REVIEW

Therapeutic Efects of Coenzyme Q10 in the Treatment of Ischemic Stroke

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Abstract

Purpose of Review Ischemic stroke is the second deadly disease worldwide, but current treatment is very limited. The brain, rich in lipids and high in oxygen consumption, is susceptible to damage from oxidative stress after ischemic stroke. Thus, antioxidants are promising neuroprotective agents for treatment and prevention of ischemic stroke. Coenzyme Q10 is the only lipophilic antioxidant that can be synthesized de novo by cells and plays a key role as an electron carrier in the oxidative phosphorylation of the mitochondrial electron transport chain. However, the reduced form of coenzyme Q10 (Ubiquinol) levels are signifcantly defcient in the brain. The aim of this article is to review the therapeutic efects and mechanisms of coenzyme Q10 in ischemic stroke.

Recent Findings Current studies have found that coenzyme Q10 protects and treats ischemic stroke through multiple mechanisms based on the evidence from in vitro experiments, in vivo experiments, and clinical observations.

Summary For the first time, we reviewed the neuroprotective effects of coenzyme Q10 in ischemic stroke. Coenzyme Q10 exerts neuroprotective efects after ischemic stroke through anti-oxidative stress, anti-nitrosative stress, anti-infammation, and anti-cell death. Here, we provided the evidence on the therapeutic and preventive efects of coenzyme Q10 in ischemic stroke and suggested the potential value of coenzyme Q10 as a medication candidate.

Keywords Coenzyme Q10 · Ischemic stroke · Antioxidant · Neuroprotective efect · Atherosclerosis · Blood-brain barrier

Introduction

Stroke is a common disease with high incidence, high mortality and high disability, and the burden of stroke ranks the second in the world $[1]$ $[1]$ $[1]$. Stroke includes hemorrhagic

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stroke and ischemic stroke (IS), with IS accounting for more than 80% [[2](#page-8-1)]. IS is a common disease in the elderly and tends to occur at a younger age in recent years. Currently, about 80 million people worldwide sufer from IS [[3\]](#page-8-2). As the population ageing processes, more than a quarter of the world's population will be over 65 years old by 2050, which will further increase the number of IS patients [[4\]](#page-8-3). Therefore, sequelae of IS has brought heavy burden to patients, families and society.

Currently, thrombolysis and mechanical thrombolysis are efective treatments for IS, but their clinical application is limited by strict contraindications, narrow treatment time windows, and severe side efects, resulting in only a small number of patients benefting from them [\[5](#page-8-4)]. In addition, the incidence of hemorrhagic transformation (HT) following the treatment of thrombolysis and mechanical thrombolysis was close to 10% [[6\]](#page-8-5). HT is an important factor in the exacerbation of brain injury and death in IS patients. Preventive strategies for IS have greatly reduced the chance of IS, however, there is still a large number of patients with risk factors such as smoking, alcoholism, diabetes, hypertension, especially in developing countries. Therefore, it is important to develop affordable drugs for the treatment of IS.

The pathological mechanism of IS is complex, and oxidative stress plays an important role in the pathological progression of IS. Thus, there is a trend to develop efective antioxidants for the treatment of IS [\[7\]](#page-9-0). Coenzyme Q10 (CoQ10) is a fat-soluble antioxidant and is widely synthesized in the body. However, in vivo synthesis of CoQ10 is often inadequate. CoQ10 plays a key role in oxidative phosphorylation in mitochondria, where it protects cell membranes from free radical-induced oxidative damage. It also plays important roles in other organelles, including lysosome, Golgi apparatus, endoplasmic reticulum, and among others. For example, CoQ10 maintains the pH in lysosomes [\[8](#page-9-1)]. In addition, CoQ10 can be obtained from a normal diet of about fve mg per day [\[9](#page-9-2)]. However, the daily requirement for CoQ10 is estimated to be about 500 mg per day [\[10](#page-9-3)]. Therefore, CoQ10 is likely to be necessary through dietary supplementation.

Many studies have reviewed the role of CoQ10 in neurological disorders, such as Parkinson's disease [\[11\]](#page-9-4), cerebral hemorrhage [[12](#page-9-5)], epilepsy [[13](#page-9-6)], migraine [[14](#page-9-7)], and multiple sclerosis [[15](#page-9-8)]. In addition, recent studies have found that serum CoQ10 levels are signifcantly lower in patients after IS and associated with clinical outcomes [[16](#page-9-9)]. However, the treatment of IS with $CoQ10$ has not been reviewed. For the frst time, we reviewed the therapeutic efects of CoQ10 for IS. Herein, we aim to provide evidences for the treatment of IS with CoQ10.

The Role of CoQ10 in the Treatment for IS

CoQ10 plays important roles in the treatment of IS. The detailed information are shown in Table [1.](#page-2-0) A study has found that in IS patients, the serum CoQ10 levels were signifcantly lower, and serum CoQ10 levels were signifcantly negatively correlated with National Institutes of Health Stroke Scale (NIHSS) scores and Modifed Rankin Scale (mRS) scores, suggesting the serum CoQ10 could be an indicator for the clinical neurological outcomes after IS [\[16](#page-9-9)]. Also, a recent clinical study has found that the intake of 300 mg/day of CoQ10 for one month has signifcantly improved IS outcomes, and increased NIHSS scores [\[17](#page-9-10)].

In addition to clinical observations, the animal model also demonstrated the role of CoQ10 in the treatment for IS. Middle cerebral artery occlusion (MCAO**)** model is the most commonly used animal model for IS. In a 10-min MACO model, 10 mg/kg CoQ10 has signifcantly decreased neurological deficit scores, brain infarct volume, and blood glucose levels [\[18](#page-9-11)]. The pretreatment of CoQ10 at 10 mg/kg alleviated the biochemical changes in both brain and serum, and the hippocampal damage in CA1 region after 60-min MACO model [\[19](#page-9-12), [20](#page-9-13)]. With the supplement at 30 mg/kg, CoQ10 has signifcantly decreased neurological defcit scores, mortality, and brain infarct volume, and attenuated motor impairment brain edema, cognitive impairment, and blood-brain barrier integrity in a 60-min MACO model [[21–](#page-9-14)[23](#page-9-15)]. Additionally, the 200 mg/kg CoQ10 has also reduced neurological defcit score and improved cerebral infarction in a 60-min MACO model [\[24\]](#page-9-16). In a bilateral common carotid artery occlusion (BCCAO) model, the post-IS treatment with CoQ10 at 200 mg/kg has signifcantly reduced brain edema, and improved anxiety and depression-like behavior, and cognitive impairments [\[25](#page-9-17)]. Additionally, in a photothrombotic stroke model for neonatal rats, pretreatment with 200 mg/kg CoQ10 decreased cerebral ischemic injury and brain infarct volume, while post-stroke treatment with CoQ10 improved motor function and reduced brain infarct volume, with a better treatment efect than the pretreatment [[26](#page-9-18)].

There are very limited studies using CoQ10 in the treatment of IS with the in vitro models. A study reported that CoQ10 could protect neural stem cells against hypoxiareperfusion by increasing cell viability and decreasing formation of free radicals in an oxygen glucose deprivation/ re-oxygenation (OGD/R) model [[27\]](#page-9-19).

The Efects and Potential Mechanisms of CoQ10 in the Treatment for IS

Here, we focus on the beneficial effects of $CoQ10$ on the prevention of IS etiology (i.e. atherosclerosis, atrial fbrillation, cerebral small vessel disease), pathogenesis (i.e. oxidative stress, infammation, endothelial damage, etc.) and other interventional risk factors of IS.

The therapeutic mechanisms of CoQ10 in IS are not clearly understood. Current studies suggest that the mechanisms of CoQ10 involves anti-oxidative stress, anti-nitrosative stress, anti-infammatory efect, anti-apoptotic efect, and anti-autophagy, mainly focusing on the benefcial efects of CoQ10 on the prevention of IS pathogenesis. The specifc mechanism and information are shown in Fig. [1](#page-2-1).

CoQ10 has been clinically proven efective in protecting ischemic stroke. Now, one of the clear therapeutic mechanisms of CoQ10 over IS is through the miRNA-149-5-p and the NF-κB to reduce infammation factors, such as MMP-2, MMP-9, IL1β, IL6, TNF-α, and iNOS. Therefore, the integrity of blood-brain barrier has improved, and brain edema and hemorrhagic transformation are reduced. In addition, CoQ10 can reduce oxidative stress to regulate apoptosis and autophagy, and increase the survival of nerve cells after IS, including through increasing the levels of glutathione and antioxidants, reducing reactive oxygen species (ROS),

Table 1 Details of included studies and results of CoQ10 in the treatment of IS

	Year Species	Dose	Model		IS Time Evidence	References
	2020 Rats	30 mg/kg/day	MCAO	60 min	Neurological deficit scores↓, mortality↓, coenzyme Q10 ₁	22
2020 Rats		100 mg/kg/day	Photothrombosis 7 d		Activated microglia, ROS, motor function ^{\uparrow} , Neuro- logical deficit scores, Brain Infarct Volume, IL-1β, $TNF-\alpha$	26
	2018 Human	300 mg/day		30d	coenzyme Q10 ^{\dagger} , NIHSS \dagger , MMSE \dagger	17
		2012 Neural stem cells 0.01, 0.1, 1, 10 μM OGD/R		8 h	ROS _L , Akt [†] , GSK3 β [†] , Bcl-2 [†] , Bax _L , cleaved cas- pase- $3\downarrow$	27
2019 Rats		200 mg/kg/day	MCAO	60 min	Neurological deficit scores \downarrow , IL-6 \downarrow , TNF- $\alpha \downarrow$, cas- pase-3, caspase-9	24
2021 Rats		30 mg/kg/day	MCAO	60 min	Neurological deficit scores, Brain Infarct Volume, Blood-brain barrier permeability, miRNA-149-5p1, MMP2↓, MMP9↓, TNF-α↓, IL-6↓, tissue damage volume.	21
2017 Rats		10 mg/kg/day	MCAO	30 min	Neurological deficit scores L, Brain Infarct Volume L, blood glucose, Lc3-II/Lc3I,	18
2022 Rats		200 mg/kg/day	BCCAO	20 min	Brain edema, depressive and anxiety-like behavior, cognitive impairments, BDNF1, SOD1	25
2010 Rats		10 mg/kg/day	MCAO	60 min	LDH \downarrow , Ca ²⁺ \downarrow , MDA \downarrow , glutathione \uparrow	19
2015 Rats		30 mg/kg/day	MCAO	60 min	Neurological deficit scores L, mortality L, motor func- tion ^{\uparrow} , coenzyme O10 \uparrow , Brain Infarct Volume	23
2017 Rats		10 mg/kg/day	MCAO	60 min	cytochrome C _k , caspase-3k, Bax _k , Bcl-2 ^t , p-Akt [*] , p -FOXO3A \uparrow , JNK3 \downarrow , c-Jun \downarrow , TNF- $\alpha \downarrow$, ICAM-1 \downarrow , MPO activity \downarrow , NF- κ Bp65 \downarrow , iNOS \downarrow , Bim \downarrow	20

 \uparrow =increased, \downarrow =decreased

OGD/R Oxygen glucose deprivation/reoxygenation, *MCAO* Middle cerebral artery occlusion, *Time* Time of ischemic stroke model

regulating the expression of Caspase3, Caspase9 and other proteins. The above are the specifc molecular mechanisms by which CoQ10 protects IS from reducing neurological deficits, infarct volume, and promoting motor function and intellectual recovery.

The Efect of CoQ10 Against Oxidative Stress

Oxidative stress is critical in the pathology of IS, and excess ROS is a key mechanism in the development of IS. In the clinical studies, CoQ10 reduced ROS levels in IS patients,

and improves prognosis [[17](#page-9-10)]. In addition, both in vivo and in vitro experiments found that CoQ10 increased glutathione levels and superoxide dismutase (SOD), and decreased ROS, lactate dehydrogenase (LDH), and malondialdehyde (MDA) levels [\[19](#page-9-12), [25](#page-9-17)[–27](#page-9-19)].

In addition to the direct evidence of CoQ10 against IS, CoQ10 also displayed protection against atherosclerosis (AS), which is the most common factor leading to IS. There are two mechanisms by which AS triggers IS. One is a stable atherosclerotic plaque that gradually increases in size

Fig. 1 Schematic illustration of the therapeutic efects of CoQ10 for IS. CoQ10 improves brain tissue damage and function after IS through antiinfammatory, anti-oxidative stress, anti-apoptosis and anti-autophagy

and encroaches on the lumen of the vessel, blocking blood flow and causing hypoperfusion or even complete ischemia. These patients were usually with symptoms and detected through examinations, often without serious consequences. In the other case, the plaque dislodgement leads to the formation of a blood clot, which blocks the blood vessel and usually causes an acute IS with serious consequences.

ROS are essential for vascular homeostasis, but excess ROS cause vascular damage [\[28\]](#page-9-20). The imbalance of oxidative stress leads to oxidative damage to endothelial cells and production of oxidized LDL (ox-LDL). The release of chemotactic proteins from damaged endothelial cells leads to migration of immune cells to the endothelium [\[29](#page-9-21)]. Further, ox-LDL increases the expression of cell adhesion molecules, leading to the aggregation of immune cells. Subsequently, monocytes differentiate into macrophages that mediate phagocytosis of ox-LDL, and these lipid-rich macrophages are known as foam cells. This is also a hallmark of early AS. These immune cells release a variety of cytokines that promote ROS production, further leading to oxidative stress. ROS and infammatory factors stimulate migration of smooth muscle cell and deposition of collagen, and the atherosclerotic plaque increases in size and invades the medial arterial. Eventually, the atherosclerotic plaque completely forms, leading to a decrease in arterial elasticity.

Recent study was found that the serum CoQ10 level was signifcantly decreased, and the 8-hydroxy-2' -deoxyguanosine/deoxyguanosine ratio, MDA level, and glutathione peroxidase (GPX) were signifcantly increased in patients with AS and hyperlipidemia [[30](#page-9-22)]. It suggests that CoQ10 may play an important role in the pathological process of AS. CoQ10 can reduce LDH and MDA levels, and increase SOD and GSH levels, suggesting that CoQ10 exerts protective effects through anti-oxidative stress in AS $[31]$ $[31]$.

Elevated levels of LDL and ox-LDL and decreased levels of high-density lipoprotein (HDL) are key factors in the development and progression of AS. Macrophage phagocytosis and uptake of ox-LDL are the basis for AS formation. Oxidation of HDL has multiple functions, including reverse cholesterol transport and blocking LDL from becoming ox-LDL. CoQ10 may be involved in inhibiting oxidative stress via miR-378, promoting macrophage cholesterol efflux and reducing ox-LDL-induced foam cell formation [\[32](#page-9-24)]. In addition, myeloperoxidase levels are elevated in patients with AS, and myeloperoxidase is involved in the oxidation of LDL, HDL and nitric oxide (NO) [[33](#page-9-25)]. CoQ10 may have reduced myeloperoxidase levels after IS [[20](#page-9-13)]. NO can be considered as a powerful antioxidant that protects cells and mitochondria during oxidative stress [\[34](#page-9-26)].

CoQ10 treatment increased serum lipocalin, NO, HDL, glutathione, and eNOS levels, and decreased lipid profle, creatine kinase activity, inducible nitric oxide synthase (iNOS), ox-LDL and lipid peroxidation [\[35](#page-9-27), [36](#page-9-28)]. Interestingly, the clinical effects of CoQ10 in combination with simvastatin are controversial. One study found that 400 mg daily of CoQ10 combined with simvastatin for eight weeks did not appear an additional augmentation efect compared to simvastatin treatment [\[37](#page-9-29)]. In contrast, another study found that simvastatin only combined with CoQ10 can achieve normalization of plasma NO levels [[38\]](#page-9-30). This diference may need to be explained by future clinical studies with larger samples.

In addition, CoQ10 may be involved in the treatment of AS through other mechanisms. One study found that lipid changes in patients treated with CoQ10 were inconsistent with a general improvement in antioxidative status, suggesting that the anti-atherosclerotic efects of CoQ10 may be mediated by other mechanisms to regulate antioxidative pathway. For example, paraoxonase 1 (PON1) was originally found to hydrolyze toxic metabolites of a variety of organophosphorus pesticides. PON1 is now also found to mediate the enzymatic protection of LDL. PON1 binds tightly to HDL and destroys ox-LDL by hydrolyzing lipid peroxides. PON1 activity was signifcantly increased during CoQ10 treatment and PON1 activity was positively correlated with HDL levels and negatively correlated with atherosclerotic coefficient [[39,](#page-9-31) [40\]](#page-9-32). In addition, the role of HIF1 α is important in IS and AS, and CoQ10 reduces oxidative stress by inhibiting HIF1 α [[41\]](#page-10-0).

As the pathological process of AS develops, atherosclerotic plaques form calcifcation. The mechanisms of atherosclerotic plaque formation are discussed further in the infammation section below. ROS generated by oxidative stress may be involved in instability and rupture of atherosclerotic plaque, and ROS are involved in AS by promoting proliferation and migration of vascular smooth muscle cell. Also, ROS induce apoptosis in vascular smooth muscle cells, which may cause instability and rupture of atherosclerotic plaque [[42](#page-10-1)]. CoQ10 can inhibit the excessive release of ROS, increase ATP content, improve mitochondrial function, control the development of AS, and facilitate the stability of atherosclerotic plaques [[31,](#page-9-23) [36\]](#page-9-28).

The Efect of CoQ10 Against Nitrosative Stress

Nitrosative stress is an important disorder of oxygen metabolism that is closely associated with the pathological processes of AS and IS. However, the role of nitrosative stress in the development of AS and IS pathology is often overlooked. This may be due to the fact that nitrosative stress and oxidative stress are very similar, and the ROS include superoxide anion (O^{2-}) , hydroxyl radical, hydrogen peroxide (H_2O_2) , and NO, which are overlapped with the formation and scavenging pathways of reactive nitrogen species (RNS) [\[43](#page-10-2), [44\]](#page-10-3). The diference is that nitrosative stress refers to the combination of NO and \cdot O^{2−} to generate peroxynitrite anion (ONOO−) when the body undergoes a disturbance in oxygen metabolism. ONOO− can nitrate biomolecules such as proteins, lipids and DNA [\[44](#page-10-3)]. Nitrosative stress-generated ONOO− promotes protein tyrosine nitration, leading to lipid peroxidation, DNA strand breaks, and cell death in macrophages and smooth muscle cells, and promotes the formation of lipid-rich necrotic cores in AS [[45\]](#page-10-4). And nitrosative stress is also an important factor in the outcome of IS patients. Since the penetration ability of ONOO− to lipid membrane is about 400 times higher than \cdot O^{2−}, it has more ability to damage DNA and more likely to cause neuronal cell death in the ischemic semidark zone [[46](#page-10-5)]. The mechanism of CoQ10 against atherosclerotic nitrosative stress, DNA damage and repair is unclear. It may be related to the inhibition of O^{2-} to further reduce the production of ONOO− by CoQ10, which needs to be confrmed in future experiments [[47\]](#page-10-6). The specifc mechanism of anti-oxidative stress and anti-nitrosative stress was shown in Fig. [2](#page-4-0).

The Efect of CoQ10 Against Infammation

Neuroinfammation plays a key role in the pathogenesis of IS, and the post-IS infammatory response is initiated by the activation of microglia. CoQ10 can inhibit infammation by reducing Interleukin-1β (IL-1β), Interleukin- 6 (IL-6), iNOS, nuclear factor-kappaB (NF-κB) and tumor necrosis factor alpha (TNF- α) levels after IS [[24,](#page-9-16) [26\]](#page-9-18). In addition, CoQ10 protects blood-brain barrier integrity by resisting autophagy and reducing matrix metalloproteinase-2 (MMP-2) and metalloproteinase-9 (MMP-9) expression [[18,](#page-9-11) [21](#page-9-14)]. These efects are inextricably linked to the anti-infammatory and antioxidative stress mechanisms of CoQ10.

The nature of AS is a chronic vascular infammatory disease characterized by the accumulation of lipids and infammatory cells [[48\]](#page-10-7). Therefore, the pathogenesis of AS in addition to oxidative stress, also includes activation of pro-infammatory signaling pathways and enhanced immune response. Moreover, oxidative stress and infammation are inextricably linked, and both are causally involved in regulating the pathological process of IS and AS. CoQ10 can reduce the release of multiple cytokines in AS, including CD11, CD35, CD62, CD63, CD40, PF4, β-TG, CCL5 [[40,](#page-9-32) [49\]](#page-10-8), sP-selectin, C-reactive protein [\[50](#page-10-9)], THP-1 [\[36](#page-9-28)], ICAM-1, VCAM-1, IL-1, IL-6, TNF-α, and NLRP3 [[31](#page-9-23)].

Diferentiation of macrophages is the most important mechanism in the development of AS, and later various types of immune cells in atherosclerotic arteries were discovered one after another [[51\]](#page-10-10). Macrophages play a crucial role in the development and progression of AS by phagocytosing ox-LDL through scavenger receptors, triggering infammation and subsequent formation of foam cells. These

Fig. 2 Schematic illustration of the anti-oxidative stress efect of CoQ10 after IS. Oxidative stress and nitrosative stress promote ROS and RNS release. Further, the combination of O^{2-} and NO generates ONOO−, leading to DNA damage. CoQ10 prevents IS by inhibiting the development of AS through anti-oxidative stress, regulation of RNS, ROS

lipid-laden macrophages are important in plaque rupture by producing various pro-infammatory factors. CoQ10 has multiple regulatory effects on immune cells. CoQ10 treatment increases tocopherol levels of monocyte and decreases CD11 and CD35 levels [\[40](#page-9-32)].

At the onset of AS, endothelial cells release signals to recruit monocytes, which enter the intima and diferentiate into macrophages to phagocytose ox-LDL, exacerbating the infammatory response and releasing chemokines, which increase plaque volume causing hemodynamic changes and further promoting plaque formation. The role of macrophages is to remove pro-infammatory ox-LDL from the endothelium. Macrophages become foam cells once they phagocytize ox-LDL [\[52](#page-10-11)]. These macrophages also increase the expression of lectin-like oxidized LDL (LOX-1) and ATP-binding cassette transporter proteins, such as ATPbinding cassette protein G1 (ABCG1), which are involved in reverse cholesterol transport [[52\]](#page-10-11). CoQ10 may be involved in atherosclerotic protection by inducing ABCG1 expression and enhancing HDL-mediated cholesterol efflux from macrophages [[53](#page-10-12)].

The metabolic status of immune cells is signifcantly altered during atherosclerotic pathology. Metabolic reprogramming plays an important role in the development of AS, and infammation promotes metabolic reprogramming of macrophages. This metabolic reprogramming promotes phagocytosis, cytokine release and ROS production, exacerbating infammation, oxidative stress and immune responses [\[54\]](#page-10-13). Under physiological conditions, monocytes and lymphocytes rely primarily on oxidative phosphorylation for energy production. The accumulation of lipids in macrophages leads to a reprogramming of cellular metabolism and a metabolic shift towards glycolysis, which enhances their pro-infammatory activity. This process includes reduced phagocytosis of macrophages and increased production of pro-infammatory cytokines. The ox-LDL also induces increased glucose uptake by macrophages in a dose- and time- dependent manner, accompanied by an increase in lactate production [[55](#page-10-14)]. Glucose over-utilization promotes IL-6 and IL-1β secretion by monocytes and macrophages, and increases infammation [[56\]](#page-10-15).

Metabolic reprogramming also plays an important role in the regulation of macrophage polarization. Macrophages are mainly classifed into pro-infammatory M1 and antiinfammatory M2 phenotypes. M1 macrophages are mainly dependent on glycolysis, whereas M2 cells are more dependent on oxidative phosphorylation [[57](#page-10-16)]. Inhibition of glycolysis reduces the M1 phenotype of macrophages. However, monocyte glycolytic enzyme expression levels appear elevated in patients with AS, exhibiting a pro-infammatory phenotype [[58](#page-10-17)]. CoQ10 plays a key role in electron transfer in mitochondrial oxidative phosphorylation [[59\]](#page-10-18). Thus, CoQ10 may be useful in the metabolic reprogramming of AS.

In addition to monocytes, multiple cell types are active in AS, such as neutrophils, lymphocytes, smooth muscle cells and endothelial cells, and they are also involved in the formation of AS. Due to the specifcity of endothelial cells (endothelial damage is the earliest event of AS), the main role of endothelial cells is discussed in the next section.

In atherosclerotic lesions, there is also a diferentiation of foam cells from smooth muscle cells (SMCs), which make

up the majority of the cells, and they are mainly involved in intimal thickening. As time progresses, foam cells infltrate into the intima, which disrupts the elastic layer of the arterial intima, and calcifcation of the artery occurs as vascular SMCs and extracellular matrix increase in mass and volume. This is a biomineralization process manifested by the deposition of insoluble calcium in the form of calcium salts in the atherosclerotic plaque, which eventually leads to arterial stifness. This process may be mediated by proteoglycans that have the ability to bind calcium $[60]$ $[60]$. In addition, glycolysis is also involved in the proliferation, migration and calcifcation of SMCs [[61\]](#page-10-20). High rates of glycolysis and low rates of glucose oxidation are observed during atherosclerotic calcifcation. The Warburg efect refers to the switch in energy metabolism from oxidative phosphorylation to glycolysis, and the Warburg efect can induce SMCs proliferation, leading to AS [\[62](#page-10-21)]. Stimulation of SMCs by ox-LDL causes a Warburg efect, resulting in increased glucose uptake and lactate formation. SMCs exhibit higher rates of glucose metabolism and lactate production than normal cells [\[63](#page-10-22)]. There are no studies related to the effect of CoQ10 on IS and AS glycolysis, and CoO10 may have beneficial effects on metabolic reprogramming. Recent studies found that CoQ10 can reduce the expression of glycolysis-related genes, but future studies are still needed to reveal the regulatory mechanism of CoQ10 on metabolic reprogramming [\[64](#page-10-23)].

Over time, neutrophils undergo apoptosis, reducing the infammatory process. However, if macrophages are not able to phagocytose neutrophils in time, then neutrophils become necrotic, which exacerbates infammation. At this point, cells can experience activation and release of neutrophil extracellular traps (NETs), a defense mechanism, but may exacerbate the infammatory response with detrimental efects. NETs are also important in IS and AS pathology, and recent studies have found that NETs are elevated in IS and AS patients and are associated with poor IS outcomes [\[65](#page-10-24), [66\]](#page-10-25). CoQ10 treatment improved neutrophil extracellular traps and diferentiation of pro-infammatory T cell [[67\]](#page-10-26).

The main role of dendritic cells is to initiate antigen-specifc adaptive immune responses, and in the early formation of atherosclerotic lesions, dendritic cells become foam cells through uptake of lipoproteins and lipid-loaded apoptotic cells, but dendritic cells have a poor eferocytosis, which AS produce adverse effects [[51\]](#page-10-10). However, there are no studies on the efect of CoQ10 on dendritic cells.

Calcifcation is important in the development of the pathological process of AS and can determine the outcome of IS. Plaque calcifcation is mainly caused by dysregulated calcium ion deposition and impaired clearance, also known as biomineralization. Rupture-prone atherosclerotic plaques usually have a thin fbrous cap. In contrast, plaques with limited lipid accumulation and a thick fbrous cap are often referred to as stable plaques. The infammatory process reduces collagen in atherosclerotic plaques. And MMPs released by infammatory cells degrade the extracellular matrix, promoting AS plaque progression and rupture [\[68](#page-10-27)]. In addition to ROS, infammatory and immune responses also determine atherosclerotic plaque stability. For example, plaques formed by the M1 phenotype of macrophages are unstable, while the M2 phenotype is stable [\[51](#page-10-10)]. Moreover, the predominant phenotype of symptomatic patients is M1 macrophage, while M2 phenotype is predominant in asymptomatic patients [\[51](#page-10-10)].

There are no clinical studies on the effects of CoQ10 on the stability of atherosclerotic plaques in the middle veins. A clinical study found that CoQ10 was benefcial for vascular stifness. Animal studies found that CoQ10 treatment of atherosclerotic rabbit for 36 weeks has increased stability of atherosclerotic plaque [[69\]](#page-10-28). CoQ10 may also increase stability of atherosclerotic plaque by elevating collagen concentrations [\[70](#page-10-29)].

The regulation of CoQ10 on stability of atherosclerotic plaque may be due to multiple factors, including the reduction of glycolysis, oxidative stress, infammatory and immune response to ROS, release of MMPs, modulation of ox-LDL and HDL levels, reduction of angiogenesis and apoptosis, and increase in collagen levels to maintain the extracellular matrix by CoQ10. These need to be confrmed in future experiments. The schematic diagram of CoQ10 for prevention of AS progression is shown in Fig. [3.](#page-6-0)

The Efect of CoQ10 Against Cell Death

Shortly after IS, the cerebral blood flow reduced, which decreased the access of brain cells to oxygen and glucose, leading to neuronal cell death. CoQ10 reduces neuronal death in IS through regulation of pro-survival and antiapoptotic pathways, including increased expression of Akt, glycogen synthase kinase 3β (GSK3β), B-cell lymphoma-2 (Bcl-2), and forkhead box O3 (FOXO3A), and decreased expression of BCL2-Associated X (Bax), Bcl-2 interacting mediator of cell death (Bim), cysteinyl aspartate specifc proteinase 3 (caspase-3), c-Jun N-terminal kinase (JNK3), C-Jun N-terminal kinase (c-Jun), cytochrome C, caspase-9, and Ca^{2+} content [[20,](#page-9-13) [24](#page-9-16), [27](#page-9-19)]. In addition, CoQ10 protects blood-brain barrier integrity by resisting autophagy and reducing matrix metalloproteinase-2 (MMP-2) and metalloproteinase-9 (MMP-9) expression^{18, 21}. These effects are inextricably linked to the anti-infammatory and anti-oxidative stress mechanisms of CoQ10. CoQ10 improves anxiety and depression-like behavior and cognitive impairment by increasing SOD and BDNF levels [\[25\]](#page-9-17). This efect may involve anti-autophagy, with CoQ10 treatment reducing the Lc3-II/Lc3I ratio.

However, the mechanism of CoQ10 in the treatment of IS needs to be revealed in more studies in the future. Furthermore, the role of CoQ10 as an endogenously produced antioxidant may be even more important for the prevention of IS.

Discussion

Although CoQ10 is often described as a vitamin-like substance, CoQ10 by defnition is not a vitamin as it is produced by various tissues in the body. CoQ10 is mainly produced by HMG Co-A reductase. In addition, CoQ10 can also be formed from Coenzyme Q and hydroxyalanine by the catalysis of polyprenyl-4-hydroxy benzoate transferase. The

Fig. 3 CoQ10 inhibits the pathological progression of AS by regulating multiple immune cells and reducing infammatory responses and cell death. CoQ10 treatment improved neutrophil extracellular traps and pro-infammatory T cell diferentiation. And CoQ10 promotes anti-infammatory M2 phenotype transformation of macrophages, and promotes ABCG1 expression to enhance HDL-mediated cholesterol efflux from macrophages and among others

decaprenyl diphosphate synthase subunit family gene is an important gene in the regulation of CoQ10 [\[71\]](#page-10-30). CoQ10, a key component in the mitochondrial respiratory chain, is involved in the electron transport process where CoQ10 accepts electrons from complexes I and II and transfers them to complex III, thereby driving ATP synthesis. Statin inhibits the 3-hydroxy-3-methylglutaryl coenzyme A reductase, resulting in decreased intracellular concentrations of CoQ10 and cholesterol [[72](#page-10-31)]. This is a potential cause of statin-related muscular symptoms. CoQ10 has many important cellular functions, particularly within the mitochondria. In mitochondria, CoQ10 plays a key role as an electron carrier in the oxidative phosphorylation of the mitochondrial electron transport chain. It is also involved in the metabolism of pyrimidines, fatty acids and mitochondrial uncoupling proteins, as well as in the regulation of mitochondrial permeability. These roles of CoQ10 should not be overlooked [\[73](#page-10-32)]. Moreover, CoQ10 inhibits lipid peroxidation in biological membranes and protects mitochondrial proteins and DNA from oxidative damage [[74](#page-11-0)]. Furthermore, it is the only lipophilic antioxidant that can be synthesized from scratch by cells and has an enzymatic mechanism to regenerate its reduced form. However, endogenous CoQ10 levels decrease with age, along with impaired oxidative phosphorylation and antioxidant capacity. It was found that the optimal production of CoQ10 occurs around age 25, with levels at age 65 being only about 50% of those at age 25 [\[73\]](#page-10-32). Because of this, CoQ10 has become the third most important nutritional supplement after fish oil and multivitamins. But is CoQ10 supplementation necessary? In this section, we will discuss the scope, dosage and safety of CoQ10.

Indications and Dosage of CoQ10

AS and IS

The multiple protective effects of CoQ10 have a positive efect on the development and progression of IS and AS. CoQ10 can be used as an adjunctive agent in the treatment of IS and AS.

Dosage and Safety

The recommended daily dose varies by indication, but is usually around 30 to 100 mg in healthy individuals, and up to 60 to 1200 mg under medical conditions [\[75](#page-11-1)]. It was found that 3600 mg/day of CoQ10 remains safe [\[75](#page-11-1)]. 2400 mg/day of CoQ10 supplementation reached equilibrium with respect to plasma CoQ10 concentrations [\[75](#page-11-1)]. Therefore, 2400 mg/ day may provide the best balance between CoQ10 safety and blood levels [[75\]](#page-11-1). CoQ10 is therefore a safe supplement, but the adverse efects and drug co-actions of CoQ10 need to be discovered in more experiments, except for the co-actions with drugs such as warfarin and antihypertensives.

Limitations and Future Perspectives

Low Bioavailability

CoQ10 has high molecular weight (863.34 g/mol) and poor water solubility. CoQ10 is almost insoluble in water, slightly soluble in ethanol, and soluble in acetone and ether. This limitation leads to its poor oral bioavailability.

Storage Difficulties

CoQ10 is a yellow crystalline powder with a melting point of about 48 \degree C. CoQ10 is unstable and susceptible to the effects of heat, light and oxygen, which limits its use in pharmaceutical and functional food formulations. Moreover, CoQ10 has a low melting point, which makes it difficult to press CoQ10 into tablets because it causes stickiness and adherence to mechanical surfaces when the temperature exceeds its melting point [[73\]](#page-10-32). In addition, CoQ10 should be stored in a cool and dark place, preferably in sealed containers, as it is afected by light, heat and oxidation [[73\]](#page-10-32).

Gradually Increasing Prices

In general, microbial biosynthesis is the preferred and most widespread method for industrial production of CoQ10 [[73\]](#page-10-32). However, microbial bioproduction is not sufficient to meet the current market demand, which leads to higher prices. Tablets are the cheapest processing method, but the physical properties of CoQ10 and the bioavailability limit the use of tablets. As a result, various formulations to improve bioavailability have increased the price of CoQ10 even more.

The Combined Efect with Other Medications

The combination of CoQ10 with other drugs has both advantages and disadvantages. It was found that CoQ10 and the anticoagulant drug fawarin produced adverse reactions, resulting in reduced anticoagulant effect [[76](#page-11-2)]. In addition, CoQ10 and Vitamin K antagonistic anticoagulants increased the risk of bleeding [[77\]](#page-11-3). CoQ10 in combination with cyclosporine can reduce renal toxicity [[78](#page-11-4)]. Also, CoQ10 may reduce the side effects of statinrelated muscular symptoms [\[79\]](#page-11-5).

The synergistic effect of CoQ10 and vitamin E is not clear, and the combination of CoQ10 and vitamin E shows different effects in many diseases, such as metabolic syndrome and alcoholism. In preclinical studies, vitamin E combined with coenzyme Q10 was found to be more effective. However, it still needs to be confirmed in future clinical trials [[19](#page-9-12)].

To address these issues, it is necessary to develop lower $cost$ industrial production systems for $CoQ10$ and more efficient delivery systems for CoQ10. In addition, more experiments are needed to demonstrate the combined efects of CoQ10 and vitamin E in the future.

Conclusion

CoQ10, an antioxidant that offers benefits to multiple body systems, has therapeutic and/or positive effects on IS and risk factors, such as AS, diabetes, hyperlipidemia, hypertension, metabolic syndrome, smoking, drinking, infection, occupational exposure, mental illness, menopause, atrial fibrillation, hyperthyroidism, chronic kidney disease, and moyamoya disease. The above people may be suitable for supplementation with CoQ10. First, CoQ10 is already present in the body and exogenous supplementation of CoQ10 does not interfere with endogenous CoQ10 synthesis. Second, CoQ10 is safe and generally does not react adversely with other drugs. Moreover, CoQ10 is affordable and easy to obtain. However, some efficacy, long-term safety and adverse reactions with other drugs of CoQ10 still need more experimental studies.

Key References

17. Ramezani, M.; Sahraei, Z.; Simani, L.; Heydari, K.; Shahidi, F. Coenzyme Q10 supplementation in acute ischemic stroke: Is it beneficial in short-term administration? *Nutritional Neuroscience* **2020**, *23* (8), 640–645, Article. [https://doi.org/10.1080/1028415x.2018.1541269.](https://doi.org/10.1080/1028415x.2018.1541269)

This is a clinical trial looking at the effect of CoQ10 on acute ischemic stroke. CoQ10 supplementation significantly improved NIHSS and MMSE scores, showing a positive therapeutic effect.

19. Abd-El-Fattah, A. A.; El-Sawalhi, M. M.; Rashed, E. R.; El-Ghazaly, M. A. Possible role of vitamin E, coenzyme Q10 and rutin in protection against cerebral ischemia/reperfusion injury in irradiated rats. *International Journal of Radiation Biology* **2010**, *86* (12), 1070–1078, Article. [https://doi.org/10.3109/09553002.](https://doi.org/10.3109/09553002.2010.501844) [2010.501844](https://doi.org/10.3109/09553002.2010.501844).

This study combined vitamin E with coenzyme Q10 for the treatment of ischemic stroke, and found that coenzyme Q10 plus vitamin E was more effective in the treatment of ischemic stroke

73. Arenas-Jal, M.; Sune-Negre, J. M.; Garcia-Montoya, E. Coenzyme Q10 supplementation: Efficacy, safety, and formulation challenges. *Comprehensive Reviews in Food Science and Food Safety* **2020**, *19* (2), 574–594, Review.<https://doi.org/10.1111/1541-4337.12539>.

This article describes the optimal therapeutic dose and safety of CoQ10.

Author Contributions Zhilei Jia contributed to conceptualization, literature search, manuscript writing, and draft preparation. Xiaoya Yu contributed to literature search and manuscript writing. Xu Wang and Jinhua Li contributed to conceptualization, drafting, guidance, and editing of the manuscript. All authors contributed to the article and approved the submitted version.

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Compliance with Ethical Standards

Competing Interests The authors declare no competing interests.

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References

- 1. Lindsay MP, Norrving B, Sacco RL, Brainin M, Hacke W, Martins S, Pandian J, Feigin V. World Stroke Organization (WSO): Global stroke fact sheet 2019. Int J Stroke. 2019;14(8):806–17.
- 2. Moretti A, Ferrari F, Villa RF. Neuroprotection for ischaemic stroke: Current status and challenges. Pharmacol Ther. 2015;146:23–34.
- 3. Carcel C, Woodward M, Wang X, Bushnell C, Sandset EC. Sex matters in stroke: A review of recent evidence on the differences between women and men. Front Neuroendocrinol. 2020;59:100870.
- 4. Liberale L, Carbone F, Montecucco F, Gebhard C, Luscher TF, Wegener S, Camici GG. Ischemic stroke across sexes: What is the status quo? Front Neuroendocrinol. 2018;50:3–17.
- 5. Macrez R, Ali C, Toutirais O, Le Mauf B, Defer G, Dirnagl U, Vivien D. Stroke and the immune system: From pathophysiology to new therapeutic strategies. Lancet Neurol. 2011;10(5):471–80.
- 6. Okada T, Suzuki H, Travis ZD, Zhang JH. The stroke-induced blood-brain barrier disruption: Current progress of inspection technique, mechanism, and therapeutic target. Curr Neuropharmacol. 2020;18(12):1187–212.
- 7. Zhou Y, Zhang SS, Fan X. Role of polyphenols as antioxidant supplementation in ischemic stroke. Oxid Med Cell Longev. 2021:5471347.
- 8. Heaton RA, Heales S, Rahman K, Sexton DW, Hargreaves I. The effect of cellular coenzyme $Q(10)$ deficiency on lysosomal acidifcation. J Clin Med. 2020;9(6):1923.
- 9. Weber C, Bysted A, Hłlmer G. The coenzyme Q10 content of the average Danish diet International journal for vitamin and nutrition research. Internationale Zeitschrift fur Vitamin- und Ernahrungsforschung. J Int Vitaminologie Nutri. 1997;67(2):123–9.
- 10. Mantle D, Heaton RA, Hargreaves IP. Coenzyme Q10, ageing and the nervous system: An overview. Antioxidants. 2022;11(1):2.
- 11. Storch A. Coenzyme Q(10) in Parkinson's disease. Nervenarzt. 2007;78(12):1378–82.
- 12. Rajdev K, Mehan S. Neuroprotective methodologies of coenzyme Q10 mediated brain hemorrhagic treatment: Clinical and pre-clinical findings. CNS Neurol Disord Drug Targets. 2019;18(6):446–65.
- 13. Kang HC, Lee YM, Kim HD. Mitochondrial disease and epilepsy. Brain Develop. 2013;35(8):757–61.
- 14. Rozen TD, Oshinsky ML, Gebeline CA, Bradley KC, Young WB, Shechter AL, Silberstein SD. Open label trial of coenzyme Q10 as a migraine preventive. Cephalalgia. 2002;22(2):137–41.
- 15. Pradhan N, Singh C, Singh A. Coenzyme Q10 a mitochondrial restorer for various brain disorders. Naunyn-Schmiedebergs Arch Pharmacol. 2021;394(11):2197–222.
- 16. Simani L, Ryan F, Hashemifard S, Hooshmandi E, Madahi M, Sahraei Z, Rezaei O, Heydari K, Ramezani M. Serum coenzyme Q10 is associated with clinical neurological outcomes in acute stroke patients. J Mol Neurosci. 2018;66(1):53–8.
- 17. Ramezani M, Sahraei Z, Simani L, Heydari K, Shahidi F. Coenzyme Q10 supplementation in acute ischemic stroke: Is it beneficial in short-term administration? Nutr Neurosci. 2020;23(8):640–5.
- 18. Lu CJ, Guo YZ, Zhang Y, Yang L, Chang Y, Zhang JW, Jing L, Zhang JZ. Coenzyme Q10 ameliorates cerebral ischemia reperfusion injury in hyperglycemic rats. Pathol Res Pract. 2017;213(9):1191–9.
- 19. Abd-El-Fattah AA, El-Sawalhi MM, Rashed ER, El-Ghazaly MA. Possible role of vitamin E, coenzyme Q10 and rutin in protection against cerebral ischemia/reperfusion injury in irradiated rats. Int J Radiat Biol. 2010;86(12):1070–8.
- 20. Abd El-Aal SA, Abd El-Fattah MA, El-Abhar HS. CoQ10 augments rosuvastatin neuroprotective efect in a model of global ischemia via inhibition of NF-κB/JNK3/Bax and activation of Akt/FOXO3A/Bim Cues. Front Pharmacol. 2017;8:735.
- 21. Ghasemloo E, Oryan S, Bigdeli MR, Mostafavi H, Eskandari M. The neuroprotective efect of MicroRNA-149-5p and coenzymeQ10 by reducing levels of infammatory cytokines and metalloproteinases following focal brain ischemia in rats. Brain Res Bull. 2021;169:205–13.
- 22. Nikolaevna OO, Aronovna GE, Igorevna KE, Alekseevna BM, Vladimirovich GM, Gennadievich MV, Andreevich PY, Stephanovich MO. Intravenous administration of coenzyme Q10 in acute period of cerebral ischemia decreases mortality by reducing brain necrosis and limiting its increase within 4 days in rat stroke model. Antioxidants. 2020;9(12):1240.
- 23. Belousova M, Tokareva OG, Gorodetskaya E, Kalenikova EI, Medvedev OS. Intravenous Treatment With Coenzyme Q10 Improves Neurological Outcome and Reduces Infarct Volume After Transient Focal Brain Ischemia in Rats. J Cardiovasc Pharmacol. 2016;67(2):103–9.
- 24. Nasoohi S, Simani L, Khodagholi F, Nikseresht S, Faizi M, Naderi N. Coenzyme Q10 supplementation improves acute outcomes of stroke in rats pretreated with atorvastatin. Nutr Neurosci. 2019;22(4):264–72.
- 25. Fatemi I, Askari PS, Hakimizadeh E, Kaeidi A, Esmaeil-Moghaddam S, Pak-Hashemi M, Allahtavakoli M. Chronic treatment with coenzyme Q10 mitigates the behavioral dysfunction of global cerebral ischemia/reperfusion injury in rats. Iran J Basic Med Sci. 2022;25(1):39–45.
- 26. Yi YY, Shin HJ, Choi SG, Kang JW, Song HJ, Kim SK, Kim DW. Preventive effects of neuroprotective agents in a neonatal rat of photothrombotic stroke model. Int J Mol Sci. 2020;21(10):3703.
- 27. Park J, Park HH, Choi H, Kim YS, Yu HJ, Lee KY, Lee YJ, Kim SH, Koh SH. Coenzyme Q10 protects neural stem cells against hypoxia by enhancing survival signals. Brain Res. 2012;1478:64–73.
- 28. Kattoor AJ, Pothineni NVK, Palagiri D, Mehta JL. Oxidative stress in atherosclerosis. Curr Atheroscler Rep. 2017;19(11):42.
- 29. Ketelhuth DFJ, Hansson GK. Cellular immunity, low-density lipoprotein and atherosclerosis: Break of tolerance in the artery wall. Thromb Haemost. 2011;106(5):779–86.
- 30. deRijke YB, Bredie SJH, Demacker PNM, Vogelaar JM, Hak-Lemmers HLM, Stalenhoef AFH. The redox status of coenzyme Q10 in total LDL as an indicator of in vivo oxidative modifcation - Studies on subjects with familial combined hyperlipidemia. Arterioscler Thromb Vasc Biol. 1997;17(1):127–33.
- 31. Xie TQ, Wang CY, Jin Y, Meng Q, Liu Q, Wu JJ, Sun HJ. Coenzyme Q10-induced activation of AMPK-YAP-OPA1 pathway alleviates atherosclerosis by improving mitochondrial function, inhibiting oxidative stress and promoting energy metabolism. Front Pharmacol. 2020;11:1034.
- 32. Wang DL, Yan X, Xia M, Yang Y, Li D, Li XR, Song FL, Ling WH. Coenzyme Q10 promotes macrophage cholesterol efflux by regulation of the activator protein-1/miR-378/ATP-binding cassette transporter g1-signaling pathway. Arterioscler Thromb Vasc Biol. 2014;34(9):1860–70.
- 33. Salonen I, Huttunen K, Hirvonen MR, Dufva J, Groundstroem K, Dufva H, Pekkanen J, Salonen RO. Serum myeloperoxidase is independent of the risk factors of atherosclerosis. Coron Artery Dis. 2012;23(4):251–8.
- 34. Robb SJ, Connor JR. Nitric oxide protects astrocytes from oxidative stress. Ann N Y Acad Sci. 2002;962:93–102.
- 35. Fouad GI. Synergistic anti-atherosclerotic role of combined treatment of omega-3 and co-enzyme Q10 in hypercholesterolemia-induced obese rats. Heliyon. 2020;6(4):e03659.
- 36. Tsai KL, Huang YH, Kao CL, Yang DM, Lee HC, Chou HY, Chen YC, Chiou GY, Chen LH, Yang YP, Chiu TH, Tsai CS, Ou HC, Chiou SH. A novel mechanism of coenzyme Q10 protects against human endothelial cells from oxidative stress-induced injury by modulating NO-related pathways. J Nutr Biochem. 2012;23(5):458–68.
- 37. Hansen M, Kuhlman ACB, Sahl RE, Kelly B, Morville T, Dohlmann TL, Chrois KM, Larsen S, Helge JW, Dela F. Infammatory biomarkers in patients in Simvastatin treatment: No efect of co-enzyme Q10 supplementation. Cytokine. 2019;113:393–9.
- 38. Chapidze GE, Kapanadze SD, Dolidze NK, Latsabidze NE, Bakhutashvili ZV. Combination treatment with coenzyme Q10 and simvastatin in patients with coronary atherosclerosis. Kardiologiya. 2006;46(8):11–3.
- 39. Ahmadvand H, Dehnoo MG, Dehghani A, Bagheri S, Cheraghi RA. Serum paraoxonase 1 status and its association with atherogenic indexes in gentamicin-induced nephrotoxicity in rats treated with coenzyme Q10. Ren Fail. 2014;36(3):413–8.
- 40. Turunen M, Wehlin L, Sjoberg M, Lundahl J, Dallner G, Brismar K, Sindelar PJ. beta 2-integrin and lipid modifcations indicate a non-antioxidant mechanism for the anti-atherogenic efect of dietary coenzyme Q10. Biochem Biophys Res Commun. 2002;296(2):255–60.
- 41. Frontinan-Rubio J, Santiago-Mora RM, Nieva-Velasco CM, Ferrin G, Martinez-Gonzalez A, Gomez MV, Moreno M, Ariza J, Lozano E, Arjona-Gutierrez J, Gil-Agudo A, De la Mata M, Pesic M, Peinado JR, Villalba JM, Perez-Romasanta L, Perez-Garcia VM, Alcain FJ, Duran-Prado M. Regulation of the oxidative balance with coenzyme Q10 sensitizes human glioblastoma cells to radiation and temozolomide. Radiother Oncol. 2018;128(2):236–44.
- 42. Fiorentino TV, Prioletta A, Zuo P, Folli F. Hyperglycemia-induced oxidative stress and its role in diabetes mellitus related cardiovascular diseases. Curr Pharm Des. 2013;19(32):5695–703.
- 43. Espey MG, Miranda KM, Feelisch M, Fukuto J, Grisham MB, Vitek MP, Wink DA. Mechanisms of cell death governed by the balance between nitrosative and oxidative stress. Ann N Y Acad Sci. 2000;899:209–21.
- 44. Wang FL, Yuan QH, Chen FY, Pang JJ, Pan C, Xu F, Chen YG. Fundamental mechanisms of the cell death caused by nitrosative stress. Front Cell Dev Biol. 2021;9:742483.
- 45. Libby P, Buring JE, Badimon L, Hansson CK, Deanfeld J, Bittencourt MS, Tokgozoglu L, Lewis EF. Atherosclerosis Nat Rev Dis Primers. 2019;5(1):56.
- 46. Pacher P, Beckman JS, Liaudet L. Nitric oxide and peroxynitrite in health and disease. Physiol Rev. 2007;87(1):315–424.
- 47. Li G, Zou LY, Cao CM, Yang ES. Coenzyme Q10 protects SHSY5Y neuronal cells from beta amyloid toxicity and oxygenglucose deprivation by inhibiting the opening of the mitochondrial permeability transition pore. BioFactors. 2005;25(1–4):97–107.
- 48. Hansson GK, Hermansson A. The immune system in atherosclerosis. Nat Immunol. 2011;12(3):204–12.
- 49. Ya FL, Xu XR, Tian ZZ, Gallant RC, Song FL, Shi YL, Wu YF, Wan JB, Zhao YM, Adili R, Ling WH, Ni HY, Yang Y. Coenzyme Q10 attenuates platelet integrin alpha IIb beta 3 signaling and platelet hyper-reactivity in ApoE-defcient mice. Food Funct. 2020;11(1):139–52.
- 50. Alehagen U, Lindahl TL, Aaseth J, Svensson E, Johansson P. Levels of sP-selectin and hs-CRP decrease with dietary intervention with selenium and coenzyme Q10 combined: A secondary analysis of a randomized clinical trial. Plos One. 2015;10(9):e0137680.
- 51. Mehu M, Narasimhulu CA, Singla DK. Infammatory cells in atherosclerosis Antioxidants. 2022;11(2):233.
- 52. Chistiakov DA, Melnichenko AA, Myasoedova VA, Grechko AV, Orekhov AN. Mechanisms of foam cell formation in atherosclerosis. J Mol Med. 2017;95(11):1153–65.
- 53. Yan X, Shen TR, Jiang XW, Tang XL, Wang DL, Li HH, Ling WH. Coenzyme Q10 consumption promotes ABCG1-mediated macrophage cholesterol efflux: A randomized, double-blind, placebo-controlled, cross-over study in healthy volunteers. Mol Nutr Food Res. 2015;59(9):1725–34.
- 54. Peace CG, O'Neill LAJ. The role of itaconate in host defense and infammation. J Clin Investig. 2022;132(2):e148548.
- 55. Li LF, Wang M, Ma QX, Ye JX, Sun GB. Role of glycolysis in the development of atherosclerosis. Am J Physiol Cell Physiol. 2022;323(2):C617–29.
- 56. Shirai T, Nazarewicz RR, Wallis BB, Yanes RE, Watanabe R, Hilhorst M, Tian L, Harrison DG, Giacomini JC, Assimes TL, Goronzy JJ, Weyand CM. The glycolytic enzyme PKM2 bridges metabolic and infammatory dysfunction in coronary artery disease. J Exp Med. 2016;213(3):337–54.
- 57. Viola A, Munari F, Sanchez-Rodriguez R, Scolaro T, Castegna A. The metabolic signature of macrophage responses. Front Immunol. 2019;10:1462.
- 58. Oatley M, Bolukbasi OV, Svensson V, Shvartsman M, Ganter K, Zirngibl K, Pavlovich PV, Milchevskaya V, Foteva V, Natarajan KN, Baying B, Benes V, Patil KR, Teichmann SA, Lancrin

C. Single-cell transcriptomics identifes CD44 as a marker and regulator of endothelial to haematopoietic transition. Nat Commun. 2020;11(1):586.

- 59. Di Lorenzo A, Iannuzzo G, Parlato A, Cuomo G, Testa C, Coppola M, D'Ambrosio G, Oliviero DA, Sarullo S, Vitale G, Nugara C, Sarullo FM, Giallauria F. Clinical evidence for Q10 coenzyme supplementation in heart failure: From energetics to functional improvement. J Clin Med. 2020;9(5):1266.
- 60. Narasimhulu CA, Fernandez-Ruiz I, Selvarajan K, Jiang XT, Sengupta B, Riad A, Parthasarathy S. Atherosclerosis - do we know enough already to prevent it? Curr Opin Pharmacol. 2016;27:92–102.
- 61. Perez J, Hill BG, Benavides GA, Dranka BP, Darley-Usmar VM. Role of cellular bioenergetics in smooth muscle cell proliferation induced by platelet-derived growth factor. Biochem J. 2010;428:255–67.
- 62. Werle M, Jahn L, Kreuzer J, Hofele J, Elsasser A, Ackermann C, Katus HA, Vogt AM. Metabolic control analysis of the Warburg-efect in proliferating vascular smooth muscle cells. J Biomed Sci. 2005;12(5):827–34.
- 63. Shi J, Yang Y, Cheng AY, Xu G, He F. Metabolism of vascular smooth muscle cells in vascular diseases. Am J Physiol Heart Circ Physiol. 2020;319(3):H613–31.
- 64. Henry ML, Velez-Irizarry D, Pagan JD, Sordillo L, Gandy J, Valberg SJ. The impact of N-Acetyl cysteine and coenzyme Q10 supplementation on skeletal muscle antioxidants and proteome in ft thoroughbred horses. Antioxidants. 2021;10(11):1739.
- 65. Denorme F, Portier I, Rustad JL, Cody MJ, de Araujo CV, Hoki C, Alexander MD, Grandhi R, Dyer MR, Neal MD, Majersik JJ, Yost CC, Campbell RA. Neutrophil extracellular traps regulate ischemic stroke brain injury. J Clin Investig. 2022;132(10):e154225.
- 66. Kim JK, Hong CW, Park MJ, Song YR, Kim HJ, Kim SG. Increased neutrophil extracellular trap formation in uremia is associated with chronic infammation and prevalent coronary artery disease. J Immunol Res. 2017;2017:8415179.
- 67. Perez-Sanchez C, Aguirre MA, Ruiz-Limon P, Abalos-Aguilera MC, Jimenez-Gomez Y, Arias-de la Rosa I, Rodriguez-Ariza A, Fernandez-del Rio L, Gonzalez-Reyes JA, Segui P, Collantes-Estevez E, Barbarroja N, Velasco F, Sciascia S, Cecchi I, Cuadrado MJ, Villalba JM, Lopez-Pedrera C. Ubiquinol efects on antiphospholipid syndrome prothrombotic profle a randomized, placebo-controlled trial. Arterioscler Thromb Vasc Biol. 2017;37(10):1923.
- 68. Senoner T, Dichtl W. Oxidative stress in cardiovascular diseases: Still a therapeutic target? Nutrients. 2019;11(9):2090.
- 69. Larijani VN, Ahrnadi N, Zeb I, Khan F, Flores F, Budoff M. Benefcial efects of aged garlic extract and coenzyme Q10 on vascular elasticity and endothelial function: The FAITH randomized clinical trial. Nutrition. 2013;29(1):71–5.
- 70. Shiomi M, Yamada S, Amano Y, Nishimoto T, Ito T. Lapaquistat acetate, a squalene synthase inhibitor, changes macrophage/lipid-rich coronary plaques of hypercholesterolaemic rabbits into fibrous lesions. Br J Pharmacol. 2008;154(5):949–57.
- 71. Yu TJ, Liu YY, Li XG, Lian B, Lu XX, Jin X, Shao ZM, Hu X, Di GH, Jiang YZ. PDSS1-Mediated activation of CAMK2A-STAT3 signaling promotes metastasis in triple-negative breast cancer. Can Res. 2021;81(21):5491–505.
- 72. Mosshammer D, Schaefeler E, Schwab M, Mörike K. Mechanisms and assessment of statin-related muscular adverse efects. Br J Clin Pharmacol. 2014;78(3):454–66.
- 73. Hargreaves I, Heaton RA, Mantle D. Disorders of human coenzyme Q10 metabolism: An overview. Int J Mol Sci. 2020;21(18):6695.
- 74. Kim JM, Park E. Coenzyme Q10 attenuated dmh-induced precancerous lesions in sd rats. J Nutr Sci Vitaminol. 2010;56(2):139–44.
- 75. Arenas-Jal M, Sune-Negre JM, Garcia-Montoya E. Coenzyme Q10 supplementation: Efficacy, safety, and formulation challenges. Compr Rev Food Sci Food Saf. 2020;19(2):574–94.
- 76. Zhou QY, Zhou SF, Chan E. Effect of coenzyme Q<sub>10</ sub> on warfarin hydroxylation in rat and human liver microsomes. Curr Drug Metab. 2005;6(2):67–81.
- 77. Chu SD, Tran MT. Anticoagulation overdose when receiving vitamin k antagonists combined with coenzyme Q10: Review of the world's medicine. Arch Pharm Pract. 2023;14(3):62–4.
- 78. Sato T, Ishikawa A, Homma Y. Efect of reduced form of coenzyme Q10 on cyclosporine nephrotoxicity. Exp Clin Transplant. 2013;11(1):17–20.

79. Skarlovnik A, Janic M, Lunder M, Turk M, Sabovic M. Coenzyme Q10 supplementation decreases statin-related mild-to-moderate muscle symptoms: A randomized clinical study. Med Sci Monit. 2014;20:2183–8.

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