BIOREV (2023) 4: 1–18 [https://doi.org/10.1007/11663_2022_19](https://doi.org/10.1007/11663_2022_19#DOI) © The Author(s), under exclusive license to Springer Nature Switzerland AG 2022 Published online: 28 September 2022

Physio-Metabolic Monitoring via Breath Employing Real-Time Mass Spectrometry: Importance, Challenges, Potentials, and Pitfalls

Pritam Sukul and Phillip Trefz

Contents

Abstract A tiny fraction of our breath contains trace volatile organics of various chemical classes. Due to their endogenous and/or exogenous origins, these volatiles can denominate many intrinsic and extrinsic effects. Exhaled volatile profiles are super dynamic in nature and their expressions may change from seconds to years. Exhaled volatile concentrations largely depend on normal or abnormal fluctuations in physiological and metabolic attributes. Minute or pronounced alterations in cardiorespiratory and other bronchopulmonary gas-exchange parameters due to simple changes in respiratory patterns, routes, and rhythms, posture, expiratory/ inspiratory flow, and upper-airway resistance can immediately affect volatile

P. Sukul (\boxtimes) and P. Trefz

Rostock Medical Breath Research Analytics and Technologies (ROMBAT), Department of Anaesthesiology and Intensive Care, University Medicine Rostock, Rostock, Germany e-mail: Pritam.Sukul@uni-rostock.de

profiles. Similarly, the subject's age, gender, sexual orientation, metabolic state or status, diet, nutrition, therapy, lifestyle habits and habitats, menstrual phases, contraception, pregnancy, menopause, as well as any acute or chronic condition and comorbidities may cause transient or long-lasting differences in breath compositions. Applications of real-time mass spectrometric techniques along with alveolar sampling enabled us to frame fast occurring and continuous changes under diverse physio-metabolic conditions. Physio-metabolic conditions affected breath components more pronouncedly than the differential expression proposed as disease biomarkers in the literature. Investigations of such regulating factors helped us to develop the present state of the art for clinical breath sampling and analysis. Besides, assessments of ventilation and hemodynamics driven changes in exhaled volatiles have depicted potential for physio-metabolic monitoring. Longitudinal personalized analysis of breath profiles may offer unconventional path toward pathophysiological and therapeutic monitoring.

Keywords Pathophysiology · Biomarkers · Breath analysis · Metabolism · Monitoring · Omics · Physiology · Sampling · Standardization · Volatile organic compounds (VOCs)

Abbreviations

1 Introduction

A very tiny fraction $\left(\langle 1\% \right)$ of our breath contains hundreds of volatile organic compounds (VOCs). These compounds belong to various substance classes and the tracible concentrations of most of these VOCs range between parts per billion and parts per trillion by volume (ppbV–pptV) levels [[1\]](#page-12-2). Many of these volatiles are exogenous – meaning that those are accumulated from our habits, habitats, diet, lifestyle, therapy, etc. For instance, if we smoke a cigarette, we will exhale considerable amounts of acetonitrile and furan [[2,](#page-13-0) [3\]](#page-13-1). Visiting a fuel station will source benzene and toluene, whereas entry to a hospital may increase breath isopropanol and formaldehyde concentrations. If we drink orange juice, limonene will become abundant in our breath and a visit to the forest will do the same for alpha-pinene [\[4](#page-13-2)]. On the other hand, some other VOCs are potentially endogenous – meaning that those are produced within us via various physiological, metabolic, biochemical, systemic microbial, and/or pathophysiological processes at the cellular/organ levels [\[5](#page-13-3), [6\]](#page-13-4). For example, acetone is known to originate from cellular and/or hepatic glycolysis and lipolysis [\[7](#page-13-5)]. Ammonia, dimethyl, and trimethylamine are linked to protein catabolism [\[8](#page-13-6), [9\]](#page-13-7). Organosulfur such as dimethyl sulfide, methanethiol, and butanethiol are produced by anerobic methylation by the systemic microbiota of our lower gut [[8,](#page-13-6) [10\]](#page-13-8). Nonetheless, some substances such as isopropanol, ethanol, acetic acid, acetaldehyde, and acrolein have mixed origins – means that they are sourced within and without. Irrespective of the origin (after being produced in vivo or being accumulated/stored from outside), all VOCs are transported via blood to our lungs, and thereafter, they are released during the bronchopulmonary gas-exchange process and thereby are found in breath. Therefore, profiling of these substances may offer non-invasive, rapid, repeatable, and beyond conventional insights into various systemic phenomena, events, or status [\[11](#page-13-9)].

While largely abundant breath gases, e.g., oxygen (O_2) and carbon dioxide (CO_2) , are conventionally applied in human medicine for diagnosis and monitoring of certain conditions, nitric oxide (NO) is the only trace gas, which is vividly investigated for point-of-care (PoC) applicability $[12-14]$ $[12-14]$ $[12-14]$ $[12-14]$. As NO is produced within the proximal airways and inflammation facilitates its production, the fractioned exhaled NO (FeNO) is well attributed to allergic asthma [[13,](#page-13-12) [15](#page-13-13)]. Nonetheless, the scenario is substantially different in case of other trace VOCs. Same substances were proposed in independent studies as biomarker for different diseases $[15-17]$ $[15-17]$ $[15-17]$ $[15-17]$ $[15-17]$. Despite many efforts to find, propose, or establish unique volatile (profiles or patterns) as diseaseor event-specific biomarkers in different studies, none of those could pass the independent validation tests [[18\]](#page-13-15). Consequently, the trace volatiles could not enter into routine clinical practice yet.

In fact, almost all the VOCs are present in everyone's breath, and if we simply measure or compare the exhaled compositions of diseased patients with that of an age- and gender-matched healthy cohort, it is very unlikely to come across any unique VOC profile or pattern [\[19](#page-13-16), [20\]](#page-13-17). After decades of breath analysis, the focus has shifted toward the detection of changes in exhaled VOC concentrations rather than expecting a unique marker. Meanwhile, a series of systematic investigations of immediate, transient, and/or persistent physio-metabolic effects on VOC profiles (mainly from healthy human subjects) helped us to realize the extremely dynamic nature of breath volatiles $[21-25]$ $[21-25]$ $[21-25]$ $[21-25]$. Changes observed in these physio-metabolic studies turned out to be even more pronounced than those published as biomarkers for diagnosis (even for early detection!) in many cross-sectional (healthy vs. sick) studies. Real-time mapping of such dynamic nature under various conditions along with the available knowledge and fundamental understanding of human physiology, metabolism, and analytical chemistry are the indispensable prerequisites to eventually translate VOCs into clinical applications [[5\]](#page-13-3).

2 The Dynamic Nature of Exhaled VOCs

Exhaled VOC concentrations are affected via immediate or precedent extrinsic and intrinsic factors such as environment exposure, diet and lifestyle habits, healthy physiological (respiratory, hemodynamic) and metabolic attributes, systemic microbial activity, as well as any health condition and/or therapy [[25](#page-14-0)–[27\]](#page-14-1). Putative breath markers not only differ interindividually but also may change instantly or over time within the same individual. Even if we succeed to rule out and exclude various extrinsic factors and/or pathological effects, simple and daily life fluctuations in normal and healthy physiology (ventilation and hemodynamics) and/or metabolism may alter our VOC profiles significantly. Besides relying upon our physio-metabolic and health status, such changes and differences are also closely related to the potential origins, physicochemical characters, exhalation kinetics, and compartmental distributions of these VOCs. Thus, while thinking of the dynamic nature of the breath volatiles, a fundamental question does arise. How dynamic are these VOCs or how long does it take to change concentrations?

Well, the actual dynamic timeframe for VOC concentrations ranges from seconds to years.

VOC concentrations can change within seconds simply due to changes in our normal breathing patterns. For example, if we hold our breath for a few seconds, sudden and profound changes are observed after the breath holding phase [\[21](#page-13-18), [28\]](#page-14-2). Such changes are substance specific and depend on the origin and physicochemical characters such as solubility, volatility, and blood-gas partition coefficients of the VOCs. Concentrations of substances with low aqueous solubility, e.g., isoprene, benzene, furan, and acetonitrile, will increase significantly due to perfusion-limited accumulation during breath holding. In contrast to that, compounds like acetone will remain almost constant due to high solubility – that will allow it to get absorbed within the surrounding lung tissue/compartments. On the other hand, oral microbiota-originated VOCs, e.g., hydrogen sulfide, will decrease via washout due to increased respiratory rate (physiological compensation) after breath holding. The physiological effects due to breath holding of 10–60 s by healthy adults are neutralized to baseline within 8–10 breaths.

VOC concentrations will change within minutes if we just switch our body positions [[22\]](#page-13-19). For instance, if you are reading this book by sitting on a chair, and you suddenly switch to supine posture, your cardiac output and pulmonary perfusion will change immediately resulting in increased exhalation of isoprene and similar substances, whereas compounds like acetone and alike compounds will remain unaffected by hemodynamic effects.

VOC concentrations may reflect metabolic changes taking place within hours or throughout the day [\[29](#page-14-3), [30\]](#page-14-4). If we continuously measure breath VOCs from early morning till evening, systemic changes will be observed on VOC exhalations. Intake of standard breakfast and lunch will cause systematic postprandial metabolic adaptation via hyperglycemia and corresponding oxidative stress, which will be reflected in the time profiles of substances such as acetone, 2-propanol, pentanal, dimethyl sulfide, and isoprene.

Changes in VOC concentrations are reported to occur within days, weeks, and also throughout the month. While looking at the natural menstrual cycle in pre-menopausal adult women, exhaled VOC concentrations largely depend on different phases of the monthly endocrine regulation [[31\]](#page-14-5). Endogenous substances such as ammonia, isoprene, acetone, and dimethyl sulfide closely mirror many wellknown effects of the female sex hormones on various metabolic pathways [[32](#page-14-6)– [35\]](#page-14-7). Natural interplay between estrogen and progesterone levels at the period, follicular, ovulation, and luteal phases is reflected distinctly on VOC profiles. Such changes differed significantly in adults undertaking daily oral contraceptive pills (i.e., comprised of supplementary female sex hormones). Similarly, longitudinal changes in VOC concentrations take place throughout pregnancy [[36,](#page-14-8) [37\]](#page-14-9). In pregnant women, various physio-metabolic effects such as gestational endocrine changes, embryonic development, fetal oxygen and nutrient demand-driven increase in cardiac output and respiratory rate, increased cholesterol biosynthesis, and altered breathing pattern via diaphragm upliftment and physiological hyperventilation cause progressive changes in VOC exhalations [\[37](#page-14-9)–[40](#page-14-10)].

If we look into the healthy aging process, exhaled expressions of many endogenous and exogenous VOCs differ significantly, based on our biological age. Recent analysis of breath VOCs from a large cohort of healthy females aged between 7 and 80 years has depicted substance-specific changes in breath composition. Breath concentrations of endogenous aldehydes, alcohols, organosulfur, short-chain fatty acids, alkene, ketones, and exogenous nitriles, aromatics, and terpenes have indicated physio-metabolic milieu between endocrine homeostasis, oxidative stress, gut and pulmonary microbial diversity/activity, energy metabolism, and lifestyle habits [\[41](#page-14-11)–[45](#page-14-12)].

Due to such dynamic nature, it is utterly critical to trace and translate the actual pathophysiological information from breath VOC expressions – as those are often overridden by the everlasting physio-metabolic effects. Continuous breath-resolved profiling of VOC concentrations under different physio-metabolic conditions may pave the path for framing the complex behavioral dynamics and exhalation kinetics of the potential VOC biomarkers in real time.

3 Mass Spectrometric Methods for Real-Time Breath Analysis

Despite the fact that gas-chromatography and mass spectrometry (GC-MS) has been applied for many years as the gold standard for trace gas analysis $[46, 47]$ $[46, 47]$ $[46, 47]$ $[46, 47]$ at low ppbV–pptV levels, punctual measurements could not provide the actual insight into the dynamic nature of breath VOCs. Further, unavoidable confounding factors related to offline mass spectrometry such as preconcentration steps, sample

collection (e.g., blowing into bags, mixed breath phases), sample storage time/ conditions, and analysis time remained as everlasting challenges. Inception of end-tidal $CO₂$ controlled manual or automated breath sampling in glass syringes or in micro-extraction-based traps (e.g., needle-trap microextraction) enabled researchers to collect the alveolar fractions of VOCs, which represents the actual systemic/blood concentrations [[48,](#page-15-2) [49](#page-15-3)]. Nevertheless, fast occurring changes (within seconds or a minute) remained untraceable via offline MS methods.

Development and application of real-time MS techniques, e.g., selected ion flowtube (SIFT)-MS, proton transfer reaction (PTR)-quadrupole-MS, and PTR-time of flight (ToF)-MS [\[50](#page-15-4)], along with online end-tidal/alveolar sampling have eventually overcome various confounding influences $[51–53]$ $[51–53]$ $[51–53]$ $[51–53]$. In principle, a SIFT and a PTR use alike ionization principles based on various primary/reagent ions such as hydronium (H₃O⁺), NO⁺, or O₂⁺. Both of the instruments allow switching between reagent ions according to diverse analytical requirements. Given the fact that most of the breath VOCs belong to the relatively lower mass range $(<500$ Da) and have higher proton affinity than water, in the field of breath analysis, soft ionization via H_3O^+ ion is desirable to have minimal fragmentation [[21](#page-13-18), [23\]](#page-14-13).

PTR-ToF-MS, H_3O^+ ions are produced via cathode discharge on pure (99.99%) water vapor. After production they are pulsed to the next chamber (drift tube) where the proton transfer reaction takes place. The breath sample is introduced to this chamber to react with H_3O^+ ions. Based on the proton affinity, VOCs react with H_3O^+ ions and get protonated (VOC + $H_3O^+ \rightarrow$ VOCH⁺ + H₂O). After that, protonated VOCs are detected via a quadrupole-MS or a ToF-MS according to their mass/charge ratio. Introduction of a ToF allows us to achieve mass resolution of 1,000–4,500 m/Δm that can assign volatiles upon their measured mass and corresponding sum formula with high precision as well as enable isobaric separation of VOC masses [[54](#page-15-7)–[56\]](#page-15-8). Unlike GC-MS, no sample preparation, preconcentration steps, and storage are required. Application of constant inlet flows in side-stream mode (in order to avoid interference to the mainstream of breathing) can uniformly introduce samples to the drift tube and measure at high time resolution in milliseconds (ms). For instance, studies have demonstrated application of 200 ms in clinical environment to simultaneously measure rapid changes of VOC concentrations in the ambient air and in the exhaled breath of healthy subjects or ventilated patients [\[24](#page-14-14), [57](#page-15-9), [58](#page-15-10)]. Here, in every 200 ms a data point was recorded and, on each data point, all protonated VOCs were measured as per mass/charge ratio. Thus, the assignment of VOCs at the exhaled alveolar plateau is also possible in real time. Besides continuous measurements, assignment of inspiratory and end-tidal breath phases (via customized data processing algorithms) helps to date the alveolar fraction of VOCs in a breath-resolved manner [[21,](#page-13-18) [54](#page-15-7)]. For instance, signal intensity of an endogenous and blood-borne VOC (e.g., acetone or isoprene – abundant in exhalation) can be used to denominate the expiratory and inspiratory phases of breath. Based on the area/mass range of interest, mass scale can be recalibrated in desired time intervals. For clinical breath studies, 21.0226 Th $(H₃O⁺$ -isotope), 29.9980 Th (NO⁺), and 59.049 Th (protonated acetone) can be used for mass scale calibration because of their natural abundances in expired and inspired air [[52\]](#page-15-11).

Secondary electrospray ionization-mass spectrometry (SESI-MS) has enabled ambient ionization via nano-electrospray-driven positive (protonated) and negative (deprotonated) ions that collide with sample analytes within the gas phase [\[59](#page-15-12)]. Detection of relatively large, semi-volatile, and even nonvolatile molecules is plausible by integrating high-resolution mass spectrometers such as Orbitrap-MS [\[60](#page-15-13)]. Nevertheless, the identification of substances with higher molecular weights is extremely challenging – especially where internal standards are not available for such mass ranges [\[61](#page-15-14)]. SESI is at an early stage and offers vivid scope for further optimizations and advancements in order to bridge other downstream omics (e.g., metabolomics, proteomics, and lipidomics) with volatolomics and exhaled breathomics.

In contrast to real-time MS techniques for nontargeted screening of VOCs, simpler methods such as electronic noses (eNOSE) or differential ion mobility spectrometry (DMS/IMS) are suitable for targeted approaches [\[62](#page-15-15)]. Artificial olfaction is often conducted via chemical, nano-optical sensors as well as via customized laser-based spectroscopy methods. Despite chemical and nano-optical methods being relatively cheap, easy to use, adaptable (e.g., integration of certain gas sensing arrays as per analytical requirements), and PoC applicable [\[63](#page-15-16)], they suffer from many demerits. These methods do not allow an unequivocal substance identification due to limited selectivity and specificity but offer promising perspectives for PoC breath tests, once marker substances are defined. Susceptibility to matrix effects, e.g., humidity, temperature, and complex sample compositions, is also an important disadvantage for eNOSEs and IMS.

4 Physiological and Metabolic Effects on Breath **Biomarkers**

Soon as it was realized that "magic bullet" biomarkers may not exist and pursuing VOC concentration changes under various pathophysiological conditions is more important and realistic than looking for unique biomarkers, the research focus was imposed on framing the factors that are affecting VOC concentrations. Gradually it is realized that "we are our actual challenge/problem." Being human, our own physiology and metabolism affect our VOC profiles more critically than other external factors.

After being produced or stored in vivo, VOCs undergo various metabolic cascades (regulated by our enzyme systems at cellular, systemic microbial, and organ levels, e.g., liver, gut, and muscle) and larger hydrocarbons often break down into smaller molecules. Thereafter, VOCs are carried by blood and pass through other body compartments where they are distributed/redistributed further. For instance, lipophilic substances are absorbed within the fatty compartments [[64\]](#page-15-17). The lung is our blood–gas interface and the alveolar gas-exchange process is largely denominated by pulmonary ventilation-perfusion (VQ) mechanism and the

distributions of blood flow and air in lung compartments. Consequently, VOC exchange is closely relying on the pulmonary ventilation/perfusion (V/Q) ratios, i.e., primarily regulated by cardiac output and minute ventilation [\[26](#page-14-15), [65](#page-15-18)–[67](#page-16-0)]. After being released in alveolar air, VOCs are further distributed/redistributed within alveolar compartments (due to collateral ventilation between fused alveoli), undergo dilution via airway dead space and substances (e.g., NO and acetone) originating from airway epithelium, and are taken up via extra-alveolar exchange [[24,](#page-14-14) [67](#page-16-0)]. Mechanisms such as pre-alveolar absorption and post-alveolar revalorization are also playing a crucial role in VOC modifications. Therefore, it is important to achieve a steady state of physio-metabolic interplay, while reproducible breath samples can be collected under minimal and systematic influence from subject's own physiology and metabolism.

Minute muscle movements during sleep or vigorous muscle activity under exercise are well known to reflect physio-metabolic effects on breath VOC profiles in real time [[68](#page-16-1)–[71\]](#page-16-2). The anerobic threshold under exhaustive exercise (e.g., by following step-wise and incremental ramp protocols) can be determined by means of VOC-based modeling of lactate threshold and ventilatory threshold. Besides the changes during movements, VOC profiles are also affected by normal physiometabolic effects at rest.

In addition to the aforementioned effects from altered breathing patterns (e.g., breath holding) and/or posture, simple changes in breathing route during breath sampling can also cause substance-specific changes in breath composition at rest. For instance, if you are breathing in and out via mouth and suddenly (even unconsciously) switch to nasal breathing, immediate changes will take place in exhaled VOC concentrations [[25\]](#page-14-0). Substances originating from the nasal cavity bacteria (e.g., methyl-propyl sulfide) will occur and substances originating from the oral cavity (e.g., H2S and allyl-methyl sulfide) will immediately reduce in concentrations. Effects will be also seen on substances regulated by ventilation and hemodynamics. For example, switching from nasal to oral breathing will significantly reduce isoprene concentrations. Isoprene is negatively correlated to minute ventilation. Therefore, bypassing the nasal cavity dead space (i.e., 70–80 ml) will increase the minute ventilation at oral breathing and thereby will reduce isoprene exhalation.

Similarly, if we simply blow our breath into bags or canisters via small straws (i.e., <1 cm of diameter), the uncontrolled upper-airway resistance from the small breathing orifice will immediately affect our exhaled constituents [[72,](#page-16-3) [73](#page-16-4)]. Reduction of the breathing mouthpiece diameter has shown substance-specific effects. Such effects depend largely on the breathing resistance-driven changes in respiratory and hemodynamic parameters [[72\]](#page-16-3). Alveolar eliminations of VOCs with relatively higher volatility are increased due to airway resistance-driven negative intrathoracic pressure (at inspiration), which instantly alters the alveolar diffusion gradient and respiratory mechanics.

Spontaneously breathing human subjects (even if healthy) start to hyperventilate once they are asked to breath normally via a mouthpiece or mask [\[74](#page-16-5)]. Application of paced breathing (i.e., metronome-controlled via sound beats or via visual guide) can be used for breath sampling in order to keep subjects within the normal respiratory rate of 10–14 breaths/min. Researchers observed that the intra- and interindividual ventilatory variations in exhaled VOCs increase significantly during paced breathing. On the other hand, switching between spontaneous and paced breathing causes immediate changes in exhaled concentration and variations of endogenous and blood-borne VOCs [\[75](#page-16-6)]. Such changes depended on minute ventilation and $CO₂$ exhalation. Any conscious and voluntary effort of breathing induces autonomic control and momentarily overwrites the natural automatic control of breathing by our respiratory center. Paced breathing, therefore, induces autonomic function that hampers the normal inspiratory: expiratory (I:E) ratio of \sim 1:2 and increases the minute ventilation. Switching to spontaneous breathing gradually resumes automatic control of breathing and thereby neutralizes the minute ventilation and associated ventilatory variations in $CO₂$ and VOC exhalations.

Moreover, simple changes in exhalation time and expiratory flow may cause profound effects on VOC exhalations [\[24](#page-14-14), [76\]](#page-16-7). For instance, if we normally expire our expiratory reserve volume (i.e., maximum exhalation), substances like isoprene, furan, dimethyl sulfide, and allyl-methyl sulfide will increase immediately by mirroring the end-tidal $CO₂$ profile. This happens mainly due to the change in alveolar slope of exhalation and increased blood-gas contact time which facilitate collateral exchange of gases and VOCs (with low aqueous solubility) between the alveolar compartments [[24\]](#page-14-14). On the other hand, if we perform a forced expiration, those VOCs will decrease instantly due to dilution effects, whereas other substances like acetone will increase significantly as the decelerating flow of exhalation may induce bronchial contribution of such VOC with aqueous miscibility and high bloodgas partition coefficient.

Alongside the above-described immediate and transient effects during sampling, long-term effects and differences due to subject's age, gender, sexual aerosol and orientation, menstrual cycle, pregnancy, menopause, and circadian metabolic rhythms are important attributes for comparison of VOC expressions between cohorts [[5,](#page-13-3) [31,](#page-14-5) [77](#page-16-8), [78](#page-16-9)]. Such factors increase the overall heterogenicity and randomness in breath data and call for more fundamental investigations to address physio-metabolic crosstalk with VOC exhalations. As physio-metabolic effects are everlasting and unavoidable during breath sampling, a basic understanding of those beyond analytical effects is extremely important for interpreting observations pragmatically.

Therefore, no matter how sophisticated and high end the analytical instrument we may use, if we cannot collect a standardized breath sample, the obvious physiometabolic effects at the time of sampling may induce unsupervised effects that are sufficient to mislead our clinical interpretations. During the last decade, the importance of standardization of breath sampling came into focus and several taskforces were formed by the International Association of Breath Research (IABR) to address the relevant factors and state of the art for clinical breath sampling and analysis [\[20](#page-13-17), [79\]](#page-16-10). The following section will briefly summarize the lessons learned during the efforts for standardization including the dos and don'ts.

5 Standardization of Real-Time Sampling for Breath Analysis

Around 20 years ago, alveolar sampling came into consideration in order to represent systemic/blood concentrations of VOCs. Thus, confounding effects from mixedalveolar (i.e., including anatomical dead space of airways, trachea, and mouth/nasal cavity) sampling were minimized significantly reduced. Certainly, the case will be different if airways are of potential interest of a study – such as in obstructive and restrictive tracheobronchial conditions.

During the last 10 years, continuous real-time monitoring of VOCs under various breathing maneuvers at rest have enabled us to knock out the key factors that are essential to collect reproducible breath samples with minimal physio-metabolic influences. Reliable sampling at rest in consciously breathing humans should consider the following crucial aspects:

- Avoid muscle movements: During sampling, subjects should maintain relaxed body postures (e.g., sitting or supine) without having any unnecessary muscle movement [[22\]](#page-13-19). These mainly include voluntary or unmindful movements of limbs and other body parts [[68\]](#page-16-1). Even minute and/or involuntary muscle movements must be recorded for data interpretations.
- Alveolar/end-tidal sampling: As indicated earlier, if airways are not of the study interest, breath sampling must extract the alveolar phase of the breath [\[64](#page-15-17), [80](#page-16-11)]. In case of continuous breath-resolved measurements, exhaled alveolar and inspiratory phases should be determined during data analysis for valid interpretations of systematic VOC concentrations.
- Subjects breathing patterns: Breathing pattern must remain as spontaneous, constant, and normal as possible during sampling [\[21](#page-13-18)]. That means parameters such as respiratory rate, minute ventilation, respiratory flow, and I:E ratio should remain constant throughout the sampling phase. Subjects should not perform prolonged (slow breathing with deeper inhalation and exhalation) or forced expiration or increased respiratory rate [[24\]](#page-14-14). Unusual breathing pattern-driven physiological hyperventilation and cardiorespiratory fluctuations must be avoided to attain steady state of breathing. Prolongation of unusual respiratory rate-driven physiological hyper- and hypoventilation is well known to cause respiratory acidosis and alkalosis, which leads to change in plasma acid–base balance (pH) and affects VOC exhalation [\[81](#page-16-12), [82](#page-16-13)].
- Subject's posture: A particular posture (e.g., sitting or lying on back) must be maintained in order to avoid effects from hemodynamic fluctuations and pulmonary distribution of ventilation and blood flow [\[22](#page-13-19), [83](#page-16-14)]. If the breath from a sick patient is sampled while he or she was at supine position, during follow-up the same supine posture must be maintained (even if the patient is recovered and can sit on a chair) for valid comparisons of pathophysiological effects beyond normal physiological noise (i.e., posture-driven differences). This is also true for

cross-sectional comparisons between healthy vs. sick cohorts. Sampling should be executed at the same posture in both cohorts.

- Subject's breathing route: Either oral or nasal breathing must be maintained without any unconscious switching between both [\[25](#page-14-0)]. This is to avoid ventilatory fluctuations and dead space ventilation and to avoid sudden contributions from oral or nasal cavity flora.
- Applied (instrumental/analytical) upper-airway resistance against breathing: Any obstruction to the mainstream of breathing must be avoided. In order to overcome unsupervised upper-airway resistance-driven change in pulmonary diffusion gradients and respiratory mechanics, smaller breathing mouthpiece should not be used. The diameter of the breathing mouthpiece should range between 1.5 and 2.0 cm [\[72](#page-16-3)]. In case of unavoidable infection safety mandates (e.g., for SARS-CoV-2 and similarly contagious pathogens), mainstream viral/ bacterial filters are applied to stop respiratory viral/bacterial transmission to room air [\[84](#page-16-15)]. In such cases, effects due to instrumental resistance must be accounted for while evaluating breath data.
- Precedent effects from wearing face masks: Researchers have demonstrated pronounced side effects of wearing medical face masks (e.g., COVID-19 protective surgical and FFP2 masks) on respiratory-hemodynamic parameters and exhaled VOC concentrations, at rest. Physiological effects from precedent mask wearing may cause significant hyperventilation (especially in older adults, aged >60 years) as a respiratory compensation process [[85\]](#page-17-0). As such effects may last for minutes to hours based on subject's age and/or health condition, observed breath compositions must account for mask-related effects (where relevant). It is reasonable to allow such subjects to sit without mask for at least 15–30 min prior to breath sampling, in order to minimize precedent physiological effects.
- Subject's ventilatory variations: Respiratory rhythms must be meticulously controlled in order to sample breath with minimal ventilatory variations. A recent study has reported that if at least a minute of paced breathing is applied (with fixed respiratory rate of $10-12$ /min) and then switch to spontaneous breathing, ventilatory variations tend to reduce significantly and attain a steady state after the 2 min of spontaneous breathing and third minute is suitable for collecting/ considering reproducible sample without physiological fluctuations [[75\]](#page-16-6).

Besides the above-indicated sampling conditions, analysis and interpretation of exhaled VOC markers should incorporate effects from acute ambient conditions and also due to long-term effects from subject's personal attributes. These are mainly associated with subject's age, gender, diet, habits, environment, and lifestyle [\[5](#page-13-3)]. Studies have proposed that application of fasting or certain standard diet prior to breath sampling may reduce effects from food intake [\[86](#page-17-1)–[88](#page-17-2)]. Nevertheless, such pre-selection is far from the real-life situation and any screening scenario or nontargeted approach [[89\]](#page-17-3). Furthermore, fasting may cause hypoglycemic adaptation and metabolic compensation effects that may induce inseparable effects onto the volatile metabolites [[29,](#page-14-3) [90\]](#page-17-4). Therefore, it is rather reasonable to carefully consider the attributes from diet, lifestyle habits (e.g., smoking, drinking, nutrition

supplement, and oral contraception), therapy, acute or chronic condition/comorbidity, and living environment as questionnaires during recruitment and retrospectively account for any suspected effects. Most importantly, simultaneous measurements of VOC concentrations in the inspiratory ambient air are extremely important to rule out acute effects.

6 Applications and Potentials of Physio-Metabolic Monitoring in Breath Analysis

Despite the fact that breath research is still in its infancy, the above-mentioned knowhows reflect a steady development toward a state of the art for clinical breath sampling. Nevertheless, the list is rather exhaustive and a lot more effort and time toward standardization of unequivocal confounders related to breath analysis has to be invested.

In principle, a disease/pathophysiological condition is nothing but a disturbed physio-metabolic state. Exhaled VOC profiles may provide rapid information on in vivo physiological or metabolic processes as the time span between marker production and exhalation of VOCs is short. Available knowledge of physiology and metabolism must be translated into disease-driven pathophysiology, for realizing effects on breath compositions. Assessments of physio-metabolic interplay can bridge the gap between our analytical and clinical expertise. Therefore, real-time breathomics holds great promise toward non-invasive monitoring of physiology, metabolism, diseases, and therapy. Screening of a large number of population (e.g., at COVID-19 test center) is also feasible via real-time breathomics. Physiometabolic and pathobiological effects induced by SARS-CoV-2 and other respiratory pathogens are well addressed in recent studies in hundreds of subjects [\[89](#page-17-3)]. Framing of systemic physio-metabolic effects also helped to optimize experimental setups and methods for safe breath analysis and patient monitoring under high safety conditions/mandated at this very time of the global pandemic [\[84](#page-16-15)].

As real-time breath analysis can rapidly deliver results directly at the point of care it is especially attractive for personalized monitoring in patients. No risk is imposed on the patient even if the analysis is done frequently or continuously. Physiometabolic monitoring can be used to follow up substances that were administered to the patient, such as volatile or intravenous anesthetics [\[26](#page-14-15), [47,](#page-15-1) [91\]](#page-17-5). Moreover, VOCs enabled continuous monitoring and immediate recognition of therapeutic efforts in intensive care unit patients [[57,](#page-15-9) [58](#page-15-10)]. Individual monitoring of selected breath VOCs facilitates recognition of metabolic transition without any delay. These findings encourage more research with respect to therapeutic monitoring, longitudinal studies, and follow-up of patients. Besides there is large scope for metabolic monitoring of aging, related life events, and health conditions such as menopause, oxidative stress, endocrine changes, and energy homeostasis. Monitoring of VOC changes during menstrual cycle and pregnancy may reflect phases of healthy natural

cycles, gestation, or any complications based on continuous changes in exhalation profiles [\[31](#page-14-5), [36](#page-14-8)]. Similarly, physio-metabolic monitoring is applicable to physical fitness tests, exercise training, and various applications in sports science/medicine [\[70](#page-16-16)]. Non-invasive assessment of anaerobic threshold, exercise capacity, combined diet, and isotopically labeled substrate interventions are of significant interest. As the putative endogenous origin of most of the VOCs is largely debatable/uncertain and a recent study has even disqualified the long-believed metabolic origin of the second most abundant VOC (i.e., isoprene) in our breath [[36](#page-14-8)], physio-metabolic monitoring under labeled substrate intervention may offer unique insights into the downstream denominators of VOCs and thereby indicate the way to their true systemic sources.

7 Conclusions and Perspective

The dynamic nature of breath VOCs offers a complex but comprehensive spectrum of immediate, transient, and chronic aspects. As breath biomarkers may provide unique and immediate physio-metabolic information on the whole-body status, new insights into normal and pathological processes may be achieved. A fundamental understanding of substance's origins, physicochemical properties, and potential regulating factors such as physiology, metabolism, microbiome, nutrition, lifestyle, and pre-exposure is essential to perceive the VOC expressions case or individual wise. If such knowledge is integrated with state-of-the-art advances in sampling and analytical techniques, observed changes or difference in VOC concentrations may translate actual effects from pathobiological and clinical conditions.

Surprisingly, normal physiology and metabolism-driven changes and variations in VOC concentrations observed in follow-up measurements (where subjects were used as his or her own control) were more pronounced than those reported as unique biomarkers in many published cross-sectional studies. Therefore, cross-sectional comparisons between healthy and ill subjects, in relation to screening or early detection of diseases via breath analysis, are far from our current abilities. In perspective, longitudinal assessments of ventilation and hemodynamics driven changes in breath compositions have depicted excellent potential for physiometabolic monitoring. Continuous and personalized analysis of breath profiles may serve as an unconventional window for monitoring disease progression and response to therapy that could become a cornerstone toward individualized medicine and therapy.

References

1. de Lacy CB, Amann A, Al-Kateb H, Flynn C, Filipiak W, Khalid T et al (2014) A review of the volatiles from the healthy human body. J Breath Res 8:014001

- 2. McKee HC, Rhoades JW, Campbell J, Gross AL (1962) Acetonitrile in body fluids related to smoking. Public Health Rep 77:553–554
- 3. Papaefstathiou E, Stylianou M, Andreou C, Agapiou A (2020) Breath analysis of smokers, non-smokers, and e-cigarette users. J Chromatogr B 1160:122349
- 4. Filipiak W, Ruzsanyi V, Mochalski P, Filipiak A, Bajtarevic A, Ager C et al (2012) Dependence of exhaled breath composition on exogenous factors, smoking habits and exposure to air pollutants. J Breath Res. [zitiert 29. January 2016];6. Verfügbar unter: [http://www.ncbi.nlm.](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3863686/) [nih.gov/pmc/articles/PMC3863686/](http://www.ncbi.nlm.nih.gov/pmc/articles/PMC3863686/)
- 5. Sukul P, Grzegorzewski S, Broderius C, Trefz P, Mittlmeier T, Fischer D-C et al (2022) Physiological and metabolic effects of healthy female aging on exhaled breath biomarkers. iScience 25:103739
- 6. Kai M, Haustein M, Molina F, Petri A, Scholz B, Piechulla B (2009) Bacterial volatiles and their action potential. Appl Microbiol Biotechnol 81:1001–1012
- 7. Kalapos MP (2003) On the mammalian acetone metabolism: from chemistry to clinical implications. Biochim Biophys Acta 1621:122–139
- 8. Pugliese G, Trefz P, Brock B, Schubert JK, Miekisch W (2019) Extending PTR based breath analysis to real-time monitoring of reactive volatile organic compounds. Anal R Soc Chem 144: 7359–7367
- 9. Spacek LA, Strzepka A, Saha S, Kotula J, Gelb J, Guilmain S et al (2018) Repeated measures of blood and breath ammonia in response to control, moderate and high protein dose in healthy men. Sci Rep 8:2554
- 10. Tangerman A (2009) Measurement and biological significance of the volatile sulfur compounds hydrogen sulfide, methanethiol and dimethyl sulfide in various biological matrices. J Chromatogr B 877:3366–3377
- 11. Miekisch W, Schubert JK, Noeldge-Schomburg GFE (2004) Diagnostic potential of breath analysis – focus on volatile organic compounds. Clin Chim Acta 347:25–39
- 12. Alving K, Weitzberg E, Lundberg JM (1993) Increased amount of nitric oxide in exhaled air of asthmatics. Eur Respir J 6:1368–1370
- 13. Ricciardolo FLM, Sterk PJ, Gaston B, Folkerts G (2004) Nitric oxide in health and disease of the respiratory system. Physiol Rev 84:731–765
- 14. American Thoracic Society, European Respiratory Society (2005) ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide. Am J Respir Crit Care Med 171:912–930
- 15. Barnes PJ, Dweik RA, Gelb AF, Gibson PG, George SC, Grasemann H et al (2010) Exhaled nitric oxide in pulmonary diseases: a comprehensive review. Chest 138:682–692
- 16. Nakhleh MK, Amal H, Jeries R, Broza YY, Aboud M, Gharra A et al (2017) Diagnosis and classification of 17 diseases from 1404 subjects via pattern analysis of exhaled molecules. ACS Nano 11:112–125
- 17. Amann A, Miekisch W, Schubert J, Buszewski B, Ligor T, Jezierski T et al (2014) Analysis of exhaled breath for disease detection. Annu Rev Anal Chem (Palo Alto, Calif) 7:455–482
- 18. Amann A, de Lacy Costello B, Miekisch W, Schubert J, Buszewski B, Pleil J et al (2014) The human volatilome: volatile organic compounds (VOCs) in exhaled breath, skin emanations, urine, feces and saliva. J Breath Res 8:034001
- 19. Miekisch W, Schubert JK, Vagts DA, Geiger K (2001) Analysis of volatile disease markers in blood. Clin Chem 47:1053–1060
- 20. Miekisch W, Herbig J, Schubert JK (2012) Data interpretation in breath biomarker research: pitfalls and directions. J Breath Res 6:036007
- 21. Sukul P, Trefz P, Schubert JK, Miekisch W (2014) Immediate effects of breath holding maneuvers onto composition of exhaled breath. J Breath Res 8:037102
- 22. Sukul P, Trefz P, Kamysek S, Schubert JK, Miekisch W (2015) Instant effects of changing body positions on compositions of exhaled breath. J Breath Res 9:047105
- 23. Trefz P, Schmidt M, Oertel P, Obermeier J, Brock B, Kamysek S et al (2013) Continuous real time breath gas monitoring in the clinical environment by proton-transfer-reaction-time-offlight-mass spectrometry. Anal Chem 85:10321–10329
- 24. Sukul P, Schubert JK, Oertel P, Kamysek S, Taunk K, Trefz P et al (2016) FEV manoeuvre induced changes in breath VOC compositions: an unconventional view on lung function tests. Sci Rep 6:28029
- 25. Sukul P, Oertel P, Kamysek S, Trefz P (2017) Oral or nasal breathing? Real-time effects of switching sampling route onto exhaled VOC concentrations. J Breath Res 11:027101
- 26. Kamysek S, Fuchs P, Schwoebel H, Roesner JP, Kischkel S, Wolter K et al (2011) Drug detection in breath: effects of pulmonary blood flow and cardiac output on propofol exhalation. Anal Bioanal Chem 401:2093–2102
- 27. Trefz P, Kamysek S, Fuchs P, Sukul P, Schubert JK, Miekisch W (2017) Drug detection in breath: non-invasive assessment of illicit or pharmaceutical drugs. J Breath Res 11:024001
- 28. Lärstad MAE, Torén K, Bake B, Olin A-C (2007) Determination of ethane, pentane and isoprene in exhaled air – effects of breath-holding, flow rate and purified air. Acta Physiol (Oxf) 189:87–98
- 29. Trefz P, Schmidt SC, Sukul P, Schubert JK, Miekisch W, Fischer D-C (2019) Non-invasive assessment of metabolic adaptation in paediatric patients suffering from type 1 diabetes mellitus. J Clin Med. [zitiert 18. Juli 2020];8. Verfügbar unter: [https://www.ncbi.nlm.nih.gov/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6912469/) [pmc/articles/PMC6912469/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6912469/)
- 30. Fink H, Maihöfer T, Bender J, Schulat J (2022) Indole as a new tentative marker in exhaled breath for non-invasive blood glucose monitoring of diabetic subjects. J Breath Res 16
- 31. Sukul P, Schubert JK, Trefz P, Miekisch W (2018) Natural menstrual rhythm and oral contraception diversely affect exhaled breath compositions. Sci Rep 8:10838
- 32. Hawkins SM, Matzuk MM (2008) Menstrual cycle: basic biology. Ann N Y Acad Sci 1135:10– 18
- 33. Mumford SL, Dasharathy S, Pollack AZ, Schisterman EF (2011) Variations in lipid levels according to menstrual cycle phase: clinical implications. Clin Lipidol 6:225–234
- 34. Wilcox AJ, Dunson D, Baird DD (2000) The timing of the "fertile window" in the menstrual cycle: day specific estimates from a prospective study. BMJ 321:1259–1262
- 35. National Institutes of Health (NIH) (2015) Women's cholesterol levels vary with phase of menstrual cycle. [zitiert 11. April 2018]. Verfügbar unter: [https://www.nih.gov/news-events/](https://www.nih.gov/news-events/news-releases/womens-cholesterol-levels-vary-phase-menstrual-cycle) [news-releases/womens-cholesterol-levels-vary-phase-menstrual-cycle](https://www.nih.gov/news-events/news-releases/womens-cholesterol-levels-vary-phase-menstrual-cycle)
- 36. Sukul P, Richter A, Schubert JK, Miekisch W (2021) Deficiency and absence of endogenous isoprene in adults, disqualified its putative origin. Heliyon 7:e05922
- 37. LoMauro A, Aliverti A (2015) Respiratory physiology of pregnancy. Breathe (Sheff) 11:297– 301
- 38. Bartels Ä, O'Donoghue K (2011) Cholesterol in pregnancy: a review of knowns and unknowns. Obstet Med 4:147–151
- 39. Soma-Pillay P, Catherine N-P, Tolppanen H, Mebazaa A, Tolppanen H, Mebazaa A (2016) Physiological changes in pregnancy. Cardiovasc J Afr 27:89–94
- 40. Lain KY, Catalano PM (2007) Metabolic changes in pregnancy. Clin Obstet Gynecol 50:938– 948
- 41. Ragonnaud E, Biragyn A (2021) Gut microbiota as the key controllers of "healthy" aging of elderly people. Immun Ageing 18:2
- 42. Bosco N, Noti M (2021) The aging gut microbiome and its impact on host immunity. Genes & Immunity Nature Publishing Group, pp 1–15
- 43. Pataky MW, Young WF, Nair KS (2021) Hormonal and metabolic changes of aging and the influence of lifestyle modifications. Mayo Clin Proc 96:788–814
- 44. Boss GR, Seegmiller JE (1981) Age-related physiological changes and their clinical significance. West J Med 135:434–440
- 45. Romano AD, Serviddio G, de Matthaeis A, Bellanti F, Vendemiale G (2010) Oxidative stress and aging. J Nephrol 23(Suppl 15):S29–S36
- 46. Intercomparison of infrared cavity leak-out spectroscopy and gas chromatography-flame ionization for trace analysis of ethane. Anal Chem. [zitiert 17. August 2022]. Verfügbar unter: <https://pubs.acs.org/doi/10.1021/ac702282q>
- 47. Miekisch W, Fuchs P, Kamysek S, Neumann C, Schubert JK (2008) Assessment of propofol concentrations in human breath and blood by means of HS-SPME-GC-MS. Clin Chim Acta 395:32–37
- 48. Miekisch W, Trefz P, Bergmann A, Schubert JK (2014) Microextraction techniques in breath biomarker analysis. Bioanalysis 6:1275–1291
- 49. Trefz P, Rösner L, Hein D, Schubert JK, Miekisch W (2013) Evaluation of needle trap microextraction and automatic alveolar sampling for point-of-care breath analysis. Anal Bioanal Chem 405:3105–3115
- 50. Smith D, Španěl P, Herbig J, Beauchamp J (2014) Mass spectrometry for real-time quantitative breath analysis. J Breath Res 8:027101
- 51. Smith D, Spanel P (2005) Selected ion flow tube mass spectrometry (SIFT-MS) for on-line trace gas analysis. Mass Spectrom Rev 24:661–700
- 52. Herbig J, Müller M, Schallhart S, Titzmann T, Graus M, Hansel A (2009) On-line breath analysis with PTR-TOF. J Breath Res 3:027004
- 53. Lindinger W, Hansel A, Jordan A (1998) On-line monitoring of volatile organic compounds at pptv levels by means of proton-transfer-reaction mass spectrometry (PTR-MS) medical applications, food control and environmental research. Int J Mass Spectrom Ion Process 173:191– 241
- 54. Schwoebel H, Schubert R, Sklorz M, Kischkel S, Zimmermann R, Schubert JK et al (2011) Phase-resolved real-time breath analysis during exercise by means of smart processing of PTR-MS data. Anal Bioanal Chem 401:2079–2091
- 55. Romano A, Hanna GB (2018) Identification and quantification of VOCs by proton transfer reaction time of flight mass spectrometry: an experimental workflow for the optimization of specificity, sensitivity, and accuracy. J Mass Spectrom 53:287–295
- 56. Pugliese G, Piel F, Trefz P, Sulzer P, Schubert JK, Miekisch W (2020) Effects of modular ion-funnel technology onto analysis of breath VOCs by means of real-time mass spectrometry. Anal Bioanal Chem 412:7131–7140
- 57. Löser B, Grabenschröer A, Pugliese G, Sukul P, Trefz P, Schubert JK et al (2020) Changes of exhaled volatile organic compounds in postoperative patients undergoing analgesic treatment: a prospective observational study. Metabolites 10:321
- 58. Brock B, Kamysek S, Silz J, Trefz P, Schubert JK, Miekisch W (2017) Monitoring of breath VOCs and electrical impedance tomography under pulmonary recruitment in mechanically ventilated patients. J Breath Res 11:016005
- 59. Singh KD, del Miguel GV, Gaugg MT, Ibañez AJ, Zenobi R, Kohler M et al (2018) Translating secondary electrospray ionization–high-resolution mass spectrometry to the clinical environment. J Breath Res 12:027113
- 60. Osswald M, Kohlbrenner D, Nowak N, Spörri J, Sinues P, Nieman D et al (2021) Real-time monitoring of metabolism during exercise by exhaled breath. Metabolites 11:856
- 61. Singh KD, Tancev G, Decrue F, Usemann J, Appenzeller R, Barreiro P et al (2019) Standardization procedures for real-time breath analysis by secondary electrospray ionization highresolution mass spectrometry. Anal Bioanal Chem 411:4883–4898
- 62. Pereira JAM, Porto-Figueira P, Taware R, Sukul P, Rapole S, Câmara JS (2020) Unravelling the potential of salivary volatile metabolites in oral diseases. Rev Mol 25:E3098
- 63. Wilson AD, Baietto M (2009) Applications and advances in electronic-nose technologies. Sensors (Basel) 9:5099–5148
- 64. Miekisch W, Kischkel S, Sawacki A, Liebau T, Mieth M, Schubert JK (2008) Impact of sampling procedures on the results of breath analysis. J Breath Res 2:026007
- 65. Lumb AB (2016) Nunn's Appl Respir Physiol 8. Aufl. Italy: Elsevier Ltd.; Verfügbar unter: [https://www.eu.elsevierhealth.com/nunns-applied-respiratory-physiology-9780702062940.](https://www.eu.elsevierhealth.com/nunns-applied-respiratory-physiology-9780702062940.html?dmnum=12449#panel1) html?dmnum=[12449#panel1](https://www.eu.elsevierhealth.com/nunns-applied-respiratory-physiology-9780702062940.html?dmnum=12449#panel1)
- 66. Anderson JC, Babb AL, Hlastala MP (2003) Modeling soluble gas exchange in the airways and alveoli. Ann Biomed Eng 31:1402–1422
- 67. Anderson JC, Hlastala MP (2007) Breath tests and airway gas exchange. Pulm Pharmacol Ther 20:112–117
- 68. King J, Kupferthaler A, Frauscher B, Hackner H, Unterkofler K, Teschl G et al (2012) Measurement of endogenous acetone and isoprene in exhaled breath during sleep. Physiol Meas 33:413–428
- 69. King J, Mochalski P, Kupferthaler A, Unterkofler K, Koc H, Filipiak W et al (2010) Dynamic profiles of volatile organic compounds in exhaled breath as determined by a coupled PTR-MS/ GC-MS study. Physiol Meas 31:1169–1184
- 70. Pugliese G, Trefz P, Weippert M, Pollex J, Bruhn S, Schubert JK et al (2022) Real-time metabolic monitoring under exhaustive exercise and evaluation of ventilatory threshold by breathomics: independent validation of evidence and advances. Front Physiol. [zitiert 18. August 2022];13. Verfügbar unter: [https://www.frontiersin.org/articles/10.3389/fphys.2022.94](https://www.frontiersin.org/articles/10.3389/fphys.2022.946401) [6401](https://www.frontiersin.org/articles/10.3389/fphys.2022.946401)
- 71. Schubert R, Schwoebel H, Mau-Moeller A, Behrens M, Fuchs P, Sklorz M et al (2012) Metabolic monitoring and assessment of anaerobic threshold by means of breath biomarkers. Metabolomics 8:1069–1080
- 72. Sukul P, Schubert JK, Kamysek S, Trefz P, Miekisch W (2017) Applied upper-airway resistance instantly affects breath components: a unique insight into pulmonary medicine. J Breath Res 11:047108
- 73. Holsbeke CV, Vos W, Hamilton M, Claes R, Prime D, Vanhevel F et al (2014) Assessment of the effect of mouthpiece design on upper airway geometry using functional respiratory imaging. Eur Respir J 44:P553
- 74. Cope KA, Watson MT, Foster WM, Sehnert SS, Risby TH (2004) Effects of ventilation on the collection of exhaled breath in humans. J Appl Physiol 96:1371–1379
- 75. Sukul P, Schubert JK, Zanaty K, Trefz P, Sinha A, Kamysek S et al (2020) Exhaled breath compositions under varying respiratory rhythms reflects ventilatory variations: translating breathomics towards respiratory medicine. Sci Rep 10:14109
- 76. Bikov A, Paschalaki K, Kharitonov S, Sinclair RL, Horvath I, Usmani O et al (2012) Expiratory flow rate and breath hold affect exhaled volatile organic compounds (VOC) in healthy subjects. Eur Respir J 40:4305
- 77. Kushch I, Arendacká B, Stolc S, Mochalski P, Filipiak W, Schwarz K et al (2008) Breath isoprene – aspects of normal physiology related to age, gender and cholesterol profile as determined in a proton transfer reaction mass spectrometry study. Clin Chem Lab Med 46: 1011–1018
- 78. Schwarz K, Pizzini A, Arendacká B, Zerlauth K, Filipiak W, Schmid A et al (2009) Breath acetone – aspects of normal physiology related to age and gender as determined in a PTR-MS study. J Breath Res 3:027003
- 79. Beauchamp J (2015) Current sampling and analysis techniques in breath research – results of a task force poll. J Breath Res 9:047107
- 80. Birken T, Schubert J, Miekisch W, Nöldge-Schomburg G (2006) A novel visually CO2 controlled alveolar breath sampling technique. Technol Health Care 14:499–506
- 81. Arbus GS, Hebert LA, Levesque PR, Etsten BE, Schwartz WB (1969) Characterization and clinical application of the significance band for acute respiratory alkalosis. N Engl J Med 280: 117–123
- 82. Cavaliere F, Volpe C, Gargaruti R, Poscia A, Di Donato M, Grieco G et al (2009) Effects of acute hypoventilation and hyperventilation on exhaled carbon monoxide measurement in healthy volunteers. BMC Pulm Med 9:51
- 83. Moreno F, Lyons HA (1961) Effect of body posture on lung volumes. J Appl Physiol 16:27–29
- 84. Sukul P, Trefz P, Schubert J, Miekisch W (2022) Advanced setup for safe breath sampling and patient monitoring under highly infectious conditions in the clinical environment. [zitiert 6. Juli 2022]. Verfügbar unter: <https://www.researchsquare.com/article/rs-1791972/v1>
- 85. Sukul P, Bartels J, Fuchs P, Trefz P, Remy R, Rührmund L et al (2022) Effects of COVID-19 protective face-masks and wearing durations onto respiratory-haemodynamic physiology and exhaled breath constituents. Eur Respir Soc 60:2200009
- 86. De Angelis M, Ferrocino I, Calabrese FM, De Filippis F, Cavallo N, Siragusa S et al (2020) Diet influences the functions of the human intestinal microbiome. Sci Rep 10:4247
- 87. Biagini D, Fusi J, Vezzosi A, Oliveri P, Ghimenti S, Lenzi A et al (2022) Effects of long-term vegan diet on breath composition. J Breath Res 16
- 88. Ajibola OA, Smith D, Španěl P, Ferns GAA (2013) Effects of dietary nutrients on volatile breath metabolites. J Nutr Sci 2:e34
- 89. Remy R, Kemnitz N, Trefz P, Fuchs P, Bartels J, Klemenz A-C et al (2022) Profiling of exhaled volatile organics in the screening scenario of a COVID-19 test center. Rochester, NY [zitiert 20. Juni 2022]. Verfügbar unter: [https://papers.ssrn.com/abstract](https://papers.ssrn.com/abstract=4055638)=4055638
- 90. Trefz P, Obermeier J, Lehbrink R, Schubert JK, Miekisch W, Fischer D-C (2019) Exhaled volatile substances in children suffering from type 1 diabetes mellitus: results from a crosssectional study. Sci Rep 9:15707
- 91. Ghimenti S, Di Francesco F, Onor M, Stiegel MA, Trivella MG, Comite C et al (2013) Postoperative elimination of sevoflurane anesthetic and hexafluoroisopropanol metabolite in exhaled breath: pharmacokinetic models for assessing liver function. J Breath Res 7:036001