates with the likelihood of distant metastases. One rare case of a very small (<3 cm) tumor that had metastasized, as confirmed at autopsy, has been reported (HELLSTEN et al. 1981). Recently, an increasing number of small renal tumors have been revealed by CT, ultrasound, and MR imag-



Fig. 1.3. Renal cell carcinoma in a 63-year-old man. Microscopic examination shows that the tumor involves the lymphatic (L) and venous (V) channels (Elastica Masson stain; original magnification, ×100).

ing. Patients diagnosed with renal tumor are classified by tumor stage, with the stage based on surgical material. We use two staging systems: the Robson system is simple and has four stages (Table 1.2; ROBSON et al. 1967). It is used mainly by urologists. The TNM staging system is more complex but provides more information (Fig. 1.4; Table 1.3; FICARRA et al. 2004; GUINAN et al. 1997). It is used by both urologists and pathologists. Tumors can also be graded according to nuclear size, shape, and nucleoli, all of which are considered prognostic factors. The most widely used nuclear grading system is that of FUHRMAN et al. (1982; Fig. 1.5; Table 1.4). In the Fuhrman system, there is a distinct prognostic difference between grade-I/II and grade-III/IV tumors. A correlation has been reported between DNA content (ploidy) and nuclear grading. Proliferation markers [Ki-67, nucleolar organizer regions (AgNORs), S-phase fraction, and proliferating cell nuclear antigen (PCNA)] have all been reported to correlate with the prognosis (SRIGLEY et al. 1997).

Table 1.2. Tumor staging system of RCC (Robson criteria)

Stage I	Stage II	Stage III	Stage IV	
Tumor confined to the kidney	Perirenal fat involvement but confined to Gerota's fascia	A: Gross renal vein or inferior vena cava involvement	A: Involvement of adjacent organs other than adrenal gland	
		<ul> <li>B: Lymphatic involvement</li> <li>C: Vascular and lymphatic involvement and metastasis to regional lymph nodes</li> </ul>	B: Distant metastases	

Table 1.3. The TNM tumor staging system of RCC. (From GUINAN et al. 1997)

#### **TNM clinical classification**

#### T: primary tumor

- TX: primary tumor cannot be assessed
- T0: no evidence of primary tumor
- T1: tumor 7.0 cm or less in greatest dimension, limited to the kidney
- T2: tumor more than 7.0 cm in greatest dimension, limited to the kidney
- T3: tumor extends into major veins or invades adrenal gland or perinephric tissues but not beyond Gerota's fascia
  - T3a: tumor invades adrenal gland or perinephric tissues but not beyond Gerota's fascia
  - T3b: tumor grossly extends into renal vein(s) or vena cava below diaphragm
  - T3c: tumor grossly extends into vena cava above diaphragm
- T4: tumor invades beyond Gerota's fascia

#### N: regional lymph nodes

- NX: regional lymph nodes cannot be assessed
- N0: no regional lymph node metastasis
- N1: metastasis in a single regional lymph node
- N2: metastasis in more than one regional lymph node

#### M: distant metastasis

- MX: distant metastasis cannot be assessed
- M0: no distant metastasis
- M1: distant metastasis

#### Other classifications

#### pTNM pathological classification:

pT, pN, and pM categories correspond to the T, N, and M categories

#### Histopathological grading:

- GX: grade of differentiation cannot be assessed
- G1: well differentiated
- G2: moderately differentiated
- G3–G4: poorly differentiated/undifferentiated

#### Stage grouping:

- Stage I: T1, N0, M0
- Stage II: T2, N0, M0
- Stage III: T1, N1, M0 T2, N1, M0 T3, N0 or N1, M0
- Stage IV: T3, N0 or N1, M0 Any T, N2, M0 Any T, any N, M1

Tal	bl	e 1.4.	Nuclear	grading	system	(Fuhrman	grade)	i
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Grade	Size (cm)	Shape of nuclei	Nucleoli
1	10	Round, uniform	Inconspicuous or absent nucleoli
2	15	Slightly irregular	Evident at high power (×400)
3	20	Obviously irregular	Prominent, large at low power (×100)
4	> 20	Bizarre, often multilobed	Heavy chromatin clump

### 1.2.2 Special Types of Renal Cell Carcinoma

### 1.2.2.1 Papillary Renal Cell Carcinoma

Papillary RCC is the second most common type of renal tubular tumor in the elderly, accounting for about 10% of all RCC. Papillary RCC is also known as "chromophil RCC" or "tubulopapillary carcinoma." THOENES et al. (1986) first and AMIN et al. (1997) more recently have developed a set of histological criteria for papillary RCC. The male-tofemale ratio is 8:1. The prognosis is better than for conventional RCC.

Macroscopic findings of papillary RCC show a rounded, heterogeneous, gray-tan, solid, well-circumscribed tumor with hemorrhage and fibrous capsule (Fig. 1.6a). The tumor size can be less than 20 cm. It is often multiple and bilateral.

Microscopic findings show that papillary RCC is characterized by papillary, tubular, and tubulopapillary growth with several cell patterns over at least 50% of its area (Fig. 1.6b). THOENES et al. (1986) further divided papillary RCC into eosinophilic, basophilic, and duophilic. Recently, DELA-HUNT and EBLE (1997) classified papillary RCC as type 1 or type 2. Type 1 has a papillary pattern of small cells with scant basophilic cytoplasm and low-grade nuclear atypia (grades 1-2). Type 2 is composed of large cells with abundant eosinophilic cytoplasm with large nuclei (grades 3-4). Foamy macrophages and neutrophils are common in the stroma adjacent the tumor. Hemosiderin, glomeruloid papillae, psammoma bodies, edema, and foamy macrophages are common in type 1, but not in type 2. Immunohistochemically, type 1 (80%) and type 2 (20%) tumors show reactivity for Kerain (CK7). Assessment of tumor growth kinet-



Fig. 1.4a,b. Renal cell carcinoma in a 60-year-old man. a Macroscopic view shows the tumor extending into the perinephric tissue (T; pT3 tumor). b Microscopic examination shows the tumor (T) involving the fatty tissue in sinus renalis (hematoxylin and eosin stain; original magnification, ×80).



**Fig. 1.5a-d.** Different grades for renal cell carcinoma by Fuhrman nuclear grading system. **a** Microscopic examination of grade 1 shows the regular, uniform nuclei comparable in size to the red blood cells (hematoxylin and eosin stain; original magnification, ×400). **b** Microscopic examination of grade 2 shows the nuclei varying in size, generally larger than in the grade-1 tumor cells. The chromatins are hyperconcentrated and the nucleoli are frequently visible (hematoxylin and eosin stain; original magnification, ×400). **c** Microscopic examination of grade 3 shows the large nuclei with hyperchromatin along with marked variability in size and shape. The nucleoli are large and conspicuous (hematoxylin and eosin stain; original magnification, ×400). **d** Microscopic examination of grade 4 shows tumor cells exhibiting large pleomorphic nuclei with chromatin clumping and conspicuous nucleoli (hematoxylin and eosin stain; original magnification, ×400).



**Fig. 1.6a,b.** Papillary renal cell carcinoma in a 67-year-old man. **a** Macroscopic view shows a rounded, heterogeneous, gray-tan, solid, well-circumscribed tumor with hemorrhage and fibrous capsule. **b** Microscopic examination shows papillary, tubular, tubulo-papillary growth (*red arrows*) with eosinophilic cells (*black arrows*) (Hematoxylin and eosin stain; original magnification,  $\times$ 200).

ics showed significant mean AgNOR scores and Ki-67 indices between the two histological types (DELAHUNT et al. 2001).

It is important to distinguish between benign and malignant renal tumors with papillary projection. Surgical pathologists use a standard protocol to make the distinction: (a) papillary, tubular, or tubulopapillary projection; (b) less than 5 mm in diameter; (c) does not histologically resemble clear cell, chromophobe, or collecting duct RCC (DELAHUNT and EBLE 1997). Collecting duct RCC is stained with lectin-binding protein [*Ulex europaeus* agglutinin-I (UEA-I), anti-keratin MA903] and expresses high molecular weight keratins, but papillary RCC is negative for these markers.

### 1.2.2.2 Chromophobe Renal Cell Carcinoma

THOENES et al. first described chromophobe RCC in 12 patients in 1985. It was first recognized in a non-human experimental tumor in 1974 (BANNASCH et al. 1974), but it was not established as a variant of human RCC until 2004 (PEYROMAURE et al. 2004). Chromophobe RCC accounts for less than 6% of renal carcinomas and has a favorable prognosis (CHEVILLE et al. 2003). It affects men and women equally. Chromophobe RCC usually presents symptoms (e.g., abdominal mass) similar to conventional RCC. Immunohistochemical, enzyme histochemical, and ultrastructural findings suggest that the intercalating cells of the collecting duct are the counterpart cells of chromophobe RCC. Macroscopic findings of chromophobe RCC show a rounded, homogenous, gray-beige, solid, well-circumscribed tumor without hemorrhage or necrosis (Fig. 1.7a). Single nodules may be present. Cystic and hyalinization changes are uncommon. Tumor size may vary. Radiologically, chromophobe RCC tumors are hypovascular, with less prominent vascularity than conventional RCC.

Microscopic findings of chromophobe RCC consist of a solid or glandular growth pattern of large cells with pale/clear cytoplasm called "pale cells" and small cells with eosinophilic cytoplasm called "eosinophilic cells" (Fig. 1.7b). Mixtures of these patterns also occur. The cell membranes are prominent in the solid or glandular pattern (COCHAND-PRIOLLET et al. 1997). These cells have little lipid or glycogen compared with conventional RCC. The nuclei are irregular and small, and appear hyperchromatic with a perinuclear halo. Chromophobe RCC usually is lower nuclear grade (grades 1-2). Ultrastructurally, chromophobe RCC commonly reveals numerous micro-cystic vesicles and mitochondria (THOENES et al. 1985). Immunohistochemically and enzyme histochemically, pale cells and eosinophilic cells stain with colloidal iron (Fig. 1.7c; THOENES et al. 1985) and show reactivity to keratin (CK18 and CK9; THOENES et al. 1988), EMA, and paxillin (KURODA et al. 2001). The tumor cells are negative for vimentin, CD10, and RCC antigen (AVERY et al. 2000).

The main differential diagnosis is between oncocytoma and conventional RCC. Colloidal iron stain can distinguish chromophobe RCC from oncocytoma and conventional RCC. CD10, RCC antibody,





Fig. 1.7a-c. Chromophobe renal cell carcinoma in a 65-year-old man. a Macroscopic view shows a rounded, homogenous, graybeige, solid, well-circumscribed tumor without hemorrhage or necrosis. b Microscopic examination shows the solid and glandular growth pattern with "pale cell" and "eosinophilic cell" with perinuclear halo (hematoxylin and eosin stain; original magnification,  $\times$ 200). c Microscopic examination shows positive stain for colloidal iron in cytoplasm (colloidal iron stain; original magnification,  $\times$ 200).

and vimentin are usually negative, and cytokeratin 18 is expressed in chromophobe RCC. On the other hand, CD10 is positive for conventional RCC, RCC antibody, and negative for vimentin.

#### 1.2.2.3 Collecting Duct Carcinoma

Collecting duct carcinoma, also known as Bellini duct carcinoma, is a rare variant of RCC, representing less than 3% of all malignant renal tumors. The tumor arises from or differentiates from the epithelium of the collecting duct, which is also known as Bellini's duct. In 1976 MANCILLA-JIMENEZ et al. first reported that collecting duct carcinoma is a form of RCC that arises from the collecting ducts. Some pathologists have suggested that collecting duct carcinoma is distinct from other types of RCC.

Collecting duct carcinoma occurs twice as often in men as in women. Initial presentation consists of hematuria and pain. An abdominal mass indicates an advanced metastatic stage (SRIGLEY and EBLE 1998). Advanced collecting duct carcinoma may be manifested by fever and leukocytosis caused by tumorderived inflammatory cytokines (NAGASHIMA et al. 2004). Approximately half of all patients die within 2 years. There is no effective therapy. Macroscopic findings of collecting duct carcinoma show a solid, white-gray tumor with invasive growth to the renal capsule, spreading to the perirenal fat (Fig. 1.8a). The tumor is usually located in the medulla or central portion of the kidney, and ranges in size from 2.5 to 12 cm. It is often necrotic and shows micro-cystic changes.

Microscopic findings show that collecting duct carcinoma is characterized by papillary, tubulopapillary, cystic, glandular, and solid patterns of tubules with desmoplastic reaction (Fig. 1.8b). Sarcomatoid/ spindle cell changes may be present (KENNEDY et al. 1990). The tumor cells have a marked nuclear atypia with eosinophilic cytoplasm and a hobnail appearance. Tumor-producing mucin may be identified. A dysplastic epithelium adjacent to the tumor also secretes mucin. These tumor cells are surrounded by desmoplastic and inflammatory stroma with many neutrophils. Immunohistochemically, collecting duct carcinoma shows reactivity to EMA and high molecular weight keratins (CK19, 34betaE12) and stains with lectin [Arachis hypogaea (PNA) and soybean agglutinin (SBA)], UEA-I (DIMOPOULOS et al. 1993).

Differential diagnosis is sometimes difficult, as macroscopically collecting duct carcinoma resembles papillary RCC. Papillary RCC is positive for lectin-binding proteins from the proximal or distal



**Fig. 1.8a,b.** Collecting duct renal cell carcinoma in a 33-year-old woman. **a** Macroscopic view shows a solid tumor with invasive growth to the renal capsule and spread to perirenal tissue. **b** Microscopic examination shows papillary, tubulopapillary, cystic patterns of tubules (hematoxylin and eosin stain; original magnification, ×200).

nephron, but collecting duct carcinoma is positive only for proteins from the distal nephron.

### 1.2.3 Other Renal Cancers

### 1.2.3.1 Cyst-Associated Renal Cell Carcinoma

Cyst-associated renal cell carcinoma (also known as multilocular cystic RCC) is an uncommon neoplasm, accounting for only a few rare cases out of all renal tumors (Fig. 1.9a). PERLMANN et al. (1928) first described cyst-associated RCC with a lymphangioma containing a small population of clear cells. The tumor appeared to be a distinct subtype of RCC with characteristic gross and microscopic features. Since then, HARTMAN et al. (1986) have suggested mechanisms by which renal cells may present carcinoma as a cystic lesion. The classification of RCC in Japan lists cyst-associated RCC rather than multilocular cystic RCC. Its etiology and behavior is uncertain, and it may be difficult to distinguish from predominantly cystic neoplasms of the kidney. Histologically, cyst-associated RCC are well-demarcated multicystic lesions containing variably sized aggregates of neoplastic clear cells with low-grade atypia (grade 1). The cyst walls are densely fibrotic and the lining is often devoid of epithelium (Fig. 1.9b). Recently, using immunohistochemistry and lectin histochemistry, it has been demonstrated that cyst-associated RCC originates from the epithelium between the distal convoluted

tubule and the collecting duct (IMURA et al. 2004). With surgery the prognosis is excellent.

However, there are many patients with long-term hemodialysis in Japan. Long-term hemodialysis frequently causes the renal tubules to dilate, and leads to malignant tumors including RCC associated with acquired cystic kidney disease (ISHIKAWA 1991). It is also known that RCC occurs as atypical cysts in patients with autosomal-dominant polycystic kidney disease (ADPKD; Fig. 1.10; KEITH et al. 1994). It is difficult to distinguish between ADPKDrelated RCC and non-related RCC, and diagnosis of ADPKD may require investigation of the patient's family medical history. It has been suggested that an RCC in ADPKD is sarcomatoid change more often than not (ISHIKAWA and KOVACS 1993).

#### 1.2.3.2

#### Sarcomatoid Renal Cell Carcinoma

Sarcomatoid RCC was first described by FARROW et al. in 1968 (Fig. 1.11a). These tumors make up less than 5% of all RCC. They have also been known as spindle cell carcinoma, anaplastic carcinoma, or carcinosarcoma. Sarcomatoid RCC is characterized by marked cytological atypia (nuclear grade 4; Fig. 1.11b) and contains enlarged pleomorphic or malignant spindle cells resembling those in sarcoma (Fig. 1.11c-e; CHEVILLE et al. 2004; PERALTA-VENTURINA et al. 2001). The aggressive behavior of sarcomatoid RCC generally leads to a poor prognosis; however, sarcomatoid RCC was not included in a recent classification scheme, because a sarcomatoid component can



**Fig. 1.9a,b.** Cyst-associated renal cell carcinoma in a 65-year-old man. **a** Macroscopic view shows a multiloculated tumor with subdividing thin septa into various-sized loculi. **b** Microscopic examination shows multicystic lesions (*C*) containing variably sized aggregates of neoplastic clear cells (*red arrow*) with fibrotic stroma (*black arrow*) (Hematoxylin and eosin stain; original magnification,  $\times$ 80).





**Fig. 1.10a,b.** Autosomal-dominant polycystic kidney disease in a 54-year-old man with hematoma. **a** Macroscopic view shows multicystic lesions (*white arrows*) with a rounded hematoma (*red arrow*). **b** Microscopic examination shows the tumor cells of renal cell carcinoma (T) are adjacent to cystic lesion (C) with intact glomeruli (*arrows*) (Hematoxylin and eosin stain; original magnification, ×80).

occur in all histological subtypes of RCC, especially chromophobe RCC (Акнтак et al. 1997).

## 1.2.3.3 Granular Renal Cell Carcinoma

Histologically, granular RCC appears granular because of the abundant eosinophilic cytoplasm and mitochondria in its cells. The only point of distinction between granular RCC and conventional RCC is the granular appearance, and distinguishing between granular RCC with low-grade nuclear atypia and benign oncocytoma is difficult. Granular RCC was eliminated from a recently proposed classification (REUTER and PRESTI 2000), and the term is no longer used in pathological diagnosis.

### 1.2.3.4 Renal Medullary Carcinoma

In 1995 DAVIS et al. first described renal medullary carcinoma in conjunction with the sickle cell trait and sickle cell hemoglobin disease. Grossly, tumors range from 4 to 12 cm in diameter and are located in the medulla. Histologically, the tumor resembles a reticular, yolk-sac tumor or an adenoid cystic carcinoma.



Peripheral satellites in the cortex and perirenal tissue with venous and lymphatic invasion are common. The tumor has a desmoplastic stroma with inflammatory cells and lymphoid follicles. Its cells react to the same stains as collecting duct carcinoma. The tumor is aggressive (SWARTZ et al. 2002).

# 1.3 Cytology

Renal cell carcinoma can be diagnosed with urine cytology. The RCC tumor cells may contain neutral lipids that stain with oil red O or Sudan IV (Fig. 1.12a). The specimen should consist of fresh, unfixed material since lipids dissolve in alcohol. Cytological examination of urine is simple and safe. Urine cytology is primarily for the diagnosis of symptomatic patients. The most common symptom of RCC is gross or micro-hematuria, but voided urine rarely yields adequate material for cytological examination, as tumor cells are not present until an advanced stage (Fig. 1.12b-d). Several investigators have suggested that retrograde catheterization with brush/lavage and fine-needle aspiration biopsy gives better results (BIBBO et al. 1974; TRUONG et al. 1999).

### 1.4 Genetic and Molecular Events in Renal Cell Carcinoma

Recent advances in genetic and molecular research have associated some types of RCC with distinct genetic and molecular abnormalities.

The von Hippel-Lindau (VHL) disease is an autosomal-dominant hereditary syndrome. Patients with VHL disease frequently develop conventional RCC (GNARRA et al. 1996). Conventional RCC has a loss of chromosome 3 and loss of function of *VHL* gene mutation (GNARRA et al. 1994). Hypermethylation of the *VHL* gene has been found in conventional RCC (HERMAN et al. 1994). Abnormalities of the *VHL* gene lead to dysfunction of the hypoxia-inducible factors–VHL protein-elongin B-C (HIF–VBC) complex pathway (MORRIS et al. 2004). Duplication and trisomy of chromosome 5q (KovAcs 1993) and



**Fig. 1.12a-d.** Metastatic renal cell carcinoma in a 63-year-old man. **a** Cytological study shows positive stain (*arrows*) for neutral lipid obtained from pleural effusion (Oil red O stain; original magnification, ×400). **b** Axial contrast-enhanced CT scan shows a large metastasis (*white arrow*) in the right lung associated with a large subcarinal lymph node (*black arrows*) and bilateral pleural effusion (*E*). **c** Macroscopic view of the resected renal neoplasm shows a large, solid tumor with invasive growth to the renal capsule. **d** Microscopic examination shows typical features of renal cell carcinoma consistent with conventional clear cell carcinoma (hematoxylin and eosin stain; original magnification, ×400).

allelic loss of chromosomes 8p, 9p, 11p, 13q, 14q, and 17p (ANGLARD et al. 1991) have been described in conventional RCC.

On the other hand, papillary RCC has trisomy and tetrasomy of chromosomes 7 and 17 (BENTZ et al. 1996). There are gains on chromosome 20 and losses of chromosome 17p and Y chromosome (DIJKHUIZEN et al. 1996). Translocations between chromosome X and 1 or 17 have been observed in 60–70% of papillary RCC (HEIMANN et al. 2001; TONK et al. 1995). Loss of heterozygosity on chromosomes 6p, 9p, 11q, 14q, and 21q (THRASH-BINGHAM et al. 1995), and the *MET* gene mutations (SCHMIDT et al. 1997), have been identified in papillary RCC.

Chromophobe RCC carcinoma has a loss of multiple chromosomes and monosomes of chromosomes 1, 2, 6, 10, 13, 17, and 20 (NAGY et al. 2004). Birt-Hogg-Dubé (BHD) syndrome has been characterized by skin tumor, pneumothorax, and renal tumor (NICKERSON et al. 2002). This renal tumor was recognized as chromophobe RCC. The *BHD* gene, located in chromosome 1p42, is reported to be a tumor suppressor gene for chromophobe RCC (OKIMOTO et al. 2004).

Cytogenetically, collecting duct RCC shows a homogeneous chromosome alteration pattern with multiple numerical and structural aberrations (mean 11.1, range 7–15) and continuous involvement of chromosomes 1 and X or Y, both as translocations and deletion/monosomy (ANTONELLI et al. 2003). Loss of heterozygosity of 1q, 2q, 6p, 8p, 13q, and 21q has been described in this RCC (FogT et al. 1998).

To our knowledge, there is little genetic and molecular information on renal medullary carcinoma. Recently, YANG et al. (2004) reported the gene expression profiles of two renal medullary carcinomas using micro-arrays containing a 21,632 cyclic DNA clone.

A good genetic and molecular marker of RCC is needed, and several investigators are working to identify markers that can differentiate the subtypes (BOER et al. 2001; BRUDER et al. 2004; LAM et al. 2004).

# 1.5 Conclusion

Management strategies for patients with kidney cancer are progressing significantly, based on advances in basic research and the growing number of clinical trials. Numerous classifications have been proposed, with varying degrees of acceptance, based on morphological and molecular futures; therefore, to arrive at the correct differential diagnosis, it is essential for the pathologist and radiologist to provide effective histological and imaging information to the urologist.

### References

- Akhtar M, Tulbah A, Kardar AH, Ali MA (1997) Sarcomatoid renal cell carcinoma: the chromophobe connection. Am J Surg Pathol 21:1188–1195
- Altaffer LF 3rd, Chenault OW Jr (1979) Paraneoplastic endocrinopathies associated with renal tumors. J Urol 122:573– 577
- Amin MB, Corless CL, Renshaw AA, Tickoo SK, Kubus J, Schultz DS (1997) Papillary (chromophil) renal cell carcinoma: histomorphologic characteristics and evaluation of conventional pathologic prognostic parameters in 62 cases. Am J Surg Pathol 21:621–635
- Anglard P, Tory K, Brauch H et al. (1991) Molecular analysis of genetic changes in the origin and development of renal cell carcinoma. Cancer Res 51:1071–1077
- Antonelli A, Portesi E, Cozzoli A et al. (2003) The collecting duct carcinoma of the kidney: a cytogenetical study. Eur Urol 43:680–685
- Avery AK, Beckstead J, Renshaw AA et al. (2000) Use of antibodies to RCC and CD10 in the differential diagnosis of renal neoplasms. Am J Surg Pathol 24:203–210
- Bannasch P, Schacht U, Storch E (1974) Morphogenesis and micromorphology of epithelial tumors of the kidney of nitrosomorpholine intoxicated rats. I. Induction and histology. Z Krebsforsch Klin Onkol Cancer Res Clin Oncol 81:311–331
- Banner BF, Burnham JA, Bahnson RR et al. (1990) Immunophenotypic markers in renal cell carcinoma. Mod Pathol 3:129–134
- Bentz M, Bergerheim US, Li C et al. (1996) Chromosome imbalances in papillary renal cell carcinoma and first cytogenetic data of familial cases analyzed by comparative genomic hybridization. Cytogenet Cell Genet 75:17–21
- Bibbo M, Gill WB, Harris MJ et al. (1974) Retrograde brushing as a diagnostic procedure of ureteral, renal pelvic and renal calyceal lesions. A preliminary report. Acta Cytol 18:137–141
- Boer JM, Huber WK, Sultmann H et al. (2001) Identification and classification of differentially expressed genes in renal cell carcinoma by expression profiling on a global human 31,500-element cDNA array. Genome Res 11:1861–1870
- Bruder E, Passera O, Harms D et al. (2004) Morphologic and molecular characterization of renal cell carcinoma in children and young adults. Am J Surg Pathol 28:1117–1132
- Bukowski RM, Negrier S, Elson P (2004) Prognostic factors in patients with advanced renal cell carcinoma: development of an international kidney cancer working group. Clin Cancer Res 10:6310S-6314S
- Cheville JC, Lohse CM, Zincke H et al. (2003) Comparisons of outcome and prognostic features among histologic subtypes of renal cell carcinoma. Am J Surg Pathol 27:612– 624
- Cheville JC, Lohse CM, Zincke H et al. (2004) Sarcomatoid

renal cell carcinoma: an examination of underlying histologic subtype and an analysis of associations with patient outcome. Am J Surg Pathol 28:435-441

- Cochand-Priollet B, Molinie V, Bougaran J et al. (1997) Renal chromophobe cell carcinoma and oncocytoma. A comparative morphologic, histochemical, and immunohistochemical study of 124 cases. Arch Pathol Lab Med 121:1081– 1086
- Colvin RB (2003) Renal parenchymal tumors: classification and new entities. Mark EJ, Young RH, Harris NL (eds) Current concepts in surgical pathology. Harvard Medical School, Boston, pp 59–80
- Davis CJ Jr, Mostofi FK, Sesterhenn IA (1995) Renal medullary carcinoma. The seventh sickle cell nephropathy. Am J Surg Pathol 19:1–11
- Delahunt B, Eble JN (1997) Papillary renal cell carcinoma: a clinicopathologic and immunohistochemical study of 105 tumors. Mod Pathol 10:537–544
- Delahunt B, Eble JN (1998) Renal tumours: the new order. N Z Med J 111:307-309
- Delahunt B, Eble JN (2002) Renal cell neoplasia. Pathology 34:13-20
- Delahunt B, Eble JN, McCredie MR et al. (2001) Morphologic typing of papillary renal cell carcinoma: comparison of growth kinetics and patient survival in 66 cases. Hum Pathol 32:590–595
- Dijkhuizen T, Van den Berg E, Van den Berg A et al. (1996) Chromosomal findings and p53-mutation analysis in chromophilic renal-cell carcinomas. Int J Cancer 68:47–50
- Dimopoulos MA, Logothetis CJ, Markowitz A et al. (1993) Collecting duct carcinoma of the kidney. Br J Urol 71:388– 391
- Eble JN, Sauter G, Epstein JI et al. (2004) WHO classification of tumours. In: Eble JN, Sauter G, Epstein JI (eds) Tumours of the urinary system and male genital organs. IARC Press, Lyon, France
- Farrow GM, Harrison EG Jr, Utz DC (1968) Sarcomas and sarcomatoid and mixed malignant tumors of the kidney in adults. Cancer 22:556–563
- Ficarra V, Novara G, Galfano A et al. (2004) Neoplasm staging and organ-confined renal cell carcinoma: a systematic review. Eur Urol 46:559–564
- Fogt F, Zhuang Z, Linehan WM et al. (1998) Collecting duct carcinomas of the kidney: a comparative loss of heterozygosity study with clear cell renal cell carcinoma. Oncol Rep 5:923–926
- Fuhrman SA, Lasky LC, Limas C (1982) Prognostic significance of morphologic parameters in renal cell carcinoma. Am J Surg Pathol 6:655–663
- Gnarra JR, Tory K, Weng Y et al. (1994) Mutations of the VHL tumour suppressor gene in renal carcinoma. Nat Genet 7:85–90
- Gnarra JR, Duan DR, Weng Y et al. (1996) Molecular cloning of the von Hippel-Lindau tumor suppressor gene and its role in renal carcinoma. Biochim Biophys Acta 1242:201–210
- Grawitz PA (1883) Die Enstehung von Nierentumoren aus Nebennierengewebe. Arch Klin Chir 30:824–830
- Guinan P, Sobin LH, Algaba F et al. (1997) TNM staging of renal cell carcinoma: Workgroup No.3. Union International Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer 80:992–993
- Hartman DS, Davis CJ Jr, Johns T et al. (1986) Cystic renal cell carcinoma. Urology 28:145–153

- Heimann P, El Housni H, Ogur G et al. (2001) Fusion of a novel gene, RCC17, to the TFE3 gene in t(X;17)(p11.2;q25.3)-bearing papillary renal cell carcinomas. Cancer Res 61:4130-4135
- Hellsten S, Berge T, Wehlin L (1981) Unrecognized renal cell carcinoma. Clinical and pathological aspects. Scand J Urol Nephrol 15:273–278
- Herman JG, Latif F, Weng Y et al. (1994) Silencing of the VHL tumor-suppressor gene by DNA methylation in renal carcinoma. Proc Natl Acad Sci USA 91:9700–9704
- Hollifield JW, Page DL, Smith C et al. (1975) Renin-secreting clear cell carcinoma of the kidney. Arch Intern Med 135:859–864
- Imura J, Ichikawa K, Takeda J et al. (2004) Multilocular cystic renal cell carcinoma: a clinicopathological, immuno- and lectin histochemical study of nine cases. APMIS 112:183– 191
- Inoue K, Kohashikawa K, Suzuki S et al. (2004) Prognostic significance of thrombocytosis in renal cell carcinoma patients. Int J Urol 11:364-367
- Ishikawa I (1991) Uremic acquired renal cystic disease. Natural history and complications. Nephron 58:257–267
- Ishikawa I, Kovacs G (1993) High incidence of papillary renal cell tumours in patients on chronic haemodialysis. Histopathology 22:135–139
- Jemal A, Tiwari RC, Murray T et al. (2004) Cancer statistics, 2004. CA Cancer J Clin 54:8–29
- Keith DS, Torres VE, King BF et al. (1994) Renal cell carcinoma in autosomal dominant polycystic kidney disease. J Am Soc Nephrol 4:1661–1669
- Kennedy SM, Merino MJ, Linehan WM et al. (1990) Collecting duct carcinoma of the kidney. Hum Pathol 21:449– 456
- Kovacs G (1993) Molecular cytogenetics of renal cell tumors. Adv Cancer Res 62:89–124
- Kovacs G, Akhtar M, Beckwith BJ et al. (1997) The Heidelberg classification of renal cell tumours. J Pathol 183:131–133
- Kuroda N, Guo L, Toi M et al. (2001) Paxillin: application of immunohistochemistry to the diagnosis of chromophobe renal cell carcinoma and oncocytoma. Appl Immunohistochem Mol Morphol 9:315–318
- Lam JS, Belldegrun AS, Figlin RA (2004) Tissue array-based predictions of pathobiology, prognosis, and response to treatment for renal cell carcinoma therapy. Clin Cancer Res 10:6304S-6309S
- Langner C, Wegscheider BJ, Ratschek M et al. (2004) Keratin immunohistochemistry in renal cell carcinoma subtypes and renal oncocytomas: a systematic analysis of 233 tumors. Virchows Arch 444:127–134
- Mancilla-Jimenez R, Stanley RJ, Blath RA et al. (1976) Papillary renal cell carcinoma: a clinical, radiologic, and pathologic study of 34 cases. Cancer 38:2469–2480
- McGregor DK, Khurana KK, Cao C et al. (2001) Diagnosing primary and metastatic renal cell carcinoma: the use of the monoclonal antibody "renal cell carcinoma marker". Am J Surg Pathol 25:1485–1492
- Morris MR, Maina E, Morgan NV et al. (2004) Molecular genetic analysis of FIH-1, FH, and SDHB candidate tumour suppressor genes in renal cell carcinoma. J Clin Pathol 57:706–711
- Motzer RJ, Bander NH, Nanus DM (1996) Renal-cell carcinoma. N Engl J Med 335:865-875
- Nagashima Y, Inayama Y, Kato Y et al. (2004) Pathological and

molecular biological aspects of the renal epithelial neoplasms, up-to-date. Pathol Int 54:377-386

- Nagy A, Buzogany I, Kovacs G (2004) Microsatellite allelotyping differentiates chromophobe renal cell carcinomas from renal oncocytomas and identifies new genetic changes. Histopathology 44:542–546
- Nickerson ML, Warren MB, Toro JR et al. (2002) Mutations in a novel gene lead to kidney tumors, lung wall defects, and benign tumors of the hair follicle in patients with the Birt-Hogg-Dube syndrome. Cancer Cell 2:157–164
- Okabe T, Urabe A, Kato T et al. (1985) Production of erythropoietin-like activity by human renal and hepatic carcinomas in cell culture. Cancer 55:1918–1923
- Okimoto K, Sakurai J, Kobayashi T et al. (2004) A germ-line insertion in the Birt-Hogg-Dube (BHD) gene gives rise to the Nihon rat model of inherited renal cancer. Proc Natl Acad Sci USA 101:2023–2027
- Peralta-Venturina M de, Moch H et al. (2001) Sarcomatoid differentiation in renal cell carcinoma: a study of 101 cases. Am J Surg Pathol 25:275–284
- Perlmann S (1928) Ubereinen Fall von Lymphangioma cysticum der Niere. Virchows Arch Pathol Anat Physiol Klin Med 268:524–535
- Peyromaure M, Misrai V, Thiounn N et al. (2004) Chromophobe renal cell carcinoma: analysis of 61 cases. Cancer 100:1406-1410
- Reuter VE, Presti JC Jr (2000) Contemporary approach to the classification of renal epithelial tumors. Semin Oncol 27:124-137
- Robson CJ, Churchill BM, Anderson W (1967) The results of radical nephrectomy for renal cell carcinoma. J Urol 101:297-301
- Schmidt L, Duh FM, Chen F et al. (1997) Germline and somatic mutations in the tyrosine kinase domain of the MET proto-oncogene in papillary renal carcinomas. Nat Genet 16:68–73
- Srigley JR, Eble JN (1998) Collecting duct carcinoma of kidney. Semin Diagn Pathol 15:54–67
- Srigley JR, Hutter RV, Gelb AB et al. (1997) Current prognostic factors-renal cell carcinoma: Workgroup No. 4. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). Cancer 80:994–996

- Swartz MA, Karth J, Schneider DT et al. (2002) Renal medullary carcinoma: clinical, pathologic, immunohistochemical, and genetic analysis with pathogenetic implications. Urology 60:1083–1089
- Takahashi M, Yang XJ, Sugimura J et al. (2003) Molecular subclassification of kidney tumors and the discovery of new diagnostic markers. Oncogene 22:6810–6818
- Thoenes W, Storkel S, Rumpelt HJ (1985) Human chromophobe cell renal carcinoma. Virchows Arch B Cell Pathol Incl Mol Pathol 48:207–217
- Thoenes W, Storkel S, Rumpelt HJ (1986) Histopathology and classification of renal cell tumors (adenomas, oncocytomas and carcinomas). The basic cytological and histopathological elements and their use for diagnostics. Pathol Res Pract 181:125–143
- Thoenes W, Storkel S, Rumpelt HJ et al. (1988) Chromophobe cell renal carcinoma and its variants: a report on 32 cases. J Pathol 155:277–287
- Thrash-Bingham CA, Salazar H, Freed JJ et al. (1995) Genomic alterations and instabilities in renal cell carcinomas and their relationship to tumor pathology. Cancer Res 55:6189– 6195
- Tonk V, Wilson KS, Timmons CF et al. (1995) Renal cell carcinoma with translocation (X;1). Further evidence for a cytogenetically defined subtype. Cancer Genet Cytogenet 81:72–75
- Truong LD, Todd TD, Dhurandhar B et al. (1999) Fine-needle aspiration of renal masses in adults: analysis of results and diagnostic problems in 108 cases. Diagn Cytopathol 20:339–349
- Waldherr R, Schwechheimer K (1985) Co-expression of cytokeratin and vimentin intermediate-sized filaments in renal cell carcinomas. Comparative study of the intermediatesized filament distribution in renal cell carcinomas and normal human kidney. Virchows Arch A Pathol Anat Histopathol 408:15–27
- Yang XJ, Sugimura J, Tretiakova MS et al. (2004) Gene expression profiling of renal medullary carcinoma: potential clinical relevance. Cancer 100:976–985
- Zambrano NR, Lubensky IA, Merino MJ et al. (1999) Histopathology and molecular genetics of renal tumors toward unification of a classification system. J Urol 162:1246–1258