

Electronic Excitation Processes in Single-Strand and Double-Strand DNA: A Computational Approach

Felix Plasser, Adélia J.A. Aquino, Hans Lischka, and Dana Nachtigallová

Abstract Absorption of UV light by nucleic acids can lead to damaging photoreactions, which may ultimately lead to mutations of the genetic code. The complexity of the photodynamical behavior of nucleobases in the DNA double-helix provides a great challenge to both experimental and computational chemists studying these processes. Starting from the initially excited states, the main question regards the understanding of the subsequent relaxation processes, which can either utilize monomer-like deactivation pathways or lead to excitonic or charge transfer species with new relaxation dynamics. After a review of photophysical processes in single nucleobases we outline the theoretical background relevant for interacting chromophores and assess a large variety of computational approaches relevant for the understanding of the nature and dynamics of excited states of DNA. The discussion continues with the analysis of calculations on excitonic and charge transfer states followed by the presentation of the dynamics of excited-state processes in DNA. The review is concluded by topics on proton transfer in DNA and photochemical dimer formation of nucleobases.

F. Plasser

Interdisciplinary Center for Scientific Computing, Ruprecht-Karls-University, Im Neuenheimer Feld 368, 69120, Heidelberg, Germany

A.J.A. Aquino and H. Lischka (✉)

Department of Chemistry and Biochemistry, Texas Tech University, Lubbock, TX 79409-1061, USA

Institute for Theoretical Chemistry, University of Vienna, Währingerstr. 17, 1090 Vienna, Austria

e-mail: hans.lischka@univie.ac.at

D. Nachtigallová

Institute of Organic Chemistry and Biochemistry AS CR, Flemingovo nam. 2, Prague, Czech Republic

e-mail: dana.nachtigallova@uochb.cas.cz

Keywords Ab initio calculations · Charge transfer excited states · Excitonic states · Interaction of excited state nucleic acid bases · Photodynamics · UV absorption spectra

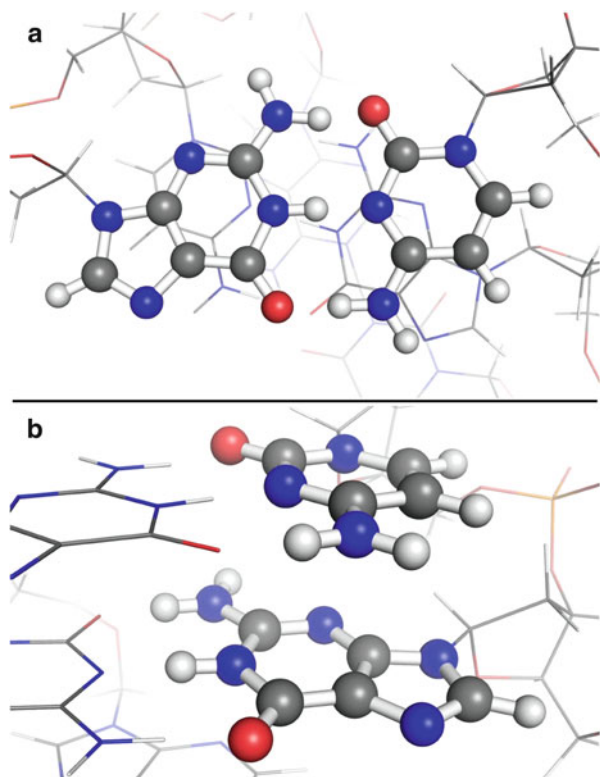
Contents

1	Introduction	2
1.1	Ultrafast Deactivation of Single Nucleobases	4
1.2	Survey of Experimental Studies of Interacting Nucleobases	8
2	Description of Electronic Coupling	10
2.1	Excimers/Exciplexes and Excitons	12
3	Assessment of Computational Methods	14
3.1	Electronic Structure Methods	15
3.2	Environmental Models, Sampling, and Dynamics	17
4	UV Absorption	19
4.1	Excitonic Delocalization	19
4.2	Charge Transfer States	21
5	Excited State Processes	23
5.1	Sterical Hindrances and Electrostatic Interactions	23
5.2	Excimer Formation and Excitation Energy Transfer	24
5.3	Proton Transfer Processes	27
5.4	Photochemical Processes	29
6	Conclusions	31
	References	31

1 Introduction

Nucleic acids play a central role in biology as carriers of the genetic code. All nucleic acid bases are strongly absorbing species in the ultraviolet (UV) region and their excitation can result in production of harmful photoproducts leading to, e.g., mutations [1–3]. A well known example is the dimerization of pyrimidine type bases, i.e., formation of thymine (T<>T), cytosine (C<>C), and thymine–cytosine (C<>T) dimers [4–7]. This process is formally analogous to the dimerization of two ethylene molecules to cyclobutane, a textbook example of a reaction which is prohibited in the ground state but allowed in the excited state according to the Woodward–Hoffmann rules. Fortunately, the formation of such photoproducts occurs very rarely. The nucleic acid bases themselves show a high degree of photostability (see, e.g., [8–11]). The decay mechanisms of isolated nucleobases have been investigated by ultrafast time resolved spectroscopic techniques [11–15] and extended computational methods [16–28]. Most of the general features and many details are largely understood by now. It is well established [11, 29] that ultrafast (on the time scale of a few picoseconds) internal conversion is responsible for the relaxation of the system into the ground state without changing its chemical identity. This process occurs at structures near the crossing seam of excited-state and ground-state potential energy surfaces (PES) under conditions of a strong

Fig. 1 Structure of Gua and Cyt allowing for inter-strand hydrogen bonding (a) and intra-strand stacking (b) interactions in the DNA double-helix

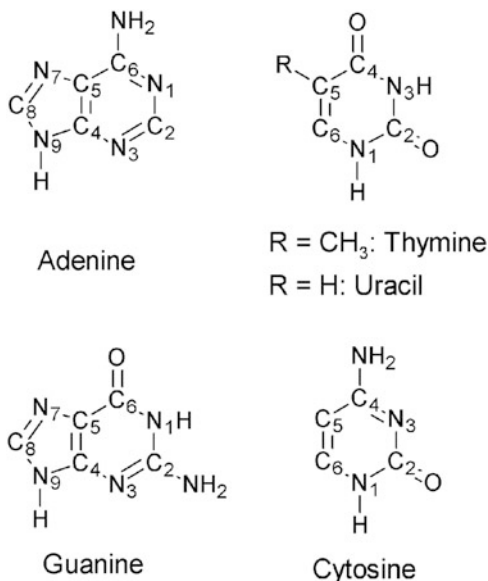


nonadiabatic coupling. Due to this mechanism nucleobases are photostable which in turn protects them against radiative damage.

The situation becomes much more complex when a nucleobase is interacting with other bases within the nucleic acid polymer. The structure of nucleic acids allows for interaction within the same strand via stacking and, in the case of double-stranded DNA, also for interactions between two strands via hydrogen bonding as illustrated in Fig. 1. These interactions are further affected by the conformation of DNA (e.g., differences can be expected in B-DNA and RNA-like A-DNA conformations) and the flexibility of the sugar–phosphate backbone which changes the mutual orientation of adjacent nucleobases and thus their interaction. This complex picture of photodynamics of nucleic acids provides a great challenge to both experimental and computational chemists.

In the present contribution the current knowledge of the photodynamics of isolated nucleobases and the survey of experimental observations and their interpretations will be provided in Sect. 1. Only a brief discussion will be given since the scope of the paper is laid on the computational approaches to the UV absorption characteristics and photodynamics of nucleic acids. The theoretical concept of electronic coupling is introduced in Sect. 2. Computational methods and their reliability for the description of excited states in DNA are discussed in Sect. 3. Section 4 discusses the interpretation of the experimentally observed absorption

Scheme 1 The numbering scheme of nucleic acid bases



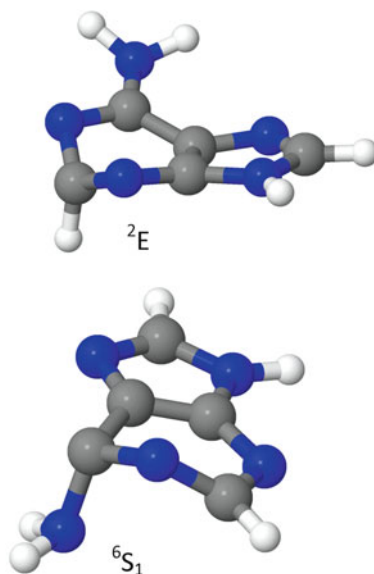
spectra of DNA and its model systems. In Sect. 5 state-of-the-art computational studies on excited states processes, including the effect of DNA environment on the photophysics of nucleobases, excimer formation, excitation energy and proton transfer, and photochemical reactions are reported. The discussion on the latter is limited to formation of cyclobutane pyrimidine dimers as the most studied process.

1.1 Ultrafast Deactivation of Single Nucleobases

The understanding of the photodeactivation mechanism of individual nucleobases is important for analysis of the more complex deactivation patterns of nucleobases embedded in DNA or RNA. It is interesting to note that all five nucleobases (adenine, guanine, thymine, cytosine, and uracil; Scheme 1) show the property of photostability due to their ultrafast, radiationless deactivation to the ground state within a few picoseconds [8, 10]. This decay provides the possibility for the nucleobase to transfer quickly the harmful excess energy accumulated by photoabsorption of UV light into heat, which further on can be dissipated to the environment. These ultrafast processes observed for all five naturally occurring nucleobases contrast with the much longer lifetimes for nucleobase analogues not found in DNA or RNA [10, 30], indicating an evolutionary pressure in early biotic ages.

The photodeactivation of nucleobases in the gas phase has been studied in detail by means of ultrafast time-resolved spectroscopy [8, 10, 11, 17]. These investigations provide the main source for relaxation times which are obtained by fitting the measured deactivation curves in terms of pump-probe delay times. Information on

Fig. 2 Adenine structures at the conical intersections 2E and 6S_1



the structural changes responsible for the photostability are not or barely available from these data. The ultrafast and radiationless character of these processes points to internal conversion governed by conical intersections located on the crossing seam between different energy surfaces. This hypothesis has been substantiated in great detail by many theoretical investigations. Starting from the Franck–Condon (FC) region, deactivation paths have been computed leading to characteristic crossing points which are usually chosen as the minima on the intersection seam. One of the nucleobases studied extensively is adenine for which a large variety of types of intersections has been found [16, 22, 23, 31–34]. The two energetically lowest S_0 – S_1 intersections show puckering at the C_2 atom and out-of-plane distortion of the NH_2 group, respectively (Fig. 2). The former structure corresponds to an S_0/π – π^* intersection characterized as envelope 2E and the latter to an S_0/n – π^* intersection with a skew boat 6S_1 puckering following the notation of ring puckering coordinates introduced by Cremer and Pople [35]. Starting at the Franck–Condon structure, pathways connecting the bright L_a state with these intersections are supposed to have no or at most very small barriers so that both are good candidates for the explanation of the ultrafast decay of adenine. For a more detailed discussion of the outcomes of different dynamics simulations see below. In the case of the other purine base guanine, two ethylenic-type intersections (I and II) with strong out-of-plane distortion of the NH_2 group and another intersection type (denoted oop-O) with puckering at C_6 and concomitant out-of-plane motion of the oxygen atom are found [20, 28, 36–39]. The bright π – π^* state is the lowest singlet excited state in the Franck–Condon region and is connected from there via a barrierless path with the ethylenic intersections [28].

In comparison to the purine bases, the pyrimidine bases are characterized by a more complex system of conical intersections and reaction paths. In the case of cytosine, three major conical intersections have been determined. The first [25] is characterized as semi-planar. It occurs in a region of mixing of the $n_O-\pi^*$, $\pi-\pi^*$ and closed shell states, which leads to a triple degeneracy as reported in [40, 41]. The second intersection leads to a puckering at N₃ connected with an out-of-plane distortion of the amino group. The third intersection involves puckering at C₆ (for more details see [42–44]). Reaction paths computed in [44] connecting the Franck–Condon region with the above-mentioned conical intersections show that these intersections are all energetically accessible and display similar qualitative features so that no a priori decision can be made concerning specific photodynamical reaction mechanisms.

Concerning theoretical investigations on the remaining nucleobases (see, e.g., [17, 21, 24, 27]), it should be mentioned at this point that not only the ring puckering motions are of relevance for the explanation of the ultrafast decay of the nucleobases but also that NH dissociation is of interest since it has been shown that this process leads to conical intersections [32] even though they are supposed to be accessible only by higher excitation energies beyond the first absorption band.

The above discussion concentrates on the singlet excited states. It should be noted that several studies discussing the triplet excited states appeared in the literature (see, e.g., [45–50]).

The large manifold of conical intersections and their reaction paths give a good picture of the different deactivation possibilities but makes conclusive predictions about the real deactivation mechanisms difficult if not impossible. Dynamics simulations have been performed to find the actual reaction paths and to obtain information about the characteristic times connected with the different processes. Owing to the strongly coupled motion involving many internal degrees of freedom, Tully surface hopping [51] and ab initio multiple spawning (AIMS) [52] appear to be methods of choice since all internal degrees of freedom are directly included in these dynamics simulations (see also [53] for a general discussion on the options and outlook of photodynamical simulations). The main bottleneck in these calculations is the necessity of performing a full quantum chemical calculation in each time step using extended methods able to describe several electronic states and their nonadiabatic coupling. Under these circumstances, mostly complete active space self-consistent field (CASSCF) or multireference configuration interaction with single excitations (MRCIS) approaches have been used [17, 19, 33, 38, 44, 54–56]. An overview of respective simulations of the photodynamics of nucleobases is available, e.g., in [18]. As an interesting alternative to the computationally expensive ab initio approaches, semi-empirical multireference methods based on the orthogonalization model 2 Hamiltonian (OM2) [34, 57, 58] and the fractional orbital occupation/AM1 model [59] have also been applied extensively in surface hopping dynamics. The performance of TDDFT using a larger variety of functionals has been investigated [60], and time-dependent tight binding density functional theory (TD-DFTB) [61] and DFTB mean field simulations [62] have been used as well. The restricted open shell Kohn–Sham (ROKS) method within the

framework of Car–Parinello dynamics [63] and quantum dynamical simulations have also been performed [64].

From this multitude of widely differing approaches, we wanted to discuss the photodynamics of adenine and cytosine in more detail. Surface hopping dynamics simulations using an MRCIS approach [33] have been performed for adenine, starting the dynamics in the $L_a \pi-\pi^*(S_3)$ state. Already after 25 fs this state is completely depopulated and practically all trajectories have arrived in the S_1 state after ~ 60 fs. By fitting an exponential function containing two decay constant, values of 22 and 538 fs were found, in good agreement with those of <50 and 750 fs measured by Ullrich et al. [9]. The analysis of the trajectories showed that almost exclusively the 2E with puckering at C_2 was accessed. In contrast, OM2 photodynamics [34] shows a strong predominance of the NH_2 out-of-plane conical intersection (1S_6). In a recent study [60] the energy profiles along the reaction coordinates leading to both mentioned intersections were computed using several methods (OM2/MRCI, MRCIS, complete active space perturbation theory to second order (CASPT2), approximate coupled cluster singles- and doubles method (CC2), and a variety of different TDDFT methods). These profiles show a preference toward the 2E intersection in the case of MRCIS, CASPT2, and CC2 whereas the opposite trend is found for OM2. It was further shown that the MRCIS calculations using one lone pair (n) orbital in the active space were subject to a certain bias toward 2E which was partially lifted by including two n orbitals into the active space [65]. Thus, according to current understanding, the real dynamics should still be dominated by the 2E pathway but significant participation of the 1S_6 should be expected as well. The investigations in [60] also show a severe failure of the TDDFT and TD-DFTB approaches since an insufficient number of hoppings to the ground state were observed. Inclusion of range-separated functionals did not improve the situation significantly. For more details see [60].

In the case of cytosine, a more complex dynamics than that described for adenine is observed [18]. All three intersections described above for cytosine participate in the dynamics. Initially, cytosine relaxes along the $\pi-\pi^*$ pathway to a region of strong mixture of $\pi-\pi^*$, $n-\pi^*$, and closed shell character, where in a first approach about 16% of trajectories switch to the ground state with a time constant of 13 fs. It should be noted that this semi-instantaneous deactivation of cytosine through the semi-planar intersection was also found in AIMS simulations [19]. In the remaining 84% of cases, cytosine quickly relaxes to the $n-\pi^*$ state from where it either can deactivate via the semi-planar intersection to the ground state or can overcome the barrier to the $\pi-\pi^*$ state and subsequently switch to the ground state. In contrast, CASSCF(2,2) AIMS [19] and OM2 [57] simulations do not show any significant deactivation in this region. The results of surface hopping dynamics at CASSCF (12,9) level [66] are similar to those found in [18] and show predominance of the semi-planar conical intersection. Due to this complex set of events it has to be expected that even rather subtle changes in the relative positions of the energy surfaces can lead to significant modifications of the photodynamical mechanisms and all discussed results have to be regarded with care. It can be expected that

further quantitative changes will occur when more sophisticated quantum chemical methods will become available for photodynamical simulations.

Notable changes in the dynamics also have to be expected when it is performed in aqueous solution. Several computations were performed to elucidate the effect of solvent on the excited states of uracil [49, 67–78], thymine [68, 76, 77, 79, 80], guanine [81–84], cytosine [75, 77, 85, 86], and adenine and its model systems [87–89]. The aqueous solvent was shown to modify the photodynamics of nucleobases by changing relative positions of excited states of different characters in the FC region, changing the heights of reaction barriers on the paths leading to conical intersections and relative energies of excited state minima. The consequences such as a blue shift of the $n-\pi^*$ states of uracil (see, e.g., [68, 70, 71, 73, 76], stabilization of a polar $S_2(\pi-\pi^*)$ [32] and $\pi-\sigma^*$ [87] states of adenine, and geometry changes of excited state minima of guanine [82] on photodynamics were discussed. Dynamics studies performed on adenine [89] have shown that the out-of-plane motion of the amino group is more pronounced in water, which can explain the faster decay of the S_1 state compared to the gas phase. The overall relaxation mechanism was however the same as observed in the gas phase. This finding is in contrast to guanine [84] where the pathway via a conical intersection with out-of-plane distortion of the carbonyl oxygen becomes dominant in water solvent.

It should be noted that in extension of the above-described dynamics simulations restricted to the singlet manifold, first surface hopping dynamics simulations have been performed for cytosine combining nonadiabatic and spin-orbit effects [90, 91].

At first sight it seems that each of the nucleobases possesses its own characteristics. A general picture emerging from the dynamics simulations [18] can be given, however, by the observation that the purine bases adenine and guanine have quite a simple deactivation pattern following basically one excited state to the intersection with the ground state whereas, in the case of the pyrimidine bases cytosine, thymine, and uracil, a significantly more complex picture appears with the participation of several excited states and a significantly more complex deactivation pattern.

1.2 Survey of Experimental Studies of Interacting Nucleobases

The question of the nature of excited states of DNA was first addressed in the 1960s. Based on theoretical models, Tinoco et al. [92] invoked a delocalized character of excited states of nucleic acids in the FC region to explain the hypochromism observed in absorption spectra of polynucleotides. In contrast to this prediction, Eisinger et al. [93] proposed that the UV photon is absorbed by a single base. This prediction was made on the basis of comparison of absorption spectra of DNA with corresponding pyrimidine and purine bases. The DNA absorption spectra closely

resembled the sum of the monomer spectra and showed theoretically predicted spectral shifts and splitting of the UV band around 260 nm. A discussion on this issue based on molecular dynamics simulations and quantum chemical calculations will be given later in this chapter.

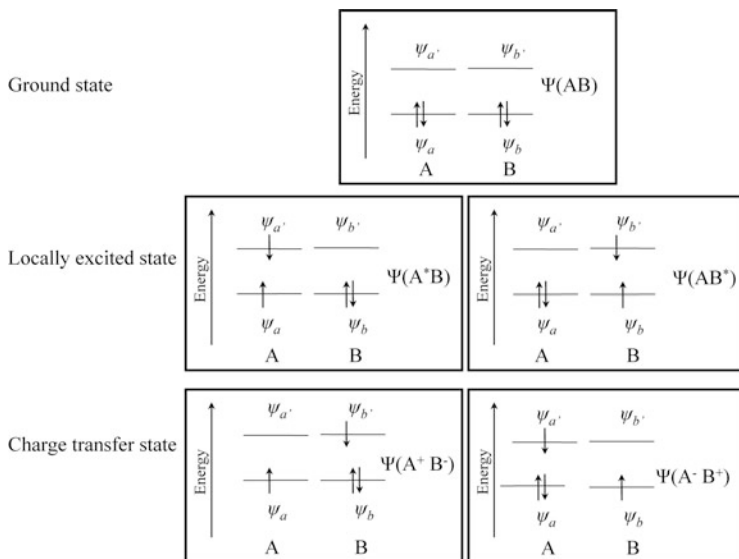
Following the prediction of the locally excited states red-shifted and broadened fluorescence spectra of di- and polynucleotides were attributed to excimers formed from these bases [5]. More pronounced occurrences of excimers observed at lower temperatures are explained by a higher degree of stacking as compared to more disordered structures at higher temperatures. Increased charge transfer (CT) contributions in these excimers were also predicted by these authors [94, 95].

In contrast to the above-discussed local character of absorption, the excitonic character in absorption spectra of di- and polynucleotides of adenine and cytosine was predicted by Kononov et al. [7], with the exciton limited to two bases. The excitonic features were, however, not observed in absorption spectra of guanine and thymine.

The experiments performed during the last two decades utilizing femtosecond time-resolved spectroscopy stimulated lively discussion on the nature of excited states of DNA. In addition to the ultrafast components decaying on the order of picoseconds, transients with lifetimes of 10–100 ps and even nanosecond time scale were observed for single- and double-stranded oligonucleotides [14, 96–103]. The existence of large decay times occurring for single-stranded oligonucleotides [98] demonstrates that stacking interactions are of primary importance for the explanation of differences in the complex decay dynamics in DNA as opposed to that observed for individual nucleic acid bases.

The interpretation of the ultrafast component observed in the experiments mentioned in the previous paragraph was based on the existence of bases undergoing monomer-like photo decay in the disordered parts of the oligo- and polymers. Different hypotheses were used to explain the slow-decay component: (1) the excitation in the FC region is localized on a single base and the interaction with an adjacent base leads to the formation of excimers [98, 101] several picoseconds after the excitation and (2) the excitation is already delocalized during the initial FC excitation, resulting in the formation of excitonic states [14, 96, 104–106]. In the latter interpretation partial emission from localized bases was suggested as well [105, 107]. Beyond the above question of the delocalization, the importance of charge transfer states is being discussed as an important issue (see, e.g., [108–113]).

A special challenge lies in interpreting and reconciling the large number of different decay times reported in the different experiments (see, e.g., [97–99, 101, 102, 114–121]). The results depend not only on the general character of the experimental technique (i.e., time-dependent absorption or emission spectroscopy) but also on the details of the preparation of the DNA samples and the base sequence, both playing an important role [118, 122]. In light of all these challenges with respect to interpreting experimental results, computational investigations play an important role in providing complementary information, shedding new light on the complex photodynamical behavior of DNA.



Scheme 2 Schematic illustration of the electronic interactions of identical chromophores *A* and *B* in terms of localized and charge transfer excited states

The extent of exciton delocalization is another issue discussed intensively. For example, Kadhane et al. suggested that excitons are delocalized over no more than two bases [123] and Bucharov et al. [14] suggested delocalization extending over three bases. However, delocalization over six and more bases was also considered [124]. Extended work concerning this issue is based mainly on dynamics simulations in connection with excitonic models or quantum chemical calculations, and will be discussed later in the chapter.

According to current evidence, inter-strand hydrogen bonding may also play an important role and add to the complexity of the photodynamical behavior of nucleic acids. A unique excited state behavior of Watson–Crick (WC) base pairs was shown in a resonant multi-photon ionization experiment performed in the gas phase [125]. Faster fluorescence decay of base pairs compared to isolated bases was also observed by other authors as shown in [98, 126–128]. Furthermore, interplay between intra-strand CT and inter-strand proton transfer has been suggested [128].

2 Description of Electronic Coupling

Before discussing the character of excited state interactions between nucleobases, the underlying theory will be briefly discussed. Scheme 2 describes the simplest situation of the electronic interactions of two identical chromophores (*A* and *B*) in terms of locally and charge transfer excited states. At infinite separation the locally

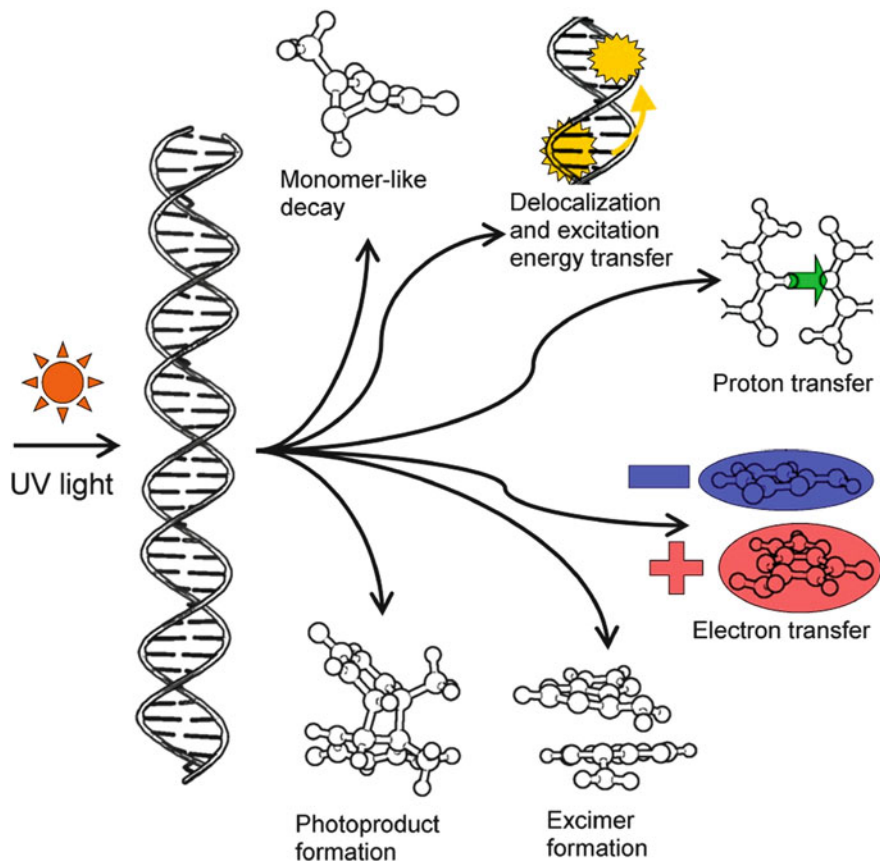


Fig. 3 Schematic representation of possible processes resulting from the interactions of nucleobases in electronically excited states

excited states (described by $\Psi(A^*B)$ and $\Psi(AB^*)$) are degenerate. At finite separation, in an adiabatic representation the degeneracy is removed and the energy splitting may be directly used to measure the excited state interaction (electronic coupling). The situation is similar for the charge transfer states described by $\Psi(A^-B^+)$ and $\Psi(A^+B^-)$ [129–131]. For non-symmetrical arrangements this approach is not so straightforward. Inside the polymer the environment of two bases is strongly non-symmetrical due to thermal fluctuations. In such a case the splitting will also reflect the difference in the environment of each chromophore, which causes a shift of the orbital and excitation energies that are not directly related to the electronic coupling. In the case of a stacked nucleobase pair with nucleobases mutually orientated as in a B-DNA structure, the changes in excitation energies observed during dimerization caused by non-symmetric effects are an order of magnitude larger than those caused by the ‘pure’ electronic interaction [132, 133].

Interaction between chromophores in electronically excited states can promote one of the following processes (Fig. 3):

1. Delocalization of the excited states which results in exciton formation.
2. Electronic excitation energy transfer.
3. Formation of a strongly interacting complex – an excimer – which results from the orbital overlap.
4. Formation of a new chemical species – a photochemical product.
5. Inter- and/or intra-strand electron or proton transfer.
6. Localization of the electronic excitation on a single base followed by a monomer-like relaxation.

The interactions during electronic excitations are governed by two main contributions: (1) Coulombic interactions which are often approximated by the interactions of transition dipole moments and higher multipole interaction terms and (2) short-range interactions, which depend on the orbital overlap between two chromophores [129–131].

While the former interactions operate over larger through-space separation R , and depend asymptotically on R^{-3} , the latter attenuate exponentially with R being unimportant for the chromophore separation larger than approximately 6 Å. Since these interactions depend on the orbital overlap they are greatly dependent on the mutual orientation of the chromophores. Thus, significantly larger short-range interactions were found for the nucleobases in the B-DNA-like orientation with almost parallel stacking as compared to A-DNA-like orientation with disordered stacking [132]. The character of the excited states also largely influences the resulting electronic interactions. For example, these interactions between the states of ($n-\pi^*$) character with negligible transition dipole moments and small orbital overlaps are much smaller in relation to states of ($\pi-\pi^*$) character which possess a larger transition dipole moment and a larger overlap of interacting orbitals [132].

2.1 Excimers/Exciplexes and Excitons

As already mentioned, the photoabsorption of two (or more) nucleobases within the nucleic acid structure can result in *excimers* or *exciplexes* being formed by two identical or different nucleobases, respectively. It should be emphasized that the excimers/exciplexes are not necessarily formed from one molecule in the ground state and the second from the excited state. They can be formed from different initial states, including excitons.

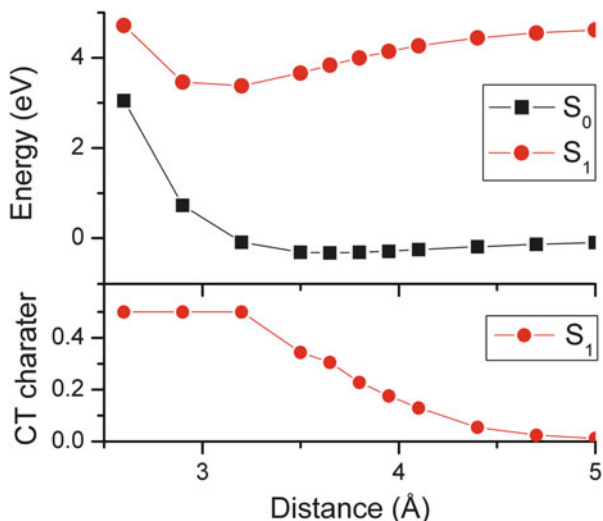
The wavefunction has the following general form [129, 134, 135]:

$$\Psi(\text{Exciplex}) = c_1\Psi(A^*B) + c_2\Psi(AB^*) + c_3\Psi(A^-B^+) + c_4\Psi(A^+B^-), \quad (1)$$

where $A = B$, $c_1 = c_2$, and $c_3 = c_4$ for excimers.

The first two terms correspond to locally excited states and their interaction results in exciton states (see below). At intermolecular separations below 5–6 Å, orbital interactions come into play [129, 134] mediating a mixing of the locally

Fig. 4 Excited state energies of the S_0 and S_1 states (*top*) and charge transfer character of the S_1 state (*bottom*) of a face-to-face stacked naphthalene dimer (see [138] for details)



excited states with the charge transfer or radical-ion-pair states, described by the third and fourth terms [135]. Stabilization of the exciplex can not only occur through excitonic coupling and Coulombic interaction, which are directly derived from the excitonic and CT contributions. It has been pointed out that with decreasing separation distance a new type of strong interaction may come into play that can be identified as a quasi-bond in the geminate radical ion pair [136]. Such quasi-bonds may go along with strong structural distortions, and experimental evidence shows that electronic and steric effects play an important role [137].

In Fig. 4, excimer formation in the face-to-face stacked naphthalene dimer computed at the ab initio algebraic diagrammatic construction to second order (ADC(2)) level is illustrated as a representative example (see [138] for more details). While the ground state potential is only weakly attractive with a shallow minimum around 3.65 Å intermolecular separation, the excited state is strongly bound with a minimum at 3.2 Å and stabilization energy of 1.38 eV with respect to infinite separation. Aside from the energy, a measure for charge transfer, representing the weight of the c_3 and c_4 coefficients in the equation (1), is shown in Fig. 4 (bottom). At separations >5.0 Å the CT value is approximately zero, showing that the excited state corresponds to a pure Frenkel exciton. When the molecules are moved closer together, orbital interactions are important (cf. [129]) and the locally excited states mix with the CT states, giving a gradual increase of the CT character. At the excimer minimum the CT character reaches 50%, showing that the excited state is now an even mixture between Frenkel excitonic and charge resonance contributions. This state is accessed by a single transition between delocalized orbitals, which corresponds to the situation of a coherent homogeneous state extended over the whole complex (cf. [138, 139]).

In extended quasi-periodic systems excited states can be delocalized over a number of chromophores. Such states are usually called *excitons*. In this context it is important to distinguish whether the exciton can be described as a linear combination of locally excited states, forming a Frenkel exciton, or whether CT configurations also play a role.

The first case can be understood in terms of Frenkel exciton theory [140, 141]. In this framework a model Hamiltonian \mathbf{H} is written as the sum of isolated chromophores H_a and a coupling term V_{ab} :

$$\mathbf{H} = \sum_a H_a + \sum_a \sum_{b>a} V_{ab}. \quad (2)$$

The singly excited states are described by the term

$$\Phi_a^i = (-1)^a \Phi_a^i \prod_{a \neq b} \Phi_b, \quad (3)$$

where Φ_a^i corresponds to the wavefunction of the state where chromophore a is in its excited state, while others are in their-ground state. The wavefunction of the excitonic state is then written as a linear combination of the wavefunction with locally excited states:

$$\Psi_k(\text{Exciton}) = \sum_a c_{ka} \Phi_a^i. \quad (4)$$

The diagonal and off-diagonal terms of the exciton matrix correspond to the excitation energies of the chromophore a and the exciton coupling terms, respectively. In this picture the excitation within some energy range populates a number of excited states delocalized over several nucleobases. Furthermore, if the initial electronic wavepacket is prepared as a non-stationary starting state, such a Hamiltonian automatically leads to excitation energy transfer. If in addition vibronic contributions are considered, the system can reach the lower part of the emission band via internal conversion (intra-band scattering) [118].

When there are strong orbital interactions between the different fragments, Frenkel exciton theory is no longer sufficient and it is necessary to include explicitly charge transfer configurations into the calculation. The theoretical treatment in such a case is more involved considering that a larger number of parameters are needed [142–144]. However, the advantage of such an approach is that charge separation processes can be easily modeled.

3 Assessment of Computational Methods

A wide range of computational methods have been applied to the description of electronically excited states of DNA fragments. In this section we review these approaches shortly. Unfortunately, no perfect method exists because of the difficulties in computing excited states and the relatively large sizes of the molecular

systems involved. In the following we will characterize the main features of the individual methods, including their strong and weak points, in an attempt to aid in assessing the substantial amount of available computational literature and to explain some of the discrepancies obtained in the different studies. This section will be strongly focused on DNA fragments; for a more general overview we refer the reader to [53].

3.1 *Electronic Structure Methods*

In this section, the different electronic structure methods that were used to describe DNA fragments will be considered. The main focus will be laid on the computation of excited states, and additionally methods for a phenomenological analysis of excited states and the computation of interaction potentials will be discussed. The main challenge relates to the large system sizes that have to be considered. Therefore, exciton models are a logical choice and were applied by several groups [142, 145–147]. In such an approach the excited states on the different molecules and their interactions are treated by a model Hamiltonian. This Hamiltonian can be easily extended to large system sizes, allowing the study of long range effects and potentially extended delocalization. However, a major challenge in such an approach is a proper parameterization. In particular the electronic couplings are of concern and a number of different methods for estimating them were used. Usually only the Coulombic contribution is computed while the direct orbital term is assumed to be smaller. The simplest approach for the estimation of this interaction is to consider only dipole–dipole interactions. This approximation was used, e.g., by Georghiou et al. [95] and Bittner [142] to estimate the efficiency of energy transfer along the DNA helix. This approach is suitable for systems in which the separation of the two chromophores is significantly greater than the dimension of the molecule. Its validity is however questionable in the case of nucleobases, because their molecular dimension is of the same order as the intermolecular distance of a neighboring nucleobase pair. Several approximations can help to overcome this difficulty using, e.g., the atomic-transition charge distribution [148, 149] or transition-density-cube models [144]. Comparison of the electronic couplings calculated using dipole–dipole and transition-density-cube approaches show that the former significantly overestimate the interactions at shorter distances [144]. Alternatively, the hybrid-multipole model which represents a combination of a truncated-multipole and the extended-dipole model [148, 150] was used to estimate Coulombic interactions [132]. Within this hybrid model, the multipole interaction up to R^{-5} is covered exactly; all terms with higher orders are considered if they arise from the multipole expansion of the extended dipole. The Coulomb part is usually the major component of the interaction potential, but a complete description also requires consideration of the direct orbital contributions. While it was shown from explicit quantum chemical calculations that at the ground state equilibrium geometry these terms only made a small contribution (<100 cm) [132], the involvement of such terms could in fact be identified through a mixing of

Frenkel excitonic and CT states [113]. An ad hoc addition of orbital overlap terms in a Frenkel exciton model is difficult as it has already been pointed out that a modest additional coupling of 100 cm had a dramatic effect by doubling the delocalization length [149]. A more extended treatment of orbital overlap and resulting charge separation is possible but requires a more involved formalism and a larger number of parameters [142, 151].

Considering the structural flexibility of DNA, an atomistic description is in many cases highly desirable. As one option semi-empirical methods were used [59, 152–155]. Due to the fact that they allow an efficient description of quite large systems they were used for extended dynamics simulations, providing interesting insights. However, similar to exciton models, semi-empirical methods rely on careful parameterization, a problem which can be avoided by the use of ab initio methods. Time-dependent density functional theory (TDDFT), which offers efficient excited state computations while relying on no or relatively few empirical parameters, has been used by a number of groups [109, 110, 156]. However, a major challenge for the application of TDDFT is the over-stabilization of charge transfer states occurring when local functionals are used [157]. This problem can be overcome by using range separated hybrid functionals or functionals containing an overall high amount of Hartree–Fock exchange but in such cases the energy of CT states crucially depends on the parameters determining the admixture of Hartree–Fock exchange [110, 112]. For a comparison of the performance of different density functionals when applied to DNA stacks see, e.g., [158]. Finally, a large number of wavefunction-based ab initio calculations was performed using single- and multi-reference methods. In the first case, in particular efficient second order models like the approximate coupled cluster method CC2 [159] and the algebraic diagrammatic construction ADC(2) [160] in connection with the resolution of the identity approximation [161] were applied [112, 113, 132]. However, computationally more demanding models like the equation-of-motion coupled-cluster for excitation energies (EOM-EE-CC) with double excitations and even perturbative triple excitations have also been used [162–165]. In the case of strongly distorted structures and intersections between different states, multi-reference methods are needed to provide a reliable description. For this purpose the complete-active space self-consistent field (CASSCF) method, second order perturbation theory on this reference (CASPT2) [166], and multi-reference configuration interaction with single and double excitations (MR-CISD) [167] were performed [168]. While wavefunction-based methods offer the attractive property of systematic improvability toward the exact solution, they suffer from high computational demands. Unless massively parallel computer systems are available heavy truncations have to be carried out as far as the excitation level and/or the one electron basis set are concerned. For this purpose wavefunction-based ab initio methods also require careful testing and analysis before they can be successfully applied.

After the computation of excitation energies the next decisive task is to get a maximum of information for interpretive and phenomenological models. In quantum chemical calculations on DNA fragments this task may be quite challenging in many cases due to many interacting configurations, partially

delocalized orbitals, and the presence of hidden charge resonance contributions. Several approaches have been taken to analyze the states in more detail. One useful indicator is Mulliken populations, which aside from simply quantifying charge transfer, can also be used to estimate the contribution of a given monomer to an electronic transition [169]. Furthermore, energetic criteria based on model Hamiltonians have been applied to estimate electronic couplings and to identify charge resonance states [132, 162]. A different route may be taken by considering that the transition density matrix (TDM) between the ground and excited states can be used to represent the structure of the exciton as an electron-hole pair [138, 139, 170]. By partitioning the TDM into blocks corresponding to the different fragments, locally excited, excitonic, and CT contributions can be readily identified [138]. This approach was applied to absorption spectra [113] and excimer formation [168].

Another critical problem of great importance in the description of excimers is the computation of interaction potentials. In particular, dispersion, which is the main force determining stacking interactions, is difficult to describe. In the case of non-correlated methods, dispersion is completely absent, while standard DFT functionals usually strongly underestimate it. By contrast, MP2 is known to overbind complexes [171] and from the pure ab initio methods usually only CCSD(T) with basis set extrapolation is considered to provide reliable results [172]. To obtain accurate interaction potentials a number of empirical corrections have been suggested, where in particular Grimme's dispersion correction for DFT is widely used [173]. While parameterized methods can provide very good results for standard ground state geometries, it is not clear how they perform for excited states and in particular for strongly bound exciplex structures. It has been pointed out that standard force field parameters severely exaggerate repulsion at short intermolecular separations [172], which means that attempts to use such parameters in the description of excimers may result in an underestimation of binding energies and an overestimation of binding distances. Another problem arising from short intermolecular separations is increased basis set superposition error (BSSE). It was pointed out that the counterpoise (CP) correction for BSSE may strongly affect excimer structures and binding energies [23]. However, it was later also shown that the CP correction, when applied to smaller basis sets, may significantly overshoot, thereby incorrectly destabilizing the resulting exciplexes [168].

3.2 *Environmental Models, Sampling, and Dynamics*

Aside from a proper description of the electronic structure, new challenges arise with respect to the description of environmental effects, proper sampling to obtain statistically significant results, and the simulation of dynamical phenomena. For representing environmental effects, continuum models as well as atomistic

descriptions have been applied. The former type of approach offers a simple well-defined way to include the main effects of environmental polarization where even a differentiation between slow and fast polarization effects is easily achieved. In particular the polarizable continuum model (PCM) [174] has been applied, e.g., in [109, 112, 175]. There are only a few adjustable parameters, most importantly the dielectric constant and the time-regime (equilibrium or non-equilibrium). However, there is some arbitrariness when choosing an effective dielectric constant to represent the effect of the heterogeneous and anisotropic surroundings (i.e., the other DNA bases, the backbone, the surrounding water molecules and counterions). To overcome this problem, quantum mechanics/molecular mechanics (QM/MM) coupling schemes for an atomistic description of the environment have been widely used [110, 113, 176, 177]. Usually the effects of the environment are considered at the level of electrostatic embedding only, and electronic polarization of the environment is neglected (see [178] for a definition of these terms). This should have an effect on vertical excitation energies, while in dynamics simulations the main solvent response can be included through orientational polarization. A critical observation, made from the PCM computations, is that the environment may have a strong impact on the stability of charge transfer states and that, aside from the time-regime (equilibrium or non-equilibrium), even the precise PCM implementation (linear-response [179] or state-specific [180]) makes a crucial difference [181]. A similar sensitivity to the environmental models is probably also present in a QM/MM framework. Thus, in summary, the importance of an accurate description of the environment should be highlighted.

The most straightforward way for sampling ground state DNA structures is by classical molecular dynamics (MD) using standard biomolecular force fields and such an approach has been taken by a number of groups [144, 149, 156, 176]. The situation becomes more complicated when effects of zero-point vibrations should also be included. For this purpose a hybrid approach based on mixed initial conditions was introduced: Large scale motions and solvent degrees of freedom are properly treated by MD while the Wigner distribution of the vibrational ground state is used to represent the central molecule of interest [182]. This approach was applied to produce initial conditions for QM/MM dynamics simulations of a DNA base [177], and a slight extension considering several active molecules was used for spectra simulations [113]. QM/MM geometry optimizations can be performed by using an averaged solvent electrostatic potential generated from MD sampling of the environmental motions [183]. This method was applied to excited state optimizations of the adenine dinucleotide [168].

To go beyond static calculations, a number of excited state dynamics studies have also been performed. A particular focus was placed on the description of non-adiabatic effects, which are essential for understanding, e.g., internal conversion, excitation energy transfer, and charge separation processes. Dynamics have been performed using exciton models [143] and wave packet propagation on parameterized surfaces [184] but in most cases on-the-fly surface hopping dynamics were applied [59, 154, 155, 176, 177]. In the latter case a QM/MM approach was often chosen to allow a real time polarization response of the environment. While

the studies presented above considered only the singlet manifold, intersystem crossings to the triplet were also already considered in surface hopping dynamics [90].

4 UV Absorption

The nature of excited states in the Franck–Condon region significantly affects the excited state behavior of nucleic acids. Thus, understanding the character of absorption spectra is crucial for the further evaluation of the photodynamics of nucleic acids. As already mentioned in the Introduction, there is still controversy as to whether during photon absorption the nucleic acids form exciton or charge transfer states or whether they remain localized. In this chapter the survey of theoretical works suggesting both possibilities of delocalization is treated separately.

4.1 Excitonic Delocalization

The concept of excitons in nucleic acids was first introduced by Tinoco et al. [92, 185] and Rhodes [186]. However, a localized character of excited states then dominated the discussions for several decades. The question of exciton character was opened again by Bouvier et al. [148]. In this work the homogeneous $(dA)_{20} \cdot (dT)_{20}$ and alternating $(dAdT)_{10} \cdot (dAdT)_{10}$ oligonucleotides in idealized B-DNA geometry are investigated. The two lowest excited states of adenine and one excited state of thymine are considered. The interaction between the states is described by the atomic transition charges model [187] in which the transition dipoles are decomposed onto atomic orbitals. This procedure results in transition charges located on each atom. Using this approach, delocalization of the excited states was found for both oligomers. In $(dA)_{20} \cdot (dT)_{20}$ the intra-strand coupling dominates with a strength of about 250 cm^{-1} . In the alternating $(dAdT)_{10} \cdot (dAdT)_{10}$ the inter-strand coupling is more important, the estimated value of the coupling being approximately 100 cm^{-1} . In the following contribution [104] the influence of structure fluctuations on the character of the Franck–Condon excited states is analyzed for oligonucleotides $(dA)_{10} \cdot (dT)_{10}$ and $(dAdT)_5 \cdot (dAdT)_5$. A ground-state molecular dynamics simulation to scan possible DNA structures resulting from the plasticity of the sugar-phosphate helix. Importantly, the diagonal energies of monomer excited states are not affected by structural fluctuations, with the change being smaller than 10 cm^{-1} . Relatively large fluctuations of the off-diagonal terms for these structures were found, with the amplitude of the variations 35% and 45% for $(dA)_{10} \cdot (dT)_{10}$ and $(dAdT)_5 \cdot (dAdT)_5$, respectively. The distribution of the oscillator strengths, on the other hand, is not significantly affected by the structural dynamics for the $(dA)_{10} \cdot$

(dT)₁₀ oligonucleotide. In fact, 90% of the oscillator strength remains concentrated on the same eigenstates. In the case of the alternating (dAdT)₅.(dAdT)₅ oligonucleotide the structural disorder results in a larger spreading of the oscillator strength among eigenstates. Studies performed on (dCdG)₅.(dCdG)₅ [149] resulted in similar conclusions as for the afore-mentioned (dAdT)₅.(dAdT)₅ case: small fluctuations of monomer excitation energies, less than 15 cm⁻¹, and a large perturbation of dipolar coupling due to the structural disorder. In all the cases mentioned above a delocalization of the excited states over at least two nucleobases was observed. Delocalization over three bases was found in the calculations of the absorption spectra using the TDDFT method in combination with ground-state MD simulations for single-stranded oligomers of adenine [156] and (adenine–thymine) oligomers [188]. Formation of delocalized exciton states was further investigated in studies which combine ground-state molecular dynamics and the evaluation of the exciton model for the poly(dA).poly(dT) [143] and (dA)₁₂.(dT)₁₂ [144]. The lattice model and the transition density cube model, based on the transition model calculated using the TDDFT method, were used in the former and later investigations, respectively. In agreement with the work of Bouvier et al. [104] and Emanuele et al. [149], changes in electronic coupling due to structural fluctuation were found. The results indicated that during the absorption process the electronic excitation is delocalized over at least six nucleobases and localizes upon relaxation on four nucleobases. Importantly, these authors predicted the presence of charge transfer excitons in their model [144]. Charge transfer character of excitons was also suggested by Starikov et al. [189]. The character of exciton states due to structural fluctuations of B-DNA conformations was investigated in short (dA)_n.(dT)_n and (dC)_n.(dG)_n oligomers (with $n = 3,4$) obtained by means of ground-state molecular dynamics [152]. The effect of different conformational modes was evaluated, among which the twist is predicted as the most powerful regulator of the exciton character. It was also shown that the effect of the twist angle on the localization of electronic states is stronger in poly(dG)-poly(dC) than in poly(dA)-poly(dT) [190]. A new study based on ab initio calculations of alternating duplexes after extended sampling [113] presented a picture of rather localized states, which were situated on one or at most two bases. Considering poly(dAdT)-poly(dAdT) and poly(dGdC)-poly(dGdC) duplexes with four stacked bases in the QM region and the remaining part of the system treated at the MM level, only about a third of the states are delocalized over 1.5 bases or more. These are, however, responsible for somewhat more than 50% of the intensity at the absorption maximum due to higher than average oscillator strengths. Intramolecular vibrations were identified as the main factor responsible for spectral broadening and for causing disorder resulting in localization. This strong coupling of intramolecular modes to the excited states suggests that these are also active in the early excited state dynamics. The picture of rather localized states was also drawn from another recent study that compared measured and computed circular dichroism spectra in (dA)_n.(dT)_n hairpins [147]. The TDDFT study on adenine stacks pointed out that

the A_5 spectrum is almost identical to that of A_4 [191], i.e., there are no relevant effects either of excitonic or indirect nature which go beyond four bases. While most studies focused on singlet states, triplet excitations were studied in poly(dA)-poly(dT) sequences, and it was reported that they are confined to single nucleobases [192, 193].

4.2 Charge Transfer States

Compared to the studies performed within the framework of exciton models where the charge transfer configurations are usually not included, supermolecular quantum chemical calculations automatically provide a simultaneous treatment of excitonic and the charge transfer states. The interpretation of the absorption spectra in terms of delocalized, charge transfer and localized characters of excited states became a matter of discussion due to the performance of different methods. Suitability of various methods to describe long-range charge transfer states between nucleic acids has been tested in recent years (see, for example, [108, 110–112, 169, 194–196]).

In the text below the calculations of the character of excited states observed at the ground state geometries, i.e., upon UV absorption, as affected by base pairing and stacking interactions will be reviewed with the emphasis on a formation of charge transfer states. In initial *ab initio* calculations the effect of base pairing was studied [42, 197–199]. Although charge transfer states were detected at higher energies when using the CIS method [197], these were the lowest in calculations at the TDDFT level employing the LDA functional [198]. Stacking interactions were considered for the first time in calculations of Varsano et al. [200]. Other calculations studying the effects of stacking on excited states formed upon UV absorption in the gas phase have been reported, e.g., in [132, 133, 162, 201–203]. The difficulties of a correct description of charge transfer states in the absorption spectra is demonstrated in the case of homologous oligomers of adenine, both single-stranded and in double stranded A–T sequences, as well as alternating A–T sequences.

Santoro et al. [109] performed the first study on the excited states of stacked nucleobases in a water environment. In this contribution the absorption spectra of the adenine dimer are studied employing TDDFT with the PBE0 functional for the (9-Met-A)₂ and (dA)₂ models using a polarizable continuum model for solvent effects. The experimentally observed features when going from monomers to multimers [97, 105, 118], i.e., a blue-shift of the maximum, a red-shift of the lower-energy part, and hypochromic effect of the absorption spectra are already reproduced in the calculations of the former model. These effects reflect the coupling due to the orbital overlap in the stacking configuration of nucleobases. The symmetric combination of monomer bright transitions gives rise to the maximum of the absorption band. The character of the states responsible for low-lying energy transitions changes with the mutual orientation of the

adjacent adenines, i.e., an increasing intra-strand charge transfer character of the first excited state was found in less symmetrical arrangements in $(dA)_2$ in comparison to $(9\text{-Met-A})_2$ [109]. The same change of the maximum absorption peak, i.e., blue shift and decrease in the intensity with increasing number of stacked nucleobases, was already observed in the gas phase in calculations of Lange et al. [110] performed with a long-range-corrected PBE0 functional (LRC- ω -PBE0) on the homologous $(A)_n$ and $(T)_n$, ($n = 1\text{--}4$). In contrast to the calculations performed with a non-corrected density functional [109], charge transfer character was not observed in the states responsible for the red tail of the absorption spectra. The effect of the solvent is studied by means of a QM/MM approach with water molecules in the first solvation shell (within 2.5 Å) treated at the QM level. Depending on the solvent configuration the energies of charge transfer states span more than 1 eV and for some configurations they overlap with bright $\pi\pi^*$ states. The average stabilization of the charge transfer states by the solvent is 0.1 eV.

A blue shift of the maximum absorption band is also observed for double-stranded $(A)_2(T)_2$ with localization on the $(A)_2$ dimer while excited states localized on the $(T)_2$ dimer are responsible for the low energy part of the spectrum [169]. The calculations performed with the M052X functional place the intra-strand ($A \rightarrow A$) and inter-strand ($A \rightarrow T$) charge transfer states in the range of bright excited states of the adenine dimer. Note that in the PBE0 calculations the charge transfer states are the most stable. As in the case of single-stranded oligomers, the gas phase calculations performed with the LRC- ω -PBE0 functional [110] place the intra-strand charge transfer states above the bright states. The inclusion of solvent causes a stabilization of these states by about 0.1 eV on the average.

For a single-strand with an alternating sequence of adenine and thymine studied with the LRC- ω -PBE0 functional in the gas phase, Lange et al. [110] found that, due to a mismatch of $\pi\pi^*$ state energies of these nucleobases, the intra-strand excitonic delocalization is missing and, consequently, the bright states are localized on a single base. In contrast to homologous oligomers, the CT states become resonant with the $\pi\pi^*$ states of adenine and thymine. When the second strand is included, the intra-strand CT states are placed about 0.4 eV below the bright absorption peak of adenine, while the inter-strand CT states appear about 0.7 eV above this bright peak. Note that the ADC(2) calculations performed by Aquino et al. [112] placed the lowest intra-strand CT state 0.5 and 0.3 eV above the lowest $\pi\pi^*$ states localized on adenine and thymine, respectively.

Recently Plasser et al. [113] reported the results of ADC(2) calculations of alternating oligomer duplexes in both gas phase and embedded in DNA environment employing a QM/MM scheme, together with a detailed analysis of the excited states. For both systems considered, poly(dAdT)-poly(dAdT) and poly(dGdC)-poly(dGdC), a similar picture was drawn: CT states were found energetically well above the bright states. Due to their low intensity they do not significantly contribute to the absorption spectra even when the DNA

environment is involved. A statistical treatment of the QM/MM simulations shows that only about half of the states exhibit Frenkel exciton character (a CT contribution below 0.1 e) while the remaining states show a non-negligible admixture of CT contributions, which means that they may be misrepresented in a pure Frenkel exciton model. About 15% of the states considered show significant charge separation (>0.5 e).

5 Excited State Processes

A number of factors may play a role in DNA photoactivity. These may be divided into external factors like sterical hindrances and electrostatic interactions on the one hand, and electronic interactions on the other. From a computational point of view the significance is that for the former class it may be sufficient to treat only one base at the QM level and the environment at a lower, e.g., molecular mechanics, level (QM/MM method). By contrast, for the second class it is indispensable to consider several bases simultaneously at the QM level, which imposes restrictions on the available approaches. In this chapter, studies concerned with external interactions will be reviewed first. Then direct electronic interactions will be considered, which may occur between stacked bases (leading to excitons, CT states, and exciplexes) as well as hydrogen bonded bases (where proton coupled electron transfer processes are of special interest). Finally, further reactions determining the photochemistry of DNA will be discussed.

5.1 Sterical Hindrances and Electrostatic Interactions

The calculations of the excited state relaxation of adenine [204] performed at the CASPT2/CASSCF/AMBER level assumed that the main reaction channel involves the 2E conical intersection for the adenine molecule (see above, Fig. 2) in both vacuo and solvated $(dA)_{10} \cdot (dT)_{10}$ duplex. In the latter case the reaction path is flatter and features a small barrier of about 0.2 eV. These characteristics are suggested to be responsible for the slow decay component (>100 ps) observed in single and double-stranded systems with stacked adenine, thus questioning the importance of delocalized excited states. Nonadiabatic dynamics simulations employing the QM/MM method with semi-empirical treatment of the QM part (using OM2/MRCI) [58, 155] performed on an adenine molecule embedded in $(dA)_{10}$ and $(dA)_{10} \cdot (dT)_{10}$ in water predict an elongation of the excited state lifetime of adenine in comparison with the gas phase but the decay still occurs within 10 ps. In particular, decay times of 5.7 and 4.1 ps for $(dA)_{10}$ and $(dA)_{10} \cdot (dT)_{10}$, respectively, were predicted. In the single-stranded system relaxation proceeds mainly via the 6S_1 conical intersection but the 2E pathway coexists, while in

double-stranded DNA the 6S_1 conical intersection is blocked due to inter-strand hydrogen bonding and only the 2E intersection is accessed.

Surface hopping dynamics simulations using the QM/MM approach with CASSCF (cytosine) and MR-CIS (guanine) wavefunction for the QM part were performed for these two nucleobases, each embedded individually in a DNA double strand helix [177]. The restraining influence of the inter-strand hydrogen bonding on the structural deformations necessary to reach conical intersections was studied. In the case of photoexcited cytosine the isolated molecule shows relatively small puckering of the structure at the conical intersections populated during the excited state relaxation. The geometrical restrictions exerted by the hydrogen bonds of the DNA environment thus do not inhibit the photodeactivation of cytosine and consequently its excited state lifetime. In contrast to this, the isolated guanine relaxes to the ground state with strong out-of-plane motions of the NH_2 group. This motion is significantly restrained by inter-strand hydrogen bonds which results in a considerable elongation of the relaxation time.

The effect of the intra-strand interaction on the photodecay of adenine in a single-stranded DNA was studied in nonadiabatic dynamics simulations using 4-aminopyrimidine (4AP) [205] as a model for adenine. In these QM/MM calculations 4AP was treated at the CASSCF level. Comparison with the previously investigated dynamics of isolated 4AP [33] shows a very similar relaxation mechanism and only slight elongation of the excited state lifetime. During the dynamics of embedded 4AP the dynamical formation and breaking of intra-strand hydrogen bonds was observed. Interestingly, these bonds contribute to a faster decay component by enhancing the out-of-plane motion of the amino-group in relevant conical intersections.

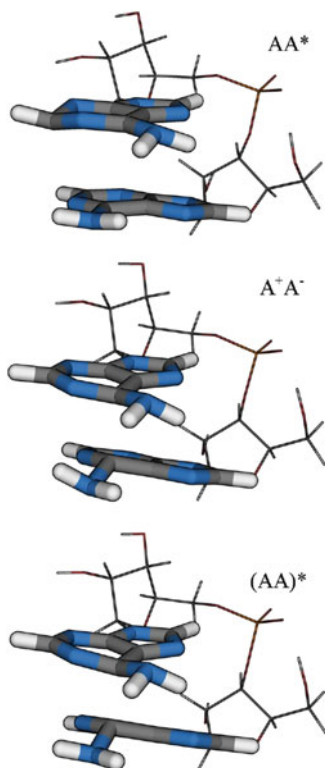
5.2 *Excimer Formation and Excitation Energy Transfer*

An initial study used static calculations of stacked adenine dimers and trimers at the TDDFT/PBE0 level with PCM solvation to discuss the effect of stacking interactions on their excited state behavior [109]. This study describes a process starting from rather delocalized absorbing states, with a subsequent localization on one base on a subpicosecond time scale. After this, the system could either deactivate to the ground state or reach a low energy CT minimum on the S_1 surface, which could act as a trapping site. A quantum dynamical study performed at the same computational level indicated that charge separation could be fast and effective [108]. Subsequent studies on a number of similar cases used, aside from PBE0, the range-separated CAM-B3LYP and the M05-2X functional containing a large percentage of non-local exchange. Consideration of the $(\text{dA})_2 \cdot (\text{dT})_2$ system showed that the interaction between stacked bases was more important than between hydrogen bonded bases. Again, charge transfer excimers were found. A subsequent study on AG and GA stacks also emphasized the importance of charge transfer in these systems [181]. Furthermore, efficient coupling of the CT states to the locally

excited states was reported. Later, excited state relaxation in (dA)₄ was considered [206]. In this investigation several excited state minima were found pertaining to localized excited states, neutral excimer states, and a CT minimum. The CT minimum was identified as the most stable one. These studies used the newly implemented state-specific PCM formalism [180] to describe the solvent response to the excitation. In these calculations it was shown that this new PCM formalism led to results significantly different from the standard linear-response approach, in particular stabilizing the CT states. Similar results were also found in a more recent combined experimental and theoretical paper [191].

In contrast to the TDDFT studies, a CASPT2/CASSCF work described the formation of neutral excimers, which were especially stabilized in maximum-overlap face-to-face configurations, whereas charge transfer excimers played only a secondary role [175]. Similar to the above analysis, a bifurcation of the initial wavepacket was considered leading either to ultrafast monomer-like decay or toward the formation of excimer states. A dynamics survey using semi-empirical methods [154] described the formation of a long-lived excimer between two adenine molecules, which was structurally characterized by a short intermolecular separation of the C2 atoms (cf. Scheme 1) to an average distance of 2.2 Å. A new decay channel of this system was found, which occurred through an additional shortening of the C2–C2 distance to 1.8 Å and concurrent ring deformations. A subsequent study by this group, using dynamics at the DFTB level [207], even described the formation of a chemical bond between two adenine molecules after excitation, which was stable for about 2 ps. Whereas similar processes in pyrimidine bases can lead to photodimerization (see below), a non-reactive deactivation was observed in these dynamics investigations. Recent *ab initio* work using the ADC(2) and MR-CISD methods in a QM/MM framework found several minima on the S₁ surface, which possessed different degrees of delocalization and charge transfer (see Fig. 5) [168]. The lowest energy minimum was of exciplex type stabilized neither by pure excitonic interactions nor charge transfer, but by direct orbital interactions, which were mediated by a close approach of the adenine C6 atoms (down to about 2.0 Å) and a concurrent strong distortion of the molecular structures. Indications for various decay channels were found, which were related to either a further approach of the two molecules (as in [154]) or a restoration of monomer-like excited states and decay through the ⁶S₁ intersection. Partial charge transfer was observed in response to solvent polarization but it did not play a dominant role. A similar conclusion was also drawn from a recent study using the RI-CIS(D) method [208] presenting a bonded exciplex mediated by an approach of two C6 atoms. In that case no excited state minimum but only an intersection was found. However, it was pointed out that the solvent effects may destabilize this intersection. These newer results may be rationalized with the bonded exciplex model [136] (see also Sect. 2.1). Experimental evidence for this type of mechanism may be found in the unexpected fact of negative fluorescence anisotropy recorded for (dA)₂₀, which was taken as evidence of an out-of-plane polarization of the emitting states [191], a phenomenon which is forbidden in planar molecules by

Fig. 5 Excited state minima of ApA localized on the S_1 surface: excitation localized on a single Ade (*top*), excitation from localized n orbital to delocalized π^* orbital (*middle*), excitation from delocalized π to delocalized π^* orbital (see [168] for details)



symmetry selection rules. Strong orbital overlap, combining two bases to one effective chromophore, can lift this selection rule, and a transition dipole moment with a strong out-of-plane component was indeed found for the exciplex [168].

Aside from explicit quantum chemical calculations, exciton models have also been used to describe excited state dynamics in DNA. The poly(dA).poly(dT) duplex was studied using a model considering both Frenkel excitonic and charge-transfer interactions [143]. The dynamics in this study were dominated by base-stacking whereas inter-strand interactions only played a minor role. It was found that on the adenine side the exciton remained in a cohesive bound state (i.e., no charge transfer occurred between stacked adenine molecules). By contrast, electron-hole separation into mobile charge carriers was reported on the thymine side. It was hypothesized that such charge separation and subsequent recombination could lead to triplet formation.

While the above studies considered adenine and thymine molecules, stacking interactions between two cytosine molecules were also investigated at the CASPT2 level [209] showing an attractive potential between the two cytosine molecules identified in the excited state, which could lead to red-shifted fluorescence. The strength of this interaction experienced strong dependence on BSSE. Using the CP

correction, an intermolecular separation of 3.08 Å and an excimer binding energy of 0.58 eV was obtained.

5.3 *Proton Transfer Processes*

The initial interest in the proton transfer process dates back to Watson and Crick pointing out that the occurrence of rare tautomeric forms of the nucleobases may lead to spontaneous mutations [210] and to Löwdin discussing that such tautomers may be formed through double proton tunneling in the DNA structure [211]. It has been pointed out by Guallar et al. [212] that such double proton transfer is not feasible in the ground state but that such a pathway could be accessed in the excited state. However, the computations suggested that it is unlikely that the rare tautomers would persist long enough to perturb the duplication of the genetic code [212]. Gorb et al. [213] reported that while the AT double proton transfer was completely inaccessible, the corresponding GC structure would show minimal population (with an equilibrium constant of about 10^{-6}). The same group also reported excited state investigations on the AT and GC pairs [197], as well as the AU pair [197]. In both cases the presence of locally excited states and higher lying charge transfer states was reported.

In 2004 Sobolewski and Domcke put forward the hypothesis that a proton coupled electron transfer (PCET) along the central hydrogen bond in the Watson–Crick base pair of guanine and cytosine could serve as an efficient decay channel to the ground state [42]. This hypothesis was based on CASSCF/CASPT2 calculations on structures and reaction paths optimized at the CIS level. It was found that in the vicinity of the FC region a stable charge transfer state existed, and that starting in this state a transfer of the central hydrogen bonding proton (yielding in total a PCET) could neutralize the system and stabilize this state. Potential curves revealed a weakly avoided crossing between the CT and ground states, which might provide an efficient channel for the relaxation. The study also found a locally excited path for cytosine to the ethylenic intersection, which was only later described for the isolated molecule (see, e.g., [44, 214]). An experimental work on GC dimers in the gas phase later found that the Watson–Crick bound base pair did indeed possess a much broader UV spectrum as opposed to other structures of this system [125]. These results were rationalized by computations at the CC2 level, where the three lowest energy conformers of the GC system were considered [215]. In all three cases a proton transfer path could be identified. However, this path was only favored in the WC case, whereas for the other conformers significant barriers would have to be overcome due to the fact that the CT states were higher in energy for these structures. Subsequently, time-resolved fluorescence experiments in chloroform were carried out, which supported the existence of such a decay channel by showing a short lifetime of about 0.36 ps for the GC base pair [127]. However, transient absorption experiments in chloroform in connection with TDDFT

calculations in PCM solvent contested this claim [126]. Regarding GC duplexes, it was pointed out from experiment that PCET may play a role in the case of alternating but not in homopolymeric sequences [128]. This idea was supported computationally by comparing TDDFT calculations of the GGG · CCC and GCG · CGC hexamer stacks [216].

Aside from static calculations, non-adiabatic dynamics simulations were also performed to get a deeper mechanistic insight into a potential proton transfer in GC and why it should or should not be effective. In particular, it is of interest to understand the coupling between the electronic and nuclear motion during the PCET process. In this regard the GC pair was studied by CASSCF using an approximate diabatic surface hopping method [176]. The simulations were performed in vacuo and embedded in a DNA double helix solvated in water using a QM/MM scheme. Dynamics were started in the S_1 (CT) state and the processes leading to the ground state decay were monitored. After a quick initial proton transfer, the back transfer was briefly inhibited by a diabatic trapping situation leading to several recrossings before reaching the closed shell configuration on a 100 fs time scale. This phenomenon was explained by an unusual intersection topology, possessing effectively only a one-dimensional branching space. The authors showed that the inclusion of the environment could speed up the decay through a stabilization of the charge transfer state. At this point it should be mentioned that the intersection topology described in [176] may be interpreted as a signature of an electron transfer process, which necessarily occurs when switching from the charge transfer state to the neutral state. In fact, similar surface topologies and diabatic trapping dynamics were also obtained in electron transfer model systems [217], further supporting this interpretation. The PCET deactivation process in the WC paired GC system was also studied using non-adiabatic dynamics at the restricted open shell Kohn–Sham (ROKS) level, considering the system in vacuo as well as in water solvation [218]. A steering technique was used to accelerate the forward PCET. In this study a fast deactivation on the order of 100 fs was also reported. To date, no study has been performed attempting to describe the dynamics starting from the bright $\pi\pi^*$ state to the CT state and to give a non-adiabatic description of the forward PCET process, considering that this would pose special challenges as a larger number of excited states would have to be described concurrently. However, a similar process was studied in the 2-pyridone dimer model system, using static calculations [219] and non-adiabatic dynamics, both performed at the TDDFT/BHLYP level [220] following time-resolved experiments [221]. The dynamics study highlighted that the PCET was not only governed by the energetics describing the proton transfer but also that non-adiabatic transition probabilities for the electron transfer (which were manifested by temporary hoppings into the adiabatic S_2 state) played a decisive role. This observation may provide also an explanation for the discrepancies between the dynamics studies and the above-mentioned experiments on the GC base pair [101, 126]. An efficient deactivation channel may indeed exist once the proton is transferred from G to C. However, it is not clear whether the initial proton transfer occurs after excitation into one of the bright states.

While most studies focused on the GC pair, proton transfer in the AT pair was also studied at the CC2 level [222]. Similar to [215], it was also found for AT that the WC structure possessed an efficient decay channel related to proton transfer. While the CT state was 1.5 eV above the lowest bright transition at the FC region, this state was strongly stabilized by proton transfer. Barrierless deactivation could occur through the CT state by a sequence of conical intersections. Internal conversion to the $n\pi^*$ state could serve as a competitive channel. As opposed to the WC structure no such efficient decay path was present in the most stable hydrogen bonded conformer of AT as the CT state was located at higher energy with respect to the minimum of the locally excited state. A TDDFT study of the WC bonded AT pair [223] highlighted the importance of solvent effects. In particular it was pointed out that a polar solvent could strongly stabilize the inter-strand CT state, thus disfavoring the subsequent proton transfer. The coupling via a $\pi\sigma^*$ state explains the time-resolved photoionization spectroscopy of adenine and the adenine dimer. In adenine this path is not dominating although it does play a role as indicated by the H-atom detection in nanosecond spectroscopic experiments. In the adenine dimer an increased population of $\pi\sigma^*$ transfer is observed. It is caused by stabilization of the relevant state in the dimer [87].

5.4 Photochemical Processes

The major photoproducts observed upon UV-irradiation of DNA include dimer or adduct formations of adjacent pyrimidine nucleobases, particularly cyclobutane pyrimidine dimers (CPDs) and the pyrimidine(6-4) pyrimidone dimers ((6-4)PP). The former products are observed more frequently (for review of experimental investigations see [224] and references therein). The dimerization proceeds via [2+2] cycloaddition of the C5–C6 double bonds of adjacent pyrimidine nucleobases and can be formed via both the singlet and triplet manifolds [224–227]. The thymine dimer (T<>T) is the major photoproduct occurring after DNA irradiation by UV light. The cytosine dimer (C<>C) on the other hand is more likely to induce mutations once it is formed [225, 228].

In the last decade, several results of experimental studies were reported, discussing the mechanism of CPD formation [226, 229–236]. For example, Schreier et al. showed that the T<>T is formed on the picosecond time scale which suggests that the formation proceeds in the singlet excited state [230]. The quantum yield of this reaction is, however, very low, of the order of 10^{-2} . Based on time-resolved femtosecond transient absorption spectroscopy, Kwok et al. [232] suggested that both pathways might simultaneously contribute to the dimerization. In this contribution the role of other excited states for this process, in particular the role of the $n\pi^*$ state, is discussed in the intersystem crossing to the triplet state.

The theoretical investigations focusing on the mechanism of the more frequently formed T<>T photoproduct considered both triplet and singlet pathways of the dimerization process. In the triplet state the stepwise mechanism was suggested

based on the results of the TDDFT [237] and CASPT2 [238] calculations. The singlet pathway was proposed to proceed via a concerted mechanism [202, 203, 236, 237, 239–241] mediated via S_1/S_0 conical intersections leading to either the original ground state or to $T\langle\rangle T$. Blancafort and Migani [240] suggested coexistence of the alternative singlet pathway where the excitation is first localized on a single thymine with a subsequent formation of quasi-minimum of the S_1 state localized energetically above the relevant conical intersection. Based on the recently published joint experimental and theoretical investigations Banyasz et al. [236] put forward that the [2+2] cycloaddition occurs from the lowest excitonic state and follows a barrierless path. No CT states were identified in the reaction mechanism, in contrast to the formation of (6-4)PP adducts (for more detailed discussion on the mechanism of (6-4)PP formation see [242, 243]). The absence of charge transfer character in the former reaction might explain a decrease of the $T\langle\rangle T$ production in the systems with purine residues flanking the TT sequences by a formation of charge transfer purine-pyrimidine(T) exciplexes and subsequent decrease of the TT exciton population observed in the experiment [244].

Much effort has been made to explain the origin of very low quantum yield of the $T\langle\rangle T$ dimerization [229, 244–250]. It is now generally accepted that the ground state conformation of adjacent thymine controls the efficiency of this reaction with inter-molecular T...T distance and torsional angle between two double bonds (C5–C6 bond) on each thymine moiety being identified as the most important. While the agreement on the criteria for the former parameter was achieved [246–248, 250], the importance of the latter parameter is not clarified yet [246, 247]. The influence of the sugar conformation on the $T\langle\rangle T$ dimerization process was discussed as well [245, 250–252]. In a recently published contribution Improta suggested that a barrierless path from the delocalized exciton state towards $T\langle\rangle T$ is most effective when both sugars have C3-endo puckered conformation since the electronic coupling between thymines is larger compared to other sugar conformations [203].

The explanation of the experimentally observed lower yields of $C\langle\rangle C$ photo-product as compared to $T\langle\rangle T$ [201, 239, 241, 253] has been the subject of several theoretical investigations. Results of these studies reveal a unified mechanism, i.e., concerted photocycloaddition of the BPD formation for both sequences. Possible competing reaction channels include monomer-like relaxation and excimer formation. The energy position of the conical intersection which mediates the cycloaddition ($CI_{\text{dimer}}(S_0/S_1)$) with respect to stable excimer structures and CI corresponding to monomer-like relaxations determines the efficiency of the BPD formation. While $CI_{\text{dimer}}(S_0/S_1)$ is the lowest-energy structure in the case of thymine, it is comparable to the energy of the excimer in the case of cytosine and the monomer-like CI, opening other reaction funnels.

6 Conclusions

The area of light absorption by DNA and subsequent photophysical and photochemical processes is a fascinating research field with many important practical implications for everyday life. It is also an outstanding example for the interaction and influences between theory and experiment in an intensity which is very unique. In writing this chapter we have made an attempt to document the large variety of theoretical approaches and different, sometimes opposing, viewpoints to the photodynamics of DNA. Looking back on the history one can observe enormous effort and progress made in experiments and in theoretical simulations by collecting many facts and putting them in the proper perspective. However, the story is certainly not over yet and many interesting and fundamental insights are still waiting to be discovered.

Acknowledgments This work was supported by the Austrian Science Fund within the framework of the Special Research Program F41, Vienna Computational Materials Laboratory (ViCoM). We also acknowledge technical support from and computer time at the Vienna Scientific Cluster (Projects 70019 and 70151). Support was also provided by the Robert A. Welch Foundation under Grant No. D-0005. FP is a recipient of a research fellowship by the Alexander von Humboldt Foundation. This work has been supported by the grants of the Grant Agency of the Czech Republic (P208/12/1318) and the grant of the Czech Ministry of Education, Youth and Sport (LH11021). The research at IOCB was part of the project RVO:61388963.

References

1. Pfeifer GP, You YH, Besaratinia A (2005) *Mutat Res Fund Mol M* 571:19
2. Coulondre C, Miller JH (1977) *J Mol Biol* 117:577
3. Brash DE, Rudolph JA, Simon JA, Lin A, McKenna GJ, Baden HP, Halperin AJ, Ponten J (1991) *Proc Natl Acad Sci U S A* 88:10124
4. Becker MM, Wang Z (1989) *J Mol Biol* 210:429
5. Rochette PJ, Therrien JP, Drouin R, Perdiz D, Bastien N, Drobetsky EA, Sage E (2003) *Nucleic Acids Res* 31:2786
6. Beletskii A, Bhagwat AS (2001) *J Bacteriol* 183:6491
7. Douki T (2006) *J Photoch Photobio B Biol* 82:45
8. Ullrich S, Schultz T, Zgierski MZ, Stollow A (2004) *Phys Chem Chem Phys* 6:2796
9. Ullrich S, Schultz T, Zgierski MZ, Stollow A (2004) *J Am Chem Soc* 126:2262
10. Canuel C, Mons M, Piuze F, Tardivel B, Dimicoli I, Elhanine M (2005) *J Chem Phys* 122:074316
11. Crespo-Hernandez CE, Cohen B, Hare PM, Kohler B (2004) *Chem Rev* 104:1977
12. Doorley GW, McGovern DA, George MW, Towrie M, Parker AW, Kelly JM, Quinn SJ (2009) *Angew Chem Int Ed* 48:123
13. Peon J, Zewail AH (2001) *Chem Phys Lett* 348:255
14. Buchvarov I, Wang Q, Raychev M, Trifonov A, Fiebig T (2007) *Proc Natl Acad Sci U S A* 104:4794
15. Markovitsi D, Sharonov A, Onidas D, Gustavsson T (2003) *ChemPhysChem* 4:303
16. Marian CM (2005) *J Chem Phys* 122:104314

17. Hudock HR, Levine BG, Thompson AL, Satzger H, Townsend D, Gador N, Ullrich S, Stolow A, Martinez TJ (2007) *J Phys Chem A* 111:8500
18. Barbatti M, Aquino AJA, Szymczak JJ, Nachtigallova D, Hobza P, Lischka H (2010) *Proc Natl Acad Sci U S A* 107:21453
19. Hudock HR, Martinez TJ (2008) *ChemPhysChem* 9:2486
20. Chen H, Li SH (2006) *J Chem Phys* 124:154315
21. Perun S, Sobolewski AL, Domcke W (2006) *J Phys Chem A* 110:13238
22. Perun S, Sobolewski AL, Domcke W (2005) *J Am Chem Soc* 127:6257
23. Serrano-Andres L, Merchan M, Borin AC (2006) *Proc Natl Acad Sci U S A* 103:8691
24. Matsika S (2004) *J Phys Chem A* 108:7584
25. Ismail N, Blancafort L, Olivucci M, Kohler B, Robb MA (2002) *J Am Chem Soc* 124:6818
26. Sobolewski AL, Domcke W (2002) *Eur Phys J D* 20:369
27. Epifanovsky E, Kowalski K, Fan PD, Valiev M, Matsika S, Krylov AI (2008) *J Phys Chem A* 112:9983
28. Serrano-Andres L, Merchan M, Borin AC (2008) *J Am Chem Soc* 130:2473
29. Hare PM, Crespo-Hernandez CE, Kohler B (2007) *Proc Natl Acad Sci U S A* 104:435
30. Blancafort L, Cohen B, Hare PM, Kohler B, Robb MA (2005) *J Phys Chem A* 109:4431
31. Chen H, Lis S (2005) *J Phys Chem A* 109:8443
32. Perun S, Sobolewski AL, Domcke W (2005) *Chem Phys* 313:107
33. Barbatti M, Lischka H (2008) *J Am Chem Soc* 130:6831
34. Fabiano E, Thiel W (2008) *J Phys Chem A* 112:6859
35. Cremer D, Pople JA (1975) *J Am Chem Soc* 97:1354
36. Marian CM (2007) *J Phys Chem A* 111:1545
37. Lan ZG, Fabiano E, Thiel W (2009) *ChemPhysChem* 10:1225
38. Barbatti M, Szymczak JJ, Aquino AJA, Nachtigallova D, Lischka H (2011) *J Chem Phys* 134:014304
39. Yamazaki S, Domcke W, Sobolewski AL (2008) *J Phys Chem A* 112:11965
40. Kistler KA, Matsika S (2008) *J Chem Phys* 128:215102
41. Blancafort L, Robb MA (2004) *J Phys Chem A* 108:10609
42. Sobolewski AL, Domcke W (2004) *Phys Chem Chem Phys* 6:2763
43. Zgierski MZ, Patchkovskii S, Fujiwara T, Lim EC (2005) *J Phys Chem A* 109:9384
44. Barbatti M, Aquino AJA, Szymczak JJ, Nachtigallova D, Lischka H (2011) *Phys Chem Chem Phys* 13:6145
45. Merchan M, Serrano-Andres L, Robb MA, Blancafort L (2005) *J Am Chem Soc* 127:1820
46. Gonzalez-Luque R, Climent T, Gonzalez-Ramirez I, Merchan M, Serrano-Andres L (2010) *J Chem Theory Comput* 6:2103
47. Etinski M, Fleig T, Marian CA (2009) *J Phys Chem A* 113:11809
48. Serrano-Perez JJ, Gonzalez-Luque R, Merchan M, Serrano-Andres L (2007) *J Phys Chem B* 111:11880
49. Marian CM, Schneider F, Kleinschmidt M, Tatchen J (2002) *Eur Phys J D* 20:357
50. Climent T, Gonzalez-Luque R, Merchan M, Serrano-Andres L (2007) *Chem Phys Lett* 441:327
51. Tully JC (1990) *J Chem Phys* 93:1061
52. Ben-Nun M, Martinez TJ (2002) *Adv Chem Phys* 121:439
53. Plasser F, Barbatti M, Aquino AJA, Lischka H (2012) *Theor Chem Acc* 131
54. Asturiol D, Lasorne B, Robb MA, Blancafort L (2009) *J Phys Chem A* 113:10211
55. Szymczak JJ, Barbatti M, Hoo JTS, Adkins JA, Windus TL, Nachtigallova D, Lischka H (2009) *J Phys Chem A* 113:12686
56. Nachtigallova D, Aquino AJA, Szymczak JJ, Barbatti M, Hobza P, Lischka H (2011) *J Phys Chem A* 115:5247
57. Lan ZG, Fabiano E, Thiel W (2009) *J Phys Chem B* 113:3548
58. Lu Y, Lan ZG, Thiel W (2012) *J Comput Chem* 33:1225
59. Alexandrova AN, Tully JC, Granucci G (2010) *J Phys Chem B* 114:12116

60. Barbatti M, Lan ZG, Crespo-Otero R, Szymczak JJ, Lischka H, Thiel W (2012) *J Chem Phys* 137:22A503
61. Mitric R, Werner U, Wohlgemuth M, Seifert G, Bonacic-Koutecky V (2009) *J Phys Chem A* 113:12700
62. Lei YB, Yuan SA, Dou YS, Wang YB, Wen ZY (2008) *J Phys Chem A* 112:8497
63. Langer H, Doltsinis NL, Marx D (2005) *ChemPhysChem* 6:1734
64. Picconi D, Barone V, Lami A, Santoro F, Improta R (2011) *ChemPhysChem* 12:1957
65. Szymczak JJ, Barbatti M, Lischka H (2011) *Int J Quantum Chem* 111:3307
66. Gonzalez-Vazquez J, Gonzalez L (2010) *ChemPhysChem* 11:3617
67. DeFusco A, Minezawa N, Slipchenko LV, Zahariev F, Gordon MS (2011) *J Phys Chem Lett* 2:2184
68. Gustavsson T, Banyasz A, Lazzarotto E, Markovitsi D, Scalmani G, Frisch MJ, Barone V, Improta R (2006) *J Am Chem Soc* 128:607
69. Gustavsson T, Sarkar N, Banyasz A, Markovitsi D, Improta R (2007) *Photochem Photobiol* 83:595
70. Improta R, Barone V (2004) *J Am Chem Soc* 126:14320
71. Santoro F, Barone V, Gustavsson T, Improta R (2006) *J Am Chem Soc* 128:16312
72. Gustavsson T, Improta R, Markovitsi D (2010) *J Phys Chem Lett* 1:2025
73. Yoshikawa A, Matsika S (2008) *Chem Phys* 347:393
74. Mercier Y, Santoro F, Reguero M, Improta R (2008) *J Phys Chem B* 112:10769
75. Kistler KA, Matsika S (2009) *J Phys Chem A* 113:12396
76. Etinski M, Marian CM (2010) *Phys Chem Chem Phys* 12:4915
77. Rasmussen AM, Lind MC, Kim S, Schaefer HF (2010) *J Chem Theory Comput* 6:930
78. Olsen JM, Aidas K, Mikkelsen KV, Kongsted J (2010) *J Chem Theory Comput* 6:249
79. Nosenko Y, Kunitzki M, Brutschy B (2011) *J Phys Chem A* 115:9429
80. Busker M, Nispel M, Haber T, Kleinermanns K, Etinski M, Fleig T (2008) *ChemPhysChem* 9:1570
81. Shukla MK, Leszczynski J (2008) *J Phys Chem B* 112:5139
82. Shukla MK, Leszczynski J (2009) *Chem Phys Lett* 478:254
83. Shukla MK, Leszczynski J (2010) *Int J Quantum Chem* 110:3027
84. Heggen B, Lan ZG, Thiel W (2012) *Phys Chem Chem Phys* 14:8137
85. Kistler KA, Matsika S (2010) *Phys Chem Chem Phys* 12:5024
86. Blancfort L, Migani A (2007) *J Photochem Photobiol A Chem* 190:283
87. Ritze HH, Lippert H, Samoylova E, Smith VR, Hertel IV, Radloff W, Schultz T (2005) *J Chem Phys* 122:224320
88. Szymczak JJ, Muller T, Lischka H (2010) *Chem Phys* 375:110
89. Lan ZG, Lu Y, Fabiano E, Thiel W (2011) *ChemPhysChem* 12:1989
90. Richter M, Marquetand P, Gonzalez-Vazquez J, Sola I, Gonzalez L (2012) *J Phys Chem Lett* 3:3090
91. Mai S, Marquetand P, Richter M, Gonzales-Vazquez J, Gonzales L (2013) *ChemPhysChem* 14:1
92. Tinoco IJ (1960) *J Am Chem Soc* 82:4785
93. Eisinger J, Shulman RG (1968) *Science* 161:1311
94. Gueron M, Shulman RG, Eisinger J (1966) *Proc Natl Acad Sci U S A* 56:814
95. Georghiou S, Zhu S, Weidner R, Huang CR, Ge G (1990) *J Biomol Struct Dyn* 8:657
96. Markovitsi D, Gustavsson T, Sharonov A (2004) *Photochem Photobiol* 79:526
97. Crespo-Hernandez CE, Kohler B (2004) *J Phys Chem B* 108:11182
98. Crespo-Hernandez CE, Cohen B, Kohler B (2005) *Nature* 436:1141
99. Crespo-Hernandez CE, de La Harpe K, Kohler B (2008) *J Am Chem Soc* 130:10844
100. Kwok WM, Ma CS, Phillips DL (2006) *J Am Chem Soc* 128:11894
101. Middleton CT, de La Harpe K, Su C, Law YK, Crespo-Hernandez CE, Kohler B (2009) *Annu Rev Phys Chem* 60:217
102. Vaya I, Miannay FA, Gustavsson T, Markovitsi D (2010) *ChemPhysChem* 11:987

103. Plessow R, Brockhinke A, Eimer W, Kohse-Hoinghaus (2000) *J Phys Chem B* 104:3695
104. Bouvier B, Dognon JP, Lavery R, Markovitsi D, Millie P, Onidas D, Zakrzewska K (2003) *J Phys Chem B* 107:13512
105. Onidas D, Gustavsson T, Lazzarotto E, Markovitsi D (2007) *J Phys Chem B* 111:9644
106. Markovitsi D, Onidas D, Gustavsson T, Talbot F, Lazzarotto E (2005) *J Am Chem Soc* 127:17130
107. Onidas D, Gustavsson T, Lazzarotto E, Markovitsi D (2007) *Phys Chem Chem Phys* 9:5143
108. Santoro F, Barone V, Improta R (2009) *J Am Chem Soc* 131:15232
109. Santoro F, Barone V, Improta R (2007) *Proc Natl Acad Sci U S A* 104:9931
110. Lange AW, Herbert JM (2009) *J Am Chem Soc* 131:3913
111. Lange AW, Rohrdanz MA, Herbert JM (2008) *J J Phys Chem B* 112:6304
112. Aquino AJA, Nachtigallova D, Hobza P, Truhlar DG, Hättig C, Lischka H (2011) *J Comp Chem* 32:1217
113. Plasser F, Aquino AJA, Hase WL, Lischka H (2012) *J Phys Chem A* 116:11151
114. Crespo-Hernandez CE, Cohen B, Kohler B (2006) *Nature* 441:E8
115. Takaya T, Su C, de La Harpe K, Crespo-Hernandez CE, Kohler B (2008) *Proc Natl Acad Sci U S A* 105:10285
116. de La Harpe K, Crespo-Hernandez CE, Kohler B (2009) *ChemPhysChem* 10:1421
117. Vaya I, Gustavsson T, Miannay FA, Douki T, Markovitsi D (2010) *J Am Chem Soc* 132:11834
118. Markovitsi D, Gustavsson T, Talbot F (2007) *Photochem Photobiol Sci* 6:717
119. Markovitsi D, Gustavsson T, Vaya I (2010) *J Phys Chem Lett* 1:3271
120. Onidas D, Gustavsson T, Lazzarotto E, Markovitsi D (2007) *Phys Chem Chem Phys* 9:1
121. Vaya I, Changenet-Barret P, Gustavsson T, Zikich D, Kotlyar AB, Markovitsi D (2010) *Photochem Photobiol Sci* 9:1193
122. Schwalb NK, Temps F (2008) *Science* 322:243
123. Kadhane U, Holm AIS, Hoffmann SV, Nielsen SB (2008) *Phys Rev E* 77:021901
124. Markovitsi D, Gustavsson T, Banyasz A (2010) *Mutat Res Rev Mutat* 704:21
125. Abo-Riziq A, Grace L, Nir E, Kabelac M, Hobza P, de Vries MS (2005) *Proc Natl Acad Sci U S A* 102:20
126. Biemann L, Kovalenko SA, Kleinermanns K, Mahrwald R, Markert M, Improta R (2011) *J Am Chem Soc* 133:19664
127. Schwalb NK, Temps F (2007) *J Am Chem Soc* 129:9272
128. de La Harpe K, Crespo-Hernandez CE, Kohler B (2009) *J Am Chem Soc* 131:17557
129. Scholes GD, Ghiggino KP (1994) *J Phys Chem* 98:4580
130. Scholes GD (1996) *J Phys Chem* 100:18731
131. Scholes GD (1999) *J Phys Chem B* 103:2543
132. Nachtigallova D, Hobza P, Ritze HH (2008) *Phys Chem Chem Phys* 10:5689
133. Ritze HH, Hobza P, Nachtigallova D (2007) *Phys Chem Chem Phys* 9:1672
134. East ALL, Lim EC (2000) *J Chem Phys* 113:8981
135. Klessinger M, Michl J (1995) *Excited states and photochemistry of organic molecules*. Wiley-VCH, New York
136. Wang YS, Haze O, Dinnocenzo JP, Farid S, Farid RS, Gould IR (2007) *J Org Chem* 72:6970
137. Wang YS, Haze O, Dinnocenzo JP, Farid S, Farid RS, Gould IR (2008) *J Phys Chem A* 112:13088
138. Plasser F, Lischka H (2012) *J Chem Theory Comput* 8:2777
139. Tretiak S, Mukamel S (2002) *Chem Rev* 102:3171
140. Frenkel J (1931) *Phys Rev* 37:1276
141. Davydov AS (1971) *Theory of molecular excitons*. McGraw-Hill, New York
142. Bittner ER (2006) *J Chem Phys* 125:094909
143. Bittner ER (2007) *J Photochem Photobio A Chem* 190:328
144. Czader A, Bittner ER (2008) *J Chem Phys* 128:035101
145. Emanuele E, Markovitsi D, Millie P, Zakrzewska K (2005) *ChemPhysChem* 6:1387

146. Conwell EM, Bloch SM, McLaughlin PM, Basko DM (2007) *J Am Chem Soc* 129:9175
147. Patwardhan S, Tonzani S, Lewis FD, Siebbeles LDA, Schatz GC, Grozema FC (2012) *J Phys Chem B* 116:11447
148. Bouvier B, Gustavsson T, Markovitsi D, Millie P (2002) *Chem Phys* 275:75
149. Emanuele E, Zakrzewska K, Markovitsi D, Lavery R, Millie P (2005) *J Phys Chem B* 109:16109
150. Czikkely V, Forsterling HD, Kuhn H (1970) *Chem Phys Lett* 6:207
151. Conwell EM, McLaughlin PM, Bloch SM (2008) *J Phys Chem B* 112:2268
152. Starikov EB, Cuniberti G, Tanaka S (2009) *J Phys Chem B* 113:10428
153. Granucci G, Persico M (2007) *J Chem Phys* 126:134114
154. Zhang W, Yuan S, Whang Z, Qi Z, ZhaO J, Dou Y, Lo GV (2011) *Chem Phys Lett* 506:303
155. Lu Y, Lan ZG, Thiel W (2011) *Angew Chem Int Ed* 50:6864
156. Tonzani S, Schatz GC (2008) *J Am Chem Soc* 130:7607
157. Dreuw A, Weisman JL, Head-Gordon M (2003) *J Chem Phys* 119:2943
158. Improta R (2008) *Phys Chem Chem Phys* 10:2656
159. Christiansen O, Koch H, Jorgensen P (1995) *Chem Phys Lett* 243:409
160. Trofimov AB, Schirmer J (1995) *J Phys B At Mol Opt* 28:2299
161. Hattig C, Weigend F (2000) *J Chem Phys* 113:5154
162. Kozak CR, Kistler KA, Lu Z, Matsika S (2010) *J Phys Chem B* 114:1674
163. Szalay PG, Watson T, Perera A, Lotrich V, Fogarasi G, Bartlett RJ (2012) *J Phys Chem A* 116:8851
164. Szalay PG, Watson T, Perera A, Lotrich V, Bartlett RJ (2013) *J Phys Chem A* 117:3149
165. Szalay PG (2013) *Int J Quantum Chem* 113:1821
166. Andersson K, Malmqvist PA, Roos BO (1992) *J Chem Phys* 96:1218
167. Szalay PG, Muller T, Gidofalvi G, Lischka H, Shepard R (2012) *Chem Rev* 112:108
168. Plasser F, Lischka H (2013) *Photochem Photobiol Sci* 12:1440
169. Santoro F, Barone V, Improta R (2008) *ChemPhysChem* 9:2531
170. Luzanov AV, Zhikol OA (2010) *Int J Quantum Chem* 110:902
171. Riley KE, Platts JA, Rezac J, Hobza P, Hill JG (2012) *J Phys Chem A* 116:4159
172. Morgado CA, Jurecka P, Svozil D, Hobza P, Sponer J (2010) *Phys Chem Chem Phys* 12:3522
173. Grimme S (2004) *J Comput Chem* 25:1463
174. Cossi M, Barone V, Cammi R, Tomasi J (1996) *Chem Phys Lett* 255:327
175. Olasso-Gonzalez G, Merchan M, Serrano-Andres L (2009) *J Am Chem Soc* 131:4368
176. Groenhof G, Schafer LV, Boggio-Pasqua M, Goette M, Grubmuller H, Robb MA (2007) *J Am Chem Soc* 129:6812
177. Zeleny T, Ruckebauer M, Aquino AJA, Muller T, Lankas F, Drsata T, Hase WL, Nachtigallova D, Lischka H (2012) *J Am Chem Soc* 134:13662
178. Bakowies D, Thiel W (1996) *J Phys Chem* 100:10580
179. Cossi M, Barone V (2001) *J Chem Phys* 115:4708
180. Improta R, Scalmani G, Frisch MJ, Barone V (2007) *J Chem Phys* 127
181. Santoro F, Barone V, Lami A, Improta R (2010) *Phys Chem Chem Phys* 12:4934
182. Ruckebauer M, Barbatti M, Muller T, Lischka H (2010) *J Phys Chem A* 114:6757
183. Galvan IF, Sanchez ML, Martin ME, del Valle FJO, Aguilar MA (2003) *J Chem Phys* 118:255
184. Improta R, Santoro F, Barone V, Lami A (2009) *J Phys Chem A* 113:15346
185. Tinoco I, Bradley DF, Woody RW (1963) *J Chem Phys* 38:1317
186. Rhodes W (1961) *J Am Chem Soc* 83:3609
187. Claverie P (1978) In: Pullman B (ed) *Intramolecular interactions – from diatomics to biopolymers*. Wiley, New York, p 69
188. Voityuk AA (2013) *Photochem Photobiol Sci* 12:1303
189. Starikov EB, Lewis JP, Sankey OF (2005) *Int J Mod Phys B* 19:4331
190. Yamada H, Starikov EB, Hennig D, Archilla JFR (2005) *Eur Phys J E* 17:149

191. Banyasz A, Gustavsson T, Onidas D, Changenet-Barret P, Markovitsi D, Improta R (2013) *Chem Eur J* 19:3762
192. Curutchet C, Voityuk AA (2011) *Chem Phys Lett* 512:118
193. Curutchet C, Voityuk AA (2011) *Angew Chem Int Ed* 50:1820
194. Jensen L, Govind N (2009) *J Phys Chem A* 113:9761
195. Shukla MK, Leszczynski J (2010) *Mol Phys* 108:3131
196. Santoro F, Barone V, Improta R (2008) *J Comput Chem* 29:957
197. Shukla MK, Leszczynski J (2002) *J Phys Chem A* 106:1011
198. Tsokalidis A, Kaxiras E (2005) *J Phys Chem A* 109:2373
199. Wesolowski TA (2004) *J Am Chem Soc* 126:11444
200. Varsano D, Di Felice R, Marques MAL, Rubio A (2006) *J Phys Chem B* 110:7129
201. Roca-Sanjuan D, Olaso-Gonzalez G, Gonzalez-Ramirez I, Serrano-Andres L, Merchan M (2008) *J Am Chem Soc* 130:10768
202. Boggio-Pasqua M, Groenhof G, Schafer LV, Grubmuller H, Robb MA (2007) *J Am Chem Soc* 129:10996
203. Improta R (2012) *J Phys Chem B* 116:14261
204. Conti I, Altoe P, Stenta M, Garavelli M, Orlandi G (2010) *Phys Chem Chem Phys* 12:5016
205. Nachtigallova D, Zeleny T, Ruckebauer M, Muller T, Barbatti M, Hobza P, Lischka H (2010) *J Am Chem Soc* 132:8261
206. Improta R, Barone V (2011) *Angew Chem Int Ed* 50:12016
207. Dou YS, Liu ZC, Yuan S, Zhang WY, Tang H, Zhao JS, Fang WH, Lo GV (2013) *Int J Biol Macromol* 52:358
208. Spata VA, Matsika S (2013) *J Phys Chem A* 117. doi:[10.1021/jp4033194](https://doi.org/10.1021/jp4033194)
209. Olaso-González G, Roca-Sanjuán D, Serrano-Andres L, Merchan M (2006) *J Chem Phys A* 125:231102
210. Watson JD, Crick FHC (1953) *Nature* 171:737
211. Lowdin PO (1963) *Rev Mod Phys* 35:724
212. Guallar V, Douhal A, Moreno M, Lluch JM (1999) *J Phys Chem A* 103:6251
213. Gorb L, Podolyan Y, Dziekonski P, Sokalski WA, Leszczynski J (2004) *J Am Chem Soc* 126:10119
214. Blancafort L (2007) *Photochem Photobiol* 83:603
215. Sobolewski AL, Domcke W, Hattig C (2005) *Proc Natl Acad Sci U S A* 102:17903
216. Ko C, Hammes-Schiffer S (2013) *J Phys Chem Lett* 4:2540
217. Plasser F, Lischka H (2011) *J Chem Phys* 134:034309
218. Markwick PRL, Doltsinis NL (2007) *J Chem Phys* 126:175102
219. Sagvolden E, Furche F (2010) *J Phys Chem A* 114:6897
220. Plasser F, Granucci G, Pittner J, Barbatti M, Persico M, Lischka H (2012) *J Chem Phys* 137:22A514
221. Muller A, Talbot F, Leutwyler S (2002) *J Chem Phys* 116:2836
222. Perun S, Sobolewski AL, Domcke W (2006) *J Phys Chem A* 110:9031
223. Dargiewicz M, Biczysko M, Improta R, Barone V (2012) *Phys Chem Chem Phys* 14:8981
224. Cadet J, Mouret S, Ravanat JL, Douki T (2012) *Photochem Photobiol* 88:1048
225. Ravanat JL, Douki T, Cadet J (2001) *J Photochem Photobiol B Biol* 63:88
226. Cuquerella MC, Lhiaubet-Vallet V, Bosca F, Miranda MA (2011) *Chem Sci* 2:1219
227. Bosca F, Lhiaubet-Vallet V, Cuquerella MC, Castell JV, Miranda MA (2006) *J Am Chem Soc* 128:6318
228. Lee DH, Pfeifer GP (2003) *J Biol Chem* 278:10314
229. Schreier WJ, Schrader TE, Koller FO, Gilch P, Crespo-Hernandez CE, Swaminathan VN, Carell T, Zinth W, Kohler B (2007) *Science* 315:625
230. Schreier WJ, Kubon J, Regner N, Haiser K, Schrader TE, Zinth W, Clivio P, Gilch P (2009) *J Am Chem Soc* 131:5038
231. Marguet S, Markovitsi D (2005) *J Am Chem Soc* 127:5780
232. Kwok WM, Ma C, Phillips DL (2008) *J Am Chem Soc* 130:5131

233. Douki T, Cadet J (2003) *Photochem Photobiol Sci* 2:433
234. Mouret S, Philippe C, Gracia-Chantegrel J, Banyasz A, Karpati S, Markovitsi D, Douki T (2010) *Org Biomol Chem* 8:1706
235. Tommasi S, Denissenko MF, Pfeifer GP (1997) *Cancer Res* 57:4727
236. Banyasz A, Douki T, Improta R, Gustavsson T, Onidas D, Vaya I, Perron M, Markovitsi D (2012) *J Am Chem Soc* 134:14834
237. Zhang RB, Eriksson LA (2006) *J Phys Chem B* 110:7556
238. Climent T, Gonzalez-Ramirez I, Gonzalez-Luque R, Merchan M, Serrano-Andres L (2010) *J Phys Chem Lett* 1:2072
239. Gonzalez-Ramirez I, Roca-Sanjuan D, Climent T, Serrano-Perez JJ, Merchan M, Serrano-Andres L (2011) *Theor Chem Acc* 128:705
240. Blancafort L, Migani A (2007) *J Am Chem Soc* 129:14540
241. Serrano-Perez JJ, Gonzalez-Ramirez I, Coto PB, Merchan M, Serrano-Andres L (2008) *J Phys Chem B* 112:14096
242. Labet V, Jorge N, Morell C, Douki T, Grand A, Cadet J, Eriksson LA (2013) *Photochem Photobiol Sci* 12:1509
243. Giussani A, Serrano-Andres L, Merchan M, Roca-Sanjuan D, Garavelli M (2013) *J Phys Chem B* 117:1999
244. Pan ZZ, Hariharan M, Arkin JD, Jalilov AS, McCullagh M, Schatz GC, Lewis FD (2011) *J Am Chem Soc* 134:3611
245. Santini GPH, Pakleza C, Auffinger P, Moriou C, Favre A, Clivio P, Cognet JAH (2007) *J Phys Chem B* 111:9400
246. Law YK, Azadi J, Crespo-Hernandez CE, Olmon E, Kohler B (2008) *Biophys J* 94:3590
247. Johnson AT, Wiest O (2007) *J Phys Chem B* 111:14398
248. McCullagh M, Hariharan M, Lewis FD, Markovitsi D, Douki T, Schatz GC (2010) *J Phys Chem B* 114:5215
249. Pan Z, McCullagh M, Schatz GC, Lewis FD (2011) *J Phys Chem Lett* 2:1432
250. Hariharan M, McCullagh M, Schatz GC, Lewis FD (2010) *J Am Chem Soc* 132:12856
251. Desnoux C, Babu BR, McFrou C, Mayo JUO, Favre A, Wengel J, Clivio P (2008) *J Am Chem Soc* 130:30
252. Ostrowski T, Maurizot JC, Adeline MT, Fourrey JL, Clivio P (2003) *J Org Chem* 68:6502
253. Yuan SA, Zhang WY, Liu LH, Dou YS, Fang WH, Lo GV (2011) *J Phys Chem A* 115:13291