



# Deciphering the Dynamics of Epithelial-Mesenchymal Transition and Cancer Stem Cells in Tumor Progression

Federico Bocci<sup>1,2</sup> · Herbert Levine<sup>1,2,3,4</sup> · José N. Onuchic<sup>1,2,4,5</sup> · Mohit Kumar Jolly<sup>1,6</sup>

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## Abstract

**Purpose of Review** The epithelial-mesenchymal transition (EMT) and the generation of cancer stem cells (CSCs) are two fundamental aspects contributing to tumor growth, acquisition of resistance to therapy, formation of metastases, and tumor relapse. Recent experimental data identifying the circuits regulating EMT and CSCs has driven the development of computational models capturing the dynamics of these circuits, and consequently various aspects of tumor progression.

**Recent Findings** We review the contribution made by these models in (a) recapitulating experimentally observed behavior, (b) making experimentally testable predictions, and (c) driving emerging notions in the field, including the emphasis on the aggressive potential of hybrid epithelial-mesenchymal (E/M) phenotype(s). We discuss dynamical and statistical models at intracellular and population level relating to dynamics of EMT and CSCs, and those focusing on interconnections between these two processes.

**Summary** These models highlight the insights gained via mathematical modeling approaches and emphasizes that the connections between hybrid E/M phenotype(s) and stemness can be explained by analyzing underlying regulatory circuits. Such experimentally curated models have the potential of serving as platforms for better therapeutic design strategies.

**Keywords** Epithelial-mesenchymal transition · Cancer stem cells · Hybrid epithelial-mesenchymal (E/M) phenotype · Mathematical modeling

## Introduction

Metastasis and tumor relapse are insuperable clinical challenges that claim most cancer-related deaths [1]. The

metastatic cascade has extremely high rates of attrition, because of the multi-step and challenging sequence of events leading to a secondary tumor. These steps include the detachment of cancer cells from their home organ, their circulation in the bloodstream, and eventually their colonization of a foreign environment, all while escaping attack by the immune system and other clinical interventions.

A first step in the metastatic cascade is a phenotypic switch called epithelial-to-mesenchymal transition (EMT). Cancer cells in a solid tumor tissue often undergo EMT, characterized by the loss of cell-cell adhesion and acquisition of migratory and invasive traits [2]. Disseminated cells travel through the bloodstream and colonize a distant organ, giving rise to macrometastases [2, 3]. EMT is not necessarily a cell-autonomous and binary process. Cells can attain one or more intermediate, or hybrid, epithelial-mesenchymal (E/M) state(s) and can involve their neighbors to form more aggressive clusters of circulating tumor cells (CTCs)—the main drivers of metastases [4–6]. EMT is regulated at multiple levels—transcriptional, translational, post-translational and epigenetic—by many context-specific factors and the tumor microenvironment. Some common traits of EMT include transcriptional repression

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✉ Mohit Kumar Jolly  
mkjolly@iisc.ac.in

<sup>1</sup> Center for Theoretical Biological Physics, Rice University, Houston, TX 77005, USA

<sup>2</sup> Department of Chemistry, Rice University, Houston, TX 77005, USA

<sup>3</sup> Department of Bioengineering, Rice University, Houston, TX 77005, USA

<sup>4</sup> Department of Physics and Astronomy, Rice University, Houston, TX 77005, USA

<sup>5</sup> Department of Biosciences, Rice University, Houston, TX 77005, USA

<sup>6</sup> Present address: Centre for BioSystems Science and Engineering, Indian Institute of Science, Bangalore 560012, India

of E-cadherin that mediates cell-cell junctions and adhesion and activation of one or more EMT-inducing transcription factors (EMT-TFs) such as SNAI1, SNAI2, ZEB1, ZEB2, TWIST1 that induce cell scattering, motility, and invasion [2].

To colonize a secondary tumor site, the disseminated tumor cells need to give rise to different cell types that constitute a tumor—a trait typical of cancer stem cells (CSCs). Cells with such stem-like properties are also typically resistant to various clinical treatments and are often implicated in tumor relapse. The conventional, so-called CSC hypothesis envisions a small fraction of CSCs that can both self-renew (symmetric division) and generate differentiated cells (asymmetric division) [7, 8]. This hypothesis implies a hierarchical lineage of tumor cells similar to stem cell hierarchy in normal tissues, such that CSCs that differentiate irreversibly lose their stem-like properties [9]. Recent studies, however, have emphasized that stemness can be a dynamic cell state that can be acquired or lost [10–12, 13]. In other words, some differentiated tumor cells can dedifferentiate and regain stemness via epigenetic and/or environmental factors such as abnormal cancer metabolism and EMT.

The interconnection between EMT and CSCs was first postulated by Brabletz et al. [14] in 2005 as “migrating cancer stem cell” by suggesting that the concepts of EMT and CSCs, considered independent of one another, were not sufficient to explain various traits of cancer progression. Afterwards, experimental evidence accumulated suggesting that stemness can be gained during EMT [2, 5, 15–18, 19]. Recent experiments have shown that cells in intermediate E/M states possess a higher metastatic potential as compared to the cells that have undergone a complete EMT. Moreover, cells in hybrid E/M phenotype have been suggested to be drug-resistant [20, 21]. Put together, these observations emphasize the clinical implications of hybrid E/M phenotypes [5, 22, 23].

Recent studies have made significant progress in identifying the molecular networks regulating EMT, CSCs, and their interconnections [24]. These networks are formidably complex and capable to give rise to emergent non-linear behavior. Identification of these networks has driven a surge in deciphering their underlying principles from a dynamical systems perspective. This approach has involved developing many computational models to capture the dynamics of these transitions. These models may deal with intracellular and intercellular circuits or may offer a population-level description without considering the detailed dynamics of signaling networks. Here, we review both of these types of models. First, we review a set of models that attempt to characterize the possible set of states for cells undergoing EMT and their possible relevance to tumor progression and metastasis. Second, we review a set of models that consider the population structure of a tumor and its implications for drug resistance. Finally, we discuss models that aim at gaining a comprehensive understanding of the connection between these two crucial axes of cancer progression.

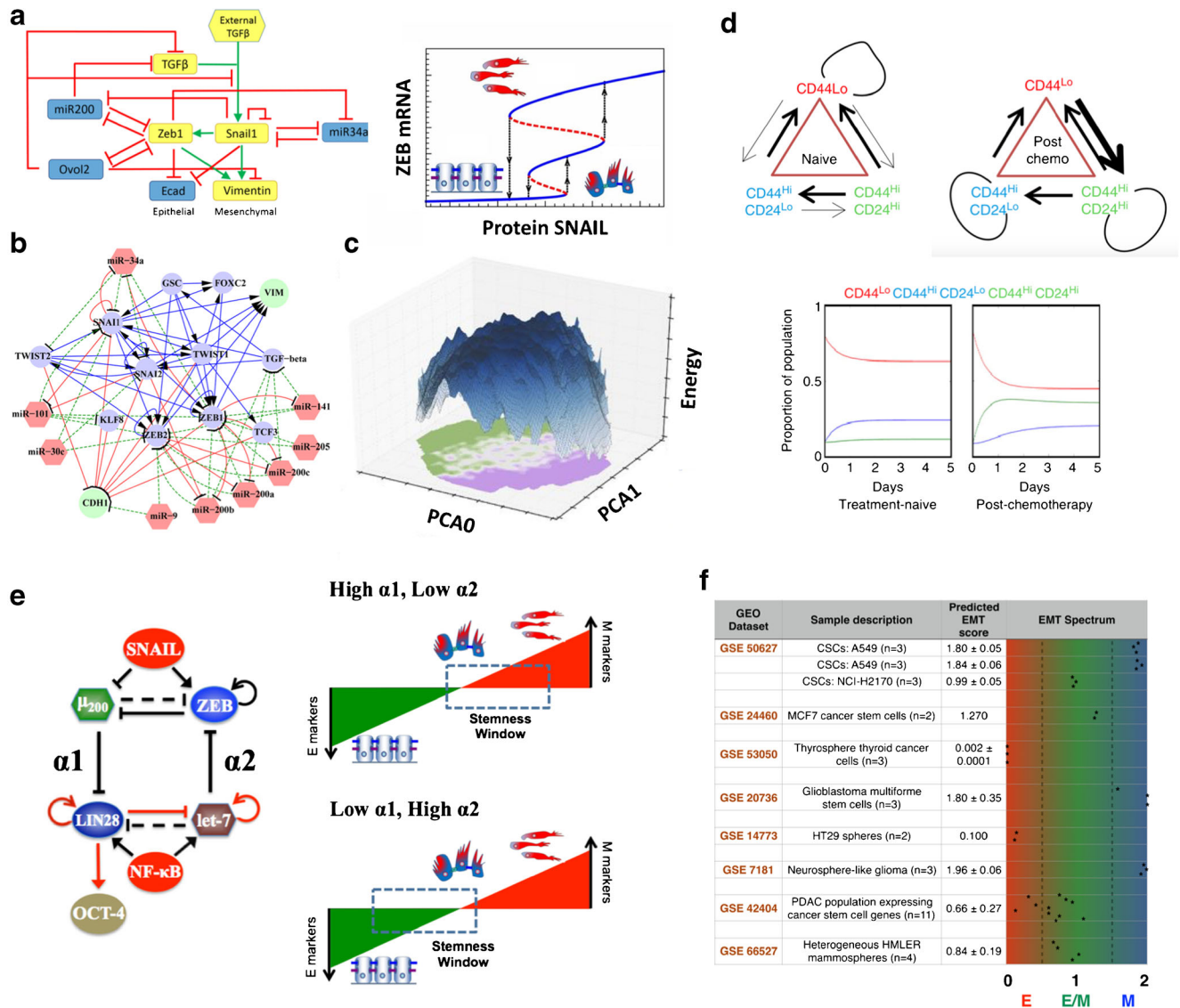
## Mathematical Models of EMT

Computational models developed for EMT can be categorized broadly into two classes: mechanism-based models and data-based models. While the first class of models adopts a “bottom-up” approach and focus on elucidating the properties of molecular networks identified experimentally, the latter adopts a “top-down” approach starting with high-dimensional data and aims to reverse engineer the networks, and/or trace the trajectories of these transitions using statistical methods.

### Decoding the Dynamics of Cellular Transitions: Mechanism-Based Models of EMT

The first set of mechanism-based models for EMT regulation—developed independently by two groups—focused on a small set of nodes and captured the dynamical features emerging from the interconnections among those nodes (Fig. 1a, left). These models included the EMT-suppressing microRNA families miR-34, miR-200, and the families of EMT-TFs ZEB and SNAIL [25, 26]. Both models predicted that this network can be tristable and could give rise to a hybrid epithelial-mesenchymal (E/M) phenotype, in addition to epithelial and mesenchymal phenotypes (Fig. 1a, right) [25, 26]. These models also suggested that more than one phenotype can be accessible to a cell due to the underlying multistability, hence giving rise to sub-populations of epithelial, hybrid E/M, and mesenchymal cells in a genetically identical population. This phenomenon was observed and later characterized in detail in multiple cancer cell lines [5, 27, 28, 29]. Due to different modeling approaches, however, these models differed on the dynamics of attaining this hybrid E/M phenotype. Experimental support for both these models has been observed [27, 30], highlighting the heterogeneity and multiplicity of hybrid E/M phenotype(s) present in different cell lines.

Further follow-up work has identified several intracellular phenotypic stability factors (PSFs) that can stabilize a hybrid E/M phenotype, including OVOL2, GRHL2, Np63 $\alpha$ , and NRF2 [31–34, 35]. Their role as PSFs have also been validated experimentally *in vitro* and *in vivo* [32, 34–36]. Moreover, higher levels of these PSFs were observed to correlate with worse patient survival, emphasizing the clinical implications of hybrid E/M phenotype(s) [4, 22]. Among those, NRF2 has been specifically proposed to be maximally expressed in hybrid E/M phenotype(s) [35]. In addition, different energy landscape approaches have been also developed for the aforementioned EMT circuit [37] as well as for related larger gene regulatory circuits [38]. This strategy allows to compute the transition rates between multiple cell states, and thus predict the relative abundance of different phenotypes (epithelial, mesenchymal, and hybrid E/M) in an isogenic population.



**Fig. 1** Mathematical models that characterize the landscape of cellular plasticity mediated via EMT and CSCs. **a** Left: a gene regulatory circuit for EMT (adapted from Hong et al. [34]). Right: a bifurcation diagram of ZEB mRNA as a function of EMT-TF SNAIL (adapted from Lu et al. [25•]) shows three stable phenotypes (i.e., continuous black curves) corresponding to epithelial (low ZEB), hybrid E/M (intermediate ZEB), and mesenchymal (high ZEB). **b** An extended EMT regulatory circuit (adapted from Huang et al. [46]). **c** The energy landscape of a large EMT regulatory circuit (adapted from Font-Clos et al. [45••]) shows two main minima (purple and green projections) corresponding to epithelial and mesenchymal phenotypes, respectively. Additionally, many local energy minima en route to EMT correspond to intermediate E/M states. PCA0 and PCA1 are the first two components of the principal component analysis of the circuit. **d** Chemotherapy increases the population of chemo-resistant cancer cells (CD44<sup>hi</sup>CD24<sup>hi</sup>) by increasing the

conversion rate from low-resistance CD44<sup>low</sup> cell population. Top: circuit schematic; bottom: temporal dynamics of cancer cell subpopulations pre- and post-treatment (adapted from Goldman et al. [67••]). **e** Left: a core gene regulatory circuit including regulation of EMT via the miR-200/ZEB axis and stemness via the LIN28/let-7 axis (adapted from Jolly et al. [12]). The parameters  $\alpha_1$ ,  $\alpha_2$  represent the strength in the regulation of the stemness circuit by the EMT circuit ( $\alpha_1$ ) and the EMT circuit by the stemness circuit ( $\alpha_2$ ). Right: varying  $\alpha_1$  and  $\alpha_2$  shifts the “stemness window” along the EMT axis adapted from Jolly et al. [12]. **f** The EMT score of different cancer stem cell lines (adapted from Bocci et al. [13•]) shows the spread of CSC properties along the EMT spectrum. The score classifies cells as epithelial (score < 0.5), hybrid E/M (0.5 < score < 1.5), or mesenchymal (score > 1.5). Each row depicts a different CSC line, and each dot depicts a different biological replicate

EMT can be also induced by biochemical signals coming from neighboring cells. Boareto et al. [39] elucidated the connection between EMT and the Notch signaling pathway, a cell-cell, contact-based, evolutionary conserved signaling mechanism that is also implicated in angiogenesis and therapy resistance. The model predicted that Notch-Jagged signaling

among cells, but not Notch-Delta signaling, can foster the formation of clusters of hybrid E/M cells by promoting a similar hybrid E/M phenotype in neighboring cells [39]. Consistently, gene expression analysis highlighted higher levels of Jagged in CTC clusters of patients as compared to single CTCs [33]. Thus, a hybrid E/M phenotype can be

stabilized not only by intracellular PSFs directly coupled to the EMT core circuit but also via cell-cell signaling. As another example, Bocci et al. [40] predicted that Numb—an inhibitor of Notch signaling—can also stabilize a hybrid E/M phenotype; this prediction was validated experimentally in multiple independent studies [40, 41].

As the network grows in size (such as going from Fig. 1a, b), identifying kinetic parameters becomes more and more challenging. Computational models that have focused on such larger networks have typically been simulated using Boolean modeling approaches, where the state of gene expression is either on (active) or off (inactive). Boolean models do not consider any kinetic parameters. Cohen et al. [42] developed a Boolean network to evaluate the combinatorial effect of different mutations on EMT and metastatic potential using transcriptome data from TGF- $\beta$ -induced EMT. Similarly, Steinway et al. [43] constructed a circuit for TGF- $\beta$ -induced EMT using data from hepatocellular carcinoma (HCC). Their model predicted the activation of several pathways during EMT such as Sonic Hedgehog and Wnt. Following up, the authors showed that certain perturbations could give rise to one or more hybrid E/M states and identified possible targets to inhibit TGF- $\beta$ -driven EMT [44]. Recently, Font-Clos et al. [45••] constructed Boolean model for a gene regulatory network that describes both EMT and its reverse, mesenchymal-to-epithelial transition (MET). An energy landscape approach showed two main attractors, or stable states, corresponding to epithelial and mesenchymal phenotypes, and multiple local minima, or relatively less stable states, corresponding to multiple hybrid E/M phenotypes (Fig. 1c). The authors further mapped RNA-seq data from both lung adenocarcinoma and embryonic differentiation during EMT/MET and compared it to the predicted phenotypic expression profiles, hence validating the existence of multiple different intermediate E/M states.

In an attempt to combine the advantages of both continuous small-scale models and Boolean large-scale models, Huang et al. [46] devised an algorithm—random circuit perturbation (RACIPE)—where the expression levels of genes are continuous, but the parameters for all regulatory links are randomly chosen within a biologically relevant range. RACIPE generates an ensemble of mathematical models, each with a different set of parameters, and identifies the robust dynamical states emerging from a given network topology. Applying RACIPE to an EMT circuit composed of 9 microRNAs and 13 TFs (Fig. 1b) highlighted two different hybrid E/M states [46] that could be stabilized further by stochasticity or noise [47].

### Reconstructing EMT Plasticity from Experiments: Data-Driven Approaches to EMT

Recent experimental techniques are capable of generating large and high-throughput (‘omics’ level) data. This deluge has driven a class of data-driven, or ‘top-down’, models,

which employ a variety of statistical tools to reconstruct correlations among genes and develop expression signatures of different EMT phenotypes.

For example, Zadran et al. [48] analyzed the temporal mRNA data of A549 lung cancer cells treated with TGF $\beta$  and identified an intermediate EMT state with a metabolic state characterized by increased cytosolic ATP levels [48]. Further, Chang et al. [49] analyzed the time course data of TGF- $\beta$ -driven EMT for A549 cells and identified three master TFs for a partial EMT state—ETS2, HNF4A, and JUNB [49]. These regulators correlate with a worse clinical outcome and their knockdown can prevent TGF $\beta$ -driven EMT [49], reminiscent of observations made for PSFs.

Besides, two different groups developed methods to analyze gene expression data of a certain cell line or tumor cell and calculate an “EMT score” to quantify the positioning of these cells along the EMT spectrum. The algorithm developed by Tan et al. uses entire transcriptomic data for a given sample [50], while that developed by George et al. considers a few E and M markers (such as E-Cadherin, Vimentin etc.) as well as PSFs of hybrid E/M phenotypes (OVOL, GRHL2 etc.) [23].

Data-driven models do not necessarily rely on omics-level data; they can also use morphological data. For instance, Mandal et al. [51] proposed a phenomenological approach to elucidate intermediate EMT states based on cell microscopy during EMT and found three intermediate states with different morphological attributes [51]. A more rigorous analysis was proposed by Leggett et al. [52] that relies on single-cell microscopy to classify cells as epithelial or mesenchymal with high precision during TGF $\beta$ -driven EMT [52]. Even further, Zhang et al. [53] classified the morphology and motility of migrating breast cancer cells using machine learning algorithms such as artificial neural networks (ANN) and random decision forest (RDF) to analyze single-cell microfluidic microscopy images [53].

As the connections between molecular and morphological traits of EMT continue to be explored in detail [54], a synergistic crosstalk among the computational models described above and their integration with experimental data can provide novel and crucial insights into the dynamics of EMT.

## Mathematical Models of CSCs

### Modeling the CSC Fraction During Tumor Progression

An important direction where mathematical approaches have offered significant insights into the CSC dynamics and its relationship with tumor progression is a set of population dynamic models that aim at understanding the temporal dynamics and the mechanisms regulating the CSC fraction (the fraction of cells with stem cell properties in a tumor) [55, 56]. Dhawan et al. [57] considered two compartments, the CSC-like cells and



the non-CSC-like cells, to elucidate the increased plasticity observed in human mammary epithelial cells under hypoxia. In their model, individual cells can both differentiate (from CSC to non-CSC) and dedifferentiate (from non-CSC to CSC). Integrating their model with gene expression analysis, the authors showed that hypoxia generates a shift toward a more CSC-like population and increases EMT features [57].

The role of cell dedifferentiation to a stem-like state has been also investigated by Jilkine et al. [58] via a hybrid model that describes the development of the differentiated cell population in a deterministic manner but considers the stochastic accumulation of mutations to better describe the small CSC population. The authors concluded that dedifferentiation to a stem-like state can speed up tumor progression by enlarging the CSC population [58].

Among the different possible mechanism for dedifferentiation, metabolic reprogramming is especially frequent in the context of cancer [59]. Liu et al. [60] devised a probabilistic framework to specifically investigate metabolic reprogramming that converts somatic cells into pluripotent stem cells. Insights gained from such analysis may be useful in understanding metabolic aspects of tumor cell dedifferentiation, given that different subsets of CSCs may have different metabolic vulnerabilities [61].

Another set of models have aimed to explain how a fraction of CSCs is maintained in a tumor. Zhou et al. [62] developed a population model of tumor growth that integrates the differential growth rate of CSCs and differentiated cells, as well as transitions among cell phenotypes. In particular, switching between phenotypes maintains a fixed ratio of cell sub-populations [62]. Extending this idea, Wang et al. [63] proposed a population model that combines hierarchical organization (irreversible loss of stem traits upon differentiation) and stochastic switching (stemness can be gained by switching to a stem-like state). In this model, CSCs can (a) self-renew (symmetric division in two CSCs), (b) differentiate (symmetric division into two non-stem cells), and (c) asymmetrically divide into a CSC and a daughter differentiated cell. Additionally, differentiated cells can proliferate (symmetric division) but also switch to the progenitor CSC state [63]. The combination of hierarchical and stochastic processes can reproduce the CSC/differentiated cell fraction observed in a human colon cancer cell population [63]. A similar idea has been proposed by Zhou et al. [64] to show that back-and-forth transitions between stem-like and non-stem states is crucial to establish an equilibrium cell fraction of CSCs [64].

A different approach to model CSC-driven tumor progression was proposed by Poleszczuk et al. [65]. They proposed an agent-based model where CSCs can gain migratory traits by stochastic mutations. Such approach enables to simulate the spatiotemporal dynamics of the cancer cell population and investigates the cell heterogeneity that arises during tumor development due to mutations. In this model, CSC can divide symmetrically or asymmetrically, and also have a migration

potential that translates into discrete movements on a two-dimensional lattice [65], reminiscent of the idea of “migrating cancer stem cell” proposed by Brabletz [14].

### CSC, Tumor Progression, and Therapy: From Modeling to the Clinic

A subset of recent models have focused their attention toward identifying optimal therapy schedules for cancer treatment [66]. In this context, CSCs are considered as important target because they are resistant to therapy and can therefore drive tumor progression and relapse. For instance, the model developed by Dhawan et al. [57] (discussed in the previous section) can be generalized to the context of drug tolerance by introducing one or more additional cell sub-populations that can resist different treatments [67••]. Specifically, the authors show by integrating *in vivo* experiments and mathematical modeling that chemotherapy can change the rates of conversions among different cell phenotypes and promote a chemotherapy-tolerant state (Fig. 1d) [67••].

A more data-driven approach aims to correlate the CSC population with tumor progression and response to therapy. For instance, Werner et al. [68] proposed a computational method to quantify the fraction of tumor-initiating cells (i.e., CSCs) by analyzing the tumor’s macroscopic growth rate as a function of time. This patient-specific method can be applied to many types of tumors and provides an estimate of the CSC fraction to rationalize the optimal therapy in a clinical setting [68]. Zhou et al. [69] applied a statistical approach to compute the transition rate between CSC and differentiated cells in colon cancer cells and showed phenotypic plasticity with back and forth transitions [69]. Furthermore, Yu et al. [70] gathered the differential response of CSCs and differentiated cells to radiotherapy for different tumor types including glioblastoma, lung, prostate, and breast cancer and fitted this tumor-specific information with a stochastic mathematical model to explain the different inter-tumor responses to radiation therapy [70].

Not all models of cancer cell-therapy interplay need to employ a population approach. Instead, Chen et al. [71] used an energy landscape approach to investigate the transitions of breast cancer cells which are sensitive, hypersensitive, or insensitive to hormone therapy regulated by the ER $\alpha$  signaling network. The authors implemented different treatment strategies including sequential treatment (multiple drugs) and intermittent treatment (alternation of treatment and “drug holiday” periods) [71]. The effects of continuous vs. intermittent treatments was also explored in the context of prostate cancer, where a small-scale model predicted that cells could oscillate between a therapy-sensitive and a therapy-resistant phenotype [72]. The authors further modeled different hormonal treatments for prostate cancer that were predicted to

synchronize oscillations among different cells, thus restricting the heterogeneity in the cell population [73].

As discussed above, most models related to CSCs have largely focused on identifying the causes underlying varying fractions of tumor population that can behave as CSCs [74]. Thus, there is much room for progress in constructing mechanistic models for CSC-driven tumor progression and the emergence of drug-resistant phenotypes. In this direction, Nazari et al. [75] recently proposed a mathematical model for the role of inflammatory cytokines in mediating CSC-driven tumor growth. This model couples the ligand-receptor interaction at the molecular scale with CSC self-renewal and proliferation at the cellular level [75] and could reproduce the observed decrease in tumor volume in mouse models with knockdown of IL-6R, an inflammatory cytokine.

## Toward an Integrated Understanding of EMT and CSC

Aside from the separate models for EMT and CSC dynamics as discussed above, multiple computational models have investigated the connection between EMT and CSC. Turner et al. [76] interrogated the connection between EMT and CSC through a phenomenological, population model with two possible scenarios where EMT enriches the CSC population. First, cells can dedifferentiate back to a CSC state while undergoing EMT. Secondly, EMT increases the probability of symmetric, self-renewal division of cells that are already stem-like [76]. The authors used the model to fit the experimental data on CSC fraction and mammosphere expansion, indicating that both processes may play an important role in supporting cancer progression [76]. Later, Gupta et al. [77••] showed that breast cancer cells SUM149 and SUM159 can exist in different subpopulations with varying functional attributes: luminal, basal, and stem-like. They demonstrated that the overall population, when perturbed, re-establishes a fixed fraction of the three cell phenotypes. This robustness could be explained by a population model where cells can undergo stochastic phenotypic transitions between the three different states [77••]. Moreover, the stem-like cell line SUM 149 has been shown to exhibit the traits of a hybrid E/M phenotype, hence suggesting a possible correlation between a partial EMT state and stemness [33].

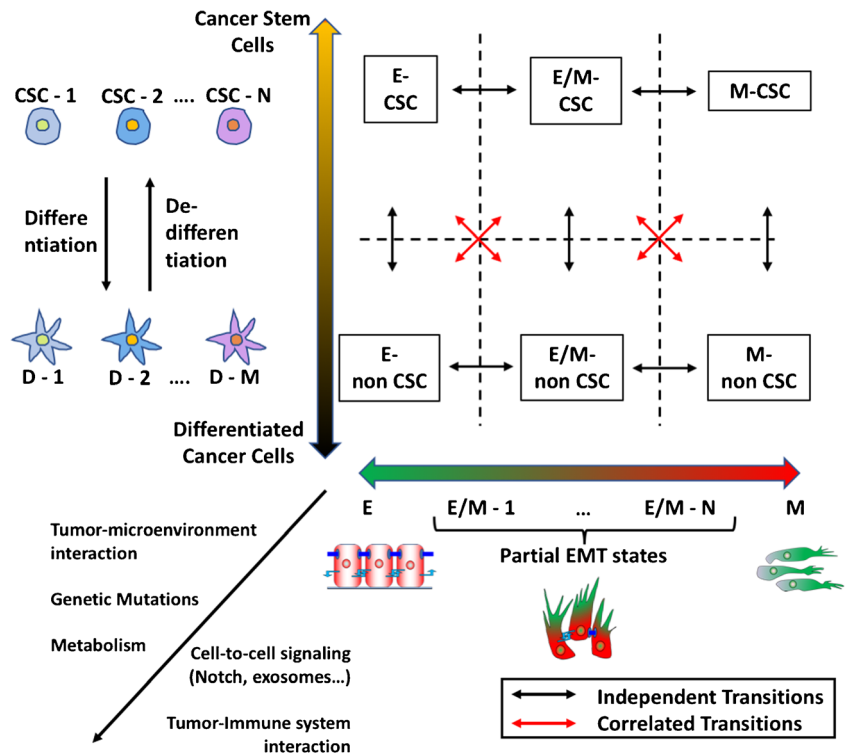
The multi-scale model proposed by Sfakianakis et al. [78], instead, focuses on resolving the spatial structure of a cancer cell population. This phenomenological model couples the aspects of CSC and EMT to describe the invasion of extracellular matrix by tumor cells. In this framework, EMT is modeled as a binary switch between an epithelial-like and a mesenchymal-like phenotype that is driven by growth factors. Therefore, this model couples EMT at the individual cell scale and the population dynamics and growth of the tumor mass at the multi-cell scale. Note, however, that the models discussed

so far proposed mechanisms for CSC-driven tumor progression and maintenance of the CSC fraction but did not provide a molecular rationale for the acquisition of CSC traits.

Li and Wang [38] reconstructed a core gene regulatory circuit with relevant players determining CSC properties such as miR-145 and OCT4 and core regulators of EMT—miR-200 and ZEB. The authors applied an energy landscape approach to predict the co-existence of multiple cellular phenotypes. In their model, a cell can either assume a “normal” state or a “cancer” state, both of which could or could not exhibit stem-like traits. Thus, a total of four possible cell phenotypes are available—normal, normal stem-like, cancer, and cancer stem-like [38]. In this framework, p53 represents a degree of cancerization and ZEB represents a degree of stemness. Notably, both the predicted “normal stem cell” state and the “CSC” state highly express ZEB, hence implicitly suggesting that stemness is gained along with EMT [38].

Finally, the models developed by Jolly and colleagues explicitly proposed a mechanism-based rationale to elucidate the connection between EMT and CSC: the stemness circuit comprising LIN28, let-7, and OCT4 is connected to the EMT circuit already discussed by Lu et al. [25•] (Fig. 1e, left). The CSC phenotype was defined as a state with intermediate levels of OCT4 that have been shown experimentally to correlate with stem-like traits [79, 80]. These models proposed that a CSC phenotype is highly correlated with a hybrid E/M phenotype [81•], but intracellular factors such as OVOL [12] or cell-cell communication via Notch signaling [13•] could move the predicted “stemness window” toward the epithelial or mesenchymal ends of the EMT spectrum (Fig. 1e, right). Experimental evidence for this dynamic stemness window concept was provided by Bocci et al. [13•] by computing the “EMT score” [23] of different human CSC lines using publicly available datasets. This analysis showed that CSC traits can be scattered along the EMT spectrum based on context-specific activation of signaling pathways, therefore resulting in epithelial, hybrid E/M, and mesenchymal CSC (Fig. 1f) [13•]. Furthermore, this model proposed a strong overlap between a hybrid E/M phenotype, CSC properties, and Notch-Jagged signaling [13•], a pathway implicated in both drug resistance and in clusters of CTCs, the key drivers of metastasis [33]. Consistently, knockdown of Jagged was shown to restrict the growth of tumor emboli in SUM149 inflammatory breast cancer cells [82••]. Given the role of Notch signaling in pattern formation in multiple contexts [83, 84], Notch signaling coupled with EMT circuitry may underlie the spatial segregation of different subsets of cells with stem-like traits, as observed experimentally in a breast cancer tissue [85]. Secretion of a diffusible EMT-inducing signal at the tumor-stroma interface (such as TGF- $\beta$ ), along with cell-cell signaling through Notch, was shown to give rise to mesenchymal CSCs at the invasive edge of the tumor and a population of hybrid epithelial-mesenchymal (E/M) CSCs in the tumor

**Fig. 2** Interconnections between EMT and CSC axes. The spectrum of EMT states can range from epithelial (E) to mesenchymal (M) phenotypes, with a variable number of partial E/M states (x-axis). Both cancer stem cells and differentiated cancer cells can assume different states depending on genetic and epigenetic factors (y-axis). Horizontal and vertical transitions represent independent EMT and stem processes, respectively. However, transitions are possible where both the EMT and stem states change, or correlated transition. The interconnection among EMT and CSC states and the transitions enabled between them depend on context-specific factors including, among others, intracellular signaling and mutations, cell-cell, and cell-environment signaling



interior [82••]. The idea that cell-cell signaling and the micro-environment can shape the spatial distribution of a cell population has been examined in different biological contexts, such as bacterial colonies [86, 87] or eukaryotic chemotaxis [88, 89], but remains largely unexplored in cancer biology, and thus demands further attention.

**Conclusion**

EMT and CSC represent two crucial biological axes that bolster tumor progression, metastasis, and tumor relapse [2, 4]. While the molecular details of multiple steps of tumor development continue to be identified, it is largely accepted that EMT often plays a crucial role in regulating epigenetic, morphological, and functional cell properties during tumor progression and metastasis formation [2, 4]. Similarly, it is well accepted that the acquisition of stem-like properties potentiates tumor maturation and enhances resistance to various treatments, driving tumor relapse. Only recently, we have been gaining insights into how, when, and where these two dynamic processes can influence one another (Fig. 2). In this context, mathematical modeling has proven itself as a potent tool to interpret existing data and formulate new predictions that can be tested experimentally.

In the context of EMT, mechanism-based computational models have suggested that cells undergoing EMT can stably acquire intermediate cell states enabling hybrid phenotypes with mixed epithelial (E) and mesenchymal (M) characters,

as opposed to a binary E/M switch scenario [2, 4]. Novel in vivo and in vitro analysis recently highlighted the existence of such hybrid states that coexpress E and M markers and often possess mixed morphological traits of cell-cell adhesion and motility [18••, 19•] and have highlighted their enhanced metastatic potential [90]. The next crucial steps will include a more comprehensive attempt to integrate data-based models, mechanism-based models, and time-course and single-cell experimental data, to formulate a more quantitative characterization of these malignant hybrid E/M state(s).

In the context of CSC, one set of models considers the dynamics of a CSC population employing the tools of population dynamics and agent-based modeling. Such class of models can provide predictions about CSC fraction or population dynamics under perturbations, hence potentially providing strategies for containing CSC-driven tumor progression. Additionally, coupling mathematical modeling with clinical data of therapy response enables predictive tools that can shed light on the CSC-therapy interplay. Such models can provide information on, among others, adaptive response, differential drug sensitivity, or phenotypic plasticity in a cancer cell population.

Recent experimental observations have led to a class of models that can offer insights into the coupling between EMT in cancer cells and the acquisition of stem-like properties. A first set of models relates phenomenologically the acquisition of stem traits with the EMT process, hence explaining how CSC-EMT interaction can support tumor progression and maintain a certain fraction of different cell phenotypes. Moreover, a second class of models investigates the

coupling between EMT and CSC at the level of gene regulatory networks, showing a correlation between the cell phenotypes enabled by an EMT regulatory circuit and the stemness regulatory circuit. A common feature across these models is envisioning the acquisition of stemness as a dynamical process correlated with EMT [12, 13, 38]. Recent mathematical modeling and experiments have suggested a correlation among hybrid E/M states and stem cell properties [5, 12, 13, 18, 19]. CSC traits, however, are not exclusively observed in intermediate states, and the crosstalk between tumor, microenvironment, and therapies is likely to play a major role in modulating the plasticity properties of cancer cells (Fig. 2), as shown by recent experiments highlighting subsets of CSCs in multiple cancer types [20, 85].

Considered together, these computational models developed for EMT, CSC, or their interconnections have contributed not only in deciphering the mechanisms underlying specific experimental observations but also have driven the next set of experiments by generating testable predictions. Such bidirectional crosstalk between theory and experiments can significantly accelerate our goal of understanding and consequently targeting these processes for therapeutic benefit.

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## Compliance with Ethical Standards

**Conflict of Interest** Federico Bocci, Herbert Levine, José N Onuchic, and Mohit Kumar Jolly declare that they have no conflict of interest.

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## References

Papers of particular interest, published recently, have been highlighted as:

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1. Fabisiwicz A, Grzybowska E, Grybowska E. CTC clusters in cancer progression and metastasis. *Med Oncol*. 2017;34:12.
2. Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT: 2016. *Cell*. 2016;166:21–45.
3. Gupta GP, Massagué J. Cancer metastasis: building a framework. *Cell*. 2006;127:679–95.
4. Jolly MK, Boareto M, Huang B, Jia D, Lu M, Ben-Jacob E, et al. Implications of the hybrid epithelial/mesenchymal phenotype in metastasis. *Front Oncol*. 2015;5:155.
5. Grosse-Wilde A, d'Hérouël AF, McIntosh E, Ertaylan G, Skupin A, Kuestner RE, et al. Stemness of the hybrid epithelial/mesenchymal state in breast cancer and its association with poor survival. *PLoS One*. 2015;10:e0126522.
6. Cheung KJ, Ewald AJ. A collective route to metastasis: seeding by tumor cell clusters. *Science*. 2016;352:167–9.
7. Tannishtha R, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature*. 2001;414:105–11.
8. Bjerkvig R, Tysnes BB, Aboody KS, Najbauer J, Terzis AJA. Opinion: the origin of the cancer stem cell: current controversies and new insights. *Nat Rev Cancer*. 2005;5:899–904.
9. Avgustinova A, Benitah SA. Epigenetic control of adult stem cell function. *Nat Rev Mol Cell Biol*. 2016;17:643–58.
10. Iliopoulos D, Hirsch HA, Wang G, Struhl K. Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. *Proc Natl Acad Sci*. 2011;108:1397–402.
11. Yang G, Quan Y, Wang W, Fu Q, Wu J, Mei T, et al. Dynamic equilibrium between cancer stem cells and non-stem cancer cells in human SW620 and MCF-7 cancer cell populations. *Br J Cancer*. 2012;106:1512–9.
12. Jolly MK, Jia D, Boareto M, Mani SA, Pienta KJ, Ben-Jacob E, et al. Coupling the modules of EMT and stemness: a tunable 'stemness window' model. *Oncotarget*. 2015;6:25161–74.
13. Bocci F, Jolly MK, George JT, Levine H, Onuchic JN. A mechanism-based computational model to capture the interconnections among epithelial-mesenchymal transition, cancer stem cells and Notch-Jagged signaling. *Oncotarget*. 2018;9:29906–20 **This study proposes a mathematical framework to couple EMT, CSC and Notch signaling, and predicted a 'plasticity window' characterized by a hybrid E/M phenotype, stem-like properties and enhanced Notch-Jagged signaling. It further evaluates the EMT phenotype of CSCs from different cancer cells to identify signaling that can give rise to E-like or M-like CSC.**
14. Brabletz T, Jung A, Spaderna S, Hlubek F, Kirchner T. Opinion: migrating cancer stem cells - an integrated concept of malignant tumour progression. *Nat Rev Cancer*. 2005;5:744–9.
15. May CD, Sphyris N, Evans KW, Werden SJ, Guo W, Mani SA. Epithelial-mesenchymal transition and cancer stem cells: a dangerously dynamic duo in breast cancer progression. *Breast Cancer Res*. 2011;13:202.
16. Shibue T, Weinberg RA. EMT, CSCs, and drug resistance: the mechanistic link and clinical implications. *Nat Rev Clin Oncol*. 2017;14:611–29.
17. Bierie B, Pierce SE, Kroeger C, Stover DG, Pattabiraman DR, Thiru P, et al. Integrin-β4 identifies cancer stem cell-enriched populations of partially mesenchymal carcinoma cells. *Proc Natl Acad Sci*. 2017;114:E2337–46.
18. Pastushenko I, Brisebarre A, Sifrim A, Fioramonti M, Revenco T, Boumahdi S, et al. Identification of the tumour transition states occurring during EMT. *Nature*. 2018;556:463–8 **This work reports, for the first time, spontaneous emergence of multiple hybrid E/M states *in vivo*, and further showed that those intermediate states are characterized by increased metastatic potential and stem-like properties.**
19. Hojo N, Huisken AL, Wang H, Chirshv E, Kim NS, Nguyen SM, Campos H. Snail knockdown reverses stemness and inhibits tumour growth in ovarian cancer. *Sci Rep*. 2018;8: 8704. **This study**



- shows that EMT inhibition via SNAIL knockdown results in loss of tumor-initiation ability in ovarian cancer cell lines.**
20. Biddle A, Gammon L, Liang X, Costea DE, Mackenzie IC. Phenotypic plasticity determines cancer stem cell therapeutic resistance in oral squamous cell carcinoma. *EBioMedicine*. 2016;4:138–45.
  21. Fustaino V, Presutti D, Colombo T, Cardinali B, Papoff G, Brandi R, et al. Characterization of epithelial-mesenchymal transition intermediate/hybrid phenotypes associated to resistance to EGFR inhibitors in non-small cell lung cancer cell lines. *Oncotarget*. 2017;8:103340–63.
  22. Andriani F, Bertolini G, Facchinetti F, Baldoli E, Moro M, Casalini P, et al. Conversion to stem-cell state in response to microenvironmental cues is regulated by balance between epithelial and mesenchymal features in lung cancer cells. *Mol Oncol*. 2016;10:253–71.
  23. George JT, Jolly MK, Xu S, Somarelli JA, Levine H. Survival outcomes in cancer patients predicted by a partial EMT gene expression scoring metric. *Cancer Res*. 2017;77:6415–28.
  24. De Craene B, Bex G. Regulatory networks defining EMT during cancer initiation and progression. *Nat Rev Cancer*. 2013;13:97–110.
  25. Lu M, Jolly MK, Levine H, Onuchic JN, Ben-Jacob E. MicroRNA-based regulation of epithelial-hybrid-mesenchymal fate determination. *Proc Natl Acad Sci U S A*. 2013;110:18174–9 **This work develops a mathematical description of transcriptional and micro-RNA mediated (post-translational) interactions to model a core EMT regulatory circuit that can exhibit an intermediate, or hybrid E/M, phenotype.**
  26. Tian XJ, Zhang H, Xing J. Coupled reversible and irreversible bistable switches underlying TGF $\beta$ -induced epithelial to mesenchymal transition. *Biophys J*. 2013;105:1079–89.
  27. Zhang J, Tian X, Zhang H, Teng Y, Li R, Bai F. TGF- $\beta$ -induced epithelial-to-mesenchymal transition proceeds through stepwise activation of multiple feedback loops. *Sci Signal*. 2015;7:ra91. **This study presents a dose-response to TGF- $\beta$  treatment and maps it onto the bifurcation diagram of EMT phenotypes.**
  28. Abshire CF, Carroll JL, Dragoi A-M. FLASH protects ZEB1 from degradation and supports cancer cells' epithelial-to-mesenchymal transition. *Oncogenesis*. 2016;5:e254.
  29. Ruscetti M, Dadashian EL, Guo W, Quach B, Mulholland DJ, Park JW, et al. HDAC inhibition impedes epithelial-mesenchymal plasticity and suppresses metastatic, castration-resistant prostate cancer. *Oncogene*. 2016;35:3781–95.
  30. Jia D, Jolly MK, Tripathi SC, Den Hollander P, Huang B, Lu M, et al. Distinguishing mechanisms underlying EMT tristability. *Cancer Converg*. 2017;1:2.
  31. Jia D, Jolly MK, Boareto M, Parsana P, Mooney SM, Pienta KJ, et al. OVOL guides the epithelial-hybrid-mesenchymal transition. *Oncotarget*. 2015;6:15436–48.
  32. Jolly MK, Tripathi SC, Jia D, Mooney SM, Celiktas M, Hanash SM, et al. Stability of the hybrid epithelial/mesenchymal phenotype. *Oncotarget*. 2016;7:27067–84 **This study identifies the stably hybrid E/M cell line H1975 and proposed different interactions between a core EMT circuit and different molecular factors (including OVOL2 and GRHL2) that help stabilize hybrid E/M phenotype(s).**
  33. Jolly MK, Boareto M, Debeb BG, Aceto N, Farach-Carson MC, Woodward WA, et al. Inflammatory breast cancer: a model for investigating cluster-based dissemination. *NPJ Breast Cancer*. 2017;3:21.
  34. Hong T, Watanabe K, Ta CH, Villarreal-Ponce A, Nie Q, Dai X. An *Ovol2-Zeb1* mutual inhibitory circuit governs bidirectional and multi-step transition between epithelial and mesenchymal states. *PLoS Comput Biol*. 2015;11(11):e1004569.
  35. Bocci F, Tripathi SC, Mercedes SV, George JT, Casabar J, Wong PK, Hanash S, Levine H, Onuchic JN, Jolly MK. NRF2 activates a partial epithelial-mesenchymal transition and is maximally present in a hybrid epithelial/mesenchymal phenotype. *bioRxiv*. 2018;390237. **This study shows via modeling and *in vitro* experiments that NRF2 activation stabilizes a partial EMT, and therefore proposes NRF2 as a hallmark of hybrid E/M phenotype(s).**
  36. Watanabe K, Villarreal-Ponce A, Sun P, Salmans ML, Fallahi M, Andersen B, et al. Mammary morphogenesis and regeneration require the inhibition of EMT at terminal end buds by *ovol2* transcriptional repressor. *Dev Cell*. 2014;29:59–74.
  37. Li C, Hong T, Nie Q. Quantifying the landscape and kinetic paths for epithelial-mesenchymal transition from a core circuit. *Phys Chem Chem Phys*. 2016;18:17949–56.
  38. Li C, Wang J. Quantifying the landscape for development and cancer from a core cancer stem cell circuit. *Cancer Res*. 2015;75:2607–18.
  39. Boareto M, Jolly MK, Goldman A, Pietilä M, Mani SA, Sengupta S, et al. Notch-jagged signalling can give rise to clusters of cells exhibiting a hybrid epithelial/mesenchymal phenotype. *J R Soc Interface*. 2016;13:20151106.
  40. Bocci F, Jolly MK, Tripathi SC, Aguilar M, Onuchic N, Hanash SM, et al. Numb prevents a complete epithelial – mesenchymal transition by modulating Notch signalling. *J R Soc Interface*. 2017;14:20170512.
  41. Kikuchi H, Sakakibara-konishi J, Furuta M, Kikuchi E. Numb has distinct function in lung adenocarcinoma and squamous cell carcinoma. *Oncotarget*. 2018;9:29379–91.
  42. Cohen DPA, Martignetti L, Robine S, Barillot E, Zinovyev A, Calzone L. Mathematical modelling of molecular pathways enabling tumour cell invasion and migration. *PLoS Comput Biol*. 2015;11:e1004571.
  43. Steinway SN, Zanudo JGT, Ding W, Rountree CB, Feith DJ, Loughran TP, et al. Network modeling of TGF- $\beta$  signaling in hepatocellular carcinoma epithelial-to-mesenchymal transition reveals joint sonic hedgehog and Wnt pathway activation. *Cancer Res*. 2014;74:5963–77.
  44. Steinway SN, Zañudo JGT, Michel PJ, Feith DJ, Loughran TP, Albert R. Combinatorial interventions inhibit TGF $\beta$ -driven epithelial-to-mesenchymal transition and support hybrid cellular phenotypes. *NPJ Syst Biol Appl*. 2015;1:15014.
  45. Font-Clos F, Zapperi S, La Porta CAM. Topography of epithelial-mesenchymal plasticity. *Proc Natl Acad Sci*. 2018;115(23):5902–7 **This study constructs a large EMT network from literature and predicts multiple intermediate states associated with hybrid E/M phenotypes.**
  46. Huang B, Lu M, Jia D, Ben-Jacob E, Levine H, Onuchic JN. Interrogating the topological robustness of gene regulatory circuits by randomization. *PLoS Comput Biol*. 2017;13:e1005456.
  47. Kohar V, Lu M. Role of noise and parametric variation in the dynamics of gene regulatory circuits. *NPJ Syst Biol Appl*. 2018;4(40).
  48. Zadran S, Arumugam R, Herschman H, Phelps ME, Levine RD. Surprisal analysis characterizes the free energy time course of cancer cells undergoing epithelial-to-mesenchymal transition. *Proc Natl Acad Sci*. 2014;111:13235–40.
  49. Chang H, Liu Y, Xue M, Liu H, Du S, Zhang L, et al. Synergistic action of master transcription factors controls epithelial-to-mesenchymal transition. *Nucleic Acids Res*. 2016;44:2514–27.
  50. Tan TZ, Miow QH, Miki Y, Noda T, Mori S, Huang RY-J, et al. Epithelial-mesenchymal transition spectrum quantification and its efficacy in deciphering survival and drug responses of cancer patients. *EMBO Mol Med*. 2014;6:1279–93.
  51. Mandal M, Ghosh B, Anura A, Mitra P, Pathak T, Chatterjee J. Modeling continuum of epithelial mesenchymal transition plasticity. *Integr Biol*. 2016;8:167–76.

52. Leggett SE, Sim JY, Rubins JE, Neronha ZJ, Williams EK, Wong IY. Morphological single cell profiling of the epithelial-mesenchymal transition. *Integr Biol*. 2016;8:1133–44.
53. Zhang Z, Chen L, Humphries B, Brien R, Wicha M, Luker KE, Luker G, Chen Y-C, Yoon E. Morphology-based prediction of cancer cell migration using artificial neural network and random decision Forest. *Integr Biol*. 2018; 10:758–67.
54. Jolly MK, Ware KE, Gilja S, Somarelli JA, Levine H. EMT and MET: necessary or permissive for metastasis? *Mol Oncol*. 2017;11:755–69.
55. Michor F. Mathematical models of cancer stem cells. *J Clin Oncol*. 2008;26:2854–61.
56. Enderling H. Cancer stem cells: small subpopulation or evolving fraction? *Integr Biol (United Kingdom)*. 2015;7:14–23.
57. Dhawan A, Madani Tonekaboni SA, Taube JH, Hu S, Sphyris N, Mani SA, et al. Mathematical modelling of phenotypic plasticity and conversion to a stem-cell state under hypoxia. *Sci Rep*. 2016;6:18074.
58. Jilkine A, Gutenkunst RN. Effect of dedifferentiation on time to mutation acquisition in stem cell-driven cancers. *PLoS Comput Biol*. 2014;10(3):e1003481.
59. Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even Warburg did not anticipate. *Cancer Cell*. 2012;21:297–308.
60. Liu LL, Brumbaugh J, Bar-Nur O, Smith Z, Stadtfeld M, Meissner A, et al. Probabilistic modeling of reprogramming to induced pluripotent stem cells. *Cell Rep*. 2016;17:3395–406.
61. Luo M, Shang L, Brooks MD, Jiagge E, Zhu Y, Buschhaus JM, et al. Targeting breast cancer stem cell state equilibrium through modulation of redox signaling. *Cell Metab*. 2018;28:69–86.
62. Zhou JX, Pisco AO, Qian H, Huang S. Nonequilibrium population dynamics of phenotype conversion of cancer cells. *PLoS One*. 2014;9:e110714.
63. Wang W, Quan Y, Fu Q, Liu Y, Liang Y, Wu J, et al. Dynamics between cancer cell subpopulations reveals a model coordinating with both hierarchical and stochastic concepts. *PLoS One*. 2014;9:e84654.
64. Zhou D, Wang Y, Wu B. A multi-phenotypic cancer model with cell plasticity. *J Theor Biol*. 2014;357:35–45.
65. Poleszczuk J, Hahnfeldt P, Enderling H. Evolution and phenotypic selection of cancer stem cells. *PLoS Comput Biol*. 2015;11:e1004025.
66. Michor F, Beal K. Improving cancer treatment via mathematical modeling: surmounting the challenges is worth the effort. *Cell*. 2015;163:1059–63.
67. Goldman A, Majumder B, Dhawan A, Ravi S, Goldman D, Kohandel M, et al. Temporally sequenced anticancer drugs overcome adaptive resistance by targeting a vulnerable chemotherapy-induced phenotypic transition. *Nat Commun*. 2015;6:6139. **This study shows that cancer cells can undergo phenotypic transitions in response to drug treatment and the vulnerability of this adaptive state can then be exploited via combinatorial therapies.**
68. Werner B, Scott JG, Sottoriva A, Anderson ARA, Traulsen A, Altrock PM. The cancer stem cell fraction in hierarchically organized tumors can be estimated using mathematical modeling and patient-specific treatment trajectories. *Cancer Res*. 2016;76:1705–13.
69. Zhou D, Mao S, Cheng J, Chen K, Cao X, Hu J. A Bayesian statistical analysis of stochastic phenotypic plasticity model of cancer cells. *J Theor Biol*. 2018;454:70–9.
70. Yu VY, Nguyen D, Pajonk F, Kupelian P, Kaprealian T, Selch M, et al. Incorporating cancer stem cells in radiation therapy treatment response modeling and the implication in glioblastoma multiforme treatment resistance. *Int J Radiat Oncol Biol Phys*. 2015;91:866–75.
71. Chen C, Baumann WT, Xing J, Xu L, Clarke R, Tyson JJ. Mathematical models of the transitions between endocrine therapy responsive and resistant states in breast cancer. *J R Soc Interface*. 2014;11(96):20140206.
72. Kulkarni P, Jolly MK, Jia D, Mooney SM, Bhargava A, Kagohara LT, et al. Phosphorylation-induced conformational dynamics in an intrinsically disordered protein and potential role in phenotypic heterogeneity. *Proc Natl Acad Sci*. 2017;114:E2644–53.
73. Lin X, Roy S, Jolly MK, Bocci F, Schafer NP, Tsai M-Y, et al. PAGE4 and conformational switching: insights from molecular dynamics simulations and implications for prostate Cancer. *J Mol Biol*. 2018;430:2422–38.
74. Hatina J. The dynamics of cancer stem cells. *Neoplasma*. 2012;59:700–7.
75. Nazari F, Pearson AT, Nör JE, Jackson TL. A mathematical model for IL-6-mediated, stem cell driven tumor growth and targeted treatment. *PLoS Comput Biol*. 2018;14(1):e1005920.
76. Turner C, Kohandel M. Investigating the link between epithelial-mesenchymal transition and the cancer stem cell phenotype: a mathematical approach. *J Theor Biol*. 2010;265:329–35.
77. Gupta PB, Fillmore CM, Jiang G, Shapira SD, Tao K, Kuperwasser C, et al. Stochastic state transitions give rise to phenotypic equilibrium in populations of cancer cells. *Cell*. 2011;146:633–44 **This study reported and explain with mathematical modeling that three main populations of breast cancer cells maintain an equilibrium fraction upon perturbation, and further shows generation of stem-like breast cancer cells from non-stem cells.**
78. Sfakianakis N, Kolbe N, Hellmann N, Lukacova-Medvid'ova M. A multiscale approach to the migration of cancer stem cells: mathematical modelling and simulations. *Bull Math Biol*. 2017;79:209–35.
79. Karwacki-Neisius V, Göke J, Osorno R, Halbritter F, Ng JH, Weiße AY, et al. Reduced Oct4 expression directs a robust pluripotent state with distinct signaling activity and increased enhancer occupancy by Oct4 and Nanog. *Cell Stem Cell*. 2013;12:531–45.
80. Niwa H, Miyazaki J, Smith AG. Quantitative expression of Oct-3 / 4 defines differentiation, dedifferentiation or self-renewal of ES cells. *Nat Genet*. 2016;24:2–6.
81. Jolly MK, Huang B, Lu M, Mani SA, Levine H, Ben-Jacob E. Towards elucidating the connection between epithelial-mesenchymal transitions and stemness. *J R Soc Interface*. 2014;11:20140962 **This study proposes a mechanism-based mathematical framework that couples the genetic circuits that regulate EMT and CSC, and first proposes the association between hybrid E/M phenotypes and stemness.**
82. Bocci F, Gaerhart-Serna L, Ribeiro M, Boareto M, Ben-Jacob E, Devi G, Levine H, Onuchic JN, Jolly MK. Toward understanding Cancer stem cell heterogeneity in the tumor microenvironment. *Proc Natl Acad Sci U S A*. 2019; 116: 148–57. **This study models the interplay among Notch signaling, TGF-beta signaling and inflammation in the tumor microenvironment to explain the spatial patterning of CSC with different EMT phenotypes in a tumor tissue reported in Ref. 85.**
83. Shaya O, Sprinzak D. From notch signaling to fine-grained patterning: modeling meets experiments. *Curr Opin Genet Dev*. 2011;21:732–9.
84. Boareto M, Jolly MK, Lu M, Onuchic JN, Clementi C, Ben-Jacob E. Jagged-Delta asymmetry in notch signaling can give rise to a sender/receiver hybrid phenotype. *Proc Natl Acad Sci*. 2015;112:402–9.
85. Liu S, Cong Y, Wang D, Sun Y, Deng L, Liu Y, et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. *Stem Cell Rep*. 2014;2:78–91.
86. Nadell CD, Drescher K, Foster KR. Spatial structure, cooperation and competition in biofilms. *Nat Rev Microbiol*. 2016;14:589–600.

87. Bocci F, Suzuki Y, Lu M, Onuchic JN. Role of metabolic spatio-temporal dynamics in regulating biofilm colony expansion. *Proc Natl Acad Sci U S A*. 2018;115:4288–93.
88. Ellison D, Mugler A, Brennan MD, Lee SH, Huebner RJ, Shamir ER, et al. Cell–cell communication enhances the capacity of cell ensembles to sense shallow gradients during morphogenesis. *Proc Natl Acad Sci*. 2016;113:E679–88.
89. Levine H, Kessler DA, Rappel W-J. Directional sensing in eukaryotic chemotaxis: a balanced inactivation model. *Proc Natl Acad Sci*. 2006;103:9761–6.
90. Jolly MK, Mani SA, Levine H. Hybrid epithelial/mesenchymal phenotype(s): the ‘fittest’ for metastasis? *BBA-Rev Cancer*. 2018. 1870; 151-7.