



Biomarkers for ADHD: the Present and Future Directions

Tejas Mehta¹ · Narmada Mannem¹ · Naveen K Yarasi² · Pradeep C. Bollu¹

Published online: 27 May 2020

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Abstract

Purpose of Review Attention deficit hyperactivity disorder (ADHD) is one of the most common neurodevelopmental disorders characterized by inattention, impulsivity, diminished executive functions, and hyperactivity. Objective criteria can be used to assess the diagnosis and response of the disease to medications.

Recent Findings Several biomarkers belonging to electrophysiological, genetic, peripheral, and miRNA-based biomarkers have shown promise in studies to be an objective aid to clinical diagnostic criteria for the diagnosis of ADHD.

Summary This review article focuses on summarizing the existing evidence for different biomarkers that have been studied in the past for diagnosing ADHD.

Keywords Biomarker · ADHD · Electrophysiological · Genetic · Neural · Peripheral

Introduction

Attention deficit hyperactivity disorder (ADHD) is a neurodevelopmental disorder, first described by Sir George F Still, as a condition persisting in children with a problem associated with “moral control” in 1902. This was followed by a neuropsychological study in 1972 that established inattention as a key feature of ADHD [1, 2]. This disorder is characterized by core symptoms of inattention, impulsivity, hyperactivity, and diminished executive functions. It most commonly affects children and adolescents, with 60–80% of these patients having persistence of these symptoms in adulthood [3]. The diagnosis of ADHD emphasizes the presence of symptoms in more than two settings with evidence of a reduction in the quality of social, academic, or occupational functioning after all other psychiatric conditions are ruled out [4]. Owing to the subjective nature of the diagnostic criteria, ADHD may be misdiagnosed as any other neurocognitive or neurodevelopmental disorder, thereby causing a delay in

diagnosis and treatment [5–7]. This calls for the need of objective markers of the disease that can be used in diagnosis, prognostication, and assessment of response to pharmacological interventions.

A biomarker is defined as “a characteristic that is measured objectively and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacological responses to a therapeutic intervention.” [8] The biomarker should have reproducibility, be heterogenous over a large population with a high test-retest reliability, and have high sensitivity and specificity to distinguish those with the disease from those that do not. For ADHD, several biomarkers from the niche of neurophysiology, neurochemistry, neuroimaging, and genetics have been reported in small and moderate studies. This review paper aims to summarize the biomarkers which have been investigated in the past and discusses briefly the challenges with their clinical applications along with future directions.

This article is part of the Topical Collection on *ADHD*

✉ Tejas Mehta
tejas.r.mehta29@gmail.com

¹ Department of Neurology, University of Missouri, Columbia, MO, USA

² Department of Psychiatry, University of Missouri, Columbia, MO, USA

Electrophysiological Biomarkers

Electroencephalography (EEG) would serve as an ideal biomarker for characterizing neurodevelopmental disorders since it provides us with a direct measure of postsynaptic activity and with a temporal resolution that is greater than functional magnetic resonance imaging (fMRI). It is also more tolerant of the motion artifacts with recordings possible in a more natural

setting for the study of infants and young children [9]. Their noninvasive nature is another advantage that makes EEG-based indices good biomarkers.

Till date, the most robust and stable finding reported in patients with ADHD has been an increase in theta band power (4–7 Hz), notably increased theta relative to beta band power (13–30 Hz) which has been referred to in the literature as “theta to beta” ratio (TBR). First proposed in 1991 by Luber and hypothesized to reflect cortical hyperarousal and slowing [10, 11], TBR quickly gained recognition as a potential electrophysiological marker of ADHD. A meta-analysis by Snyder and Hall in 2006 reported a sensitivity and specificity of 94% for this index in the diagnosis of ADHD [12]. The same group later published an empirical study that reported a specificity of 94%, sensitivity of 87%, and an 89% overall accuracy for ADHD diagnosis [13]. In July of 2013, the US Food and Drug Association (FDA) approved the Neuropsychiatric EEG-Based Assessment Aid Health (FDA, 2013), for the assessment of ADHD, which was marketed as “Brain Wave Diagnostic Tool.” However, the usage of TBR as a biomarker was questioned by several studies that have rendered this index controversial in its implementation to diagnose ADHD [14–20]. This was also backed by a meta-analysis performed by Arns et al. in 2013 who concluded that increased TBR should not be considered a reliable diagnostic measure for ADHD [21].

Decreased event-related potentials (ERPs) components including the attentional cue and target P300, preparatory contingent negative variation (CNV), and the inhibitory NoGo P300 are considered good markers of ADHD in children and adults [22–26]. A recent study performed a visual stimulus go/no-go task in two groups of age matched adults (75 control vs 75 ADHD) and aimed to investigate the use of ERPs to differentiate adult ADHD patients from nonclinical controls using a classification method originating from machine learning. The study reported that using a tenfold cross-validation approach, the classification accuracy was 91% and showed that ERPs can contribute to the diagnosis of ADHD [27]. A study by Liechti et al. in 2012 showed that the inclusion of ERP markers improved discrimination although they were not diagnostically relevant [28]. These studies show the benefits and power of advanced multivariate methods, which may contribute to the discovery of more reliable electrophysiological biomarkers of ADHD.

miRNA-Based Biomarkers

Genetic factors play a vital role in the causation of ADHD, a disorder that has heritability rates as high as 75–90% [29, 30]. The heritability of the disease is also influenced by epigenetic factors, of which microRNAs (miRNAs) are known to play a key role. They are non-coding RNAs that negatively regulate

gene expression in human cells [31] and have emerged as possible biomarkers due to their implicated role in dysregulation of gene expression [32, 33]. It has been predicted that approximately one-third of all the genes in the genome are directly targeted by miRNA, and they play a vital role in the central nervous system where they are involved in neuroplasticity as well as development [34]. Owing to their contribution to the development and functioning of the central nervous system, miRNAs have been implicated in several psychiatric and neurological disorders [35–37].

Earlier in the decade, animal studies have reported that miRNA target gene—Homer 1a is associated with phenotypes of ADHD models [38•, 39–42]. Early human studies, although focused on the domain of genetics and genomics, focused on the role of gene polymorphisms within miRNA and miRNA target sites in ADHD pathogenesis [38•, 39–42].

In human studies, lower levels of miRNA 18a-5p, 22-3p, 106b-5p, 24-3p, and 107 have been observed, whereas miRNA 155a-5p and miRNA let-7d were reported to be higher in individuals with ADHD [43, 44]. In 2010, miRNA let-7d was reported by Wu et al. to be overexpressed in the prefrontal cortex and was reported to target galectin-3, leading to a downregulation of tyrosine hydroxylase which plays a key role in dopamine metabolism as well as ADHD development [45]. A common denominator in all the above-mentioned studies is that the selection of candidate miRNA relied heavily on mechanisms implicated in the etiopathogenesis of the disease rather than screening for genes at a global level. This disadvantage was avoided in a recent study conducted by Wang et al. where they used next-generation technique sequencing (NGS) to create a pooled patient library from controls and ADHD-affected patients and found 13 potential miRNA ADHD biomarkers which were efficient in differentiating ADHD-affected individuals from healthy controls with sensitivity and specificity of 86.8% and 88.9%, respectively [46••]. These 13 miRNAs had not been reported to be involved in the pathophysiology of ADHD previously, which favors the argument of conducting studies that search for implicated miRNA biomarkers globally rather than in a preselected fashion based on previously published literature and etiopathological mechanisms implicated in ADHD.

Genetic Biomarkers

The dopamine transporter gene (DAT1 gene) has been linked to the etiopathogenesis of ADHD with many studies on knockout mice for DAT1 showing deficits in inhibitory behavior and hyperactivity. The same gene has also been mapped near a susceptibility locus for ADHD, i.e., 5p13 [47, 48]. Along with this, methylphenidate and amphetamine, which are drugs used to manage ADHD, are known to bring

about their effects by targeting the DAT1 protein. The most studied variant of DAT1 is variable number tandem repeats (VNTR) of 40 base pairs located at the 3' untranslated region (3'-UTR) of which 10 repeat (10R) and 9 repeat (9R) alleles are the most common [49]. A recent meta-analysis showed a positive association between 10R allele and ADHD in the pediatric population, although a reverse association is shown to be present for adults [49, 50]. Studies have shown an association between higher commission errors, increased reaction time variability (RTV), and increased impulsive responses in continuous performance test (CPT) and sustained attention to response test (SART) with 10R allele, although there are other studies which show no or opposite association [51, 52]. Some pharmacogenetic studies have also shown increased response to methylphenidate among homozygous 10R, while others have conflicting results [53, 54]. A more recent meta-analysis showed that VNTR polymorphism is not a reliable predictor of MPH treatment success in ADHD patients [55]. A DAT1 VNTR at intron-8 containing 5R and 6R alleles has also been associated with increased susceptibility of ADHD [49]. Other variants such as rs6350 have been reported to be associated with alerting and executive control performance on the attention network test [56]. Another haplotype, 9rs403636(G)/rs463379 (C)/rs393795 (C)/rs37020 (G), has been reported to be associated with spatial working memory in ADHD [57].

Located on chromosome 11p15.5, the DRD4 gene has a high expression in the anterior cingulate cortex, which is associated with attention and inhibition and thereby also implicated in ADHD [58]. This association is further backed by an earlier study that reported lower mRNA expression levels of DRD4 in ADHD [59]. A highly polymorphic functional VNTR comprising of 11 copies of 48 bp repeat sequences has been studied, and the 7R allele was reported to be associated with ADHD [49, 60]. Other papers have emphasized the association of the 7R allele with processing speed, cognitive impulsiveness, and set-shifting but not with response inhibition [61, 62]. Pharmacogenetic studies have also reported an enhanced response to methylphenidate in 7R carriers even though other studies contradict this finding [53, 54]. Another interesting aspect that was found regarding this issue is that different DRD4 genotypes exhibit different methylphenidate response curves [53, 54]. There have been other variants in the promoter region that have been studied, including the -521 C/T (rs1800955), 120 base pair duplication (120-bp dup) located in the 5' untranslated region but showed no association with ADHD [49, 60].

The DRD2 gene is located on chromosome 11q23.1 and is known to be expressed profoundly in brain areas relevant for regulation of mesolimbic pathways and the other regions known to be associated with ADHD [63]. A genome-wide association study reported a nominal association of ADHD susceptibility with this gene [64]. There are three variants that

have been studied in association with ADHD—rs2075654, rs10795696, and rs1800497. Higher commission errors have been associated with rs207654 and rs1079596 variants [61], whereas the study trying to find a relation of MPH response with rs1800497 showed no association [65].

The DRD5 gene is located on chromosome 4p15.3 and has been reported to be expressed in high concentration in the hippocampus, a brain area implicated in the pathogenesis of ADHD, and functionally affects synaptic strength in memory formation [66]. Studies have also reported that the 148 bp allele is associated with omission errors, commission errors, RTs, and RTVs [67]. Although no association was found between 148 bp allele and methylphenidate response, the 151 bp allele has been linked to a favorable response [68].

The SLC6A2 codes the norepinephrine transporter and is targeted by one of the medications from the non-stimulant class—atomoxetine—while the NET1 is a gene that is highly expressed in the frontal lobe where it is involved in dopaminergic and noradrenergic reuptake [69]. A genome-wide association study reported a frequently located SNP—rs5569 on exon 9 to have a nominal association with ADHD [70]. Another such study reported two SNPs in the SCL6A2 gene to be associated with methylphenidate response and rs3785143 to be associated with atomoxetine response [71, 72].

Peripheral Biomarkers

One of the most studied protein lines includes the neurotrophins, which are a group of growth factors that are known to play a key role in several psychiatric disorders, including anxiety disorders, major depressive disorder, bipolar disorder, schizophrenia, and autism spectrum disorders [73–79]. The brain-derived neurotrophic factor (BDNF) is found primarily in the central nervous system and is distributed throughout the brain and peripheral blood. Several studies have reported increased circulating levels of BDNF in patients with ADHD and have also shown an association between BDNF gene polymorphisms and ADHD [80–82]. Many studies also reported increased levels of BDNF in patients after being started on treatment with methylphenidate and a decrease in their levels when started on atomoxetine [83, 84]. Animal studies also back the role of BDNF in characteristics that are a key feature of ADHD, including aggression, learning deficiency, and increased locomotion activities [85, 86]. However, there is another set of studies that show no association of BDNF levels with ADHD [87, 88]. Another member of the neurotrophin family is the glial-derived neurotrophic factor (GDNF), which plays a pivotal role in the maintenance and survival of dopaminergic and serotonergic neurons by its actions against oxidative damage and neuroinflammation [89]. Functionally, it is known to be an integral component

of learning and memory-related functions [90]. Although evidence reports that no association exists between ADHD and GDNF levels, a recent study in children reported higher levels of circulating GDNF levels in children with ADHD as compared to controls [91••]. Neurotrophin growth factor (NGF) which is known to play a vital role in learning processes and the executive control of attention [92, 93] has been reported to have a borderline significant association with its polymorphisms and ADHD in a family-based sample [94]. Neurotrophin-3 (NTF3) plays a critical role in dopaminergic neurons, noradrenergic neurons, and glutamatergic neurons in the mesolimbic pathways, locus coeruleus, and hippocampal areas, respectively [95]. Polymorphisms involving NTF3 have been reported to be associated with selective attention deficits in ADHD, with a recent study backing the role of NTF3 in ADHD by endorsing a relationship between the adverse emotional effects of methylphenidate in the pediatric population with ADHD and the NTF3 genotype [96•]. A relatively recent study by Bilgic et al. in 2016 also reported elevated levels of GDNF and NTF3 levels in children with ADHD [97].

A key pathogenic factor that has been implicated in the etiopathogenesis of ADHD is the disturbance of neurotransmitter systems in ADHD [98–102]. Noradrenaline and dopamine deficiency in the prefrontal cortex, putamen, striatum, and limbic regions of the brain were found in patients with ADHD [103–108]. This was followed by a study that reported a two-and-a-half-fold increase in levels of glutamate with relative GABA deficiency in the frontal lobe of ADHD patients [109]. These initial studies were complemented by studies involving disturbances of amino acid and their metabolites, which was first reported by Hoshino et al. in 1985 and reported increased plasma concentration of tryptophan in patients with ADHD [110]. A recent study by Dolina et al. studied pyridoxal phosphate-dependent tryptophan degradation and obtained concentration of compounds formed and metabolized in the pathway of tryptophan degradation and the ratios between these compounds. The group reported low values of 4PA (4-pyridoxic acid)/TRP (tryptophan), IND (indole)/TRP, and IND/KYN (kyneurine) ratios in patients with ADHD, which did not change despite treatment with Ritalin [111••].

A recent study also hypothesized an association between inflammation and its contribution to the etiopathogenesis of ADHD by measuring cytokines and reported increased levels of proinflammatory cytokines and finding cytokine gene polymorphisms in patients with ADHD [112]. A study by Oades et al. reported a high but nonsignificant increase in cytokines, including interleukin 6, interleukin 10, and interferon-gamma [113]. Another study reported increased cytokines with certain symptoms of ADHD, viz., increased interleukin 16 with hyperactivity and interleukin 13 with inattention [114]. This association between inflammation and its role in the pathogenesis of ADHD was strengthened by a study conducted in Turkey by Avcil who reported higher values of neutrophil-

lymphocyte ratio (NLR), monocyte lymphocyte ratio (MLR), platelet lymphocyte ratio (PLR), and mean platelet volume (MPV) in patients with ADHD as compared to healthy controls in the study [115].

Conclusion

This review summarizes the biomarkers for ADHD that have been reported in the literature for diagnosis and the response of patients to pharmacological interventions. Although our review article attempted all of the well-studied biomarkers, there might be some that escaped our mention. A common trend noticed in this respect is the fact that with each of these biomarkers, there can be some confounding factors that can lead to elevations or reductions in the level of the biomarker in question. With the onset of the “omic era” and the continuous evolution of machine learning and artificial intelligence, the future of biomarker discovery is promising. The need for a worldwide database that would integrate and store all known patterns of biomarkers that have been assessed in the past would prove to be very useful in the diagnosis of ADHD and its response to drugs.

Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no conflicts of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
 - Of major importance
1. Still G. The Goulstonian lectures on some abnormal physical conditions in children. Lecture 1. *Lancet*. 1902;i 1008–0102:1077–82 1163–1168.
 2. Douglas VI. Stop, look and listen: the problem of sustained attention and impulse control in hyperactive and normal children. *Can J Behav Sci*. 1972;4:259–82.
 3. Childress AC, Berry SA. Pharmacotherapy of attention-deficit hyperactivity disorder in adolescents. *Drugs*. 2012;72:309–25. <https://doi.org/10.2165/11599580-000000000-00000>.
 4. American Psychiatric Association. Attention-deficit and disruptive behavior disorders. In: *Diagnostic and Statistical Manual of Mental Disorders*. 5th ed. Arlington: American Psychiatric Association; 2013.
 5. Zimetkin AJ, Ernst M. Problems in the management of attention-deficit-hyperactivity disorder. *N Engl J Med*. 1999;340:40–6.

6. Mirsky AF, Duncan CC. A nosology of disorders of attention. *Ann N Y Acad Sci.* 2001;931:17–32.
7. Daley KC. Update on attention-deficit/hyperactivity disorder. *Curr Opin Pediatr.* 2004;16:217–26.
8. Biomarkers Definition Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clin Pharmacol Therapeutics.* 2001;69:89–95.
9. Jeste SS, Nelson CA 3rd. Event related potentials in the understanding of autism spectrum disorders: an analytical review. *J Autism Dev Disord.* 2009;39:495–510.
10. Lubar JF. Discourse on the development of EEG diagnostics and biofeedback for attention-deficit/hyperactivity disorders. *Biofeedback Self Regul.* 1991;16:201–25.
11. Monastra VJ, Lubar JF, Linden M. The development of a quantitative electroencephalographic scanning process for attention deficit-hyperactivity disorder: reliability and validity studies. *Neuropsychology.* 2001;15:136–44.
12. Snyder SM, Hall JR. A meta-analysis of quantitative EEG power associated with attention-deficit hyperactivity disorder. *J Clin Neurophysiol.* 2006;23:440–55.
13. Snyder SM, Quintana H, Sexson SB, Knott P, Haque AFM, Reynolds DA. Blinded, multicenter validation of EEG and rating scales in identifying ADHD within a clinical sample. *Psychiatry Res.* 2008;159:346–58.
14. Buyck I, Wiersma JR. Resting electroencephalogram in attention deficit hyperactivity disorder: developmental course and diagnostic value. *Psychiatry Res.* 2014;216:391–7.
15. Liechti MD, Valko L, Muller UC, et al. Diagnostic value of resting electroencephalogram in attention-deficit/hyperactivity disorder across the lifespan. *Brain Topogr.* 2013;26:135–51.
16. Loo SK, Cho A, Hale TS, McGough J, McCracken J, Smalley SL. Characterization of the theta to beta ratio in ADHD: identifying potential sources of heterogeneity. *J Atten Disord.* 2013;17:384–92.
17. Nazari MA, Wallois F, Aarabi A, Berquin P. Dynamic changes in quantitative electroencephalogram during continuous performance test in children with attention-deficit/hyperactivity disorder. *Int J Psychophysiol.* 2011;81:230–6.
18. Ogrim G, Kropotov J, Hestad K. The quantitative EEG theta/beta ratio in attention deficit/hyperactivity disorder and normal controls: sensitivity, specificity, and behavioral correlates. *Psychiatry Res.* 2012;198:482–8.
19. van Dongen-Boomsma M, Lansbergen MM, Bekker EM, Sandra Kooij JJ, van der Molen M, Kenemans JL, et al. Relation between resting EEG to cognitive performance and clinical symptoms in adults with attention-deficit/hyperactivity disorder. *Neurosci Lett.* 2010;469:102–6.
20. Williams LM, Hermens DF, Thein T, Clark CR, Cooper NJ, Clarke SD, et al. Using brain-based cognitive measures to support clinical decisions in ADHD. *Pediatr Neurol.* 2010;42:118–26.
21. Arns M, Conners CK, Kraemer HC. A decade of EEG theta/beta ratio research in ADHD: a meta-analysis. *J Atten Disord.* 2013;17:374–83.
22. Banaschewski T, Brandeis D, Heinrich H, Albrecht B, Brunner E, Rothenberger A. Association of ADHD and conduct disorder—brain electrical evidence for the existence of a distinct subtype. *J Child Psychol Psychiatry Allied Discip.* 2003;44(3):356–76.
23. Banaschewski T, Brandeis D, Heinrich H, Albrecht B, Brunner E, Rothenberger A. Questioning inhibitory control as the specific deficit of ADHD—evidence from brain electrical activity. *J Neural Transm.* 2004;111(7):841–64. <https://doi.org/10.1007/s00702-003-0040-8>.
24. Valko L, Doehner M, Muller UC, Schneider G, Albrecht B, Drechsler R, et al. Differences in neurophysiological markers of inhibitory and temporal processing deficits in children and adults with ADHD. *J Psychophysiol.* 2009;23(4):235–46. <https://doi.org/10.1027/0269-8803.23.4.235>.
25. van Leeuwen TH, Steinhausen HC, Overtom CC, Pascual-Marqui RD, van't Klooster B, Rothenberger A, et al. The continuous performance test revisited with neuroelectric mapping: impaired orienting in children with attention deficits. *Behav Brain Res.* 1998;94(1):97–110.
26. Fallgatter AJ, Ehlis AC, Rosler M, Strik WK, Blocher D, Herrmann MJ. Diminished prefrontal brain function in adults with psychopathology in childhood related to attention deficit hyperactivity disorder. *Psychiatry Res.* 2005;138(2):157–69. <https://doi.org/10.1016/j.psychres.2004.12.002>.
27. Mueller A, Candrian G, Grane VA, Kropotov JD, Ponomarev VA, Baschera GM. Discriminating between ADHD adults and controls using independent ERP components and a support vector machine: a validation study. *Nonlinear Biomed Phys.* 2011;5:5. <https://doi.org/10.1186/1753-4631-5-5>.
28. Liechti MD, Maurizio S, Heinrich H, Jäncke L, Meier L, Steinhausen HC, et al. First clinical trial of tomographic neurofeedback in attention-deficit/hyperactivity disorder: evaluation of voluntary cortical control. *Clin Neurophysiol.* 2012;123(10):1989–2005. <https://doi.org/10.1016/j.clinph.2012.03.016>.
29. Thapar A, Cooper M, Eyre O, Langley K. What have we learnt about the causes of ADHD? *J Child Psychol Psychiatry.* 2013;54:3–16. <https://doi.org/10.1111/j.1469-7610.2012.02611.x>.
30. Hawi Z, Cummins TD, Tong J, Johnson B, Lau R, Samraoui W, et al. The molecular genetic architecture of attention deficit hyperactivity disorder. *Mol Psychiatry.* 2015;20:289–97. <https://doi.org/10.1038/mp.2014.183>.
31. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell.* 2009;136:215–33. <https://doi.org/10.1016/j.cell.2009.01.002>.
32. Mill J, Petronis A. Pre- and peri-natal environmental risks for attention-deficit hyperactivity disorder (ADHD): the potential role of epigenetic processes in mediating susceptibility. *J Child Psychol Psychiatry.* 2008;49:1020–30. <https://doi.org/10.1111/j.1469-7610.2008.01909.x>.
33. Schuch V, Utsumi DA, Costa TV, Kulikowski LD, Muszkat M. Attention deficit hyperactivity disorder in the light of the epigenetic paradigm. *Front Psychiatry.* 2015;6:126. <https://doi.org/10.3389/fpsy.2015.00126>.
34. Kosik KS. The neuronal microRNA system. *Nat Rev Neurosci.* 2006;7:911–20. <https://doi.org/10.1038/nrn2037>.
35. Miller BH, Wahlestedt C. MicroRNA dysregulation in psychiatric disease. *Brain Res.* 2010;1338:89–99. <https://doi.org/10.1016/j.brainres.2010.03.035>.
36. Geaghan M, Cairns MJ. MicroRNA and posttranscriptional dysregulation in psychiatry. *Biol Psychiatry.* 2015;78:231–9. <https://doi.org/10.1016/j.biopsych.2014.12.009>.
37. Issler O, Chen A. Determining the role of microRNAs in psychiatric disorders. *Nat Rev Neurosci.* 2015;16:201–12. <https://doi.org/10.1038/nrn3879>.
38. Wu L, Zhao Q, Zhu X, Peng M, Jia C, Wu W, et al. A novel function of microRNA let-7d in regulation of galectin-3 expression in attention deficit hyperactivity disorder rat brain. *Brain Pathol.* 2010;20:1042–54. <https://doi.org/10.1111/j.1750-3639.2010.00410.x> **This study reported that Rno-let-7d was increased in animal models of ADHD also regulated galectin-3.**
39. Hong Q, Yang L, Zhang M, Pan XQ, Guo M, Fei L, et al. Increased locomotor activity and non-selective attention and impaired learning ability in SD rats after lentiviral vector-mediated RNA interference of Homer 1a in the brain. *Int J Med Sci.* 2013;10:90–102. <https://doi.org/10.7150/ijms.4892>.
40. Yang L, Hong Q, Zhang M, Liu X, Pan XQ, Guo M, et al. The role of Homer 1a in increasing locomotor activity and non-selective

- attention, and impairing learning and memory abilities. *Brain Res.* 2013;1515:39–47. <https://doi.org/10.1016/j.brainres.2013.03.030>.
41. Pietrzykowski AZ, Spijker S. Impulsivity and comorbid traits: a multi-step approach for finding putative responsible microRNAs in the amygdala. *Front Neurosci.* 2014;8:389. <https://doi.org/10.3389/fnins.2014.00389>.
 42. Wu LH, Cheng W, Yu M, He BM, Sun H, Chen Q, et al. Nr3C1-Bhlhb2 axis dysregulation is involved in the development of attention deficit hyperactivity. *Mol Neurobiol.* 2017;54:1196–212. <https://doi.org/10.1007/s12035-015-9679-z>.
 43. Kandemir H, Erdal ME, SeleK S, Ay ÖI, Karababa IF, Kandemir SB, et al. Evaluation of several micro RNA (miRNA) levels in children and adolescents with attention deficit hyperactivity disorder. *Neurosci Lett.* 2014;580:158–62. <https://doi.org/10.1016/j.neulet.2014.07.060>.
 44. Wu LH, Peng M, Yu M, Zhao QL, Li C, Jin YT, et al. Circulating microRNA let-7d in attention-deficit/hyperactivity disorder. *NeuroMolecular Med.* 2015;17:137–46. <https://doi.org/10.1007/s12017-015-8345-y>.
 45. Wu L, Zhao Q, Zhu X, Peng M, Jia C, Wu W, et al. A novel function of microRNA let-7d in regulation of galectin-3 expression in attention deficit hyperactivity disorder rat brain. *Brain Pathol.* 2010;20(6):1042–54.
 46. Wang L-J, Li S-C, Lee M-J, Chou M-C, Chou W-J, Lee S-Y, et al. Blood-borne microRNA biomarker evaluation in attention-deficit/hyperactivity disorder of Han Chinese individuals: an exploratory study. *Front Psychiatry.* 2018;9:227. <https://doi.org/10.3389/fpsy.2018.00227> **This study identified 13 miRNAs as potential ADHD biomarkers that would aid in the diagnosis of ADHD.**
 47. Cortese S. The neurobiology and genetics of attention-deficit/hyperactivity disorder (ADHD): what every clinician should know. *Eur J Paediatr Neurol.* 2012;16(5):422–33.
 48. Friedel S, Saar K, Sauer S, Dempfle A, Walitza S, Renner T, et al. Association and linkage of allelic variants of the dopamine transporter gene in ADHD. *Mol Psychiatry.* 2007;12(10):923–33.
 49. Gizer IR, Ficks C, Waldman ID. Candidate gene studies of ADHD: a meta-analytic review. *Hum Genet.* 2009;126(1):51–90.
 50. Franke B, Faraone SV, Asherson P, Buitelaar J, Bau CHD, Ramos-Quiroga JA, et al. The genetics of attention deficit/hyperactivity disorder in adults, a review. *Mol Psychiatry.* 2012;17(10):960–87.
 51. Kebir O, Joobar R. Neuropsychological endophenotypes in attention-deficit/hyperactivity disorder: a review of genetic association studies. *Eur Arch Psychiatry Clin Neurosci.* 2011;261(8):583–94.
 52. Barnes JJM, Dean AJ, Nandam LS, O'Connell RG, Bellgrove MA. The molecular genetics of executive function: role of monoamine system genes. *Biol Psychiatry.* 2011;69(12):E127–43.
 53. McGough JJ. Attention deficit hyperactivity disorder pharmacogenetics: the dopamine transporter and D4 receptor. *Pharmacogenomics.* 2012;13(4):365–8.
 54. Bruxel EM, Akutagawa-Martins GC, Salatino-Oliveira A, Contini V, Kieling C, Hutz MH, et al. ADHD pharmacogenetics across the life cycle: new findings and perspectives. *Am J Med Genet B Neuropsychiatr Genet.* 2014;165:263–82.
 55. Kambeitz J, Romanos M, Ettinger U. Meta-analysis of the association between dopamine transporter genotype and response to methylphenidate treatment in ADHD. *Pharm J.* 2014;14(1):77–84.
 56. Konrad K, Dempfle A, Friedel S, Heiser P, Holtkamp K, Walitza S, et al. Familiality and molecular genetics of attention networks in ADHD. *Am J Med Genet B Neuropsychiatr Genet.* 2010;153B(1):148–58.
 57. Shang CY, Gau SSF. Association between the DAT1 gene and spatial working memory in attention deficit hyperactivity disorder. *Int J Neuropsychopharmacol.* 2014;17(1):9–21.
 58. Noaín D, Avale ME, Wedemeyer C, Calvo D, Peper M, Rubinstein M. Identification of brain neurons expressing the dopamine D4 receptor gene using BAC transgenic mice. *Eur J Neurosci.* 2006;24(9):2429–38.
 59. Taurines R, Grunblatt E, Schecklmann M, Schwenck C, Albantakis L, Reefschlager L, et al. Altered mRNA expression of monoaminergic candidate genes in the blood of children with attention deficit hyperactivity disorder and autism spectrum disorder. *World J Biol Psychiatry.* 2011;12:104–8.
 60. Wu J, Xiao HF, Sun HJ, Zou L, Zhu LQ. Role of dopamine receptors in ADHD: a systematic meta-analysis. *Mol Neurobiol.* 2012;45(3):605–20.
 61. Kebir O, Joobar R. Neuropsychological endophenotypes in attention-deficit/hyperactivity disorder: a review of genetic association studies. *Eur Arch Psychiatry Clin Neurosci.* 2011;261(8):583–94.
 62. Barnes JJM, Dean AJ, Nandam LS, O'Connell RG, Bellgrove MA. The molecular genetics of executive function: role of monoamine system genes. *Biol Psychiatry.* 2011;69(12):E127–43.
 63. Usiello A, Baik JH, Rouge-Pont F, Picetti R, Dierich A, LeMeur M, et al. Distinct functions of the two isoforms of dopamine D2 receptors. *Nature.* 2000;408(6809):199–203.
 64. Lasky-Su J, Neale BM, Franke B, Anney RJJ, Zhou KX, Maller JB, et al. Genome-wide association scan of quantitative traits for attention deficit hyperactivity disorder identifies novel associations and confirms candidate gene associations. *Am J Med Genet B.* 2008;147B(8):1345–54.
 65. Winsberg BG, Comings DE. Association of the dopamine transporter gene (DAT1) with poor methylphenidate response. *J Am Acad Child Adolesc Psychiatry.* 1999;38(12):1474–7.
 66. Beischlag TV, Marchese A, Meador-Woodruff JH, Damask SP, O'Dowd BF, Tyndale RF, et al. The human dopamine D5 receptor gene: cloning and characterization of the 5'-flanking and promoter region. *Biochemistry.* 1995;34(17):5960–70.
 67. Manor I, Corbex M, Eisenberg J, Gritsenko I, Bachner-Melman R, Tyano S, et al. Association of the dopamine D5 receptor with attention deficit hyperactivity disorder (ADHD) and scores on a continuous performance test (TOVA). *Am J Med Genet B.* 2004;127B(1):73–7.
 68. Froehlich TE, McGough JJ, Stein MA. Progress and promise of attention-deficit hyperactivity disorder pharmacogenetics. *CNS Drugs.* 2010;24(2):99–117.
 69. Stahl SM. Neurotransmission of cognition, part 3. Mechanism of action of selective NRIs: both dopamine and norepinephrine increase in prefrontal cortex. *J Clin Psychiatry.* 2003;64(3):230–1.
 70. Lasky-Su J, Neale BM, Franke B, Anney RJJ, Zhou KX, Maller JB, et al. Genome-wide association scan of quantitative traits for attention deficit hyperactivity disorder identifies novel associations and confirms candidate gene associations. *Am J Med Genet B.* 2008;147B(8):1345–54.
 71. Mick E, Neale B, Middleton FA, McGough JJ, Faraone SV. Genome-wide association study of response to methylphenidate in 187 children with attention-deficit/hyperactivity disorder. *Am J Med Genet B.* 2008;147B(8):1412–8.
 72. Yang L, Qian Q, Liu L, Li H, Faraone SV, Wang Y. Adrenergic neurotransmitter system transporter and receptor genes associated with atomoxetine response in attention-deficit hyperactivity disorder children. *J Neural Transm.* 2013;120(7):1127–33.
 73. Meng WD, Sun SJ, Yang J, Chu RX, Tu W, Liu Q. Elevated serum brain-derived neurotrophic factor (BDNF) but not BDNF gene Val66Met polymorphism is associated with autism spectrum disorders. *Mol Neurobiol.* 2016;54:1167–72. <https://doi.org/10.1007/s12035-016-9721-9>.
 74. de Azevedo CT, Mondin TC, Wiener CD, Marques MB, Fucolo Bde A, Pinheiro RT, et al. Neurotrophic factors, clinical features and gender differences in depression. *Neurochem Res.* 2014;39:1571–8.

75. Domingos da Silveira da Luz AC, Pereira Dias G, do Nascimento Bevilacqua MC, Cocks G, Gardino PF, Thuret S, et al. Translational findings on brain-derived neurotrophic factor and anxiety: contributions from basic research to clinical practice. *Neuropsychobiology*. 2013;68:129–38.
76. Kotyuk E, Keszler G, Nemeth N, Ronai Z, Sasvari-Szekely M, Szekely A. Glial cell line-derived neurotrophic factor (GDNF) as a novel candidate gene of anxiety. *PLoS One*. 2013;8:e80613.
77. Vargas HE, Gama CS, Andreazza AC, Medeiros D, Stertz L, Fries G, et al. Decreased serum neurotrophin 3 in chronically medicated schizophrenic males. *Neurosci Lett*. 2008;440:197–201.
78. Walz JC, Andreazza AC, Frey BN, Cacilhas AA, Cereser KM, Cunha AB, et al. Serum neurotrophin-3 is increased during manic and depressive episodes in bipolar disorder. *Neurosci Lett*. 2007;415:87–9.
79. Wysokinski A. Serum levels of brain-derived neurotrophic factor (BDNF) and neurotrophin-3 (NT-3) in depressed patients with schizophrenia. *Nord J Psychiatry*. 2016;70:267–71.
80. Bergman O, Westberg L, Lichtenstein P, Eriksson E, Larsson H. Study on the possible association of brain-derived neurotrophic factor polymorphism with the developmental course of symptoms of attention deficit and hyperactivity. *Int J Neuropsychopharmacol*. 2011;14:1367–76.
81. Shim SH, Hwangbo Y, Kwon YJ, Jeong HY, Lee BH, Lee HJ, et al. Increased levels of plasma brain-derived neurotrophic factor (BDNF) in children with attention deficit-hyperactivity disorder (ADHD). *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2008;32:1824–8.
82. Li H, Liu L, Tang Y, Ji N, Yang L, Qian Q, et al. Sex-specific association of brain-derived neurotrophic factor (BDNF) Val66Met polymorphism and plasma BDNF with attention-deficit/hyperactivity disorder in a drug-naïve Han Chinese sample. *Psychiatry Res*. 2014;217:191–7.
83. Amiri A, Torabi Parizi G, Kousha M, Saadat F, Modabbernia MJ, Najafi K, et al. Changes in plasma brain-derived neurotrophic factor (BDNF) levels induced by methylphenidate in children with attention deficit-hyperactivity disorder (ADHD). *Prog Neuro-Psychopharmacol Biol Psychiatry*. 2013;47:20–4.
84. Ramos-Quiroga JA, Corominas-Roso M, Palomar G, Gomez-Barros N, Ribases M, Sanchez-Mora C, et al. Changes in the serum levels of brain-derived neurotrophic factor in adults with attention deficit hyperactivity disorder after treatment with atomoxetine. *Psychopharmacology*. 2014;231:1389–95.
85. Rios M, Fan G, Fekete C, Kelly J, Bates B, Kuehn R, et al. Conditional deletion of brain-derived neurotrophic factor in the postnatal brain leads to obesity and hyperactivity. *Mol Endocrinol*. 2001;15:1748–57.
86. Chourbaji S, Hellweg R, Brandis D, Zorner B, Zacher C, Lang UE, et al. Mice with reduced brain-derived neurotrophic factor expression show decreased choline acetyltransferase activity, but regular brain monoamine levels and unaltered emotional behavior. *Brain Res Mol Brain Res*. 2004;121:28–36.
87. Scassellati C, Zanardini R, Tiberti A, Pezzani M, Valenti V, Effedri P, et al. Serum brain-derived neurotrophic factor (BDNF) levels in attention deficit-hyperactivity disorder (ADHD). *Eur Child Adolesc Psychiatry*. 2014;23:173–7.
88. Tzang RF, Hsu CD, Liou YJ, Hong CJ, Tsai SJ. Family based association of the brain-derived neurotrophic factor gene in attention-deficit hyperactivity disorder. *Psychiatr Genet*. 2013;23(4):177–8.
89. Naumenko VS, Bazovkina DV, Semenova AA, Tsybko AS, Il'chibaeva TV, Kondaurova EM, et al. Effect of glial cell line-derived neurotrophic factor on behavior and key members of the brain serotonin system in mouse strains genetically predisposed to behavioral disorders. *J Neurosci Res*. 2013;91:1628–38.
90. Gerlai R, McNamara A, Choi-Lundberg DL, Armanini M, Ross J, Powell-Braxton L, et al. Impaired water maze learning performance without altered dopaminergic function in mice heterozygous for the GDNF mutation. *Eur J Neurosci*. 2001;14:1153–63.
91. Shim SH, Hwangbo Y, Yoon HJ, Kwon YJ, Lee HY, Hwang JA, et al. Increased levels of plasma glial-derived neurotrophic factor in children with attention deficit hyperactivity disorder. *Nord J Psychiatry*. 2015;69:546–51 **This study was conducted on 86 drug naïve pediatric patients with ADHD which showed increased levels of plasma GDNF levels. The plasma GDNF levels showed a positive correlation with inattention and hyperactivity-impulsivity.**
92. Aloe L, Rocco ML, Bianchi P, Manni L. Nerve growth factor: from the early discoveries to the potential clinical use. *J Transl Med*. 2012;10:239.
93. Sarter M, Gehring WJ, Kozak R. More attention must be paid: the neurobiology of attentional effort. *Brain Res Rev*. 2006;51:145–60.
94. Syed Z, Dudbridge F, Kent L. An investigation of the neurotrophic factor genes GDNF, NGF, and NT3 in susceptibility to ADHD. *Am J Med Genet B Neuropsychiatr Genet*. 2007;144B:375–8.
95. Maness LM, Kastin AJ, Weber JT, Banks WA, Beckman BS, Zadina JE. The neurotrophins and their receptors: structure, function, and neuropathology. *Neurosci Biobehav Rev*. 1994;18:143–59.
96. Park S, Kim BN, Kim JW, Shin MS, Cho SC, Kim JH, et al. Neurotrophin 3 genotype and emotional adverse effects of osmotic-release oral system methylphenidate (OROS-MPH) in children with attention-deficit/hyperactivity disorder. *J Psychopharmacol*. 2014;28:220–6 **This study provided evidence that genetic variation of the NTF3 gene was related to susceptibility of emotional side effects in response to treatment with methylphenidate.**
97. Bilgiç A, Tokar A, Işık Ü, Kılınç İ. Serum brain-derived neurotrophic factor, glial-derived neurotrophic factor, nerve growth factor, and neurotrophin-3 levels in children with attention deficit/hyperactivity disorder. *Eur Child Adolesc Psychiatry*. 2017;26(3):355–63. <https://doi.org/10.1007/s00787-016-0898-2>.
98. Oades RD. Role of the serotonin system in ADHD: treatment implications. *Expert Rev Neurother*. 2007;7(10):1357–74.
99. Oades RD. Dopamine-serotonin interactions in attention-deficit hyperactivity disorder (ADHD). *Prog Brain Res*. 2008;172:543–65.
100. Ludolph AG, Kassubek J, Schmeck K, Glaser C, Wunderlich A, Buck AK, et al. Dopaminergic dysfunction in attention deficit hyperactivity disorder (ADHD), differences between pharmacologically treated and never treated young adults: a 3,4-dihydroxy-6 [18F]fluorophenyl-l-alanine PET study. *Neuroimage*. 2008;41(3):718–27.
101. Seo D, Patrick CJ, Kennealy PJ. Role of serotonin and dopamine system interactions in the neurobiology of impulsive aggression and its comorbidity with other clinical disorders. *Aggress Violent Behav*. 2008;13(5):383–95.
102. Nordquist N, Creland L. Serotonin, genetic variability, behavioral, and psychiatric disorders-a review. *Ups J Med Sci*. 2010;115(1):2–10.
103. Arnsten AF. Catecholamine influences on dorsolateral prefrontal cortical networks. *Biol Psychiatry*. 2011;69(12):89–99.
104. Volkow ND, Wang GJ, Kollins SH, Wigal TL, Newcorn JH, Telang F, et al. Evaluating dopamine reward pathway in ADHD: clinical implications. *JAMA*. 2009;302(10):1084–91.
105. Volkow ND, Wang GJ, Newcorn J, Telang F, Solanto MV, Fowler JS, et al. Depressed dopamine activity in caudate and preliminary evidence of limbic involvement in adults with attention deficit/hyperactivity disorder. *Arch Gen Psychiatry*. 2007;64(8):932–40.

106. Pliska SR, McCracken JT, Maas JW. Catecholamines in attention-deficit hyperactivity disorder: current perspective. *J Am Acad Child Adolesc Psychiatry*. 1996;35:264–72.
107. Ernst M, Zametkin AJ, Matochik JA, Jons PH, Cohen RM. Dopa decarboxylase activity in attention deficit hyperactivity disorders adults. A [fluorine-18] flourodopa positron emission tomography study. *J Neurosci*. 1998;18(15):5901–7.
108. Zametkin AJ, Ernst M. Current concepts: problems in the management and treatment of attention deficit hyperactivity disorder. *N Engl J Med*. 1999;340(1):40–8.
109. Courvoisier H. Neurometabolic functioning and neuropsychological correlates in children with ADHD-H: preliminary findings. *J Neuropsychiatr Clin Neurosci*. 2004;16:63–9.
110. Hoshino Y, Ohno Y, Yamamoto T, Kaneko M, Kumashiro H. Plasma free tryptophan concentration in children with attention deficit disorder. *Folia Psychiatr Neurol Jpn*. 1985;39(4):531–5.
111. Dolina S, Margalit D, Malitsky S, Rabinkov A. Attention-deficit hyperactivity disorder (ADHD) as a pyridoxine-dependent condition: urinary diagnostic biomarkers. *Med Hypotheses*. 2014;82(1):111–6. <https://doi.org/10.1016/j.mehy.2013.11.018> **This study reported ratios of levels of certain enzymes to be biomarkers of ADHD and reported low concentrations of monoamines and disordered amino acid metabolism to be an inherent cause of ADHD.**
112. Anand D, Colpo GD, Zeni G, Zeni CP, Teixeira AL. Attention-deficit/hyperactivity disorder and inflammation: What does current knowledge tell us? A systematic review. *Front Psychiatry*. 2017;8:228.
113. Oades RD, Dauvermann MR, Schimmelmann BG, Schwarz MJ, Myint AM. Attention-deficit hyperactivity disorder (ADHD) and glial integrity: S100B, cytokines and kynurenine metabolism effects of medication. *Behav Brain Funct*. 2010;6:29.
114. Oades RD, Myint AM, Dauvermann MR, Schimmelmann BG, Schwarz MJ. Attention-deficit hyperactivity disorder (ADHD) and glial integrity: an exploration of associations of cytokines and kynurenine metabolites with symptoms and attention. *Behav Brain Funct*. 2010;6:32.
115. Avcil S. Evaluation of the neutrophil/lymphocyte ratio, platelet/lymphocyte ratio, and mean platelet volume as inflammatory markers in children with attention-deficit hyperactivity disorder. *Psychiatry Clin Neurosci*. 2018;72(7):522–30. <https://doi.org/10.1111/pcn.12659>.

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