

Chapter 14

Energy Homeostasis and the Tumor/Host Interaction: The role of the Brain

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Abstract: The defensive regulation of energy homeostasis by neural and endocrine systems is examined to evaluate the role of the brain in macroenvironmental metabolic control systems that help counterattack the aggressive tumor. Brain homeostatic mechanisms (neural and hormonal) discussed are those linked to metabolic rhythms, food intake and adiposity. Homeostasis is discussed in terms of rheostasis, the low probability of dysregulation, and the potential risks for the defense. The perspective of this review is that many of the metabolic alterations observed in tumor progression may be due to appropriate central homeostatic regulation. Clear deficits in homeostatic regulation during tumor growth have not been unequivocally demonstrated. Yet the brain of the host is clearly under duress due to the tumor. Clarification of homeostatic macroenvironmental regulatory responses may be useful in developing strategies that collaborate with these brain mechanisms. Further analysis of these regulatory systems may identify key changes that ultimately subserve the lethal failure of all host responses. Strategies that protect the brain from the pathological consequences of tumor growth (glucoprivation, oxidative stress, ketogenic diets) and thereby enable a stronger defense are discussed.

Key words: Anorexia, cachexia, circadian rhythms, glucoprivation, ketogenic diet, leptin, melatonin, neuropeptide Y, oxidative stress, suprachiasmatic nucleus

1. ENERGY HOMEOSTASIS

Considerable attention has been focused in recent years on the microenvironmental interaction between the host and tumor, yet relatively little attention has been focused on the macroenvironmental interaction between the host and tumor, particularly with the role of the brain. This is curious because the brain is the site of potent metabolic homeostatic defense systems. Much of what occurs locally is monitored and adjusted centrally. Indeed, disparity in outcomes between *in vitro* and *in vivo* analyses is often due to interactive contributions by the brain.

Homeostasis is regulated through dynamic control mechanisms that defend the host from

perturbations that threaten the milieu intérieur (1-4). These energy-related phenotypes are central to the evolved organization of the higher organism that may host a tumorous neoplasm. The macroenvironmental management of homeostasis occurs in the brain where there are regulatory mechanisms for the complex control of food intake (5-11), adiposity (6-12), glucose (13-14), and metabolic rhythms (14-22) that rely in part on peripheral information derived from hormonal and neural input and in part on the local environment within the brain (e.g., glucose sensing). During the progression of aggressive tumor growth these defended parameters in the host are often altered to manifest a metabolic syndrome that features unusual rhythms (23-26), glucose intolerance (27-28), insulin

resistance (27-28), increased gluconeogenesis (29), increased free fatty acid turnover, oxidation and clearance (30), cachexia (31-32) and anorexia (31-32).

The perspective of this review is that many of the metabolic alterations observed in tumor progression may be due to central homeostatic regulation that is often appropriate, sometimes disturbed, but seldom dysregulated. Whether the response is appropriate or disturbed has been difficult to establish. For example, food restriction inhibits tumor growth and the host response to the tumor is to inhibit food intake (anorexia/cachexia) (33-36). If food restriction is additionally reduced by fasting the tumor grows faster³⁷. Thus the brain seems to have achieved just the right balance of food intake and food restriction to minimize the growth potential of the tumor. Yet there are risks for this strategy due to the dire long-term consequences of anorexia/cachexia. Similarly, the brain and the tumor compete for glucose and the brain can enhance gluconeogenesis to meet its glucose needs.

Clarification of homeostatic macroenvironmental regulatory responses may be useful in developing strategies that collaborate with these brain mechanisms. Moreover, further analysis of these regulatory systems may identify the key events that ultimately subserve the lethal failure of all host responses. For example, homeostatic systems may be misled by or blind to the re-activated embryogenic systems often observed in malignant tumors (e.g., proteolysis-inducing factor (31)). Thus the etiology of lethality in cancer pathology may plausibly be due to tumor factors that disturb, foil, or overwhelm homeostatic regulation, even when that regulation is working appropriately.

Progress has been made in the last decade that furthers our understanding of some key neural circuits utilized for energy homeostasis, in particular the integrated hypothalamic and brainstem circuits for metabolic rhythms, feeding, fuel, adiposity. Key hormonal metabolic signals to the brain (e.g., leptin) have been identified and extensively examined. Cross-talk between the research fields of energy homeostasis and cancer may be fruitful for both disciplines.

Evidence on the role of macroenvironmental regulatory mechanisms residing in the brain for

energy homeostasis as an adaptive and/or maladaptive response by the host to tumor progression will be briefly reviewed in this chapter.

2. HOMEOSTASIS, RHEOSTASIS AND DYSREGULATION

Much of medicine is assisted homeostasis. But parsing malignant metabolism and parceling out the contribution of the host and the tumor can be difficult. The tumor perturbs, the host responds. Most of the metabolic characteristics of malignant metabolism probably reflect the interaction of host and tumor.

The metabolic syndrome that occurs during cancer malignancy has some features of starvation, infection and late term pregnancy³³ as well as some features of another metabolic syndrome, diabetes. In each analogy however there are significant differences and in the final analysis cancer appears to induce its own metabolic syndrome³⁸. As with diabetes the contribution of host homeostatic mechanisms to each of these metabolic alterations has been difficult to assess. And certainly there are significant differences in etiology for each metabolic syndrome. But just as certainly there are common features and a limited set of control system weapons to work with in the defense of energy homeostasis. To understand that set of control systems is one aim of those who study energy homeostasis.

Altered homeostatic parameters alone do not constitute dysregulatory pathology. Indeed it may be more appropriate to reconsider homeostasis as rheostasis where the defended level changes, but the defense of the level remains intact. For example, as Mrosovsky (4) has eloquently articulated, hibernation, starvation, infection, and pregnancy are physiological conditions where appropriate regulatory changes occur. The organism might run a fever, not eat, conserve energy and/or reduce activity. Each of these adaptations may become pathological but each represents a risk that favors survival. Obesity, for example, derived as a phenotype from adaptive selection that favored survival during sustained food deprivation (39). The fat survived. Hypertension may be required to adequately provide circulation especially when there

is about a mile of additional capillary length for each pound of fat. Hyperglycemia may assure glucose delivery during insulin insufficiency (40). Fever fights infection but may itself become pathological. These examples suggest that pathology does not require dysregulation.

Indeed true dysregulation is probably rare and would best be characterized by no defense at all. The host seldom misinterprets or is blind to the homeostatic insult, especially in chronic conditions. One important example may help clarify this point. Hypoglycemia is an acute risk in insulin treated diabetes and can be fatal. The phenomenon is called hypoglycaemia-associated autonomic failure (HAAF) and is often referred to as hypoglycemia unawareness (41). In HAAF the regulatory responses transduced in the medullary brainstem (e.g., glucoprivic feeding, stimulation of epinephrine, glucocorticoid and glucagon secretion, enhanced gluconeogenesis) do not occur (42). When studied in rats HAAF was produced by an acute prior induction of hypoglycemia during which the normal regulatory responses occurred, followed in a few hours by a second bout of hypoglycemia. During the second bout the normal regulatory responses did not occur. Hypoglycemia unawareness in these rats, however, did not occur when glucocorticoids were removed by prior adrenalectomy (42). Thus we see that hypoglycemia unawareness may reflect either 1. homeostatic unawareness; 2. a regulated, yet adverse, reaction to a competition between homeostatic responses (acquisition of glucose versus stress induced glucocorticoids); or 3. a regulated, yet adverse, reaction to components of the same homeostatic response (acquisition of glucose) in which glucocorticoids may play a direct role.

The lesson then is that dysregulation can not be assumed to occur. A demonstration of a changed response, or even an inappropriate non-response, is not necessarily a demonstration of dysregulation. In addition, integrated responses to complex challenges can create competition between regulatory subsystems, which may result in a lack of metabolic stability⁴. When not extreme such a competition is usually resolved with metabolic stability. For example, rats will leave a warm environment to acquire food in a cold environment (-15° C) but they

will eat that food as quickly as possible (43). The suggestion from this perspective on homeostasis during cancer is that the identification, and experimental manipulation, of environmental impediments (e.g., glucoprivation, cyclic AMP deficits, and oxidative stress) on function is teleologically necessary in order to understand the brain-derived defense of the energy realm. In addition, experiments on the direct impact of tumor-derived molecules (e.g., proteolysis-inducing factor and Zn-alpha (2)-glycoprotein (31) on brain outcomes are needed, as are further analysis of brain energy systems under the influence of a malignant tumor.

Clearly, without further study, the complex perturbations caused by cancer that render the host inadequately defended will continue to a lethal outcome. Assisted homeostasis through a better understanding of the role of the brain in cancer may be critical in tipping the counterbalance of this defense in favor of the host.

3. ENERGY HOMEOSTASIS CIRCUITS IN THE BRAIN

Neural and neuroendocrine mechanisms for the defense of energy homeostasis have been extensively elaborated on during the last decade (5-12, 14-22, 44-48). These reviews have identified the regulatory signal to the brain from adipose tissue as the cytokine leptin, a hormone derived primarily from adipose tissue. Leptin levels in the circulation reflect the level of adiposity. The administration of exogenous leptin to rodents potently reduces body weight (49-55). This reduction in body weight is almost entirely due to loss of stored lipid fuels and is not due to loss of protein (56-58). This pattern differs from starvation, during which both lipid and protein are lost (59-60).

Leptin's central actions are critical for its effects on feeding and body weight. First, hypophagia can be produced by central administration of leptin at doses several fold lower than equally-effective systemic doses, and this effect is achieved without any increase in the circulating levels of leptin (56, 61), either by efflux from the cerebral spinal fluid into venous blood or by enhancing peripheral leptin

secretion. Second, central administration of leptin is sufficient to reverse the obesity of ob/ob (*Lepr^{ob}/Lepr^{ob}*) mice lacking the gene for leptin (50). Although peripheral administration of leptin also causes anorexia and weight loss, these actions of leptin may be mediated centrally since leptin is able to gain access to the brain from the circulation. Finally, selective deletion of neuronal leptin receptors abolished leptin's anorexic effect and resulted in obesity in mice (62); and epigenetic expression of leptin receptors only in the brain reversed the obese phenotype of the db/db mouse (*Lepr^{db}/Lepr^{db}*), which is without leptin receptors due to an autosomal recessive mutation (63).

The related cytokines ciliary neurotrophic factor (CNTF), interleukin-6 (IL-6) and leukemia inhibitory factor (LIF), which signal through the highly related glycoprotein-130 (gp-130) JAK/STAT receptor circuitry, also dramatically decrease fat pads to a similar degree when chronically given centrally into the ventricles (64-68). These members of the interleukin (IL)-6-receptor family of cytokines appear to redundantly activate a brain system that controls fat mass to the extent that rodent models live in good health without adipose tissue for most of their adult life (68). Redundancy in signalling is a common feature of this system (69-71).

Yet these cytokines that employ the gp-130 signal transducing subunit also interact with specificity (69). Oncostatin M (OSM) regulated IL-6 expression in glia, but LIF did not (72). OSM did not produce anorexia, but LIF and interleukin-11 (IL-11) did produce anorexia in short-term feeding tests (68).

Numerous neuropeptide effectors in the brain that are implicated in the control of feeding occur downstream of leptin and have been extensively studied (5-12, 14-22, 45-48, 73, 74). Orexigenic neuropeptides inhibited by leptin include neuropeptide Y (NPY) (75-76), galanin (77), melanin concentrating hormone (MCH) (75) and agouti-related peptide (AGRP) (75-76). Anorexigenic neuropeptides stimulated by leptin include corticotropin releasing hormone (77-78), glucagon-like peptide 1 (GLP-1) (79), alpha-melanocyte stimulating hormone (80), neurotensin (75, 81) and cocaine- and amphetamine-regulated

transcript (CART) (82). Neurotrophins such as brain-derived neurotrophin factor (BDNF) (83, 84), CNTF (85-88) and the neurotrophin regulated VGF (90-91) have emerged as important in this matrix of neural signals necessary for energy homeostasis, as has leukemia inhibitory factor (LIF) (85, 89). The specific role of each signalling agent is beginning to be clarified (83). Thus there is an extensive brain circuitry that subserves the control of food intake (e.g., anorexia, hypogeusia, hyposmia, nausea, satiation, aversion, and appetite), metabolism (e.g., gluconeogenesis, free fatty acid release, compartmentalization of fuels), adiposity (fat cachexia) and perhaps muscle mass (muscle cachexia).

4. WHY IS BRAIN CIRCUITRY FOR ENERGY HOMEOSTASIS OF INTEREST IN CANCER?

In the sections that follow six particular topics will be briefly considered with each section providing only a partial, and necessarily incomplete, suggested answer to this question. More complex brain functions that affect the host/tumor interaction but involve external events and behaviors that transcend energy homeostasis (e.g., biopsychosocial oncology (93) or psychoneuroimmunology (94), such as social stress (95), social defeat (96) or social dominance (97) will not be discussed because these topics are beyond the scope of this review.

4.1 Regulatory feeding responses appear intact during tumor progression

Although anorexia, cachexia and numerous metabolic disturbances are manifest in many cancer models, the normal controls of feeding are still substantially intact (33-34). That is, the anorexia can be returned to normophagia under some experimental conditions. In a classic review Seoras Morrison asked his version of the central question of cachexia/anorexia: "Why the host does not respond to change in need with change in intake?" (33). Why doesn't the energy-depleted host simply invoke hyperphagia? For example, cold exposure (5° C vs.

24° C) for two days normalized food intake in rats with tumor-induced anorexia (98). And the hyperphagia induced by insulin-induced glucoprivation was intact in Walker 256 carcinoma-bearing Sprague-Dawley rats (34). Interestingly, obese mice maintained on a food for which they had developed a conditioned taste aversion lived longer with B16 melanoma than those maintained on a food that was not aversive (99). These examples indicate that the metabolic changes observed during tumor progression may be due to a regulatory response to the tumor and are not due to incapacity of the regulatory system.

Moreover cachexia and anorexia do not require the presence of the tumor (34). As with parabiosis (100), direct transfer of blood from a cancer-bearing host to a non-cancer-bearing host results in cachexia/anorexia without a tumor (101). And, indeed, removal of the tumor, except in the late stages of cachexia, reverses both the anorexia and cachexia (34, 102). Thus the cachectic/anorectic response interaction by the host to the tumor does not require the tumor and does not require structural alterations to the host that can not be rapidly adjusted.

The answer to Seoras Morrison's central question, therefore, may be that the host has already responded appropriately to the tumor with just the right amount of food restriction.

4.2 There is an altered energy environment within the brain during tumor progression

Alteration in homeostatic function may occur because of alterations in the metabolic environment within the brain, as a distant function of tumor growth. Tumor growth involves vascular leakiness within the tumor induced by a unique tumor angiogenesis that produces a necrotic condition characterized by hypoxia, elevated glycolysis even under aerobic conditions, high glucose turnover, glucose deprivation, adverse acidity and increased interstitial fluid pressure (103-104). These elements of tumor growth alter the brain environment by reducing glucose availability and increasing oxidative stress.

The high glucose turnover of the aggressive tumor reduced glucose utilization by the brain and increased utilization of lactate and 3-hydroxybutyrate (ketones) (105). Indeed it is probable that the proximal cause of death with rapidly growing non-metastasizing tumors is often acute hypoglycemia (106). Survival for sarcoma-bearing hypoglycemic mice was not enhanced by administration of glucagon, but drinking of glucose by food-deprived mice did enhance survival. And adrenalectomy shortened survival time (106).

The homeostatic responses to glucoprivation are transduced in the ventrolateral medulla of the brainstem (107-108) and include hyperphagia, gluconeogenesis and thus the sympathoadrenal stimulation of epinephrine, glucagon and glucocorticoids. The need to provide nutrients to the brain quickly is paramount during acute hypoglycemia.

One way to reduce the brain's need for glucose, and the adverse consequences of oxidative stress, is to substitute ketones for glucose as fuel for the brain (109). Several reports (110-116) have examined the mostly positive effects of a ketogenic diet based on medium-chain triglycerides on non-brain tumor cachexia and tumor growth but not on survival rates or brain function. Glycerol supplementation may also be helpful (117). And recently a direct role for glucose supplementation has been beneficial (118). Additional studies with diets that may support brain function during cancer are warranted. From the perspective of this review the support of brain function should be considered a different objective than ameliorating anorexia/cachexia.

Oxidative stress was increased within the brain, with highest levels of increased stress observed in the hypothalamus, during the growth of the Walker-256 tumor, due to the elevated rate of oxygen consumption, the high level of endogenous polyunsaturated fats (119) and iron (120). TNF α has been implicated in this increased stress (122). Specific sites within the hypothalamus are rich in TNF- α receptors, especially the paraventricular nucleus, the supraoptic nucleus and the arcuate nucleus (123). Central catecholamines are particularly susceptible to oxidative stress (123) and critical participants in energy homeostasis and especially in glucose regulation (107-108).

Oxidative injury to catecholamines has been reported in the locus coeruleus (124) and ventrolateral medulla (VLM) (125). The VLM is a critical site for transduction of numerous homeostatic responses, including those stimulated by hypoxia and glucoprivation. Melatonin functions as a neuroprotectant during oxidative stress to catecholamines in part due to upregulation of glial derived neurotrophic factor (GDNF) (124). Oxidative stress induced by the tumor stimulated glycose-6-phosphate dehydrogenase activity, pentose phosphate pathway activity and elevated flux of substrates, while brain mitochondrial activity was inhibited (119).

Thus in these two ways, and probably others (e.g., the aberrant presence in the brain of tumor-derived molecules), the brain environment is altered by the presence of the tumor.

4.3 Altered expression of neuropeptides and monoamines in brain circuitry for energy homeostasis during tumor malignancy

The hypothalamus is an important site for integrating metabolic information and regulating energy homeostasis and metabolism (5-9, 12, 14-15, 20, 73-78, 80, 82-83). Alterations in signalling within the hypothalamus during tumor progression have usually been reviewed in the context of cancer anorexia (31, 126-137). For example, there are reportedly decreased NPY fibers in anorectic tumor-bearing rats in the parvocellular region of the paraventricular nucleus, as well as in the supraoptic, suprachiasmatic and arcuate nuclei (138). Most of the NPY fibers in the hypothalamus are located in these nuclei. This decrease in NPY fibers tends to support these authors' hypothesis that the anorexia induced by the tumor alters the structure of the neuronal signalling pathway for food intake, of which NPY is one important part. Indeed, when NPY was acutely delivered directly into the perifornical hypothalamus in MCA sarcoma-bearing rats before the expression of anorexia hyperphagia in a four-hour food intake test was at first induced comparable to non-tumor bearing controls (139). While elevated food intake in tumor-bearing rats

who received acute hypothalamic injections of NPY continued to occur during the ongoing experiment over the next two weeks and five injections, the elevation was greatly attenuated in tumor bearing rats compared to controls without the MCA sarcoma that received hypothalamic injections of NPY. A similar refractory response to perifornical NPY was observed with chronic minipump infusion (139). This refractory feeding response induced by perifornical NPY in tumor-bearing rats may reflect refractory adenylate cyclase AMP formation (131).

The effect of a tumor on NPY expression and signalling in the hypothalamus is far from clear, which may reflect differences between tumors, in the brain site examined, experimental designs, rodent strains, multiple roles for NPY and/or other factors. Some studies report decreased expression of NPY (137, 141-143), other studies report increased expression (144-148) and yet other studies report normal expression (149-150). A more complete critical review of the NPY feeding system and its alteration by a malignant tumor is overdue but beyond the scope of this review. However, further exploration of the NPY feeding system, and related systems within the prevue of brain regulated energy homeostasis, seems warranted. Indeed, the direct examination in tumor bearing rodents of the effects of infusion into the brain of either energy related molecules such as this orexigenic neuropeptide NPY, or of tumor-related molecules into non-tumor bearing hosts, seems to be a neglected area of cancer research with only a few examples (139, 151-152), even when experiments with proinflammatory cytokines (66, 67, 126, 131) are included. Perhaps to refocus research on brain pathology engendered by a malignant tumor and on brain responses to that tumor would reenergize research in this neglected area.

In addition to NPY, there are expression changes that occur for other neuropeptides and monoamines critical to neural communication in the circuitry for energy homeostasis during the progression of malignant tumors. Alterations in brain serotonin (141, 143), dopamine (141, 143), MCH (145), orexin (145) and interleukin-1 beta (150) have been reported.

Which subset of homeostatic controls these changes reflect (anorexia, cachexia, the regulation of

adiposity or muscle, metabolites) is unclear. But clearly the brain is reacting to the presence of the tumor with alterations in this critical neural network that serves energy homeostasis.

4.4 Destruction of the medial basal hypothalamus with gold thioglucose, or agouti blockade of the melanocortin-4 receptor, accelerates lipid wasting and demise in mice

In 1971 Liebelt and co-workers induced hyperphagia and obesity in two strains of mice by destruction of the medial basal hypothalamus with gold thioglucose (GTG) (153). They predicted that the hyperphagia induced by this treatment would override the hypophagia induced by either of two strains of tumor, the CBA 2663 stomach tumor or the C57Bl sarcoma. To their surprise a profound anorexia and cachexia were accelerated in these obese mice, who died by day twenty of tumor growth. The lean mice lived until the end of the experiment, at day thirty of tumor growth, and never expressed anorexia or cachexia. Tumor growth was not accelerated in the obese mice and the tumors remained small but lethal. Carcass weight loss was primarily lipid in the GTG tumor-bearing mice. A parallel experiment utilized the agouti yellow (Ay/a) obese mice, which have a dominant mutation of the agouti gene so that there is a blockade of the melanocortin-4 receptor and insensitivity to α MSH (154), produced the same pattern to tumor bearing.

There are several factors in common to these two mouse models of obesity. The ability of central leptin to reduce body weight, adiposity and food intake is attenuated in both the GTG mouse and the agouti yellow obese mouse (155-156). In both models there is insensitivity to POMC and α MSH157. And both obesity models require the neurotrophin-induced polypeptide VGF (158), which is also important in energy homeostasis. Thus the circuitry of energy homeostasis in the hypothalamus is blocked with catastrophic consequence in tumor bearing mice. Further work in cancer with these models is warranted, but clearly both the circuitry and the site of action are important in the host defense against tumor lethality.

4.5 Suprachiasmatic nucleus-induced rhythms inhibit tumor growth

Time-keeping is important for energy homeostasis. Metabolism is processed with rhythm (14-22). Food is anticipated with activity (22). Insulin is secreted in anticipation of food (20). There are circadian rhythms for metabolites such as glucose (14), and metabolic hormones such as ghrelin and leptin (159). Food (21), or glucose, but not lipid, reset the biological clock (160). Disrupted sleep also disrupts these rhythms (161).

The suprachiasmatic nucleus (SCN) in the hypothalamus serves as a site for photic transduction of the light/dark cycle; as an entrainer for its endogenous clock; and, as a site for rhythm synchronization with non-photoc stimuli (14, 17-20, 162, 163). Ablation of the SCN in rodents removes photic entrained rhythms and accelerates tumor growth (164, 165). In a recent experiment, Filipski and co-workers demonstrated that a simple advance of the light/dark cycle by eight hours every two days accelerated Glasgow osteosarcoma tumor growth (166).

Circadian rhythms are often disrupted in cancer and chronotherapies are currently under investigation for improved management of cancer (24, 26). Sleep disorders are a common feature of cancer (23). Brain derived rhythms inhibit tumor growth and sustain normal sleep. And the tumor may disrupt those rhythms. Thus there is a need for further investigation of the role of circadian rhythms in tumor growth and malignancy.

4.6 The pineal gland and melatonin

The pineal gland is located within the brain and is the source of the hormone melatonin (167-168). Both the pineal gland and melatonin are involved in energy homeostasis (169-170). And both melatonin and the pineal can interact with tumor growth, which has stimulated an interest in the antitumor properties of this hormonal system (171-176).

In one study the removal of the pineal gland stimulated the growth of melanoma (177). And in another study administration of melatonin shortened the survival rate of mice with the Ehrlich ascites

tumor (178). And overexpression of the melatonin MT1 receptor suppressed mammary tumor formation (179).

These examples, supported by correlational studies (171-176), have generated a recent surge of interest in the role of the pineal axis in the treatment of malignant tumors.

5. SUMMARY

The brain is the ultimate defender of energy homeostasis and cancer is the ultimate aggressor. The metabolic syndrome expressed in this terrible struggle is due in part to both a stout defense and a violent aggression. Disentangling this interaction requires further experiment but may prove to be important in the management of malignancy. Fundamental components of the defense are apparent and suggest that the brain is responding appropriately, with roles for particular sites (e.g., the medial basal hypothalamus, the suprachiasmatic nucleus, the pineal gland), that mediate the control of rhythms, food intake, and adiposity. The environment within the brain is altered by the tumor, which may compromise the defense. Analysis of the altered environment (e.g., glucoprivation, oxidative stress, tumor-derived molecules, alterations in neuropeptides and monamines) may lead to an improved defense. And, finally, analysis of the antitumor strategies of this brain-derived defense may lead to assisted homeostasis: interventions that serve to tip the counterbalance in favour of the host.

REFERENCES

- Bernard, C., 1878, *Leçons sur les phénomènes de la vie communs aux animaux et aux végétaux*. J-B Baillière et fils, Paris, France.
- Cannon, W.B., 1929, Organization for physiological homeostasis. *Physiol Rev*, 9:399-431.
- Cannon, W.B., 1939, *The wisdom of the body*. W.W. Norton & Co. Inc., New York, NY, USA.
- Mrosovsky, N., 1990, *Rheostasis: the physiology of change*. : Oxford University Press. New York
- Saper, C.B., Chou, T.C., and Elmquist, J.K. 2002 The need to feed: homeostatic and hedonic control of eating. *Neuron* 36: 199-211.
- Berthoud, H.R. 2002 Multiple neural systems controlling food intake and body weight. *Neurosci Biobehav Rev* 26: 393-428.
- Blevins, J.E., Schwartz, M.W., and Baskin, D.G. 2002 Peptide signals regulating food intake and energy homeostasis. *Can J Physiol Pharmacol* 80: 396-406.
- Horvath, T.L., Diano, S., and Tschop, M. 2004 Brain circuits regulating energy homeostasis. *Neuroscientist* 10: 235-46.
- Flier, J., and Maratos-Flier, E. 2000 Energy homeostasis and body weight. *Curr Biol* 10: R215-7.
- Havel, P.J. 2001 Peripheral signals conveying metabolic information to the brain: short-term and long-term regulation of food intake and energy homeostasis. *Exp Biol Med (Maywood)* 226: 963-77.
- Meier, U., and Gressner, A.M. 2004 Endocrine regulation of energy metabolism: review of pathobiochemical and clinical chemical aspects of leptin, ghrelin, adiponectin, and resistin. *Clin Chem* 50: 1511-25.
- Cowley, M.A., Pronchuk, N., Fan, W., Dinulescu, D.M., Colmers, W.F., and Cone, R.D. 1999 Integration of NPY, AGRP, and melanocortin signals in the hypothalamic paraventricular nucleus: evidence of a cellular basis for the adipostat. *Neuron* 24: 155-63.
- Epstein, A.N., Nicolaidis, S. and Miselis, R. 1975. The glucoprivic control of food intake and the glucostatic theory of feeding behaviour. Pp 146-168 in *Neural integration of physiological mechanisms and behavior*, Mogenson G.J. and Calaresu, F.R., eds. Toronto: University of Toronto Press.
- La Fleur, S.E. 2003 Daily rhythms in glucose metabolism: suprachiasmatic nucleus output to peripheral tissue. *J Neuroendocrinol* 15: 315-22.
- Kreier, F., Kalsbeek, A., Ruiten, M., Yilmaz, A., Romijn, J.A., Sauerwein, H.P., Fliers, E., and Buijs, R.M. 2003 Central nervous determination of food storage--a daily switch from conservation to expenditure: implications for the metabolic syndrome. *Eur J Pharmacol* 480: 51-65.
- Schibler, U., Ripperger, J., and Brown, S.A. 2003 Peripheral circadian oscillators in mammals: time and food. *J Biol Rhythms* 18: 250-60.
- Challet, E., Caldelas, I., Graff, C., and Pevet, P. 2003 Synchronization of the molecular clockwork by light- and food-related cues in mammals. *Biol Chem* 384: 711-9.
- Rutter, J., Reick, M., and McKnight, S.L. 2002 Metabolism and the control of circadian rhythms. *Annu Rev Biochem* 71: 307-31.
- Nagai, K., Nagai, N., Sugahara, K., Nijijima, A., and Nakagawa, H. 1994 Circadian rhythms and energy metabolism with special reference to the

- suprachiasmatic nucleus. *Neurosci Biobehav Rev* 18: 579-84.
20. Strubbe, J.H., and van Dijk, G. 2002 The temporal organization of ingestive behaviour and its interaction with regulation of energy balance. *Neurosci Biobehav Rev* 26: 485-98.
 21. Stephan, F.K. 2002 The "other" circadian system: food as a Zeitgeber. *J Biol Rhythms* 17: 284-92.
 22. Mistlberger, R.E. 1994 Circadian food-anticipatory activity: formal models and physiological mechanisms. *Neurosci Biobehav Rev* 18: 171-95.
 23. Lee, K., Cho, M., Miaskowski, C., and Dodd, M. 2004 Impaired sleep and rhythms in persons with cancer. *Sleep Med Rev* 8: 199-212.
 24. Sephton, S., and Spiegel, D. 2003 Circadian disruption in cancer: a neuroendocrine-immune pathway from stress to disease? *Brain Behav Immun* 17: 321-8.
 25. Fu, L., and Lee, C.C. 2003 The circadian clock: pacemaker and tumour suppressor. *Nat Rev Cancer* 3: 350-61.
 26. Mormont, M.C., and Levi, F. 2003 Cancer chronotherapy: principles, applications, and perspectives. *Cancer* 97: 155-69.
 27. Argiles, J.M., Almendro, V., Busquets, S., and Lopez-Soriano, F.J. 2004 The pharmacological treatment of cachexia. *Curr Drug Targets* 5: 265-77.
 28. Tayek, J.A. 1992 A review of cancer cachexia and abnormal glucose metabolism in humans with cancer. *J Am Coll Nutr* 11: 445-56.
 29. Thompson, M.G, and Palmer, R.M. 1998 Signalling pathways regulating protein turnover in skeletal muscle. *Cell Signal* 10: 1-11.
 30. Baron, A., Migita, T., Tang, D., and Loda, M. 2004 Fatty acid synthase: a metabolic oncogene in prostate cancer? *J Cell Biochem* 91: 47-53.
 31. Tisdale, M.J. 2004 Tumor-host interactions. *J Cell Biochem* 93: 871-7.
 32. van Halteren, H.K., Bongaerts, G.P., and Wagener, D.J. 2003 Cancer cachexia: what is known about its etiology and what should be the current treatment approach? *Anticancer Res* 23: 5111-5.
 33. Morrison, S.D. 1976 Control of food intake in cancer cachexia: a challenge and a tool. *Physiol Behav* 17: 705-714.
 34. Morrison, S.D. 1981 Control of food intake in experimental tumor growth. *Cancer Treat Rep* 65(Suppl 5): 9-14.
 35. Molotkov, A., Satoh, M., and Tohyama, C. 1998 Tumor growth and food intake in interleukin-6 gene knock-out mice. *Cancer Lett* 132: 187-92.
 36. Thompson, C.I., Kreider, J.W., and Margules, D.L. 1984 Food intake during tumor growth: anorexia in genetically obese ob/ob mice and hyperphagia in lean mice. *Physiol Behav* 32: 935-9.
 37. Rofe, A.M., Porter, S.J., Bais, R., and Coyers, R.A.J. 1985 The metabolic response of tumour-bearing mice to fasting. *Br J Cancer* 52: 619-623.
 38. Devereux, D.F., Redgrave, T.G., Loda, M.F., Clowes, G.H. Jr, and Deckers, P.J. 1985 Tumor-associated metabolism in the rat is a unique physiologic entity. *J Surg Res* 38: 149-53.
 39. Coleman, D.L. 1979 Obesity genes: beneficial effects in heterozygous mice. *Science* 203: 663-5.
 40. Cabanac, M. and Russek, M. 1982 Régulation et contrôle en biologie. Les Presses de l'Université Laval, Québec.
 41. Cryer, P.E. 2002 Hypoglycaemia: the limiting factor in the glycaemic management of Type I and Type II diabetes. *Diabetologia* 45: 937-48.
 42. Sanders, N.M., and Ritter, S. 2001 Acute 2DG-induced glucoprivation or dexamethasone abolishes 2DG-induced gluco regulatory responses to subsequent glucoprivation. *Diabetes* 50: 2831-6.
 43. Johnson, K.G., and Cabanac, M. 1982 Homeostatic competition between food intake and temperature regulation in rats. *Physiol Behav* 28: 675-9.
 44. Mrosovsky, N. 1986 Body fat: what is regulated? *Physiol Behav* 38: 407-14.
 45. Cancellato, R., Tounian, A., Poitou, Ch., and Clement, K. 2004 Adiposity signals, genetic and body weight regulation in humans. *Diabetes Metab* 30: 215-27.
 46. Wynne, K., Stanley, S., and Bloom, S. 2004 The gut and regulation of body weight. *J Clin Endocrinol Metab* 89: 2576-82.
 47. Tremblay, A. 2004 Dietary fat and body weight set point. *Nutr Rev* 62: S75-7.
 48. Cone, R.D., Cowley, M.A., Butler, A.A., Fan, W., Marks, D.L., and Low, M.J. 2001 The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obes Relat Metab Disord* 25 Suppl 5: S63-7.
 49. Pelleymounter, M.A., Cullen, M.J., Baker, M.B., Hecht, R., Winters, D., Boone, T., and Collins, F. 1995 Effects of the obese gene product on body weight regulation in ob/ob mice. *Science* 269: 540-543
 50. Campfield, L.A., Smith, F.J., Guisez, Y., Devos, R., and Burn, P. 1995 Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science* 269: 546-549.
 51. Halaas, J.L., Gajiwala, K.S., Maffei, M., Cohen, S.L., Chait, B.T., Rabinowitz, D., Lallone, R.L., Burley, S.K., and Friedman, J.M. 1995 Weight regulation effects of the plasma protein encoding the obese gene. *Science* 269: 543-6.
 52. Matson, C.A., Wiater, M.F., and Weigle, D.S. 1996 Leptin and the regulation of body adiposity: A critical review. *Diabetes Rev* 4: 488-508.
 53. Baile, C.A., Della-Fera, M.A., and Martin, R.J. 2000 Regulation of metabolism and body fat mass by leptin. *Annu Rev Nutr* 20: 105-27.

54. Spiegelman, B.M., and Flier, J.S. 2001 Obesity and the regulation of energy balance. *Cell* 104: 531-543.
55. Friedman, J.M., and Halaas, J.L. 1998 Leptin and the regulation of body weight in mammals. *Nature* 395: 763-70.
56. Azain, M.J., Wang, T., Hulsey, M.G., Qian, H., Hartzell, D.L., and Baile, C.A. 1999 Effects of intracerebroventricularly administered leptin on protein selection in the rat. *Physiol Behav* 66: 537-541.
57. Kaibara, A., Moshyedi, A., Auffenberg, T., Abouhamze, A., Copeland, E.M., III, Kalra, S., and Moldawer, L.L. 1998 Leptin produces anorexia and weight loss without inducing an acute phase response or protein wasting. *Amer J Physiol* 274: R1518-R1525.
58. Chen, Y., and Heiman, M.L. 2000. Chronic leptin administration promotes lipid utilization until fat mass is greatly reduced and preserves lean mass of normal female rats. *Reg Peptides* 92: 113-119.
59. Dallman, M.F., Akana, S.K., Bhatnagar, S., Bell, M.E., Choi, S., Chu, A., Horsley, C., Levin, N., Meijer, O., Soriano, L.R., Strack A.M., and Viau, V. 1999 Starvation: early signals, sensors, and sequelae. *Endocrinology* 140: 4015-4023.
60. Goodman, M.N., and Ruderman, N.B. 1980 Starvation in the rat. I. Effect of age and obesity on organ weights, RNA, DNA, and protein. *Am J Physiol* 239: E269-E276.
61. van Dijk, G., Seeley, R.J., Thiele, T.E., Friedman, M.I., Ji, H., Wilkinson, C.W., Burn, P., Campfield, L.A., Tenenbaum, R., Baskin, D.G., Woods, S.C., and Schwartz, M.W. 1999 Metabolic, gastrointestinal, and CNS neuropeptide effects of brain leptin administration in the rat. *Am J Physiol* 276: R1425-33.
62. Cohen, P., Zhao, C., Cai, X., Montez, J.M., Rohani, S.C. Feinstein, P., Mombaerts, P., and Friedman, J.M. 2001 Selective deletion of leptin receptor in neurons leads to obesity. *J Clin Invest* 108: 1113-21.
63. Kowalski, T.J., Liu, S.M., Leibel, R.L., and Chua, S.C. Jr. 2001 Transgenic complementation of leptin-receptor deficiency. I. Rescue of the obesity/diabetes phenotype of LEPR-null mice expressing a LEPR-B transgene. *Diabetes* 50: 425-35.
64. Duff, E., Li, C.L., Hartzell, D.L., Choi, Y.H., Della-Fera, M.A., and Baile, C.A. 2004 Ciliary neurotrophic factor injected icv induces adipose tissue apoptosis in rats. *Apoptosis* 9: 629-34.
65. Beretta, E., Dhillon, H., Kalra, P.S., and Kalra, S.P. 2002 Central LIF gene therapy suppresses food intake, body weight, serum leptin and insulin for extended periods. *Peptides* 23: 975-84.
66. Jansson, J.O., Wallenius, K., Wernstedt, I., Ohlsson, C., Dickson, S.L., and Wallenius, V. 2003 On the site and mechanism of action of the anti-obesity effects of interleukin-6. *Growth Horm IGF Res* 13 Suppl A: S28-32.
67. Wallenius, K., Wallenius, V., Sunter, D., Dickson, S.L., and Jansson, J.O. 2002 Intracerebroventricular interleukin-6 treatment decreases body fat in rats. *Biochem Biophys Res Commun* 293: 560-5.
68. Plata-Salaman, C.R. 1996 Anorexia induced by activators of the signal transducer gp 130. *Neuroreport* 7: 841-4.
69. Ishihara, K., and Hirano, T. 2002 Molecular basis of the cell specificity of cytokine action. *Biochim Biophys Acta* 1592: 281-96.
70. Taga, T., and Kishimoto, T. 1997 gp130 and the interleukin-6 family of cytokines. *Annu. Rev. Immunol.* 15: 797-819.
71. Heinrich, P.C., Behrmann, I., Haan, S., Hermanns, H.M., Muller-Nowen, G., and Schaper, F. 2003 Principles of interleukin (IL)-6-type cytokine signalling and its regulation. *Biochem J* 374: 1-20.
72. Van Wagoner, N.J., Choi, C., Repovic, P., and Benveniste, E.N. 2000 Oncostatin M regulation of interleukin-6 expression in astrocytes: biphasic regulation involving the mitogen-activated protein kinases ERK1/2 and p38. *J Neurochem* 75: 563-75.
73. Hakansson, M.L., Brown, H., Ghilardi, N., Skoda, R.C., and Meister, B. 1998. Leptin receptor immunoreactivity in chemically defined target neurons of the hypothalamus. *J Neurosci* 18: 559-72.
74. Schwartz, M.W., Seeley, R.J., Campfield, L.A., Burn, P., and Baskin, D.G. 1996 Identification of targets of leptin action in rat hypothalamus. *J Clin Invest* 98: 1101-6.
75. Sahu, A. 1998 Evidence suggesting that galanin (GAL), melanin-concentrating hormone (MCH), neurotensin (NT), proopiomelanocortin (POMC) and neuropeptide Y (NPY) are targets of leptin signaling in the hypothalamus. *Endocrinology* 139: 795-8.
76. Korner, J., Savontaus, E., Chua, S.C. Jr, Leibel, R.L., and Wardlaw, S.L. 2001 Leptin regulation of *Agrp* and *Npy* mRNA in the rat hypothalamus. *J Neuroendocrinol* 13: 959-66.
77. Costa, A., Poma, A., Martignoni, E., Nazppi, G., Ur, E., and Grossman, A. 1997 Stimulation of corticotrophin-releasing hormone release by the obese (ob) gene product, leptin, from hypothalamic explants. *NeuroReport* 8: 1131-34.
78. Uehara, Y., Shimizu, H., Ohtani, K., Sato, N., and Mori, M. 1998 Hypothalamic corticotropin-releasing hormone is a mediator of the anorexigenic effect of leptin. *Diabetes* 47: 890-93.
79. Mercer, J.G., Moar, K.M., Findlay, P.A., Hoggard, N., and Adam, C.L. 1998 Association of leptin receptor (OB-Rb), NPY, and GLP-1 gene expression in the ovine and murine brainstem. *Regul Pept* 75-76: 271-278.

80. Forbes, S., Bui, S., Robinson, B.R., Hochgeschwender, U., and Brennan, M.B. 2001 Integrated control of appetite and fat metabolism by the leptin-proopiomelanocortin pathway. *Proc Natl Acad Sci USA* 98: 4233-4237.
81. Sahu, A., Carraway, R.E., and Wang, Y.P. 2001 Evidence that neurotensin mediates the central effect of leptin on food intake in rat. *Brain Res* 888: 343-347.
82. Kristensen, P., Judge, M.E., Thim, L., Ribel, U., Christjansen, K.N., Wulff, B.S., Clausen, J.T., Jensen, P.B., Madsen, O.D., Vrang, N., Larsen, P.J., and Hastrup, S. 1998 Hypothalamic CART is a new anorectic peptide regulated by leptin. *Nature* 393: 72-76.
83. Pellemounter, M.A., Cullen, M.J., and Wellman, C.L. 1995 Characteristics of BDNF-induced weight loss. *Exp Neurol* 131: 229-38.
84. Kernie, S.G., Liebl, D.J., and Parada, L.F. 2000 BDNF regulates eating behavior and locomotor activity in mice. *EMBO J* 19: 1290-300.
85. Prima, V., Tennant, M., Gorbatyuk, O.S., Muzyczka, N., Scarpace, P.J., and Zolotukhin, S. 2004 Differential modulation of energy balance by leptin, ciliary neurotrophic factor, and leukemia inhibitory factor gene delivery: microarray deoxyribonucleic acid-chip analysis of gene expression. *Endocrinology* 145: 2035-45.
86. Mattson, M.P. 2001 Lose weight STAT: CNTF tops leptin. *Trends Neurosci* 24: 313-4.
87. Duff, E., Li, C.L., Hartzell, D.L., Choi, Y.H., Della-Fera, M.A., and Baile, C.A. 2004 Ciliary neurotrophic factor injected icv induces adipose tissue apoptosis in rats. *Apoptosis* 9: 629-34.
88. Anderson, K.D., Lambert, P.D., Corcoran, T.L., Murray, J.D., Thabet, K.E., Yancopoulos, G.D., and Wiegand, S.J. 2003 Activation of the hypothalamic arcuate nucleus predicts the anorectic actions of ciliary neurotrophic factor and leptin in intact and gold thioglucose-lesioned mice. *J Neuroendocrinol* 15: 649-60.
89. Beretta, E., Dhillon, H., Kalra, P.S., and Kalra, S.P. 2002 Central LIF gene therapy suppresses food intake, body weight, serum leptin and insulin for extended periods. *Peptides* 23: 975-84.
90. Levi, A., Ferri, G.L., Watson, E., Possenti, R., and Salton, S.R. 2004 Processing, distribution, and function of VGF, a neuronal and endocrine peptide precursor. *Cell Mol Neurobiol* 24: 517-33.
91. Salton, S.R. 2003 Neurotrophins, growth-factor-regulated genes and the control of energy balance. *Mt Sinai J Med* 70: 93-100.
92. Leibowitz, S.F., and Wortley, K.E. 2004 Hypothalamic control of energy balance: different peptides, different functions. *Peptides* 25: 473-504.
93. Temoshok, L.R., and Wald, R.L. 2002 Change is complex: rethinking research on psychosocial interventions and cancer. *Integr Cancer Ther* 1: 135-45.
94. Ben-Eliyahu, S. 2003 The promotion of tumor metastasis by surgery and stress: immunological basis and implications for psychoneuroimmunology. *Brain Behav Immun* 17 Suppl 1: S27-36.
95. Strange, K.S., Kerr, L.R., Andrews, H.N., Emerman, J.T., and Weinberg, J. 2000 Psychosocial stressors and mammary tumor growth: an animal model. *Neurotoxicol Teratol*. 2000 Jan-Feb;22(1):89-102.
96. Stefanski, V., and Ben-Eliyahu, S. 1996 Social confrontation and tumor metastasis in rats: Defeat and β -adrenergic mechanisms. *Physiol Behav* 60: 277-282.
97. Temoshock, L., Peeke, H.V.S., and Mehard, C.W. 1988 Individual behaviour differences related to induced tumor growth in the female syrian hamster: two studies. *Int. J. Neurosci* 38: 199-209.
98. Stevenson, J.A., Box, B.M., and Wright, R.B. 1963 The effect of a cold environment on malignant anorexia. *Can J Biochem Physiol* 41: 531-2.
99. Thompson, C.I., Margules, D.L., Kreider, J.W., Boha, S.P., Rejer, R.E. Jr, Quirey, R.A., and Reitz, J.A. 1993 Propensity to form conditioned taste aversions augments anorexia in obese (ob/ob) mice with B16 melanoma. *Behav Neurosci* 107: 786-98.
100. Norton, J.A., Moley, J.F., Green, M.V., Carson, R.E., and Morrison, S.D. 1985 Parabolic transfer of cancer anorexia/cachexia in male rats. *Cancer Res* 45: 5547-52.
101. Illig, K.A., Maronian, N., and Peacock, J.L. 1992 Cancer cachexia is transmissible in plasma *J Surg Res* 52: 353-358.
102. Donovan, H. 1954 Malignant cachexia. *Proc R Soc Med* 47: 27-31.
103. McDonald, D.M., and Choyke, P.L. 2003 Imaging of angiogenesis: from microscope to clinic. *Nat Med* 9: 713-25.
104. Måseide, K., Kalliomäki, T., and Hill, R.P. Microenvironmental Effects on Tumour Progression and Metastasis. Chapter in this book.
105. Mulligan, H.D., and Tisdale, M.J. 1991 Metabolic substrate utilization by tumour and host tissues in cancer cachexia. *Biochem J* 277: 321-6.
106. Svaninger, G., Gelin, J., and Lundholm, K. 1989 The cause of death in non-metastasizing sarcoma-bearing mice. A study with relevance for tumor treatment experiments in mice. *Eur J Cancer Clin Oncol* 25: 1295-302.
107. Ritter, S., Dinh, T.T., and Zhang, Y. 2000 Localization of hindbrain glucoreceptive sites controlling food intake and blood glucose. *Brain Res* 856: 37-47.
108. Ritter, S., Bugarith, K., and Dinh, T.T. 2001 Immunotoxic destruction of distinct catecholamine subgroups produces selective impairment of

- glucoregulatory responses and neuronal activation. *J Comp Neurol* 432: 197-216.
100. Veech, R.L. 2004 The therapeutic implications of ketone bodies: the effects of ketone bodies in pathological conditions: ketosis, ketogenic diet, redox states, insulin resistance, and mitochondrial metabolism. *Prostaglandins Leukot Essent Fatty Acids*. 2004 70: 309-19.
 110. Beck, S.A., and Tisdale, M.J. 1989 Nitrogen excretion in cancer cachexia and its modification by a high fat diet in mice. *Cancer Res* 49: 3800-4.
 111. Tisdale, M.J., Brennan, R.A., and Fearon, K.C. 1987 Reduction of weight loss and tumour size in a cachexia model by a high fat diet. *Br J Cancer* 56: 39-43.
 112. Fearon, K.C., Borland, W., Preston, T., Tisdale, M.J., Shenkin, A., and Calman, K.C. 1988 Cancer cachexia: influence of systemic ketosis on substrate levels and nitrogen metabolism. *Am J Clin Nutr* 47: 42-8.
 113. Fearon, K.C. 1988 Nutritional pharmacology in the treatment of neoplastic disease. *Baillieres Clin Gastroenterol* 2: 941-9.
 114. Tisdale, M.J., and Brennan, R.A. 1988 A comparison of long-chain triglycerides and medium-chain triglycerides on weight loss and tumour size in a cachexia model. *Br J Cancer* 58: 580-3.
 115. Beck, S.A., and Tisdale, M.J. 1989 Effect of insulin on weight loss and tumour growth in a cachexia model. *Br J Cancer* 59: 677-81.
 116. Nebeling, L.C., and Lerner, E. 1995 Implementing a ketogenic diet based on medium-chain triglyceride oil in pediatric patients with cancer. *J Am Diet Assoc* 95: 693-7.
 117. Wang, H.Y., Hochwald, S., Port, J., Harrison, L.E., Ng, B., and Burt, M. 1995 Hypoglycemia with glycerol salvage: a role in anti-neoplastic therapy? *Anticancer Res* 15: 1343-8.
 118. Bozzetti, F., Gavazzi, C., Mariani, L., and Crippa, F. 2004 Glucose-based total parenteral nutrition does not stimulate glucose uptake by humans tumours. *Clin Nutr* 23: 417-21.
 119. Freitas, J.J., Pompeia, C., Miyasaka, C.K., and Curi, R. 2001 Walker-256 tumor growth causes oxidative stress in rat brain. *J Neurochem* 77: 655-63.
 120. Zaleska, M.M., and Floyd, R.A. 1985 Regional lipid peroxidation in rat brain in vitro: possible role of endogenous iron. *Neurochem Res* 10: 397-410.
 121. Theologides, A., Ingersoll-Stroubos, A.M., and Apple, F.S. 1994 TNF-alpha effect on oxygen free radical scavenging and generating enzymes in rat liver. *Biochem Mol Biol Int* 33: 205-10.
 122. Nadeau, S., and Rivest, S. 1999 Effects of circulating tumor necrosis factor on the neuronal activity and expression of the genes encoding the tumor necrosis factor receptors (p55 and p75) in the rat brain: a view from the blood-brain barrier. *Neuroscience* 93: 1449-64.
 123. Smythies, J., and Galzigna, L. 1998 The oxidative metabolism of catecholamines in the brain: a review. *Biochim Biophys Acta* 1380: 159-62.
 124. Chen, K.B., Lin, A.M., and Chiu, T.H. 2003 Oxidative injury to the locus coeruleus of rat brain: neuroprotection by melatonin. *J Pineal Res* 35: 109-17.
 125. Kishi, T., Hirooka, Y., Kimura, Y., Ito, K., Shimokawa, H., and Takeshita, A. 2004 Increased reactive oxygen species in rostral ventrolateral medulla contribute to neural mechanisms of hypertension in stroke-prone spontaneously hypertensive rats. *Circulation* 109: 2357-62.
 126. Ramos, E.J., Suzuki, S., Marks, D., Inui, A., Asakawa, A., and Meguid, M.M. 2004 Cancer anorexia-cachexia syndrome: cytokines and neuropeptides. *Curr Opin Clin Nutr Metab Care* 7: 427-34.
 127. Davis, M.P., Dreicer, R., Walsh, D., Lagman, R., and LeGrand, S.B. 2004 Appetite and cancer-associated anorexia: a review. *J Clin Oncol* 22: 1510-7.
 128. Laviano, A., Russo, M., Freda, F., and Rossi-Fanelli, F. 2002 Neurochemical mechanisms for cancer anorexia. *Nutrition* 18: 100-5.
 129. Inui, A., and Meguid, M.M. 2003 Cachexia and obesity: two sides of one coin? *Curr Opin Clin Nutr Metab Care* 6: 395-9.
 130. Fanelli, F.R. and Laviano, A. Cancer anorexia: a model for the understanding and treatment of secondary anorexia. *Int J Cardiology* 85: 67-72.
 131. Turrin, N.P., Ilyin, S.E., Gayle, D.A., Plata-Salaman, C.R., Ramos, E.J., Laviano, A., Das, U.N., Inui, A., and Meguid, M.M. 2004 Interleukin-1beta system in anorectic catabolic tumor-bearing rats. *Curr Opin Clin Nutr Metab Care* 7: 419-26.
 132. Inui, A. 1999 Cancer anorexia-cachexia syndrome: are neuropeptides the key? *Cancer Res* 59: 4493-501.
 133. Inui, A. 1999 Neuropeptide Y: a key molecule in anorexia and cachexia in wasting disorders? *Mol Med Today* 5: 79-85.
 134. Wisse, B.E., Schwartz M.W., and Cummings, D.E. 2003 Melanocortin signaling and anorexia in chronic disease states. *Ann N Y Acad Sci* 994: 275-81.
 135. Chance, W.T., Balasubramaniam, A., and Fischer, J.E. 1995 Neuropeptide Y and the development of cancer anorexia. *Ann Surg* 221: 579-87.
 136. Norton, J.A., Peacock, J.L., and Morrison, S.D. 1987 Cancer cachexia. *Crit Rev Oncol Hematol* 7: 289-327.
 137. Chance, W. T., Balasubramaniam, A., Sheriff, S., and Fischer, J. E. 1994 Possible role of neuropeptide Y in experimental cancer anorexia. Pp 185-201 in:

- Jacobs, M., ed. Diet and cancer: Markers, prevention and treatment. New York: Plenum Press.
138. Makarenko, I.G., Meguid, M.M., Gatto, L., Chen, C., and Ugrumov, M.V. 2003 Decreased NPY innervation of the hypothalamic nuclei in rats with cancer anorexia. *Brain Res* 961: 100-8.
139. Chance, W.T., Balasubramaniam, A., Thompson, H., Mohapatra, B., Ramo, J., and Fischer, J.E. 1996 Assessment of feeding response of tumor-bearing rats to hypothalamic injection and infusion of neuropeptide Y. *Peptides* 17: 797-801.
140. Chance, W.T., Balasubramaniam, A., Borchers, M., and Fischer, J.E. 1995 Refractory hypothalamic adenylate cyclase in anorectic tumor-bearing rats: implications for NPY-induced feeding. *Brain Res* 691: 180-4.
141. Meguid, M.M., Ramos, E.J., Laviano, A., Varma, M., Sato, T., Chen, C., Qi, Y., and Das, U.N. 2004 Tumor anorexia: effects on neuropeptide Y and monoamines in paraventricular nucleus. *Peptides* 25: 261-6.
142. Chance, W.T., Balasubramaniam, A., Dayal, R., Brown, J., and Fischer, J.E. 1994 Hypothalamic concentration and release of neuropeptide Y into microdialysates is reduced in anorectic tumor-bearing rats. *Life Sci* 54: 1869-74.
143. Ramos, E.J., Suzuki, S., Meguid, M.M., Laviano, A., Sato, T., Chen, C., and Das, U. 2004 Changes in hypothalamic neuropeptide Y and monoaminergic system in tumor-bearing rats: pre- and post-tumor resection and at death. *Surgery* 136: 270-6.
144. McCarthy, H.D., McKibbin, P.E., Perkins, A.V., Linton, E.A., and Williams, G. 1993 Alterations in hypothalamic NPY and CRF in anorectic tumor-bearing rats. *Am J Physiol* 264: E638-43.
145. Nara-ashizawa, N., Tsukada, T., Maruyama, K., Akiyama, Y., Kajimura, N., and Yamaguchi, K. 2001 Response of hypothalamic NPY mRNAs to a negative energy balance is less sensitive in cachectic mice bearing human tumor cells. *Nutr Cancer* 41: 111-8.
146. Nara-ashizawa, N., Tsukada, T., Maruyama, K., Akiyama, Y., Kajimura, N., Nagasaki, K., Iwanaga, T., and Yamaguchi, K. 2001 Hypothalamic appetite-regulating neuropeptide mRNA levels in cachectic nude mice bearing human tumor cells. *Metabolism* 50: 1213-9.
147. Chance, W.T., Sheriff, S., Kasckow, J.W., Regmi, A., and Balasubramaniam, A. 1998 NPY messenger RNA is increased in medial hypothalamus of anorectic tumor-bearing rats. *Regul Pept* 75-76: 347-53.
148. Jensen, P.B., Blume, N., Mikkelsen, J.D., Larsen, P.J., Jensen, H.I., Holst, J.J., and Madsen, O.D. 1998 Transplantable rat glucagonomas cause acute onset of severe anorexia and adipsia despite highly elevated NPY mRNA levels in the hypothalamic arcuate nucleus. *J Clin Invest* 101: 503-10.
149. Bing, C., Taylor, S., Tisdale, M.J., and Williams, G. 2001 Cachexia in MAC16 adenocarcinoma: suppression of hunger despite normal regulation of leptin, insulin and hypothalamic neuropeptide Y. *J Neurochem* 79: 1004-12.
150. Plata-Salaman, C.R., Ilyin, S.E., and Gayle, D. 1998 Brain cytokine mRNAs in anorectic rats bearing prostate adenocarcinoma tumor cells. *Am J Physiol* 275: R566-73.
151. Wisse, B.E., Frayo, R.S., Schwartz, M.W., and Cummings, D.E. 2001 Reversal of cancer anorexia by blockade of central melanocortin receptors in rats. *Endocrinology* 142: 3292-301.
152. Chance, W. T., van Lammeren, F. M., and Fischer, J. E. 1988 Feeding elicited by cholinergic and adrenergic hypothalamic stimulation of anorectic tumor-bearing rats. *Pharmacol. Biochem. Behav* 31: 209-213.
153. Liebelt, R.A., Liebelt, A.G., Johnston, H.M. 1971 Lipid mobilization and food intake in experimentally obese mice bearing transplanted tumors. *Proc Soc Exp Biol Med* 138: 482-90.
154. Boston, B.A. 2001 Pro-opiomelanocortin and weight regulation: from mice to men. *J Pediatr Endocrinol Metab* 14 Suppl 6: 1409-16.
155. Rahmouni, K., Haynes, W.G., Morgan, D.A., and Mark A.L. 2002 Selective resistance to central neural administration of leptin in agouti obese mice. *Hypertension* 39: 486-90.
156. Fei, H., Okano, H.J., Li, C., Lee, G.H., Zhao, C., Darnell, R., and Friedman, J.M. 1997 Anatomic localization of alternatively spliced leptin receptors (Ob-R) in mouse brain and other tissues. *Proc Natl Acad Sci U S A* 94: 7001-5.
157. Anderson, K.D., Lambert, P.D., Corcoran, T.L., Murray, J.D., Thabet, K.E., Yancopoulos, G.D., and Wiegand, S.J. 2003 Activation of the hypothalamic arcuate nucleus predicts the anorectic actions of ciliary neurotrophic factor and leptin in intact and gold thioglucose-lesioned mice. *J Neuroendocrinol* 15: 649-60.
158. Hahm, S., Fekete, C., Mizuno, T.M., Windsor, J., Yan, H., Boozer, C.N., Lee, C., Elmquist, J.K., Lechan, R.M., Mobbs, C.V., and Salton, S.R. 2002 VGF is required for obesity induced by diet, gold thioglucose treatment, and agouti and is differentially regulated in pro-opiomelanocortin- and neuropeptide Y-containing arcuate neurons in response to fasting. *J Neurosci* 22: 6929-38.
159. Kalra, S.P., Bagnasco, M., Otukonyong, E.E., Dube, M.G., and Kalra, P.S. 2003 Rhythmic, reciprocal ghrelin and leptin signaling: new insight in the development of obesity. *Regul Pept* 111: 1-11.

160. Stephan, F.K., and Davidson, A.J. 1998 Glucose, but not fat, phase shifts the feeding-entrained circadian clock. *Physiol Behav* 65: 277-88.
161. Everson, C.A., and Wehr, T.A. 1993. Nutritional and metabolic adaptations to prolonged sleep deprivation in the rat. *Am J Physiol* 264: R376-87.
162. Hastings, M.H., and Herzog, E.D. 2004 Clock genes, oscillators, and cellular networks in the suprachiasmatic nuclei. *J Biol Rhythms* 19: 400-13.
163. Buijs, R.M., van Eden, C.G., Goncharuk, V.D., and Kalsbeek, A. 2003 The biological clock tunes the organs of the body: timing by hormones and the autonomic nervous system. *J Endocrinol* 177: 17-26.
164. Filipski, E., King, V.M., Li, X., Granda, T.G., Mormont, M.C., Claustrat, B., Hastings, M.H., and Levi, F. 2003 Disruption of circadian coordination accelerates malignant growth in mice. *Pathol Biol (Paris)* 51: 216-9.
165. Filipski, E., King, V.M., Li, X., Granda, T.G., Mormont, M.C., Liu, X., Claustrat, B., Hastings, M.H., and Levi, F. 2002 Host circadian clock as a control point in tumor progression. *J Natl Cancer Inst* 94: 690-7.
166. Filipski, E., Delaunay, F., King, V.M., Wu, M.W., Claustrat, B., Grechez-Cassiau, A., Guettier, C., Hastings, M.H., and Francis, L. 2004 Effects of chronic jet lag on tumor progression in mice. *Cancer Res* 64: 7879-85.
167. Csernus, V., and Mess, B. 2003 Biorhythms and pineal gland. *Neuro Endocrinol Lett* 24: 404-11.
168. Simonneaux, V., and Ribelayga, C. 2003 Generation of the melatonin endocrine message in mammals: a review of the complex regulation of melatonin synthesis by norepinephrine, peptides, and other pineal transmitters. *Pharmacol Rev* 55: 325-95.
169. Morgan, P.J., Ross, A.W., Mercer, J.G., and Barrett, P. 2003 Photoperiodic programming of body weight through the neuroendocrine hypothalamus. *J Endocrinol* 177: 27-34.
170. Barrenetxe, J., Delagrange, P., and Martinez, J.A. 2004 Physiological and metabolic functions of melatonin. *J Physiol Biochem* 60: 61-72.
171. Sanchez-Barcelo, E.J., Cos, S., Fernandez, R., and Mediavilla, M.D. 2003 Melatonin and mammary cancer: a short review. *Endocr Relat Cancer* 10: 153-9.
172. Blask, D.E., Dauchy, R.T., Sauer, L.A., Krause, J.A., and Brainard, G.C. 2002 Light during darkness, melatonin suppression and cancer progression. *Neuro Endocrinol Lett* 23 Suppl 2: 52-6.
173. Lissoni, P. 2002 Is there a role for melatonin in supportive care? *Support Care Cancer* 10: 110-6.
174. Karasek, M., and Pawlikowski, M. 1999 Pineal gland, melatonin and cancer. *NEL Review. Neuro Endocrinol Lett* 20: 139-144.
175. Sauer, L.A., Dauchy, R.T., and Blask, D.E. 2001 Polyunsaturated fatty acids, melatonin, and cancer prevention. *Biochem Pharmacol* 61: 1455-62.
176. Blask, D.E., Sauer, L.A., and Dauchy, R.T. 2002 Melatonin as a chronobiotic/anticancer agent: cellular, biochemical, and molecular mechanisms of action and their implications for circadian-based cancer therapy. *Curr Top Med Chem* 2: 113-32.
177. El-Domeiri, A.A., and Das Gupta, T.K. 1976 The influence of pineal ablation and administration of melatonin on growth and spread of hamster melanoma. *J Surg Oncol* 8: 197-205.
178. Catrina, S.B., Curca, E., Catrina, A.I., Radu, C., and Coculescu, M. 2002 Melatonin shortens the survival rate of Ehrlich ascites-inoculated mice. *Neuro Endocrinol Lett* 22: 432-4.
179. Collins, A., Yuan, L., Kiefer, T.L., Cheng, Q., Lai, L., and Hill, S.M. 2003 Overexpression of the MT1 melatonin receptor in MCF-7 human breast cancer cells inhibits mammary tumor formation in nude mice. *Cancer Lett* 189: 49-57.