PEDIATRIC FUNGAL INFECTIONS (E ROILIDES, SECTION EDITOR)



# Antifungal Prophylaxis in Children Receiving Antineoplastic Chemotherapy

Elio Castagnola<sup>1</sup> · Alessio Mesini<sup>2</sup>

Published online: 2 March 2018 © Springer Science+Business Media, LLC, part of Springer Nature 2018

#### Abstract

**Purpose of the review** The purpose of this study was to summarize data on available antifungal prophylaxis of invasive fungal disease (IFD) in children and when it should be administered during antineoplastic chemotherapy.

**Recent findings** Antifungal prophylaxis should be considered when incidence of IFD is  $\geq 10\%$ , as acute myeloblastic leukemia, high-risk acute lymphoblastic leukemia, and second-line therapy for any relapsing leukemia. In absence of specific pediatric studies, data from adults indicate that triazoles, especially posaconazole tablets, could represent the most attractive option, even if some troubles (mainly regarding drug interactions and intestinal absorption) must be underlined. Echinocandins and liposomal amphotericin B (intravenous or nebulized) can represent alternatives in specific conditions. Other infection control measures (hand hygiene, respiratory masks) can represent adjunctive and effective measures.

**Summary** Antifungal prophylaxis should be implemented in children receiving aggressive chemotherapy for acute leukemia, and triazoles represent the first choice for this purpose.

Keywords Antifungal prophylaxis · Children · Cancer · Acute leukemia · Invasive fungal disease

# Introduction

Invasive fungal diseases (IFD) are a major cause of morbidity and mortality in children with cancer [1]. *Candida* spp. and *Aspergillus* spp. are the most frequently identified pathogens [2, 3], but other fungi, e.g., *Fusarium* spp., *Geotrichum* spp., and also *Pneumocystis jirovecii*, can cause severe infections even if with a lower frequency [4, 5].

The epidemiology of IFD is strictly related with the aggressiveness of antineoplastic chemotherapy, being the highest in children receiving first-line treatment for acute myeloblastic leukemia and high-risk acute lymphoblastic leukemia (depending on protocol and risk profile), second-line therapy

This article is part of the Topical Collection on *Pediatric Fungal* Infections

Elio Castagnola eliocastagnola@gaslini.org for any relapsing leukemia, or an allogeneic hemopoietic stem transplant (HSCT) from an alternative donor or in presence of severe acute or chronic graft vs. host disease (GvHD) [1, 6-11]. In spite of the progresses in diagnosis and treatment, mortality is still very high especially in children at highest risk [12]. Therefore, the development of strategies aimed to prevent these infections could be of pivotal importance. Setting up any prophylactic strategy, including against IFD, has to take in account different factors as follows [13]: (i) frequency of the disease in any given patients' population and any single center [1, 6, 9, 14–19], (ii) number of patients that must receive prophylaxis to prevent one single event (number needed to treat—NNT) [20], and (iii) availability of drugs suitable for prophylactic purposes (with adequate spectrum of activity, easy to be administered, effective, associated with few or no adverse event, without any interaction). Availability of pharmacological data (pharmacokinetic/pharmacodynamic, PK/ PD), registration for pediatric use, and formulations suitable for administration in children (especially the youngest) are other important conditions not always available [21-23].

In 2014, the 4th European Conference on Infections in Leukemia (ECIL-4) published a comprehensive guideline for the management of IFD in children with cancer [22]. The aim of the present review is to integrate and update these

<sup>&</sup>lt;sup>1</sup> Infectious Diseases Unit, Istituto Giannina Gaslini, Via G.Gaslini, 5, 16147 Genoa, Italy

<sup>&</sup>lt;sup>2</sup> Infectious Diseases Unit, DISSal, University of Genoa, Ospedale Policlinico San Martino–IRCCS per l'Oncologia, Genoa, Italy

recommendations focusing on children receiving antineoplastic chemotherapy or autologous HSCT (also called megatherapy with autologous stem cell rescue).

# **Population at Risk**

Besides the well-known role of prolonged (defined as longer than 10 days) [8, 24, 25] and profound neutropenia (absolute neutrophil lower than 100 cells/µL) [8] in IFD development [26], other risk factors as mucositis, steroid treatment [8], highly intensive antineoplastic protocols [10, 26], and presence of central venous catheter [27] have been classically associated with an increasing incidence of IFD. More recently, other patient-related, genetic factors, like toll-like receptors and dectin-1 polymorphisms [28-31], have been identified as possible further risk factors for the development of invasive mycoses in cancer or allogeneic HSCT patients. The analysis of the epidemiology of IFD observed in different groups of patients suggested a stratification of the risk of IFD in children as high if incidence is  $\geq 10\%$ , low if < 5%, or sporadic [22]. For patients undergoing antineoplastic chemotherapy or autologous HSCT, this stratification is confirmed in the everyday clinical practice where the highest incidence of IFD is observed in acute myeloblastic leukemia or any relapsing leukemia [1, 9–11, 14, 18, 32, 33], while IFD are infrequent in solid tumors [19, 34] or after autologous HSCT [16, 19]. Noteworthy, mortality is still high, especially in infections due to molds or in high-risk patients [1, 8, 12]. All these data should be kept in mind when deciding to start a prophylactic strategy, never forgetting than also local epidemiology can play an important role [15].

## **Primary Antifungal Prophylaxis**

Several compounds are available for antifungal prophylaxis, and their effectiveness has been demonstrated in many adults' studies. Unfortunately, adequate clinical trials are lacking in pediatrics as well as approval by Regulatory Agencies of antifungal drugs for this indication in children. Therefore, the use of antifungal prophylaxis in pediatrics is a procedure derived from adults and frequently off-label for indication and/or drug administered [22, 23]. Anyway, at present, pediatric PK/PD data are available for many drugs and therefore, in the case of implementation of an antifungal prophylaxis program administration of antifungal drugs in children, can be considered feasible and safe, especially when therapeutic drug monitoring (TDM) is available to supervise effectiveness and toxicity [22, 35, 36].

In the last few years, several guidelines have tried to establish a correct pharmacological approach to antifungal prophylaxis in pediatric population with hematological or oncological diseases [21, 22, 37], and Table 1 summarizes the presently available drugs and dosages recommended for prophylaxis of IFD in cancer children.

Triazoles represent the principal drug class involved in prophylaxis, but all available options have some issues. Fluconazole is approved for all ages, but it is active only against yeasts (mainly Candida spp. and Cryptococcus spp), and not against molds. Fluconazole prophylaxis determined a reduction in IFD development in different studies, but no one in children of age less than 12 years [23, 42, 43]. Itraconazole is active against both molds and yeasts, and it is superior to fluconazole in preventing IFD in patients undergoing allogeneic HSCT [44], but it is not approved for patient aged less than 18 years [22]. Moreover, administration of oral solution is necessary to obtain effective plasma concentrations [22], compound that is often discontinued because of side effects [23]. Voriconazole is active against yeasts and molds, and its prophylactic use in children has been described as well tolerated [45], with improved survival in children with acute myeloid leukemia compared with historical controls, even if associated with changing of epidemiology versus more rare fungi [46]. But voriconazole is not approved for use in patients < 2 years of age. Posaconazole is active against molds and yeasts and has been initially demonstrated effective in preventing IFD in adults receiving chemotherapy for myelodisplastic syndrome [47]. Even if not approved in patients < 18 years of age, it has been administered for prevention of IFD in pediatrics [38, 48, 49]. Posaconazole was first commercialized as oral suspension, but with this, compound treatment was complicated by variable absorption, problem potentially solved by fatty meal and/or other "bundle" measures [50, 51]. Moreover, the use of posaconazole oral solution in patients younger than 13 years has been frequently associated with side effects as rash, abdominal pain, nausea, and vomitus [52]. Tablets containing 100 mg of posaconazole have been approved in 2013 and they show no absorption issue. Furthermore, pediatrics PK/PD data show that the use of this formulation determines effective concentrations also in children [38]. However, tablets are slightly less than 2 cm long and "should be swallowed whole with water and should not be crushed, chewed, or broken" as reported in the "Product Information Document" [53]. Therefore, this approach could not be easy to be applied in children receiving aggressive chemotherapy (mainly for acute myelogenous leukemia or relapsing acute leukemia) because of the presence of mucositis with obvious difficulties in swallowing. Furthermore, this could represent a problem when "a fraction" of a tablet (e.g., 50 mg, 0.5 tablet) is prescribed [38] (Table 1). In this case, a possible trickery could be represented by alternation of the number of tablets (i.e., one a day and two the next day, e.g., 100 mg alternated to 200 mg, in order to obtain a "mean" dose of 150 mg over 2 days) coupled with TDM. When tablets cannot be swallowed but oral therapy is feasible, recent data

Risk level	Condition	Prophylaxis <sup>a</sup>		
High (≥ 10%)	Acute myeloid leukemia High-risk acute lymphoblastic leukemia Relapsing acute leukemia	Fluconazole: poor evidence to support a recommendation because of the narrow spectrum (yeasts only); dosage: 6 mg/kg per day (maximum 400 mg per day) intravenously or orally q24h Itraconazole: moderate evidence to support a recommendation for use; dosage: oral solution 2.5 mg/kg per day orally (in children aged ≥2 years) q12h, with empty stomach. TDM is recommended Posaconazole: <u>oral suspension</u> dosage: 120 mg/m <sup>2</sup> q8h for children who can not swallow tablets. For this formulation a "posaconazole bundle" can be necessary (see Table 2); <u>tablets</u> dosage: loading dose of 300 mg q12h (1st day) then maintenance 300 q24h, independently from meal. A dosage for younger children using tables has been suggested by means of pharmacokinetic studies:		
		Body weight	Loading dose (1st day)	Maintenance dose
		>15–21 kg	100 mg q12h	100 mg q24h
		22–30 kg	150 mg q12h	150 mg q24h
		31–35 kg	200 mg q12h	200 mg q24h
		35–40 kg	250 mg q12h	250 mg q24h
		>40 kg or 13 years old	300 mg q12h	300 mg q24h
		<ul> <li>TDM is recommended</li> <li>Voriconazole: no evidence to support recommendation; dosage: children aged 2-&lt;12 years or 12-14 years and weighing &lt; 50 kg 9 mg/kg q12h orally; children aged ≥15 years or 12-14 years and weighing ≥50 kg: 4 mg/kg q12h (1st day, 6 mg/kg)</li> <li>TDM is recommended</li> <li>Liposomal amphotericin B: intravenous, moderate evidence to support a recommendation for use; dosage 1 mg/kg q24h every other day or 2.5 mg/kg q24h twice weekly; nebulized: no grading, 25 mg q12h on 2 consecutive days per week</li> <li>Micafungin: no grading; dosage: 1 mg/kg (in children weighing ≥50 kg, 50 mg) q24h intravenously</li> </ul>		
Low (≤5%) Sporadic	Acute lymphoblastic leukemia Non-Hodgkin's lymphoma Autologous HSCT Solid tumors	Not recommended		
Sporadic	Brain tumors Hodgkin's lymphoma	Not recommended		

Table 1 Primary antifungal prophylaxis in children according to risk levels, evidences, and pharmacological data [22, 38-40]

<sup>a</sup> Prophylaxis for *P. jirovecii* pneumonia should be always considered in all patients' populations

Legend: qxh = every "x" hours; TDM = therapeutic drug monitoring

showed that a dosage of the suspension based on the body surface area allowed to reach effective concentrations too [39]. At present, TDM should be performed for all triazoles (with the possible exclusion of fluconazole) [22, 36, 54] in cancer children receiving antifungal prophylaxis in order to control effectiveness and side effects. Furthermore, TDM with maintenance of adequate plasma concentrations has been associated not only with reduction of adverse events but also in preventing the onset of resistance both in molds and in yeasts [35]. TDM should be considered mandatory for voriconazole in the youngest patients [41, 55–57] and in presence of laboratory signs of inflammation [58–60] or treatment with dexamethasone [61]. For posaconazole, TDM should be considered mandatory if tablets are used in the youngest patients. When changes in dosages are needed because of toxicity or sub-therapeutic plasma levels, specific strategies should be implemented [36, 41] in order to obtain the desired concentrations. Table 2 summarizes target plasma concentrations (trough,  $C_{min}$ ) and gives some clues for dosages changes and TDM. Triazoles have many interactions with other drugs [54, 62], with the risk of absence of therapeutic effect or adverse events for all the compounds interacting. Sometimes, these interactions signify absolute contraindications for the concomitant use of triazoles with some antineoplastic drugs (e.g., posaconazole and vinca alkaloids that represent a cornerstone for the treatment of acute lymphoblastic leukemia in children). The possibility of drug interactions must always be evaluated any time triazoles are considered for antifungal prophylaxis. Also, in this case, TDM could be very helpful.

Drug	Target trough ( $C_{min}$ ) concentration and strategies for monitoring	Strategies for dosage modifications in case of low serum concentrations	Strategies for dosage modifications in case of high serum concentrations: empiric changes before plasma concentration available
itraconazole	<ul> <li>target concentration for prophylaxis</li> <li>0.5 mg/L at steady state</li> <li>Measure serum concentrations</li> <li>5–7 days after initiation of therapy</li> <li>following any dose adjustment,</li> <li>when interacting drugs start or stop</li> <li>in case of uncertain compliance with oral therapy</li> <li>in presence of concerns about gastrointestinal absorption</li> <li>in case of potential clinical or laboratory manifestations of toxicity</li> </ul>	<ul> <li>Increase total daily dose (e.g., from 200 mg q12h to 300 mg q12h) by 50% and/or</li> <li>use oral solution (if not already administered)</li> <li>check if the drug is given in the fasting state</li> <li>check compliance</li> <li>s check for top interacting drugs</li> </ul>	_
voriconazole	<ul> <li>target concentration for prophylaxis &gt; 1 and &lt; 6 mg/L at steady state</li> <li>Measure serum concentrations</li> <li>before the 5th dose (2 days of treatment),</li> <li>before the 5th dose following any dose adjustment</li> <li>routine every 1–2 weeks after achievement of steady-state</li> <li>when interacting drugs start or stop</li> <li>in case of uncertain compliance for oral therapy</li> <li>in case of concerns about gastrointestinal absorption, especially for prolonged periods of time</li> <li>in case of potential clinical or laboratory</li> </ul>	<ul> <li>Increase total daily dose by 50% (e.g., from 200 mg q12h to 300 mg q12h), and/or</li> <li>use a pro/kg dose instead of a fixed one</li> <li>for patients &lt; 2 years increase dose by 1 mg/kg/day for every 0.5 mg/l rise in trough concentration desired, divided in 3 equal doses</li> <li>check if the drug is given in the fasting state</li> <li>check compliance</li> <li>check for and stop interacting drugs</li> </ul>	<ul> <li>Withdraw 1 dose, then decrease daily dose by 25%</li> <li>In case of hepathotoxicity</li> <li>mild-moderate hepatic dysfunction (Child-Pugh score 5–9): start with normal loading dose, reduce maintenance dose by 50%</li> <li>severe hepatic dysfunction (Child-Pugh score 10–15) do not administer</li> </ul>
posaconazole	<ul> <li>manifestations of toxicity</li> <li>target concentration for prophylaxis</li> <li>0.7 mg/L at steady state</li> <li>Measure serum concentrations</li> <li>7 days after initiation of therapy</li> <li>following dose adjustment</li> <li>when interacting drugs start or stop</li> <li>in case of uncertain compliance</li> <li>in case of concerns about gastrointestinal absorption, especially for prolonged periods of time</li> <li>in case of potential clinical or laboratory manifestations of toxicity</li> </ul>	<ul> <li>Remove acid suppression if possible (i.e., stop or reduce H<sub>2</sub> antagonists or proton pump inhibitors</li> <li>check compliance</li> <li>check for stop interacting drugs</li> <li>For oral solution use check adherence to "posaconazole bundle"</li> <li>ascorbic acid 500 mg per os with each dose of posaconazole.</li> <li>120–180 ml of carbonated soda beverage (i.e., cola or ginger ale) or acidic fruit juice (e.g., cranberry or orange juice) with each dose of posaconazole.</li> <li>heavy snack or food with each dose, preferably high-fat, including commercially available nutritional supplements.</li> </ul>	

Table 2Triazole plasma trough ( $C_{min}$ ) concentrations effective for antifungal prophylaxis and strategies to modify dosages in case of low or highserum concentrations. Modified from [36, 41]

Echinocandins have been less frequently used as antifungal prophylaxis, but they could represent a possible option in order to avoid drug-drug interactions or when oral route is not feasible [22, 23]. Micafungin has an indication for prevention of invasive candidiasis during prolonged neutropenia preceding engraftment in allogeneic HSCT [63]. Caspofungin is not registered for prophylactic use, but it has been demonstrated to be not inferior to liposomal amphotericin B for this indication in allogeneic HSCT patients [64]. No data are available for anidulafungin prophylaxis in children receiving chemotherapy or autologous HSCT. Liposomal amphotericin B (L-AmB) can be another option in children who cannot receive prophylaxis with azoles or echinocandins, but its use could be limited by lack of registration for this indication and clinical trials with adequate power, beyond some severe side effects. A retrospective study demonstrated effectiveness of intravenous L-AmB administered two times/week compared with historical controls in children with hematologic malignancies [65], and nebulized L-AMB has been administered for IFD prophylaxis in patients with prolonged neutropenia following chemotherapy or allogeneic HSCT [66]. In this last case, efficacy is, obviously, restricted to prevention of pulmonary IFD, and therefore, it should be associated with a drug with systemic effect [22]. Both these class of drugs have no oral formulation forcing the patient to alternative hospitalization (e.g., day-hospital, home care) to receive intravenous prophylaxis.

A last annotation regards correct timing for starting antifungal prophylaxis. This aspect is not well defined, but generally, it is suggested to start together with chemotherapy, or at its end if interacting drugs are used, and to stop it after the resolution of the risk period [21, 22].

#### Pneumocystis jirovecii Pneumonia

*Pneumocystis jirovecii* pneumonia (PCP) is a peculiar, severe, life-threatening fungal infection in immunocompromised hosts, both children and adults. The 5th European Conference on Infections in Leukaemia (ECIL-5) provided indications for its prophylaxis in all ages. Trimethoprim/ sulfamethoxazole prophylaxis is highly recommended in children affected with acute lymphoblastic leukemia, from induction to end of maintenance, and it is considered optional in acute myelogenous leukemia or solid tumors for all the duration of chemotherapy [67], but should be considered also after autologous HSCT [68]. Prophylaxis is highly effective, and in case of documented failure, especially in adolescents, compliance must be checked [5, 69].

## Secondary Antifungal Prophylaxis

Patients with a history of invasive mycosis are at high risk of reactivation when undergoing further aggressive chemotherapy or HSCT (mainly allogeneic). Therefore, secondary antifungal prophylaxis is recommended during aggressive treatments for patients with previous IFD, especially in presence of deep organ localizations. The drug for secondary prophylaxis should be chosen according to the etiology of the primary infection, the localization, the drugs available and their formulations, and the risks of interactions with other therapies, especially those for the treatment of the underlying disease.

# **Other Prophylactic Measures**

Administration of granulocyte (G-CSF) or granulocyte macrophage-colony stimulating factor (GM-CSF) has been advocated as a possible prophylactic measure for

IFD by reducing duration of neutropenia, but wide studies on their use in pediatrics did not show any effectiveness on mortality [70, 71].

*Candida* spp. colonize the intestinal tract and the skin, including health workers hands [72–74] that can become vectors for *Candida* colonization and for infections, e.g., by vascular access manipulation. Therefore, correct hands hygiene procedures are an essential practice to reduce also the risk of IFD [75, 76]. Products for hand hygiene containing chlorhexidine gluconate or isopropyl alcohol are more effective in yeast reduction compared to water and soap (4 vs. 50%) [76]. In spite of the low frequency of vascular access-related fungal infections [77, 78], a correct manipulation of these devices performed both in the hospital and at home represents a further aspect for the prevention of IFD [77, 79].

Aspergillus spp. colonization is a necessary condition to develop disease during immunosuppression, and conidia inhalation represents the classical way for acquisition of this pathogen [80]. Inhalation can occur outside the hospital or during hospital admissions, especially during building reconstruction [81, 82]. Use of high-efficiency particulate air (HEPA) filtration, better if associated with positive pressure isolation rather than laminar airflow, could reduce invasive diseases [81], but regular systems inspections and maintenance must be performed [81, 83-86]. Face masks could be utilized too, especially when patients are outside from a hospital room with HEPA filtration or near to building work areas [85], but the use of these devices could be difficult especially in younger children [87]. Aspergillus spp. and other filamentous fungi have been found also in hospital water systems [88], which therefore should be periodically controlled. However, there is growing evidence that in many patients, colonization is present before hospitalization, since many of these opportunistic pathogens are present in potted plants, flower arrangements, carpet, and home water supplies [80]. All these conditions should be monitored, and possibly avoided or amended both in the hospital and at patient's home, for an effective program of IFD prophylaxis. These strategies have the advantage of absence of adverse events (if any) and drug interactions, and effectiveness also against other, nonfungal pathogens.

#### Conclusion

In conclusion, IFD have a high incidence in children receiving aggressive antineoplastic chemotherapy for acute leukemia, but not for solid tumors or undergoing autologous HSCT, even if local epidemiological peculiarities can be present. Mortality is high especially in cases due to molds and in patients at the highest risk. No clinical trial on prophylaxis of IFD with adequate design and power has been performed in children, and many of the pediatric indications are derived

from adults or inferred from pediatric observational studies. In spite of the absence of formal registration for antifungal prophylaxis and/or for the pediatric age, triazoles represent the easiest to use and actually administered drugs for antifungal prophylaxis in children. In any case, the availability of oral formulations and of specific PK/PD pediatric data allows safe and (realistically) effective choices. However, TDM should be considered mandatory for monitoring their effectiveness and safety both in terms of adverse events and resistance selection. Other drugs like echinocandins or L-AmB can be safe and effective, especially in presence of important drug interaction or difficulties in assuming oral drugs, but the lack of an oral formulation reduces their utilization. Finally, nonpharmacological measures as hand hygiene and use of HEPA filters might represent effective and "safe" prophylactic tools not only for the prevention of IFD.

#### **Compliance with Ethical Standards**

**Conflict of Interest** Elio Castagnola has received personal fees from Astellas Pharma and Basilea Pharmaceutica. Alessio Mesini declares no conflict of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

# References

- Cesaro S, Tridello G, Castagnola E, Calore E, Carraro F, Mariotti I, et al. Retrospective study on the incidence and outcome of proven and probable invasive fungal infections in high-risk pediatric oncohematological patients. Eur J Haematol. 2017;99:240–8.
- Steinbach WJ, Roilides E, Berman D, Hoffman JA, Groll AH, Bin-Hussain I, et al. Results from a prospective, international, epidemiologic study of invasive candidiasis in children and neonates. Pediatr Infect Dis J. 2012;31:1252–7.
- Wattier RL, Dvorak CC, Hoffman JA, Brozovich AA, Bin-Hussain I, Groll AH, et al. A prospective, international cohort study of invasive mold infection in children. J Pediatr Infect Dis Soc. 2015;4:313–22.
- Zajac-Spychala O, Gowin E, Fichna P, et al. Pneumocystis pneumonia in children - the relevance of chemoprophylaxis in different groups of immunocompromised and immunocompetent paediatric patients. Cent Eur J Immunol. 2015;40:91–5.
- Caselli D, Petris MG, Rondeli R, et al. Single-day trimethoprim/ sulfamethoxazole prophylaxis for pneumocystis pneumonia in children with cancer. J Pediatr. 2014;164:389–92.
- Castagnola E, Bagnasco F, Bandettini R, et al. Role of acute graftversus-host disease in the risk of bacteremia and invasive fungal disease after allogeneic Hemopoietic stem cell transplantation in children. Results from a single-center observational study. Biol Blood Marrow Transplant. 2014;20:1056–73.
- Castagnola E, Cesaro S, Giacchino M, Livadiotti S, Tucci F, Zanazzo G, et al. Fungal infections in children with cancer: a prospective, multicenter surveillance study. Pediatr Infect Dis J. 2006;25:634–9.

- Fisher BT, Robinson PD, Lehrnbecher T, Steinbach WJ, Zaoutis TE, Phillips B, et al. Risk factors for invasive fungal disease in pediatric cancer and hematopoietic stem cell transplantation: a systematic review. J Pediatric Infect Dis Soc. 2017; https://doi.org/10. 1093/jpids/pix030.
- Hale KA, Shaw PJ, Dalla-Pozza L, MacIntyre CR, Isaacs D, Sorrell TC. Epidemiology of paediatric invasive fungal infections and a case-control study of risk factors in acute leukaemia or post stem cell transplant. Br J Haematol. 2010;149:263–72.
- Sung L, Gamis A, Alonzo TA, Buxton A, Britton K, DeSwarte-Wallace J, et al. Infections and association with different intensity of chemotherapy in children with acute myeloid leukemia. Cancer. 2009;115:1100–8.
- Styczynski J, Czyzewski K, Wysocki J, et al. Increased risk of infections and infection-related mortality in children undergoing haematopoietic stem cell transplantation compared to conventional anticancer therapy: a multicentre nationwide study. Clin Microbiol Infect. 2016;22:179–88.
- Castagnola E, Bagnasco F, Amoroso L, Caviglia I, Caruso S, Faraci M, et al. Role of management strategies in reducing mortality from invasive fungal disease in children with cancer or receiving hemopoietic stem cell transplant: a single center 30-year experience. Pediatr Infect Dis J. 2014;33:233–7.
- McQuay HJ, Moore RA. Issues involved in making choices in prophylaxis. Ann Intern Med. 1997;126:712–20.
- Castagnola E, Rossi MR, Cesaro S, Livadiotti S, Giacchino M, Zanazzo G, et al. Incidence of bacteremias and invasive mycoses in children with acute non-lymphoblastic leukemia: results from a multi-center Italian study. Pediatr Blood Cancer. 2010;55:1103–7.
- De Pauw BE, Donnelly JP. Prophylaxis and aspergillosis-has the principle been proven? N Engl J Med. 2007;356:409–11.
- Tatarelli P, Faraci M, Caviglia I, Bandettini R, Cangemi G, Magnano GM, et al. Epidemiology of invasive fungal diseases in children with solid tumours undergoing autologous haematopoietic stem cell transplantation: a 10-year experience in a tertiary Italian centre. Mycoses. 2017;60:517–20.
- Castagnola E, Fontana V, Caviglia I, Caruso S, Faraci M, Fioredda F, et al. A prospective study on the epidemiology of febrile episodes during chemotherapy-induced neutropenia in children with cancer or after hemopoietic stem cell transplantation. Clin Infect Dis. 2007;45:1296–304.
- Castagnola E, Caviglia I, Pistorio A, Fioredda F, Micalizzi C, Viscoli C, et al. Bloodstream infections and invasive mycoses in children undergoing acute leukaemia treatment: a 13-year experience at a single Italian institution. Eur J Cancer. 2005;41:1439–45.
- Haupt R, Romanengo M, Fears T, Viscoli C, Castagnola E. Incidence of septicaemias and invasive mycoses in children undergoing treatment for solid tumours: a 12-year experience at a single Italian institution. Eur J Cancer. 2001;37:2413–9.
- Sinclair JC, Cook RJ, Guyatt GH, Pauker SG, Cook DJ. When should an effective treatment be used? Derivation of the threshold number needed to treat and the minimum event rate for treatment. J Clin Epidemiol. 2001;54:253–62.
- Fleming S, Yannakou CK, Haeusler GM, Clark J, Grigg A, Heath CH, et al. Consensus guidelines for antifungal prophylaxis in haematological malignancy and haemopoietic stem cell transplantation, 2014. Intern Med J. 2014;44:1283–97.
- 22. Groll AH, Castagnola E, Cesaro S, Dalle JH, Engelhard D, Hope W, et al. Fourth European conference on infections in Leukaemia (ECIL-4): guidelines for diagnosis, prevention, and treatment of invasive fungal diseases in paediatric patients with cancer or allogeneic haemopoietic stem-cell transplantation. Lancet Oncol. 2014;15:e327–40.
- Lehmbecher T. Antifungal prophylaxis in pediatric patients undergoing therapy for cancer: drugs and dosing. Curr Opin Infect Dis. 2015;28:523–31.

- Dvorak CC, Steinbach WJ, Brown JM, Agarwal R. Risks and outcomes of invasive fungal infections in pediatric patients undergoing allogeneic hematopoietic cell transplantation. Bone Marrow Transplant. 2005;36:621–9.
- Lai HP, Chen YC, Chang LY, Lu CY, Lee CY, Lin KH, et al. Invasive fungal infection in children with persistent febrile neutropenia. J Formos Med Assoc. 2005;104:174–9.
- Mor M, Gilad G, Kornreich L, Fisher S, Yaniv I, Levy I. Invasive fungal infections in pediatric oncology. Pediatr Blood Cancer. 2011;56:1092–7.
- Tragiannidis A, Dokos C, Lehrnbecher T, Groll AH. Antifungal chemoprophylaxis in children and adolescents with haematological malignancies and following allogeneic haematopoietic stem cell transplantation: review of the literature and options for clinical practice. Drugs. 2012;72:685–704.
- Lanciotti M, Pigullo S, Lanza T, Dufour C, Caviglia I, Castagnola E. Possible role of toll-like receptor 9 polymorphism in chemotherapy-related invasive mold infections in children with hematological malignancies. Pediatr Blood Cancer. 2008;50:944.
- Bochud PY, Chien JW, Marr KA, Leisenring WM, Upton A, Janer M, et al. Toll-like receptor 4 polymorphisms and aspergillosis in stem-cell transplantation. N Engl J Med. 2008;359:1766–77.
- Koldehoff M, Beelen DW, Elmaagacli AH. Increased susceptibility for aspergillosis and post-transplant immune deficiency in patients with gene variants of TLR4 after stem cell transplantation. Transpl Infect Dis. 2013;15:533–9.
- Fischer M, Spies-Weisshart B, Schrenk K, Gruhn B, Wittig S, Glaser A, et al. Polymorphisms of Dectin-1 and TLR2 predispose to invasive fungal disease in patients with acute myeloid leukemia. PLoS One. 2016;11:e0150632.
- Zaoutis TE, Heydon K, Chu JH, Walsh TJ, Steinbach WJ. Epidemiology, outcomes, and costs of invasive aspergillosis in immunocompromised children in the United States, 2000. Pediatrics. 2006;117:e711–6.
- Crassard N, Hadden H, Piens MA, Pondarré C, Hadden R, Galambrun C, et al. Invasive aspergillosis in a paediatric haematology department: a 15-year review. Mycoses. 2008;51: 109–16.
- Castagnola E, Conte M, Parodi S, Papio F, Caviglia I, Haupt R. Incidence of Bacteremias and invasive mycoses in children with high risk neuroblastoma. Pediatr Blood Cancer. 2007;49:672–7.
- 35. Stergiopoulou T, Walsh TJ. Clinical pharmacology of antifungal agents to overcome drug resistance in pediatric patients. Expert Opin Pharmacother. 2015;16:213–26.
- Ashbee HR, Barnes RA, Johnson EM, Richardson MD, Gorton R, Hope WW. Therapeutic drug monitoring (TDM) of antifungal agents: guidelines from the British Society for Medical Mycology. J Antimicrob Chemother. 2014;69:1162–76.
- Science M, Robinson PD, MacDonald T, Rassekh SR, Dupuis LL, Sung L. Guideline for primary antifungal prophylaxis for pediatric patients with cancer or hematopoietic stem cell transplant recipients. Pediatr Blood Cancer. 2014;61:393–400.
- Doering M, Cabanillas Stanchi KM, Queudeville M, et al. Efficacy, safety and feasibility of antifungal prophylaxis with posaconazole tablet in paediatric patients after haematopoietic stem cell transplantation. J Cancer Res Clin Oncol. 2017;143:1281–92.
- Vanstraelen K, Colita A, Bica AM, Mols R, Augustijns P, Peersman N, et al. Pharmacokinetics of Posaconazole oral suspension in children dosed according to body surface area. Pediatr Infect Dis J. 2016;35:183–8.
- Green MR, Woolery JE. Optimising absorption of posaconazole. Mycoses. 2011;54:e775–9.
- Zembles TN, Thompson NE, Havens PL, Kaufman BA, Huppler AR. An optimized Voriconazole dosing strategy to achieve therapeutic serum concentrations in children younger than 2 years old. Pharmacotherapy. 2016;36:1102–8.

- Goodman JL, Winston DJ, Greenfield RA, Chandrasekar PH, Fox B, Kaizer H, et al. A controlled trial of fluconazole to prevent fungal infections in patients undergoing bone marrow transplantation. N Engl J Med. 1992;326:845–51.
- 43. Slavin MA, Osborne B, Adams R, Levenstein MJ, Schoch HG, Feldman AR, et al. Efficacy and safety of fluconazole prophylaxis for fungal infections after marrow transplantation–a prospective, randomized, double-blind study. J Infect Dis. 1995;171:1545–52.
- 44. Winston DJ, Maziarz RT, Chandrasekar PH, Lazarus HM, Goldman M, Blumer JL, et al. Intravenous and oral itraconazole versus intravenous and oral fluconazole for long-term antifungal prophylaxis in allogeneic hematopoietic stem-cell transplant recipients. A multicenter, randomized trial. Ann Intern Med. 2003;138: 705–13.
- Pana ZD, Kourti M, Vikelouda K, et al. Voriconazole antifungal prophylaxis in children with malignancies: a Nationwide study. J Pediatr Hematol Oncol. 2018;40:22–6.
- Maron GM, Hayden RT, Rodriguez A, Rubnitz JE, Flynn PM, Shenep JL, et al. Voriconazole prophylaxis in children with cancer: changing outcomes and epidemiology of fungal infections. Pediatr Infect Dis J. 2013;32:e451–5.
- Cornely OA, Maertens J, Winston DJ, Perfect J, Ullmann AJ, Walsh TJ, et al. Posaconazole vs. fluconazole or itraconazole prophylaxis in patients with neutropenia. N Engl J Med. 2007;356:348–59.
- Cesaro S, Milano GM, Aversa F. Retrospective survey on the offlabel use of posaconazole in pediatric hematology patients. Eur J Clin Microbiol Infect Dis. 2011;30:595–6.
- Vicenzi EB, Calore E, Decembrino N, Berger M, Perruccio K, Carraro F, et al. Posaconazole oral dose and plasma levels in pediatric hematology-oncology patients. Eur J Haematol. 2017;100: 315–22. https://doi.org/10.1111/ejh.13017.
- Ullmann AJ, Lipton JH, Vesole DH, Chandrasekar P, Langston A, Tarantolo SR, et al. Posaconazole or fluconazole for prophylaxis in severe graft-versus-host disease. N Engl J Med. 2007;356:335–47.
- Krishna G, Moton A, Ma L, Medlock MM, McLeod J. Pharmacokinetics and absorption of posaconazole oral suspension under various gastric conditions in healthy volunteers. Antimicrob Agents Chemother. 2009;53:958–66.
- Gwee A, Cranswick N, Curtis N. Posaconazole: promising but problematic in practice in pediatric patients. Pediatr Infect Dis J. 2015;34:604–6.
- 53. European, Medicine, Agency. Posaconazole Summary of Product Characteristics. In.
- Chau MM, Kong DC, van Hal SJ, et al. Consensus guidelines for optimising antifungal drug delivery and monitoring to avoid toxicity and improve outcomes in patients with haematological malignancy, 2014. Intern Med J. 2014;44:1364–88.
- Allegra S, Fatiguso G, De Francia S, et al. Therapeutic drug monitoring of voriconazole for treatment and prophylaxis of invasive fungal infection in children. Br J Clin Pharmacol. 2018;84:197– 203.
- 56. Bartelink IH, Wolfs T, Jonker M, de Waal M, Egberts TCG, Ververs TT, et al. Highly variable plasma concentrations of voriconazole in pediatric hematopoietic stem cell transplantation patients. Antimicrob Agents Chemother. 2013;57:235–40.
- Boast A, Curtis N, Cranswick N, Gwee A. Voriconazole dosing and therapeutic drug monitoring in children: experience from a paediatric tertiary care centre. J Antimicrob Chemother. 2016;71:2031–6.
- Dote S, Sawai M, Nozaki A, Naruhashi K, Kobayashi Y, Nakanishi H. A retrospective analysis of patient-specific factors on voriconazole clearance. J Pharm Health Care Sci. 2016;2:10.
- 59. van Wanrooy MJ, Span LF, Rodgers MG, et al. Inflammation is associated with voriconazole trough concentrations. Antimicrob Agents Chemother. 2014;58:7098–101.

- Wallace KL, Filipek RL, La Hoz RM, Williamson JC. Subtherapeutic voriconazole concentrations associated with concomitant dexamethasone: case report and review of the literature. J Clin Pharm Ther. 2016;41:441–3.
- Bruggemann RJ, Alffenaar JW, Blijlevens NM, et al. Clinical relevance of the pharmacokinetic interactions of azole antifungal drugs with other coadministered agents. Clin Infect Dis. 2009;48:1441– 58.
- 63. van Burik JA, Ratanatharathorn V, Stepan DE, et al. Micafungin versus fluconazole for prophylaxis against invasive fungal infections during neutropenia in patients undergoing hematopoietic stem cell transplantation. Clin Infect Dis. 2004;39:1407–16.
- Doring M, Hartmann U, Erbacher A, et al. Caspofungin as antifungal prophylaxis in pediatric patients undergoing allogeneic hematopoietic stem cell transplantation: a retrospective analysis. BMC Infect Dis. 2012;12:151.
- Bochennek K, Tramsen L, Schedler N, Becker M, Klingebiel T, Groll AH, et al. Liposomal amphotericin B twice weekly as antifungal prophylaxis in paediatric haematological malignancy patients. Clin Microbiol Infect. 2011;17:1868–74.
- Hullard-Pulstinger A, Holler E, Hahn J, Andreesen R, Krause SW. Prophylactic application of nebulized liposomal amphotericin B in hematologic patients with neutropenia. Onkologie. 2011;34:254–8.
- 67. Maertens J, Cesaro S, Maschmeyer G, Einsele H, Donnelly JP, Alanio A, et al. ECIL guidelines for preventing pneumocystis jirovecii pneumonia in patients with haematological malignancies and stem cell transplant recipients. J Antimicrob Chemother. 2016;71:2397–404.
- Dallorso S, Castagnola E, Garaventa A, Rossi GA, Giacchino R, Dini G. Early onset of pneumocystis carinii pneumonia in a patient receiving bone marrow transplantation from a matched unrelated donor. Bone Marrow Transplant. 1994;13:106–7.
- Castagnola E, Zarri D, Caprino D, Losurdo G, Micalizzi C. Cotrimoxazole prophylaxis of pneumocystis carinii infection during the treatment of childhood acute lymphoblastic leukemia– beware non compliance in older children and adolescents. Support Care Cancer. 2001;9:552–3.
- Sung L, Nathan PC, Lange B, Beyene J, Buchanan GR. Prophylactic granulocyte colony-stimulating factor and granulocyte-macrophage colony-stimulating factor decrease febrile neutropenia after chemotherapy in children with cancer: a metaanalysis of randomized controlled trials. J Clin Oncol. 2004;22: 3350–6.
- Sung L, Aplenc R, Alonzo TA, Gerbing RB, Lehrnbecher T, Gamis AS. Effectiveness of supportive care measures to reduce infections in pediatric AML: a report from the Children's oncology group. Blood. 2013;121:3573–7.
- Delfino D, Scordino F, Pernice I, Lo Passo C, Galbo R, David A, et al. Potential association of specific Candida Parapsilosis genotypes, bloodstream infections and colonization of health workers' hands. Clin Microbiol Infect. 2014;20:O946–51.
- Yildirim M, Sahin I, Kucukbayrak A, et al. Hand carriage of Candida species and risk factors in hospital personnel. Mycoses. 2007;50:189–92.
- 74. Storti LR, Pasquale G, Scomparim R, Galastri AL, Alterthum F, Gambale W, et al. Candida spp. isolated from inpatients, the environment, and health practitioners in the pediatric unit at the

Universitary Hospital of the Jundiai Medical College, state of Sao Paulo, Brazil. Rev Soc Bras Med Trop. 2012;45:225–31.

- 75. WHO Guidelines Approved by the Guidelines Review Committee. In WHO Guidelines on Hand Hygiene in Health Care: First Global Patient Safety Challenge Clean Care Is Safer Care. Geneva: World Health Organization 2009.
- Yildirim M, Sahin I, Oksuz S, Sencan I, Kucukbayrak A, Cakir S, et al. Hand carriage of Candida occurs at lesser rates in hospital personnel who use antimicrobial hand disinfectant. Scand J Infect Dis. 2014;46:633–6.
- Zakhour R, Chaftari AM, Raad II. Catheter-related infections in patients with haematological malignancies: novel preventive and therapeutic strategies. Lancet Infect Dis. 2016;16:e241–50.
- Zakhour R, Hachem R, Alawami HM, Jiang Y, Michael M, Chaftari AM, et al. Comparing catheter-related bloodstream infections in pediatric and adult cancer patients. Pediatr Blood Cancer. 2017;64
- Lo Vecchio A, Schaffzin JK, Ruberto E, Caiazzo MA, Saggiomo L, Mambretti D, et al. Reduced central line infection rates in children with leukemia following caregiver training: a quality improvement study. Medicine (Baltimore). 2016;95:e3946.
- Castagnola E, Viscoli C. Invasive aspergillosis in malignancy and stem cell transplant recipients. In: Largè JP, Steinbach WJ, editors. Aspergillus fumigatus and aspergillosis. Washington: ASM Press; 2009. p. 519–30.
- Benet T, Nicolle MC, Thiebaut A, Piens MA, Nicolini FE, Thomas X, et al. Reduction of invasive aspergillosis incidence among immunocompromised patients after control of environmental exposure. Clin Infect Dis. 2007;45:682–6.
- Pokala HR, Leonard D, Cox J, Metcalf P, McClay J, Siegel J, et al. Association of hospital construction with the development of healthcare associated environmental mold infections (HAEMI) in pediatric patients with leukemia. Pediatr Blood Cancer. 2014;61: 276–80.
- Hahn T, Cummings KM, Michalek AM, Lipman BJ, Segal BH, McCarthy PL. Efficacy of high-efficiency particulate air filtration in preventing aspergillosis in immunocompromised patients with hematologic malignancies. Infect Control Hosp Epidemiol. 2002;23:525–31.
- Humphreys H. Positive-pressure isolation and the prevention of invasive aspergillosis. What is the evidence? J Hosp Infect. 2004;56:93–100. quiz 163
- Berthelot P, Loulergue P, Raberin H, Turco M, Mounier C, Tran Manh Sung R, et al. Efficacy of environmental measures to decrease the risk of hospital-acquired aspergillosis in patients hospitalised in haematology wards. Clin Microbiol Infect. 2006;12:738–44.
- Cornet M, Levy V, Fleury L, Lortholary J, Barquins S, Coureul MH, et al. Efficacy of prevention by high-efficiency particulate air filtration or laminar airflow against aspergillus airborne contamination during hospital renovation. Infect Control Hosp Epidemiol. 1999;20:508–13.
- Maschmeyer G, Neuburger S, Fritz L, Bohme A, Penack O, Schwerdtfeger R, et al. A prospective, randomised study on the use of well-fitting masks for prevention of invasive aspergillosis in high-risk patients. Ann Oncol. 2009;20:1560–4.
- Anaissie EJ, Stratton SL, Dignani MC, Lee CK, Summerbell RC, Rex JH, et al. Pathogenic molds (including aspergillus species) in hospital water distribution systems: a 3-year prospective study and clinical implications for patients with hematologic malignancies. Blood. 2003;101:2542–6.