

Perivascular Fat and its Role in Vascular Disease, Insulin Resistance and Diabetes

Chengyu Xu · Gianluca Iacobellis

Published online: 9 January 2014
© Springer Science+Business Media New York 2014

Abstract Perivascular adipose tissue is a visceral fat depot with an anatomical and functional contiguity to the vasculature system. Recent evidence suggests that perivascular adipose tissue could mechanically and functionally affect the vasculature, thereby possibly playing a role in adiposity-related atherosclerosis. Experimental and clinical observations suggest both favorable and unfavorable effects of perivascular fat. This review focuses on the emerging physiological and pathophysiological aspects of the perivascular fat and its role in vascular disease, insulin resistance and diabetes.

Keywords Perivascular fat · Epicardial fat · Diabetes · Insulin resistance

Introduction

A growing body of evidence suggests that regional fat distribution plays an important part in the development of an unfavorable metabolic and cardiovascular risk profile. Thus, the increased accumulation of visceral fat is now widely seen as a defining characteristic of the so-called metabolic syndrome [1, 2]. The recognition that adipose tissue is a highly complex endocrine organ that generates various molecules with profound local and systemic effects has spawned a remarkable interest in adipose-tissue research [3, 4]. Despite their similar qualitative properties, different types of adipose tissue, particularly subcutaneous and visceral adipose depots,

are now recognized as having distinct quantitative characteristics [5, 6]. While much of the interest has focused on the importance of intra-abdominal visceral fat, some extra-abdominal visceral fat depots, including epicardial [7–9] and perivascular fat, have also been studied [10]. In this review paper we focus on the biomolecular properties of perivascular adipose tissue (PAT), and its role as cardiovascular risk factor.

Anatomy of Perivascular Fat

All vascular vessels are surrounded by a perivascular sheath of adipose tissue called perivascular fat. Histologically, the perivascular fat differs according to which type of vessel they are surrounding: white adipose tissue in resistance vessels, whereas white and brown adipose tissue (BAT) in larger vessels like aorta [11]. At first it was suggested that this depot would offer mechanical support for the vasculature system, but as it has been discovered this is also an active endocrine organ which is thoroughly related to muscle tone control.

Physiology of Perivascular Fat

PAT plays a fundamental role in the “brain-vessel axis” control of vascular muscle tone [12]. The control of vascular function is dual, and depends on the stimuli around the vessel [13]. Vascular tone control by PAT is a complicated network of vasoactive substances, pathways and cellular subtypes that have recently begun to be described [14, 15, 16, 17, 18]. Adipose-derived substances can modulate vascular tone. Insulin can also exert a dual role in modulating the vascular tone, by controlling endothelin-1 secretion via ERK1/2 (extracellular signal regulated kinases) and endothelial derived nitric oxide (eNOS) activation with subsequent production of NO via phosphatidylinositol-3-kinase and protein kinase B (PI3K/Akt)

This article is part of the Topical Collection on *Diabetes and Insulin Resistance*

C. Xu · G. Iacobellis (✉)
Division of Diabetes, Endocrinology and Metabolism, Department of Medicine, Miller School of Medicine, University of Miami, 1400 NW 10th Ave, Dominion Tower suite 805-807, Miami, FL 33136, USA
e-mail: giacobellis@med.miami.edu

[14]. Vascular tone also depends on adipose-derived relaxing factors (ADRFs) which are capable of activating inward rectifying K^+ channels to inhibit vasoconstriction [19]. Leptin and adiponectin have been considered to function as ADRFs. Sahin et al. [20] reported that leptin was capable of inducing vessel relaxation via endothelium derived nitric oxide (AMPK and Akt phosphorylation of eNOS), hydrogen peroxide, and also by inducing endothelium-derived hyperpolarizing factor [21], mechanisms which are exacerbated during the initial adapting period during over-nutrition and obesity to preserve vascular tone [22]. On the other hand, adiponectin is known to open K channels through its receptors adiponectin receptor 1 (AdipoR1) and AdipoR2 inducing relaxation in aortic and mesenteric rings [23]. Other relaxing factors include Angiotensin, which induces vasodilation by antagonizing angiotensin 1 (AT1) receptors in a bradykinin-NO pathway [24], and H_2O_2 through activation of several types of K^+ channels (calcium-dependent [25••], ATP-dependent [26], and voltage-dependent [27]). In summary, PAT has been recognized as a very active metabolic organ responsible for mechanical and functional tasks. PAT offers mechanical support during the arterial wave pulse and it can even maintain the integrity of the vascular wall during the pathological expansion during the formation of an atheromatous plaque.

Pathophysiology of Perivascular Fat

PAT & Atherosclerosis

Atherosclerosis and its consequent coronary artery disease (CAD) are the leading cause of death worldwide [28]. Current evidence has suggested that the basic pathogenesis includes lipid metabolism abnormalities and an awry immune response characterized by increased leukocyte trafficking, enhanced inflammatory microenvironment and progressive plaque formation [29]. Given the close anatomical proximity and its importance in vessel embryology, PAT has been placed in a very compromising place, with evidence that points to a local source of adipokines that seems to be the culprit of the very initial vascular dysfunction that ends in atherosclerosis.

PAT & Adipokines

PAT secretes adipokines that modulate and affect vascular function. Inflammatory cell infiltration is markedly increased in PAT surrounding atherosclerotic human aorta as compared with a non-diseased aorta, and the inflammatory gene expression is upregulated in this location [30]. PAT is able to promote constriction via induction of NADPH and synthesis of superoxide anion, which enhances mitogen activated protein kinase/extracellular signal regulated kinases (MAPK/ERK) pathway [31], suggesting a never ending battle of control of vascular tone; only when one is truly defective will the other

be able to sustain its corresponding effect. Additionally, superoxide anion is capable of uncoupling the eNOS activity, maintaining only the enzymes production of superoxide, which is an actual intermediary in the synthesis of NO [32]. The overproduction of superoxide anion and oxygen singlet (ROS) enhances the levels of nitrogen species such as peroxynitrite, which is a potent vasoconstrictor and inductor of apoptosis in cardiomyocytes, endothelium and vascular smooth muscle [33]. Additionally, PAT can secrete a number of constrictors like angiotensin-II (Ang-II), resistin and visfatin.

Ang-II is not only a potent vasoconstrictor via AT1 [34], but also an inductor of nicotinamides adenine dinucleotide (NADPH) activity and superoxide production [35]. Moreover, this hormone is capable of inducing CD44 and C chemokine receptor 5 (CCR5) in T cells and production of regulated upon activation normal T cell expressed and secreted (RANTES) in vascular endothelium, creating a migration gradient for $CD4^+$ and in a lesser extent to $CD8^+$, although 30 % of T cells present in perivascular fat are $CD4^+CD8^-$ [36], capable of secreting IL-17 - a proinflammatory cytokine [37]. These $CCR5^+CD44^{hi}$ cells are in fact effector T cells with a lower threshold for activation, releasing TNF-alpha and IFN-gamma which contribute to endothelial dysfunction and hypertension [36]. Ang-II has also been related to the formation of "neoantigens", such as heat shock protein (HSP) 70 and HSP25 [38], responsible for the production of autoantibodies in an endothelial-injury manner and enhanced atherogenesis in animal models [39, 40].

Resistin, another adipokine fairly recently added to the adipocyte secretome, has been linked to type 2 diabetes and insulin resistance [41]. Several pro-atherogenic properties have been described, including inducing oxidative stress via uncoupling of eNOS and activation of Jun N-terminal protein kinase (JNK) [42], vascular smooth muscle cell hyperplasia promotion with subsequent thickening of the intima [43], increased expression of IL-1beta, IL-6, TNF-alpha, alpha4beta1 integrin (VLA-4) and vascular cell adhesion molecule 1 (VCAM-1) [44•], becoming a useful marker for vascular inflammation [45•] and acute coronary syndrome [46]. Another adipokine involved in this scenario is visfatin (pre-B cell colony enhancing factor or nicotinamide phosphoribosyltransferase [Nampt]), a controversial protein with catalytic activity, capable of improving insulin sensitivity [47], inducing B cell differentiation [48], and mediating vessel relaxation through NO synthesized by endothelial cells [49]. Even though these properties seem beneficial, during type 2 diabetes and obesity visfatin has been related to impairment of vasodilation due to stimulation of NADPH via its Nampt activity [50] and synthesis of proinflammatory cytokines (IL-6 and C-reactive Protein) from VAT during obesity [51], using as main source incoming and circulating leukocytes [52].

Leptin also has a principal role in PAT physiology. As previously reported, leptin induces vasodilation, yet there is a concept – the “*leptin paradox*” – where this cytokine is associated with inhibition of NO-dependent relaxation [53]. Tune and Considine [53] reported that the vasodilating effects of leptin are dose-dependent, where leptin at 10–90 ng/ml impairs acetylcholine-mediated relaxation, yet at levels higher than 160 ng/ml induces vasodilation. These authors conclude that leptin induced vasodilation might just be a pharmacological phenomenon, nevertheless, other studies advocate for a physiological role [54–56]. There are other novel adipokines [57] that have vascular tone modulating functions like omentin (described previously), chemerin, vaspin, and nesfatin. Chemerin is a protein which has been described as a potent chemoattractant for macrophages and immature dendritic cells [58]. This adipokine has been related to degree of coronary atherosclerosis and epicardial fat [59], arterial stiffness [60], severity of coronary arterial disease in patients with metabolic syndrome [61], and insulin resistance and glucose intolerance [62, 63]. Its ability to attract immunocytes to the subendothelium, contributes to the progression of atherosclerosis plaque and the vicious circle of localized insulin resistance (see the following section). Vaspin (SERPIN12) is an adipocyte-derived serine protease inhibitor with insulin sensitizing properties [64] and capable of protecting endothelial cells from apoptosis via PI3K/Akt pathway [65] protecting them from lipid toxicity. Hypovaspinemia has been related to ischemic cardiovascular events [66] and severity of CAD [67], while normal secretion inhibits TNF-alpha-induced expression of ICAM-1 preventing the subsequent activation of NF-kB and PKC protecting the vascular smooth muscle cells from an inflammatory profile [68]. Finally, nesfatin is an anorexigenic hormone with influence in the energy expenditure control circuit [69], and capable of inducing glucagon secretion [70]. It has been recently published that it can also inhibit soluble guanylate cyclase, which reduces NO-dependent vasodilation, contributing to vascular dysfunction [71].

PAT & Vascular Smooth Muscle Cells

There is an undeniable relationship between vascular smooth muscle cells and the white adipose tissue sheath that surrounds them. These cells are the basic motor units of the vessels, but they are also capable of altering its structure and modifying the extracellular matrix composition. Their phenotypes vary according to different stimuli and are considered not terminally differentiated, being classified into 2 classes: the spindle-shaped and the epithelioid vascular smooth muscle cells. The crosstalk between PAT and vascular smooth muscle cells is very complex and fairly new in concept. PAT is capable of modulating neointimal hyperplasia by increased secretion of TNF-alpha with parallel decreased secretion of adiponectin [72•]. Free fatty acids are released which can induce

inflammation, as was proven using oleic acid in an adipocyte-conditioned media, where NF-kB and mTOR pathways are activated inducing proliferation of the smooth muscle cells [73, 74]. Insulin-like growth factor-I is able to induce vascular muscle cells migration via ERK1/2, signaling which is inhibited by adiponectin implicating AMPK activation [75]. Interestingly, PAT can also modulate the local renin-angiotensin-aldosterone system by inducing JNK, p38MAPK, and ERK1/2 via mineralocorticoid receptor, glucocorticoid receptor and AT1 receptor [76, 77].

PAT & Localized Insulin Resistance

Obesity is a disease closely associated with an array of cardiovascular risk factors such as diabetes, hypertension, proinflammatory and prothrombotic states, and obstructive sleep apnea. Remarkably these five risk factors are also related to insulin resistance [78–82]. Obesity milieu induces inflammation of the PAT, with the subsequent release of adipocytokines and the start and amplification of several pathways that will conclude in the development of the atherosclerotic plaque and the risk of rupture [83]. In fact, Rittig et al. [84] reported that PAT is an independent player in insulin resistance and it correlates negatively with insulin sensitivity. Moreover, they also reported that PAT secretes higher amounts of angiogenic factors like MC-1, insulin-like growth factor-binding protein-3, thrombospondin-1, fibroblastic growth factor and hepatocyte growth factor [85•], contributing to plaque rupture risk [86]. Obesity has systemic effects which affect short and long axis loops during the development of the disease, but at the vascular level obesity has an ample role in its pathophysiology like enhanced vasoconstriction, functional hyperemia, sympathetic neural hyperactivity, microvascular rarefaction, vascular remodeling, and local inflammation [87]. Insulin is a Janus-type controller in tone control: it can induce vasodilation via PKB/Akt and eNOS induction [88], while it induces vasoconstriction via MAPK and ERK1/2 pathways culminating in endothelin-1 synthesis [88]; nevertheless, the overall result is vasodilation [89]. As insulin resistance develops hyperinsulinemia occurs, stimulating MAPK pathways while PI3K/Akt is blunted, inducing an increase production of endothelin-1 and enhanced arterial and terminal arterioles constriction. This phenomenon alters the uptake of glucose and perfusion of the vascular muscles, describing a vicious circle concerning local insulin resistance and vascular tone control [90].

Conclusions

The general hypothesis for atherosclerosis is an “inside-out” model which is focused on lipid accumulation and monocyte attraction toward vascular intima. However, accumulating evidence supports an outside-in hypothesis in which the vascular *adventitia* promotes vascular inflammation and tone

deregulation [91] which would suggest that PAT have a role in this signaling landscape. Results from *in vitro*, *ex vivo*, animal and clinical studies support the double role of PAT in the pathogenesis of atherosclerosis. It is possible that a mass-dependent mechanism could influence the equilibrium between harmful and protective effects of this fat depot; however, a causal effect of PAT on atherosclerosis remains to be demonstrated. Therefore, future studies in this direction are needed to confirm or refute the relationship between PAT and cardiovascular risk.

Compliance with Ethics Guidelines

Conflict of Interest Chengyu Xu and Gianluca Iacobellis declare no conflicts of interest.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by the author.

References

Papers of particular interest, published recently, have been highlighted as

- Of importance
- Of major importance

1. Grundy SM et al. Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association Conference on scientific issues related to definition. *Circulation*. 2004;109:433–8.
2. Carr DB et al. Intra-abdominal fat is a major determinant of the National Cholesterol Education Program Adult Treatment Panel III criteria for the metabolic syndrome. *Diabetes*. 2004;53:2087–94.
3. Kershaw EE et al. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab*. 2004;89:2548–56.
4. Sharma AM. Adipose tissue: a mediator of cardiovascular risk. *Int J Obes Relat Metab Disord*. 2004;26 Suppl 4:S5–7.
5. Dusserre E et al. Differences in mRNA expression of the proteins secreted by the adipocytes in human subcutaneous and visceral adipose tissues. *Biochim Biophys Acta*. 2000;1500:88–96.
6. Wajchenberg BL. Subcutaneous and visceral adipose tissue: their relation to the metabolic syndrome. *Endocr Rev*. 2000;2:697–738.
7. Iacobellis G, Bianco AC. Epicardial adipose tissue: emerging physiological, pathophysiological and clinical features. *Trends Endocrinol Metab*. 2011;22:450–7.
8. Iacobellis G, Malavazos AE, Corsi MM. Epicardial fat: from the biomolecular aspects to the clinical practice. *Int J Biochem Cell Biol*. 2011;43:1651–4.
9. Iacobellis G, Corradi D, Sharma AM. Epicardial adipose tissue: anatomical, biomolecular and clinical relation to the heart. *Nat Cardiovasc Clin Pract Med*. 2005;2:536–43.
10. Sharma AM. Mediastinal fat, insulin resistance, and hypertension. *Hypertension*. 2004;44:117–8.
11. Gao YJ. Dual modulation of vascular function by perivascular adipose tissue and its potential correlation with adiposity/lipoatrophy-related vascular dysfunction. *Curr Pharm Des*. 2007;13:2185–92.
12. Guzik TJ, Marvar PJ, Czesnikiewicz-Guzik M, Korbut R. Perivascular adipose tissue as a messenger of the brain-vessel axis: role in vascular inflammation and dysfunction. *J Physiol Pharmacol*. 2007;58:591–610.
13. Takemori K, Gao YJ, Ding L, Lu C, Su LY, An WS, et al. Elevated blood pressure in transgenic lipoatrophic mice and altered vascular function. *Hypertension*. 2007;49:365–72.
14. Lohn M, Dubrovskaja G, Lauterbach B, Luft FC, Gollasch M, Sharma AM. Periadventitial fat releases a vascular relaxing factor. *FASEB J*. 2002;16:1057–63.
15. Sahin AS, Bariskaner H. The mechanisms of vasorelaxant effect of leptin on isolated rabbit aorta. *Fundam Clin Pharmacol*. 2007;21:595–600.
16. Beltowski J. Leptin and the regulation of endothelial function in physiological and pathological conditions. *Clin Ex Pharmacol Physiol*. 2012;39:168–78. *An overview of leptin and its regulation of endothelial function*.
17. Gil-Ortega M, Stucchi P, Guzmán-Ruiz R, Cano V, Arribas S, González MC, et al. Adaptive nitric oxide overproduction in perivascular adipose tissue during early diet-induced obesity. *Endocrinology*. 2010;151:3299–306.
18. Fésus G, Dubrovskaja G, Gorzelnik K, Kluge R, Huang Y, Luft F, et al. Adiponectin is a novel humoral vasodilator. *Cardiovasc Res*. 2007;75:719–27.
19. Ferrario CM, Chappell MC, Tallant EA, Brosnihan KB, Diz DI. Counterregulatory actions of angiotensin-(1–7). *Hypertension*. 1997;30:535–41.
20. Barlow RS, White RE. Hydrogen peroxide relaxes porcine coronary arteries by stimulating BK_{Ca} channel activity. *Am J Physiol*. 1998;275:H1283–9.
21. Wei EP, Kontos HA, Beckman JS. Mechanisms of cerebral vasodilation by superoxide, hydrogen peroxide, and peroxynitrite. *Am J Physiol*. 1996;271:H1262–6.
22. Gao YJ, Hirota S, Zhang D, Janssen LJ, Lee RMKW. Mechanisms of hydrogen peroxide-induced biphasic response in rat mesenteric artery. *Br J Pharmacol*. 2003;138:1085–92.
23. Yudkin JS, Eringa E, Stehouwer CDA. “Vasocrine” signalling from perivascular fat: a mechanism linking insulin resistance to vascular disease. *Lancet*. 2005;365:1817–20.
24. Maenhaut N, Van de Voorde J. Regulation of vascular tone by adipocytes. *BMC Med*. 2011;9:25.
25. •• Houben AJ, Eringa EC, Jonk AM, Serne EH, Smulders YM, Stehouwer CD. Perivascular fat and microcirculation: relevance to insulin resistance, diabetes, and cardiovascular disease. *Curr Cardiovasc Risk Rep*. 2012;6:80–90. *An update review of perivascular fat and microcirculation*.
26. Chaldakov GN. Cardiovascular adipobiology: a novel heart-associated adipose tissue in cardiovascular disease. *Ser J Exp Clin Res*. 2008;9:81–8.
27. Rajsheker S, Manka D, Blomkalns AL, Chatterjee TK, Stoll LL, Weintraub NL. Crosstalk between perivascular adipose tissue and blood vessels. *Curr Opin Pharmacol*. 2010;10:191–6.
28. Cooney MT, Dubina AL, Graham IN. Value and limitations of existing scores for the assessment of cardiovascular risk. *J Am Coll Cardiol*. 2009;54:1209–27.
29. Weber C, Noels H. Atherosclerosis: current pathogenesis and therapeutic options. *Nat Med*. 2011;17:1410–22.
30. Henrichot E, Juge-Aubry CE, Permin A, Pache JC, Velebit V, Dayer JM, et al. Production of chemokines by perivascular adipose tissue: a role in the pathogenesis of atherosclerosis? *Arterioscler Thromb Vasc Biol*. 2005;25:2594–9.
31. Gao YJ, Takemori K, Su LY, An WS, Lu C, Sharma AM, et al. Perivascular adipose tissue promotes vasoconstriction: the role of superoxide anion. *Cardiovasc Res*. 2006;71:363–73.
32. Zou MH, Shi C, Cohen RA. Oxidation of the zinc-thiolate complex and uncoupling of endothelial nitric oxide synthase by peroxynitrite. *J Clin Invest*. 2002;109:817–26.

33. Pacher P, Beckman JS, Liaudet L. Nitric oxide and peroxynitrite in health and disease. *Physiol Rev.* 2007;87:315–424.
34. Gustafsson F, Holstein-Rathlou NH. Angiotensin II modulates conducted vasoconstriction to norepinephrine and local electrical stimulation in rat mesenteric arterioles. *Cardiovasc Res.* 1999;44:176–84.
35. Griendling KK, Minieri CA, Ollerenshaw JD, Alexander RW. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res.* 1994;74:1141–8.
36. Guzik TJ, Hoch NE, Brown KA, McCann LA, Rahman A, Dikalov S, et al. Role of the T cell in the genesis of angiotensin II-induced hypertension and vascular dysfunction. *J Exp Med.* 2007;204:2449–60.
37. Umemura M, Kawabe T, Shudo K, Kidoya H, Fukui M, Asano M, et al. Involvement of IL-17 in Fas ligand-induced inflammation. *Int Immunol.* 2004;16:1099–108.
38. Ishizaka N, Aizawa T, Ohno M, Usui Si S, Mori I, Tang SS, et al. Regulation and localization of HSP70 and HSP25 in the kidney of rats undergoing long-term administration of angiotensin II. *Hypertension.* 2002;39:122–8.
39. Leng X, Zhan R, Wang Y, Liu X, Gong J, Gao X, et al. Anti-heat shock protein 70 autoantibody epitope changes and BD091 promotes atherosclerosis in rats. *Cell Stress Chaperones.* 2010;15:947–58.
40. Ghayour-Mobarhan M, Lamb DJ, Tavallaie S, Ferns GA. Relationship between plasma cholesterol, von Willebrand factor concentrations, extent of atherosclerosis and antibody titres to heat shock proteins-60, -65 and -70 in cholesterol-fed rabbits. *Int J Exp Pathol.* 2007;88:249–55.
41. Kusminski CM, McTernan PG, Kumar S. Role of resistin in obesity, insulin resistance and type II diabetes. *Clin Sci.* 2005;109:243–56.
42. Chen C, Jiang J, Lu JM, Chai H, Wang X, Lin PH, et al. Resistin decreases expression of endothelial nitric oxide synthase through oxidative stress in human coronary artery endothelial cells. *Am J Physiol Heart Circ Physiol.* 2010;299:H193–201.
43. Shyu KG, Lien LM, Wang BW, Kuan P, Chang H. Resistin contributes to neointimal formation via oxidative stress after vascular injury. *Clin Sci.* 2011;120:121–9.
44. Cho Y, Lee SE, Lee HC, Hur J, Lee S, Youn SW, et al. Adipokine resistin is a key player to modulate monocytes, endothelial cells, and smooth muscle cells, leading to progression of atherosclerosis in rabbit carotid artery. *J Am Coll Cardiol.* 2011;57:99–109. *It provides insights on the role of resistin in the atherosclerosis.*
45. Choi HY, Kim S, Yang SJ, Yoo HJ, Seo JA, Kim SG, et al. Association of adiponectin, resistin, and vascular inflammation: analysis with 18 F-fluorodeoxyglucose positron emission tomography. *Arterioscler Thromb Vasc Biol.* 2011;31:944–9. *It explains the relation of adiponectin, resistin, and vascular inflammation.*
46. Langheim S, Dreas L, Veschini L, Maisano F, Foglieni C, Ferrarello S, et al. Increased expression and secretion of resistin in epicardial adipose tissue of patients with acute coronary syndrome. *Am J Physiol Heart Circ Physiol.* 2010;298:H746–53.
47. Fukuhara A, Matsuda M, Mizushima M, Segawa K, Tanaka M, Kishimoto K, et al. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. *Sci.* 2005;307:426–30.
48. Samal B, Sun Y, Stearns G, Xie C, Suggs S, McNiece I. Cloning and characterization of the cDNA encoding a novel human pre-B cell colony-enhancing factor. *Mol Cell Biol.* 1994;14:1431–7.
49. Yamawaki H, Hara N, Okada M, Hara Y. Visfatin causes endothelium-dependent relaxation in isolated blood vessels. *Biochem Biophys Res Commun.* 2009;383:503–8.
50. Vallejo S, Romacho T, Angulo J, Villalobos LA, Cercas E, Leivas A, et al. Visfatin impairs endothelium-dependent relaxation in rat and human mesenteric microvessels through nicotinamide phosphoribosyltransferase activity. *PLoS ONE.* 2011;6:e27299.
51. Terra X, Auguet T, Quesada I, Aguilar C, Luna AM, Hernández M, et al. Increased levels of adipose tissue expression of visfatin in morbidly obese women. The relationship with pro-inflammatory cytokines. *Clin Endocrinol.* 2011. doi:10.1111/j.1365-2265.2011.04327.
52. Friebe D, Neef M, Kratzsch J, Erbs S, Dittich K, Garten A, et al. Leucocytes are a major source of circulating nicotinamide phosphoribosyltransferase (NAMPT)/pre-B cell colony (PBEF)/visfatin linking obesity and inflammation in humans. *Diabetologia.* 2011;54:1200–11.
53. Tune JD, Considine RV. Effect of leptin on cardiovascular physiology. *J Am Soc Hypertens.* 2007;1:231–41.
54. Busch HJ, Schirmer SH, Jost M, van Stijn S, Peters SL, Piek JJ, et al. Leptin augments cerebral hemodynamic reserve after three-vessel occlusion: distinct effects on cerebrovascular tone and proliferation in a nonlethal model of hyperperfused rat brain. *J Cereb Blood Flow Metab.* 2011;31:1085–92.
55. Biasucci LM, Graziani F, Rizzello V, Liuzzo G, Guidone C, De Caterina AR, et al. Paradoxical preservation of vascular function in severe obesity. *Am J Med.* 2010;123:727–34.
56. Leung YM, Kwan CY. Dual vascular effects of leptin via endothelium: hypothesis and perspective. *Chin J Physiol.* 2008;51:1–6.
57. Hideyuki Y. Vascular effects of novel adipocytokines: focus on vascular contractility and inflammatory responses. *Biol Pharm Bull.* 2011;34:307–10.
58. Wittamer V, Franssen JD, Vulcano M, Mirjole JF, Le Poul E, Migeotte I, et al. Specific recruitment of antigen-presenting cells by chemerin, a novel processed ligand from human inflammatory fluids. *J Exp Med.* 2003;198:977–85.
59. Gao X, Mi S, Zhang F, Gong F, Lai Y, Gao F, et al. Association of chemerin mRNA expression in human epicardial adipose tissue with coronary atherosclerosis. *Cardiovasc Diabetol.* 2011;10:87.
60. Yoo HJ, Choi HY, Yang SJ, Kim HY, Seo JA, Kim SG, et al. Circulating chemerin level is independently correlated with arterial stiffness. *J Atheroscler Thromb.* 2012;19:59–68.
61. Dong B, Ji W, Zhang Y. Elevated serum chemerin levels are associated with the presence of coronary artery disease in patients with metabolic syndrome. *Intern Med.* 2011;50:1093–7.
62. Ernst MC, Haidl ID, Zúñiga LA, Dranse HJ, Rourke JL, Zabel BA, et al. Disruption of the chemokine-like receptor-1 (CMKLR1) gene is associated with reduced adiposity and glucose intolerance. *Endocrinology.* 2012;153:672–82.
63. Shin HY, Lee DC, Chu SH, Jeon JY, Lee MK, Im JA, et al. Chemerin levels are positively correlated with abdominal visceral fat accumulation. *Clin Endocrinol.* 2011. doi:10.1111/j.1365-2265.2011.04217.x.
64. Hida K, Wada J, Eguchi J, Zhang H, Baba M, Seida A, et al. Visceral adipose tissue-derived serine protease inhibitor: a unique insulin-sensitizing adipocytokine in obesity. *Proc Natl Acad Sci U S A.* 2005;102:10610–5.
65. Jung CH, Lee WJ, Hwang JY, Seol SM, Kim YM, Lee YL, et al. Vaspin protects vascular endothelial cells against free fatty acid-induced apoptosis through a phosphatidylinositol 3-kinase/Akt pathway. *Biochem Biophys Res Commun.* 2011;413:264–9.
66. Aust G, Richter O, Rohm S, Kerner C, Hauss J, Klötting N, et al. Vaspin serum concentrations in patients with carotid stenosis. *Atherosclerosis.* 2009;204:262–6.
67. Li HL, Peng WH, Cui ST, Lei H, Wei YD, Li WM, et al. Vaspin plasma concentrations and mRNA expressions in patients with stable and unstable angina pectoris. *Clin Chem Lab Med.* 2011;49:1547–54.
68. Phalitakul S, Okada M, Hara Y, Yamawaki H. Vaspin prevents TNF- α -induced intracellular adhesion molecule-1 via inhibiting reactive oxygen species-dependent NF- κ B and PKC θ activation in cultured rat vascular smooth muscle cells. *Pharmacol Res.* 2011;64:493–500.
69. Kőnczöl K, Pintér O, Ferenczi S, Varga J, Kovács K, Palkovits M, et al. Nesfatin-1 exerts long-term effect on food intake and body temperature. *Int J Obes.* 2012. doi:10.1038/ijo.2012.2.

70. Riva M, Nitert MD, Voss U, Sathanoori R, Lindgvist A, Ling C, et al. Nesfatin-1 stimulates glucagon and insulin secretion and beta cell NUCB2 is reduced in human type 2 diabetic subjects. *Cell Tissue Res.* 2011;346:393–405.
71. Yamawaki H, Takahashi M, Mudohda M, Morita T, Okada M, Hara Y. A novel adipocytokine, nesfatin-1 modulates peripheral arterial contractility and blood pressure in rats. *Biochem Biophys Res Commun.* 2012;418:676–81.
72. Miao CY, Li ZY. The role of perivascular adipose tissue in vascular smooth muscle cell growth. *Br J Pharmacol.* 2012;165:643–58. *It nicely describes the role of perivascular adipose tissue in vascular smooth muscle cell growth.*
73. Lamers D, Schlich R, Greulich S, Sasson S, Sell H, Eckel J. Oleic acid and adipokines synergize in inducing proliferation and inflammatory signalling in human vascular smooth muscle cells. *J Cell Mol Med.* 2011;15:1177–88.
74. Barandier C, Montani JP, Yang Z. Mature adipocytes and perivascular adipose tissue stimulate vascular smooth muscle cell proliferation: effects of aging and obesity. *Am J Physiol Heart Circ Physiol.* 2005;289:H1807–13.
75. Motobayashi Y, Izawa-Ishizawa Y, Ishizawa K, Orino S, Yamaguchi K, Kawazoe K, et al. Adiponectin inhibits insulin-like growth factor-1-induced cell migration by the suppression of extracellular signal-regulated kinase $\frac{1}{2}$ activation, but not Akt in vascular smooth muscle cells. *Hypertens Res.* 2009;32:188–93.
76. Nguyen Dinh Cat A, Briones AM, Callera GE, Yogi A, He Y, Montezano AC, et al. Adipocyte-derived factors regulate vascular smooth muscle cells through mineralocorticoid and glucocorticoid receptors. *Hypertension.* 2011;58:479–88.
77. Herrmann J, Lerman LO, Rodriguez-Porcel M, Holmes Jr DR, Richardson DM, Ritman EL, et al. Coronary vasa vasorum neovascularization precedes epicardial endothelial dysfunction in experimental hypercholesterolemia. *Cardiovasc Res.* 2001;51:762–6.
78. Madonna R, De Caterina R. Atherogenesis and diabetes: focus on insulin resistance and hyperinsulinemia. *Rev Esp Cardiol.* 2012. doi:10.1016/j.recresp.2011.11.010.
79. Rojas J, Bermúdez V, Leal E, Cano R, Luti Y, Acosta L, et al. Insulinorresistencia e hiperinsulinemia como factores de riesgo para enfermedad cardiovascular. *AVFT.* 2008;27:29–39.
80. Galli-Tsinopoulou A, Kyrgios I, Maggana I, Giannopoulou EZ, Kotanidou EP, Stylianou C, et al. Insulin resistance is associated with at least threefold increased risk for prothrombotic state in severely obese youngsters. *Eur J Pediatr.* 2011;170:879–86.
81. Monte SV, Caruana JA, Ghanim H, Sia CL, Korzeniewski K, Schentag JJ, et al. Reduction in endotoxemia, oxidative and inflammatory stress, and insulin resistance after Roux-en-Y gastric bypass surgery in patients with morbid obesity and type 2 diabetes mellitus. *Surgery.* 2012;151:587–93.
82. Bonsignore MR, Esquinas C, Barceló A, Sanchez-de-la-Torre M, Paternó A, Duran-Cantolla J, et al. Metabolic syndrome, insulin resistance and sleepiness in real-life obstructive sleep apnoea. *Eur Respir J.* 2011. doi:10.1183/09031936.00151110.
83. Takaoka M, Nagata D, Kihara S, Shimomura I, Kimura Y, Tabata Y, et al. Periadventitial adipose tissue plays a critical role in vascular remodeling. *Circ Res.* 2009;105:906–11.
84. Rittig K, Staib K, Machann J, Böttcher M, Peter A, Schick F, et al. Perivascular fatty tissue at the brachial artery is linked to insulin resistance but not to local endothelial dysfunction. *Diabetologia.* 2009;51:2093–9.
85. Rittig K, Dolderer JH, Balletshofer B, Machann J, Schick F, Meile T, et al. The secretion pattern of perivascular fat cells is different from that of subcutaneous and visceral fat cells. *Diabetologia.* 2012;55:1514–25. *It helps to understand the difference in the secretosome of perivascular fat cells from subcutaneous and visceral fat.*
86. Cicha I, Wömer A, Urschel K, Beronov K, Goppelt-Struebe M, Verhoeven E, et al. Carotid plaque vulnerability: a positive feedback between hemodynamic and biochemical mechanisms. *Stroke.* 2011;42:3502–10.
87. Stapleton PA, James ME, Goodwill AG, Frisbee JC. Obesity and vascular dysfunction. *Pathophysiology.* 2008;15:79–89.
88. Kim JA, Montagnani M, Koh KK, Quon MJ. Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms. *Circulation.* 2006;113:1888–904.
89. Jonk AM, Houben AJ, Schaper NC, de Leeuw PW, Serné EH, Smulders YM, et al. Meal-related increases in microvascular vasomotion are impaired in obese individuals: a potential mechanism in the pathogenesis of obesity-related insulin resistance. *Diabetes Care.* 2011;34 Suppl 2:S342–8.
90. Levy BI, Ambrosio G, Pries AR, Struijker-Boudier HA. Microcirculation in hypertension: a new target for treatment? *Circulation.* 2001;104:735–40.
91. Maiellaro K, Taylor WR. The role of the adventitia in vascular inflammation. *Cardiovasc Res.* 2007;75:640–8.