TOLL-LIKE RECEPTORS: LINKING INNATE AND ADAPTIVE IMMUNITY

Chandrashekhar Pasare and Ruslan Medzhitov*

1. ABSTRACT

Work in recent years has shown an essential role for Toll-like receptors (TLRs) in the activation of innate and adaptive immunity in vertebrate animals. These germ-line encoded receptors, expressed on a diverse variety of cells and tissues, recognize conserved molecular products derived from various classes of pathogens, including Gram-positive and -negative bacteria, DNA and RNA viruses, fungi and protozoa. Ligand recognition induces a conserved host defense program, which includes production of inflammatory cytokines, upregulation of costimulatory molecules, and induction of antimicrobial defenses. Importantly, activation of dendritic cells by TLR ligands is necessary for their maturation and consequent ability to initiate adaptive immune responses. How responses are tailored by individual TLRs to contain specific classes of pathogens is not yet clear.

2. INTRODUCTION

In all animals, the innate immune system provides essential protection against invading pathogens. A key component of this system is a collection of germ-line encoded receptors called pathogen recognition receptors (PRRs), which recognize a highly conserved set of molecular structures specific to microbes (Pathogen associated molecular patterns, or PAMPs) [1]. In addition to this system, vertebrates have a second line of defense called the adaptive immune system, which employs a diverse set of somatically rearranged receptors (T- cell receptors [TCRs] and B-cell receptors [BCRs]) with the ability to recognize a large spectrum of antigens.

^{*}Howard Hughes Medical Institute, Section of Immunobiology, 300 Cedar Street, TAC S660, Yale University School of Medicine, New Haven, CT 06510.

Mechanisms of Lymphocyte Activation and Immune Regulation X Edited by Gupta et al., Springer, 2005

C. PASARE and R. MEDZHITOV

The best understood and perhaps the most important subgroup of PRRs is the Toll-like receptor family. These receptors have the ability to recognize pathogens or pathogen derived products and initiate signaling events leading to activation of innate host defenses. Signaling by TLRs initiates acute inflammatory responses by induction of anti-microbial genes and inflammatory cytokines and chemokines [2,3]. In addition, TLRs have an important role in activation of adaptive immune responses [4,5]. Although T and B cells of the adaptive immune system express receptors of enormous diversity, activation of these cells depends on induction of co-stimulatory molecules and secretion of cytokines and chemokines by the cells of the innate immune system. Effective response to microbial infection requires several levels of interactions between innate and adaptive immune systems. A variety of cell surface receptors, secreted cytokines and chemokines participate in the induction of protective immunity. We will discuss here current paradigms of the importance of innate immune recognition by TLRs and the significance of that recognition for the outcome of adaptive immune responses. We will also discuss how inappropriate activation of TLRs under certain circumstances can lead to autoimmune diseases.

3. TLRs AND THEIR LIGANDS

The mammalian TLR family consists of 10 members with distinct ligand specificities and gene targets [2,3]. TLR4 recognizes lipopolysaccharide (LPS) [6,7] from gram-negative bacteria, TLR2 recognizes peptidoglycan from gram-positive bacteria [8], TLR3 recognizes double-stranded RNA from double stranded and negative strand viruses [9], TLR7 and 8 recognize RNA from single stranded viruses [10,11], and TLR9 recognizes unmethylated CpG DNA found abundantly in prokaryotic genomes and DNA viruses [12,13]. A comprehensive list of ligands and signaling events downstream of various TLRs is described elsewhere [3,14]. In addition, several reports have suggested that some TLRs can also recognize host-derived ligand. One example is the recognition of heat shock proteins by TLR2 and TLR4 [15-17]; however, it remains possible that the recombinant heat shock proteins used in these studies were contaminated with endotoxin (or other TLR ligands), consistent with more recent reports that more stringent purification of the hsps resulted in a loss of stimulatory activity [18-20]. Another example of recognition of a host ligand by a TLR (chromatin associated DNA by TLR9) is discussed in more detail below.

4. ADAPTIVE IMMUNE SYSTEM: MECHANISMS OF TOLERANCE AND IMPORTANCE OF INNATE IMMUNE RECOGNITION

Although most auto-reactive T and B lymphocytes undergo clonal deletion during their development in primary lymphoid organs (central tolerance), a few nevertheless escape into the periphery and must be held in check by mechanisms of peripheral tolerance [21].

12

TOLL-LIKE RECEPTORS: LINKING INNATE AND ADAPTIVE IMMUNITY

Clonal deletion of auto-reactive B cells is achieved in the bone marrow. Immature B cells that interact with membrane bound self-antigens are promptly deleted, while those that interact with soluble self-antigens are subject to a much more prolonged process of anergy followed by apoptosis [22-25]. In the case of T cells, clonal deletion is the fate of thymocytes with high affinity to self-antigens expressed in the thymus [21]. To make this an effective process, it is important that genes that function only in peripheral tissues and organs are expressed in thymic epithelium and DCs resident in thymus. Recent findings identify a transcriptional regulator called autoimmune regulator (AIRE) that permits thymic expression of many genes that function or are otherwise expressed only in peripheral tissues and organs [26,27]. Mice and humans with defective AIRE have an increased number of self-reactive T cells, which underscores the important role of AIRE in central tolerance [26-28]. Because expression of co-stimulatory molecules on thymic APCs is essential for induction of negative selection, it will be interesting to address the role of TLRs in controlling co-stimulatory molecule upregulation in this context [29-32].

Tolerance to self-antigens in the peripheral tissues and secondary lymphoid organs is achieved by at least two distinct and equally important mechanisms. The first one is through controlled expression of co-stimulatory molecules on professional APCs. The second is by the suppressor activity of a specialized group of cells called suppressor or regulatory T cells (Tr cells). Priming of naive T cells can be induced only when TCR-MHC/peptide interaction is coupled with a second (co-stimulatory) signal provided by interaction between CD28 on T cells and CD80/CD86 molecules on APCs [33,34]. This second signal, a result of dendritic cell (DC) maturation [35], is regulated by Toll-like receptors, and in effect, flags the antigen as being pathogenic [36]. Another consequence of TLR-induced DC maturation is the elaboration of a variety of cytokines and chemokines [2,3]. These soluble signals play an important role in the outcome of adaptive immune responses. Some cytokines are particularly important for overcoming suppression mediated by Tr cells; others like IL-12 are responsible for directing the T cell responses towards a Th1 phenotype.

Recently, several studies have suggested that peripherally-resident DCs, in addition to TLR-induced migration to draining lymph nodes during infection, may also traffick to lymph nodes even in the absence of infection. This homeostatic migration enables DCs expressing self-peptides either to tolerize self-reactive T cells [37,38] or to promote these T cells to acquire properties of suppressor T cells [39,40], thereby promoting peripheral tolerance.

The second mechanism by which peripheral tolerance is maintained is through the activity of Tr cells. These cells can be divided into two major classes based on their function. The first class, which express CD4 and CD25, [41] reside in all the secondary lymphoid organs and develop in the thymus under control of a specialized transcription factor Foxp3 [42,43]. This class of suppressor cells may have evolved to prevent activation of auto-reactive T cells that escape thymic deletion; humans and mice with defective Foxp3 expression develop generalized, fatal autoimmune [42,44]. The second class is characterized by the secretion of either of two anti-inflammatory cytokines, IL-10 (Tr1) [45], or TGF-beta (Th3) [46]. These suppressor cells typically reside in mucosal tissues of the body such as the gut and lung, and may have evolved to prevent destruction of host tissues caused by chronic inflammatory processes.

5. ROLE OF TLRs IN PROTECTION FROM INFECTIONS

5.1. Induction of Innate Immune Responses

The primary function of pattern recognition receptors is to provide an immediate protection from invading pathogens [3,47]. This is achieved through activation of a plethora of defense mechanisms, including inflammatory cytokines, complement, phagocytosis, and killing via anti-microbial proteins and peptides [48]. In addition, many members of the TLR family are potent inducers of type I interferons and the consequent suite of interferon-inducible genes in response to viral DNA and RNA [9-11,13,49]. Below we will discuss data indicating an essential role for TLRs in control of adaptive immunity.

5.2. Induction of Adaptive Immune Responses: Role of Co-stimulation and Cytokines

As described earlier, TLR-induced DC maturation during infection is essential for naive T cell activation. This has been demonstrated in mice lacking MyD88, and adapter protein downstream of all TKRs. optimal activation of naive T cells requires signals through both TCR and CD28 molecules [33,34]. These requirements are met only by a fully mature DC exposed to TLR ligands either in the secondary lymphoid organ or the peripheral tissues. Engagement of TLRs on DCs by TLR ligands (presence of infection) leads to up-regulation of both MHC and co-stimulatory molecules [35]. Migration of this professional APC to the draining lymph node and subsequent interaction with naive T cells ensures that immune responses are mounted only to pathogen-derived antigens. As discussed earlier interaction of immature DC and naive T cells leads to tolerance or induction of suppressor T cells [50]. The scenario of exposure to pathogens is mimicked in most experimental conditions by use of TLR ligands as adjuvants. The hypothesis that induction of DC maturation by microbial stimuli is essential for naive T cell activation was experimentally confirmed using MyD-88 deficient mice. MyD88 is an adapter protein that functions downstream of all TLRs and deletion of this adapter abolish signal transduction downstream of most TLRs

As described earlier, TLR-induced DC maturation during infection is essential for naive T cell activation. This has been demonstrated in mice lacking MyD88, and adapter protein downstream of all TLRs. [3]. Unlike wild-type mice, MyD88-deficient mice failed to induce T cell activation and interferon-gamma production when immunized with antigens emulsified in complete Freund's adjuvant (CFA), which is essentially heat killed Mycobacterium and contains several TLR ligands [4]. Further analysis showed that mycobacterial extracts fail to induce maturation of DCs derived from MyD88-deficient mice [4], underscoring the importance of TLR-induced DC maturation for T cell priming.

In addition to controlling the co-stimulatory pathway, DCs seem to contribute to T cell activation by overcoming suppression mediated by Tr cells. This was shown in experiments using LPS-treated MyD88-deficient DCs, which can still undergo maturation, but cannot produce inflammatory cytokines [51]. Surprisingly, when these DCs were used as APCs in T cell priming assays, they failed to induce effective priming. Additional experiments revealed that the cytokine IL-6, produced by DCs upon TLR ligation, is essential to overcome the function of Tr suppressor T cells, by making responder T cells refractory to

suppression [5]. Importantly, TLR-induced cytokines, in the absence of co-stimulatory molecule induction, seem to be insufficient by themselves to induce T cell activation (unpublished observations, CP and RM), thereby ensuring that bystander T cells are not activated non-specifically by DCs during infection.

6. TLRs AND AUTOIMMUNITY

6.1. Control of B Cell Activation

In addition to their clonally-expressed B cell receptors, B lymphocytes express most known TLRs. TLR ligation on B cells induces polyclonal proliferation and expression of co-stimulatory molecules, and also promotes plasma cell differentiation, but the significance of these events (with regard to inducibility by TLR signaling) is not yet clear. However, that TLR activation on B cells can contribute to pathology in at least some contexts has been shown. In this study, linked recognition of ligands by BCR and TLR9 has been shown to initiate and exacerbate at least one autoimmune disease. While TLR9 evolved to recognize unmethylated CpG motifs found in bacterial genomes and DNA viruses, TLR9 can also be activated by self DNA under certain special circumstances. One such case occurs when apoptotic cells are inefficiently removed, leading to secondary necrosis and release of high quantities of chromatin-bound DNA fragments. B cells specific to these fragments efficient take them up via BCR-mediated endocytosis and deliver them to intracellular sites containing TLR9. Signaling through TLR9 in such a scenario can lead to activation of B cells and differentiation into plasma cells that secrete antibodies specific to chromatin or chromatin-associated antigens and lead to immune pathology. A mouse model has been described recently that demonstrates that uptake of such complexes by rheumatoid factor positive (RF+) B cells leads to highly efficient activation of such B cells [52]. This activation was shown to be dependent on TLR9-mediated recognition of DNA present in the mammalian chromatin. Dual signaling through BCR and TLR9 therefore led to enhanced production of RF antibodies and associated immune pathology. This example illustrates the potential danger inherent in expression of both BCRs and TLRs on B cells, and may explain why continuous signaling through the BCR, which may indicate engagement of self antigens, leads to inhibition of TLR9-triggered plasma cell differentiation [53].

6.2. Role of TLR-Induced Cytokines

As discussed earlier, cytokines secreted by DCs and macrophages in response to TLR ligands contribute significantly to induction of T cell responses. However, many of these cytokines, including IL-6, TNF, and interferons, can in some contexts contribute to autoimmunity [54]. IL-6 has been implicated in autoimmune diseases such as pristine induced lupus, collagen induced arthritis and experimental autoimmune encephalomyelitis (EAE) [55-58]. The fact that IL-6-deficient animals are resistant to several auto-immune diseases suggest that this may be, at least in part, due to the inability of its effector T cells to overcome suppression mediated by Tr cells. During chronic infections, persistence of pathogens and their products can lead to enhanced production of several pro-inflammatory

cytokines including IL-6, leading to a situation conducive to activation of self-reactive T cells. Indeed, several studies have shown a link between chronic infections and autoimmune diseases [59].

7. REFERENCES

- 1. Janeway CA, Jr.: Approaching the asymptote? Evolution and revolution in immunology. *Cold Spring Harb Symp Quant Biol* 1989, **54 Pt 1**:1-13.
- 2. Janeway CA, Jr., Medzhitov R: Innate immune recognition. Annu Rev Immunol 2002, 20:197-216.
- 3. Takeda K, Kaisho T, Akira S: Toll-like receptors. Annu Rev Immunol 2003, 21:335-376.
- Schnare M, Barton GM, Holt AC, Takeda K, Akira S, Medzhitov R: Toll-like receptors control activation of adaptive immune responses. *Nat Immunol* 2001, 2:947-950.
- Pasare C, Medzhitov R: Toll pathway-dependent blockade of CD4+CD25+ T cell-mediated suppression by dendritic cells. Science 2003, 299:1033-1036.
- Poltorak A, He X, Smirnova I, Liu MY, Van Huffel C, Du X, Birdwell D, Alejos E, Silva M, Galanos C, et al.: Defective LPS signaling in C3H/HeJ and C57BL/10ScCr mice: mutations in Tlr4 gene. *Science* 1998, 282:2085-2088.
- Qureshi ST, Lariviere L, Leveque G, Clermont S, Moore KJ, Gros P, Malo D: Endotoxin-tolerant mice have mutations in Toll-like receptor 4 (Tlr4). J Exp Med 1999, 189:615-625.
- Takeuchi O, Hoshino K, Kawai T, Sanjo H, Takada H, Ogawa T, Takeda K, Akira S: Differential roles of TLR2 and TLR4 in recognition of gram-negative and gram-positive bacterial cell wall components. *Immunity* 1999, 11:443-451.
- Alexopoulou L, Holt AC, Medzhitov R, Flavell RA: Recognition of double-stranded RNA and activation of NF-kappaB by Toll-like receptor 3. *Nature* 2001, 413:732-738.
- 10. Heil F, Hemmi H, Hochrein H, Ampenberger F, Kirschning C, Akira S, Lipford G, Wagner H, Bauer S: Species-Specific Recognition of Single-Stranded RNA via Toll-like Receptor 7 and 8. *Science* 2004.
- 11. Diebold SS, Kaisho T, Hemmi H, Akira S, Reis ESC: Innate Antiviral Responses by Means of TLR7-Mediated Recognition of Single-Stranded RNA. *Science* 2004.
- Hemmi H, Takeuchi O, Kawai T, Kaisho T, Sato S, Sanjo H, Matsumoto M, Hoshino K, Wagner H, Takeda K, et al.: A Toll-like receptor recognizes bacterial DNA. *Nature* 2000, 408:740-745.
- 13. Lund J, Sato A, Akira S, Medzhitov R, Iwasaki A: Toll-like receptor 9-mediated recognition of Herpes simplex virus-2 by plasmacytoid dendritic cells. *J Exp Med* 2003, **198**:513-520.
- 14. Barton GM, Medzhitov R: Toll-like receptor signaling pathways. Science 2003, 300:1524-1525.
- 15. Asea A, Rehli M, Kabingu E, Boch JA, Bare O, Auron PE, Stevenson MA, Calderwood SK: Novel signal transduction pathway utilized by extracellular HSP70: role of toll-like receptor (TLR) 2 and TLR4. *J Biol Chem* 2002, **277**:15028-15034.
- 16. Ohashi K, Burkart V, Flohe S, Kolb H: Cutting edge: heat shock protein 60 is a putative endogenous ligand of the toll-like receptor-4 complex. *J Immunol* 2000, **164**:558-561.
- 17. Vabulas RM, Ahmad-Nejad P, Ghose S, Kirschning CJ, Issels RD, Wagner H: HSP70 as endogenous stimulus of the Toll/interleukin-1 receptor signal pathway. *J Biol Chem* 2002, **277**:15107-15112.
- Bausinger H, Lipsker D, Ziylan U, Manie S, Briand JP, Cazenave JP, Muller S, Haeuw JF, Ravanat C, de la Salle H, et al.: Endotoxin-free heat-shock protein 70 fails to induce APC activation. *Eur J Immunol* 2002, 32:3708-3713.
- Gao B, Tsan MF: Recombinant Human Heat Shock Protein 60 Does Not Induce the Release of Tumor Necrosis Factor {alpha} from Murine Macrophages. J Biol Chem 2003, 278:22523-22529.
- Gao B, Tsan MF: Endotoxin contamination in recombinant human heat shock protein 70 (Hsp70) preparation is responsible for the induction of tumor necrosis factor alpha release by murine macrophages. J Biol Chem 2003, 278:174-179.
- 21. Starr TK, Jameson SC, Hogquist KA: Positive and negative selection of T cells. *Annu Rev Immunol* 2003, **21**:139-176.
- 22. Goodnow CC, Crosbie J, Adelstein S, Lavoie TB, Smith-Gill SJ, Brink RA, Pritchard-Briscoe H, Wotherspoon JS, Loblay RH, Raphael K, et al.: Altered immunoglobulin expression and functional silencing of self-reactive B lymphocytes in transgenic mice. *Nature* 1988, **334**:676-682.

TOLL-LIKE RECEPTORS: LINKING INNATE AND ADAPTIVE IMMUNITY

- 23. Hartley SB, Crosbie J, Brink R, Kantor AB, Basten A, Goodnow CC: Elimination from peripheral lymphoid tissues of self-reactive B lymphocytes recognizing membrane-bound antigens. *Nature* 1991, **353**:765-769.
- 24. Cyster JG, Hartley SB, Goodnow CC: Competition for follicular niches excludes self-reactive cells from the recirculating B-cell repertoire. *Nature* 1994, **371**:389-395.
- 25. Cyster JG, Goodnow CC: Antigen-induced exclusion from follicles and anergy are separate and complementary processes that influence peripheral B cell fate. *Immunity* 1995, 3:691-701.
- 26. Anderson MS, Venanzi ES, Klein L, Chen Z, Berzins SP, Turley SJ, von Boehmer H, Bronson R, Dierich A, Benoist C, et al.: Projection of an immunological self shadow within the thymus by the aire protein. *Science* 2002, **298**:1395-1401.
- Liston A, Lesage S, Wilson J, Peltonen L, Goodnow CC: Aire regulates negative selection of organ-specific T cells. *Nat Immunol* 2003, 4:350-354.
- Peterson P, Nagamine K, Scott H, Heino M, Kudoh J, Shimizu N, Antonarakis SE, Krohn KJ: APECED: a monogenic autoimmune disease providing new clues to self-tolerance. *Immunol Today* 1998, 19:384-386.
- 29. Punt JA, Osborne BA, Takahama Y, Sharrow SO, Singer A: Negative selection of CD4+CD8+ thymocytes by T cell receptor-induced apoptosis requires a costimulatory signal that can be provided by CD28. *J Exp Med* 1994, **179**:709-713.
- Punt JA, Havran W, Abe R, Sarin A, Singer A: T cell receptor (TCR)-induced death of immature CD4+CD8+ thymocytes by two distinct mechanisms differing in their requirement for CD28 costimulation: implications for negative selection in the thymus. J Exp Med 1997, 186:1911-1922.
- 31. Kishimoto H, Cai Z, Brunmark A, Jackson MR, Peterson PA, Sprent J: Differing roles for B7 and intercellular adhesion molecule-1 in negative selection of thymocytes. *J Exp Med* 1996, **184**:531-537
- 32. Sprent J, Kishimoto H: The thymus and negative selection. Immunol Rev 2002, 185:126-135.
- Lenschow DJ, Walunas TL, Bluestone JA: CD28/B7 system of T cell costimulation. Annu Rev Immunol 1996, 14:233-258.
- Liu Y, Janeway CA, Jr.: Cells that present both specific ligand and costimulatory activity are the most efficient inducers of clonal expansion of normal CD4 T cells. Proc Natl Acad Sci U S A 1992, 89:3845-3849
- 35. Banchereau J, Steinman RM: Dendritic cells and the control of immunity. Nature 1998, 392:245-252
- 36. Medzhitov R: Toll-like receptors and innate immunity. Nat Rev Immunol 2001, 1:135-145.
- Belz GT, Behrens GM, Smith CM, Miller JF, Jones C, Lejon K, Fathman CG, Mueller SN, Shortman K, Carbone FR, et al.: The CD8alpha(+) dendritic cell is responsible for inducing peripheral self-tolerance to tissue-associated antigens. J Exp Med 2002, 196:1099-1104.
- Hawiger D, Inaba K, Dorsett Y, Guo M, Mahnke K, Rivera M, Ravetch JV, Steinman RM, Nussenzweig MC: Dendritic cells induce peripheral T cell unresponsiveness under steady state conditions in vivo. J Exp Med 2001, 194:769-779.
- Menges M, Rossner S, Voigtlander C, Schindler H, Kukutsch NA, Bogdan C, Erb K, Schuler G, Lutz MB: Repetitive injections of dendritic cells matured with tumor necrosis factor alpha induce antigen-specific protection of mice from autoimmunity. J Exp Med 2002, 195:15-21.
- 40. Wakkach A, Fournier N, Brun V, Breittmayer JP, Cottrez F, Groux H: Characterization of dendritic cells that induce tolerance and T regulatory 1 cell differentiation in vivo. *Immunity* 2003, **18**:605-617.
- 41. Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M: Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. *J Immunol* 1995, **155**:1151-1164.
- 42. Fontenot JD, Gavin MA, Rudensky AY: Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol* 2003, **4**:330-336.
- 43. Hori S, Nomura T, Sakaguchi S: Control of regulatory T cell development by the transcription factor Foxp3. *Science* 2003, **299**:1057-1061.
- 44. Asano M, Toda M, Sakaguchi N, Sakaguchi S: Autoimmune disease as a consequence of developmental abnormality of a T cell subpopulation. *J Exp Med* 1996, **184**:387-396.
- 45. Groux H, O'Garra A, Bigler M, Rouleau M, Antonenko S, de Vries JE, Roncarolo MG: A CD4+ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature* 1997, **389**:737-742.
- 46. Chen Y, Kuchroo VK, Inobe J, Hafler DA, Weiner HL: Regulatory T cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. *Science* 1994, **265**:1237-1240.
- 47. Thoma-Uszynski S, Stenger S, Takeuchi O, Ochoa MT, Engele M, Sieling PA, Barnes PF, Rollinghoff M, Bolcskei PL, Wagner M, et al.: Induction of direct antimicrobial activity through mammalian toll-like receptors. *Science* 2001, **291**:1544-1547.

- 48. Ayabe T, Satchell DP, Wilson CL, Parks WC, Selsted ME, Ouellette AJ: Secretion of microbicidal alpha-defensins by intestinal Paneth cells in response to bacteria. *Nat Immunol* 2000, 1:113-118.
- 49. Krug A, Luker GD, Barchet W, Leib DA, Akira S, Colonna M: Herpes simplex virus type 1 activates murine natural interferon-producing cells through toll-like receptor 9. *Blood* 2004, **103**:1433-1437.
- 50. Steinman RM, Hawiger D, Nussenzweig MC: Tolerogenic dendritic cells. Annu Rev Immunol 2003, 21:685-711.
- Kaisho T, Takeuchi O, Kawai T, Hoshino K, Akira S: Endotoxin-induced maturation of MyD88-deficient dendritic cells. J Immunol 2001, 166:5688-5694.
- Leadbetter EA, Rifkin IR, Hohlbaum AM, Beaudette BC, Shlomchik MJ, Marshak-Rothstein A: Chromatin-IgG complexes activate B cells by dual engagement of IgM and Toll-like receptors. *Nature* 2002, 416:603-607.
- Rui L, Vinuesa CG, Blasioli J, Goodnow CC: Resistance to CpG DNA-induced autoimmunity through tolerogenic B cell antigen receptor ERK signaling. *Nat Immunol* 2003, 4:594-600.
- 54. Drakesmith H, Chain B, Beverley P: How can dendritic cells cause autoimmune disease? *Immunol Today* 2000, **21**:214-217.
- 55. Alonzi T, Fattori E, Lazzaro D, Costa P, Probert L, Kollias G, De Benedetti F, Poli V, Ciliberto G: Interleukin 6 is required for the development of collagen-induced arthritis. *J Exp Med* 1998, **187**:461-468.
- 56. Kobayashi H, Ohshima S, Nishioka K, Yamaguchi N, Umeshita-Sasai M, Ishii T, Mima T, Kishimoto T, Kawase I, Saeki Y: Antigen induced arthritis (AIA) can be transferred by bone marrow transplantation: evidence that interleukin 6 is essential for induction of AIA. J Rheumatol 2002, 29:1176-1182.
- 57. Ohshima S, Saeki Y, Mima T, Sasai M, Nishioka K, Nomura S, Kopf M, Katada Y, Tanaka T, Suemura M, et al.: Interleukin 6 plays a key role in the development of antigen-induced arthritis. *Proc Natl Acad Sci U S A* 1998, **95**:8222-8226.
- Richards HB, Satoh M, Shaw M, Libert C, Poli V, Reeves WH: Interleukin 6 dependence of anti-DNA antibody production: evidence for two pathways of autoantibody formation in pristane-induced lupus. J Exp Med 1998, 188:985-990.
- 59. Rose NR: The role of infection in the pathogenesis of autoimmune disease. Semin Immunol 1998, 10:5-13.

18