# **Rhizosphere Biology – an Overview** Chakravarthula Manoharachary, Krishna G. Mukerji

# 1.1 Introduction

Fungi, bacteria and actinomycetes are known to colonize diverse habitats and substrates and they are known to play substantial role in plant health and productivity besides producing diseases. The specialized ecological niches where the microbial association and their activity amply evidence are soil, rhizosphere, rhizoplane and phylloplane. These microbes may interact with the same plant simultaneously either independently, synergistically and/or antagonistically resulting sometimes in beneficial effect and at other times in harmful consequences. The microbes living in the complex region of rhizosphere influence the crop health and yield. The fungal biotechnologists have forgotten that soil, rhizosphere, rhizoplane and phyllosphere are the natural resources for microbial metabolites, products and other of biotechnological importance.

# 1.2 Soil Microbes

There is vast microbial flora inheriting the earth and they are found in all types of soils which are virgin or cultivated, sands, deserts, thermal soils or of volcanic origin, in bogs and moors, in snow covered soils, in sediments, semi aquatic ecosystems, on rocks and in rock crevices. The dominating groups of microorganisms are bacteria, actinomycetes, fungi, soil nematodes and protozoans. However, most of these organisms share a common character in being heterotrophic in their nutrition and thereby depend on other organic, dead or living organism and inorganic source of nutrition for their survival and multiplication. Most of the fungi, bacteria and actinomycete are microscopic and show vast variation quantitatively and qualitatively in different sites of collection and at different

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depths. Considerable variation occurs even between soil samples taken a few inches apart. Biotrophic, saprobic and symbiotic microbes are found in soil. The microbial population of soil is, therefore, dependent on composite micro-ecological environments. While most of the microorganisms live as saprophytes, others may be parasitic and symbiotic in relation to living organic substrates. Depending upon their enzymatic activity, they may be cellulolytic, keratinophilic, chitinophilic, pectinolytic, lignicolous or humicolous. The discovery of several parasitic fungi in causing root infection of several economically important crop plants paved way for intensive study of soil microorganisms in different niches-soil, rhizosphere or rhizoplane. Many of the parasitic fungi belong to genera *Pythium, Rhizoctonia* and *Fusarium*. Fusaria are known to cause root rots and wilts in young seedlings and some of them act as saprophytes on some dead plants tissues (Deacon 1996).

The discovery of several isolating techniques by several workers such as dilution plate techniques (Waksman 1911, 1944), soil plate method and modified soil plate methods (Warcup 1950), agar film method (Jones and Mollison 1948), immersion tube method (Chester 1948), immersion plate technique (Thornton 1952), baiting method (Harvey 1925), dilution frequency method (Allen 1957), direct microscopic examination (Conn 1981), uncoated glass slide (Rossi and Riccardo 1927; Cholodny 1930), root maceration (Stover and Waite 1953) and many other techniques for assaying of microbes resulted in their quantitative and qualitative estimations in different niches of the soil. Subsequently specific and nonspecific media were used selectively for isolation of bacteria, fungi and actinomycetes. Utilization of these isolating techniques and culture media contributed immensely in our present day understanding of soil, rhizosphere or rhizoplane microorganisms. The microorganisms in the soil are diverse and they differ quantitatively and qualitatively with the variations in the climatic, edaphic and biotic factors. The population dynamics of microflora of the soil depends on initial substrate and changes in substrate over period of time, nature and changes in other microflora and the genetic ability of a particular microorganisms to utilize or modify the components of the substrate and their interaction with biotic and abiotic components of the soil. Microorganisms in the soil are in constant state of flux and play an active role in recycling of organic wastes, nitrogen, phosphorus cycle and mineralization and other phenomenon and also produce several metabolites useful for humans. Bacteria from the soil were enumerated quantitatively by several workers. Agrobacterium tumefaciens was isolated by Schroth and Snyder (1961).

Soil fungi in relation to distinct fungal communities, physico-chemical factors, seasonal variations, biotic and other ecological parameters have been studied by several workers (Behera and Mukerji 1985a,b; Manohara-

chary 1977; Manoharachary and Rama Rao 1975; Mukerji 1966; Rama Rao 1970; Saksena 1967; Warcup 1967). Soil fungi in relation to microhabitat, interaction, physiology, biotic factors, role in decomposition of soil organic matter, antagonism, and role of dilution plate method were some of the facts reviewed extensively at International Symposium on ecology of soil fungi held at Liverpool (Parkinson and Waid 1960). Fungi isolated from different ecological niches including soils have been compiled in several publications by Barron (1968), Gilman (1957), Rai et al. (1961, 1963, 1964a,b,c, 1969), Rai and Mukerji (1962a,b).

The effect of genetically altered *Pseudomonas solanacearum* in soil and rhizosphere of tomato was studied by Hartel et al. (1990) and it was revealed in their studies that the genetically altered *Pseudomonas* slowed down the growth of unaltered *Pseudomonas* as a consequence of recombination. The biological control of soil-borne diseases of crops was achieved by treating soils with VAM fungi (Sharma and Adholeya 2000). It was observed that damping off caused by *Pythium* and *Rhizoctonia* on cucumber seedlings and bacterial wilt caused by *Pseudomonas solanaceraum* was controlled by charcoal compost and VAM combination but control was unsatisfactory with VAM fungi alone. Microorganisms isolated from the soil and the rhizosphere of wheat plants produced growth promoting substances whose effect was similar to those of gibberellins (Voker and Birnstiel 1989).

Pigmented actinomycetes were isolated from Cerrado soils of Brazil using starch agar nutrient medium (Copelhnas and Linhares 1993). The tests revealed that 90% of the pigmented actinomycetes were melanin producers. Melanin production was induced by supplementary inorganic or organic nitrogen source. These strains may have a role in soil organic matter formation. An actinomycetes strain – TB isolated from Korean soils – showed strong antimicrobial activity against Gram-positive bacteria and tumor cell growth inhibition (Jin-Ho et al. 1993).

Survey of actinomycetes from mud and water samples for a period of five years from 12 lakes in China was carried out by Jiang and Ku (1989) and they have found that the number and composition of actinomycetes was related to physical and chemical factors of the water body. Five new species *Micromonospora phaeovivda, Saccharomonospora yumnanensis, Microtetraspora flavorosea, Actinomadura viridoflava* and *Micropholyspora* were reported.

# 1.3 Rhizosphere Soil

The rhizosphere may be defined as that portion of the soil which is adjacent to the root system of a plant and is influenced by the root exudates. The area of this zone depends on the soil type and host plant under study and soil environment conditions. The roots exert influences on various type of microorganisms. The stimulatory effect on microorganisms is known as the "Rhizosphere effect" as indicated by the interaction of soil and rhizosphere microbes and their ratio. The chemical and physical nature of the root zone is quite different from the soil away from the root zone and the biology of this complex zone has been studied extensively. The term 'Rhizosphere' was proposed by Hiltner (1904). The phenomenon of accumulation of microorganisms around the root zone was reported by a number of earlier workers (Agnihothrudu 1955; Starkey 1958; Rouatt 1959; Parkinson 1957; Katznelson 1946; Sadasivan 1965; Jackson 1960; Mukerji and Ranga Rao 1968; Ranga Rao and Mukerji 1969, 1971a; Sorensen et al. 1997). Various compounds such as amino acids, vitamins, sugars, tannins etc. are exuded by the roots. These compounds have selective effect on microorganisms with in the root zone. Some root exudates are also known to affect certain microbial species adversely leading to their decrease in the root zone and, in return, microorganisms are known to exert profound influence on the plant itself by decomposition, affecting nutrient uptake, antagonistic effect on other microbes and parasitism.

Rhizosphere is a soil ecological region where soil is subjected to specific influence by plant root due to the exudates from root cells and sloughing of root tissue (Curl and Truelove 1986; Giddens and Todd 1984; Harley and Russell 1968). According to Pinton et al. (2001), rhizosphere represents a poorly defined zone of soil with a microbiological gradient in which maximum changes in the population of microflora in soil is evident adjacent to root and decline with distance away from it (Newman 1978; Bowen 1991; Mukerji 2002). The region of the external surface of plant root together with any closely adhering particles of soil or debris is differentiated from it and has been called rhizoplane. Root exudates stimulate microbial activity selectively in rhizosphere and rhizoplane regions (Bansal and Mukerji 1996). There is an intense competitive activity by the obligate saprobes, unspecialized root parasites and root inhabiting fungi depending on their behaviour towards exudates. In case of root diseases the pathogen has to react with the rhizosphere and rhizoplane fungi before entering the root tissues. These may show antagonism and check its advancement. The various factors and mechanism responsible for fungistasis, although already investigated, still need in-depth investigation. Plant microbe interaction is a regular and continuous feature of the biological world. The beneficial fallouts of such interactions have been extensively exploited for economic gain in recent years.

Once we develop a microbial technology for nif gene transfer and genes of mycorrhizal fungi and chitinase, genes offering resistance to plant pathogens, will definitely boost the crops production and pave the way for sustainable agriculture, making man's efforts for freedom from hunger a realizable end. This technology will reduce artificial fertilizer application and pesticide use.

Many have emphasized the importance of rhizosphere in plant-microbe interaction (Pinton et al. 2001). It has been documented by several researchers that microorganisms are stimulated in the rhizosphere (Katznelson 1946, 1965). Significant differences in the rhizosphere effect have also been reported. The metabolic state of the plant and the nature of soil appear to influence the extent of the rhizosphere effect (Rovira 1991). There is a need to learn more about the beneficial rhizosphere microorganisms. Ultimately the work on rhizosphere microbes has to aim at augmenting the biomass production (Yang-Ching et al. 2000).

Rhizosphere studies are fascinating and interesting leading to many beneficial consequences though some microbes have harmful effects. Some microbial metabolites like antibiotics and toxins, are crucial factors in determining plant-microbe relations. Microbial enzymes also play an effective role. The interesting relationship between plant root and microbes has attracted the attention of molecular biologists, microbiologists and biochemists throughout the world. Interestingly different types of microbes like fungi, bacteria, nematodes and viruses may interact with the same plant simultaneously either independently, synergistically or antagonistically.

Factors such as soil type, soil moisture, pH, temperature, plant age, relative humidity and several other factors are known to influence the rhizosphere effect. The rhizosphere effect is expressed in terms of R/S ratio (Katznelson 1946; Timonin 1966). The term 'rhizoplane' was proposed by Clark (1949) to refer to the immediate surface of plant roots together with any closely adhering particles of soil or debris. Using different isolation techniques microorganisms were isolated and identified by a number of workers.

In recent studies on rhizosphere, the emphasis was laid on interactions of microflora in the root zone due to effect of systemic or aerial sprays of herbicides or insecticides or fungicides on rhizosphere microflora (Ranga Rao et al. 1972). Data on microflora in Relation to various economically important crop plants or medicinal plants, studies on root diseases (Ranga Rao and Mukerji 1971b), soil environment and various factors of biotic, edaphic, environmental variations on the rhizosphere microflora and Dynamics Of Microorganisms In The Rhizosphere are meagre.

Mineralogical studies of rhizosphere of forest plants from six different sites in north and south of United States (Lynch and Whipps 1991) revealed that mineral grains within these zones are affected mechanically, chemically and mineralogically by invading root microorganisms than compare with non-rhizosphere forest soils. The mineral grains abutting root surfaces were significantly more fractured. This process of weathering in the root zone micro-environment should have a positive effect on soil complex formation that is biologically active.

Using soil biology methods on rhizosphere soils of six different plant species grown in four different soils and combinations of species were studied (Lynch 1982) to get information on organic metabolites and their origin in the rhizosphere. The amounts of sugar, amino acids and enzyme activity of some metabolites suggested a closer interrelationship with the involved organisms. Sometimes the nature of the soil was the dominating factor while it was host plant species which played a major role.

Recently plant growth promoting rhizobacteria (PGPR) were isolated from a number of economically important plants such as barley, bean, cotton, maize, groundnut, rice, various vegetables, wheat and wood species. In addition to increasing crop yield the strains of PGPR can also affect pathogenic fungi in reducing population densities.

With a higher nitrogen input a higher amount of <sup>14</sup>C was released from roots as photosynthates. This indicates that root exudates were significantly used by soil microbes in the rhizosphere. Using different nitrogen regimes, the rhizosphere bacteria of wheat was investigated (Keister and Cregan 1991). The bacterial population in relation to different nitrogen concentrations in the soil application resulted in higher amounts of bacterial populations in the rhizosphere but fluorescent pseudomonas population were low at higher nitrogen rates. The number of species of soil microorganisms, such as bacteria and actinomycetes were significantly correlated with cumulative temperature and rainfall in different agroclimatic zones. Similarly, higher correlations were obtained between rhizosphere microorganisms and pH, K, Ca and Mg. Acremonium, Alternaria, Cladosporium, Cephalosporium and Fusarium were the dominant fungal species.

Molecular cross talk seems to be the prerequisite mechanism for most root microbial infections. Just as the rhizodeposition can affect the composition of rhizosphere microflora, microbial metabolites can also affect the rhizosphere deposition. Rhizobacteria promote plant growth due to the production of plant growth regulators like auxin derivatives. The most studied molecular cross talk has been between rhizobia and the leguminous and non-leguminous host plant. The root exudates or chemical composition of rhizosphere solution can affect plant growth. It is very much the case that uptake of nutrients may be considerably influenced by the ionic concentration of the rhizosphere solution. However, we do not have the complete picture that takes into account the relative weight of each factor regarding molecular analysis of the interaction between plants, microbes and soil components.

#### 1.3.1 Rhizoplane

Rhizoplane is an ecological niche harbouring a number of bacteria, actinomycetes and fungi. The term rhizoplane was proposed by Clark (1949) to refer to the immediate external surface of plant roots together with any closely adhering particles of soil or debris and microbial communities. It was proposed by Katznelson (1946) that the proportion of microorganisms in the rhizosphere with the population found outside the rhizosphere in numerical terms may be expressed as an R/S ratio. Using serial root washings, Harley and Waid (1955) successfully isolated rhizoplane microorganisms. Stover and Waite (1953) and Singh (1965) using a root maceration technique isolated Fusaria and other root mycoflora. Using baiting technique several pathogenic fungi like *Phytophthora* (Eckert and Tsao 1962) and *Pythium* (Drechsler 1929; Johnson and Curl 1972) have been isolated from infected plant roots and the soil around them.

#### 1.3.2 Root Exudates

Root exudates are the substances released by roots and may affect growth and activity of soil microorganisms in rhizosphere and on rhizoplane (Bansal and Mukerji 1994, 1996; Kapoor 1999; Schottendreier and Falkengren-Greup 1999). Several substances are reported as released by roots in the form of exudates such as amino acids, sugars, vitamins, organic acids, nucleotides, flavones, enzymes, hydrocyanic acid, glycosides, auxins and saponins (Gupta and Mukerji 2002). Root exudates are known to influence the proliferation and survival of root infecting pathogens. Spores or other propagules of many pathogenic fungi such as Rhizoctonia, Fusarium, Sclerotium, Aphanomyces, Pythium, Colletotrichum, Verticillium and Phytophthora are shown to germinate as a result of stimulation and or food sources provided by root exudates of susceptible cultivars of host plants (Bhuvaneshwari and Subba Rao 1957; Vancura 1964). Root exudates also provide food base for antagonists which in turn suppress the growth of pathogenic microorganisms in soil. The resistant varieties are known to harbour a higher number of Streptomyces and Trichoderma which in turn may show antagonism against pathogenic fungi, thus rendering host plant resistant (Katznelson and Rouatt 1957; Starkey 1958; Subba Rao and Bailey 1961; Walker 1975).

Root exudates containing toxic substances such as glycosides and hydrocyanic acid may inhibit the growth of pathogens (Rangaswami 1988). It is also possible that the root exudates alter soil pH in rhizosphere and thus alter the micro-climate or the rhizosphere which possibly reflected in changes and growth of microorganisms.

Age of the plant, temperature and calcium in relation to root exudation and its rhizosphere effect was studied by Vancura (1964). Schroth and Snyder (1961) observed trace amounts of nitrogenous substances in addition to sugar in the root exudates of bean plants.

The compounds released by plant root are diverse and only a few of them have direct effect on the growth of soil-grown plants. Furthermore very few root secretion and root products can be expected to be effective unless the right set of circumstances exist around the root.

#### 1.3.3 Rhizosphere and Compounds Released by Roots

A vast array of compounds gets released into the rhizosphere by plant roots. However few have a direct effect on the growth of soil grown plant root products including organic compounds and these have a direct effect on the growth of the plant. Roots are not only organs for nutrients uptake, but are able to release a wide range of organic and inorganic compounds into the root environment (Giddens and Todd 1984). Soil-chemical changes related to the presence of these compounds and products of their microbial turnover are important factors affecting microbial populations, availability of nutrients, solubility of toxic elements in the rhizosphere and thereby the ability of plants to cope with adverse conditions (organic rhizodepositions – yeast).

In the last two decades due attention was given to functional characterization of plant root exudates. In order to understand the complex interactions at the root-soil interface a multidisciplinary approach is necessary. A number of investigators have suggested that plants treated with humic molecules differ in growth and morphology from control plants. Microbes associated with plant roots are related to plants growth regulators using the precursors released by the roots. Though much work has been done in the stimulation of microbial populations antagonistic to pathogens, manipulation of rhizosphere populations using plant breeding programs has not been seriously pursued. Further, the degradation of pollutants by the rhizosphere microorganisms will open greater opportunities to decontaminate land more effectively. The plant-microbe performance in the rhizosphere is an important aspect. In this regard mineralization and immobilization are the processes which will help in the evaluation. The increasing use of transgenic plants and the release into the environment of genetically modified microorganisms could change the plant-microbe interactions and microbial potential. Four groups of signal organic molecules namely flavonoids, antibiotics, lipochitooligosaccharides and vitamins have different functions in plant growth promotions, plant defence and plant symbiosis.

Siderophores are iron chelating agents that are secreted by microorganisms in response to iron deficiency. Siderophores have been studied for their importance in plant disease suppression. Recently some studies have been taken up in examining the role of siderophore and phytosiderophores in facilitating heavy metal uptake and food chain transfer of metals. In spite of having wealth of information, there is still considerable debate as to how they function in the rhizosphere and the degree to which they accumulate in soils.

Rhizosphere is considered as a dynamic environment where microbial balance is maintained in spite of having constellation of soil physicochemical factors. Mycorrhizae, an important component of the rhizosphere, form a mutual plant-fungus association which is an essential feature of biology and ecology of most terrestrial plants, since it influences their growth as well as their water and nutrient absorption and protects them from root diseases. Mycorrhizosphere has still to be understood in detail. Loss of carbon compounds from roots is the driving force for the development of enhanced microbial populations in the rhizosphere in comparison with the bulk soil. There is a need to learn more about carbon flow and microbial communities in natural environments.

The organic substrates released in rhizosphere soil, including plant and animal debris, has the potential to determine the high diversity of the soil microbial community inhabiting the root zone. Physico-chemical factors among rhizosphere soils of different plant species at different developmental stages of plants may dispose rhizosphere microbial communities toward preferential C-metabolite pathways which will impact differently on mineralization or immobilization of nutrients. Productivity of an ecosystem and its performance are dependent on mineralization and immobilization. Estimates of the diversity of DNA-based soil organisms so far have not been able to answer the questions concerning the functions of the soil ecosystem, although they can serve as an efficient tool for explaining DNA diversity in situ. The presence of a certain microbial gene does not assure that the metabolic process for which the gene is responsible is carried out. The increasing utility of transgenic plants and also the release of genetically modified microbes may affect plant-microbe interactions and microbial potential.

Flavonoids are identified as very specific molecules in the rhizobia/legume host plant communication. By activating specific nod genes, a new type of signal molecule, nod factors are produced by the bacteria, triggering host plant reactions such as root hair curling and meristem induction. Antibiotic substances are also released into the rhizosphere ecosystem. Nitrogen fixation is the result of co-evolution of atmosphere and life; one has to understand the molecular aspects of rhizobia in relation to symbiotic relationship and nitrogen fixation in symbiotic and non-symbiotic plants.

The important function of plant and microbial siderophores in the rhizosphere is for iron acquisition under iron-limiting conditions. Microbes produce siderophores and compete for iron. Plants and microbes also compete for iron. Siderophores also suppress soil-borne and root-borne plant pathogens, besides employing them for phytoremediation.

Mycorrhizae are the rhizosphere soil components and form symbiotic interaction between plant and fungal genomes. Molecular events related to mycorrhization are not understood clearly. Mycorrhizal fungi act as biofertilizer to be used for sustainable agriculture and forestry. One of the interesting fields in microbial ecology is interaction between mycorrhizal fungi and rhizosphere microorganisms.

The rhizosphere microbial community is different from the bulk soil community and differs between plant species. Mathematical models do generate quantitative answers and allow the integration of many individual plants and soil processes to predict a single outcome. There is a requirement for such models to work on rhizosphere ecosystems. Loss of carbon compounds from roots or rhizodeposition is the driving force for the development of enhanced microbial populations in the rhizosphere in comparison with the bulk soil – use of <sup>13</sup>C techniques for measuring carbon flow is the best example besides use of biochemical or molecular signatures.

# 1.4 Decomposition of Organic/Inorganic Compounds in Rhizosphere

Plants and trees interact with soil to produce an area around the roots called the rhizosphere. This is an area of high microbial activity, lower pH, increased organic matter, higher carbon dioxide and lower water and nutrient content. Particularly because of increased microbial activity this area also contributes a unique environment, conducive to the breakdown of organic compounds. Because plants are constantly taking up water the rhizosphere is an area where concentration of metals and organic compounds occurs. Once in the rhizosphere, inorganic and organic compounds, including metals, may be deposited, taken up or broken down by plants.

Active degradation of contaminants involves enzymes, or metabolic pathways, which accommodate the pollutant. The plant or microorganism may use the pollutant as a source of nutrients, carbon, nitrogen, phosphorus etc., as a source of energy or both. Common short-lived insecticides and herbicides are broken down and used in such a manner. Nitrates can be removed from water under anaerobic conditions by a process called denitrification. During this process nitrate is used as an electron acceptor and thus the organism is using the nitrate in energy production.

Passive removal could be by uptake along with water and release into the atmosphere by transpiration. Gases dissolved in water may be taken up by plants, transported up the plant system and released into the atmosphere during transpiration. On the other hand one might simply wish to have trees remove water from soil to prevent it and associated pollutants from moving into or out of the area.

Passive removal can also take place in the rhizosphere. This could be by concentration or by concentration and precipitation. If a plant can be found which takes up the contaminant, then the plant can be grown and the pollutant "harvested" along with the plant. If the pollutant is a metal the plant material can be burned and the metal recovered. If the pollutant is a heavy metal and is precipitated in the rhizosphere then the plant, roots and surrounding soils may need to be "harvested".

Bioremediation using any type of organism is a valuable remediation tool. It has many advantages including the fact that it is often carried out in situ and thus involves minimal disturbance to the site. In addition microorganisms and plant roots can explore every recess of soil and thus carry out a more complete remediation.

# 1.5 Root Sensing Signals

Recent advances show that organic compounds present in the rhizosphere can have a specific role in plant-microbe-soil interactions. Signal molecules exchanged between plants and microorganisms have been identified that favour beneficial plant colonization (Giovannetti et al. 1996). Some compounds present in soils, e.g. humic molecules and nutrients, metabolise through stimulation or inhibition of biochemical reactions or processes of root cells and triggering of specific signal transduction pathways. However, molecular information on how the plant senses the environmental conditions of the rhizosphere is still lacking.

# 1.6 Conclusions

The rhizosphere is the zone of soil adjacent to and influenced by roots. The surface of the root is known as the rhizoplane and the endorhizosphere refers to the root cortex, in the context of its colonisation by bacteria. The rhizosphere is colonised by bacteria, fungi and other microorganisms and thus constitutes a living interface between roots and the soil. The rhizosphere, rhizoplane and endorhizosphere form a continuum that is difficult to distinguish in terms of microbial ecology. The functional and spatial complexity of the rhizosphere is heightened by mycorrhizal fungi, which in turn support a mycorrhizosphere that is also colonised by bacteria. The rhizosphere generally refers to the zone of soil surrounding the roots with higher microbial activity than in soil remote from roots.

### References

- Agnihothrudu V (1955) State in which fungi occur in the rhizosphere. Naturwissenschefen 42:515–516
- Allen LE (1957) Experiments in soil bacteriology. Burgess, Minneapolis, Minn
- Bansal M, Mukerji KG (1994) Positive correlation between VAM induced changes in root exudation and mycorrhizosphere mycoflora. Mycorrhiza 5:39–44
- Bansal M, Mukerji KG (1996) Root exudation in rhizosphere biology. In: Mukerji KG, Singh VP, Suvercha (eds) Concepts in applied microbiology and biotechnology. Aditya Books, New Delhi, pp 97–119
- Barron GO (1968) The genera of Hyphomycetes from soil. The Williams and Wilkins Company, Baltimore
- Behera N, Mukerji KG (1985a) Seasonal variation and distribution of microfungi in forest soils of Delhi. Folia Geobot Phytotaxon 20:291–311
- Behera N, Mukerji KG (1985b) Ecology of micromycetes in forest soils of Delhi. Acta Mycol 21:101–108
- Bhuvaneshwari K, Subba Rao NS (1957) Root exudates in relation to rhizosphere effect. Proc Indian Acad Sci 45B:299–301
- Bowen GD (1991) Microbial dynamis in the rhizosphere: possible strategies in managing rhizosphere population. In: Keister DL, Cregan B (eds) The rhizosphere and plant growth. Kluwer Academic Publishers, Netherlands, pp 25–32
- Chester CGC (1948) A contribution to the study of fungi in the soil. Trans Brit Mycol Soc 30:100–117
- Cholodny N (1930) Uber eine neue methode zur untersuchung der Boden microflora. Arch Microb 1:620–652
- Clark FE (1949) Soil microorganisms and plant roots. Adv Agron 1:241-288
- Conn HJ (1981) The microscopic study of bacteria and fungi in the soil. NY Agric Exp St Tech Bull 64
- Copelhnas RRR, Linhares LE (1993) Melanogenic actinomycetes (*Streptomyces* sp.) from Brazilion soils. Biol Fertil Soils 15:220–224
- Curl EA, Truelove B (1986) The rhizosphere. Springer, Berlin Heidelberg New York
- Deacon JA (1996) Ecological implications of recognition events in the pre-infection stages of root pathogens. New Phytol 133:135–145
- Drechsler C (1929) The best water mold and several related root parasites. J Agr Res 38: 309–392
- Eckert JW, Tsoa PH (1962) A slective antibiotic medium for isolation of *Phytophthora* and *Pythium* from plant roots. Phytopathology 52:771–777
- Giddens JE, Todd RL (1984) Rhizosphere microorganisms overview. In : Todd RL, Giddens JE (eds) Microbial-plant interactions. Proc Soil Sci Soc Am, Madison, pp 51–68
- Gilman JC (1957) A manual of soil fungi. Iowa Sate University Press, Ames, Iowa

- Giovannetti M, Sbrana C, Citernesi AS, Avio L (1996) Analysis of factors involved in fungal recognition responses to host-desired signals by arbuscular mycorrhizal fungi. New Phytol 133:65–71
- Gupta R, Mukerj KG (2002) Root exudate biology. In: Mukerji KG et al. (eds) Techniques in Mycorrhizal studies. Kluwer Academic Publishers, Netherlands, pp 103–131
- Harley JL, Russell RS (1968) The soil-root interface. Academic Press, London
- Harley HL, Waid JS (1955) A method of studying active mycelia on living roots and other surfaces in the soil. Trans Br Mycol Soc 38:104–118
- Hartel PG, Williams JW, Schnell MA (1990) Growth of genetically altered *Pseudomonas* solanacearum in soil and rhizosphere. Soil Sci Soc Am J 54:1021–1025
- Harvey TV (1925) A survey of the water molds and Pythiums occurring in the soils of Chapel Hill. J Eisha Mitchell Sci Soc 41:151–164
- Hiltner L (1904) Über neuere Erfahrungen und Probleme auf dem Gebiet der Bodenbakteriologie und unter besonderer Berücksichtigung der Gründüngung und Brache. Arb Dtsch Landwirt Ges 98:59–78
- Jackson TM (1960) Soil fungistasis and rhizosphere. In: Parkinson D, Waid JS (eds) The ecology of soil fungi. Liverpool Univ Press, Liverpool, pp 168–176
- Jiang C, Ku L (1989) Studies on actinomycetes flora and resources in the plateau lakes in Yunnan (China). Acta Microbiol Sin 29:7–14
- Jin-Ho H, Kim S-C, Chang Y, Konsyeom (1993) Antimicrobial activity and tuner cell growth inhibition of on actinomycetes isolated from Korean soil. Yakhak Hoeji 37:389–396
- Johnson LF, Curl EA (1972) Methods for research on the ecology of soil-borne plant pathogens. Burgess Publ, Minneapolis, Minnesota
- Jones PCT, Mollison JE (1948) A technique for quantitative estimation of soil microorganisms. J Gen Microbiol 2:54–69
- Kapoor R (1999) Root exudation and its implication or rhizospehre mycoflora. In: Tewari JP, Lakhanpal TN, Singh J, Gupta R, Chamola BP (eds) Advances in microbial biotechnology. APH Publ Corp, New Delhi, pp 351–362
- Katznelson H (1946) The "Rhizosphere effect" of mangles on certain groups of soil microorganisms. Soil Sci 62:343–354
- Katznelson H (1965) Nature and importance of the rhizosphere. In: Baker KF, Snyder WC (eds) Ecology of soil-borne plant pathogens. John Murray Press
- Katznelson H , Rouatt JW (1957) Studies on the microflora of the rhizosphere. Trans 6th Int Cong Soil Sci Paris, pp 151–156
- Keister DL, Cregan PB (1991) Rhizosphere and plant growth. Kluwer Academic Publishers, Boston
- Lynch JM (1982) The rhizosphere. In: Burns RG, Slatter JH (eds) Experimental microbial ecology. Blackwell Sci Publ, Oxford, pp 295–411
- Lynch JM, Whipps JM (1991) Substrate flow in the rhizosphere. In: Keister DL, Cregan PB (eds) The rhizosphere and plant growth. Kluwer Academic, Dordrecht, pp 15-24
- Manoharachary C (1977) Microbial ecology of scrub jungle and dry wasteland soils from Hyderabad District, Andhra Pradesh (India). Proc Indian Nat Sci Acad 43B:6–18
- Manoharachary C, Rama Rao P (1975) Two new additions to the Indian soil fungi. Indian Phytopathol 28:427–428
- Mukerji KG (1966) Ecological studies on the microorganic population of usar soils. Mycopath Mycol Appl 29:339–349
- Mukerji KG (2002) Rhizosphere biology. In: Mukerji KG et al. (eds) Techniques in Mycorrhizal studies. Kluwer Academic Publishers, Netherlands, pp 87–101
- Newman EI (1978) Root microorganisms: their significance in the ecosystem. Biol Rev 53:511-554

- Parkinson D (1957) New methods for qualitative and quantitative study of fungi in the rhizosphere. Pedologi Gand 7:146–154
- Parkinson D, Waid JS (1960) The ecology of soil fungi. Liverpool Univ Press, Liverpool
- Pinton R, Varanini Z, Nannipieri P (2001) The rhizosphere-biochemistry and organic subtances at the soil-plant interface. Marcel Dekker. New York Basel
- Rai JN, Mukerji KG (1962a) A new species of *Chaetomium* from Indian soils. Can J Bot 40:857-860
- Rai JN, Mukerji KG (1962b) *Sprotrichum carthusioviride* Rai and Mukerji, a new species from Indian soils. Mycopath Mycol Appl 18:122–126
- Rai JN, Mukerji KG, Tewari JP (1961) A new *Helicostylum* from Indian soils. Can J Bot 39:1282–1284
- Rai JN, Mukerji KG, Tewari JP (1963) *Tripterospora tetraspora* sp. nov., a new cleistothecial Ascomycetes. Can J Bot 41:327–329
- Rai JN, Tewari JP, Mukerji KG (1964a) *Achaetomium*, a new genus of Asconycetes. Can J Bot 42:693–697
- Rai JN, Tewari JP, Mukerji KG (1964b) A new *Aspergillus* from Indian soils *A. striatus* sp. nov. Can J Bot 42:1521–1524
- Rai JN, Tewari JP, Mukerji KG (1964c) Cultural and texonomic studies on two rare species of Aspergillus – A. paradoxus and an interesting strain of A. variecolor from Indian soils. Mycopath Mycol Appl 26:369–376
- Rai JN, Tewari JP, Mukerji KG (1969) Mycoflora of mangrove mud. Mycopath Mycol Appl 38:17–31
- Rama Rao P (1970) Studies on soil fungi III. Seasonal variation and distribution of microfungi in some soils of Andhra Pradesh (India). Mycopath Mycol Appl 40:277–298
- Ranga Rao V, Mukerji KG (1969) Fungi of Delhi. IX. Additions to our knowledge of soil fungi. J Ind Bot Soc 48:258–261
- Ranga Rao V, Mukerji KG (1971a) Fungi in the root zone of four cultivars of wheat. Ann Inst Pasteur 121:533–544
- Ranga Rao V, Mukerji KG (1971b) Studies on charcoal rot disease of *Abelmoschus esculentus*. II. Fungal flora of the root zone of healthy and infected plants. Ann Inst Pasteur 122:81–90
- Ranga Rao V, Jayakar M, Sharma KR, Mukerji KG (1972) Effect of foliar spray of Morphactin on fungi in the root zone of *Capsicum annum*. Plant Soil 37:179–182
- Rangaswami G (1988) Soil-plant microbe interrelationships. Indian Phytopath 41:165-172
- Rossi G, Ricardo S (1927) L'seae microscopico ebacteriologico iretto del ferreno agrario. Nuovi Ann Minist Agric 7:457–470
- Rouatt JW (1959) Initiation of Rhizosphere effect. Can J Microbiol 5:67-71
- Rovira AD (1991) Rhizosphere research 85 years of progress and frustration. In: Keister DL, Cregan PB (eds) Rhizosphere and plant growth. Kluwer Academic Publishers, Netherlands, pp 3–13
- Sadasivan TS (1965) Root and its environment. J Sci Ind Res 24:11-113
- Saksena RK (1967) The soil fungi a biological appraisal. Indian Phytopath 24:13-25
- Schottendreier M, Falkengren-Greup U (1999) Plant induced alteration in the rhizosphere and the utilization of soil heterogenicity. Plant Soil 209:297–309
- Schroth MN, Snyder WC (1961) Effect of host exudation, chalamydospore germination of the bean root rot fungus *F. solani, F. phaseoli.* Phytopathology 51:389–393
- Sharma MP, Adholeya A (2000) Sustainable management of Arbuscular Mycorrhizal fungi in the biocontrol of soil-borne plant disease. In: Upadhyay RK, Mukerji KG, Chamola BP (eds) Biocontrol potential and its exploitation in sustainable agriculture. Kluwer Academic/Plenum Publishers, Dordrect, New York, pp 117–138
- Singh KG (1965) Comparison of techniques for the isolation of root infecting fungi. Nature 206:1169–1170

- Sorensen J, van Elsas JD, Trevors JT (1997) The rhizosphere as a habitat for soil microorganisms. In: Wellington EMH (ed) Modern soil microbiology. Marcel Dekker, New York, pp 21–45
- Starkey RL (1958) Inter relation between microorganisms and plant roots in rhizosphere. Bot Rev 22:154-172
- Stover RH, Waite BH (1953) An improved method of isolating *Fusarium* sp. from plant tissues. Phytopathology 43:700-701
- Subba Rao NS, Bailey DL (1961) Rhizosphere studies in relation to varietal resistance or susceptibility of tomato to *Verticillium* wilt. Can J Bot 39:1747–1758
- Thornton RH (1952) The screened immersion plate. A method of isolating soil microorganisms. Research 5:190–191
- Timonin MI (1966) Rhizosphere effect of healthy and diseased lodgepole pine seedlings. Can J Microbiol 12:531–537
- Vancura V (1964) Root exudates of plants. I. Analysis of root exudates of barley and wheat in their initial phases of growth. Plant Soil 21:231–248
- Voker L, Birnstiel H (1989) Plant growth substances produced by microorganisms of the rhizosphere and the soil. J Basic Microbiol 29:473–476
- Waksman SA (1911) Do fungi live and produce mycelium in the soil? Soil Sci NS 44:320–322 Waksman SA (1944) Three decades with soil fungi. Soil Sci 58:89–114
- Walker N (1975) Soil microbiology a critical review. Butterworths, London
- Warcup JH (1950) The soil plate method for isolation of soil fungi. Nature, London, 166:117–118
- Warcup JH (1967) Fungi in soil. In: Burgess A and Raw F (eds) Soil Biology Academic Press, London, pp 51–110
- Yang-Ching H, Crowley DE, Yang CH (2000) Rhizosphere microbial community structure in relation to root location and Plant iron nutritional status. Appl Environ Microbiol 66:345–351