Anaphylaxis (M Sánchez Borges, Section Editor)

Drug-Induced Anaphylaxis

Natalia Blanca-López, MD, PhD¹
María del Carmen Plaza-Serón, BSc²
José Antonio Cornejo-García, PhD^{2,3}
James Richard Perkins, PhD²
Gabriela Canto, MD, PhD¹
Miguel Blanca, MD, PhD^{3,4,*}

Address

¹Allergy Service, Infanta Leonor Hospital, Madrid, Spain

²Research Laboratory, IBIMA, Regional University Hospital of Malaga, UMA, Malaga, Spain

³Allergy Unit, IBIMA, Regional University Hospital of Malaga, UMA, Málaga, Spain * ⁴Allergy Unit, Plaza del Hospital Civil s/n, Hospital Civil, Pabellón 6, 1° planta,

29009, Málaga, Spain Email: mblancago@gmail.com

Published online: 2 June 2015

© Springer International Publishing AG 2015

This article is part of the Topical Collection on Anaphylaxis

 $\textbf{Keywords} \ \ \textbf{Drug-induced} \ \ \textbf{anaphylaxis} \cdot \ \ \textbf{Beta-lactams} \cdot \ \ \textbf{NSAIDs} \cdot \ \ \textbf{Immunological} \ \ \textbf{reactions} \cdot \ \ \textbf{Non-immunological} \ \ \textbf{reactions}$

Opinion statement

Drug-induced anaphylaxis can be a life-threatening condition occurring after drug intake by any route, including non-therapeutic exposure due to accidental contact. The most common culprit drugs are non-steroidal anti-inflammatory drugs and beta-lactam antibiotics. Other antibiotics, such as guinolones, are also becoming important elicitors. In the hospital setting, neuromuscular blocking agents and contrast media play an important role. In addition to specific immunological mechanisms (IgE-mediated), other non-immunological mechanisms may participate, including the generation of vasoactive mediators through the arachidonic acid metabolic pathway, direct stimulation of mast cells and the activation of other inflammatory cascades. Diagnosis of drug-induced anaphylaxis includes skin testing, in vitro testing in some instances, and very often requires drug provocation to search for alternatives or in some cases for diagnosis confirmation. Often, the diagnosis must be performed in the context of confounding agents. Factors contributing to the degree of severity include previous diseases (e.g. cardiovascular disease) and the intake of other drugs such as betablockers. In circumstances where the culprit drug must be administered, patients should be desensitised if possible. This may be carried out for drug-induced anaphylaxis caused by both immunological and non-immunological reactions, such as cross-intolerance to NSAIDs.

Introduction

Drug-induced anaphylaxis (DIA) consists of a systemic reaction occurring as the result of a quick release of histamine and other vasoactive mediators with multiple effects in different organs and can be life threatening [1]. In the past, several terminologies have been used to identify this clinical entity. The term DIA should be applied to all cases where acute symptoms indicative of a hypersensitivity episode appear after drug intake [2]. If the mechanism is IgE-mediated, we are dealing with allergic anaphylaxis. Non-allergic anaphylaxis was previously known as anaphylactoid; anaphylactic-like or pseudoallergic reactions and are now referred to as nonallergic hypersensitivity [3]. In adults the most common agents involved in anaphylaxis are drugs, whereas for children drugs are the second most common cause after food allergens, although in some studies, they both carry equal weight $[1, 4, 5 \bullet \bullet]$.

Nowadays, there is great concern about the high prevalence of allergic diseases in general [6], including drug allergy and DIA [7.0, 8]. Although the responsible agents for DIA can include vaccines [9] and biological agents [10] including humanised antibodies [11, 12], for didactic purposes and in order to reduce complexity, in this review, we will deal with anaphylactic reactions induced by drugs of small molecular weight (<1000 kDa). These are organic structures that require

protein binding and multivalence to induce an immunological response [1]. We will also omit anti-neoplastic drugs and direct the readers to reviews in the literature dealing with this problem [13]. Given that over 80 % of anaphylactic drug reactions occur in ambulatory settings [14], we will therefore focus on the most relevant drugs involved: beta-lactam antibiotics (BLs) and non-steroidal anti-inflammatory drugs (NSAIDs).

From a mechanistic point of view, specific IgEmediated DIA occurs very soon after contact has been made between the drug and the immunological system, through cross-linking of IgE antibodies at the surface of mast cells/basophils and the subsequent triggering of a mechanism that leads to the release of inflammatory mediators that induce the episode. The kinetics of this process is very well established [1]. In general, subjects react to therapeutic or very low concentrations of the drug that often occur after an inadvertent exposure [1, 15]. Some IgE-mediated DIA reactions can occur after a longer time period, for example, when the culprits are drug metabolites, where the drug must be metabolised before IgE recognition can occur [16]. In terms of administration, all routes can lead to DIA; however, reactions tend to be more severe when the agent is administered by parenteral route $[1, 7 \bullet \bullet]$.

Epidemiology and Drugs Involved

DIA is responsible for over 230,000 hospital admissions in the USA annually and prevalence is increasing [1, $5 \bullet \bullet$, 8]. In a study of 2458 anaphylaxis-related deaths in the USA between 1999 and 2010, medications were the most common cause, responsible for more than 50 % of the cases, followed by unspecified triggers, venoms and food. A significant increase over recent years was also noted [$5 \bullet \bullet$]. Very often, anaphylaxis occurs in ambulatory practice and figures are likely to be underreported [17].

For many years, antibiotics (in particular BLs) have been considered the most frequent culprit for DIA [14]; however, recent studies have questioned this view. In a nationwide study carried out by allergists in Portugal during a 4-year period, NSAIDs were shown to be the leading cause, followed by antibiotics. The mean age at which the reaction occurred ranged from 2 to 89 years, with a mean of 17.4, indicating that DIA may appear at any age [18•].

There is often disagreement between population-based surveys and studies carried out in specialised centres. The former generally find antibiotics as the most frequent cause of hypersensitivity drug reactions including anaphylaxis [14]; however, in specialised centres, NSAIDs predominate over BLs [$7 \bullet \bullet$]. In a

study carried out by Messaad et al., around 10 % of subjects evaluated for penicillin allergy were confirmed as such, compared to over 20 % for NSAIDs [19]. Similar data have been recently reported by Doña et al. [7••].

Concerning neuromuscular blocking agents, a study carried out by Reiter et al. consulting data from the French National Pharmacovigilance Database showed that from 2022 cases registered, 70 % developed anaphylaxis at grades 3 and 4 [20•].

Within NSAID hypersensitivity reactions, we must differentiate two main groups of patients, those who are cross-intolerant and those who are selective responders [21••]. Both groups can show anaphylaxis, although it is significantly more frequent amongst the latter [22, 23]. Out of all drug reactions attended to an emergency department, anaphylaxis to NSAIDs has been shown to be the most common entity [24•]; however, the most severe reactions are likely to be induced by BLs. Of these, amoxicillin is the most frequent culprit in many countries [25, 26], although clavulanic acid is becoming more prevalent in the induction of immediate hypersensitivity BL reactions including anaphylaxis [27, 28].

Case series, particularly hospital surveys, have shown muscle relaxants to be also relevant for an important number of drug reactions [14] as well as contrast media [4], which are potential inducers of anaphylaxis [4, 5••]. Other drugs that can elicit DIA include proton pump inhibitors [29, 30•], quinolones [31], heparins [32], codeine [33], amiodarone [34], *N*-acetyl cysteine [35], chlorhexidine [36], vitamins [37–39] and corticoids [40–42]. Many of these drugs are frequently prescribed and cannot be easily substituted, warranting further study of these reactions.

Underlying Mechanisms

Anaphylaxis involves the abrupt release of pro-inflammatory mediators with vasoactive properties that induce an often severe systemic response involving many organs. It has long been known that in addition to IgE-mediated responses, other mechanisms can be involved [1].

The most highly studied model of DIA mediated by specific immunological mechanisms is that of penicillin. The availability of diagnostic tests and determination of specific IgE antibodies has made it possible to study this model in great detail [1]. Clinical experience with such patients indicates that reactions can be induced by very low concentrations of drugs through the oral route, skin testing or inadvertent exposures [43–45]. If a subject takes amoxicillin or another BL derivative and develops an anaphylactic response after an interval of over 1 h, we are likely dealing with an accelerated reaction for which the proposed mechanism is a T cell response [46•]. In a study carried out by Torres et al., it was shown that for those patients who developed a positive response after amoxicillin challenge, the longer the interval between intake and reaction, the greater the dose required to elicit a positive response, usually consisting of urticaria with some systemic manifestations [44].

The situation is more complex for NSAIDs. According to the recent classification proposed by the ENDA, an anaphylactic reaction to dipyrone or diclofenac, two drugs frequently prescribed at all ages ranges, can be either a selective response or due to cross-intolerance (Table 1) [21••]. If these drugs are

NSAIDs
reactions to
persensitivity
of h
Classification
able 1.

	Type of reaction	Clinical manifestations	Timing	Underlying disease	Mechanism
Cross-intolerance reactions (non-immunologic)	NSAID-exacerbated respiratory disease	Bronchial obstruction, dyspnoea and/or nasal congestion/	Acute (immediate to several hours after exposure)	Asthma rhinosinusitis	COX-1 inhibition
(26)	NSAID-exacerbated	Wheals and/or angioedema		Chronic urticaria	COX-1 inhibition
	NSAID-induced urticaria/	Wheals and/or angioedema			Unknown, probably COX-1 inhibition
Selective	Single-NSAID-induced	Wheals/angioedema/anaphylaxis	Delayed onset (usually		lgE-mediated
(immunologic)	Single-NSAID-induced	Maculopapular exathema	exposure)		T cell-mediated
	מכנמלכת וכמרנוסווז	Fixed drug eruption			
		Acute generalized exanthematous pustulosis Drug reaction with eosinophilia and systemic symptoms Stevens-Johnson syndrome/Toxic epidermal necrolysis Organ-specific reactions			

administered to patients with chronic spontaneous urticaria, this can also lead to an anaphylactic reaction or an accelerated reaction similar to those observed with amoxicillin [46•]. In summary, when dealing with an NSAID reaction indicative of DIA, there are four potential mechanisms to consider: selective and cross-intolerance in subjects without any underlying disease, and immediate and accelerated reactions in subjects with chronic spontaneous urticaria.

As stated above, the most highly studied model is that of BLs, where it has been shown that different penicillin structures can produce an epitope that induces selective or cross-reactive responses [47, 48]. When symptoms occur over an hour after drug intake, this may be due to a T cell response [46•, 49]. If anaphylaxis occurs following a long interval, as can occur with several drugs, the possibility of metabolite generation must be considered [15, 50]. This is thought to be relevant for several NSAIDs [50, 51].

Selective immediate reactions have been reported for all NSAIDs, with the most common being pyrazolones [23]. Since histamine and other mediators are released in NSAID reactions, an anaphylactic event may occur. It is now clear that when mediators are released, their metabolism and interactions with specific receptors contribute to the severity of the response [52, 53].

Serum sickness attributed to the formation of immune complexes has been proposed to be induced by penicillin [1], other antibiotics and other drugs [54]. However, protein-drug complexes with IgG or other antibodies have never been shown. Complement activation in the acute phase of these reactions has been reported, although the mechanism requires further clarification [55–57].

Monitoring the Acute Phase

Anaphylaxis involves the release of vasoactive mediators including histamine, tryptase and prostaglandin-leukotriene mediators [1]. Histamine appears very rapidly and can be detected between 5 and 10 min after the reaction initiation in peripheral blood [58]. It then becomes undetectable, but its metabolites can be measured in urine by taking samples at different time intervals after the episode [59]. Serum tryptase levels peak 60–90 min after the onset of anaphylaxis and can persist for up to 24 h [60, 61]. Both parameters can be used to monitor the anaphylactic response.

In addition, arachidonic acid derivatives can be monitored in peripheral blood or urine, although as with histamine, this is more feasible in sequential urine samples [1]. Levels of these mediators are indicative of mast cell activation but not the initial underlying mechanisms, since both IgE and other mechanisms can lead to their release [58]. Kinin generation, complement system activation and nitric oxide synthesis may also occur during anaphylaxis [1].

IgE-mediated reactions are caused by antigenic stimulation, leading to a boosting of the immunological response, which can be monitored. More specifically, the IgE response shows a Th2 pattern with an increase of IL-4 and IL-13 and a downregulation of IFN- γ and IL-12 [62, 63]. Involvement of TNF- α in anaphylactic reactions has also been reported [62].

Clinical Manifestations

According to recently published work, the most common clinical manifestations appear in the skin, followed by respiratory, cardiovascular and gastrointestinal involvement [1, 4, 18•]. Histamine and the other vasoactive mediators are responsible for flushing, urticaria and angioedema in the skin, wheezing and upper airway involvement including laryngeal oedema, a decrease in blood pressure, cardiac arrhythmia in the cardiovascular system and abdominal cramps, nausea, vomiting and diarrhoea. DIA can be divided into several categories according to the extent of the reaction [64]. Isolated symptoms involving a single organ can also occur, such as cardiovascular collapse, syncope and seizure [1, 17]. Biphasic anaphylaxis can occur—this involves the aggravation of symptoms several hours after the initial episode. Sometimes, this may evolve into protracted shock [1, 65]. Although drug allergy may appear in subjects with mastocytosis, it is still unclear whether these patients are more prone to developing anaphylactic reactions after the intake of drugs, particularly for NSAIDs, codeine and muscle relaxants [66].

Risk Factors

Predictors of serious outcomes include a previous history of DIA, a history of allergic disease, multiorgan involvement, and old age [1, 4]. Diabetes, hypertension and other cardiovascular diseases can also be considered risk factors for developing anaphylaxis [1]. Patients with these conditions may develop complications during the treatment of anaphylaxis because some drugs (e.g. epinephrine or similar drugs) may induce adverse cardiovascular effects [2]. There is no consensus as to whether atopy is a risk factor for DIA [1]. Patients with penicillin allergy are more likely to develop allergy to other drugs [31].

NSAIDs, particularly aspirin, and food additives may exacerbate allergic symptoms in patients with chronic idiopathic urticaria and food-dependent exercise-induced anaphylaxis (FDEIA) [67–69]. In these cases, these drugs act as cofactors for triggering a response.

The worsening of symptoms in patients with chronic urticaria and FDEIA can be due to aspirin-enhanced histamine release from basophils via increased Syk kinase activation [70–72]. Confounding factors also exist leading to drugs being falsely identified as the culprit. For example, cold stimuli such as water, air or physical objects may sometimes induce anaphylaxis [73–75]. This can also occur following the infusion of fluids during surgery [74].

Cross-Reactivity Versus Cross-Intolerance

The term cross-reactivity refers to immunological reactions and implies that drugs with similar chemical structures will be recognised by the immunological

system and induce a response. It is known that in IgE-mediated reactions to penicillin, subjects can react to all penicillin determinants or to those with specific side chains [47, 49]. The production of monoclonal antibodies indicates that different parts of the amoxicillin molecule bound to a protein carrier can be recognised [48], and selective mediator release following drug administration has been shown using experimental sensitisation models [76]. It is very unlikely that a subject sensitised to the major benzyl penicillin determinant will also respond to BLs with very different chemical structures such as aztreonan [77]. This concept must be considered when desensitising BLs patients, as will be outlined below.

The concept of cross-intolerance applies only to NSAIDs and is not immunological in nature but rather related to the inhibition of the COX-1 and COX-2 enzymes [51]. Different drugs with similar inhibitory capacity may trigger the same response. Therefore, an anaphylactic reaction induced by cross-intolerance to ibuprofen can also be induced by diclofenac, aspirin or indomethacin [22]. Although subjects generally respond to strong COX-1 inhibitors, reactions have also been reported for weaker inhibitors like oxicams [78] or selective COX-2 inhibitors [78, 79]. In some cases, NSAID responses can also be immunologically mediated and therefore selective to specific drugs [23]. In these circumstances, subjects usually respond to a single drug/drug subgroup but tolerate other NSAIDs, including strong COX inhibitors [80–82].

Diagnosis

DIA diagnosis is performed using clinical history [1]. A precise description of symptoms is also essential to decide which in vivo and in vitro tests should be carried out, including a drug provocation test (DPT) [7••, 19, 49]. In allergic reactions to BLs, if the clinical history is indicative of an immediate reaction, i.e. anaphylaxis, a skin test is performed with penicillin determinants as recommended. The concept of major/minor determinants of benzyl penicillin (PPL and MDM) has now been surpassed since these drugs are not the most frequent elicitors of anaphylaxis [26], and it is now recommended to include amoxicillin determinants, and if the culprit drug is a cephalosporin, this must also be included. In those countries where clavulanic acid is becoming an important elicitor, the inclusion of this drug is also recommended (clavulanic for skin prick test) [27, 28]. For anaphylactic reactions to NSAIDs, if cross-intolerance is ruled out, skin testing is of very little value, except for pyrazolones [23, 81]. Skin test positivity to other NSAIDs is almost anecdotal, and only a few instances have been reported [23].

In vitro tests are considered useful alternatives in the case of penicillin determinants. Although they are generally less sensitive than skin testing [26], there are instances in which the in vitro test can be positive in the absence of an in vivo positive response [44]. Experimental prototypes have been developed for some drugs, such as pyrazolones and ASA; however, they are not yet available in routine clinical practice [80, 82]. Another in vitro option is the use of basophil activation tests. Although these have been proposed for subjects with NSAID hypersensitivity caused by cross-intolerance [83], they show a high number of false positives [84, 85]. However, the basophil activation has proved to be useful for beta-lactam [86], selective reactions to pyrazolones [81, 85], omeprazole [87], ranitidine [88], and other agents [89].

Other techniques, such as lymphocyte-stimulation-based tests, originally developed for T cell-mediated reactions [90, 91], have also been applied to IgE-dependent reactions [91, 92].

Muscle relaxants are particularly relevant for anaphylaxis occurring in hospital settings [93]. In these situations, skin testing, quantitation of specific IgE antibodies and basophil activation tests have been used to establish diagnosis [94, 95]. Cross-reactivity can occur for some of these drugs [93, 96].

In daily practice, a controlled challenge is often necessary to assess tolerance and confirm diagnosis, due to the low sensitivity and availability of many of the above-mentioned diagnostic tests $[1, 7 \bullet \bullet, 19]$. This is commonly used when the clinical history is unclear or when many drugs have been taken simultaneously [1]. The approach taken depends on the putative mechanism involved.

Controlled challenge is made with increasing drug concentrations until a therapeutic dose is achieved. In DIA the initial concentration should be very low and increased at intervals of 30 min [26]. This approach is different for cross-intolerant NSAID-induced DIA. In these circumstances, the incremental steps can be performed using higher doses [22].

Differential Diagnosis

Anaphylaxis may occur in the context of a clear-cut episode induced by a single drug; however, in many cases, a patient will have taken several drugs simultaneously alongside other potential triggers such as foods [67]. To make matters even more complex, anaphylaxis can occur following the intake of drugs and foods and undertaking exercise [67, 69, 72]. Therefore, a detailed clinical history with a precise description of the timing of drug intake, other potential factors, and appearance of symptoms is essential. As previously stated, cold stimuli can also lead to anaphylaxis, and these must also be considered in the diagnosis of DIA [73–75]. Other complications that may mimic DIA include scombroidoisis due to histamine poisoning in spoiled fish. If this occurs alongside drug intake, it can be incorrectly diagnosed as drug allergy [97].

Treatment and Desensitisation

For this section, we refer the reader to different position statements that provide protocols for the treatment and management of DIA. Once anaphylaxis has been recognised, the sooner the treatment is administered, the more effective it will be. Intramuscular injection in adults of 0.3–0.5 ml (1 mg/ml), or 0.01 mg/kg of body weight for children, epinephrine to the mid outer thigh has been recommended [1, 2]. This can be repeated every 5–15 min if necessary. Protocols used for treating DIA are the same as those recommended for anaphylaxis in general. In addition to epinephrine, these include antihistamines and corticosteroids plus administration of parenteral fluids and other supporting measures. We refer the reader to references [1, 2] for a full description of treatment options. DIA can occur in allergy units where drug allergy diagnosis is performed, even with very low concentrations (e.g. through skin prick or intradermal testing). For BLs, it is recommended, according to severity, to begin with doses of less than one thousandth of the initial concentration and in some cases not testing all

determinants simultaneously [19, 26, 45]. The use of this measure has led to an important reduction in the number of reactions induced in such units [26]. Often, symptoms mainly affect the lung or upper airways, for example, laryngeal oedema, with mild skin involvement and no systemic cardiovascular effects. In these cases, particularly for older patients, administration of epinephrine is particularly risky, and a careful balance between drug administration and blood pressure control must be achieved.

The aim of desensitisation is to achieve the situation where basophils and mast cells become unresponsive to the drug [1, 98, 99]. Desensitisation in DIA is indicated when the drug is crucial to the patient and no alternatives are available. In the past, this was almost exclusively performed for BLs and NSAIDs [1] but is now used for chemotherapeutic agents, contrast media and many other drugs [100].

Desensitisation to BLs is antigen specific. It consists of administering progressive doses of the drug every 30-60 min until the therapeutic dose is achieved. If minor symptoms occur, they should be treated adequately [98, 99]. A reduction in skin test sensitivity during desensitisation has been observed in some patients [100, 101], although the mechanism has not been fully elucidated [102].

Whenever possible, rapid oral desensitisation is preferable to intravenous desensitisation since the cost is much lower and the procedure has similar efficacy [17]. However, the oral route is less feasible due to difficulties in obtaining the precise dose due to absorption and other factors related to drug distribution.

If adverse effects occur during desensitisation, the time interval and number of doses should be extended and treated when necessary [1]. Once desensitisation is completed, patients should be able to tolerate the full dose of the drug with minimal side effects [100–102].

Desensitisation can also be performed for DIA due to NSAIDs [103, 104]. Although most studies of NSAID desensitisation have been performed for patients with respiratory airway involvement, it can also be used for DIA. It is critical to establish whether we are dealing DIA due to cross-intolerance or to a selective response in order to choose the correct dosage regime and to see if alternative NSAIDs are available.

An increasing concentration of the drug is given during a period of 2–3 days until the required dose has been acquired. This is then maintained. Subjects with cross-intolerance generally respond to relatively high doses, and doses below 300 mg may be tolerated without the need for desensitisation [105, 106].

For selective reactions, desensitisation is not indicated unless there is a very specific need for a particular NSAID, for example, aspirin for the treatment of stroke [23].

Acknowledgments

JA Cornejo-García is a researcher from the Miguel Servet Program (Ref CP14/00034) and JR Perkins from the Sara Borrell Program (Ref CD14/00242) (Carlos III National Health Institute, Spanish Ministry of Economy and Competitiveness. This work has been supported by grants from the Carlos III National Health Institute RD12/0013 (RIRAAF

Network), FIS PI12/02247, and FIS PI13/02598; Marie Curie (IAPP 7th Framework Program Mr.SymBioMath, no. 324554); and the Health Government of Andalusia (PI-0279-2012).

Compliance with Ethics Guidelines

Conflict of Interest

Natalia Blanca-López, María del Carmen Plaza-Serón, José Antonio Cornejo-García, James Richard Perkins, Gabriela Canto, and Miguel Blanca declare no conflicts of interest.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

References and Recommended Reading

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- •• Of major importance
- Celik G, Pichler WJ, Adkinson NF. Drug allergy in allergy: principles and practice. 7th ed. In: Adkison NF, Bochner BS, Busse WW, Holgate ST, Lemaske RF, Simons EF, editors. Philadelphia: Mosby Elsevier; 2009. p 1205-1226.
- Simons FE, Ardusso LR, Bilo MB, et al. International consensus on (ICON) anaphylaxis. World Allergy Organ J. 2014;7:1–19.
- 3. Johansson S, Bieber T, Dahl R, et al. Revised nomenclature for allergy for global use: report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. J Allergy Clin Immunol. 2004;113:832–6.
- Ye YM, Kim MK, Kang HR, et al. Predictors of the severity and serious outcomes of anaphylaxis in Korean adults: a multicenter retrospective case study. Allergy Asthma Immunol Res. 2015;7:22–9.
- 5.•• Jerschow E, Lin RY, Scaperotti MM, et al. Fatal anaphylaxis in the United States, 1999-2010: temporal patterns and demographic associations. J Allergy Clin Immunol. 2014;134:1318–28.

In this manuscript trends in fatal anaphylaxis were analysed between 1999 and 2010 showing that medications were the most common cause. They found an increase in medication-related deaths caused by anaphylaxis, which was likely associated with different factors, including increases in medication and contrast media usage.

- Papadopoulos, Agache I, Bavbek S, et al. Research needs in allergy: an EAACI position paper, in collaboration with EFANG. Clin Transl Allergy. 2012;2:1–23.
- 7.•• Doña I, Blanca-Lopez N, Torres MJ, et al. Drug hypersensitivity reactions: response patterns, drug involved, and temporal variations in a large series of patients. J Investig Allergol Clin Immunol. 2012;22:363–71.

In this study all patients attending an allergy service for drug allergy between 2005 and 2010 were evaluated. From a total of more than 4000 cases, 37 % were confirmed as allergic. The most frequent diagnosis was hypersensitivity reactions to multiple NSAIDs in 47 % of the cases, followed by immediate reactions to BLs in 18 %.

- 8. Doña I, Barrionuevo E, Blanca-Lopez N, et al. Trends in hypersensitivity drug reactions: more drugs, more response patterns, more heterogeneity. J Investig Allergol Clin Immunol. 2014;24:143–53.
- 9. Mayorga, Torres MJ, Corzo JL, et al. Immediate allergy to tetanus toxoid vaccine: determination of immuno-globulin E and immunoglobulin G antibodies to allergenic proteins. Ann Allergy Asthma Immunol. 2003;90:238–43.
- 10. Komericki P, Grims RH, Aberer W, et al. Near-fatal anaphylaxis caused by human serum albumin in fibrinogen and erythrocyte concentrates. Anaesthesia. 2014;69:176–8.
- 11. Hopps S, Medina P, Pant S, et al. Cetuximab hypersensitivity infusion reactions: incidence and risk factors. J Oncol Pharm Pract. 2013;19:222–7.
- Baldo BA. Adverse events to monoclonal antibodies used for cancer therapy: focus on hypersensitivity responses. Oncoimmunology. 2013;2:e26333.
- Weiss RB, Baker Jr JR. Hypersensitivity reactions to antineoplastic agents. Cancer Metastasis Rev. 1987;6:413–32.
- 14. Renaudin JM, Beaudouin E, Ponvert C, et al. Severe drug-induced anaphylaxis: analysis of 333 cases recorded by the Allergy Vigilance Network from 2002 to 2010. Allergy. 2013;68:929–37.
- 15. Lavergne SN, Park BK, Naisbitt DJ. The roles of drug metabolism in the pathogenesis of T-cell mediated

- drug hypersensitivity. Curr Opin Allergy Clin Immunol. 2008;8:299–307.
- Sicherer SH, Leung DY. Advances in allergic skin disease, anaphylaxis, and hypersensitivity reactions to foods, drugs, and insect stings. J Allergy Clin Immunol. 2004;114:118–24.
- 17. Stone SF, Phillips EJ, Wiese MD, et al. Immediate-type hypersensitivity drug reactions. Br J Clin Pharmacol. 2014;78:1–13.
- 18. Faria E, Rodrigues-Cernadas J, Gaspar E, et al. Druginduced anaphylaxis survey in Portuguese Allergy Departments. J Investig Allergol Clin Immunol. 2014;24:40–8.

During a 4 years period a nationwide notification system for anaphylaxis was implemented by data taken from allergists in Portugal. The main culprit drugs were NSAIDs followed by antibiotics. They conclude that these drugs were the most common cause of anaphylaxis.

- Messaad D, Sahla H, Benahmed S, et al. Drug provocation tests in patients with a history suggestive of immediate drug hypersensitivity. Ann Intern Med. 2004;140:1001-6.
- 20. Reitter M, Petitpain N, Latarche C, et al. Fatal anaphylaxis with neuromuscular blocking agents: a risk factor and management analysis. Allergy. 2014;69:954–9.

This study evaluated the mortality rate in France from anaphylactic reactions to neuromuscular blocking agents. From a total of two thousand and twenty two cases, they found that 4.1 % was fatal. Sixty one percent of the cases had severe anaphylaxis.

21. •• Kowalski ML, Asero R, Blanca M, et al. Classification and practical approach to the diagnosis and t of hypersensitivity to nonsteroidal anti-inflammatory drugs. Allergy. 2013;68:1219–32.

A new classification for hypersensitivity reactions to NSAIDs is provided. Three major groups of entities are diagnosed for cross-intolerants and 2 for selective responders. This new classification represents a consensus document provided by the expert committee on drug allergy of the European Academy of Allergy and Immunology.

- Doña I, Blanca-López N, Cornejo-García JA, et al. Characteristics of subjects experiencing hypersensitivity to non-steroidal anti-inflammatory drugs: patterns of response. Clin Exp Allergy. 2011;41:86–95.
- Canto G, Andreu I, Fernandez J, et al. Selective immediate hypersensitivity reactions to NSAIDs. Curr Opin Allergy Clin Immunol. 2009;9:293–7.
- 24. Aun MV, Blanca M, Garro LS, et al. Nonsteroidal antiinflammatory drugs are major causes of drug-induced anaphylaxis. J Allergy Clin Immunol Practice. 2014;2:414–20.

This work investigates a total of 800 cases who attended emergencies of a hospital because of drug allergy. Anaphylaxis was diagnosed in 14 % of the cases and NSAIDs were the drugs more frequently implicated.

 Blanca M. Allergic reactions to penicillins. A changing world? Allergy. 1995;50:777–82.

- Blanca M, Mayorga C, Torres MJ, et al. Side-chainspecific reactions to betalactams: 14 years later. Clin Exp Allergy. 2002;32:192–7.
- 27. Longo N, Gamboa PM, Gastaminza G, et al. Diagnosis of clavulanic acid allergy using basophil activation and leukotriene release by basophils. J Investig Allergol Clin Immunol. 2008;18:473–5.
- 28. Torres MJ, Ariza A, Mayorga C, et al. Clavulanic acid can be the component in amoxicillin-clavulanic acid responsible for immediate hypersensitivity reactions. J Allergy Clin Immunol. 2010;125:502–5.
- Chang YS. Hypersensitivity reactions to proton pump inhibitors. Curr Opin Allergy Clin Immunol. 2012;12:348–53.
- 30. Bonadonna P, Lombardo C, Bortolami O. Hypersensitivity to proton pump inhibitors: diagnostic accuracy of skin tests compared to oral provocation test. J Allergy Clin Immunol. 2012;130:547–9.

This work presents a study conducted between 2008 and 2010 concerning immediate reactions to protein pump inhibitors. Skin tests were positive in 22% of the cases reported. Patients sensitised to pantoprazole also were skin test positive to omeprazole but patients sensitised to lanzoprazol and rabeprazole were negative to omeprazole.

- 31. Blanca-López N, Ariza A, Doña I, et al. Hypersensitivity reactions to fluoroquinolones: analysis of the factors involved. Clin Exp Allergy. 2013;43:560–7.
- 32. Phan C, Vial-Dupuy A, Autegarden JE, et al. A study of 19 cases of allergy to heparins with positive skin testing. Ann Demerol Venereol. 2014;141:23–9.
- 33. Yoo HS, Yang M, Kim MA, et al. A case of codein induced anaphylaxis by oral route. Allergy Asthma Immunol Res. 2014;6:95–7.
- 34. Fransi S, Briedis J. Anaphylaxis to intravenous amiodarone. Anaesth Intensive Care. 2004;32:578–9.
- 35. Schmidt LE, Dalhoff K. Risk factors in the development of adverse reactions to N-acetylcysteine in patients with paracetamol poisoning. Br J Clin Pharmacol. 2001;51:87–91.
- 36. Dyer JE, Nafie S, Mellon JK, Khan MA. Anaphylactic reaction to intraurethral chlorhexidine: sensitisation following previous repeated uneventful administration. Ann R Coll Surg Engl. 2013;95:e105–6.
- 37. Juel J, Pareek M, Langfrits CS, Jensen SE. Anaphylactic shock and cardiac arrest caused by thiamine infusion. BMJ Case Rep. 2013;12:2013. doi:10.1136/bcr-2013-009648.
- 38. Jethava A, Mathew J, Dasanu CA. Anaphylactic reactions with intravenous vitamin K: lessons from the bedside. Conn Med. 2012;76:549–50.
- 39. Fernandez M, Barcelo M, Muñoz C, et al. Anaphylaxis to thiamine (vitamin B1). Allergy. 1997;52:958–9.
- Calogiuri GF, Nettis E, Di Leo E, Muratore L, Ferrannini A, Vacca A. Long-term selective IgE-mediated hypersensitivity to hydrocortisone sodium succinate. Allergol Immunopathol. 2013;41:206–8.
- Torres MJ, Canto G. Hypersensitivity reactions to corticosteroids. Curr Opin Allergy Clin Immunol. 2010;10:273–9.

- 42. Aranda A, Mayorga C, Ariza A, et al. IgE-mediated hypersensitivity reactions to methylprednisolone. Allergy. 2010;65:1376–80.
- Blanca M, Garcia J, Vega JM, et al. Anaphylaxis to penicillins after non-therapeutic exposure: an immunological investigation. Clin Exp Allergy. 1996;26:335–40.
- Torres MJ, Mayorga C, Cornejo-García JA, et al. IgE antibodies to penicillin in skin test negative patients. Allergy. 2002;57:965.
- 45. Torres MJ, Sánchez-Sabaté E, Alvarez J, et al. Skin test evaluation in nonimmediate allergic reactions to penicillins. Allergy. 2004;59:219–24.
- 46. Gómez E, Blanca-Lopez N, Salas M, et al. Induction of accelerated reactions to amoxicillin by T-cell effector mechanisms. Ann Allergy Asthma Immunol. 2013;110:267–73.

Three cases with accelerated reactions to amoxicillin are reported. Monitoring the acute phase of the reaction as well as the in vitro lymphocyte studies showed that the mechanism involved was a T cell effector response.

- Moreno F, Blanca M, Mayorga C, et al. Studies of the specificities of IgE antibodies found in sera from subjects with allergic reactions to penicillins. Int Arch Allergy Immunol. 1995;108:74–81.
- Mayorga C, Obispo T, Jimeno L, et al. Epitope mapping of β-lactam antibiotics with the use of monoclonal antibodies. Toxicology. 1995;97:225–34.
- 49. Torres MJ, Mayorga C, Leyva L, et al. Controlled administration of penicillin to patients with a positive history but negative skin and specific serum IgE tests. Clin Exp Allergy. 2002;32:270–6.
- Castell JV, Castell M. Allergic hepatitis induced by drugs. Curr Opin Allergy Clin Immunol. 2006;6:258–65.
- 51. Cornejo-Garcia JA, Blanca-López N, Doña I, et al. Hypersensitivity reactions to non steroidal anti-inflammatory drugs. Curr Drug Metab. 2009;10:971–80.
- García-Martín E, Ayuso P, Martínez C, et al. Histamine pharmacogenomics. Pharmacogenomics. 2009;10:867–83.
- 53. Agúndez JAG, Ayuso P, Cornejo-García JA, et al. The diamine oxidase gene is associated with hypersensitivity response to non-steroidal anti-inflammatory drugs. PLoS ONE. 2012;11:e47571.
- 54. Kim DH, Choi YH, Kim HS, et al. A case of serum sickness-like reaction and anaphylaxis induced simultaneously by rifampin. Allergy Asthma Immunol Res. 2014;6:183–5.
- Lasser EC, Lang JH, Lyon SG, et al. Complement and contrast material reactors. J Allergy Clin Immunol. 1979;64:105–12.
- Szebeni J. Complement activation-related pseudoallergy: a stress reaction in blood triggered by nanomedicines and biologicals. Mol Immunol. 2014;61:163–73.
- 57. Szebeni J, Alving CR, Savay S, et al. Formation of complement-activating particles in aqueous solutions of taxol: possible role in hypersensitivity reactions. Intern Immunopharm. 2001;1:721–35.

- 58. Eberlein-König B, Ullmann S, Thomas P, et al. Tryptase and histamine release due to a sting challenge in bee venom allergic patients treated successfully and unsuccessfully with hyposensitisation. Clin Exp Allergy. 1995;25:704–12.
- 59. Fernandez J, Blanca M, Moreno C, et al. Role of tryptase, eosinophil cationic protein and histamine in immediate allergic reactions to drugs. Int Arch Allergy Immunol. 1995;107:160–2.
- 60. Moreno F, Blanca M, Fernandez J, et al. Determination of inflammatory mediators in allergic reactions to drugs. Allergy Asthma Proc. 1995;16:119–22.
- 61. Galvão VR, Giavina-Bianchi P, Castells M. Perioperative anaphylaxis. Curr Allergy Asthma Rep. 2014;14:452.
- 62. Posadas SJ, Leyva L, Torres MJ, et al. Subjects with allergic reactions to drugs show in vivo polarized patterns of cytokine expression depending on the chronology of the clinical reaction. J Allergy Clin Immunol. 2000;106:769–76.
- 63. Cornejo-Garcia JA, Fernandez TD, Torres MJ, et al. Differential cytokine and transcription factor expression in patients with allergic reactions to drugs. Allergy. 2007;62:1429–38.
- Simons FE, Ardusso LR, Dimov V, et al. World Allergy Organization Anaphylaxis Guidelines: 2013 update of the evidence base. Int Arch Allergy Immunol. 2013;162:193–204.
- 65. Banerji A, Rudders S, Clark S, et al. Retrospective study of drug-induced anaphylaxis treated in the emergency department or hospital: patient characteristics, management, and 1-year follow-up. J Allergy Clin Immunol Pract. 2014;2:46–51.
- Brockow K, Bonadonna P. Drug allergy in mast cell disease. Curr Opin Allergy Clin Immunol. 2012;12:354–60.
- 67. Asero R. Multiple nonsteroidal anti-inflammatory drug-induced cutaneous disease: what differentiates patients with and without underlying chronic spontaneous urticaria? Int Arch Allergy Immunol. 2014:163:114–8.
- 68. Matsuo H, Kaneko S, Tsujino Y, et al. Effects of nonsteroidal anti-inflammatory drugs (NSAIDs) on serum allergen levels after wheat ingestion. J Dermatol Sci. 2009;53:241–3.
- 69. Pascal M, Muñoz-Cano R, Reina Z, et al. Lipid transfer protein syndrome: clinical pattern, cofactor effect and profile of molecular sensitization to plant-foods and pollens. Clin Exp Allergy. 2012;42:1529–39.
- Matsukura S, Aihara M, Sugawara M, et al. Two cases of wheat-dependent anaphylaxis induced by aspirin administration but not by exercise. Clin Exp Dermatol. 2010;35:233–7.
- 71. Harada S, Horikawa T, Ashida M, et al. Aspirin enhances the induction of type I allergic symptoms when combined with food and exercise in patients with food-dependent exercise-induced anaphylaxis. Br J Dermatol. 2001;145:336–9.

- 72. Matsuo H, Yokooji T, Morita H. Aspirin augments IgEmediated histamine release from human peripheral basophils via Syk kinase activation. Allergol Int. 2013;62:503–11.
- 73. Wanderer AA, Grandel KE, Wasserman SI, et al. Clinical characteristics of cold-induced systemic reactions in acquired cold urticaria syndromes: recommendations for prevention of this complication and a proposal for a diagnostic classification of cold urticaria. J Allergy Clin Immunol. 1986;78:417–23.
- 74. Lockhart CH, Brownrigg JC. Anesthetic hazards of cold urticaria. Anesthesiology. 1973;38:96–7.
- 75. Park HJ, Park SY, Lee SH, et al. Cold-induced systemic reactions caused by infusion of intravenous fluid. Acta Derm Venereol. 2013;93:469–70.
- Fernandez M, Warbrick, Blanca M, et al. Activation and hapten inhibition of mast cells sensitized with monoclonal IgE anti-penicillin antibodies: evidence for twosite recognition of the penicillin derived determinant. Eur J Immunol. 1995;25:2486–91.
- Vega JM, Blanca M, Garcia JJ, et al. Tolerance to aztreonam in patients allergic to betalactam antibiotics. Allergy. 1991;46:96–102.
- 78. Dona I, Blanca-López N, Jagemann LR, et al. Response to a selective COX-2 inhibitor in patients with urticaria/ angioedema induced by nonsteroidal anti-inflammatory drugs. Allergy. 2011;66:1428–33.
- Asero R, Quarantino D. Cutaneous hypersensitivity to multiple NSAIDs: never take tolerance to selective COX-2 inhibitors for granted. Eur Ann Allergy Clin Immunol. 2013;45:3–6.
- 80. Blanca M, Perez E, Garcia JJ, et al. Angiodema and IgE antibodies to aspirin: a case report. Ann Allergy. 1989;62:295–8.
- 81. Gómez E, Blanca-Lopez N, Torres MJ, et al. Immunogloblin E-mediated immediate allergic reactions to dipyrone: value of basophil activation test in the identification of patients. Clin Exp Allergy. 2009;39:1217–24.
- 82. Himly M, Jahn-Schmid B, Pittertschatscher K, et al. IgEmediated immediate-type hypersensitivity to the pyrazolone drug propyphenazone. J Allergy Clin Immunol. 2003;111:882–8.
- 83. De Weck AL, Sanz ML, Gamboa PM, et al. Nonsteroidal anti-inflammatory drug hypersensitivity syndrome: a multicenter study. II. Basophil activation by nonsteroidal anti-inflammatory drugs and its impact on pathogenesis. J Investig Allergol Clin Immunol. 2010;20:39–57.
- 84. Ariza A, Fernandez TD, Doña I, et al. Basophil activation after nonsteroidal anti-inflammatory drugs stimulation in patients with immediate hypersensitivity reactions to these drugs. Cytometry A. 2014;85:400–7.
- 85. Gamboa PM, Sanz ML, Caballero MR, et al. Use of CD63 expression as a marker of in vitro basophil activation and leukotriene determination in metamizol allergic patients. Allergy. 2003;58:312–7.
- 86. Torres MJ, Padial A, Mayorga C, et al. The diagnostic interpretation of basophil activation test in immediate

- allergic reactions to betalactams. Clin Exp Allergy. 2004;34:1768–75.
- 87. Musset B, Morgan D, Cherny VV, MacGlashan Jr DW, et al. A pH-stabilizing role of voltage-gated proton channels in Ig-E mediated activation of human basophils. Proc Natl Acad Sci U S A. 2008;105:11020–5.
- 88. Makris M, Aggelides X, Chliva C, et al. High baseline blood histamine levels and lack of cross-reactivity in a patient with ranitidine induced anaphylaxis. J Investig Allergol Clin Immunol. 2014;24:361–3.
- 89. Mayorga C, Sanz ML, Gamboa PM, et al. In vitro diagnosis of immediate allergic reactions to drugs: an update. J Investig Allergol Clin Immunol. 2010;20:103–9.
- 90. Adam J, Pichler WJ, Yerly D. Delayed drug hypersensitivity: models of T-cell stimulation. Br J Clin Pharmacol. 2011;71:701–7.
- Luque I, Leyva L, Torres MJ, et al. In vitro T-cell responses to β-lactam drugs in immediate and nonimmediate allergic reactions. Allergy. 2001;56:611–8.
- 92. Rodriguez-Pena R, Lopez S, Mayorga C, et al. Potential involvement of dendritic cells in delayed type hypersensitivity reactions to beta-lactams. J Allergy Clin Immunol. 2006;118:949–56.
- 93. Dong SW, Mertes PM, Petitpain N, et al. Hypersensitivity reactions during anesthesia. Results from the ninth French survey (2005-2007). Minerva Anestesiol. 2012;78:868–78.
- 94. Leysen J, Sabato V, Verweij MM, et al. The basophil activation test in the diagnosis of immediate drug hypersensitivity. Expert Rev Clin Immunol. 2011;7:349–55.
- 95. Fisher MM, Baldo BA. Immunoassays in the diagnosis of anaphylaxis to neuromuscular blocking drugs: the value of morphine for the detection of IgE antibodies in allergic subjects. Anaesth Intensive Care. 2000;28:167–70.
- 96. Laroche D, Chollet-Martin S, Léturgie P, et al. Evaluation of a new routine diagnostic test for immunoglobulin E sensitization to neuromuscular blocking agents. Anesthesiology. 2011;114:91–7.
- 97. Tortorella V, Masciari P, Pezzi M, et al. Histamine poisoning from ingestion of fish or scombroid syndrome. Case Rep Emerg Med. 2014;2014:482531.
- 98. Stark BJ, Earl HS, Gross GN, et al. Acute and chronic desensitization of penicillin-allergic patients using oral penicillin. J Allergy Clin Immunol. 1987;79:523–32.
- 99. Turvey SE, Cronin B, Arnold AD, et al. Antibiotic desensitization for the allergic patient: 5 years of experience and practice. Ann Allergy Asthma Immunol. 2004;92:426–32.
- Wendel Jr GD, Stark BJ, Jamison RB, et al. Penicillin allergy and desensitization in serious infections during pregnancy. N Engl J Med. 1985;312:1229–32.
- Naclerio R, Mizrahi EA, Adkinson Jr NF. Immunologic observations during desensitization and maintenance of clinical tolerance to penicillin. J Allergy Clin Immunol. 1983;71:294–301.

- 102. Sobotka AK, Dembo M, Goldstein B, et al. Antigenspecific desensitization of human basophils. J Immunol. 1979;122:511–7.
- 103. Pleskow WW, Stevenson DD, Mathison DA, et al. Aspirin desensitization in aspirin-sensitive asthmatic patients: clinical manifestations and characterization of the refractory period. J Allergy Clin Immunol. 1982;69:11–9.
- 104. Makowska J, Makowski M, Kowalski ML. NSAIDs hypersensitivity: when and how to desensitize? Curr
- Treat Options Allergy. 2015. doi:10.1007/s40521-015-0049-x.
- 105. Lee JY, Simon RA, Stevenson DD. Selection of aspirin dosages for aspirin desensitization treatment in patients with aspirin-exacerbated respiratory disease. J Allergy Clin Immunol. 2007;119:157–64.
- 106. Macy E, Bernstein JA, Castells MC, et al. Aspirin challenge and desensitization for aspirin-exacerbated respiratory disease: a practice paper. Ann Allergy Asthma Immunol. 2007;98:172–4.