

17 Allergic Contact Dermatitis and Atopic Eczema

A. Schnuch, W. Uter, K. Reich

Patients with atopic eczema (AE) share an increased susceptibility to widespread or severe cutaneous infections. This phenomenon is considered to be due to impaired cellular immunity [1, 2]. Hence, a decreased baseline risk to become sensitized to delayed-type contact allergens (haptens) could be expected and has, indeed, been postulated by many [3–7]. Conversely, an increased risk to acquire contact allergy (CA) has been claimed by some researchers [8–11]. This controversial issue has been reviewed several times (e.g., [12–18]). On the other hand, atopic eczema modulates exposure to these very allergens, most obviously in the case of emollients and topical therapeutics used for eczema therapy. In the following chapter, we will outline and discuss clinical findings reported to date and review basic mechanism of allergic contact dermatitis (ACD) in relation to relevant pathogenetic characteristics of AE, possibly interfering with the pathogenesis of ACD.

List of Definitions

- ACD** Allergic contact dermatitis. A diagnosis based on history (allergen exposure correlating with course of ACD), clinical picture (exposed sites affected), and a relevant contact allergy.
- AE** Atopic eczema. Usually diagnosed on clinical grounds, with some variation of usage to be anticipated, notwithstanding current efforts for increased standardization [221, 222].
- CA** Contact allergy. Diagnosed by patch testing; clinical relevance not considered, if not stated otherwise.
- PT** Patch test. Occlusive application of contact allergens for 24–48 h, with test readings at least until 72 h after start of exposure. PT is still the gold standard tool to diagnose contact allergy.

17.1 Clinical Findings

17.1.1 Experimental Sensitization

The notion of decreased susceptibility to ACD in AE patients is based on several experimental studies, dating back some decades (review [14]). First sensitization experiments were done with 3-pentadecylcatechol (PDC), one of the allergenic molecules of *Rhus toxicodendron* [19]. Patients with AE, regardless of current severity, were not sensitized more often than healthy controls. However, in patients tested for a second time with the *Rhus* allergen, the frequency of active sensitization (i.e., negative in the first patch test but positive in the second) was markedly reduced (6% vs 31%) [20]. Several studies used dinitrochlorobenzene (DNCB) as experimental contact allergen in patients with AE vs controls [21–23]. In these studies with DNCB and also with PDC, a lower sensitization rate was primarily observed in patients with severe AE, defined by high total serum IgE levels: only 18% of patients with a serum IgE over 1,000 kU/l became sensitized compared with 42% with a serum IgE under 1,000 kU/l [21]. Subsequent studies using different DNCB concentrations for elicitation and including AE patients with milder symptoms, however, claimed to have found “unequivocally reduced” reactivity in these patients, too [22]. In conclusion, experimental sensitization studies in humans do not provide a conclusive answer to the question as to whether AE patients are generally less prone to developing CA.

17.1.2

Population-Based Studies

While there are numerous epidemiological studies on AE morbidity and risk factors for AE, mostly in children and adolescents, only very few of these address the association between AE and ACD or CA:

- The Glostrup Allergy Study is possibly the only population-based study addressing the incidence of CA. None of the following were risk factors for acquisition of CA during the observation period: the “history of flexural eczema” (OR 1.06), a commonly used marker of (previous) AE, an elevated IgE level (OR 1.0), or the “at least one out of 10 prick tests positive” (OR 1.0), the latter factor being weakly related with AE [24, 25].
- In the Odense study with 12- to 16-year-old adolescents, Mortz et al. reported that “of those with ACD, 37% had a history of AE,” while in the whole study sample ($n = 1,340$, 1,146 of these patch tested) this proportion was 21.3% [26].
- The KORA Allergy Study found 28% of the general adult population (25–75 years) to be sensitized to at least one standard series allergen [27]; AE was not a risk factor in this older sample.

In conclusion, there is no convincing evidence of a significant association between AE and CA from population-based epidemiological studies.

17.1.3

Clinical Epidemiology

Population-based studies are often preferable over patient-based studies by virtue of unbiased estimation of morbidity and risk factor impact. However, in the field of CA they have drawbacks that impair their usefulness: (a) the positive predictive value of PT results is low due to a low prevalence of CA, i.e., a large proportion of false-positives will result; (b) for the sake of feasibility, sample size and thus power are usually limited; (c) participation rates are not in an order of magnitude that could rule out selection bias. Hence, patient-based analyses necessarily provide the bulk of evidence in the field of CA (and its association with AE).

However, due to between-center differences in the indication for patch testing AE patients, reflected by varying proportions of AE patients among patients undergoing patch testing [28], the crude prevalences of CA found in the subgroup with AE are hard to com-

pare, both between centers and between AE and non-AE patients in one given center. In this situation, adjusted, multifactorial analyses of pooled data may offer the most valid insight into the association between AE and CA. Additionally, PT screening data obtained from whole AE populations (e.g., in an AE clinic setting) may give useful information, notwithstanding the problem of an adequate control group for comparison: a population sample is probably the best possible reference, because in patients with suspected ACD, attending a PT clinic, the prevalence of CA will be higher than normal due to this very selection [29].

17.1.4

Comparisons Between Contact Allergy Patients with or Without Atopic Eczema

A multitude of case series has been published detailing the spectrum of CA in the subset of AE patients, partly comparing CA prevalences with PT results of patients without AE [6, 7, 30, 31]. While these descriptive studies can give valuable information on the patch-reaction pattern (allergic vs doubtful reactions) [31] and the spectrum of the involved contact allergens causing CA in AE patients, their results are hardly comparable for the reasons outlined above. Furthermore, unadjusted analyses are usually heavily confounded at least by age, because (a) AE PT patients tend to be younger than patients without AE and (b) age is an important surrogate marker of a multitude of exposures to allergens, including nickel. Recently, however, an age- and sex-adjusted analysis focusing on CA to topical antibiotics and antiseptics has been published, which did not find an elevated risk of these CA in AE patients, despite presumably higher exposure [32].

Because of the presumable impact of the proportion of AE patients on the overall pattern of sensitization in a PT population – both quantitatively (increased vs decreased susceptibility to CA) and qualitatively (particular allergens in topical therapeutics) – the MOHL index [33] was extended to the MOAHL index, with “A” originally including rhinitis, asthma, or AE [34]. These indices, as well as the recent extension to the MOAHLFA index (MOAHLFA: M = Men, O = Occupational Dermatitis, A = Atopic Dermatitis, H = Hand Dermatitis, L = Leg Dermatitis, F = Face Dermatitis, A = Age > 40 years) [28], with the first “A” now denoting the proportion of patch-tested patients with previous or current AE, irrespective of mucosal symptoms, intend

to summarize important patient characteristics as background information to PT results reported.

17.1.5 Prevalence of Contact Allergy in Atopic Eczema Patients

If a group of AE patients is screened for the presence of CA, the biasing effect of selection as discussed above is not a major concern. However, selection may have a certain effect in terms of a spectrum bias, in that more severe cases of AE may be overrepresented in a clinical population of AE patients, compared with the severity spectrum on a population level. In case the common notion of “reduces susceptibility to CA” should hold true, CA prevalences should be low in such studies. However, this is not the case [35–39]. Of 73 adult patients attending a specially provided AE clinic, 42% showed one or more positive PT reactions, with a striking female preponderance [36]. Of 114 children under the age of 16 years, presenting as sequential clinic attenders with AE (42.7% mild, 47% moderate, and 10.3% severe), CA was demonstrated in 43% [38]. In this study, there was no statistically significant negative correlation between the severity of AE and CA. In a study with 251 nonselected patients with moderate or severe AE, CA was frequently found on patch testing with strong age dependency: 11% of children age 2 and below, 43% of children age 7–15, and 58% of older AE patients were diagnosed with CA [35]. The authors emphasized that the diagnosis of atopic dermatitis must not lead to focusing on IgE-dependent sensitizations without PT, because ACD may often be misdiagnosed as a flare-up of AE [13].

It was discussed that CA may be a characteristic of those AE patients who have a continuing problem with their AE [36, 40]. Therefore, the high number of patients found to be sensitized to contact allergens cannot be regarded as representative for AE patients in general, but may be a marker of a specific subgroup to be further characterized, e.g., by certain immunological features. Finally, in a prospective study in 65 patients with AE and a noneczematous control group, there was no significant difference in the occurrence of CA, except of an increased risk for sensitization to nickel [39]. The few population-based studies available found 28% of the general adult population (25–75 years) to be sensitized to at least one standard series allergen [27], or 26.4% females and 7.3% males (15- to 41-year-old Danes) [41]. Assuming that the pro-

portion of false-positive PT results was not exceedingly high in the AE study [35], the age-stratified prevalences in AE patients thus appear high, and seem to indicate even an increased risk of CA in AE patients, or at least in a certain subpopulation of AE patients [36, 35, 13].

17.1.6 Multifactorial Analyses

The first analysis of this kind, performed by the Danish Contact Dermatitis Group, considered personal atopy, i.e., included not only AE, but also rhinitis and asthma [42]. Interestingly, despite this “dilution” of the effect of AE alone, Christophersen et al. found a decreased risk for nickel allergy, which contradicted notions of “nickel CA as minor criterion for AE” [15, 43]. In a multifactorial analysis of the North American Contact Dermatitis Group (NACDG) PT data (1985–1989), Nethercott et al. assessed the association of age, sex, site of dermatitis, coexistent irritant contact dermatitis, and AE with positive test results to standard series contact allergens [44]. With regard to AE as a risk factor for CA, only an “underrepresentation of AE in patients sensitized to p-phenylenediamine” was noted ([44], p. 15). The most current analysis, based on data collected by the Information Network of Departments of Dermatology (www.ivdk.org), found no significant association between AE as a risk factor for CA to 9 of 18 selected standard series contact allergens [45]. In seven instances (methyl[chloro]isothiazolinone, formaldehyde, fragrance mix, potassium dichromate, lanolin alcohols, thiuram mix and mercaptobenzothiazole and derivatives) a slightly increased risk for AE patients was identified, which may reflect higher exposure and/or increased false-positive test reactivity. In two allergens, a significantly lower risk for CA was found in AE patients, namely epoxy resin and nickel [46], the latter finding corroborating the results of Christophersen et al. [42].

17.1.7 Allergens 17.1.7.1 Nickel

The issue of nickel CA in patients with AE has long been the focus of dermatological and immunological research [14]. As one approach, the frequency of AE in

patients who are sensitized or not sensitized to nickel has been compared. However, without adjustment for age and sex as confounding factors, this approach is invalid. A recent adjusted analysis of clinical data, also taking into account other potential confounding factors, found no evidence of past or present AE being a risk factor for nickel CA [46]. As another approach, the frequency of nickel CA was compared between persons with AE and persons without AE (healthy controls). In the Odense Adolescence Cohort Study on Atopic Diseases and Dermatitis (TOACS), no association between nickel CA and AE was found [47]. In a study with university students from Turku, 34.0% (25.4%) of persons with (without) current AE had nickel CA. However, in adjusted analyses, a nonsignificantly reduced risk of nickel CA was found for atopics in this study [48] (alas, AE was not addressed).

Recently, nickel CA in relation to AE was studied not only clinically, but also immunologically [49, 50]. In view of the suppressive role of IL-10, the observation of an unchanged intralesional expression of IL-10 mRNA in AE with nickel CA, compared to increased expression after epicutaneous nickel challenge in nonatopics with nickel CA, may warrant further study; otherwise, the cytokine pattern was largely similar [49]. Recently it was shown that the *in vitro* proliferative (DNA synthesis) and secretory (IL-2, IL-5) response was impaired in nickel-stimulated peripheral blood mononuclear cells from patients with AE who were allergic to nickel, which might be interpreted as a hampered downregulation of ACD in AE [50].

17.1.7.2

Topical Drugs, Emulsifiers

For the treatment and prophylaxis of AE, various topical preparations are used, comprising a vast number of potential contact allergens. Hence, patients with severe and long-standing AE are heavily exposed to these allergens. Despite this, CA to various agents has not been found to be overrepresented in AE patients. For instance, while stasis dermatitis has been found to be a significant risk factor for CA to the antipruritic agent polidocanol, used (in Germany) to prevent and treat subchronic eczema, AE was not a risk factor, despite presumably similar exposure in both groups of eczema patients [51]. In a similar age- and sex-adjusted analysis, in this case excluding patients with stasis dermatitis, the prevalences of CA to a whole range of topical

antibiotics and antiseptics was found to be similar in patients with or without AE. Among the allergens considered, only CA to neomycin and in particular buprenorphine was (strongly) associated with AE [32]. The strong association between AE and buprenorphine CA can easily be explained by the fact that this agent is predominantly used for the treatment of AE. In contrast, AE was found to be significantly, albeit weakly, associated with CA, e.g., to lanolin alcohols [45] and to the fragrance mix [52]. However, as these two (and other) contact allergens are marginal irritants under PT conditions, and patients with AE presumably are more prone to irritation, the higher proportion of irritant, false-positive PT reactions in AE patients may at least have contributed to this finding [31].

17.2

Preimmunologic Mechanisms in Allergic Contact Dermatitis

ACD is an inflammatory reaction of the skin to a xenobiotic (the allergen). Being an allergic reaction, the immune system of the skin is involved. Allergen penetration through the horny layer, into the viable epidermis, and final absorption by the lymphatic system are thus necessary prerequisites [53, 54]. Penetration of an allergen is mainly determined by two primarily independent, but interrelated factors: the molecule and the state of the skin barrier [55–59].

17.2.1

The Molecule

The principal factors determining the kinetics of the diffusion into the skin of a xenobiotic are the physicochemical properties of the molecule. Small, nonpolar and moderately lipophilic substances penetrate best, highly water-soluble compounds least. However, some degree of aqueous solubility is also required, since the chemical must later be diffused in the relatively aqueous viable tissue. One parameter measuring lipophilicity is the octanol/water partition coefficient (P or $\log P$). The influence of a low lipophilicity was demonstrated in an experiment with the alkylating agent streptozotocin (STZ), composed of *N*-methyl-nitrosourea (the alkylating function) and a sugar moiety reducing the $\log P$. After dermal exposure in the local lymph node assay (LLNA), STZ failed to induce a response, presum-

ably because the sugar inhibits the passage of the chemical across the stratum corneum, but after intradermal injection, STZ was able to induce proliferation of draining lymph node cells, thus confirming that the chemical is inherently allergenic, provided it can access the viable epidermis [60]. However, permeability does not necessarily correlate directly with log P because other properties such as molecular weight and the molecular size may also play a role. The threshold above which penetration should become impossible was suggested to be at 500 D [61]. Combining molecular weight and lipophilicity resulted in mathematical models to predict the penetration rate of a given molecule in terms of a quantitative structure–activity relationship [62, 63].

The “500-D rule” was recently challenged, as skin contact to proteins with weights in the range of 5,000–20,000 (in the case of heveins, for example [64]) was well known to cause immediate and late symptoms [56]. In food industry workers, occupational contact urticaria (protein contact dermatitis) due to several foods (e.g., meat, baking additives) is not uncommon [65]. In the atopy patch test, penetration of high-molecular-weight aeroallergens into the skin must have taken place to elicit the eczematous reaction [66]. However, the arguments in favor of the penetration of larger molecules put forward by Berard et al. [56] are based mainly on exceptional preconditions, namely the presence of an already damaged/compromised stratum corneum [65, 67]. Many individuals sensitized to natural latex are atopic [68, 69] or had pre-existing skin lesions or dermatitis [70]. Moreover, occlusion (by gloves) as well as protective creams may enhance permeation of xenobiotics [71, 72]. Dextrans (4–10 kD) were shown to penetrate only in conjunction with the vigorous permeation enhancer *n*-octyl- β -D-thioglucoside [67]. In conclusion, the general 500-D rule [61] still seems valid in most cases.

17.2.2

Skin Barrier Function

The anatomical correlate of the epidermal permeability barrier is the stratum corneum (SC), a heterogeneous, two compartment tissue, characterized as “bricks” (corneocytes consisting of bundled, water-insoluble proteins), embedded in a “mortar” of lipids, organized into characteristic lamellar structures [55, 56, 58, 59, 73–75]. Although the permeability of corne-

ocytes is normally low, it was shown that several compounds (e.g., water, surfactants, low-molecular-weight moisturizers) can penetrate the corneocytes and thereby alter their water-binding capacities, which are normally controlled by the “natural moisturizing factor” (amino acids, potassium lactate, and others) [76]. Elevation of the water content (hydration) of the stratum corneum, e.g., after occlusion, causes increased permeability and physical/chemical changes. Barrier function, however, is mainly mediated by the lipid-enriched matrix, organized in stacked membrane sheets, with coexisting liquid crystalline and gel phase domains, which has been described by different models (the domain mosaic model, sandwich model, or single gel phase model) [58]. These structures are particularly suited for barrier function: diffusion of lipidic substances is more than 1,000-fold less than that found in cellular membranes. However, at least as conceived in the domain-mosaic model, water transport should not be excluded entirely, due to lacunar domains embedded within the lipid bilayers. After a permeabilizing stimulus (e.g., occlusion), they are thought to expand until they interconnect, forming a continuous pore pathway (extended macrodomain mosaic) [77]. The lipids account for approximately 20% of the volume of the stratum corneum. This matrix is composed of roughly equimolar mixtures of ceramides (45%–50% by weight), cholesterol (25%), and long-chain fatty acids (10%–15%), plus less than 5% of several other lipids, the most important being cholesterol sulfate.

In addition to this transepidermal route, there may be a second route via appendages (pilosebaceous follicles and sweat glands). They are a potential site of discontinuity of skin barrier integrity, which are, compared to the stratum corneum, considered as zones of less resistance (shunts) to the penetration of larger molecules, such as possibly bulky proteins. Particularly the forehead and the lower leg can be regarded as such zones of lower resistance [78], which may partly explain the high sensitization risk in lower leg dermatitis [79].

17.2.3

Regulation of Epidermal Barrier Homeostasis

Although the stratum corneum has been generally viewed as an inert structure, modern concepts of the living SC comprise a persistent metabolic activity (e.g., proteolysis of proteins, cytokine activation, lipid for-

mation, acidification), homeostatic/ metabolic links to deeper cell layers, an external biosensor function (e.g., external humidity has an impact on proteolysis, DNA synthesis and inflammation), and pathophysiologic links to deeper skin layers (barrier abrogation initiates inflammation) [80, 81]. The concept of the stratum corneum functioning as a biosensor to internal and external stimuli implies the existence of signaling mechanisms between the stratum corneum and deeper cell layers. Several processes are stimulated by barrier abrogation (Table 17.1), most importantly (relative to ACD), the activation of cellular signaling via MAP kinases (MAPK) p44/42 MAPK, and p38 MAPK [82], and the release (from preformed pools) and synthesis of several cytokines (Table 17.1). In particular, TNF- α was shown to increase via the TNF receptor p55 and induction of sphingomyelinase activity the synthesis of ceramides [83]. The role of TNF- α as a danger signal in the pathogenesis of ACD is crucial. The inflammatory cytokines remain increased in chronic perturbation, resulting in a cytokine cascade, with downstream stimulation of chemokines, adhesion molecules, and Langerhans cells [59]. Stimulation of class-I nuclear recep-

tors with steroids (glucocorticoids, estrogens, androgens) as ligands may provoke a decline in barrier function or a delay in barrier recovery. The class-II family comprises not only receptors for ligands such as thyroid hormone, retinoic acid, vitamin D3, but also orphan receptors, including peroxisome proliferator-activated receptor-(PPAR-) α (with free fatty acids as natural ligand), PPAR- γ (eicosanoids), PPAR- δ (unknown natural ligand), and LXR- α,β (oxygenated sterols). When activated by, for example, coproducts of the increased lipid synthesis after barrier disruption, namely free fatty acids, they are involved in epidermal growth, differentiation, and barrier function. Furthermore, PPAR- α activation may be involved in an increased synthesis of ceramides and cholesterol derivatives [84], and may have anti-inflammatory effects in irritant and allergic contact dermatitis [85].

17.3 Atopic Eczema and Impairment of the Epidermal Skin Barrier

Although the existence of a defect in skin barrier function in AE is well accepted, whether this defect is innate and pre-exists or whether it is a consequence of chronic cutaneous inflammation, or both, is still being debated. To deal with this controversy, a distinction should be made between function (and the operationalized indicators) and structure and its biochemical/ultrastructural indicators, such as a decrease in total lipids, a different composition of ceramides, and a different epidermal differentiation [75, 86, 87].

The barrier function is most frequently measured as water permeability and water retention by means of the transepidermal water loss (TEWL), and it was shown to be elevated in nonlesional dry skin in AE [88, 89]. It was also shown that the elevated TEWL was confined to patients with active AE [90], correlated with the acuteness of dermatitis, and was said to be normal in completely healed (and not necessarily normal appearing) skin [90, 91, 92]. However, after epidermal insults through solvents, irritants, and surfactants, TEWL increases – less in healthy controls, slightly more in inactive AE and dry skin, and significantly more in active AE – indicating an increased susceptibility of barrier function to irritants like sodium lauryl sulfate (SLS) [89, 90]. However, AE is a less reliable marker of susceptibility than TEWL itself [93, 94], indicating only

Table 17.1. Signals in response to barrier disruption (modified after [81])

Signal	Regulated response
Ions: Ca ²⁺ , K ⁺	Activation of p44/42 and p38 MAP kinases [82] Lamellar body secretion Keratinocyte differentiation
Cytokines: TNF- α , IL-1 α , β , IL-1Ra, GM-CSF, IL-6, IL-8	DNA synthesis Lipid synthesis (IL-1 α)
Growth factors: NGF, TGF- β 1, amphiregulin	DNA synthesis
Sterol regulatory element-binding proteins	Cholesterol/fatty acid synthesis LDLr expression
Nuclear hormone receptor: Class I (steroids), class II (PPAR)	Epidermal differentiation Epidermal proliferation Lipid (ceramide and sterol) synthesis [84] Anti-inflammatory effects in irritant and allergic contact dermatitis [85].

It is hypothesized that the first event after barrier disruption is an increase in transepidermal water loss (TEWL), leading to hypertonicity of epidermal cells and subsequent change in the balance of ions [82]. A number of signals may be involved in the pathogenesis of allergic contact dermatitis as well

a moderate correlation between AE and the generally used TEWL as a measure of permeability. It would certainly be premature to generalize the findings of an impaired barrier to only one single molecule (water) to other compounds, and additional work will be needed to explore whether TEWL serves as a universal, accurate, and reproducible predictor for transdermal penetration of xenobiotics. Supporting this notion, when using caffeine and lidocaine as model permeants, the extent of changes in TEWL correlated linearly with transdermal penetration of both drugs [77]. The concept of a (more or less) general barrier impairment of atopic skin could be supported further: it was shown that the skin barrier defect in AE extends to other substances such as dimethyl sulfoxide and theophylline [95]. Finally, larger protein molecules like inhalant allergens of the atopy patch test and natural latex proteins can penetrate into the skin of AE patients [66, 96]. An important prerequisite for proteins to penetrate into the skin seems to be the enzyme activity exhibited at least by some protein allergens [56]. The allergens of house dust mites, for example, are proteolytic enzymes which are able to increase permeability. The molecular targets of the Der p are occludins, members of the claudin family (transmembrane proteins of the tight junctions) [56]. And in fact, the barrier function was seriously disturbed in atopy patch test reactions, in contrast to contact allergic patch test reactions. Altogether, these findings may be taken as hints on a more general, inherent barrier impairment of atopic skin, which in turn may be further enhanced in a vicious circle by aeroallergens allowing the penetration of further allergens [97].

Beyond a simple mechanistic view of a window more or less open, substantiated by the different morphology and biochemistry of the atopic skin, the penetration could differ in a biochemical aspect as well, depending on the biophysical properties of the allergen. Substances with, for example, a specific partition coefficient (log P) could be “attracted” by the differing lipid composition of atopic skin, and “rejected” (or less attracted) by the lipids of normal skin, and vice versa. Based on such purely theoretical assumptions, it would be difficult to establish a general rule on the penetration of xenobiotic compounds in AE. Regardless of a different or similar immunologic processing of contact allergens, the susceptibility to sensitization to specific compounds could be different in AE, due to a different, allergen-specific penetration behavior.

17.4 Immunologic Mechanisms in Allergic Contact Dermatitis

The immunology of ACD has classically been divided into sensitization and elicitation phases. The sensitization phase (also called the induction phase) refers to those events that lead up to the activation of T lymphocytes, whereas the elicitation phase is the term applied to events that occur once activated T cells are re-exposed to the same allergen.

17.4.1 Sensitization Phase

Sensitization begins with the entrance of haptens into the skin [98–100]. Those haptens participating in the induction phase conjugate to epidermal and dermal molecules, generally referred to as hapten-carrier complex. The critical binding structures have not yet been identified unequivocally. Probably depending on their chemical nature, haptens may bind directly to peptides bound on MHC molecules of antigen-presenting cells, or bind to proteins, which are processed by antigen-presenting cells, or bind directly to MHC molecules. Sensitizing organic compounds are generally electrophilic and bind covalently to nucleophilic groups, such as thiol, amino, or hydroxyl groups, whereas metal ions, e.g., nickel cations, form stable metal-protein chelate complexes [101]. However, some xenobiotics (prohaptens) only enter these first steps of sensitization after conversion to protein-reactive haptens, i.e., the original compound is a nonsensitizer. Examples are limonene and colophony. Their induction capacity relies on oxidation by air [102, 103]. In addition, xenobiotic metabolizing enzymes in the skin can convert prohaptens to electrophilic compounds [104, 105]. One example is the activation of cinnamic alcohol to the presumed allergen cinnamic aldehyde [106]. For effective sensitization, a chemical must therefore be inherently protein-reactive or must be converted in the skin to a protein-reactive metabolite. For the latter compounds, genetic differences in metabolism may play a role in the differential susceptibility of individuals to develop contact allergy [107, 108].

17.4.1.1

Activation, Maturation, and Migration of Langerhans Cells

Immature dendritic cells (DCs) bearing the antigen are first activated by antigen nonspecific stimuli, (as irritants also activate LCs). Following activation, these cells are stimulated to leave the epidermis and migrate to the local lymph node. During migration, LCs undergo functional maturation such that they lose the ability to process antigen and acquire instead the characteristics of mature antigen-presenting DCs, e.g., increased expression of MHC and of costimulatory molecules (ICAM-1, LFA-3, B7-1, and B7-2) [109].

The whole process is orchestrated by several important changes in the skin, involving cytokines and chemokines and their receptors (IL-1 β , TNF- α , IFN- γ , IP-10, MIP-2, IL-12, IL-15, IL-18), adhesion molecules (E-cadherin, ICAM-1, α 6 integrin, CD44 variants), lipid mediators (PGE₂), and matrix metalloproteinases (e.g., MMP-9) [100–112]. The first and probably most crucial step in the induction phase is the early upregulation of IL-1 β mRNA and synthesis of the IL-1 β precursor, which is cleaved by the protease IL-1 β -converting enzyme (ICE; caspase-1). Caspase-1 activation is induced either by haptens or irritants (SLS) [113]. IL-1 β was also referred to as the master cytokine, as it was able to initiate the whole cytokine profile, in particular TNF- α synthesis by adjacent keratinocytes, and second, to supply signals for the activation, maturation, and mobilization of LCs [114]. TNF- α provides LCs with the second cytokine signal necessary for successful migration. These stimuli are delivered to LCs via both types of the TNF- α receptor (p55 TNFR and p75 TNFR) [115], and the type 1, signal-transducing, receptor for IL-1, IL-1RI [116]. Furthermore, IL-1 β and TNF- α weaken and break the E-cadherin bonds that bind LCs to adjacent keratinocytes, thereby allowing LCs to move through the layers of the epidermis. To facilitate the penetration of LCs into the dermis, the production of several matrix metalloproteinases is upregulated (by TNF- α), which participate in the degradation of E-cadherin and degrade the macromolecules of the epidermal basement membrane [109]. The movement via the extracellular matrix and lymphatic endothelial cells is guided by several chemokines and their respective receptors (e.g., CCR 7) [111].

Mobilization and migration of LCs seem, however, subject to counter-regulatory influences [110]. One important candidate is IL-10, which is upregulated fol-

lowing skin sensitization. It has been suggested that in the absence of IL-10 (in IL-10 knockout mice) the pro-inflammatory cytokines IL-1 β and TNF- α are overexpressed. Another cytokine that may have the potential to regulate LC migration is TGF- β 1, which is able to inhibit the upregulation by TNF- α of CCR7 expression on DCs and to increase the expression by DCs of E-cadherin [117]. As further regulators of LC migration, lactoferrin (LF) and peroxisome proliferator-activated receptor γ (PPAR- γ) can be mentioned. LF is an iron-binding protein, which is found in exocrine secretions, known to be expressed in healthy skin. Exogenous topical (recombinant) LF was shown to be able to inhibit allergen-induced LC migration, secondary to suppression of the de novo synthesis of TNF- α , and possibly of other proinflammatory cytokines [116, 118, 119]. PPARs belong to the nuclear hormone receptor superfamily [120]. PPAR- γ is involved in macrophage maturation and modulation of immune and inflammatory reactions [120, 121]. Recently, it was shown that LCs express PPAR- γ and that activation of PPAR- γ by rosiglitazone, an antidiabetic drug acting as a synthetic ligand, specifically impairs the departure of LCs from the epidermis [122].

17.4.1.2

The Role of (Nonspecific) Inflammation

In many instances, it appears that topical administration of a contact allergen alone is sufficient to trigger the induction or upregulation of those cytokines necessary for the effective acquisition of sensitization. Under these conditions of exposure, the chemical itself causes sufficient cutaneous inflammation and irritation and hence the production of proinflammatory cytokines. However, chemicals that do not provoke proinflammatory changes may fail to induce the necessary cytokine responses. Furthermore, endosomal processing (MHC peptide ligand formation) and LC activation depend on inflammatory stimuli [123, 124]. The intimate relationship between irritation and sensitization was substantiated by, for example, studies with the contact allergen DNCB together with the irritant SLS in mice [125]. At high (irritant) doses of DNCB, SLS did not influence the levels of immune activation induced by the allergen. However, at lower (nonirritant) concentrations of DNCB, responses were augmented by SLS, which is thought to provide the necessary exogenous inflammatory stimuli. Coadministra-

tion of an irrelevant hapten reduced the doses still sufficient to elicit CA by a factor of 10^3 [126]. It was proposed that the chemical irritancy of a hapten activates the innate immune system, an activation step necessary for development of specific immunity in the skin [127] (The innate immune response is a defense mechanism through which invariant molecular patterns of infectious agents – Toll-like receptors – are recognized [128]).

The concept referring to the necessary danger signals [129] is supported by clinical observation. Patients with a lower threshold of sensitivity to SLS seem to be more susceptible to sensitization to a contact allergen (colophony) [130]. In summary, a certain level of skin irritation seems to be required, at least for weak allergens. Chemicals that fail to trigger sufficient local cytokine production may – in the absence of additional exogenous stimuli – be unable to realize their full potential as allergens.

17.4.1.3

Langerhans Cell–T Cell Interaction and the Role of T Cell Subsets

The induction of skin sensitization and the subsequent elicitation of allergic contact dermatitis depend on the development of hapten-specific T lymphocytes.

Primary hapten presentation to naive T cells together with costimulatory signals results in the generation of cutaneous hypersensitivity (CHS) effector cells. In contrast to other types of delayed-type hypersensitivity (DTH) responses, which are mediated by CD4+ cells, most haptens evoke a response consisting mainly of CD8+ effector cells. However, besides CHS effector cells, T cell populations that downmodulate CHS are also induced, namely hapten-specific suppressor cells. Reduction of the hapten dose results in gradual loss of T suppressor cell induction but retained sensitization. Further dose reduction finally results in low-zone tolerance [131, 132]. This dose-dependent activation of different T cell subsets might result from different antigen presentation. Whereas insufficient antigen-presenting cell (APC) activation or inadequate costimulation results in T cell anergy, inadequate ligation of T cell receptors may result in generation of T suppressor cells. High doses of hapten may lead to antigen presentation by LCs and also by less efficient APCs, the latter generating only inadequately primed T suppressor cells. Lower doses of hapten might result in antigen presentation

exclusively by LCs, and therefore induce CHS effector cells only. Very low doses might result in suboptimal hapten concentration, even on professional APCs (LCs) [126], or bypass the involvement of LCs [132], again generating T suppressor cells that mediate low-zone tolerance, which were characterized as CD8 helper-type 2 cells [133].

For activation and proliferation, T cell receptor triggering (signal 1) is insufficient, but hapten-presenting APCs also provide the required costimulation (signal 2), which involves, for example, IL-1 α , OX40 ligand, and cellular adhesion molecules (e.g., CD80 and CD86) [134–136]. The latter molecules bind to their counterparts on T cells, CD28, and CD152 (CTLA-4, functioning as a negative regulator [137]). These interactions promote mutual activation of both hapten-presenting APCs and hapten-reactive T cells. To promote T cell proliferation, cellular adhesion stimuli need to be complemented by several cytokines (e.g., IL-2, a highly potent T cell growth factor). Primary skin contact with most contact allergens leads to differentiation and expansion of allergen-specific effector T cells, particularly CD8-positive cells displaying the type-1 cytokine profile, whereas a subgroup of CD4-positive T cells produces IL-2, IL-4, and large amounts of IL-10, regulating the immune reaction principally mounted by CD8-positive T cells. However, prolonged allergenic contact ultimately leads to a predominance of type-2 allergen-specific T cells, which may take over the role of type-1 cells in causing contact allergic hypersensitivity. It seems likely that the expression of IL-4 (and possibly other type-2 cytokines), particularly at sites of dermal challenge, regulates what is considered to be a largely Th1- or Tc1-dependent immune response, although the factors governing whether it is upregulated or downregulated are still unclear [138–140]. Finally, on maturation T cells acquire (in an IL-12-dependent manner) molecular keys that allow extravasation, one of the important ones being CLA, the (cutaneous lymphocyte associated antigen), which is formed from the glycosylation of P-selectin glycoprotein ligand 1.

17.4.2

Elicitation Phase

The elicitation phase of ACD is triggered by re-exposure of the skin to the relevant hapten. As in the induction phase, antigen-presenting cells are required to reactivate specific T cells [126, 98, 100].

17.4.2.1

The Movement of Nonspecific and Specific T Cells to the Site of Hapten Re-exposure

The first events initiated by the hapten in the skin after contact with keratinocytes are nonspecific inflammatory reactions caused by inherent inflammatory/irritant properties of the hapten (danger signals). Inflammatory and vasoactive mediators from, for example, mast cells (C5a and serotonin), cytokines (TNF- α , IL-1 α , GM-CSF, IL-18, from keratinocytes, and later from infiltrating monocytes and DCs), and chemokines (CXCL1, MCP1 [CCL2], RANTES [CCL5], Mig [CXCL9], CTACK [CCL27], IP-10 [CXCL10], MIP3- α [CCL20]) are released, which is followed by an activation of endothelial cells and an increased expression of adhesion molecules. All these first inflammatory responses are nonspecific, due to the inherent proinflammatory properties of the hapten (or accompanying inflammatory stimuli). Nonspecific leukocyte recruitment is largely under the control of chemokines, released in a sequential and coordinated manner from resident and immigrating cells [111]. As the hapten can only be presented in the extravasal tissue, T cells have to move from circulation to the hapten-exposed regions. The process is initiated by selectins expressed on T cells (L-selectin), endothelial cells (P-selectin and E-selectin) and activated platelets (P-selectin). Selectins form bonds between endothelial surfaces and T cells, moderating the rapid motion of T cells to a slow roll ("tethering"). In a second stage of extravasation, T cells receive chemokine signals, which are required for integrin activation. Integrins bind to ICAM-1 and VCAM-1, which halts T cell motion. Now cell extravasation into the dermis and migration to the site of the hapten are possible.

17.4.2.2

Specific T Cell–Antigen-Presenting Cell Interaction and Inflammatory Response

The accumulation of mostly nonspecific and much less specific T cells on the site and their activation by APCs, macrophages probably playing a key role [141], is followed by the release of various cytokines (IFN- γ , TNF- α , TNF- β , GM-CSF, IL-1). The main effector cytokine in ACD is IFN- γ , acting in concert with TNF- β to upregulate the expression of ICAM-1 [100]. Another cytokine, IL-12, was shown to be important in the induction and the elicitation phase of ACD. The cytokines in turn stim-

ulate keratinocytes to produce IP-10, Mig, and I-TAC (CXCL11), the ligands for CXCR3. These chemokines selectively attract T lymphocytes. Keratinocytes continuously produce large amounts of CXCR3 ligands, thus contributing to further accumulation of CXCR3-bearing T cells. The result is that more than 70% of ACD-infiltrating T cells are CXCR3+. Mast cells and thrombocytes are activated and enhance the inflammatory reaction. The final steps are, therefore, as the first steps of elicitation, nonspecific inflammatory processes.

17.4.2.3

The Effector T Cells

Effector T cells may be CD4+ (Th) or CD8+ (Tc) cells. CD8+ cells producing IFN- γ , no IL-4, and no IL-10 (Tc1 cells), and activated under the influence of IL-12 [142–144], are now considered to be the main effector cells in contact allergy, together with some (IFN- γ -producing) CD4+ cells, but other CD4+ cells (producing IL-4 and IL-10 but no IFN- γ) mainly seem to have a regulatory function. It seems now established that both CD4+ (Th1) and CD8+ (Tc1) cells are necessary for the full expression of ACD [145]. Furthermore, the inflammatory reaction in CHS depends on CD8+ cytotoxic activity mediated by perforin and FasL [146] and is responsible for the lysis of keratinocytes [147–149]. Beside CD8+ cells, CD4+ cells in concert with IFN- γ may also exert cytotoxic activity [143]. It was further hypothesized that CD8+ T cells lyse CD4+ cells (bystander cytotoxicity), responsible for the predominance of CD8+ effector T cells [147]. However, an increased apoptosis of Th1 cells was observed only in atopic patients (leading to a predominance of Th2 cells), and not in ACD patients [150].

T cell subsets, whether CD4+ or CD8+, release not only type-1 cytokines (IFN- γ , IL-12), an opinion held for a long time, but also type-2 cytokines, particularly IL-4 [151, 152]. Loss of IL-4 expression in BALB/c mice was associated with impairment of the ACD reaction to DNCB. On the other hand, Ni-specific T cell clones prepared from nonallergic patients displayed low IFN- γ and a high IL-10 production, compared with T cell clones from allergic patients, again indicating a regulator role for IL-10 on an individual basis [153]. With regard to the regulation of the balance between these two responses, it is likely that the properties of the allergen play the major role in controlling the equilibrium between Th1 and Th2. The response to DNFB is

Th1-predominant, the contact allergen MCI/MI elicits, as metals, a mixed (Th1 and Th2) profile [154], whereas the fluorescein isothiocyanate response is Th2-predominant.

17.4.2.4

The Resolution of Allergic Contact Dermatitis

The elicitation phase of ACD is self-limiting. IL-4 and IL-10, secreted in the late elicitation phase by CD4+ Th2 cells, have both been implicated in its downregulation. T regulatory (Tr) lymphocytes producing predominantly IL-10 may play a central role [143]. They migrate in response to various chemokines including I-309 (CCL1), MCP-1, MIPs, and TARC. Tr cells express higher levels of CCR8 (the receptor of I-309). I-309 from keratinocytes and activated T cells with an earlier kinetics than IL-4 and IL-10 attracts Tr cells more vigorously than Th2 cells. This indicates that I-309/CCR8 may contribute relevantly to the termination of ACD through the recruitment of Tr lymphocytes [111]. IL-10 blocks DC maturation, including IL-12 release, thus impairing activation of T cells. In addition, the release of factors such as PGE2 and TGF- β , derived from activated keratinocytes and leukocytes, contributes to dampening the immune response. PGE2 inhibits the production of pro-inflammatory cytokines, probably through an enhanced production of thrombospondin1, an endogenous antiinflammatory regulator stimulated by nonspecific danger signals and released by DCs [155]. TGF- β silences activated T cells and inhibits further infiltration by downregulating the expression of adhesion molecules on endothelial and skin cells. Further suppressive effects were observed with certain neuropeptides, especially α -MSH and VIP (in contrast to other neuropeptides such as substance P and CGRP, which enhance the inflammatory response) [156, 157].

17.5

The Immunopathogenesis of Atopic Eczema – Possible Interference with Allergic Contact Dermatitis

17.5.1

Background

AE is a chronic eczematous skin disorder with a complex polygenetic background that occurs as cutaneous manifestation of the atopy syndrome. Accordingly, the

majority of patients with AE show high levels of IgE antibodies, which usually react with a limited spectrum of typical allergens, such as food components, house dust mite, or birch pollen, and many patients suffer from concomitant allergic rhinoconjunctivitis or asthma. A significant percentage of affected individuals, however, do not show IgE hyper-responsiveness. These patients, who are usually referred to as “intrinsic” or “nonallergic” AE patients, might be genetically and immunologically different from “extrinsic” AE patients [158, 159], although some principle mechanisms, such as the activation of IL-5- and IL-13-producing CD4+ and CD8+ T effector cells, appear to be comparable [160].

IgE-related inflammatory pathways were originally thought to play a critical role, especially in immediate-type hypersensitivity reactions (type I) with (contact) urticaria as a typical skin symptom. Because AE clinically and histologically corresponds to a cutaneous DTH response (type IV), and AE lesions are in fact often indistinguishable from those of ACD, the role of IgE in this type of atopic skin lesion has long been theoretical. A possible link between the increased production of IgE antibodies and the development of eczematous skin lesions has been provided by the demonstration of high levels of high-affinity IgE receptors, Fc ϵ RI, on epidermal dendritic cells of AE patients, and the finding that these molecules contribute to a preferential and highly efficient uptake of IgE-targeted allergens and subsequent activation of specific T cell responses, the latter step resembling the sensitization phase of ACD. The highest levels of Fc ϵ RI expression have been observed on a subset of epidermal DCs, the so-called inflammatory epidermal dendritic cells (IDECs), which seem to be specifically recruited into the epidermis of active AE lesions and might contribute to the increased number of epidermal DCs observed in this condition [161]. Upon activation, these cells release large amounts of pro-inflammatory cytokines, thereby possibly amplifying the inflammatory immune reaction in AE [159]. Because aggregation of Fc ϵ RI on monocytes and DCs induces NF- κ B activation [162], increased expression of this receptor may directly contribute to abnormal APC function in AE.

In both types of eczematous skin disease, AE and ACD, activated T cells have been identified as the main effector cell type. According to current pathogenetic concepts, however, a characteristic feature of AE is the preferential activation of CD4+ and CD8+ T cells that

produce Th2 cytokines, such as IL-4, IL-5 and IL-13 [163], while the majority of ACD reactions, under the influence of IL-12 derived from APCs and other cell types, involve the activation of the Th1 and Tc1 subsets with IFN- γ as the leading cytokine [164], although some ACD responses may also require the activation of Th2 cells. The bias toward activation of Th2-type pathways is regarded as a principal immune deviation in AE, which is likely to result from complex interactions between genetic and environmental factors. The preferential priming of Th2 cells in AE, which is reflected in the preponderance of (allergen-specific) Th2 cells in the peripheral blood, acute skin lesions, and the early phase of atopy patch test reactions, is likely to occur as the consequence of several abnormalities present at different cellular and molecular levels of the immunologic cascade. On the other hand, a lack of sufficient Th1-inducing stimuli during a critical learning phase of the immune system in early childhood has been postulated as an important environmental component involved in the impaired ability of patients with atopic diseases to generate allergen-specific Th1 responses. In line with this concept, changes in the infectious environment and in the pattern of microbial exposure of children associated with westernization might be a critical factor underlying the increased prevalence and severity of atopic diseases that has been observed in Western countries over the last decades (the hygiene hypothesis) [165, 166].

17.5.2

Immune Deviation in Atopic Eczema

The presently available experimental data on the possible mechanisms involved in the preferential generation and recruitment of Th2 cells and induction of skin inflammation in AE have recently been reviewed [167, 165]. They may be summarized as follows:

1. Keratinocytes in AE show different abnormalities including overexpression of thymic stromal lymphopietin (TSLP). TSLP activates DCs and induces the expression of chemokines that selectively attract CCR4-expressing Th2 cells such as TARC (CCL17) and MDC (CCL22). Moreover, TSLP-primed DCs induce IL-4, IL-5, IL-13, and TNF- α in responding T cells, but downregulate IFN- γ and IL-10 [168]. There is also evidence that, in addition to an exaggerated expression of Th2-selective chemokines [169, 170] and other chemotactic factors involved in the recruitment of DCs and T cells, such as MIP-3a (CCL20) [171, 172] and CTACK (CCL27) [173], keratinocytes in AE patients, compared with nonatopics, release higher amounts of pro-inflammatory cytokines, including IL-1 and TNF- α , either spontaneously or in response to stimuli such as IFN- γ [174, 175]. The higher baseline activation of keratinocytes may be related to the disturbed epidermal barrier function in AE. Alternatively, a state of keratinocyte preactivation could result from endogenous abnormalities regarding the inhibitory effects provided by cytokines such as TGF- β [176, 177, 178, 214], a product, for example, of T cells with suppressor functions, and other negative regulators of IFN- γ signaling, including members of the suppressor of cytokine signaling (SOCs) family [179]. Finally, increased keratinocyte apoptosis in AE may trigger the release of several factors that induce chemotactic responses in CXCR3-expressing T cells, which are highly increased in AE lesions [180].
2. At the level of APCs, including monocytes and monocyte-derived DCs, several functional alterations have been described in AE patients, including an increased immunostimulatory capacity [181] with preferential induction of Th2 cells [182]. This finding is possibly related to a reduced capacity to secrete IL-12 and increased release of IL-10, at least under certain conditions of DC activation [183]. Furthermore, monocyte-derived DCs from AE patients display an enhanced production of chemokines such as MDC (CCL-22) [184] and IL-16 [185], a cytokine involved in the selective recruitment of CD4+ T cells, DCs, and eosinophils. The results of other studies indicate a defective synthesis of the Th1-activating cytokine IL-18 in monocytes of AE patients as a possible mechanism underlying decreased IFN- γ production in response to bacterial toxins [186, 187].
3. Th1 and Th2 cells develop from the same naïve T cell under the influence of various factors, the majority of which are active during the interaction with APCs. These include ligation of the T cell receptor, binding of costimulatory molecules, and presence of regulatory cytokines in the micromilieu of the responding Th cell. The functional balance between Th1 and Th2 cells in a given immune response will also depend on the presence of regulatory T cell subsets that may specifically suppress one

or the other Th subset, either via release of mediators or the selective induction of T cell apoptosis. Notably, a preferential apoptosis of circulating Th1 memory effector cells has recently been shown to contribute to the predominance of Th2 cells in atopic diseases [150]. Signals through contact molecules, as well as through cytokine receptors, induce a complex series of secondary molecular events that ultimately lead to the binding of cell type-specific transcription factors to regulatory elements in the promoters of sets of genes implicated in the functional program of the activated T cell. These signal transduction cascades have recently been identified to play an important role in determining Th1 or Th2 differentiation because of their possible antagonism [165]. At the molecular level activation of STAT6, the proto-oncogene c-Maf and GATA-3 are associated with Th2 development. GATA-3 not only plays a role in the upregulation of the Th2 cytokines IL-4, IL-5, and IL-13, but also inhibits the production of IFN- γ , thereby preventing Th1 development [188]. While the interaction of IL-4 with its receptor on the surface of naïve T cells initiates the activation of STAT6, binding of IL-12 to the IL-12R results in the activation of STAT4. Another transcription factor involved in Th1 lineage commitment and expression of IFN- γ is the protein T-box expressed in T cells (T-bet), which simultaneously represses IL-4 and IL-5 [188]. While there is solid evidence for a Th2 polarization, the exact relation between altered signaling cascades and the immune deviation in AE is less clear. However, from animal models it can be concluded that the activity of transcription factors may play a crucial role in the manifestation of atopic diseases including AE [189; 190].

4. There is no doubt that T regulatory cells (Treg) are important elements in maintaining a physiological immune homeostasis of the skin, and a disturbance of Treg functions may contribute to abnormal immune responses underlying AE and ACD. Treg cells have been divided into natural and adaptive subpopulations [191]. The former are generated in the thymus and later migrate to peripheral tissues, where they normally function to prevent the activation of self-reactive T cells that have the potential to develop into effector cells. They are characterized by a CD4+CD25+ phenotype, expression of Foxp3 and glucocorticoid-induced TNF receptors, produce little or no cytokines, and mainly act through T cell-T

cell or T cell-APC contact-dependent mechanisms. Among CD4+ cells, Th3 cells, which mainly produce TGF- β , and Tr1 cells, which mainly produce IL-10, with or without TGF- β , are adaptive Treg cells that also originate from the thymus but differentiate further and acquire their suppressive activity in the periphery under certain conditions of antigenic stimulation [191]. Their expression of CD25 is variable, and their mechanism of suppression is mediated by inhibitory cytokines, such as IL-10 and TGF- β . Natural and adaptive Treg cells might function in different immunologic settings, depending, for example, on the context of antigen exposure and the nature of the inflammatory response. Whether Th1 and Th2 responses are equally susceptible to the suppressive activities of Treg cells, and whether they are controlled by the same or different types of Treg cells is presently not completely clear. There is evidence to suggest that CD4+CD25+ Treg cells effectively suppress Th1 responses, but have an impaired suppressive [192] or even activating effect on Th2 responses [193]. In fact, a recent study found an increased frequency of CD4+CD25+ Treg cells in the peripheral blood of AE patients compared to patients with asthma and healthy controls [194]. The authors also suggested that stimulation of CD4+CD25+ Treg cells with staphylococcal superantigens may reverse their suppressive function. Furthermore, IL-10, a central mediator of adaptive Treg cells, has been implicated in the control [195] as well as in the induction of Th2 allergic reactions [196].

5. It is noteworthy that in chronic AE lesions, the expression of IL-4 and IL-13 decreases, whereas expression of IFN- γ is upregulated. The switch toward a Th1-type response is probably mediated by an increased dermal recruitment of eosinophils, macrophages, and DCs expressing IL-12, and chemokines derived from keratinocytes in chronic AE lesions may further enhance the local accumulation of Th1 cells (reviewed in [164]). IL-4 itself and the increased colonization of AE skin by *Staphylococcus aureus* may represent important stimuli that activate DCs and macrophages to release IL-12. The microbial induction of toll-like receptors may also initiate other events in favor of a Th1-type response, including the inhibition of Th1-suppressing Treg cells and upregulation of T-bet, which are able to convert polarized Th2 cell into IFN- γ -producing

Th1 cells. Thus a variety of changes in the local microenvironment may finally lead to a cutaneous cytokine milieu reminiscent of ACD that promotes the activation of cytotoxic lymphocytes. Interestingly, in a certain analogy to ACD, a T cell-mediated apoptosis of keratinocytes has recently been proposed as a pathogenic mechanism in AE [148].

17.5.3

How Atopic Eczema Can Affect Allergic Contact Dermatitis

Because AE was regarded as a Th2-type disease, contrary to Th1-driven ACD, a lower prevalence of ACD reactions in AE patients compared to healthy controls was expected. On the other hand, the local microenvironment in AE, particularly that of chronic AE lesions, may be generally regarded as a so-called danger signal that should facilitate the development of ACD response to an absorbed allergen. However, recent epidemiologic studies suggest that ACD is about as common in AE patients as it is among nonatopic individuals. Based on the increasing knowledge of the specific immunologic abnormalities in AE, interactions with the pathogenesis of ACD may hypothetically occur at multiple levels (Table 17.2). Among others, these include:

- The increased release of chemokines and cytokines from keratinocytes
- The presence of high numbers of preactivated DCs that are able to secrete large amounts of chemotactic and pro-inflammatory mediators
- The presence of increased numbers of activated memory effector and cytotoxic T cells
- Changes in the local control of inflammatory responses by Treg cells

The specific immunologic changes associated with AE may either facilitate or hamper the development of

ACD responses, depending on the type of AE (intrinsic vs extrinsic) [197, 159], the duration of AE skin lesions (acute vs chronic), the presence of co-factors, such as epidermal barrier dysfunction (dry skin) and microbial colonization (*S. aureus*) and the type of the potential contact allergen (Th1- or Th2-type ACD). Clearly, the exact consequences of different immunologic alterations present in AE for the manifestation of ACD reactions remain to be determined. It is also possible that ACD reactions to a given allergen differ immunologically between AE patients and nonatopic individuals, as suggested by recent studies on the cutaneous response [49] and the release of cytokines from peripheral blood nuclear cells to nickel in patients with ACD to nickel with or without concomitant AE [50].

17.6

Conclusion

Susceptibility to sensitization to contact allergens may vary with the clinical severity of AE [7, 198, 199, 200], e.g., has been found remarkably low in patients with high serum IgE levels (above 1,000 kU/l [21, 35]. Furthermore, the group of patients with AE has been found to be heterogeneous concerning responses to immediate type hypersensitivity allergens (extrinsic vs intrinsic type of AE); such heterogeneity, albeit not yet clearly identifiable, may also exist with regard to contact allergens, both specifically in AE patients and generally. Possible mitigating effects of AE on the pathogenic process of ACD on the immunological level may be compensated by the established barrier dysfunction, facilitating the penetration of haptens in AE. In summary, taking recent evidence into account, it appears reasonable to assume a largely similar susceptibility to CA in persons with or without AE.

Table 17.2. Preimmunologic and immunologic factors that may interfere with allergic contact dermatitis in atopic eczema

	Atopic eczema	Possible (hypothetical) effect on ACD in AE
Preimmunologic parameters	Disturbed epidermal barrier function: TEWL increased in nonlesional dry skin [88, 89]; permeation of drugs [77, 95] and macromolecules [66, 96] (increase) Biochemical: different lipid composition [75]	Penetration of an allergen into the skin facilitated, possibly also haptens with a mol weight > 500 D Different permeation of the hapten depending on its biophysical properties (e.g., log P)
	Repair signals also acting as inflammatory signals (e.g., TNF- α) [81, 83]	Danger signals in the immunology of ACD

(Abbreviations see p. 193)

Table 17.2. (cont.)

Atopic eczema		Possible (hypothetical) effect on ACD in AE
Immunologic parameters	<p>Keratinocytes Increased MMP activity with increased serum levels of TIMP-1 that resolve during treatment [201]</p> <p>Altered caspase activation probably due to genetic variations in the caspase recruitment domain containing protein 15 (CARD15) [202]</p> <p>Animal model: evidence for a crucial role of increased caspase-1 activity [203]</p> <p>Indirect evidence for an altered pattern of toll-like receptors [204]</p>	<p>Increased sensitivity of keratinocytes to activation by allergen contact</p>
	<p>Evidence for an increased cytokine response (including IL-1, TNF-α) of keratinocytes from healthy-appearing skin after stimulation with IFN-γ [174] and in response to topical application of irritants (SDS)/allergens (HDM) [175]</p> <p>Decreased levels of IFN-γ inhibiting transcription factors SOCS1, 2, 3 in lesional AE [179]</p>	<p>Modulation of cutaneous immune responses</p> <p>Enhanced keratinocyte-driven stimulation of epidermal DC</p>
	<p>Increased production of TARC/CCL-17 and MDC/CCL-22 [205] possibly mediated by IFN-γ [169] and inhibited by TGF-β [176]</p>	<p>Augmented attraction of T cells to sites of allergen challenge</p>
	<p>Overexpression of human thymic stromal lymphopoietin (TSLP): activates DC and induces the expression of chemokines (TARC/CCL17 and MDC/CCL22) that selectively attract Th2 cells; TSLP-primed DC induce IL-4, IL-5, IL-13, and TNF-α in responding T cells, but downregulate IFN-γ and IL-10 [168]</p>	<p>The Th2-attracting milieu in acute lesions may negatively affect certain types of ACD reactions</p>
	<p>Increased keratinocyte apoptosis resulting in the release of several factors that are overexpressed in AE lesions and attract CXCR3+ T cells [180]</p>	
	<p>Antigen-presenting cells: dendritic cells, monocytes/macrophages Increased stimulatory capacity of MoDC from atopic donors [181] with enhanced induction of Th2 responses in T cells from atopic donors [182]</p>	<p>Factors that facilitate DC migration and DC-dependent T cell activation may trigger the manifestation of ACD</p>
	<p>Increased production of IL-10 but decreased production of IL-12p40 from MoDC of AE patients after LPS stimulation [183] with IL-10 being a central mediator in the induction of Th2 responses and eosinophilia in a murine model of AE [196]</p>	<p>Preferential stimulation of Th2 cells may antagonize ACD</p>
	<p>Disturbed maturation of MoDC in response to CD40 cross-linking [183]</p> <p>IDEC, a subpopulation of inflammatory epidermal DC with very high expression of FcϵRI, are selectively recruited into the epidermis; upon ligation of FcϵRI, IDEC secrete IL-1α, IL-1β, MCP-1, MCP-3, RANTES TNF-α, and MIP-1α [161]; preactivation of DC might be related to induction of NFκB signaling following ligation of FcϵRI [162]</p>	
	<p>Increased expression of IL-16 in epidermal DC in active lesions and positive atopy patch test reactions, possibly induced by engagement of FcϵRI [206, 185]</p> <p>Strong expression of MDC (CCL22) in cutaneous DC [184]</p>	<p>DC-derived lymphocyte-attracting factors may augment ACD responses</p>
	<p>Effector T cells and cytokine milieu Increased numbers of CD4+ and CD8+ T effector cells with increased expression of Th2 cytokines (IL-4, IL-5, IL-13) and IL-10 in acute skin lesions compared with the skin of healthy individuals, but predominance of Th1 cytokines (IFN-γ) in chronic lesions; cytokine switch is likely to occur under the influence of IL-12 released from DC and macrophages (stimulated by IL-4, bacterial antigens) and eosinophils [164]</p>	<p>A pro-inflammatory cytokine milieu (chronic lesions) established by skin infiltrating cells may act as danger signal and trigger ACD reactions</p> <p>Decreased capacity to mount Th1 responses and preferential attraction of Th2 cell may prevent ACD</p>

Table 17.2. (cont.)

	Atopic eczema	Possible (hypothetical) effect on ACD in AE
Immunologic parameters	<p>Comparative DNA microarray analysis shows overexpression of MCP-4 (CCL-13), PARC (CCL-18), and CTACK (CCL-27) in AE vs psoriasis [207]</p> <p>Several studies indicate specifically elevated serum/plasma levels of Th2-selective chemokines TARC (CCL17), MDC (CCL22), and CTACK (CCL27) and correlation of these markers with disease severity [208, 209, 210]; elevated serum levels of IL-16 [211]</p> <p>Increased expression of CCR3 and CCR4 in (acute) lesions, the receptors for the eosinophil/Th2-recruiting chemokines TARC (CCL17) and MDC (CCL22) [212, 170]</p> <p><i>Extrinsic vs intrinsic</i> Comparative RT-PCR analysis indicates three groups of cytokines: IL-1β, IL-5, and IL-13 are increased in AE compared to healthy skin and higher in extrinsic than in intrinsic forms; IFNγ, IL-12, GM-CSF, IL-4, IL-10 are higher in AE compared to healthy skin with similar levels in extrinsic and intrinsic forms; decreased levels of TNF-α in both AE variants compared to healthy controls [158]</p> <p><i>Acute vs chronic</i> In addition to the Th2/Th1 switch: TGF-β is enhanced in acute and even more in chronic lesions; IL-17 is increased in acute lesions, IL-11 increased in chronic lesions [213]; it is unclear how these findings correlate with:</p> <ul style="list-style-type: none"> - TGF-β suppresses AE-like skin lesions in an established mouse model of AE [177] - TGF-β+/CD4+ T cells suppress Th1- and Th2-mediated allergen-induced skin inflammation in animal models [178] - AE is associated with a low-producer TGF-β1 cytokine genotype [214] 	
	<p>T regulatory cells Role of Treg subpopulations and related cytokines (IL-10, TGF-β) remains to be explored; increased expression of IL-10 and TGF-β have been reported; however, IL-10 may enhance certain mechanisms of allergic inflammation (see text); natural CD4+ CD25+ Treg cells seem to be increased in AE, their suppressive effects may, however, be restricted to Th1 cells; bacterial products may break anergy of these cells [194]</p>	<p>High numbers of Treg cells may counterbalance ACD responses</p> <p>Disturbed expression of anti-inflammatory cytokines (IL-10), that physiologically antagonize ACD, may also influence ACD</p>
	<p>Adhesion molecules Variable upregulation of VCAM-1 on dermal DC and endothelial cells [215]</p> <p>Strong expression of ICAM-3 on CD1+ epidermal and dermal DC [216]</p> <p>Overexpression of α6-integrin in active lesions and ACD reactions in AD patients [217]</p>	<p>Increased expression of adhesion molecules may trigger the influx of inflammatory cells in ACD</p>
	<p>Neuropeptides Evidence for an increased number of substance P (SP) and calcitonin gene-related peptide positive fibers [218]; increase in plasma levels of SP and NGF correlate with disease activity [219]; SP might aggravate AE by increasing the production of TNF α and IL-10 rather than by affecting IL-4 and IFN-γ [220]</p>	<p>Neurogenic inflammation may contribute to the manifestation of ACD</p>

ACD allergic contact dermatitis, APC antigen-presenting cell, AE atopic eczema, DC dendritic cells, HDM house dust mites, SDS sodium dodecyl sulfate, TEWL transepidermal water loss

References

- Nicolas JF, Thivolet J (1988) Immunologic features of atopic dermatitis. *Semin Dermatol* 7:156–162
- Strannegard Ö, Strannegard I-L (1991) Changes in cell-mediated immunity in atopic eczema. In: Ruzicka T, Ring J, Przybilla B (eds) *Handbook of Atopic eczema*. Springer Verlag Berlin Heidelberg New York, pp 221–231
- Bandmann H-J, Breit R, Leutgeb C (1972) Kontaktallergie und Dermatitis atopica. *Arch Derm Forsch* 244:332–334
- Breit R (1981) Positive Epikutantestreaktionen bei Dermatitis atopica. *Hautarzt* 32 [Suppl 5]:147–148
- Blondeel A, Achten G, Dooms-Goossens A, Buekens P, Broeckx W, Oleeff J (1987) Atopie et allergie de contact. *Ann Dermatol Venerol* 114:203–209
- Edman B, Möller H (1992) Contact allergy and contact allergens in atopic skin disease. *Am J Contact Dermatitis* 3:27–29
- Cronin E, McFadden JP (1993) Patients with atopic eczema do become sensitized to contact allergens. *Contact Dermatitis* 28:225–228
- Dotterud LK, Falk ES (1995) Contact allergy in relation to hand eczema and atopic diseases in north Norwegian schoolchildren. *Acta Paediatr* 84:402–406
- Stables GI, Forsyth A, Lever RS (1996) Patch testing in children. *Contact Dermatitis* 34:341–344
- Sanz-Ortega J, DeLaCuadra-Oyanguren J, Martorell-Aragones A, Torro-Demenech I, Cerda-Mir JC, Alvarez-Angel V (1990) Prevalencia de la sensibilizacion a alergenosen de contacto entre la poblacion infantil atopica y no atopica sin dermatitis. *Ann Esp Pediatr* 33:339–342
- Romaguera C, Vilaplana J (1998) Contact dermatitis in children: a 6-year experience (1992–1997). *Contact Dermatitis* 39:277–280
- Bieber T (1995) Pathogenese des atopischen Ekzems und des allergischen Kontakttekzems. Plewig G, Korting HC (Hrsg), *Fortschritte der praktischen Dermatologie und Venerologie*, Springer Verlag, Berlin Heidelberg New York 14:25–29
- Guillet MH, Guillet G (1997) Overview of expected sensitizations in atopic dermatitis. In: Grob JJ, Stern RS, MacRMKie, Weinstock WA (eds) *Epidemiology, causes and prevention of skin diseases*. Blackwell Science, Oxford, pp 245–248
- Neumann C, Marghescu S (1991) Allergic contact eczema and atopic eczema. In: Ruzicka T, Ring J, Przybilla B (eds) *Handbook of atopic eczema*. Springer Verlag Berlin Heidelberg New York, pp 98–106
- Rajka G (1989) *Essential aspects of atopic dermatitis*. Springer, Berlin Heidelberg New York
- Ring J (1990) Atopisches Ekzem und Allergie. In: Braun O-Falco, Ring J (Hrsg), *Fschr prakt Dermatol Venerol* 12: 103–113
- Ring J, Abeck D, Vieluf D (1995) Atopisches Ekzem und Kontakttekzem – Gemeinsamkeiten, Unterschiede und praktische Konsequenzen. Plewig G, Korting HC (Hrsg), *Fortschritte der praktischen Dermatologie und Venerologie*. Springer Verlag, Berlin Heidelberg New York 14: 30–36
- Akhavan A, Cohen SR (2003) The relationship between atopic dermatitis and contact dermatitis. *Clin Dermatol* 21:158–162
- Skog E (1960) Primary irritant and allergic eczematous reactions induced in patients with different dermatoses. *Acta Derm Venereol (Stockh)* 40:307–316
- Jones HE, Lewis CW, McMartin SL (1973) Allergic contact sensitivity in atopic dermatitis. *Arch Dermatol* 107:217–222
- Forsbeck M, Hovmark A, Skog E (1976) Patch testing, tuberculin testing and sensitization with dinitrochlorobenzene and nitrosodimethylanilini of patients with atopic dermatitis. *Acta Derm Venereol (Stockh)* 56:135–138
- Rees J, Friedmann PS, Matthews JNS (1990) Contact sensitivity to dinitrochlorobenzene is impaired in atopic subjects – controversy revisited. *Arch Dermatol* 126:1173–1175
- Uehara M, Sawai T (1989) A longitudinal study of contact sensitivity in patients with atopic dermatitis. *Arch Dermatol* 125:366–368
- Nielsen NH, Linneberg A, Menne T, Madsen F, Frolund L, Dirksen A, Jorgensen T (2002) Incidence of allergic contact sensitization in Danish adults between 1990 and 1998; the Copenhagen Allergy Study, Denmark. *Br J Dermatol* 147:487–492
- Nielsen NH, Menné T (1996) The relationship between IgE-mediated and cell-mediated hypersensitivities in an unselected Danish population: The Glostrup Allergy Study, Denmark. *Br J Dermatol* 134:669–672
- Mortz CG, Lauritsen JM, Bindsløv-Jensen C, Andersen KE (2001) Prevalence of atopic dermatitis, asthma, allergic rhinitis, and hand and contact dermatitis in adolescents. The Odense Adolescence Cohort Study on Atopic Diseases and Dermatitis. *Br J Dermatol* 144:523–532
- Schäfer T, Bohler E, Ruhdorfer S, Weigl L, Wessner D, Filipiak B, Wichmann HE, Ring J (2001) Epidemiology of contact allergy in adults. *Allergy* 56:1192–1196
- Schnuch A, Geier J, Uter W, Frosch PJ, Lehmacher W, Aberer W, Agathos M, Arnold R, Fuchs T, Laubstein B, Lischka G, Pietrzyk PM, Rakoski J, Richter G, Rueff F (1997) National rates and regional differences in sensitization to allergens of the standard series. Population-adjusted frequencies of sensitization (PAFS) in 40,000 patients from a multicenter study (IVDK). *Contact Dermatitis* 37:200–209
- Seidenari S, Manzini BM, Danese P, Motolese A (1990) Patch and prick test study of 593 healthy subjects. *Contact Dermatitis* 23:162–167
- DeGroot AC (1990) The frequency of contact allergy in atopic patients with dermatitis. *Contact Dermatitis* 22: 273–277
- Brasch J, Schnuch A, Uter W (2003) Patch test reaction patterns in patients with a predisposition to atopic dermatitis. *Contact Dermatitis* 49:197–201
- Jappe U, Schnuch A, Uter W (2003) Frequency of sensitization to antimicrobials in patients with atopic eczema compared with non-atopic individuals: analysis of multicentre surveillance data, 1995–1999. *Br J Dermatol* 149:87–93
- Wilkinson JD, Hambly EM, Wilkinson DS (1980) Comparison of patch test results in two adjacent areas of England. II. Medicaments. *Acta Derm Venereol Stockh* 60:245–249
- Andersen KE, Veien NK (1985) Biocide patch tests. *Contact Dermatitis* 12:99–103

35. Guillet MH, Guillet G (1996) Enquête allergologique chez 251 malades atteints de dermatite atopique modérée ou sévère – fréquence et intérêt du dépistage de l'eczéma de contact, de l'allergie alimentaire et de la sensibilisation aux aéroallergènes. *Ann Dermatol Venereol* 123:157–164
36. Lever R, Forsyth A (1992) Allergic contact dermatitis in atopic dermatitis. *Acta Derm Venereol (Stockh)* 71 [Suppl 176]:95–98
37. Giordano-Labadie F, Rance F, Pellegrin F, Bazex J, Dutau G (1997) Fréquence de l'allergie de contact au cours de la dermatite atopique de l'enfant: résultat d'une étude prospective de 137 cas (abstract C23). *Ann Dermatol Venereol* 124 [Suppl 1]:S17
38. Giordano-Labadie F, Rance F, Pellegrin F, Bazex J, Dutau G, Schwarze HP (1999) Frequency of contact allergy in children with atopic dermatitis: results of a prospective study of 137 cases. *Contact Dermatitis* 40:192–195
39. Huber A, Fartasch M, Diepgen TL, Baurle G, Hornstein OP (1987) Auftreten von Kontaktallergien beim atopischen Ekzem. Zusammenhänge mit gleichzeitig gefundenen atopischen Merkmalen. *Dermatosen* 35:119–123
40. Lewis FM, Shah M, Gawkrödger DJ (1995) Contact sensitivity in atopic dermatitis. *Am J Contact Dermatitis* 6:150–152
41. Nielsen NH, Linneberg A, Menne T, Maden F, Frolund L, Dirksen A, Jørgensen T (2001) Allergic contact sensitization in an adult Danish population: two cross-sectional surveys eight years apart (The Copenhagen Allergy Study). *Acta Derm Venereol* 81:31–34
42. Christophersen J, Menne T, Tanghoj P, Andersen KE, Brandrup F, Kaaber K, Osmundsen PE, Thestrup Pedersen K, Veien NK (1989) Clinical patch test data evaluated by multivariate analysis. Danish Contact Dermatitis Group. *Contact Dermatitis* 21:291–299
43. Diepgen TL, Fartasch M, Hornstein OP (1989) Evaluation and relevance of atopic basic and minor features in patients with atopic dermatitis and in the general population. *Acta Derm Venereol (Stockh) Suppl* 144:50–54
44. Nethercott JR, Holness DL, Adams RM, Belsito DV, DeLeo VA, Emmett EA, Fowler JF, Fisher AM, Larsen WG, Maibach HI, Marks JG, Rietschel RL, Rosenthal L, Schorr W, Storrs FJ, Taylor JS (1994) Multivariate analysis of the effect of selected factors on the elicitation of patch test response to 28 common environmental contactants in North America. *Am J Contact Dermatitis* 5:13–18
45. Uter W, Gefeller O, Geier J, Lessmann H, Pfahlberg A, Schnuch A (2003) Untersuchungen zur Abhängigkeit der Sensibilisierung gegen wichtige Allergene von arbeitsbedingten sowie individuellen Faktoren. Schriftenreihe der Bundesanstalt für Arbeitsschutz und Arbeitsmedizin Wirtschaftsverlag NW, Bremerhaven, Fb 949
46. Uter W, Pfahlberg A, Gefeller O, Geier J, Schnuch A (2003) Risk factors for contact allergy to nickel – results of a multifactorial analysis. *Contact Dermatitis* 48:33–38
47. Mortz CG, Lauritsen JM, Bindselev-Jensen C, Andersen KE (2002) Contact allergy and allergic contact dermatitis in adolescents: prevalence measures and associations. *Acta Derm Venereol (Stockh)* 82:352–358
48. Mattila L, Kilpeläinen M, Terho EO, Koskenvuo M, Helenius H, Kalimo K (2001) Prevalence of nickel allergy among Finnish university students in 1995. *Contact Dermatitis* 44:218–223
49. Szepletowski JC, McKenzie RC, Keohane SG, Aldrige RD, Hunter JAA (1997) Atopic and non-atopic individuals react to nickel challenge in a similar way. A study of the cytokine profile in nickel-induced contact dermatitis. *Br J Dermatol* 137:195–200
50. Buchvald D, Lundberg L (2004) Impaired responses of peripheral blood mononuclear cells to nickel in patients with nickel-allergic contact dermatitis and concomitant atopic dermatitis. *Br J Dermatol* 150:484–492
51. Uter W, Geier J, Fuchs T (2000) Contact allergy to polidocanol, 1992 to 1999. *J Allergy Clin Immunol* 106:1203–1204
52. Uter W, Schnuch A, Geier J, Pfahlberg A, Gefeller O (2001) Association between occupation and contact allergy to the fragrance mix: a multifactorial analysis of national surveillance data. *Occup Environ Med* 58:392–398
53. Singh P, Roberts MS (1996) Local deep tissue penetration of compounds after dermal application: structure-tissue penetration relationships. *J Pharmacol Exp Ther* 279:908–917
54. Pior J, Vogl T, Sorg C, Macher E (1999) Free hapten molecules are dispersed by way of the bloodstream during contact sensitization to fluorescein isothiocyanate. *J Invest Dermatol* 113:888–893
55. Schaefer H, Redelmeier TE (2001) Skin penetration. In: Rycroft RJG, Menne T, Frosch PJ, Lepoittevin J-P (eds), *Textbook of contact dermatitis*, 3rd edn. Springer, Berlin Heidelberg New York, pp 209–225
56. Berard F, Marty JP, Nicolas JF (2003) Allergen penetration through the skin. *Eur J Dermatol* 13:324–330
57. Monteiro-Riviere NA (1996) Anatomical factors affecting barrier function. In: Marzulli FN, Maibach HI (eds), *Dermatotoxicology*, 5th edn. Taylor & Francis, Washington DC, pp 3–17
58. Madison KC (2003) Barrier function of the skin: “La Raison d’Etre” of the epidermis. *J Invest Dermatol* 121:231
59. Elias PM, Wood LC, Feingold KR (1999) Epidermal pathogenesis of inflammatory dermatoses. *Am J Contact Dermatitis* 10:119–126
60. Ashby J, Hilton J, Dearman RJ, Kimber I (1995) Streptozotocin: inherent but not expressed skin sensitizing activity. *Contact Dermatitis* 33:165–167
61. Bos JD, Meinardi MMHM (2000) The 500-Dalton rule for the skin penetration of chemical compounds and drugs. *Exp Dermatol* 9:165–169
62. Barratt MD (1995) Quantitative structure-activity relationships for skin permeability. *Toxic in Vitro* 9:27–37
63. Hostynek JJ (1995) Predicting absorption of fragrance chemicals through human skin. *J Soc Cosmet Chem* 46:221–229
64. Lahti A, Basketter D (2001) Immediate contact reactions. In: Rycroft RJG, Menne T, Frosch PJ, Lepoittevin J-P (eds), *Textbook of contact dermatitis*, 3rd edn. Springer, Berlin Heidelberg New York, pp 111–132
65. Ale SI, Maibach HI (2000) Occupational contact urticaria. In: Kanerva L, Elsner P, Wahlberg JE, Maibach HI (eds) *Handbook of occupational dermatology*. Springer Verlag, Berlin Heidelberg New York, pp 200–216
66. Ring J, Kunz B, Bieber T, Vieluf D, Przybilla B (1989) The

- 'atopy patch test' with aeroallergens in atopic eczema. *J Allergy Clin Immunol* 82:195
67. Smith Pease CK, White IR, Basketter DA (2002) Skin as a route of exposure to protein allergens. *Clin Exp Dermatol* 27:296–300
 68. Field EA (1998) Atopy and other risk factors for UK dentists reporting an adverse reaction to latex gloves. *Contact Dermatitis* 38:132–136
 69. Porri F, Lemiere C, Birnbaum J, Guilloux L, Didelot R, Vervoet D, Charpin D (1995) Prevalence of latex allergy in atopic and non-atopic subjects from the general population (abstract 56). *J Allergy Clin Immunol* 95:154
 70. Boxer M (1996) Hand dermatitis: a risk factor for latex hypersensitivity. *J All Clin Immunol* 98:855–856
 71. Hotchkiss SAM (1998) Absorption of fragrance ingredients using in vitro models with human skin. In: Frosch PJ, Johansen JD, White IR (eds) *Fragrances – beneficial and adverse effects*. Springer, Berlin Heidelberg New York, pp 125–135
 72. Baur X, Chen Z, Allmers H, Raulf-Heimsoth M (1998) Results of wearing test with two different latex gloves with and without the use of skin-protection cream. *Allergy* 53:441–444
 73. Schaefer H, Redelmeier ThE (1996) Skin barrier. Principles of percutaneous absorption. Karger, Basel
 74. Lademann J, Sterry W (eds) (2001) Structure and function of the stratum corneum as border organ. *Skin Pharmacol Appl Skin Physiol* 14 [Suppl 1]. Karger, Basel
 75. Proksch E, Jensen JM, Elias PM (2003) Skin lipids and epidermal differentiation in atopic dermatitis. *Clin Dermatol* 21:134–144
 76. Nakagawa N, Sakai S, Matsumoto M, Yamada K, Nagano M, Yuki T, Sumida Y, Uchiwa H (2004) Relationship between NMF (lactate and potassium) content and the physical properties of the stratum corneum in healthy subjects. *J Invest Dermatol* 122:755–763
 77. Elias PM, Tsai J, Menon GK, Holleran WM, Feingold KR (2002) The potential of metabolic interventions to enhance transdermal drug delivery. *J Invest Dermatol Symp Proc* 7:79–85
 78. Lademann J, Otberg N, Richter H, Jacobi U, Schaefer H, Blume-Peytavi U, Sterry W (2003) Follikuläre Penetration. Ein entscheidender Penetrationsweg von topisch applizierten Substanzen. *Hautarzt* 54:321–323
 79. Schaefer H, Lademann J (2001) The role of follicular penetration. *Skin Pharmacol Appl Skin Physiol* 14:23–27
 80. Elias PM, Feingold KR (2001) Coordinate regulation of epidermal differentiation and barrier homeostasis. *Skin Pharmacol Appl Skin Physiol* 4:28–34
 81. Elias PM (2004) The epidermal permeability barrier: from the early days at Harvard to emerging concepts. *J Invest Dermatol* 122: xxxvi–xxxix
 82. Kobayashi H, Aiba S, Yoshino Y, Tagami H (2003) Acute cutaneous barrier disruption activates epidermal p44/42 and p38 mitogen-activated protein kinases in human and hairless guinea pig skin. *Exp Dermatol* 12:734–746
 83. Jensen JM, Schutze S, Forl M, Kronke M, Proksch E (1999) Roles for tumor necrosis factor receptor p55 and sphingomyelinase in repairing the cutaneous permeability barrier. *J Clin Invest* 104:1761–1770
 84. Rivier M, Castiel I, Safonova I, Ailhaud G, Michel S (2000) Peroxisome proliferator-activated receptor-alpha enhances lipid metabolism in skin equivalent model. *J Invest Dermatol* 114:681–687
 85. Sheu MY, Fowler AJ, Kao J, Schmuth M, Schoonjans K, Auwerx J, Fluhr JW, Man MQ, Elias PM, Feingold KR (2002) Topical peroxisome proliferator activated receptor-alpha activators reduce inflammation in irritant and allergic contact dermatitis models. *J Invest Dermatol* 118: 94–101
 86. Sator P-G, Schmidt JB, Hönigsmann H (2003) Comparison of epidermal hydration and skin surface lipids in healthy individuals and in patients with atopic dermatitis. *J Am Acad Dermatol* 48:352–358
 87. Bleck O, Abeck D, Ring J, Hoppe U, Vietzke JP, Wolber R, Brandt O, Schreiner V (1999) Two ceramide subfractions detectable in Cer(AS) position by HPTLC in skin surface lipids of non-lesional skin of atopic eczema. *J Invest Dermatol* 113:894–900
 88. Fartasch M, Diepgen TL (1992) The barrier function in atopic dry skin. Disturbance of membrane-coating granule exocytosis and formation of epidermal lipids? *Acta Derm Venereol (Stockh) Suppl* 176:26–31
 89. Tupker RA, Pinnagoda J, Coenraads PJ, Nater JP (1990) Susceptibility to irritants: role of barrier function, skin dryness and history of atopic dermatitis. *Br J Dermatol* 123:199–205
 90. Löffler H, Effendy I (1999) Skin susceptibility of atopic individuals. *Contact Dermatitis* 40:239–242
 91. Sakurai K, Sugiura H, Matsumoto M, Uehara M (2002) Occurrence of patchy parakeratosis in normal-appearing skin in patients with active atopic dermatitis and in patients with healed atopic dermatitis: a cause of impaired barrier function of the atopic skin. *J Dermatol Sci* 30: 37–42
 92. Matsumoto M, Sugiura H, Uehara M (2000) Skin barrier function in patients with completely healed atopic dermatitis. *J Dermatol Sci* 23:178–182
 93. Stolz R, Hinnen U, Elsner P (1997) An evaluation of the relationship between 'atopic skin' and skin irritability in metalworker trainees. *Contact Dermatitis* 36:281–284
 94. Tupker RA, Coenraads PJ, Pinnagoda J, Nater JP (1989) Baseline transepidermal water loss (TEWL) as a prediction of susceptibility to sodium lauryl sulphate. *Contact Dermatitis* 20:265–269
 95. Yoshiike T, Aikawa Y, Sindhvananda J, Suto H, Nishimura K, Kawamoto T, Ogawa (1993) Skin barrier defect in atopic dermatitis: increased permeability of the stratum corneum using dimethyl sulfoxide and theophylline. *J Dermatol Sci* 5:92–96
 96. Junghans V, Gutgesell C, Jung T, Neumann C (1997) Epidermal cytokines IL-1-beta, TNF-alpha and IL-12 in patients with atopic dermatitis: response to application of house dust mite antigens. *Arch Dermatol Res* 289 [Suppl]: A45
 97. Gfesser M, Rakoski J, Ring J (1996) The disturbance of epidermal barrier function in atopy patch test reactions in atopic eczema. *Br J Dermatol* 135:560–565
 98. Rustemeyer T, van Hoogstraten IMW, von Blomberg BME, Scheper RJ (2001) Mechanisms in allergic contact dermati-

- tis. In: Rycroft RJG, Menne T, Frosch PJ, Lepoittevin J-P (eds) *Textbook of contact dermatitis*, 3rd edn. Springer, Berlin Heidelberg New York, pp 13–58
99. Dearman RJ, Kimber I (2003) Factors influencing the induction phase of skin sensitization. *Am J Contact Dermatitis* 14:188–194
100. Watanabe H, Unger M, Tuvel B, Wang B, Sauder DN (2002) Contact hypersensitivity: the mechanism of immune responses and T cell balance. *J Interferon Cytokine Res* 22:407–412
101. Lepoittevin JP, Basketter DA, Goossens A, Karlberg AT (eds) (1998) *Allergic contact dermatitis. The molecular basis*. Springer, Berlin Heidelberg New York
102. Sadhra S, Foulds IS, Gray CN (1998) Oxidation of resin acids in colophony (rosin) and its implications for patch testing. *Contact Dermatitis* 39:58–63
103. Karlberg A-T, Shao LP, Nilsson U, Gäfvert E, Nilsson JLG (1994) Hydroperoxides in oxidized delta-limonene identified as potent contact allergens. *Arch Dermatol Res* 286:97–103
104. Smith Pease CK, Basketter DA, Patlewicz GY (2003) Contact allergy: the role of skin chemistry and metabolism. *Clin Exp Dermatol* 28:177–183
105. Smith CK, Hotchkiss SHAM (2001) *Allergic contact dermatitis. Chemical and metabolic mechanisms*. Taylor & Francis, London
106. Cheung C, Hotchkiss SA, Pease CK (2003) Cinnamic compound metabolism in human skin and the role metabolism may play in determining relative sensitisation potency. *J Dermatol Sci* 31:9–19
107. Schnuch A, Westphal GA, Müller MM, Schulz TG, Geier J, Brasch J, Merk HF, Kawakubo Y, Richter G, Koch P, Fuchs Th, Gutgesell C, Reich K, Gebhardt M, Becker D, Grabbe J, Szliska C, Lischka G, Aberer W, Hallier E (1998) Genotype and phenotype of N-acetyltransferase 2 (NAT2) polymorphism in patients with contact allergy. *Contact Dermatitis* 38:209–211
108. Westphal G-A, Schnuch A, Schulz T-G, Reich K, Aberer W, Brasch J, Koch P, Wessbecher R, Szliska C, Bauer A, Hallier E (2000) Homozygous gene deletions of the glutathione S-transferases M1 and T1 are associated with thimerosal sensitization. *Int Arch Occup Environ Health* 73:384–388
109. Staquet MJ, Piccardi N, Msika P, Schmitt D (2002) [Langerhans cell migration. An essential step in the induction of contact hypersensitivity]. *Ann Dermatol Venerol* 129:1071–1077
110. Cumberbatch M, Dearman RJ, Griffiths CE, Kimber I (2003) Epidermal Langerhans cell migration and sensitization to chemical allergens. *APMIS* 111:797–804
111. Sebastiani S, Albanesi C, De PO, Puddu P, Cavani A, Girolmoni G (2002) The role of chemokines in allergic contact dermatitis. *Arch Dermatol Res* 293:552–559
112. Kobayashi Y, Matsumoto M, Kotani M, Makino T (1999) Possible involvement of matrix metalloproteinase-9 in Langerhans cell migration and maturation. *J Immunol* 163:5989–5993
113. Zepter K, Häffner A, Soohoo LF, De-Luca D, Tang HP, Fisher P, Chavinson J, Elmets CA (1997) Induction of biologically active IL-1-beta-converting enzyme and mature IL-1 beta in human keratinocytes by inflammatory and immunologic stimuli. *J Immunol* 159:6203–6208
114. Enk AH (2003) Pathophysiologie der allergischen Kontaktdermatitis. *Allergo J* 12:501–507
115. Becke FM, Hehlhans T, Brockhoff G, Mannel DN (2001) Development of allergic contact dermatitis requires activation of both tumor necrosis factor receptors. *Eur Cytokine Netw* 12:45–50
116. Griffiths CEM, Cumberbatch M, Tucker SC, Dearman RJ, Andrew S, Headon DR, Kimber I (2001) Exogenous topical lactoferrin inhibits allergen-induced Langerhans cell migration and cutaneous inflammation in humans. *Br J Dermatol* 144:715–725
117. Sato K, Kawasaki H, Nagayama H, Enomoto M, Morimoto C, Tadokoro K, Juji T, Takahashi TA (2000) TGF-beta 1 reciprocally controls chemotaxis of human peripheral blood monocyte-derived dendritic cells via chemokine receptors. *J Immunol* 164:2285–2295
118. Cumberbatch M, Tucker SC, Andrew S et al (1999) Influence of lactoferrin on allergen-induced Langerhans cell migration in humans. *Br J Dermatol* 140:789
119. Bhushan M, Cumberbatch M, Dearman RJ, Kimber I, Griffiths CEM (2001) Human epidermal Langerhans cell migration in response to interleukin-1 and regulation by topical lactoferrin. *Br J Dermatol* 145:96
120. Kuenzly S, Saurat J-H (2003) Peroxisome proliferator-activated receptors in cutaneous biology. *Br J Dermatol* 149:229–236
121. Zhang X, Young HA (2002) PPAR and immune system—what do we know? *Int Immunopharmacol* 2:1029–1044
122. Angeli V, Hammad H, Staels B, Capron M, Lambrecht BN, Trottein F (2003) Peroxisome proliferator-activated receptor gamma inhibits the migration of dendritic cells: consequences for the immune response. *J Immunol* 170:5295–5301
123. Inaba K, Turley S, Iyoda T, Yamaide F, Shimoyama S, Reis e-Sousa C, Germain RN, Mellman I, Steinman RM (2000) The formation of immunogenic major histocompatibility complex class II-peptide ligands in lysosomal compartments of dendritic cells is regulated by inflammatory stimuli. *J Exp Med* 191:927–936
124. McLellan AD, Bröcker E-B, Kämpgen E (2000) Dendritic cell activation by danger and antigen-specific T-cell signalling. *Exp Dermatol* 9:313–322
125. Cumberbatch M, Scott RC, Basketter DA, Scholes EW, Hilton J, Dearman RJ, Kimber I (1993) Influence of sodium lauryl sulphate on 2,4-dinitrochlorobenzene-induced lymph node activation. *Toxicology* 77:181–191
126. Grabbe S, Schwarz T (1998) Immunoregulatory mechanisms involved in elicitation of allergic contact hypersensitivity. *Immunol Today* 19:37–44
127. Zhang L, Tinkle SS (2000) Chemical activation of innate and specific immunity in contact dermatitis. *J Invest Dermatol* 115:168–176
128. Sabroe I, Parker LC, Wilson AG, Whyte MK, Dower SK (2002) Toll-like receptors: their role in allergy and non-allergic inflammatory disease. *Clin Exp Allergy* 32:984–989
129. McFadden JP, Basketter DA (2000) Contact allergy, irritancy and 'danger'. *Contact Dermatitis* 42:123–127

130. Smith HR, Holloway D, Armstrong DK, Basketter DA, McFadden JP (2000) Irritant thresholds in subjects with colophony allergy. *Contact Dermatitis* 42:95–97
131. Lepoittevin J-P, Goossens A (1998) Molecular basis for the recognition of haptens by T lymphocytes. In: Lepoittevin J-P, Basketter DA, Goossens A, Karlberg A-T (eds) *Allergic contact dermatitis the molecular basis*. Springer Verlag, Berlin Heidelberg New York, pp 112–128
132. Steinbrink K, Kolde G, Sorg C, Macher E (1996) Induction of low zone tolerance to contact allergens in mice does not require functional Langerhans cells. *J Invest Dermatol* 107:243–247
133. Steinbrink K, Sorg C, Macher E (1996) Low zone tolerance to contact allergens in mice: a functional role for CD8. T helper type 2 cells. *J Exp Med* 183:759–768
134. Nakae S, Naruse-Nakajima C, Sudo K, Horai R, Asano M, Iwakura Y (2001) IL-1 alpha, but not IL-1 beta, is required for contact-allergen-specific T cell activation during the sensitization phase in contact hypersensitivity. *Int Immunol* 13:1471–1478
135. Hendriks J, Xiao Y, Borst J (2003) CD27 promotes survival of activated T cells and complements CD28 in generation and establishment of the effector T cell pool. *J Exp Med* 198:1369–1380
136. Chen A, McAdam AJ, Buhlmann JE, Scott S, Lupher ML Jr, Greenfield EA, Baum PR, Fanslow WC, Calderhead DM, Freeman GJ, Sharpe AH (1999) OX40-ligand has a critical costimulatory role in dendritic cell: T-cell interactions. *Immunity* 11:689–698
137. Nuriy S, Enomoto S, Azuma M (2001) The role of CTLA-4 in murine contact hypersensitivity. *J Invest Dermatol* 116:764–768
138. Wang L-F, Sun C-C, Wu J-T, Lin R-H (1999) Epicutaneous administration of hapten through patch application augments TH2 responses which can downregulate the elicitation of murine contact hypersensitivity. *Clin Exp Allergy* 29:271–279
139. Kitagaki H, Ono N, Hayakawa K, Kitazawa T, Watanabe K, Shiohara T (1997) Repeated elicitation of contact hypersensitivity induces a shift in cutaneous cytokine milieu from a T helper cell type 1 to a T helper cell type 2 profile. *J Immunol* 159:2484–2491
140. Kitagaki H, Kimishima M, Teraki Y, Hayakawa J, Hayakawa K, Fujisawa S, Shiohara T (1999) Distinct in vivo and in vitro cytokine profiles of draining lymph node cells in acute and chronic phases of contact hypersensitivity: importance of a type 2 cytokine-rich cutaneous milieu for the development of an early-type response in the chronic phase. *J Immunol* 163:1265–1273
141. Nasorri F, Sebastiani S, Mariani V, De Pita O, Puddu P, Girolomoni G, Cavani A (2002) Activation of nickel-specific CD4+ T lymphocytes in the absence of professional antigen-presenting cells. *J Invest Dermatol* 118:172–179
142. Kimber I, Dearman RJ (2002) Allergic contact dermatitis: the cellular effectors. *Contact Dermatitis* 46:1–5
143. Cavani A, Albanesi C, Traidl C, Sebastiani S, Girolomoni G (2001) Effector and regulatory T cells in allergic contact dermatitis. *Trends Immunol* 22:118–120
144. Yawalkar N, Egli F, Brand CU, Pichler WJ, Braathen LR (2000) Antigen-presenting cells and keratinocytes express interleukin-12 in allergic contact dermatitis. *Contact Dermatitis* 42:18–22
145. Wang B, Fujisawa H, Zhuang L, Freed I, Howell BG, Shahid S, Shivji GM, Mak TW, Sauder DN (2000) CD4+ Th1 and CD8+ type 1 cytotoxic T cells both play a crucial role in the full development of contact hypersensitivity. *J Immunol* 165:6783–6790
146. Xu B, Bulfone-Paus S, Aoyama K, Yu S, Huang P, Morimoto K, Matsushita T, Takeuchi T (2003) Role of Fas/Fas ligand-mediated apoptosis in murine contact hypersensitivity. *Int Immunopharmacol* 3:927–938
147. Martin S, Simon JC (2001) Effector T cells and regulatory T cells in allergic contact dermatitis. *ACI Int* 13:117–121
148. Trautmann A, Akdis M, Kleemann D, Altnauer F, Simon HU, Graeve T, Noll M, Brocker EB, Blaser K, Akdis CA (2000) T cell-mediated Fas-induced keratinocyte apoptosis plays a key pathogenetic role in eczematous dermatitis. *J Clin Invest* 106:25–35
149. Kehren J, Desvignes C, Krasteva M et al (1999) Cytotoxicity is mandatory for CD8+ T cell-mediated contact hypersensitivity. *J Exp Med* 189:779–786
150. Akdis M, Trautmann A, Klunker S, Daigle I, Kucuksezer UC, Deglmann W, Disch R, Blaser K, Akdis CA (2003) T helper (Th) 2 predominance in atopic diseases is due to preferential apoptosis of circulating memory/effector Th1 cells. *FASEB J* 17:1026–1035
151. Asherson GL, Dieli F, Sireci G, Salerno A (1996) Role of IL-4 in delayed hypersensitivity. *Clin Exp Immunol* 103:1–4
152. Traidl C, Jugert F, Krieg T, Merk H, Hunzelmann N (1999) Inhibition of allergic contact dermatitis to DNCB but not to oxazolone in interleukin-4-deficient mice. *J Invest Dermatol* 112:476–482
153. Cavani A, Mei D, Guerra E, Corinti S, Giani M, Pirrotta L, Puddu P, Girolomoni G (1998) Patients with allergic contact dermatitis to nickel and nonallergic individuals display different nickel-specific T cell responses. Evidence for the presence of effector CD8+ and regulatory CD4+ T cells. *J Invest Dermatol* 111:621–628
154. Masjedi K, Ahlborg N, Gruvberger B, Bruze M, Karlberg AT (2003) Methylisothiazolinones elicit increased production of both T helper (Th)1- and Th2-like cytokines by peripheral blood mononuclear cells from contact allergic individuals. *Br J Dermatol* 149:1172–1182
155. Doyen V, Rubio M, Braun D, Nakajima T, Abe J, Saito H, Delespesse G, Sarfati M (2003) Thrombospondin 1 is an autocrine negative regulator of human dendritic cell activation. *J Exp Med* 198:1277–1283
156. Bondesson L, Nordlind K, Mutt V, Lidén S (1996) Inhibitory effect of vasoactive intestinal polypeptide and ketanserine on established allergic contact dermatitis in man. *Acta Derm Venereol (Stockh)* 76:102–106
157. Grabbe S, Bhardwaj RS, Mahnke K, Simon MM, Schwarz T, Luger TA (1996) alpha-Melanocyte-stimulating hormone induces hapten-specific tolerance in mice. *J Immunol* 156:473–478
158. Jeong CW, Ahn KS, Rho NK, Park YD, Lee DY, Lee JH, Lee ES, Yang JM (2003) Differential in vivo cytokine mRNA expression in lesional skin of intrinsic vs. extrinsic atopic dermatitis patients using semiquantitative RT-PCR. *Clin Exp Allergy* 33:1717–1724

159. Novak N, Bieber T (2003) Allergic and nonallergic forms of atopic diseases. *J Allergy Clin Immunol* 112:252–262
160. Akdis CA, Akdis M, Simon D, Dibbert B, Weber M, Gratzl S, Kreyden O, Disch R, Wuthrich B, Blaser K, Simon HU (1999) T cells and T cell-derived cytokines as pathogenic factors in the nonallergic form of atopic dermatitis. *J Invest Dermatol* 113:628–634
161. Novak N, Kraft S, Haberstick J, Geiger E, Allam P, Bieber T (2002) A reducing microenvironment leads to the generation of FcεRI high inflammatory dendritic epidermal cells (IDEC). *J Invest Dermatol* 119:842–849
162. Kraft S, Novak N, Katoh N, Bieber T, Rupec RA (2002) Aggregation of the high-affinity IgE receptor Fc(ε)RI on human monocytes and dendritic cells induces NF-κB activation. *J Invest Dermatol* 118:830–837
163. Akdis M, Simon HU, Weigl L, Kreyden O, Blaser K, Akdis CA (1999) Skin homing (cutaneous lymphocyte-associated antigen-positive) CD8+ T cells respond to superantigen and contribute to eosinophilia and IgE production in atopic dermatitis. *J Immunol* 163:466–475
164. Girolomoni G, Sebastiani S, Albanesi C, Cavani A (2001) T-cell subpopulations in the development of atopic and contact allergy. *Curr Opin Immunol* 13:733–737
165. Romagnani S (2004) Immunologic influences on allergy and the TH1/TH2 balance. *J Allergy Clin Immunol* 113:395–400
166. Von Mutius E (2004) Influences in allergy: epidemiology and the environment. *J Allergy Clin Immunol* 113:373–379
167. Biedermann T, Röcken M, Carballido JM (2004) TH1 and TH2 lymphocyte development and regulation of TH cell-mediated immune responses of the skin. *J Invest Dermatol Symp Proc* 9:5–14
168. Soumelis V, Reche PA, Kanzler H, Yuan W, Edward G, Homey B, Gilliet M, Ho S, Antonenko S, Lauerma A, Smith K, Gorman D, Zurawski S, Abrams J, Menon S, McClanahan T, Waal-Malefyt Rd R, Bazan F, Kastelein RA, Liu YJ (2002) Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. *Nat Immunol* 3:673–680
169. Horikawa T, Nakayama T, Hikita I, Yamada H, Fujisawa R, Bito T, Harada S, Fukunaga A, Chantry D, Gray PW, Morita A, Suzuki R, Tezuka T, Ichihashi M, Yoshie O (2002) IFN-γ-inducible expression of thymus and activation-regulated chemokine/CCL17 and macrophage-derived chemokine/CCL22 in epidermal keratinocytes and their roles in atopic dermatitis. *Int Immunol* 14:767–773
170. Zheng X, Nakamura K, Furukawa H, Nishibu A, Takahashi M, Tojo M, Kaneko F, Kakinuma T, Tamaki K (2003) Demonstration of TARC and CCR4 mRNA expression and distribution using in situ RT-PCR in the lesional skin of atopic dermatitis. *J Dermatol* 30:26–32
171. Nakayama T, Fujisawa R, Yamada H, Horikawa T, Kawasaki H, Hieshima K, Izawa D, Fujii S, Tezuka T, Yoshie O (2001) Inducible expression of a CC chemokine liver- and activation-regulated chemokine (LARC)/macrophage inflammatory protein (MIP)-3 α/CCL20 by epidermal keratinocytes and its role in atopic dermatitis. *Int Immunol* 13:95–103
172. Schmutz M, Neyer S, Rainer C, Grassegger A, Fritsch P, Romani N, Heufler C (2002) Expression of the C-C chemokine MIP-3 α/CCL20 in human epidermis with impaired permeability barrier function. *Exp Dermatol* 11:135–142
173. Kakinuma T, Saeki H, Tsunemi Y, Fujita H, Asano N, Mitsui H, Tada Y, Wakugawa M, Watanabe T, Torii H, Komine M, Asahina A, Nakamura K, Tamaki K (2003) Increased serum cutaneous T cell-attracting chemokine (CCL27) levels in patients with atopic dermatitis and psoriasis vulgaris. *J Allergy Clin Immunol* 111:592–597
174. Pastore S, Corinti S, La Placa M, Didona B, Girolomoni G (1998) Interferon-γ promotes exaggerated cytokine production in keratinocytes cultured from patients with atopic dermatitis. *J Allergy Clin Immunol* 101:538–544
175. Junghans V, Gutgesell C, Jung T, Neumann C (1998) Epidermal cytokines IL-1β, TNF-α, and IL-12 in patients with atopic dermatitis: response to application of house dust mite antigens. *J Invest Dermatol* 111:1184–1188
176. Zheng X, Nakamura K, Tojo M, Oyama N, Nishibu A, Satoh M, Kakinuma T, Wakugawa M, Tamaki K, Kaneko F (2002) TGF-β1-mediated regulation of thymus and activation-regulated chemokine (TARC/CCL17) synthesis and secretion by HaCaT cells co-stimulated with TNF-α and IFN-γ. *J Dermatol Sci* 30:154–160
177. Sumiyoshi K, Nakao A, Ushio H, Mitsuishi K, Okumura K, Tsuboi R, Ra C, Ogawa H (2002) Transforming growth factor-β1 suppresses atopic dermatitis-like skin lesions in NC/Nga mice. *Clin Exp Allergy* 32:309–314
178. Terui T, Sano K, Shirota H, Kunikata N, Ozawa M, Okada M, Honda M, Tamura G, Tagami H (2001) TGF-β-producing CD4+ mediastinal lymph node cells obtained from mice tracheally tolerized to ovalbumin (OVA) suppress both Th1- and Th2-induced cutaneous inflammatory responses to OVA by different mechanisms. *J Immunol* 167:3661–3667
179. Federici M, Giustizieri ML, Scarponi C, Girolomoni G, Albanesi C (2002) Impaired IFN-γ-dependent inflammatory responses in human keratinocytes overexpressing the suppressor of cytokine signaling 1. *J Immunol* 169:434–442
180. Klunker S, Trautmann A, Akdis M, Verhagen J, Schmid-Grendelmeier P, Blaser K, Akdis CA (2003) A second step of chemotaxis after transendothelial migration: keratinocytes undergoing apoptosis release IFN-γ-inducible protein 10, monokine induced by IFN-γ, and IFN-γ-inducible α-chemoattractant for T cell chemotaxis toward epidermis in atopic dermatitis. *J Immunol* 171:1078–1084
181. Den Heuvel MM van, Vanhee DD, Postmus PE, Hoefsmit EC, Beelen RH (1998) Functional and phenotypic differences of monocyte-derived dendritic cells from allergic and nonallergic patients. *J Allergy Clin Immunol* 101:90–95
182. Bellinghausen I, Brand U, Knop J, Saloga J (2000) Comparison of allergen-stimulated dendritic cells from atopic and nonatopic donors dissecting their effect on autologous naive and memory T helper cells of such donors. *J Allergy Clin Immunol* 105:988–996

183. Aiba S, Manome H, Yoshino Y, Tagami H (2003) Alteration in the production of IL-10 and IL-12 and aberrant expression of CD23, CD83 and CD86 by monocytes or monocyte-derived dendritic cells from atopic dermatitis patients. *Exp Dermatol* 112:86–95
184. Vulcano M, Albanesi C, Stoppacciaro A, Bagnati R, D'Amico G, Struyf S, Transidico P, Bonocchi R, Del Prete A, Allavena P, Rucio LP, Chiabrando C, Girolomoni G, Mantovani A, Sozzani S (2001) Dendritic cells as a major source of macrophage-derived chemokine/CCL22 in vitro and in vivo. *Eur J Immunol* 31:812–822
185. Reich K, Heine A, Hugo S, Blaschke V, Middel P, Kaser A, Tilg H, Blaschke S, Gutgesell C, Neumann C (2001) Engagement of the Fc epsilon RI stimulates the production of IL-16 in Langerhans cell-like dendritic cells. *J Immunol* 167:6321–6329
186. Higashi N, Gesser B, Kawana S, Thestrup-Pedersen K (2001) Expression of IL-18 mRNA and secretion of IL-18 are reduced in monocytes from patients with atopic dermatitis. *J Allergy Clin Immunol* 108:607–614
187. Habu Y, Seki S, Talayama E, Ohkawa T, Koike Y, Ami K, Majima T, Hiraide H (2001) The mechanism of a defective IFN-gamma response to bacterial toxins in an atopic dermatitis model, NC/Nga mice, and the therapeutic effect of IFN-gamma, IL-12, or IL-18 on dermatitis. *J Immunol* 166:5439–5447
188. Rengarajan J, Szabo SJ, Glimcher LH (2000) Transcriptional regulation of Th1/th2 polarization. *Immunol Today* 21:479–483
189. Yagi R, Nagai H, Iigo Y, Akimoto T, Arai T, Kubo M (2002) Development of atopic dermatitis-like skin lesions in STAT6-deficient NC/Nga mice. *J Immunol* 168:2020–2027
190. Finotto S, Neurath MF, Glickman JN, Qin S, Lehr HA, Green FH, Ackerman K, Haley K, Galle PR, Szabo SJ, Drazen JM, de-Sanctis GT, Glimcher LH (2002) Development of spontaneous airway changes consistent with human asthma in mice lacking T-bet. *Science* 295:336–338
191. Bluestone JA, Abbas AK (2003) Natural versus adaptive regulatory T cells. *Nat Rev Immunol* 3:253–257
192. Ling EM, Smith T, Nguyen XD, Pridgeon C, Dallman M, Arbery J, Carr VA, Robinson DS (2004) Relation of CD4+CD25+ regulatory T-cell suppression of allergen-driven T-cell activation to atopic status and expression of allergic disease. *Lancet* 363:608–615
193. Suto A, Nakajima H, Kagami SI, Suzuki K, Saito Y, Iwamoto I (2001) Role of CD4(+)CD25(+) regulatory T cells in T helper 2 cell-mediated allergic inflammation in the airways. *Am J Respir Crit Care Med* 164:680–687
194. Ou LS, Goleva E, Hall C, Leung DY (2004) T regulatory cells in atopic dermatitis and subversion of their activity by superantigens. *J Allergy Clin Immunol* 113:756–763
195. Akdis CA, Blaser K (2001) Role of IL-10 in allergen-specific immunotherapy and normal response to allergens. *Microbes Infect* 3:891–898
196. Laouini D, Alenius H, Bryce P, Oettgen H, Tsitsikov E, Geha RS (2003) IL-10 is critical for Th2 responses in a murine model of allergic dermatitis. *J Clin Invest* 112:1058–1066
197. Akdis CA, Akdis M (2003) Immunological differences between intrinsic and extrinsic types of atopic dermatitis. Editorial. *Clin Exp Allergy* 33:1618–1621
198. DiLandro A, Valsecchi R, Imberti G, Cainelli T (1993) Dermatite atopica e sensibilizzazione a nichel solfato. *G Ital Dermatol Venereol* 128:95–99
199. Lammintausta K, Kalimo K, Fagerlund VL (192) Patch test reactions in atopic patients. *Contact Dermatitis* 26:234–240
200. Pons-Guiraud A (1996) Intérêt de la batterie standard chez l'atopique. *Progrès en Dermato-Allergologie*. Vol II. Mediscript Editions, Paris, pp 43–53
201. Katoh N, Hirano S, Suehiro M, Ikenaga K, Yasuno H (2002) Increased levels of serum tissue inhibitor of metalloproteinase-1 but not metalloproteinase-3 in atopic dermatitis. *Clin Exp Immunol* 127:283–288
202. Kabesch M, Peters W, Carr D, Leupold W, Weiland SK, von Mutius E (2003) Association between polymorphisms in caspase recruitment domain containing protein 15 and allergy in two German populations. *J Allergy Clin Immunol* 111:813–817
203. Konishi H, Tsutsui H, Murakami T, Yumikura-Futatsugi S, Yamanaka K, Tanaka M, Iwakura Y, Suzuki N, Takeda K, Akira S, Nakanishi K, Mizutani H (2002) IL-18 contributes to the spontaneous development of atopic dermatitis-like inflammatory skin lesion independently of IgE/stat6 under specific pathogen-free conditions. *Proc Natl Acad Sci U S A* 99:11340–11345
204. Ahmad-Nejad P, Mrabet-Dahbi S, Breuer K, Klotz M, Werfel T, Herz U, Heeg K, Neumaier M, Renz H (2004) The toll-like receptor 2. R753Q polymorphism defines a subgroup of patients with atopic dermatitis having severe phenotype. *J Allergy Clin Immunol* 113:565–567
205. Hijnen D, De Bruin-Weller M, Oosting B, Lebre C, De Jong E, Bruijnzeel-Koomen C, Knol E (2004) Serum thymus and activation-regulated chemokine (TARC) and cutaneous T cell-attracting chemokine (CTACK) levels in allergic diseases: TARC and CTACK are disease-specific markers for atopic dermatitis. *J Allergy Clin Immunol* 113:334–340
206. Reich K, Hugo S, Middel P, Blaschke V, Heine A, Gutgesell C, Williams R, Neumann C (2002) Evidence for a role of Langerhans cell-derived IL-16 in atopic dermatitis. *J Allergy Clin Immunol* 109:681–687
207. Nomura I, Gao B, Boguniewicz M, Darst MA, Travers JB, Leung DY (2003) Distinct patterns of gene expression in the skin lesions of atopic dermatitis and psoriasis: a gene microarray analysis. *J Allergy Clin Immunol* 112:1195–1202
208. Hijnen D, De Bruin-Weller M, Oosting B, Lebre C, De Jong E, Bruijnzeel-Koomen C, Knol E (2004) Serum thymus and activation-regulated chemokine (TARC) and cutaneous T cell-attracting chemokine (CTACK) levels in allergic diseases: TARC and CTACK are disease-specific markers for atopic dermatitis. *J Allergy Clin Immunol* 113:334–340
209. Hon KL, Leung TF, Ma KC, Li AM, Wong Y, Fok TF (2004) Serum levels of cutaneous T-cell attracting chemokine (CTACK) as a laboratory marker of the severity of atopic dermatitis in children. *Clin Exp Dermatol* 29:293–296
210. Leung TF, Ma KC, Hon KL, Lam CW, Wan H, Li CY, Chan IH (2003) Serum concentration of macrophage-derived chemokine may be a useful inflammatory marker for

- assessing severity of atopic dermatitis in infants and young children. *Pediatr Allergy Immunol* 14:296–301
211. Frezzolini A, Paradisi M, Zaffiro A, Provini A, Cadoni S, Ruffelli M, De Pita O (2002) Circulating interleukin 16 (IL-16) in children with atopic/eczema dermatitis syndrome (AEDS): a novel serological marker of disease activity. *Allergy* 57:815–820
212. Yawalkar N, Uguccioni M, Schärer J et al (1999) Enhanced expression of eotaxin and CCR3 in atopic dermatitis. *J Invest Dermatol* 113:43–48
213. Toda M, Leung DY, Molet S, Boguniewicz M, Taha R, Christodoulopoulos P, Fukuda T, Elias JA, Hamid QA (2003) Polarized in vivo expression of IL-11 and IL-17 between acute and chronic skin lesions. *J Allergy Clin Immunol* 111:875–881
214. Arkwright PD, Chase JM, Babbage S, Pravica V, David TJ, Hutchinson IV (2001) Atopic dermatitis is associated with a low-producer transforming growth factor beta(1) cytokine genotype. *J Allergy Clin Immunol* 108:281–284
215. Groves RW, Ross EL, Barker JN, MacDonald DM (1993) Vascular cell adhesion molecule-1: expression in normal and diseased skin and regulation in vivo by interferon gamma. *J Am Acad Dermatol* 29:67–72
216. Griffiths CE, Railan D, Gallatin WM, Cooper KD (1995) The ICAM-3/LFA-1 interaction is critical for epidermal Langerhans cell alloantigen presentation to CD4+ T cells. *Br J Dermatol* 133:823–829
217. Jung K, Imhof BA, Linse R, Wollina U, Neumann C (1997) Adhesion molecules in atopic dermatitis: upregulation of alpha6 integrin expression in spontaneous lesional skin as well as in atopen, antigen and irritative induced patch test reactions. *Int Arch Allergy Immunol* 113:495–504
218. Jarvikallio A, Harvima IT, Naukkarinen A (2003) Mast cells, nerves and neuropeptides in atopic dermatitis and nummular eczema. *Arch Dermatol Res* 295:2–7
219. Toyoda M, Nakamura M, Makino T, Hino T, Kagoura M, Morohashi M (2002) Nerve growth factor and substance P are useful plasma markers of disease activity in atopic dermatitis. *Br J Dermatol* 147:71–79
220. Kim KH, Park KC, Chung JH, Choi HR (2003) The effect of substance P on peripheral blood mononuclear cells in patients with atopic dermatitis. *J Dermatol Sci* 32:115–124
221. Bos JD, Van Leent EJ, Sillevius Smitt JH (1998) The millennium criteria for the diagnosis of atopic dermatitis. *Exp Dermatol* 7:132–138
222. Williams HC, Burney PG, Hay RJ et al (1994) The U.K.'s Working Party criteria for atopic dermatitis. I. Derivation of a minimum set of discriminators for atopic dermatitis. *Br J Dermatol* 131:383–396